Electronic Supplementary Material (ESI) for Biomaterials Science. This journal is © The Royal Society of Chemistry 2022

Supplementary Information

The impact of Gd-DOTA spatial distribution within PLGA-PEG micelles encapsulated IR-1061 on bimodal over–1000 nm near-infrared fluorescence and magnetic resonance imaging

Thi Kim Dung Doan,^{*ab} Masakazu Umezawa,^c Kyohei Okubo,^c Masao Kamimura,^c Masayuki

Yamaguchi,^b Hirofumi Fujii,^b Kohei Soga.^{ac}

 ^a Research Institute for Biomedical Science, Tokyo University of Science, 2669 Yamazaki, Noda, Chiba 278-0022 Japan.
 ^b Division of Functional Imaging, Exploratory Oncology Research & Clinical Trial Center, National Cancer Center, 6-5-1 Kashiwanoha, Kashiwa 277-8577, Japan.

^cDepartment of Material Science and Technology, Tokyo University of Science, 6-3-1 Niijuku, Katsushika-ku, Tokyo 125-8585, Japan.

Methods

Micelle preparation

 $D[m(\%)] = \frac{m(D)}{m[(D)] + m[(PLGA - PEG)]} \times 100 \ (1)$

PL-D-P/PL-N-P micelles: PLP–D were mixed with PLGA–PEG at various special concentrations (D [m (%)]) from the equation (1): where D = PL-D-P/PL-N-P, and [m (%)] = 70 % of 30 mg (total amount of polymers). 30 mg of mixed polymers were dissolved in a solution containing 996 μ l ACN and 4 μ l IR–1061 in ACN (5 mg/ml), and 2 mL of Milli-Q water was added into PL-D-P samples. The solutions were stirred for about 20 hours to evaporate the ACN. Ultimately, all products were purified using a gel filtration column in water.

Micelle characterization

OTN–NIR absorption: An ultraviolet/ visible/ NIR spectrophotometer (V770, JASCO, Inc., Japan) was used to measure the absorption spectra.

OTN–NIR emission: A spectrometer (NIR–256–1.7, Avantes, Apeldoorn, The Netherlands) equipped with a fiber-coupled laser diode (SP–976–5–1015–7, Laser Components GmBH, Olching, Germany) of 980–nm excitation source was used to obtain the emission spectra. The optical set-up includes a 980–nm excitation source (0.67 W) irradiated the solution sample in a 10–mm–path–length cuvette, and the emission was collected perpendicularly to the excitation plane in a Q–pod (Quantum Northwest, WA, USA) integrated with NIR spectroscopy.

(a)

$$Yield (\%) = \frac{weight of PLP - D}{weight of PLGA - PEG - COOH + Gd - DOTA} \times 100 (1.a)$$

(b)

$$Efficience (\%) = \frac{mol \ number \ of \ Gd(III)ion}{mol \ number \ of \ PLP - D} \times 100 \ (1.b)$$

(c)

(d)

Conjugation	Yield ^a (%)	Conjugation efficiency ^b (%)
PLP-D	>37	>90
PL-D-P	>30	>95
PL-N-P	>56	>96

^a Yield = weight of polymer before and after conjugation; ^b efficiency = Gd molar number per molecular weight

Micelles	Estimated Gd (III) mass (mg/mL) ª	Estimated Gd (III) mass (mg/mL) ^b	Estimated Gd ion [%] ^c	Actual Gd ion [%]) ^d
PLP-D	0.23	0.168±0.01	70	514
PL-D-P	0.23	0.198±0.01	70	60
PLP	0	0	0	0

^a Gd (III) mass (calculated)/10mg of micelle ^b actual Gd (III) mass (from ICP) /10mg of the micelle. ^c Gd (III) wt% (calculated)/10mg of the micelle. ^d Actual Gd (III) wt% (from ICP) /10mg of the micelle.

Figure S.1: Yield (a) and efficiency (b) formulas and (c) actual values from ICP. (d) Gd ion estimated and the actual values from ICP.



Figure S.2 (a) The cross-coordination induced by Coulomb interaction between Gd-DOTA molecules at extremely close contact. The structure of (no Gd)-DOTA (b) and Gd-DOTA (c). The absorption of PL-N-P (d) and PL-D-P (e) in water and albumin. [PL-N-P made from PLGA-PEG conjugated (no Gd) - DOTA in the middle, PL-D-P made from PLGA-PEG conjugated Gd -DOTA in the middle.] The fluorescence loss (g) difference in mouse serum between PL-N-P [70] and PL-D-P [70].



Figure S.3: (a) OTN-NIR images of PL-D-P[70] and (b) the corresponding emission intensity in four media observed within 3 days. (c-f) Absorption spectra of the probe in four media were observed within 3 days.



Figure S.4: (a) OTN-NIR image of the mouse injected by PL-D-P [70] at 5 min and the plot profile (b) of the blood vessel corresponding to the fluorescence image.