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Electronic Supplementary Material

Simple and practical solvent system selection strategy for high-speed countercurrent chromatography based on the HPLC

polarity parameter model

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Column	Reference compound	р	Flow rate (mL/min)	B% (Methanol)	t ₀	t _R	P_m^N	$(\log k)_0$	P_s^N
Calumn 1	Honokiol	4.31			2 150	5.250	0.2267	5.9474	1.5217
Column 1	Magnolol	3.923			2.150	7.267	0.2267		1.5217
Column 2	Honokiol	4.31	1.0	000/	1 714	5.224	0.22(7	4.9171	1 2012
Column 2	Magnolol	3.923	1.0	80%	1./14	6.832	0.2267		1.2013
	Honokiol	4.31			1.000	4.017	0.2267	5.8428	1 22/2
Column 3	Magnolol	3.923			1.000	5.617	0.2267		1.3262

Table S1 Parameters of stationary phase of different columns under the same conditions

			Mobile phase composition (B% ν/ν)																		
Flow rate (mL/min)	Reference compound	р		75 (P_m^N	= 0.2625)			$80 (P_m^N)$	= 0.2267)			85 (P_m^N)	= 0.1922)			90 (P_n^N	n= 0.1588)				
			t ₀	t _R	$(\log k)_0$	P_S^N	t ₀	t _R	$(\log k)_0$	P_S^N	t ₀	t _R	$(\log k)_0$	P_S^N	t ₀	t _R	$(\log k)_0$	P_S^N			
0.8	Honokiol	4.31	1.25	7.483	6 9246	1 4755	1.25	5.033	5 9021	1 2162	1.25	3.633	4 7110	1 1256	1.25	2.817	2 7497	0.0761			
0.8	Magnolol	3.923	1.25	11.217	0.8340	1.4755	7.017	7.017	5.8051 1.5162	1.3102	1.25	4.683	4.7110	1.1330	1.25	3.400	5./48/	0.9761			
0.0	Honokiol	4.31	1 1 1	1 1 1	1 11	6.700	6.700	6 9669	1 4800	1 1 1	4.467	5 01/1	5 9141 1 2102	1 1 1	3.233	4 7002	1 1249	1 1 1	2.517	2 9152	0.0804
0.9	Magnolol	3.923	1.11	10.067	0.0000	1.4809	6.233	6.233	5.8141	1.3193	1.11	4.167	4.7093	1.1348	1.11	3.0	3.050	5.6152 0.9894			
Honc 1.0 Magr	Honokiol	4.31	6.050 1.00 9.083	6.050			4.017 1.00 5.617	4.017				2.900				2.200					
	Magnolol	3.923		9.083	6.8579	1.4779		5.8428	1.3262	1.00	3.733	4.6906 1.	1.1316	1.00	2.633	3.6136	0.9549				

Table S2 Parameters of stationary phase under different flow rates and methanol concentrations for compounds of group 1

							Mobile phase composition (B% ν/ν)													
Flow rate (mL/min)	Reference compound	р	$60 \ ({}^{P_m^N} = 0.3775)$				$65 \left({{P_m^N} = 0.3378} \right)$				$70 (P_m^N = 0.2995)$				75 ($P_m^N = 0.2625$)					
		ind ²	t ₀ (min)	t _R (min)	$(\log k)_0$	P_S^N	t ₀ (min)	t _R (min)	$(\log k)_0$	P_S^N	t ₀ (min)	t _R (min)	$(\log k)_0$	P_S^N	t ₀ (min)	t _R (min)	$(\log k)_0$	P_S^N		
	vitexicarpin	4.31		19.716				16.589				12.500				7.917				
0.8	apigenin	3.923	1.25	33.635	8.9491 1.8	1.8291 1.25	27.635	8.5479 1.739	1.7393	393 1.25	19.917	/.8369	1.6080	1.25	12.000	6.9945	1.4969			
	vitexicarpin	4.31		18.623 .11 31.826	18.623	1.0204	8294 1.11	15.573	0.5640	5649 1.7296	1.11	11.617	20115	1 (120	1.1.1	7.017	7.0000	1 5012		
0.9	apigenin	3.923	1.11		9.0150	1.8294		25.894	8.5649			18.583	7.9115	1.6138	1.11	10.650	7.0099	1.5013		
	vitexicarpin	4.31		17.603	17.603	17.603				14.457				10.150				6.283		
1.0	apigenin	apigenin 3.923	1.00	29.946	9.0000	1.8138	1.00	24.089	8.6119	1.7328	1.00	16.333	7.9632	1.6335	1.00	9.500	6.9609	1.4913		

Table S3 Parameters of stationary phase under different flow rates and methanol concentrations for compounds of group 2

Mobile phase composition (B%, v/v)	Flow rate (mL/min)	Column temperature (°C)	UV detector temperature (°C)	Wavelengths of the UV detector (nm)
90% Methanal	1.0	30	30	254, 280, 365
60% Methanal	1.0	30	30	254, 280, 365

 Table S4
 HPLC analysis conditions for mangosteen peel and H. sampsonii Hance

	Compound 1			Compound 2	
Position	$\delta_{\mathrm{H}}(J \text{ in Hz})$	δς	Position	$\delta_{\rm H}(J \text{ in Hz})$	δς
1		160.1	1		
2		109.9	2		157.8
3		162.1	3		134.8
4	6.20, s	91.7	4		178.2
4a		154.7	5		161.8
4b		156.4	6	6.22, d, <i>J</i> = 1.9 Hz	93.3
5	6.65, s	101.3	7		164.6
6		155.2	8	6.38, d, <i>J</i> = 2.0 Hz	98.4
7		143.3	9		157.1
			10		104.4
8		136.9	1′		121.5
8a		110.7	2′	7.36, d, $J = 2.0$ Hz	114.9
8b		102.3	3′		145.0
9		181.6	4′		148.4

Table S5 ¹H-NMR (4000MHz) data for Compound 1 and 2

1′	3.27, d, <i>J</i> = 7.2 Hz	25.7	5′	6.93, d, <i>J</i> = 8.2 Hz	115.5
2'	5.24, m	123.8	6′	7.33, d, <i>J</i> = 8.4 Hz	121.4
3′		130.2	1′′	5.37, s	102.1
4′	1.67, s	24.6	2′′		70.6
5′	1.79, s	16.5	3''		70.7
1′′	4.04, d, <i>J</i> = 4.6 Hz	20.8	4′′		71.8
2′′	5.24, m	122.5	5''		70.5
3''		130.2	6''	0.96, d, J = 6.0 Hz	16.2
4′′	1.68, s	24.6			
5′′	1.82, s	16.9			
OCH ₃ -7	3.75, s	59.9			



Figure S1. Chemical structure of standard compounds.



Figure S2. Representative HPLC chromatograms. Experimental conditions: column: Agilent ZORBAX SB-C18 column (5 μ m particle size, 4.6 × 250 mm, marked as column 1); mobile phase: 80% methanol; flow rate: 1.0 mL/min; column temperature: 30°C; detector temperature: 30°C; detector



Figure S3. The HPLC chromatogram of mangosteen peel. Experimental conditions: column: Kromasil 100-5-C18 column (5 μ m particle size, 4.6 × 150 mm); mobile phase: 90% methanol; column temperature: 30°C; detector temperature: 30°C; detection wavelength: 254 nm.



Figure S4. The HPLC chromatogram of *H. sampsonii* Hance. Experimental conditions: column: Agilent Eclipse XDB-C18 column (5 μ m particle size, 9.4 × 250 mm); mobile phase: 60% methanol; column temperature: 30°C; detector temperature: 30°C; detection wavelength: 254 nm.



Figure S5. The HSCCC chromatogram of mangosteen peel and *H. sampsonii* Hance. Experimental conditions: solvent system: n-hexane-ethyl acetate-methanol-water (6:3:2:1 for mangosteen peel and 1:4:1:4 for *H. sampsonii* Hance, v/v/v/v); resolution speed: 900 rpm; separation temperature: 30°C; detection wavelength: 254 nm. The peaks marked in two chromatogram are mangostin and quercitrin.



Figure S6. The structure of mangostin and quercitrin.



Figure S7. ¹H-NMR (400 MHz, CDCl3) Spectrum of Compound 1.



Figure S8. ¹³C-NMR (100 MHz, CDCl₃) Spectrum of Compound 1.



Figure S9. ¹H-NMR (400 MHz, CD₃OD) Spectrum of Compound 2.



Figure S10. ¹H-NMR (100 MHz, CD₃OD) Spectrum of Compound 2.