Supporting Information

In Situ Self-Assembly of Polydopamine inside Injectable Hydrogels: Antibacterial Activity and Photothermal Therapy for Superbug-Infected Wound Healing

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Figure S1. Schematic image of preparing specimens underwater.



Figure S2. The standard curve of the production of ROS.



Figure S3. Photographs of colour changes of the GCPP-PDA gel along with the continuous self-assembly of dopamine.



Figure S4. Photographs of the coagulation effect of control and GCPP-PDA gel in whole blood.



Figure S5. a) UV-vis spectra of PDA/PBS solution with different concentration (1 μ g~100 μ g). b) The standard curve of the PDA. c) UV-vis spectra of gel/PBS solution with different extraction times at 37 °C, the volume of PBS was three time of the gel.



Figure S6. The self-assembly of PDA NPs. The steps are presented in the electropolymerization representing the starting reaction for PDA formation.



Figure S7. The ABTS⁺ scavenging activity of the PDA NPs at different assembly time.



Figure S8. a) The stable PAM-PDA solution (positive charged) presents pink. b) The SEM image of the PAM-PDA solution (vacuum freeze drying), and the hydrodynamic diameter of the PDA nanoparticles in the PAM-PDA solution. c) FT-IR test results of PAM and PAM-PDA.



Figure S9. Raman spectra of Dopamine, PAM, PAM-PDA.



Figure S10. a) H₂O₂ generated by GCPP-PDA gel after self-assembly for different time at 37 °C.
b) The antibacterial rates of the GCPP-PDA gel after immersing in PBS for different time.



Figure S11. Crystal violet staining image and its corresponding absorbance for integrated MRSA biofilm incubated with the SC-PDA NPs and the GCPP-PDA gel. The biofilm without incubation with the sample was used as the control.



Figure S11. EDS of the GCPP-PDA gel.



Figure S12. The FT-IR spectra of the samples.



Figure S13. Photograph of the injectable GCPP-PDA hydrogel.



Figure S14. Rheological property of the GCPP-PDA gel. a) Strain sweep measurements of G' (storage modulus) and G" (loss modulus) of the GCPP-PDA gel. b) Dynamic step-strain measurements of G' (storage modulus) and G" (loss modulus) under repeated deformation of 1% strain and 100% strain. c) G' (storage modulus) and G" (loss modulus) of the GCPP-PDA gel with different self-assembly time of PDA.



Figure S15. TGA curves of a series of the GCPP-PDA gels with different self-assembly time of dopamine.



Figure S16. The microscopy images of the MC3T3-E1 cells cultured in the presence of unconditioned media and GCPP-PDA gel conditioned medium for 72 hours, respectively.



Figure S17. The growth curves of the MC3T3-E cells cultured in each group by the the Incucyte Live-Cell Analysis system.



Figure S18. The immunogenicity results of the GCPP-PDA gel using CFSE as maker and analysing by flow cytometry (IV ConA as positive group, culture medium as negative, PE antimouse CD 3 and PE Rat IgG2b Isotype as antibodies).



Figure S19. Photographs of MRSA colonies after treatments with PBS, GCPP gel, GCPP-Van gel, GCPP-PDA gel, GCPP-PDA gel (NIR) *in vivo*.

Group	Rabbit blood	GCPP-PDA	CaCl ₂	
	(natrium citricum)	gel	(0.25 M)	Clotting time
1	1 mL	0 mg	0 µL	Non
2	1 mL	0 mg	100 µL	365 s
3	1 mL	50 mg	0 µL	780 s
4	1 mL	50 mg	100 µL	180 s

Table S1. Clotting time of the GCPP-PDA gel and Ca^{2+} to whole blood.