Supporting Information

Dual-Responsive and NIR-Driven Free Radical Nanoamplifier with Glutathione Depletion for Enhanced Tumor-Specific Photothermal/Thermodynamic/Chemodynamic Synergistic Therapy

Fanghui Chen,^a Xichen Zhang,^a Zining Wang,^a Chensen Xu,^a Jinzhong Hu,^a Ling Liu, ^{*,c} Jiancheng Zhou, ^{*,a} Baiwang Sun, ^{*,a,b}

^a School of Chemistry and Chemical Engineering, Southeast University, Nanjing
 211189, China

^b Jiangsu Province Hi-Tech Key Laboratory for Biomedical Research, Southeast University, Nanjing 211189, China

 ^c Department of Infectious Diseases, Hospital of Integrated Traditional Chinese and Western Medicine Affiliated with Nanjing University of Chinese Medicine, Nanjing 210028, China

* Corresponding authors, E-mail addresses: lingliumed@163.com (Ling Liu); jczhou@seu.edu.cn (Jiancheng Zhou); chmsunbw@seu.edu.cn (Baiwang Sun)

Supplementary Methods

Materials. 1,3,5-trimethylbenzene (TMB), dopamine hydrochloride, ammonia hydroxide (NH₄OH, 28-30 wt%), Tris-HCl buffer (pH 8.5), methylene blue (MB), glutathione (GSH), 2,2'-azobis[2-(2-imidazolin-2-yl) propane] dihydrochloride (AIPH), hydrogen peroxide (H₂O₂, 30 wt%), and fluorescein isothiocyanate (FITC) were purchased from Aladdin Biochemical Technology Co. Ltd. (Shanghai, China). Pluronic F127, copper (II) chloride (CuCl₂), tannic acid (TA), hyaluronate (HA), 5,5'dithiobis (2-nitrobenzoic acid) (DTNB), indocyanine green (ICG), and 2,2'-azinobis(3-ethylbenzothiazoline-6-sulfonicacid) (ABTS) were acquired from Sigma-Aldrich Trading Co. Ltd. (Shanghai, China). RPMI-1640 medium, DMEM medium, fetal bovine serum (FBS), phosphate-buffered saline (PBS), calcein-AM/propidium iodide (PI) stain kit, 2',7'-dichlorodihydrofluorescein diacetate (DCFH-DA), Annexin V-FITC/PI apoptosis detection kit, 4',6-Diamidino-2-phenylindole (DAPI), Lyso-Tracker Red, GSH assay kit, and 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) were obtained from Jiangsu KeyGen Biotech. Co. Ltd. (Nanjing, China).

Calculation of the Photothermal Conversion Efficiency. In order to evaluate the photothermal conversion efficiency (η), the MACTH dispersion (100 µg mL⁻¹, 1.0 mL) was irradiated with 808 nm laser (1.0 W cm⁻²) for 600 s, followed by switching off the laser and naturally cooling to ambient temperature. The photothermal conversion efficiency was calculated by the following equation:

$$\eta = \frac{hS(T_{max} - T_{sur}) - Q_{dis}}{I(1 - 10^{-A_{808}})}$$

Where *h* is the heat transfer coefficient, *S* is the surface area of the container, T_{max} is the maximum equilibrium temperature of the sample solution, T_{sur} is the surrounding temperature, Q_{dis} is the heat generated by the container and solution under laser irradiation, *I* is the laser power, and A_{808} is the absorbance of the sample solution at 808 nm.

Cell Culture. Human breast adenocarcinoma cell line (MDA-MB-231) and human normal liver cell line (LO2) were obtained from the Cell Bank of the Chinese Academy of Science (Shanghai, China). The MDA-MB-231 and LO2 cells were cultured in DMEM and RPMI-1640 medium respectively, containing 10% FBS and 1% penicillin/streptomycin at 37 °C under 5% of CO₂.

Supplementary Figures



Fig. S1. (a) N_2 absorption-desorption isotherms and (b) corresponding pore size distribution of MPDA, MACT and MACTH.



Fig. S2. (a) UV-vis absorption spectra of AIPH at different concentrations. (b) Standard curve of AIPH (absorbance *vs.* concentration).



Fig. S3. (a) TGA curves and (b) normalized weight loss diagram of MPDA, MA, MACT and MACTH.



Fig. S4. The hydrodynamic size changes of MACTH in various physiological media, including water, PBS, DMEM and FBS for 7 days.



Fig. S5. Normalized absorbance of MACTH aqueous solution at 808 nm with different concentrations.



Fig. S6. (a) TEM images and (b) hydrodynamic size distributions of MACTH after various treatments for 24 h (scale bars: 100 nm).



Fig. S7. GSH depletion by MACT (50 µg mL⁻¹) at different time points.



Fig. S8. UV-vis absorption spectra of MB after treatment with different samples (AIPH:
20.3 μg mL⁻¹, MACT: 200 μg mL⁻¹, H₂O₂: 8 mM).



Fig. S9. MB degradation by GSH-treated MACT plus different concentrations of H_2O_2 at pH 6.0 (MACT: 200 μ g mL⁻¹, GSH: 2 mM).



Fig. S10. Generation of ABTS⁺⁻ as induced by the free radicals produced from AIPH at different temperatures and time points.



Fig. S11. CLSM images of MDA-MB-231 cells treated with FITC-labeled MACT and

MACTH (scale bars: 50 μ m).



Fig. S12. GSH level in MDA-MB-231 cells after different treatments.



Fig. S13. The quantitative mean fluorescence intensity (MFI) of intracellular DCF corresponding to Fig. 4c.



Fig. S14. Cell viability of LO2 and MDA-MB-231 cells after treatment with gradient concentrations of MACTH for 24 h.



Fig. S15. Fluorescence intensity of the tumor sites at different time points corresponding to Fig. 5b.



Fig. S16. Fluorescence intensity of major organs and tumor corresponding to Fig. 5c.



Fig. S17. Temperature elevation curves of tumor regions after various treatments corresponding to Fig. 5d.



Fig. S18. Tumor inhibition rate of MDA-MB-231 tumor-bearing nude mice with various treatments.



Fig. S19. Histopathological examination of the main organs after different treatments (scale bar: $100 \ \mu m$).