

SUPPORTING INFORMATION

Electroconductive and biocompatible hydrogels based on gelatin and PEDOT:PSS for a mini-invasive approach in neural tissue regeneration

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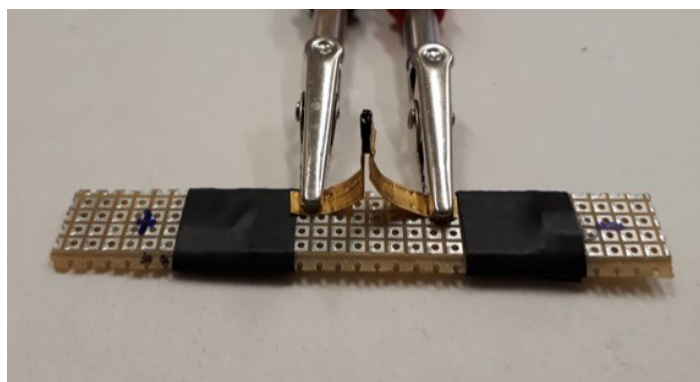
monica.sandri@istec.cnr.it

Gelatin % (w/w respect to the total weight of the sample)	Genipin % (w/w respect to gelatin)	Gelation (yes/no)
5	0.25	no
5	0.5	no
5	1	no
10	0.1	no
10	0.2	no
10	0.5	no
10	1	yes

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25 **Table S1.** Composition of samples tested. Concentrations of gelatin, genipin and features of resulting
26 samples at 37°C are reported.

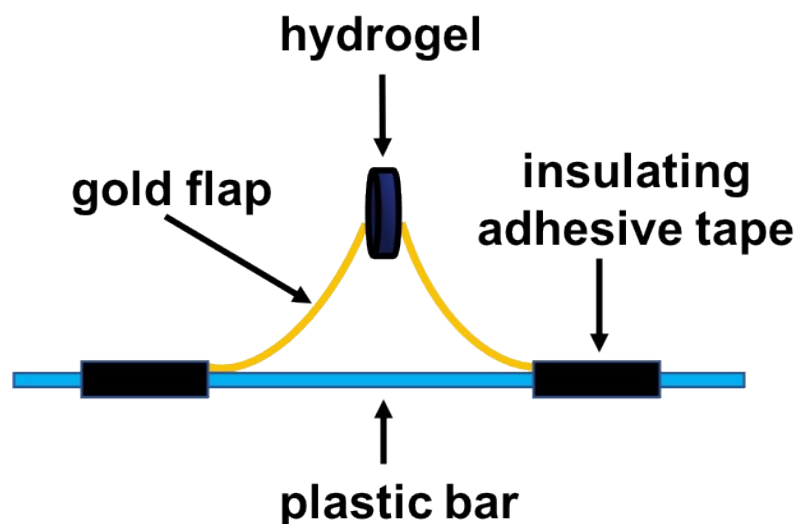
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29 **Figure S1.** Experimental set-up used for Electrochemical Impedance Spectroscopy (EIS) analyses. The
30 electroconductive hydrogel sample was tested between the gold electrodes. Experimental conditions:
31 [gelatin] = 10% (w/w respect to the total weight of the sample), [genipin] = 0.1% (w/w respect to gelatin),
32 and [PEDOT:PSS] = 0.3% (w/w respect to the total weight of the sample).

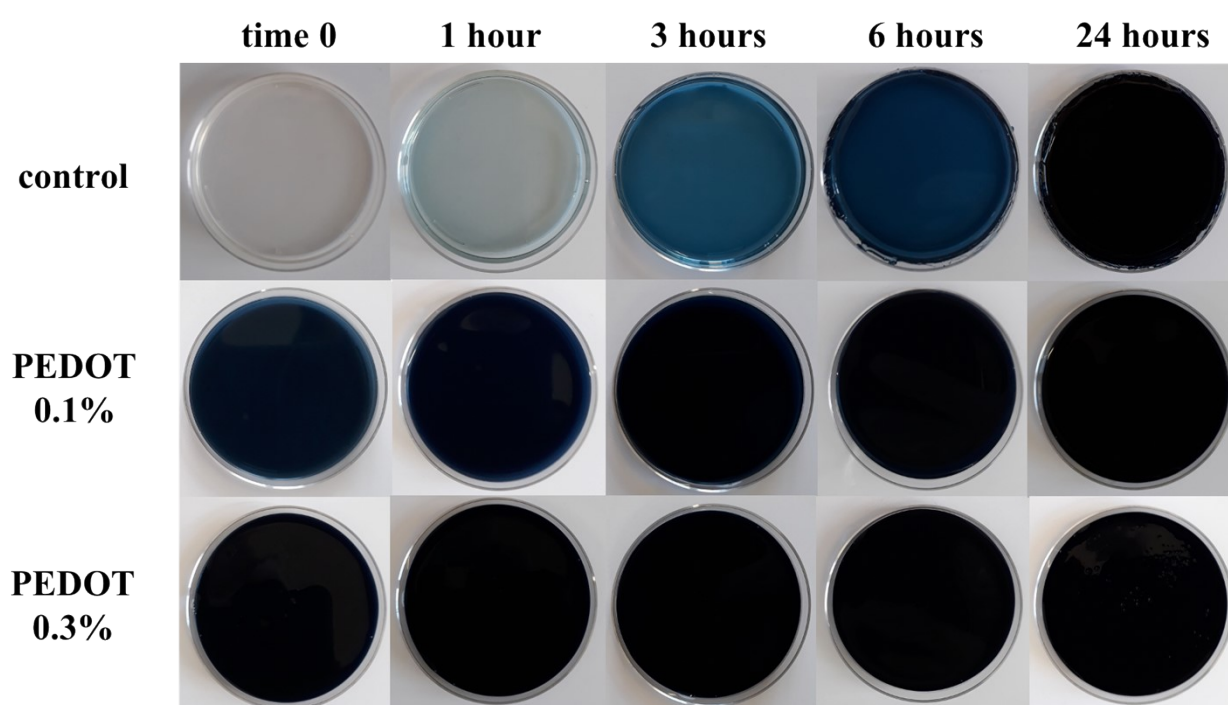
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35 **Scheme S1.** Schematic representation of experimental set-up used for Electrochemical Impedance
 36 Spectroscopy analyses.

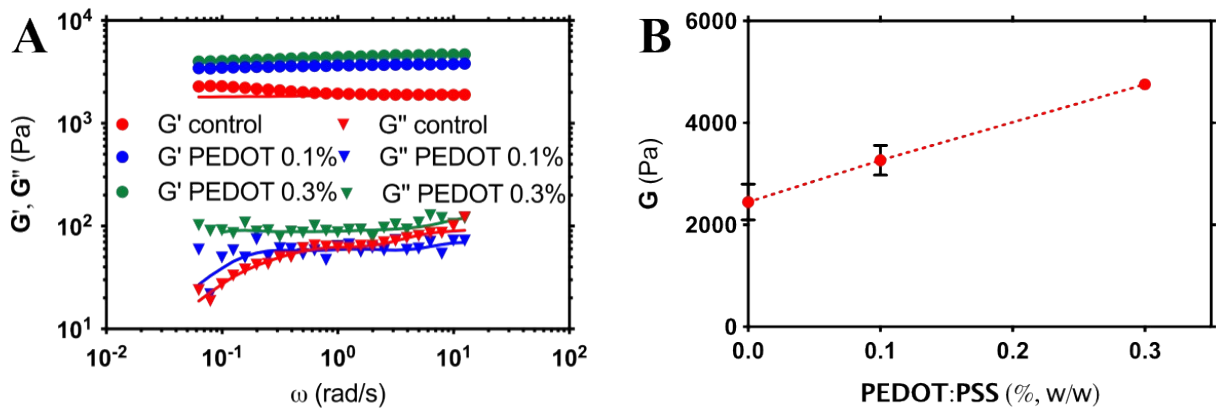
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39 **Figure S2.** Visual analyses of gelatin-based matrices at different timeframes, *i.e.* immediately after genipin
 40 addition (time 0) and after 1, 3, 6 and 24 hours. Experimental conditions: [gelatin] = 10% (w/w respect to the
 41 total weight of the sample), [genipin] = 1% (w/w respect to gelatin), and [PEDOT:PSS] = 0 - 0.3% (w/w
 42 respect to the total weight of the sample), $T = 37^\circ\text{C}$.

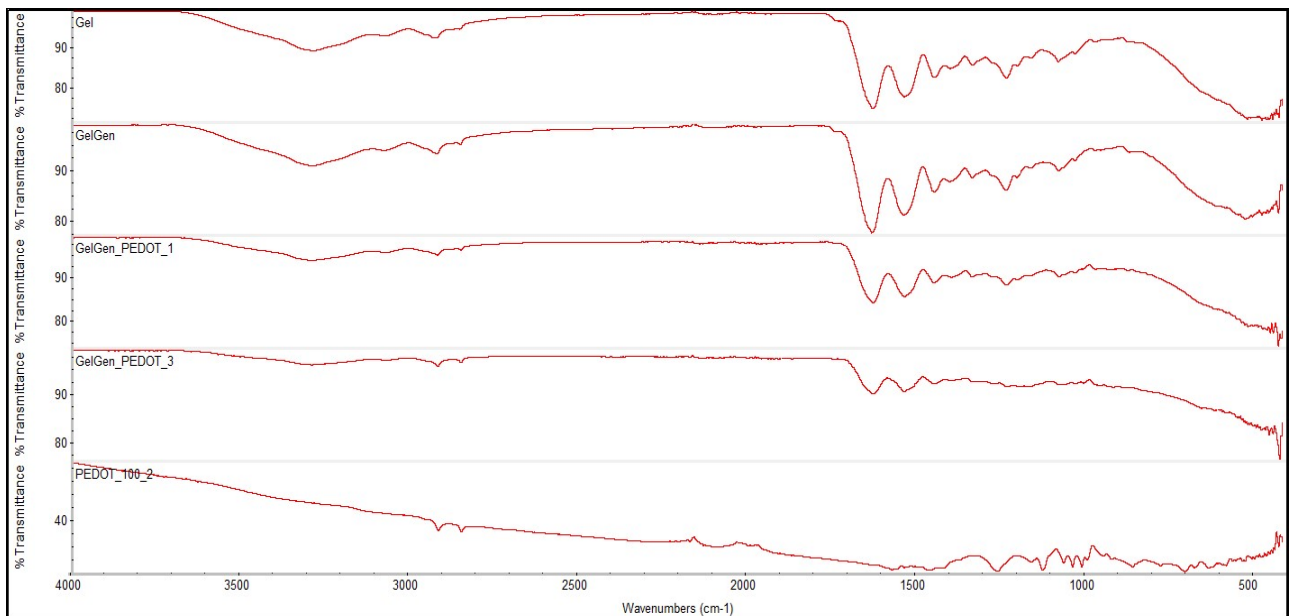
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45 **Figure S3.** Mechanical characterization by rheology. (A) Dependence of storage (G' , circles) and loss
 46 moduli (G'' , triangles) on angular frequency, ω , for gelatin-based hydrogels with different amounts of
 47 PEDOT:PSS: [PEDOT:PSS] = 0% (w/w) (red), [PEDOT:PSS] = 0.1% (w/w) (blue), and [PEDOT:PSS] =
 48 0.3% (w/w) (green). Solid lines are the best fit of experimental points according to eq. 1 and 2. (B)
 49 Dependence of the shear modulus, G , on the amount of PEDOT:PSS. G values were determined according to
 50 eq. 3. data are reported as means \pm standard deviations (SD) of at least three measurements. The dotted line
 51 is drawn to guide the eye. Experimental conditions: [gelatin] = 10% (w/w respect to the total weight of the
 52 sample), [genipin] = 1% (w/w respect to gelatin), and [PEDOT:PSS] = 0 - 0.3% (w/w respect to the total
 53 weight of the sample), $T = 37^\circ\text{C}$.

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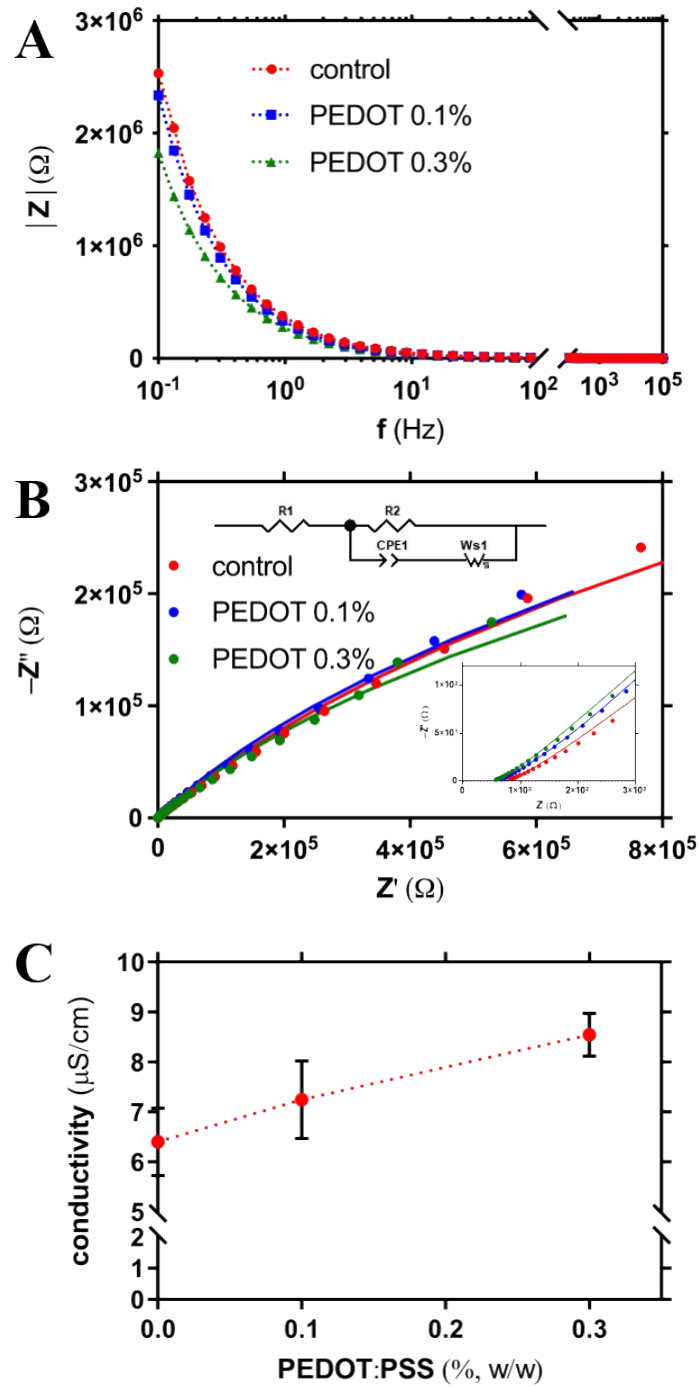


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56 **Figure S4.** Stacked FTIR-ATR of freeze-dried samples. From the top to the bottom are reported samples
 57 based on gelatin (Gel), genipin-crosslinked gelatin (GelGen), genipin-crosslinked gelatin (GelGen) with
 58 PEDOT:PSS = 0.1% w/w (GelGen_PEDOT_1), genipin-crosslinked gelatin (GelGen) with PEDOT:PSS =

59 0.3% w/w (GelGen_PEDOT_3), and pure PEDOT:PSS (PEDOT_100_2).

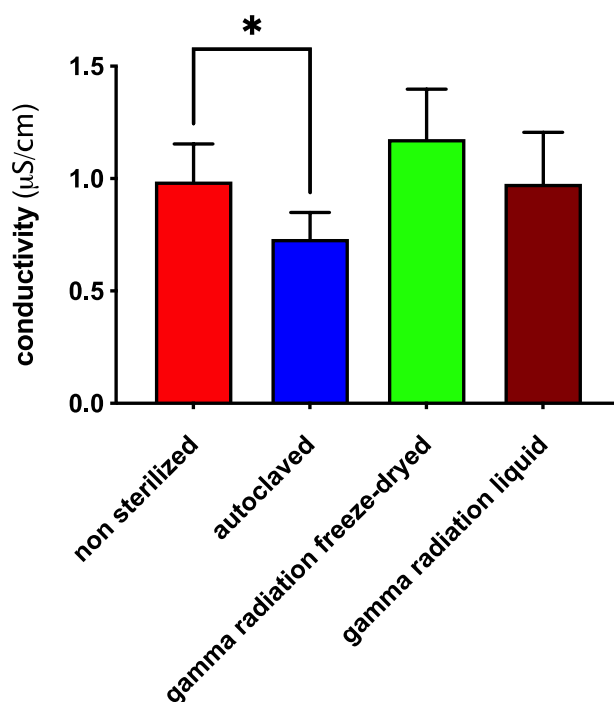
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62 **Figure S5.** (A) Electrochemical Impedance Spectroscopy measurements of gelatin-based hydrogels with
63 different amounts of PEDOT: [PEDOT:PSS] = 0% (w/w) (red), [PEDOT:PSS] = 0.1% (w/w) (blue), and
64 [PEDOT:PSS] = 0.3% (w/w) (green). (B) Nyquist plot for gelatin-based hydrogels with different amounts of
65 PEDOT: [PEDOT:PSS] = 0% (w/w) (red), [PEDOT:PSS] = 0.1% (w/w) (blue), and [PEDOT:PSS] = 0.3%
66 (w/w) (green). Colored solid lines are the best fit of experimental points. (C) Dependence of the electronic
67 conductivity on the amount of PEDOT. Data are reported as means \pm standard deviations (SD) of seven
68 measurements. All dotted lines are drawn to guide the eye. Experimental conditions: [gelatin] = 10% (w/w
69 respect to the total weight of the sample), [genipin] = 1% (w/w respect to gelatin), and [PEDOT:PSS] = 0 -
70 0.3% (w/w respect to the total weight of the sample), PBS 1X as solvent, T = room temperature.

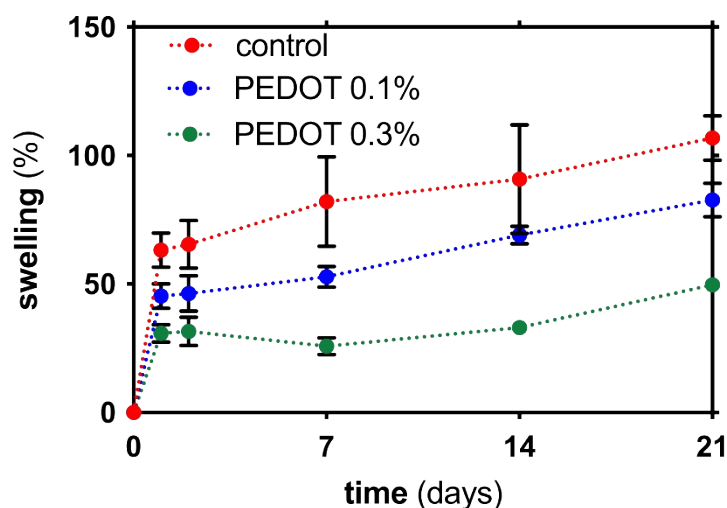
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73 **Figure S6.** Dependence of conductivity on different sterilization techniques for PEDOT:PSS. Experimental
 74 conditions: [gelatin] = 10% (w/w respect to the total weight of the sample), [genipin] = 1% (w/w respect to
 75 gelatin), and [PEDOT:PSS] = 0.3% (w/w respect to the total weight of the sample). One-way ANOVA
 76 followed by Dunnett post-hoc test was performed to compare all groups (*: p value < 0.05). In all cases, data
 77 are reported as means \pm standard deviations (SD) of at least five measurements.

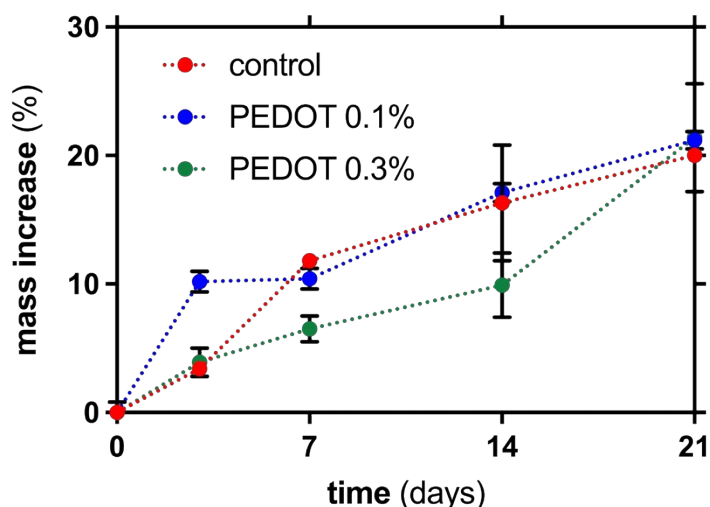
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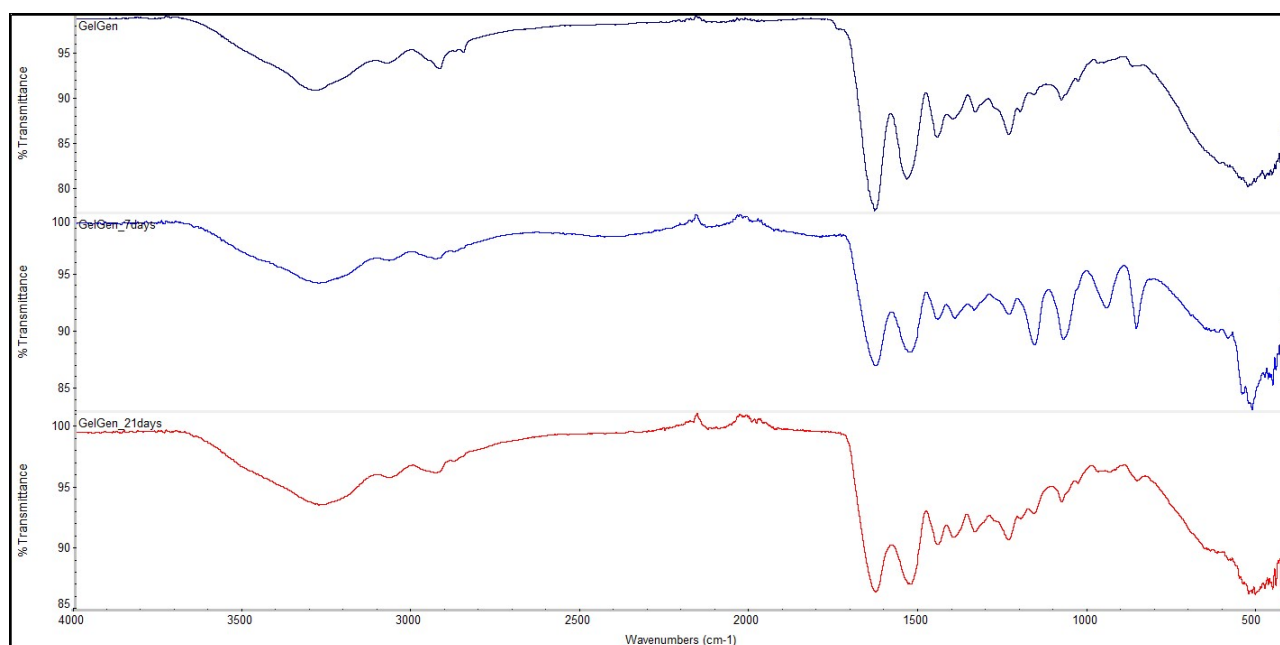
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80 **Figure S7.** Dependence of swelling, *i.e.* PBS uptake, on time of gelatin-based hydrogels with different
 81 amounts of PEDOT:PSS: [PEDOT:PSS] = 0% (w/w) (red), [PEDOT:PSS] = 0.1% (w/w) (blue), and
 82 [PEDOT:PSS] = 0.3% (w/w) (green). Experimental conditions: [gelatin] = 10% (w/w respect to the total
 83 weight of the sample), [genipin] = 1% (w/w respect to gelatin), and [PEDOT:PSS] = 0 - 0.3% (w/w respect

84 to the total weight of the sample), $T = 37\text{ }^{\circ}\text{C}$. In all cases, data are reported as means \pm standard deviations
 85 (SD) of three measurements.
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 88 **Figure S8.** Dependence of mass increase on time of gelatin-based hydrogels with different amounts of
 89 PEDOT:PSS: [PEDOT:PSS] = 0% (w/w) (red), [PEDOT:PSS] = 0.1% (w/w) (blue), and [PEDOT:PSS] =
 90 0.3% (w/w) (green). Experimental conditions: [gelatin] = 10% (w/w respect to the total weight of the
 91 sample), [genipin] = 1% (w/w respect to gelatin), and [PEDOT:PSS] = 0 - 0.3% (w/w respect to the total
 92 weight of the sample), $T = 37\text{ }^{\circ}\text{C}$. In all cases, data are reported as means \pm standard deviations (SD) of two
 93 measurements.
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 97 **Figure S9.** Stacked FTIR-ATR of genipin-crosslinked gelatin hydrogels used for degradation test and freeze-
 98 dried. From the top to the bottom are reported samples based on the native sample and the same sample after
 99 the incubation for 1 (7 days) and 3 weeks (21 days) in PBS. Experimental conditions: [gelatin] = 10% (w/w
 100 respect to the total weight of the sample), [genipin] = 1% (w/w respect to gelatin).
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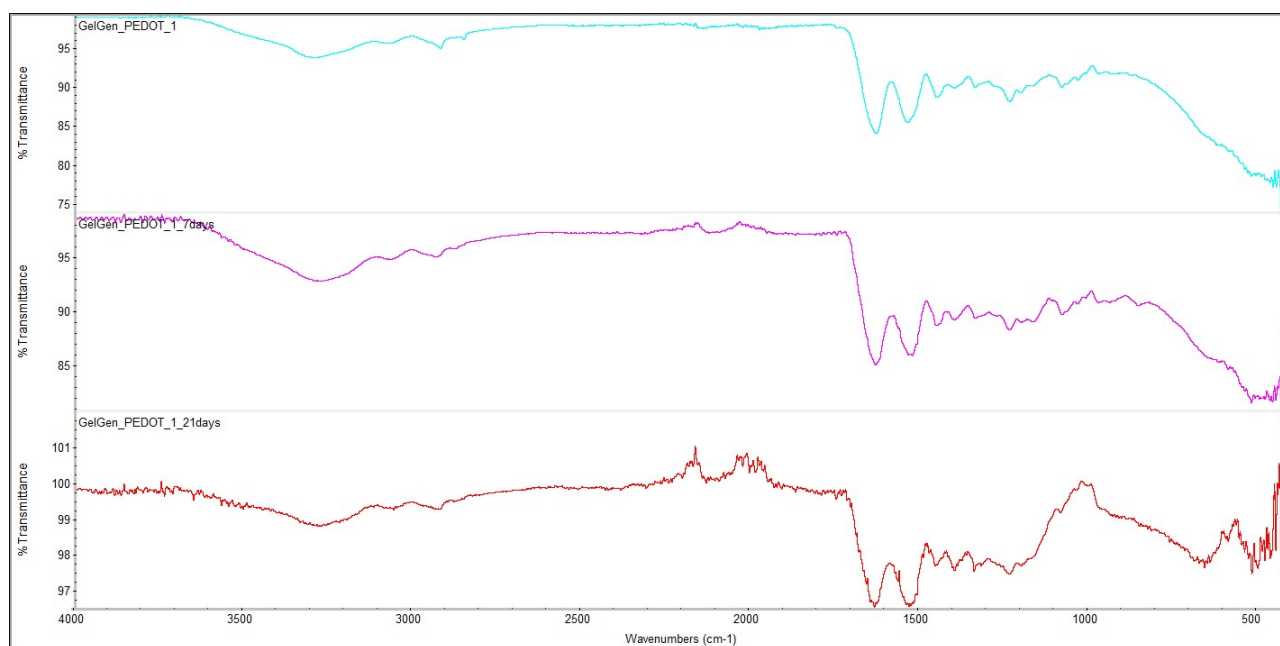


Figure S10. Stacked FTIR-ATR of genipin-crosslinked gelatin hydrogels used for degradation test and freeze-dried. From the top to the bottom are reported samples based on the native sample and the same sample after the incubation for 1 (7 days) and 3 weeks (21 days) in PBS. Experimental conditions: [gelatin] = 10% (w/w respect to the total weight of the sample), [genipin] = 1% (w/w respect to gelatin), and [PEDOT:PSS] = 0.1% (w/w respect to the total weight of the sample).

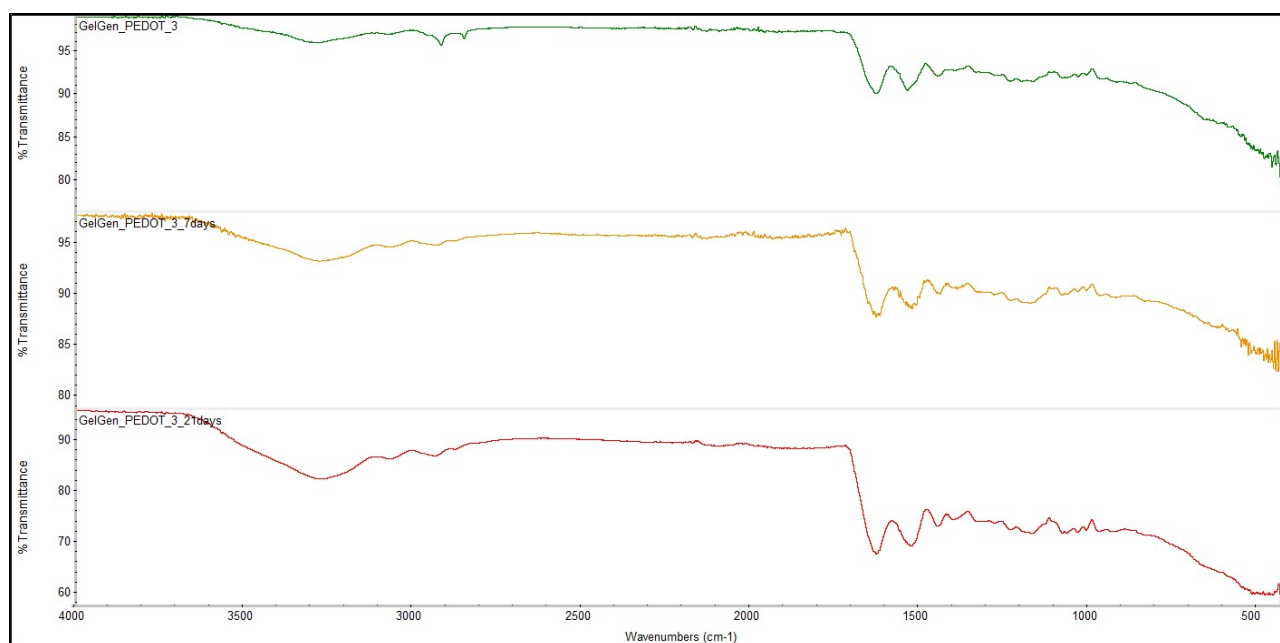
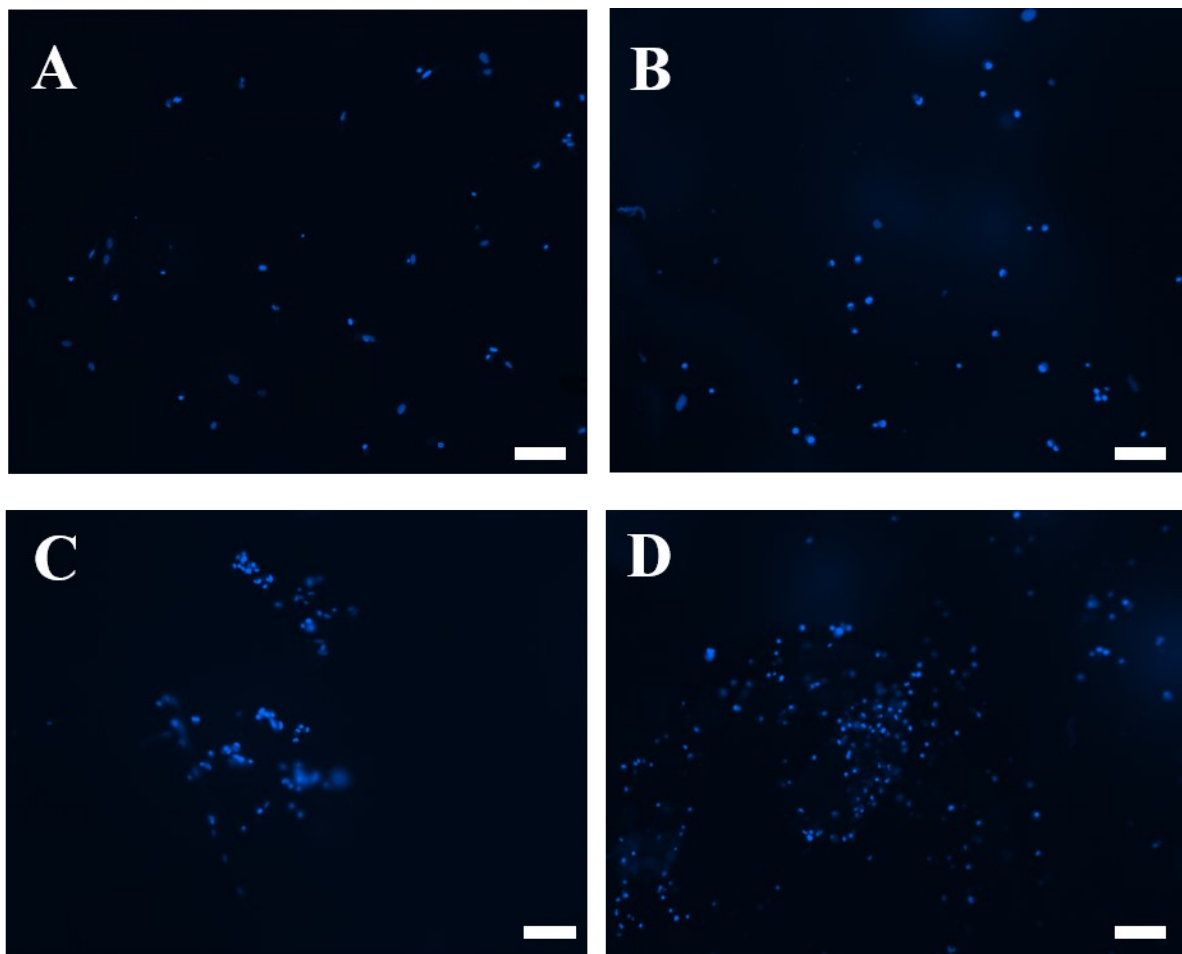
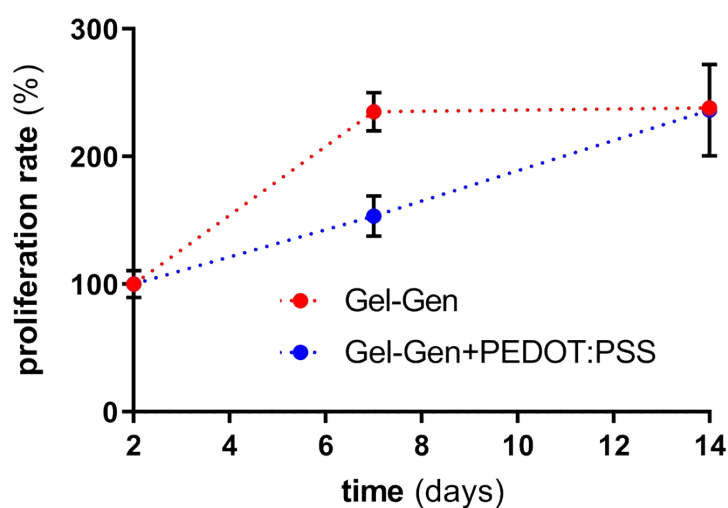


Figure S11. Stacked FTIR-ATR of genipin-crosslinked gelatin hydrogels used for degradation test and freeze-dried. From the top to the bottom are reported samples based on the native sample and the same sample after the incubation for 1 (7 days) and 3 weeks (21 days) in PBS. Experimental conditions: [gelatin] = 10% (w/w respect to the total weight of the sample), [genipin] = 1% (w/w respect to gelatin), and [PEDOT:PSS] = 0.3% (w/w respect to the total weight of the sample).



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121 **Figure S12.** Fluorescence microscopy analyses of astrocytes after 2 (**A, B**) and 7 days (**C, D**) from cells
122 seeding on control hydrogels (**A, C**) and on electroconductive hydrogels (**B, D**). DAPI staining of astrocytes'
123 nuclei is revealed in blue. The scale bar stands for 100 μm for all figures.
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125
126 **Figure S13.** Proliferation rate of astrocytes on electroconductive and control as function of time by Alamar
127 blue assay. Data are plotted as the averaged percentages of reduced AB \pm Standard Error (SE).
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