Supporting Information

An allosteric DNA switch-mediated catalytic DNA circuit for ratiometric and sensitive nucleic acid detection

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Name	Sequence from 5' to 3'			
Α	Cy5-GCCTGACGTATAAAGTGAGGTTGATGG			
в	CACCTGCCTGACCCTCACTTTATACGTCAGGCT-BHQ- GTCCTTTCTCCTTGCGAATC			
С	GATTCGCAAGGAGAAAGGAC-Cy3-TTAGGTG			
D	CCATCGATTCGCGTCCTTTCTCCTTGCGAATCCCTCACTTTATACGTCAGGC			
Target	CCATCAACCTCACTTTATACGTCAGGC			
D2	CCATCGATTCGCGTCCTTTCTCCTTGCGAATCTTCTGTCCCGCGTACTGAAT			
1MT-3	CCTTCAACCTCACTTTATACGTCAGGC			
1MT-7	CCATCA <u>T</u> CCTCACTTTATACGTCAGGC			
1MT-13	CCATCAACCTCATTTTATACGTCAGGC			
1MT-22	CCATCAACCTCACTTTATACG <u>A</u> CAGGC			
2MT	CCATCAACATCACTTTAGACGTCAGGC			

Table S1. The used nucleic acid sequences in the experiment



Figure S1. Fluorescence experiments investigating each separate step (I to III)of the proposed allosteric DNA switch-mediated catalytic DNA circuit strategy.(A) Fluorescence responses of Cy5 for probe AB and its mixing with targetDNA. (B) Fluorescence responses of Cy3 for probe CD and its mixing withstrand B. (C) Fluorescence responses of Cy5 for probe AB and its mixing withstrand D. Inset in Figure S1 A to C shows the corresponding workingmechanismforeachstep.



Figure S2. Gel electrophoresis characterizations for the allosteric DNA switch mediated catalytic DNA circuit reaction strategy: lane 1, 20 bp DNA ladder; lane 2, hybrid AB; lane 3, hybrid CD; lane 4, hybrid BC; lane 5, hybrid AD; lane 6, hybrids AB and CD; lane 7, target DNA; lane 8, hybrids AB and CD in the presence of target DNA. The DNA concentrations used in the lanes were 200 nM except the target DNA concentration of 10 nM in lane 8.

Table S2. Detection performance comparison between current sensing

Ref.	Detection	Detection	Strategy		
	limit	range	Strategy		
[1]	0.7 nM	2.0 nM – 40 nM	Hybridization chain reaction and fluorescence resonance energy transfer		
[2]	10 pM	10 pM – 1 nM	Bimolecular beacons-based enzyme-free strategy		
[3]	10 pM	10 pM – 400 nM	Target-catalyzed dynamic assembly and pyrene excimer		
[4]	220 fM	500 fM – 10 nM	Entropy-driven strand displacement and DNAzyme		
[5]	7.0 pM	10 pM – 1nM	Entropy-driven target recycling and hybridization		
			chain reaction		
[6]	13 pM	0.1 nM – 50 nM	Helper probes-based strand displacement and G-		
[0]			quadruplex		
[7]	1 pM	1 pM – 250 pM	Hybridization-triggered DNAzyme cascade		
[8]	56 pM	500 pM – 20 nM	Catalyzed hairpin assembly		
[9]	1.9 pM	10 pM – 1 µM	Cascade toehold-mediated strand displacement		
[10]	0.58 pM	0.5 pM – 3 pM	Helper probes-assisted target recycling		
[11]	5 pM	5 pM –10 nM	Entropy beacon		
[12]	72 pM	0.5 nM – 50nM	Catalyzed hairpin assembly		
This work	0.1 pM	0.1 pM – 100 nM	Allosteric DNA switch-medicated reciprocal strand		
			displacement amplification		

system and those reported enzyme-free fluorescence methods.



Figure S3. Schematic illustration of allosteric DNA switch-mediated strand displacement strategy. In this case, the strand D2 was used as the substitute of strand D. The released D2 from hybrid probe CD2 could not further interact with probe AB via toehold-mediated strand displacement reaction for recycling amplification. It thus could be denoted in current study as an allosteric DNA switch-mediated strand displacement strategy and used for the performance comparison with the developed allosteric DNA switch-mediated catalytic DNA circuit strategy.



Figure S4. Time dependence of fluorescence responses of Cy5 (A) and Cy3 (B) toward 1 nM target DNA by using the allosteric DNA switch-mediated catalytic DNA circuit strategy (red or blue squares) and the allosteric DNA switch-mediated strand displacement strategy (black circle).



Figure S5. Fluorescence spectra of Cy5 (A) and Cy3 (B) for the sensing system fabricated by the allosteric DNA switch-mediated strand displacement strategy toward different concentrations of target DNA. The concentrations for the curves a to g were 0, 1, 10, 100 pM, 1, 10, 100 nM, respectively. (C) The calibration curves for fluorescence intensities of Cy5 and Cy3 versus target DNA concentration. (D) The relationship of the fluorescence ratios of Cy5 to Cy3 with different target DNA concentrations.

Samples	Added (pM)	Detected (pM)	Recovery
1	5.00×10 ¹	(4.14 ± 0.59) ×10 ¹	83%
2	5.00×10 ²	$(5.38 \pm 0.47) \times 10^2$	107%
3	5.00×10 ³	(5.99 ± 0.42) ×10 ³	119%

 Table S3. Recovery experiments of the sensing system toward spiked target

DNA in diluted serum

The results were based on three repetitive experiments.

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