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Supporting Information

2 **A disposable and sensitive non-enzymatic glucose sensor**
3 **based on 3D-Mn doped NiO nano-flower modified flexible**
4 **electrode**

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20 **Experimental section**

21 **Materials**

22 Carbon cloth (CFC) was obtained from TORAY Japan. Nickel chloride hexahydrate
23 ($\text{NiCl}_2 \cdot 6\text{H}_2\text{O}$) was purchased from the Kelon Chemical Reagent Factory (Chengdu,
24 China). Manganese chloride tetrahydrate ($\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$) and hexamethylene tetramine ($\text{C}_6\text{H}_{12}\text{N}_4$)
25 were bought from the Cologne Chemicals Co. Ltd (Chengdu, China). Absolute ethanol
26 ($\text{C}_2\text{H}_5\text{OH}$), acetone (CH_3COCH_3) and urea were purchased from the Chuandong Chemical
27 Industry Group (Chongqing, China). Sodium hydroxide (NaOH) was bought from the
28 Sinopharm Chemical Reagent Co. Ltd. Glucose (Glu) was purchased from ACROS.
29 Ascorbic acid (AA), acetylsalicylic acid (ASA), acetaminophen (AMP), NaCl and glycine
30 (Gly) were obtained from Sigma-Aldrich (St. Louis, USA). Dopamine hydrochloride (DA)
31 was purchased from the Yuancheng Chemical Co. Ltd (Wuhan, China). Ultrapure water
32 was produced from a Milli-Q system ($18.25 \text{ M}\Omega \text{ cm}^{-1}$).

33 **Characterization**

34 The structure of the as-prepared Bare CFC, NiO/CFC and Mn-NiO/CFC was
35 characterized by a Nova 400 scanning electron microscope (SEM). The element content
36 and composition of the electrode materials were analyzed quantitatively and qualitatively
37 by EDS. The x-ray diffraction (XRD) analysis was performed using Bruker AXS D5005
38 to obtain the crystal structure and phase information of the samples. The chemical
39 composition and electronic structure of samples were conducted on an ESCALAB250Xi
40 X-ray photoelectron spectroscopy (XPS).

41 **Electrochemical measurements**

42 A CHI 760E (Shanghai, CH Instruments, Inc.) electrochemical workstation was used
43 to conduct all electrochemical measurements. A conventional three-electrode system was
44 employed for the experiments, with the as-prepared Mn-NiO/CFC as the working
45 electrode, a platinum wire as the counter electrode and Ag/AgCl (3 M KCl) as the reference
46 electrode. The techniques of cyclic voltammetry (CV), electrochemical impedance
47 spectroscopy (EIS) and Amperometric *i-t* curves were applied to investigate the
48 electrochemical performance of the prepared Mn-NiO/CFC electrode in 100 mM NaOH

49 solution or 10 mL 5 mM·L⁻¹ [Fe(CN)₆]^{3-/4-} solution (including 0.1 M KCl). In addition, all
50 tests were performed at room temperature.

51 **Electrochemical characterization**

52 The conductivity of the bare CFC, NiO/CFC and Mn-NiO/CFC electrodes was tested
53 by CV and EIS in 10 mL 5 mM·L⁻¹[Fe(CN)₆]^{3-/4-} solution (including 0.1 M KCl). The CVs
54 were carried out at a scan rate 50 mV/s from -0.2 V to 0.6 V. The frequency range
55 investigated by EIS was 10⁻¹ to 10⁵ Hz. The glucose catalytic performance of the three
56 electrodes was tested by amperometry in 10 mL NaOH with adding 150 μM glucose every
57 50 s for seven times. The CVs of Mn-NiO/CFC at different scan rate (25-500 mV/s) were
58 recorded to investigate the reaction kinetics. Furthermore, the scanning potential of CVs
59 varied from 0 to 0.8V, and the scan rate was 50 mV/s.

60 **Optimization**

61 CVs were conducted by dropping 1 mM glucose into different concentrations of
62 NaOH solutions (25, 50, 75, 100, 125, 150 mM) to obtain the optimal concentration of
63 NaOH. The electrocatalytic performance of Mn-NiO/CFC to different concentrations of
64 glucose (0, 1, 2, 4, 6 mM) was tested with CV at optimal concentrations, with a scan rate
65 50 mV/s from 0.2 V to 1.0 V. The optimal working potential and preparation calcination
66 time of the electrode materials were obtained through comparing a series of potentials by
67 adding 150 μM glucose continuously for seven times.

68 **Linear detection research**

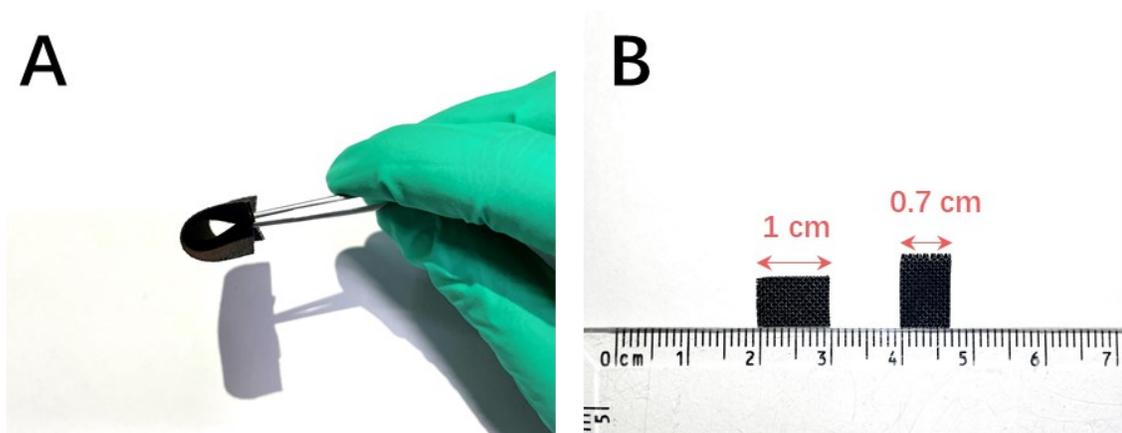
69 At the optimal working potential of 0.6 V, the linear study of the electrode was carried
70 out by amperometry in 10 mL 100 mM NaOH solution system with continuously adding
71 different concentrations of glucose every 50 s, three times for each concentration.

72 **Selectivity and reproducibility study of the Mn-NiO/CFC**

73 Under the optimal experimental conditions, selectivity of the Mn-NiO/CFC was
74 evaluated with continuously adding 500 μM glucose and 50 μM biological interfering
75 species such as AA, UA, Fru, Suc, urea and DA by using amperometric method. The
76 reproducibility of this sensor was confirmed at 0.6 V via assessing the current response of
77 150 μM glucose at five Mn-NiO/CFC electrodes prepared by the same way. Each electrode
78 was repeated three times.

79 **Figure S1:**

80

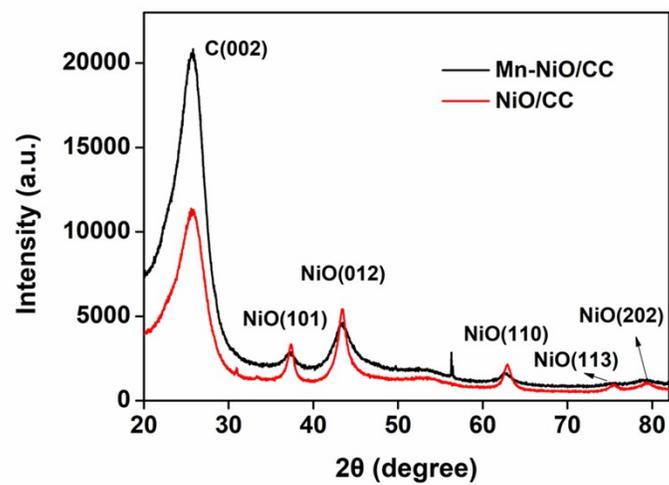


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82 **Figure S1** (A) Images of the prepared flexible CFC ($5 \times 5 \text{ cm}^2$); the CFCs with a size of $1 \times 0.7 \text{ cm}^2$

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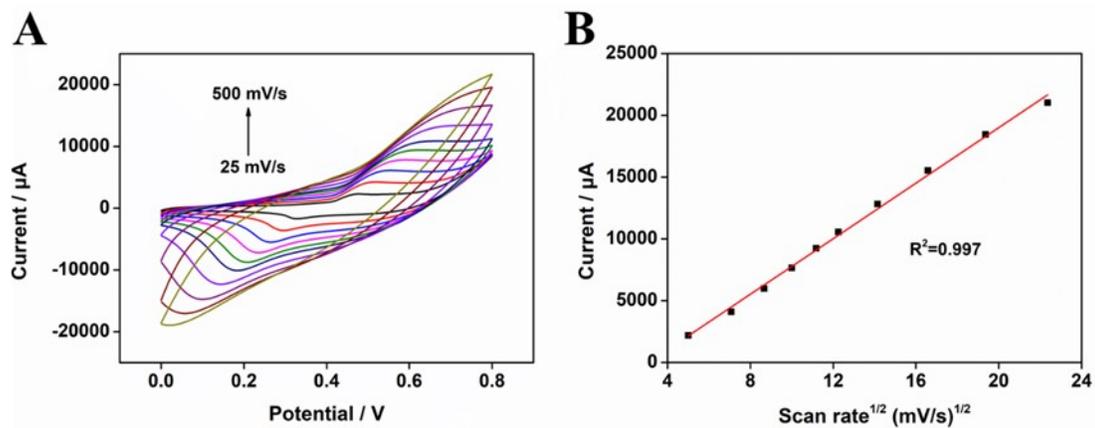
84 **Figure S2:**



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86 **Figure S2** XRD patterns for the Mn-NiO/CFC and NiO/CFC

87 **Figure S3:**



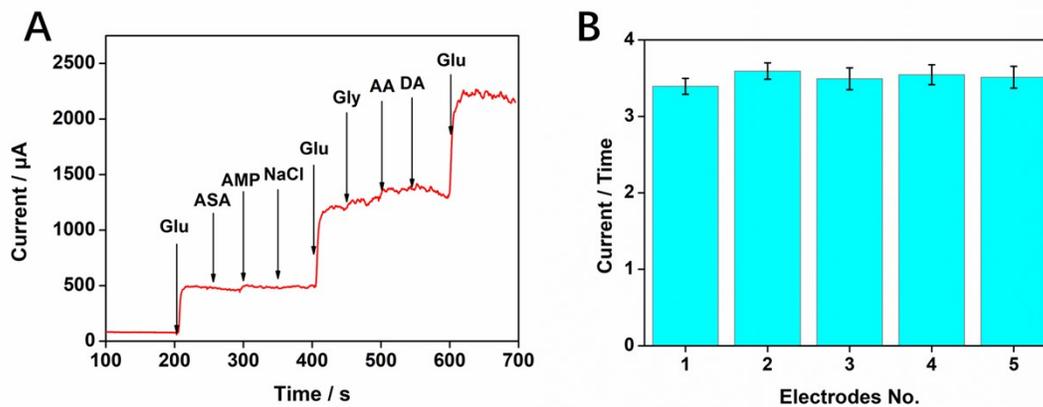
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89 **Figure S3** (A) CVs of the Mn-NiO/CFC at different scan rates (solution: 100 mM NaOH containing 1
90 mM glucose); (B) The linear fitting diagram of its oxidation peak current and the square root of the
91 sweep speed.

92 **Table S1** Comparison of the performance of Mn-NiO /CFC sensor electrode and other sensors of the
 93 similar type for glucose detection.

Electrode materials	Linear range (μM)	LOD (μM)	Documents
Ni ₇ S ₆ /NiO	90-3120	0.3	1
Ni/Cu(OH) ₂ NS	10-1490	1.27	2
Ni/Co-TCPP MOFs	1-3800	0.3	3
Ni/PANI coaxial nanowires	0-7000	10	4
CuO polyhedrons/CFC	0.5-800	0.46	5
Co ₃ O ₄ /CuO NRA/CFC	1-500	0.38	6
MOF-74(Cu) NS-CFC	1-1000	0.41	7
CuO/Ni(OH) ₂ /CFC	50-8500	0.31	8
CuCo ₂ O ₄ @NiCo ₂ O ₄ /CFC	1-1158	0.35	9
GCE/WMCNT/ZnO NPs	100~10000	0.82	10
GCE/CuO NWs-MoS ₂ /Au NPs	0.5~5670	0.5	11
GCE/Hollow cage-like NiO	0.1~5000	0.1	12
Cu ₃ Se ₂ film	2.8~1500	2.8	13
GCE/Cu/ZnO NC	10~7000	57	14
GCE/MXene-Cu ₂ O	10~30000	2.83	15
Mn-NiO/CFC	3-5166	0.28	This work

95 **Figure S4:**



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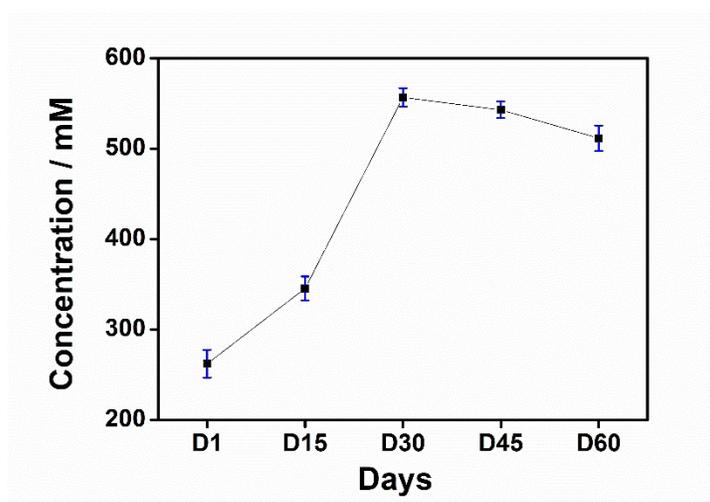
97 **Figure S4 (A)** Amperometric response of the the Mn-NiO/CFC to the selectivity of glucose; **(B)**

98 Reproducibility of five Mn-NiO/CFC electrodes. (The error bar was calculated based on the standard

99 deviation of 3 measurements, n=3)

100

101 **Figure S5:**



102

103 **Figure S5** Determination of glucose in sorghum fermentation broth of different cycle by
104 Mn-NiO/CFC sensors. (The error bar was calculated based on the standard deviation of 3
105 measurements, n=3)

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