### Fac-to-mer isomerization triggers hydride transfer from Mn(I) complex fac-[(Ph<sub>2</sub>PCH<sub>2</sub>PPh<sub>2</sub>)Mn(CO)<sub>3</sub>H]

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#### General considerations and synthesis of Mn(I) hydride complexes.

All manipulations were performed using standard Schlenk techniques under an atmosphere of dry nitrogen or argon. Dry and oxygen-free organic solvents (toluene,  $CH_2Cl_2$ ) used for synthesis were obtained using LabSolv (Innovative Technology) solvent purification system. Commercially available dry ethanol was degassed by nitrogen bubbling for 10 min and stored under 4 Å molecular sieves.  $CH_2Cl_2$  and *n*BuCl used for IR spectroscopy studies were stored over  $CaH_2$  and distilled under an argon atmosphere prior to use. Deuterated solvents for NMR were degassed before use by three freeze-pump-thaw cycles and kept over 3 Å molecular sieves. A liquid nitrogen/ethanol or isopropanol slush bath was used to maintain samples at the desired low temperature. Manganese hydride complexes *fac*-[(dppm)Mn(CO)<sub>3</sub>H] (*fac*-1) and *fac*-[(dppmD<sub>2</sub>)Mn(CO)<sub>3</sub>D] (*fac*-1-*d*<sub>3</sub>) were prepared from the corresponding bromide precursor *fac*-[(dppm)Mn(CO)<sub>3</sub>Br]<sup>1</sup> and NaBH<sub>4</sub> using the modified literature procedure.<sup>2</sup> Commercially available B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> and [Ph<sub>3</sub>C](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) were purified by vacuum sublimation and recrystallization from CH<sub>2</sub>Cl<sub>2</sub>/hexane mixture, respectively. All other reagent grade chemicals purchased from commercial sources were used as received.

Routine <sup>1</sup>H, <sup>31</sup>P and <sup>13</sup>C NMR spectra were recorded on Bruker Avance 400 and Avance III HD 400 spectrometers at 400.1, 162.0 and 100.6 MHz, respectively. Low-temperature NMR experiments were carried out using Bruker Avance NEO 600 spectrometer in the 183–303 K temperature range. <sup>1</sup>H and <sup>13</sup>C signals reported in parts per million (ppm) were calibrated against the residual signals of the deuterated solvent,<sup>3</sup> whereas <sup>31</sup>P and <sup>11</sup>B NMR spectra were referenced to 85% H<sub>3</sub>PO<sub>4</sub> and BF<sub>3</sub>×OEt<sub>2</sub>, respectively. When required, signal attribution in <sup>13</sup>C spectra was based on decoupled <sup>13</sup>C{<sup>31</sup>P, <sup>1</sup>H} and <sup>13</sup>C–<sup>1</sup>H HSQC experiments. Variable temperature (160–293 K) IR spectra were recorded on Nicolet 6700 spectrometer using a home-modified cryostat (Carl Zeiss Jena, the accuracy of temperature adjustment ±0.5 K) and 0.05 cm CaF<sub>2</sub> cells.

**Modified procedure for the synthesis** *fac*-[(Ph<sub>2</sub>PCH<sub>2</sub>PPh<sub>2</sub>)Mn(CO)<sub>3</sub>H] (*fac*-1). A solution of (dppm)Mn(CO)<sub>5</sub>Br (1.1 g, 1.82 mmol) and NaBH<sub>4</sub> (345 mg, 9.12 mmol) in EtOH (30 mL) were heated at 80 °C during 1 h until the disappearance of v<sub>co</sub> bands of the precursor ( $v_{co}$  2020 (vs), 1950 (s), 1917 (s) cm<sup>-1</sup>). The resulting suspension was cooled to room temperature and the volatiles were removed under vacuum. The product was extracted with toluene (3×15 mL) and combined extracts were filtered through Celite and evaporated under reduced pressure. The residue was dissolved in CH<sub>2</sub>Cl<sub>2</sub> (10 mL), the solution was again filtered through Celite and hexane (60 mL) was added under stirring. The resulting suspension was concentrated to *ca*. 30% of the initial volume and the supernatant was removed by decantation. The precipitate was washed with hexane (2×20 mL) and dried under vacuum to afford *fac*-1 (860 mg, 90%) as pale-yellow solid.

*fac*-1: <sup>1</sup>H NMR (400.1 MHz, 298 K, CD<sub>2</sub>Cl<sub>2</sub>):  $\delta$  7.70–7.65 (m, 4H, CH<sub>Ar</sub>), 7.59–7.54 (m, 4H, CH<sub>Ar</sub>), 7.43–7.41 (m, 12H, CH<sub>Ar</sub>), 4.42 (dtd, <sup>2</sup>J<sub>HH</sub> = 15.0 Hz, <sup>2</sup>J<sub>PH</sub> = 9.4 Hz, <sup>4</sup>J<sub>HH</sub> = 5.5 Hz, 1H, PCH<sub>2</sub>P), 4.03 (dt, <sup>2</sup>J<sub>HH</sub> = 15.2 Hz, <sup>2</sup>J<sub>PH</sub> = 11.0 Hz, 1H, PCH<sub>2</sub>P), -5.56 (td, <sup>2</sup>J<sub>PH</sub> = 44.0 Hz, <sup>4</sup>J<sub>HH</sub> = 5.5 Hz, 1H, Mn–H). <sup>31</sup>P{<sup>1</sup>H} NMR (162.0 MHz, 298 K, CD<sub>2</sub>Cl<sub>2</sub>):  $\delta$  32.6 (br s). IR (CH<sub>2</sub>Cl<sub>2</sub>): *v*<sub>CO</sub> 1995, 1915, 1905 cm<sup>-1</sup>. IR (*n*BuCl): *v*<sub>CO</sub> 1993, 1911, 1902 cm<sup>-1</sup>. IR (toluene): *v*<sub>CO</sub>: 1996, 1916, 1909 cm<sup>-1</sup>.

**Synthesis** *fac*-[(Ph<sub>2</sub>PCD<sub>2</sub>PPh<sub>2</sub>)Mn(CO)<sub>3</sub>D] (*fac*-1-*d*<sub>3</sub>). A solution of (dppm)Mn(CO)<sub>5</sub>Br (60 mg, 0.10 mmol) and NaBH<sub>4</sub> (21 mg, 0.50 mmol) in EtOD (2 mL) was heated at 80 °C during 30 min and then kept overnight at 50 °C. The resulting suspension was cooled to room temperature and the volatiles were removed under vacuum. The product was extracted with toluene (3×5 mL), combined extracts were filtered through Celiteand evaporated under reduced pressure. The residue was triturated with pentane (2×2 mL) and dried under vacuum to afford *fac*-1-*d*<sub>3</sub> (31 mg, 59%) as pale-yellow solid. According to <sup>1</sup>H NMR data hydride and PCH<sub>2</sub>P positions had more than 94% of deuterium incorporation.

*fac*-**1**-*d*<sub>3</sub>: <sup>1</sup>H NMR (400.1 MHz, C<sub>6</sub>D<sub>6</sub>, 298 K):  $\delta$  7.71–7.66 (m, 4H, *CH*<sub>Ar</sub>), 7.46–7.41 (m, 4H, *CH*<sub>Ar</sub>), 7.03–6.92 (m, 12*H*, CH<sub>Ar</sub>). <sup>31</sup>P{<sup>1</sup>H} NMR (162.0 MHz, C<sub>6</sub>D<sub>6</sub>, 298 K):  $\delta$  34.0 (s). <sup>13</sup>C{<sup>1</sup>H}{<sup>31</sup>P} NMR (100.6 MHz, C<sub>6</sub>D<sub>6</sub>, 298 K):  $\delta$  225.1 (br s, Mn–CO), 222.4 (br s, Mn–CO), 138.0, 135.8 (s, *C<sub>ipso</sub>* PPh<sub>2</sub>), 132.1, 130.3, 130.2, 128.8 (s, *CH*<sub>Ar</sub>), 48.0 (quint, <sup>1</sup>*J*<sub>CD</sub> = 21.7 Hz, PCD<sub>2</sub>P). IR (CH<sub>2</sub>Cl<sub>2</sub>): *v*<sub>CO</sub> 1993, 1911, 1899 cm<sup>-1</sup>. IR (*n*BuCl): *v*<sub>CO</sub> 1995, 1915, 1904 cm<sup>-1</sup>.

#### General procedure for variable temperature IR studies.

The solution (c = 0.003 M) of fac-[(dppm)Mn(CO)<sub>3</sub>H] (fac-1, 1.3 mg) prepared at room temperature in 0.8 mL of the corresponding solvent (CH<sub>2</sub>Cl<sub>2</sub>, *n*BuCl, toluene) was cannulated under an inert atmosphere into the cryostat and cooled to 160 K or 180 K. After the acquisition of reference IR spectrum of initial complex, the solution was removed from the cryostat to the Schlenk tube and cooled to 160 K or 180 K in liquid nitrogen/*i*PrOH or EtOH slush bath. The corresponding Lewis acid ([Ph<sub>3</sub>C](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>): 2.2 mg, 1 equiv.; B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>: 1.2-1.8 mg, 1-1.5 equiv.) dissolved in a small amount of solvent (0.05-0.1 mL) was then added at low temperature and the resulting mixture was quickly mixed and returned into the cryostat and monitored by IR spectroscopy in the 160–290 K temperature range.



**Figure S1**. IR spectroscopy monitoring of the reaction of complex *fac*-**1** with 1 equiv. of  $[Ph_3C](BAr_4)$ , Ar =  $C_6F_5$  in CH<sub>2</sub>Cl<sub>2</sub> at 180-300 K temperature range (*c* = 0.003 M, *l* = 0.05 cm).



**Figure S2**. IR spectroscopy monitoring of the reaction of complex *fac*-**1** with 1 equiv. of  $[Ph_3C](BAr_4)$  (Ar =  $C_6F_5$ ) in *n*BuCl at 160-300 K temperature range (*c* = 0.004 M, *l* = 0.05 cm).



**Figure S3**. IR spectrum of the mixture *of fac*-**1** with 1.5 equiv. of BAr<sub>3</sub> (Ar =  $C_6F_5$ ) in CH<sub>2</sub>Cl<sub>2</sub> at 180 K with the mathematical subtraction of the initial hydride spectrum and its peak deconvolution.

# Determination of thermodynamic characteristics of hydride transfer from *fac*-1 to B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> using low temperature IR studies.

For the reaction of *fac*-**1** with 1.3 equiv. of B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> in toluene experimental equilibrium constants were obtained by IR spectroscopy at 210-240 K range (Figure S4, Table S1). Every temperature point was kept for 20 minutes. Thermodynamic parameters of hydride transfer ( $\Delta H^\circ = -3.3$  kcal/mol,  $\Delta S^\circ = 5.5$  cal/(mol·K) were obtained by Van't-Hoff method from the RlnK<sub>p</sub> – 1/*T* dependency (Figure S5) according to the formula:  $\Delta H/T - \Delta S = -R \cdot \ln K_p$ 



**Figure S4**. Temperature dependence of intensity of  $v_{CO}$  bands of *fac*-**1** and *mer*-[**2**](BHAr<sub>3</sub>) (Ar = C<sub>6</sub>F<sub>5</sub>). Experimental conditions: *fac*-**1** (*c* = 0.004 M), B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> (*c* = 0.005 M), toluene, 190-220 K, *l* = 0.05 cm.

**Table S1**. Experimental equilibrium constants for  $fac-1 + B(C_6F_5)_3 \leftrightarrow mer-[2](BH(C_6F_5)_3)$  process obtained from low temperature IR spectroscopy data.



Figure S5. RInK vs. 1/T plot for determination of thermodynamic parameters for hydride transfer.

# Determination of kinetic parameters of hydride transfer from *fac*-1 and *fac*-1- $d_3$ to B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> using low temperature IR studies

The interaction between fac-[(dppm)Mn(CO)<sub>3</sub>H] (fac-1) and BAr<sub>3</sub> (Ar = C<sub>6</sub>F<sub>5</sub>) as a Lewis acid could be described as the second order reaction with two pre-equilibria ( $K_1$  and  $K_2$ , see below), since the formation of mer-[(dppm)Mn(CO)<sub>3</sub>( $\kappa^1$ (Cl)-nBuCl)](BAr<sub>4</sub>) (mer-[**3'**]<sup>+</sup>) is reversible at low temperatures, and warming from 160 K to 190 K shifts this equilibrium to the formation of initial manganese hydride fac-1. At the same time, the mer-to-fac isomerization of mer-[**3'**]<sup>+</sup> is almost irreversible due to much higher stability of fac-[(dppm)Mn(CO)<sub>3</sub>( $\kappa^1$ (Cl)-nBuCl)](BAr<sub>4</sub>) (fac-[**3'**]<sup>+</sup>). Excluding the process of fac-to-mer isomerization for starting hydrides 1 and hydrogen bonded adducts 1···BAr<sub>3</sub>, the mechanism could be simplified to following scheme with appropriate kinetic equations. Taking into account that the rate-determining step is mer-[**3'**]<sup>+</sup>  $\rightarrow fac$ -[**3'**]<sup>+</sup> isomerization, the overall process rate will be r = k[mer-[**3'**]<sup>+</sup>], where concentration of mer-[**3'**]<sup>+</sup> =  $K_1K_2$ [1][BAr<sub>3</sub>]. The reaction rate in this case will be  $r = k_{eff}$ [1<sup>H</sup>][BAr<sub>3</sub>], in which  $k_{eff} = K_1K_2k$ .

$$1 + BAr_{3} \stackrel{K_{1}}{\longleftarrow} 1 \cdots BAr_{3} \qquad K_{1} = \frac{[1 \cdots BAr_{3}]}{[1] [BAr_{3}]} \qquad K_{2} = \frac{[mer-[3']^{+}]}{[1 \cdots BAr_{3}]} \qquad r = k[mer-[3']^{+}]$$

$$1 \cdots BAr_{3} \stackrel{K_{2}}{\longleftarrow} mer-[3]^{+} \qquad r = k K_{1} K_{2} [1] [BAr_{3}] = k_{eff} [1] [BAr_{3}]$$

$$r = k K_{1} K_{2} [1] [BAr_{3}] = k_{eff} [1] [BAr_{3}]$$

Experimental effective rate constants for the reaction of fac-1 with 1.2 equiv. of BAr<sub>3</sub> (Table S2) were obtained in *n*BuCl at 220-250 K temperature range with integrated rate equation for second order reaction when initial reagents concentrations are not equal:

$$kt = \frac{1}{[1]_0[BAr_3]_0} \ln \frac{[BAr_3]_0[1]}{[1]_0[BAr_3]}$$

Then the corresponding activation parameters (Figure S6) were calculated with Eyring's equation for bimolecular reaction in solution (see below). Analysis of the experimental data for *fac*-**1** and *fac*-**1**-*d*<sub>3</sub> under the same conditions at 230 K (Figure S7) revealed the negligible kinetic isotopic effect ( $k_{\rm H}/k_{\rm D} = 0.9 \pm 0.1$ ).

$$k_{eff} = \frac{k_B T}{\hbar} e^{-\frac{\Delta H}{RT} + \frac{\Delta S}{T}}$$

**Table S2**. Experimentally determined effective rate constants for the interaction between *fac*-1 and 1.2 equiv. of BAr<sub>3</sub> in *n*BuCl at 220-250 K range.

	$k_{eff} = K_1 K_2 k, \ \mathrm{S} \cdot \mathrm{M}^{-1}$
<i>Т,</i> К	<i>fac</i> - <b>1</b> /B(C <sub>6</sub> F <sub>5</sub> ) <sub>3</sub>
220	2.4 · 10 <sup>-3</sup>
230	6.9 · 10 <sup>-3</sup>
240	$1.9 \cdot 10^{-2}$
250	4.4 · 10 <sup>-2</sup>



**Figure S6.**  $-\ln(k_{eff} \cdot \hbar/(T \cdot k_B))$  vs 1/T plot used for the determination of activation parameters for hydride transfer.



⊿H <sup>≠</sup> , kcal/mol	10.1 ± 0.5			
ΔS <sup>≠</sup> , cal/(mol·K)	-24 ± 2			
ΔG <sub>230</sub> <sup>≠</sup> , kcal/mol	15.6			

**Figure S7.**  $-\ln(k_{eff}\cdot\hbar/(T\cdot k_B))$  vs t plots for the determination of effective rate constants of for hydride (*fac*-**1** + B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>, blue) and deuteride (*fac*-**1**-*d*<sub>3</sub> + B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>, orange) transfer processes at 230K.

#### General procedure for variable temperature NMR experiments

A solution of *fac*-[(dppm)Mn(CO)<sub>3</sub>H] (*fac*-1) (20 mg, 0.04 mmol) in  $CD_2CI_2$  (0.2 mL) was filtered through Celite directly into the NMR tube and the filter pad was washed with additional amount of solvent (0.1 mL). The resulting solution was frozen in a liquid nitrogen and the solution of corresponding Lewis acid ([Ph<sub>3</sub>C](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>): 36.9 mg, 1 equiv.; B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>: 20 mg, 1 equiv.) in  $CD_2CI_2$  (0.2 mL) was carefully added on the wall of frozen NMR tube. Two frozen solutions were simultaneously melted and mixed in slush liquid nitrogen/EtOH bath at *ca*. 180 K and the resulting NMR sample was inserted into pre-cooled NMR probe at 183 K and then monitored with multi-nuclear NMR spectroscopy in 183-273 K temperature range.

# NMR spectroscopy characterization of Mn(I) cationic complexes *fac*-[3](BAr<sub>4</sub>), *mer*-[4](BAr<sub>4</sub>), *fac*-[5](BAr<sub>4</sub>) and *fac*-[2](HBAr<sub>3</sub>).

#### Spectroscopic characterization of complex fac-[3](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>).

The most pertinent NMR spectra for this compound are shown in Figures S13-S15. Based on <sup>1</sup>H and <sup>31</sup>P NMR data complex *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) exists in solution as a mixture of two isomers in *ca.* 4:1 ratio at room temperature attributed to *fac*,*anti*- (**A**, major) and *fac*,*syn*- (**B**, minor) forms according to the results of DFT calculations (see page S41). The equivalent amount of Ph<sub>3</sub>CH was also observed in <sup>1</sup>H and <sup>13</sup>C spectra, but their signals in <sup>1</sup>H and <sup>13</sup>C spectra are not listed.

#### *fac*-[3](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>):

<sup>1</sup>H NMR (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 298 K)  $\delta$  7.47-7.08 (m, 20H, CH<sub>Ar</sub>, **A**+**B**), 5.05 (dt overlapped with another dt, <sup>2</sup>J<sub>HH</sub> = 16.3 Hz, <sup>2</sup>J<sub>PH</sub> = 11.7 Hz, 0.2H, PCH<sub>2</sub>P, **B**), 5.03 (dt overlapped with another dt, <sup>2</sup>J<sub>HH</sub> = 16.4 Hz, <sup>2</sup>J<sub>PH</sub> = 11.5 Hz, 0.8H, PCH<sub>2</sub>P, **A**), 4.78 (dt, <sup>2</sup>J<sub>HH</sub> = 16.4 Hz, <sup>2</sup>J<sub>PH</sub> = 11.5 Hz, 0.8H, PCH<sub>2</sub>P, **A**), 4.66 (dt, <sup>2</sup>J<sub>HH</sub> = 16.3 Hz, <sup>2</sup>J<sub>PH</sub> = 11.7 Hz, 0.2H, PCH<sub>2</sub>P, **B**).

<sup>1</sup>H{<sup>31</sup>P} NMR (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 298 K)  $\delta$  7.08-7.47 (m, CH<sub>Ar</sub>, **A**+**B**), 5.05 (d overlapped with another d, <sup>2</sup>J<sub>HH</sub> = 16.2 Hz, PCH<sub>2</sub>P, **B**), 5.03 (d overlapped with another d, <sup>2</sup>J<sub>HH</sub> = 16.3 Hz PCH<sub>2</sub>P, **A**), 4.78 (d, <sup>2</sup>J<sub>HH</sub> = 16.3 Hz, PCH<sub>2</sub>P, **A**), 4.66 (d, <sup>2</sup>J<sub>HH</sub> = 16.2 Hz, PCH<sub>2</sub>P, **B**).

<sup>31</sup>P{<sup>1</sup>H} NMR (243.0 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 298 K): δ 13.8 (s, **B**), 10.0 (s, **A**).

<sup>13</sup>C{<sup>1</sup>H} NMR (150.9 MHz, 243 K, CD<sub>2</sub>Cl<sub>2</sub>): 219.7 (t, <sup>2</sup>J<sub>PC</sub> = 18.8 Hz, Mn–CO, **A**), 215.2 (br s, Mn–CO, **B**), 214.8 (br s, Mn–CO, **A**), 147.9 (br d, <sup>1</sup>J<sub>BF</sub> = 239.5 Hz, *C*<sub>ortho</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 138.1 (br dt, <sup>1</sup>J<sub>BF</sub> = 239.5 Hz, <sup>2</sup>J<sub>BF</sub> = 11.5 Hz, *C*<sub>para</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 136.2 (br d, <sup>1</sup>J<sub>BF</sub> = 245.7 Hz, *C*<sub>meta</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 133.0, 132.9 (s, *C*H<sub>PPh2</sub>, **B**), 132.7, 132.5 (s, *C*H<sub>PPh2</sub>, **A**), 131.5 (vt, *J*<sub>PC</sub> = 5.9 Hz, *C*H<sub>PPh2</sub>, **B**), 131.3 (vt, *J*<sub>PC</sub> = 5.6 Hz, *C*H<sub>PPh2</sub>, **A**), 131.0 (vt, *J*<sub>PC</sub> = 5.3 Hz, *C*H<sub>PPh2</sub>, **A**), 130.9 (vt, *J*<sub>PC</sub> = 5.8 Hz, *C*H<sub>PPh2</sub>, **B**), 130.6 (vt, *J*<sub>PC</sub> = 4.9 Hz, *C*H<sub>PPh2</sub>, **B**), 130.4 (vt, *J*<sub>PC</sub> = 5.7 Hz, *C*H<sub>PPh2</sub>, **B**), 130.3 (vt, *J*<sub>PC</sub> = 5.2 Hz, *C*H<sub>PPh2</sub>, **A**), 130.1 (vt, *J*<sub>PC</sub> = 5.3 Hz, *C*H<sub>PPh2</sub>, **A**), 129.6 (vt, *J*<sub>PC</sub> = 25.2 Hz, *C*<sub>ipso-PPh2</sub>, **A**), 127.2 (vt, *J*<sub>PC</sub> = 19.4 Hz, *C*<sub>ipso-PPh2</sub>, **A**), 123.6 (br m, *C*<sub>ipso</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 63.0 (br m, Mn–*C*D<sub>2</sub>Cl<sub>2</sub>, **A**+**B**), 39.7 (t, <sup>1</sup>*J*<sub>PC</sub> = 24.3 Hz, PCH<sub>2</sub>P, **A**+**B**). IR (CH<sub>2</sub>Cl<sub>2</sub>, 290 K): *v*<sub>CO</sub> 2037, 1968, 1946 cm<sup>-1</sup>; IR (nBuCl, 290 K): *v*<sub>CO</sub> 2040, 1970, 1943 cm<sup>-1</sup>; IR (toluene, 290 K):

*v*<sub>co</sub> 2037, 1967, 1942 cm<sup>-1</sup>.

## Characterization of complex *mer*-[4](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) observed by low-temperature NMR monitoring of the reaction between *fac*-1 and [Ph<sub>3</sub>C](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>).

The approximate ratio between *mer*-[**4**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), *fac*, *anti*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) and Ph<sub>3</sub>CH in the reaction mixture at 183 K was determined by the integration for well-separated signals at  $\delta_{\rm H}$  5.78, 4.74 and 5.58 ppm, respectively (Figure S18). A quantitative <sup>1</sup>H NMR yield of two latter products was observed at 223 K using the signal of residual solvent protons as an internal standard (Figure S18, inset). The existence of CH=CHCH<sub>2</sub>CH=CH structural motif in the molecule of *mer*-[**4**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) was unambiguously confirmed by <sup>1</sup>H TOCSY NMR spectrum (Figure S20) and their exact attribution was inferred using <sup>1</sup>H–<sup>1</sup>H COSY (Figure S19) and <sup>1</sup>H–<sup>13</sup>C HSQC (Figure S23) spectra. Based on the general trends known in the literature, it was proposed that more upfield CH resonances observed at  $\delta_{\rm H}/\delta_c$  5.78/107.4 and 4.41/88.5 ppm belong to the CH=CH moiety coordinated to the manganese atom (Figures S22-23), whereas non-coordinated CH=CH 1,4-cyclohexadiene fragment was attributed to the signals at  $\delta_{\rm H}/\delta_c$  6.44/126.5 and 5.51/127.7 ppm. Upon warming a fast transformation of *mer*-[**4**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) was observed by <sup>1</sup>H and <sup>31</sup>P NMR spectroscopy (Figure S24-S25) together with a detection of non-coordinated cyclic diene **5** isomerizing into triphenylmethane.

#### *mer*-[4](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>):

<sup>1</sup>H NMR (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K):  $\delta$  7.72 (br dd,  $J_{PH} = 12.2$ ,  ${}^{3}J_{HH} = 7.3$  Hz, 2H,  $CH_{PPh_{2}}$ ), 7.67-7.28 (m, overlapped with signals of *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 24H,  $CH_{PPh_{2}}+CH_{Ph}$ ), 6.98 (d,  ${}^{3}J_{HH} = 7.3$  Hz, 2H,  $CH_{Ph}$ ), 6.87 (br dd,  $J_{PH} = 11.4$ ,  ${}^{3}J_{HH} = 7.5$  Hz, 2H,  $CH_{PPh_{2}}$ ), 6.44 (d,  ${}^{3}J_{HH} = 10.5$  Hz, 1H,  $CH_{2}CH=CH$ ), 5.78 (br d,  ${}^{3}J_{HH} = 8.5$  Hz, 1H,

CH<sub>2</sub>CH=CH...Mn), 5.51 (br d,  ${}^{3}J_{HH}$  = 10.8 Hz, 1H, CH<sub>2</sub>CH=CH), 5.05 (dt,  ${}^{2}J_{HH}$  = 18.0 Hz,  ${}^{2}J_{PH}$  = 9.3 Hz, 1H, PCH<sub>2</sub>P), 4.53-4.40 (m overlapped with singlet, 2H, PCH<sub>2</sub>P+CH<sub>2</sub>CH=CH...Mn), 2.23 (d,  ${}^{2}J_{HH}$  = 25.7 Hz, 1H, CH<sub>2</sub>), 2.06 (d,  ${}^{3}J_{HH}$  = 25.7 Hz, 1H, CH<sub>2</sub>(CH=CH)<sub>2</sub>).

 $^{31}P{^{1}H} NMR (243.0 \text{ MHz}, 183 \text{ K}, CD_2Cl_2): \delta 10.6 (d, {}^{2}J_{PP} = 44.4 \text{ Hz}), 7.10 (d, {}^{2}J_{PP} = 44.3 \text{ Hz}).$ 

<sup>13</sup>C{<sup>1</sup>H} NMR (150.9 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K):  $\delta$  218.9 (br s, Mn–CO), 218.1 (br s, Mn–CO), 213.6 (br s, Mn–CO), 147.9 (br d, <sup>1</sup>J<sub>BF</sub> = 238.5 Hz, *C*<sub>ortho</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 140.2, 139.3 (s, *C*<sub>ipso-Ph<sub>2</sub>C=C</sub>), 138.1 (br dt, <sup>1</sup>J<sub>BF</sub> = 239.0 Hz, <sup>2</sup>J<sub>BF</sub> = 11.5 Hz, *C*<sub>para</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 136.2 (br d, <sup>1</sup>J<sub>BF</sub> = 245.7 Hz, *C*<sub>meta</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 132.5-129.0 (numerous CH<sub>Ph</sub> signals overlapping with those of *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 127.7 (s, CH<sub>2</sub>CH=CH), 126.5 (s, CH<sub>2</sub>CH=CH), 126.3 (vt, *J*<sub>PC</sub> = 19.0 Hz, *C*<sub>ipso-PPh<sub>2</sub>), 125.7, 124.4 (s, *C*<sub>Ph<sub>2</sub>C=C</sub>), 123.6 (br m, *C*<sub>ipso</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 107.6 (br s, CH<sub>2</sub>CH=CH...Mn), 88.5 (br s, CH<sub>2</sub>CH=CH...Mn), 39.8 (br d, <sup>1</sup>J<sub>PC</sub> = 20 Hz, PCH<sub>2</sub>P), 27.5 (br s, CH<sub>2</sub>CH=CH)<sub>2</sub>).</sub>

IR (CH<sub>2</sub>Cl<sub>2</sub>, 180 K): v<sub>CO</sub> 2063 cm<sup>-1</sup>; IR (*n*BuCl, 160 K): v<sub>CO</sub> 2064 cm<sup>-1</sup>.

**Compound 5**: <sup>1</sup>H NMR (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 213 K):  $\delta$  7.66-7.45 (m overlapped with signals of *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 20H, CH<sub>Ar</sub>), 6.46 (d, <sup>3</sup>J<sub>HH</sub> = 10.2 Hz, 2H, CH<sub>2</sub>(CH=CH)<sub>2</sub>), 5.88 (br dt, <sup>3</sup>J<sub>HH</sub> = 9.2 Hz, 1H, <sup>3</sup>J<sub>HH</sub> = 4.2 Hz, 2H, CH<sub>2</sub>(CH=CH)<sub>2</sub>), 3.07 (s, 2H, CH<sub>2</sub>(CH=CH)<sub>2</sub>).

## Low temperature NMR monitoring of the reaction between complex *fac*-1 and $B(C_6F_5)_3$ and spectroscopic characterization of complexes *fac*-[3](BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>) and *fac*-[2](BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>).

The evidence of Lewis acid adducts formation by <sup>1</sup>H and <sup>31</sup>P NMR spectroscopy is illustrated in Figures S26 and S29, respectively. Several very broad signals presented in low-temperature <sup>1</sup>H NMR spectra in 5-to-3 ppm region (Figure 28) may be attributed to PCH<sub>2</sub>P resonances of these adducts and/or cationic *mer*-[**3**](BHAr<sub>3</sub>) intermediate involved into fluxional process. While two isomers of cationic *fac*-[**3**](BHAr<sub>3</sub>) products (*ca.* 3:1 ratio at 183 K) attributed to *fac,anti*- (major, **A**) and *fac,syn*- (minor, **B**) forms according to the results of DFT calculations (see page S41) were observed in 183-233 K temperature range, these species at 243 K starts to transform into *fac*-[**2**](BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>) being the principal component at 273 K. The spectroscopic characteristics of *fac*-[**3**](BAr<sub>4</sub>) and *fac*-[**2**](BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>) are quite similar and a sole notable difference between them deals with strong shielding of PCH<sub>2</sub>P protons by 0.5-1.0 ppm in the latter (Figures S27-S28).

#### fac-[3](BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>):

<sup>1</sup>H NMR (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K)  $\delta$  5.23 (dt, <sup>2</sup>J<sub>HH</sub> = 16.7 Hz, <sup>2</sup>J<sub>PH</sub> = 10.2 Hz, 0.7H, PCH<sub>2</sub>P, **A**), 4.93-4.80 (m, 0.6H, PCH<sub>2</sub>P, **B**), 4.76 (dt, <sup>2</sup>J<sub>HH</sub> = 16.7 Hz, <sup>2</sup>J<sub>PH</sub> = 11.9 Hz, 0.7H, PCH<sub>2</sub>P, **A**).

<sup>1</sup>H{<sup>31</sup>P) NMR (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K) δ 5.26 (d, <sup>2</sup>*J*<sub>HH</sub> = 16.7 Hz, PC*H*<sub>2</sub>P, **A**), 4.89 (d, <sup>2</sup>*J*<sub>HH</sub> = 16.6 Hz, PC*H*<sub>2</sub>P, **B**), 4.81 (d, <sup>2</sup>*J*<sub>HH</sub> = 16.6 Hz, PC*H*<sub>2</sub>P, **B**), 4.76 (d, <sup>2</sup>*J*<sub>HH</sub> = 16.7 Hz, PC*H*<sub>2</sub>P, **A**).

<sup>31</sup>P{<sup>1</sup>H} NMR (243.0 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K): δ 11.1 (s, **B**), 10.1 (s, **A**).

#### fac-[2](BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>):

<sup>1</sup>H NMR (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 273 K)  $\delta$  7.46-7.36 (m, 12H, CH<sub>Ph</sub>), 7.21-7.14 (m, 4H, CH<sub>Ph</sub>), 7.12-7.06 (m, 4H, CH<sub>Ph</sub>), 4.58 (dt, <sup>2</sup>J<sub>HH</sub> = 15.6 Hz, <sup>2</sup>J<sub>PH</sub> = 10.6 Hz, 1H, PCH<sub>2</sub>P), 3.85 (dt, <sup>2</sup>J<sub>HH</sub> = 15.6 Hz, <sup>2</sup>J<sub>PH</sub> = 10.9 Hz, 1H, PCH<sub>2</sub>P), 3.68 (very br, 1H, BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>).

 $^{31}P{^{1}H} NMR (243.0 MHz, CD_2Cl_2, 273 K): \delta 10.1 (s).$ 

<sup>11</sup>B NMR (192.5 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 273 K): δ 25.4 (br s).

<sup>13</sup>C{<sup>1</sup>H} NMR (150.9 MHz, 253 K, CD<sub>2</sub>Cl<sub>2</sub>): 219.5 (t, <sup>2</sup>J<sub>PC</sub> = 18.4 Hz, Mn–CO), 217.2 (br s, Mn–CO), 148.3 (br d, <sup>1</sup>J<sub>BF</sub> = 243.6 Hz, *C*<sub>ortho</sub> BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>), 139.3 (br d, <sup>1</sup>J<sub>BF</sub> = 248.0 Hz, *C*<sub>para</sub> BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>), 136.2 (br d, <sup>1</sup>J<sub>BF</sub> = 245.7 Hz, *C*<sub>meta</sub> B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>), 131.9 (vt, *J*<sub>PC</sub> = 5.2 Hz, *C*H<sub>PPh<sub>2</sub></sub>), 131.7, 131.5 (s, *C*H<sub>PPh<sub>2</sub></sub>), 131.4 (vt, *J*<sub>PC</sub> = 5.4 Hz, *C*H<sub>PPh<sub>2</sub></sub>), 130.7 (vt, *J*<sub>PC</sub> = 21.8 Hz, *C*<sub>ipso-PPh<sub>2</sub></sub>), 130.4 (vt, *J*<sub>PC</sub> = 20.0 Hz, *C*<sub>ipso-PPh<sub>2</sub></sub>), 129.4 (vt overlapped with another vt, *J*<sub>PC</sub> = 4.9 Hz, *C*H<sub>PPh<sub>2</sub></sub>), 129.3 (vt overlapped with another vt, *J*<sub>PC</sub> = 5.1 Hz, *C*H<sub>PPh<sub>2</sub></sub>), 119.0 (br m, *C*<sub>ipso</sub> BH(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub>), 38.4 (t, <sup>1</sup>J<sub>PC</sub> = 20.6 Hz, PCH<sub>2</sub>P).

IR (CH<sub>2</sub>Cl<sub>2</sub>, 290 K): *v*<sub>CO</sub> 2037, 1968, 1946 cm<sup>-1</sup>; IR (*n*BuCl, 290 K): *v*<sub>CO</sub> 2040, 1970, 1943 cm<sup>-1</sup>; IR (toluene, 290 K): *v*<sub>CO</sub> 2037, 1967, 1942 cm<sup>-1</sup>.



**Figure S8**. <sup>31</sup>P{<sup>1</sup>H} NMR spectrum of complex *fac*-**1** (162.0 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 298 K).



Figure S9. <sup>1</sup>H NMR spectrum of complex *fac*-1 (400.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 298 K).



**Figure S10**. <sup>31</sup>P{<sup>1</sup>H} NMR spectrum of complex *fac*-**1** (162.0 MHz, C<sub>6</sub>D<sub>6</sub>, 298 K).



**Figure S11**. <sup>1</sup>H NMR spectrum of complex *fac*-**1**-*d*<sub>3</sub> (400.1 MHz, C<sub>6</sub>D<sub>6</sub>, 298 K).



**Figure S12**. <sup>13</sup>C{<sup>1</sup>H, <sup>31</sup>P} NMR spectrum of complex *fac*-**1**-*d*<sub>3</sub> (100.6 MHz, C<sub>6</sub>D<sub>6</sub>, 298 K).



Figure S13.  ${}^{31}P{}^{1}H{}$  NMR spectrum of complex *fac*-[3](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) (243.0 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 298 K).



**Figure S14**. <sup>1</sup>H NMR spectrum of complex fac-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 298 K), the signals of triphenylmethane are marked with asterisk. Inset spectrum corresponds to the section of PCH<sub>2</sub>P protons in <sup>1</sup>H{<sup>31</sup>P} decoupled experiment.



**Figure S15**. <sup>13</sup>C{<sup>1</sup>H} NMR spectrum of complex *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) (150.9 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 243 K), the signals of triphenylmethane are marked with asterisk. Inset spectrum corresponds to the section of <sup>13</sup>C{<sup>1</sup>H} NMR spectrum for the same compound at 183 K.



**Figure S16**. <sup>31</sup>P{<sup>1</sup>H} NMR spectrum of the reaction between complex *fac*-**1** and  $[Ph_3C](B(C_6F_5)_4)$  (243 MHz,  $CD_2Cl_2$ , 183 K).



**Figure S17**. <sup>1</sup>H NMR spectrum of the reaction between complex *fac*-**1** and  $[Ph_3C](B(C_6F_5)_4)$  (600.1 MHz,  $CD_2Cl_2$ , 183 K), only the signals of *mer*-[**4**]( $B(C_6F_5)_4$ ) are identified and integrated. The signals of non-coordinated Ph<sub>3</sub>CH are marked with asterisk, and those of *fac*-[**3**]( $B(C_6F_5)_4$ ) with red dots.



**Figure S18**. Section of <sup>1</sup>H NMR spectrum (600.1 MHz,  $CD_2Cl_2$ ) of the mixture of *mer*-[**4**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) and *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) at 183 K indicating the amount of *fac*-[**2**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) and free Ph<sub>3</sub>CH (the signal of residual CD<sub>2</sub>Cl<sub>2</sub> protons was used for internal standard). Inset corresponds to the sample after full conversion into *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) at 223 K.



**Figure S19**.  ${}^{1}H-{}^{1}H$  COSY NMR spectrum with  ${}^{31}P$  decoupling of the reaction between complex *fac*-**1** and  $[Ph_{3}C](B(C_{6}F_{5})_{4})$  (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K) and correlation scheme in 1,4-cyclohexadiene moiety (chemical shifts are shown in red and the observed correlation between the corresponding protons with green arrows).



Figure S20. 1D <sup>1</sup>H TOCSY spectrum of mer-[4](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K, irradiated at 6.44 ppm) and proposed scheme of spin transfer (green arrows)



**Figure S21**. Section of  ${}^{1}H{}^{31}P{}$  NMR spectrum of the reaction between complex *fac*-**1** and  $[Ph_{3}C](B(C_{6}F_{5})_{4})$  (600.1 MHz,  $CD_{2}Cl_{2}$ , 183 K), only the signals of *mer*-**[4**]( $B(C_{6}F_{5})_{4}$ ) are identified. The signals of non-coordinated  $Ph_{3}CH$  are marked with asterisk, and those of *fac*-**[3**]( $B(C_{6}F_{5})_{4}$ ) with red dots.



**Figure S22**. <sup>13</sup>C{<sup>1</sup>H} NMR spectrum of mixture of *mer*-[**4**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) and *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) at 183 K (150.9 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K), the signals of triphenylmethane are marked with asterisk, and those of *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) with red dots.



**Figure S23**. <sup>13</sup>C–<sup>1</sup>H HSQC NMR spectrum for the mixture of *mer*-[**4**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) and *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) at 183 K (150.9 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 183 K, cross-peaks for CH an CH<sub>2</sub> fragments are shown in red and blue, respectively) and correlation scheme in 1,4-cyclohexadiene moiety.



**Figure S24**. <sup>31</sup>P{<sup>1</sup>H} NMR monitoring of the reaction between *fac*-**1** and [Ph<sub>3</sub>C](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) in 183-213 K temperature range (243 MHz, CD<sub>2</sub>Cl<sub>2</sub>).



**Figure S25**. Section of <sup>1</sup>H NMR spectra (600.1 MHz,  $CD_2Cl_2$ ) in 183-213 K temperature range illustrating the transformation of *mer*-[**4**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) into *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) and cyclic diene **5**, the signals of triphenylmethane are marked with asterisk, and those of *fac*-[**3**](B(C<sub>6</sub>F<sub>5</sub>)<sub>4</sub>) are marked with red dots.



**Figure S26**. <sup>1</sup>H NMR monitoring of the reaction between *fac*-**1** and B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> in 183-243 K temperature range in hydride signals region (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>).



**Figure S27**. <sup>1</sup>H NMR monitoring of the reaction between *fac*-**1** and B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> in 183-243 K temperature range in PCH<sub>2</sub>P signals region (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>).



**Figure S28**. Superposition of <sup>1</sup>H NMR spectra of the reaction between *fac*-**1** and  $B(C_6F_5)_3$  at 243 and 273 K in PCH<sub>2</sub>P signals region illustrating the transformation of *fac*-[**3**](BHPh<sub>3</sub>) into *fac*-[**2**](BHPh<sub>3</sub>) (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>).



**Figure S29**. Section of <sup>31</sup>P{<sup>1</sup>H} NMR spectra of the reaction between *fac*-1 and B( $C_6F_5$ )<sub>3</sub> in 183-243 K temperature range illustrating the formation of non-covalent adducts *fac*-1···BAr<sub>3</sub> and *mer*-1···BAr<sub>3</sub>.



**Figure S30**. Section of <sup>31</sup>P{<sup>1</sup>H} NMR spectra of the reaction between *fac*-**1** and B(C<sub>6</sub>F<sub>5</sub>)<sub>3</sub> in 243-273 K temperature range illustrating the formation of meridional cationic species *mer*-[**3**](BHAr<sub>3</sub>) and/or *mer*-[**2**](BHAr<sub>3</sub>).



**Figure S31**. <sup>1</sup>H NMR spectrum of complex *fac*-[**2**](BHAr<sub>3</sub>) (600.1 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 273 K), inset correspond to the section of <sup>1</sup>H{<sup>11</sup>B} NMR spectrum evidencing broad signal of the B–H moiety coordinated to the metal atom.



Figure S32. <sup>31</sup>P{<sup>1</sup>H} NMR spectrum of complex *fac*-[2](BHAr<sub>3</sub>) (243.0 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 273 K).



**Figure S33**. <sup>11</sup>B NMR spectrum of complex *fac*-[**2**](BHAr<sub>3</sub>) (192.5 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 273 K).



**Figure S34**. <sup>13</sup>C{<sup>1</sup>H} NMR spectrum of complex *fac*-[**2**](BHAr<sub>3</sub>) (150.9 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 253 K).



**Figure S35**. <sup>13</sup>C{<sup>1</sup>H, <sup>31</sup>P} NMR spectrum of complex *fac*-[**2**](BHAr<sub>3</sub>) (150.9 MHz, CD<sub>2</sub>Cl<sub>2</sub>, 253 K).

#### **DFT calculations.**

Structures of reactants and complexes were optimized at the  $\omega$ B97-XD level<sup>4</sup> applying def2-TZVP basis set<sup>5</sup> by Gaussian 09.<sup>6</sup> Optimizations were done in CH<sub>2</sub>Cl<sub>2</sub> introduced by SMD solvent model.<sup>7</sup> Infrared frequencies were scaled by a factor 0.950.<sup>8</sup> Manganese hydride complex (dppm)Mn(CO)<sub>3</sub>H (**1**) has two minima on PES having facial and meridional arrangement of three carbonyl groups (Figure S36). At that *fac*-**1** has *syn*-arrangement of CH<sub>2</sub>-bridge of dppm-ligand respective to the hydride ligand, the minima corresponding to the *anti-fac*-**1** isomer was not revealed.



**Figure S36.** Optimized structures of two isomers for Mn(I) hydride complex (dppm)Mn(CO)<sub>3</sub>H (**1**). Hydrogen atoms except hydride are removed for clarity and phenyl rings are shown as a wireframe. Energies in kcal/mol are relative to *fac-***1**.



**Figure S37.** Optimized structures of the possible isomers of **1**...BAr<sub>3</sub> complexes. Hydrogens (except hydride) and fluorine removed, phenyl carbons shown as a wireframe. Energies in kcal/mol are relative to *fac*-**1** and free BAr<sub>3</sub>.



**Figure S38.** Simulated IR spectra for isomers of hydride **1**, their adducts with BAr<sub>3</sub> and cations [**3**](BHAr<sub>3</sub>). Frequencies scales by 0.950, FWHH set to 9.5.

The frequency analysis for **1**, **3** and [mer-1]···BAr<sub>3</sub> adducts shows that vibrations which are higher than v(CO) of facial cation **3** is unambiguously belongs to the meridional isomers.



**Figure S39.** Optimized structures of non-stabilized  $[(dppm)Mn(CO)_3]^+$  cations  $[2]^+$  with key geometric parameters. Hydrogens are removed, phenyl carbons shown as a wireframe. Energies in kcal/mol are relative to most stable *fac,anti*-2<sup>+</sup>.

The *fac,anti*-[**3**]<sup>+</sup> and *fac,syn*-[**2**]<sup>+</sup> cations retain C<sub>s</sub> symmetry of the parent facial hydride (Mn-P distances differs by <0.01 Å), while difference of Mn–P bonds is 0.072 Å for *mer*-[**2**]<sup>+</sup> isomer. Due to non-planarity of MnPCP cycle two "apical" CO groups becomes non-equivalent.



**Figure S40**. Simulated IR spectra for isomers of complex [(dppm)Mn(CO)<sub>3</sub>]<sup>+</sup> [**2**]<sup>+</sup>. Frequencies scales by 0.950, FWHH set to 9.5.



**Figure S41**. Optimized structures of fac-[(dppm)Mn(CO)<sub>3</sub>( $\kappa^1Cl$ -CH<sub>2</sub>Cl<sub>2</sub>)](BAr<sub>4</sub>) ([**3**]<sup>+</sup>) with key geometric parameters. Hydrogens except those of CH<sub>2</sub>Cl<sub>2</sub> are removed, phenyl carbons are shown as a wireframe. Energies in kcal/mol are relative to the *fac*, *anti*-[**3**]<sup>+</sup> isomer.



**Figure S42.** Comparison of simulated IR spectra for cationic complexes with coordinated  $CH_2Cl_2$  [**3**]<sup>+</sup> vs. their ion pairs [**3**](BHAr<sub>3</sub>). Frequencies scales by 0.950, FWHH set to 9.5.

Specific  $CH_2Cl_2$  solvatation significantly changes the energetic pattern of cationic complexes. The most favored isomer at this conditions being the *fac,syn*-one (in the Gibbs energy scale). Notably the relative preference of *syn*- and *anti*-isomers are clearly depending on the coordination of Mn atom – if covalent bond is formed – in hydride or bromide – only *syn*-form could be found, when the coordination site is fully opened – in the free cation – the *anti*-isomer is strongly preferred.  $CH_2Cl_2$  complexes occupy the intermediate position in this row and in this case *fac,syn*- and *fac,anti*-isomers become almost isoergic.

Another notable result is a unfavorable character of  $fac, anti-[(dppm)Mn(CO)_3(\kappa^1Cl-CH_2Cl_2)](BAr_4)$ ( $fac, anti-[3]^+$ ) complex formation in the Gibbs energy scale, even at 190 K. It should be noted that  $\Delta G=+2 \div$ +3 kcal/mol still allow *ca*. 10% of CH<sub>2</sub>Cl<sub>2</sub> complex when using CH<sub>2</sub>Cl<sub>2</sub> as a solvent due to the concentration shift of the equilibrium. Therefore we could suggest that formed as a result of hydride transfer *mer*-[2]<sup>+</sup> cation at low temperatures readily coordinates CH<sub>2</sub>Cl<sub>2</sub> molecule to form *mer*-[(dppm)Mn(CO)<sub>3</sub>( $\kappa^1Cl$ -CH<sub>2</sub>Cl<sub>2</sub>)](BAr<sub>4</sub>) (*mer*-[3]<sup>+</sup>) solvate. The isomerization to the preferred *fac, syn*-[3]<sup>+</sup> complex could proceeds both directly via dissociation to the free [2]<sup>+</sup> cation. At the higher temperatures CH<sub>2</sub>Cl<sub>2</sub> complexes are tend to dissociate with the further isomerization to the thermodynamic product *fac, anti*-[3]<sup>+</sup>.

	fac,anti-[ <b>3</b> ]+	fac,syn-[ <b>3</b> ]+	mer-[ <b>3</b> ]+
ΔE	-7.5	-10.6	-11.6
ΔН	-5.8	-9.0	-9.9
$\Delta G^{298}$	6.8	2.9	3.0
$\Delta G^{190}$	2.2	-1.5	-1.7

**Table S2.** Energies of  $CH_2Cl_2$  complexes formation for  $[3]^+([2]^+ + CH_2Cl_2 \rightarrow [3]^+)$ .

Commission	(dppm)Mn(CO)₃H		(dppm)Mn(CO)₃H…BAr₃		[(dppm)Mn(CO)₃]⁺		$[(dppm)Mn(CO)_3(CH_2Cl_2)]^+$			[(dppm)Mn(CO)₃(CH₂Cl₂)]⁺…[HBAr₃] <sup>-</sup> [ <b>3</b> ](BHAr₃)			
Complex	1"		1 <sup>H</sup> ····BAr₃		[ <b>2</b> ]⁺		[ <b>3</b> ]⁺						
	mer-	fac,syn-	mer-	fac,syn-	mer-	fac,syn-	fac,anti-	mer-	fac,syn-	fac,anti-	mer-	fac,syn-	fac,anti-
<i>v</i> <sub>CO1</sub> , cm <sup>-1</sup>	1914	1914	1968	1959	1978	1970	1966	1977	1974	1969	1974	1970	1966
A <sub>CO1</sub> , km/mol	2215	1901	1366	1317	1560	1331	1331	1330	1276	1297	1343	1313	1322
<i>v</i> <sub>CO2</sub> , cm <sup>-1</sup>	1918	1918	1975	1974	1992	1980	1982	1997	1982	1984	1999	1977	1980
A <sub>CO2</sub> , km/mol	2338	1785	2417	1252	2889	1629	1567	2418	1308	1356	2278	1272	1338
<i>v</i> <sub>CO3</sub> , cm <sup>-1</sup>	1999	1996	2056	2037	2073	2047	2045	2069	2043	2044	2072	2040	2041
A <sub>CO3</sub> , km/mol	664	1588	467	1566	368	1495	1576	249	1498	1509	257	1527	1487

**Table S3.** Calculated  $v_{CO}$  frequencies (scaled by factor 0.95) and intensities for Mn(I) complexes **1**, [**2**]<sup>+</sup> and [**3**]<sup>+</sup> and [**3**](BHAr<sub>3</sub>)



**Figure S43.** Computed (DFT/ $\omega$ B97XD/def2-TZVP/SMD(CH<sub>2</sub>Cl<sub>2</sub>)) Gibbs energy profiles for the *fac*-1 and BAr<sub>3</sub> reaction at 298 K (top) and 190 K (bottom).

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