Supplemental Information

RT-qPCR as a screening platform for mutational and small molecule impacts on structural stability of RNA tertiary structures

Martina Zafferani; Dhanasheel Muralidharan; Nadeska I. Montalvan; Amanda E. Hargrove*

Department of Chemistry, Duke University, 124 Science Drive, Durham, NC 27705, USA

*Corresponding Author; Contact: amanda.hargrove@duke.edu

Table S1. Nucleic acid sequences for synthesis of the RNA constructs. SHAPE cassette added to the 3'-end of MALAT1 is shown in red.

Name	Sequence
MALAT1-WT	GAAGGTTTTTCTTTCCTGAGAAAACAACACGTATTGTTTTCTCAGGTTTTG
DNA template	CTTTTTGGCCTTTTTCTAGCTTAAAAAAAAAAAAAGCAAAATCGATCCGGTT
-	CGCCGGATCCAAATCGGGCTTCGGTCCGGTTC
MALAT1-WT	GAAATTAATACGACTCACTATAGGAAGGTTTTTCTTTCCTGAGAAAACAAC
Forward Primer	ACGTATT
MALAT1 SHAPE	mG[mA]ACCGGACCGAAGCCC
cassette Reverse	
Primer	
MALAT1-U13C	GGAAGGTTTTTCTCTTCCTGAGAAAACAACACGTATTGTTTTCTCAGGTTT
DNA template	TGCTTTTTGGCCTTTTCTAGCTTAAAAAAAAAAAAAAAA
	ATCGATCCGGTTCGCCGGATCCAAATCGGGCTTCGGTCCGGTTC
MALAT1-U13C	GAAATTAATACGACTCACTATAGGAAGGTTTTTCTCTTCCTGAGAAAACAAC
Forward Primer	ACGTATT
SARS-CoV-2	GCAGCTTAATACGACTCACTATAGTTTGCGGTGTAAGTGCAGCCCGTCTTA
pseudoknot	CACCGTGCGGCACAGGCACTAGTACTGATGTCGTATACAGGGCTTTTGAC
' template	ATC
SARS-CoV-2	GCAGCTTAATACGACTCACTATAGTTTGCGGTGTAAGTGC
nseudoknot	
forward perimer	
SAPS CoV 2	mGmATGTCAAAAGCCCTGTATACGACATCAGTACTAGTGCC
SANS-COV-2	
pseudoknot	
reverse primer	



Figure S1. Representation of structure and sequence of the SHAPE cassette (orange) as reported by Wilkinson and co-workers.¹



Figure S2. Purity analysis of constructs used in the study. (A) Agilent DNA high-resolution Chip of the PCR products indicate purify of all constructs. The samples are arranged as SARS-CoV-2 pseudoknot, MALAT1 wild-type, and MALAT1 mutant, respectively. (B) Small RNA Agilent chip of the RNA constructs used in the RT-qPCR assay. Samples are arranged as MALAT1 wild type, MALAT1 mutant, and MALAT1 triple helix without the SHAPE cassette for reference, respectively. (C) 10% TBE denaturing gel with the RNA constructs used in the RT-qPCR assay. Samples are arranged as MALAT1 mutant, SARS-CoV-2 pseudoknot, and ladder, respectively.



Figure S3. Raw curve of *z*'-factor experiment with amplification of MALAT1_WT or control. The experiment was run in two independent replicates with 10 wells of RNA and 10 wells of controls (no RNA) each. Z-factor was calculated as previously reported.²



Figure S4. Denatured control of the MALAT1 wild type ensures the RT-qPCR assay reports on construct structuredness. The MALAT1 WT sample was either not annealed (light blue), denatured in 100μ M DTT and 10μ M RNA prior to dilution to 100nM and addition to the plate, or annealed as performed with small molecules treatment. Decrease in C_t value of the not annealed and DTT-denatured conditions confirm that the assay reports on and is influenced by structuredness of the construct.

References

1. Wilkinson, K. A.; Merino, E. J.; Weeks, K. M., Selective 2'-hydroxyl acylation analyzed by primer extension (SHAPE): quantitative RNA structure analysis at single nucleotide resolution. *Nature Protocols* **2006**, *1* (3), 1610-1616.

2. Patwardhan, N. N.; Cai, Z.; Newson, C. N.; Hargrove, A. E., Fluorescent peptide displacement as a general assay for screening small molecule libraries against RNA. *Org Biomol Chem* **2019**, *17* (7), 1778-1786.