Supporting Information

Copper ferrite Nanoparticles loaded on Reduced Graphene Oxide Nanozymes for the Ultrasensitive Colorimetric Assay of Chromium ions

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1. Reagents and apparatus

Ferric nitrate (Fe(NO₃)₃), copper nitrate (Cu(NO₃)₂·3H₂O), Chromic Chloride hexahydrate (CrCl₃·6H₂O), sodium hydroxide (NaOH), potassium permanganate (KMnO₄) were procured from Aladdin Reagent Co.,Ltd (Shanghai, China). NaNO₃, hydrogen peroxide (30% H₂O₂), Sulfuric acid (H₂SO₄, 98%), 3,3′,5,5′ tetramethylbenzidine (TMB) were supplied from Macklin Biochemical Co., Ltd (Shanghai, China). Graphite powder was procured from Tianjin Reagent Factory. All reagents used in this research are analytically grade. Ultrapure water (18.2 MΩ) was used throughout the whole experiments. The working buffer was 0.2 M HAc-NaAc buffer with various pH in the present study.

The morphology of CuFe₂O₄/rGO nanozyme was recorded by TEM (JEM-2100 (JEOL, Japan)). The Fourier transform infrared (FT-IR) spectra were obtained on TENSOR II infrared spectrometer (Bruker Germany). X-ray photoelectron spectroscopy (XPS) of CuFe₂O₄/rGO was obtained on a Thermo Scientific, ESCALAB 250Xi with Al K α radiation (USA). UV-vis spectra were carried out on a Perkin Elmer, Lambda 950 UV-vis spectrophotometer.

2. Results and discussion



Fig. S1 Time dependent absorbance evolution at 652 nm of TMB with different concentrations of $CuFe_2O_4/rGO$.



Fig. S2 UV-vis absorbance spectra of the reaction system at 652 nm at different reaction temperatures (A) and pH (B).

$$\frac{1}{\nu} = \frac{K_m}{V_m} \left(\frac{1}{[S]} + \frac{1}{K_m} \right)$$
 (eq S1)^[1]

where v is the rate of conversion, V_m the maximum rate of conversion, [S] the substrate concentration, and K_m the Michaelis–Menten constant, which is equivalent to the substrate concentration, since the rate of conversion is half of V_m .



Fig. S3 The fluorescence intensity of the H_2O_2 + CuFe₂O₄/rGO systems generated hydroxyl radicals captured by PTA changes with time.



Scheme S2 PTA is combined with hydroxyl radicals to form 2-hydroxy phenylene terephthalic acid.

Due to CuFe₂O₄/rGO can be used as a mimetic enzyme, it is obviously an important work to explore its internal catalytic mechanism. Then, we explore the possible peroxidase-like catalytic mechanism of CuFe₂O₄/rGO by fluorescence method. Simply, the formation of hydroxyl radicals during the reaction of H₂O₂ catalyzed by CuFe₂O₄/rGO was verified by using a p-phthalic acid (PTA) as a fluorescence probe ^[2]. Non-fluorescent PTA is very easy to bind to hydroxyl radicals to form 2-hydroxy terephthalic acid (HTA) having high fluorescent, which typically emits unique blue fluorescence near 436 nm by excitation at 315 nm ^[3] (see in scheme S2). In Fig. S3 indicates the fluorescence intensity of H₂O, PTA-H₂O₂ and PTA-H₂O₂ -catalyst with time. As can be seen, no fluorescence emission was observed without the catalyst. However, in the presence of catalyst, the fluorescence intensity revealed a significant increase with increasing time, confirming that·OH is derived from the decomposition of H₂O₂ by the catalysis of CuFe₂O₄/rGO nanozyme and resulted in the oxidation of PTA to form HTA. This result indicates that the CuFe₂O₄/rGO has high catalytic activity with other materials having a consistent like-peroxidase properties.

At the same time, reactive oxygen species (\cdot OH, O₂⁻⁻, etc.) that may be generated by CuFe₂O₄/rGO in the catalytic H₂O₂ reaction can be proved by radical inhibition experiments. The introduction of TBA as a scavenging agent of \cdot OH can be seen from Figure S4 (A), with the addition of TBA, the absorbance of the solution decreased significantly, further proving the existence of \cdot OH in the system ^[4,5]. PBQ was taken as the scavenging agent of O₂⁻⁻, as shown in Figure S4 (B), the absorbance of the solution did not change significantly, indicating the O₂⁻⁻ was almost not produced during the catalytic oxidation process ^[6]. To further confirm the generation of \cdot OH in the catalytic system, EPR analysis was performed with 5,5- dimethyl-1-pyrroline N-oxide (DMPO),

as presented in Figure S5. In DMPO trapping spectra of the EPR tests, the intensified peaks of DMPO-HO \cdot could be identified, indicating the existence of \cdot OH radicals^[7].



Fig. S4 Absorbance changes of TMB+H₂O₂+CuFe₂O₄/rGO system after addition of different concentrations of TBA and PBQ.



Fig. S5 EPR spectra in H₂O₂+CuFe₂O₄/rGO system with DMPO as the trapping agent.

Table S1. Comparison of kinetic parameters for the oxidation of TMB by different nanomaterials

Catalyst	$K_{\rm m} [{ m mM}]$		$V_{\rm m}[10^{-8}{ m Ms}^{-1}]$		Defeneres
	H_2O_2	TMB	H_2O_2	TMB	
HRP	3.70	0.434	8.71	10.0	[8]
Au/HZIF-8@TCPP(Fe)	1.74	0.79	9.60	7.26	[9]
Cu-MOF	6.41	4.11	10.21	55.56	[10]
Cu-Ag/rGO	8.6245	0.6340	4.2553	7.0175	[11]
Au-NCs+ ZIF-8/CQDs	16.86	13.29	1.429	2.941	[12]
MoS ₂ -Fe ₃ O ₄	2.50	0.35	3.3	35	[13]
CoFe ₂ O ₄	8.89	0.387	1.93	2.90	[14]
Graphene nanoribbons	3.52	0.42	3.09	1.58	[15]
CuFe ₂ O ₄ /rGO	0.453	0.47	1.778	65.1	this work

Table S2 Comparison of differently colorimetric methods for determination of Cr³⁺

Materials	Linear range(µM)	LOD(nM)	Ref.
TADA@AuNPs	0.5-5	5.89	[16]
silver nanoflakes	-	1.4/11.5	[17]
gold nanoparticles	12.5-75	1.15×10 ⁴	[18]
Silver nanopentagons	80-10000	5×10 ⁴	[19]
4-MBA–AuNPs	20-25	5000	[20]
Citrate- and thiourea-AuNPs	-	500	[21]
PMAA@Au NPs	-	4×10^{4}	[22]
NiCNF-RhB	-	203	[23]
AuNBP@Ag NRs	0.01-100	8.7	[24]
N-T/AuNPs	0.5-2.5	30	[25]
MUA-AuNPs	0.0001-1	0.017	[26]
CTP- AgNPs	18.75-62.5	6.25×10 ³	[27]
CuFe ₂ O ₄ /rGO	0.1-25	35	This work

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