

Acquisition of quantitative ^{13}C NMR spectra in a short time with EXACT (EXtended ACquisition Time) acquisition Method (SI)

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1 Experimental

Camphor, Strychnine, and DEET (Diethyl meta toluamide) samples were prepared as follows: 30 mg of Camphor and 30 mg of Strychnine were dissolved separately in 800 μl CDCl_3 each and used for the analysis. DEET (100 μl) sample was mixed with 900 μl of deuterated chloroform to make a 10% sample.

All sequences were programmed using C and MAGICAL languages compatible with the VNMRJ4 software. Spectra were acquired without the need for any window function processing after acquisition and FT. Windowed acquisition with the default constant sampling is used.

The EXACT and the SRT NMR experiments were carried out on Varian VNMR5 500 MHz spectrometers equipped with either a OneNMR probe or an Auto_X_DB_9070 probe. The duration of the 90° pulse on the ^{13}C observed channel was calibrated to 9 μs and the 180° refocusing pulse on the proton decoupler channel was calibrated to 25 μs using the auto_X_DB_9070 probe equipped spectrometer. Using the OneNMR probe equipped spectrometer, the 180° refocusing pulse was calibrated to 12 μs and the 90 pulse was calibrated to 9 μs .

All experiments were run at 25°C . All ^{13}C spectra were collected with 248.7 ppm spectral width (31250 Hz) and 32768 complex points and used Waltz-16 decoupling for the CPD (indicated by black rectangles in the pulse sequence in Figure 1). Acquisition times (including all chunks and gaps) were 1s for all the experiments. Relaxation delays and chunk lengths were varied as described in the main text,

IST was used to reconstruct the unmeasured datapoints in the EXACT spectra. IT is an algorithm of the Compressed Sensing family, based on the assumption of spectral sparsity: the

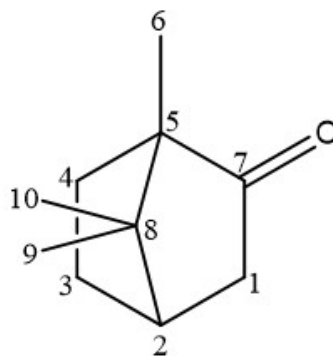
fact that an NMR spectrum has much more baseline points than “meaningful” ones, i.e. those contributing to peaks. It is an iterative algorithm in which we take the Fourier transform of the FID in each iteration; perform a thresholding operation, i.e. “cut” the peak tops which are above a pre-defined threshold; take the inverse Fourier transform of the thresholding result; and, finally, redefine non-measured points keeping the registered ones. When the output stops improving considerably from iteration to iteration, the algorithm is considered to be converged and yielding a reconstructed spectrum that is close enough to the one we would get if we had no gaps. More details on how to set the parameters of the algorithm can be found in reference, see also Fig.3 there for an illustration. Scripts for performing the reconstructions are available in the Supporting Information of references 9-11 of the main paper and were performed using Python wrappers of the MDDNMR software package (can be downloaded at <http://mddnmr.spektrino.com/download>).

Data processing and analysis were performed in Mestrenova 11.0. Processing of the SRT FIDs was performed by adding the two processed FIDs prior to the Fourier transform. The spectra were zero-filled to 256K datapoints prior to integration and baseline corrected with a 3rd order polynomial fit.

2 Camphor experimental results

2.1 Table 1: Camphor ^{13}C numbers and assignment in ppm.

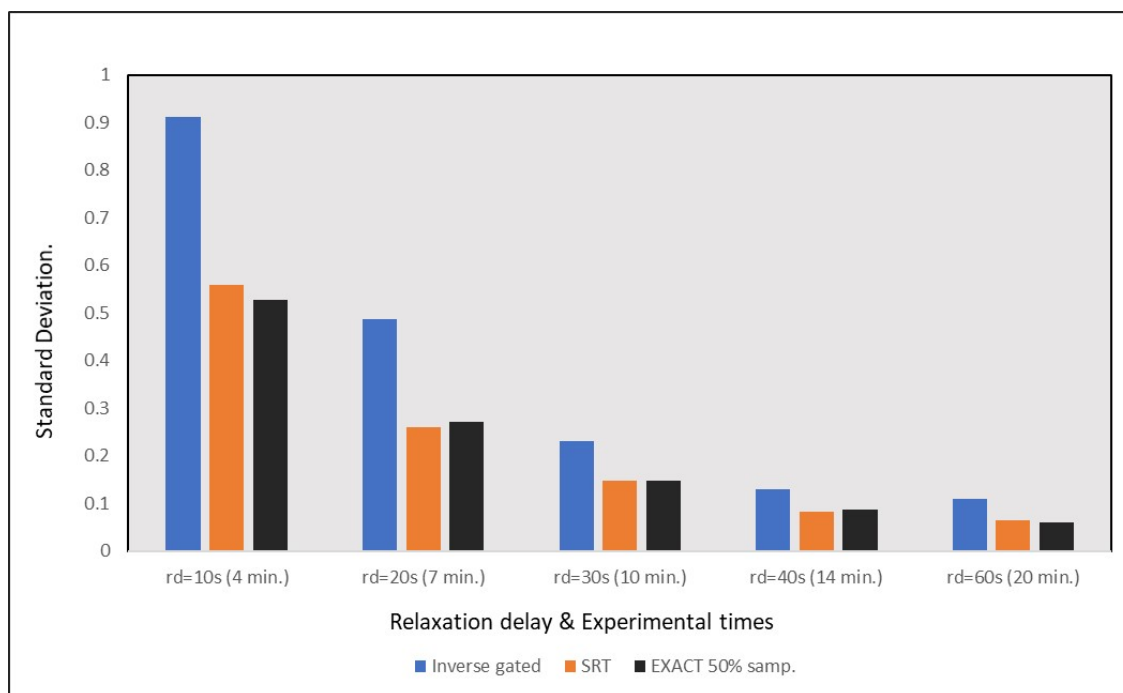
Carbon	^{13}C Assignment (ppm)
C1	43.04
C2	43.29
C3	27.03
C4	29.9
C5	57.68
C6	9.22
C7	219.65
C8	46.77
C9	19.12
C10	19.76



2.2 Table 2: Integrals and standard deviations of Camphor sample at different relaxation delays using inverse gated (IG), SRT, and EXACT acquisition methods.

Inverse-gated Acquisition (Camphor)					
	rd=10s	rd=20	rd=30s	rd=40s	rd=60s
C1	3.13	2.00	1.41	1.19	1.13
C2	2.83	1.95	1.30	1.06	1.16
C3	2.90	1.80	1.50	1.16	0.97
C4	2.50	1.71	1.44	1.28	1.01
C5	0.81	0.74	0.88	0.88	0.77
C6	2.57	1.92	1.26	1.14	1.07
C7	1.00	1.00	1.00	1.00	1.00
C8	0.89	0.85	0.88	0.88	1.00
C9	2.60	1.79	1.24	1.06	1.10
C10	2.71	1.76	1.41	1.11	0.95
STDEV.	0.910	0.480	0.230	0.129	0.110
Semi-Real Time Acquisition (Camphor)					
	rd=10s	rd=20	rd=30s	rd=40s	rd=60s
C1	2.27	1.54	1.22	1.20	1.0
C2	2.03	1.42	1.25	1.06	1.08
C3	1.99	1.35	1.06	1.01	1.02
C4	2.09	1.29	1.08	1.06	1.05
C5	0.85	0.76	0.76	0.92	0.87
C6	1.88	1.42	1.11	0.95	1.05

C7	1.00	1.00	1.00	1.00	1.00
C8	0.91	0.91	0.90	0.95	0.92
C9	2.14	1.40	1.15	1.07	1.04
C10	1.99	1.40	1.13	1.08	1.03
STDEV.	0.560	0.260	0.147	0.080	0.060
EXACT Acquisition 50% sampling density (Camphor)					
	rd =10s	rd =20s	rd =30s	rd =40s	rd =60s
C1	2.15	1.55	1.17	1.19	0.99
C2	1.95	1.45	1.24	1.07	1.04
C3	2.01	1.36	1.07	1.02	1.00
C4	1.92	1.32	1.06	1.06	1.04
C5	0.80	0.74	0.76	0.90	0.86
C6	1.86	1.42	1.11	0.95	1.03
C7	1.00	1.00	1.00	1.00	1.00
C8	0.94	0.93	0.85	0.94	0.94
C9	2.04	1.47	1.15	1.09	1.04
C10	2.01	1.40	1.14	1.11	1.05
STDEV.	0.528	0.270	0.148	0.088	0.059
EXACT Acquisition 33% sampling density (Camphor)					
	rd =10s	rd =20s	rd =30s	rd =40s	rd =60s
C1	2.1	1.42	1.05	1.07	0.97
C2	1.83	1.13	1.10	1.014	1.12
C3	1.72	1.28	1.16	1.01	0.97
C4	1.61	1.11	1.29	1.14	1.17
C5	0.89	0.8	0.73	0.9	0.9
C6	1.82	1.38	1.27	1.21	0.94
C7	1	1	1	1	1
C8	1	0.96	0.83	0.91	0.92
C9	1.84	1.32	1.06	1.14	0.97
C10	1.85	1.42	1.16	1.12	0.94
STDEV.	0.434	0.215	0.178	0.102	0.087

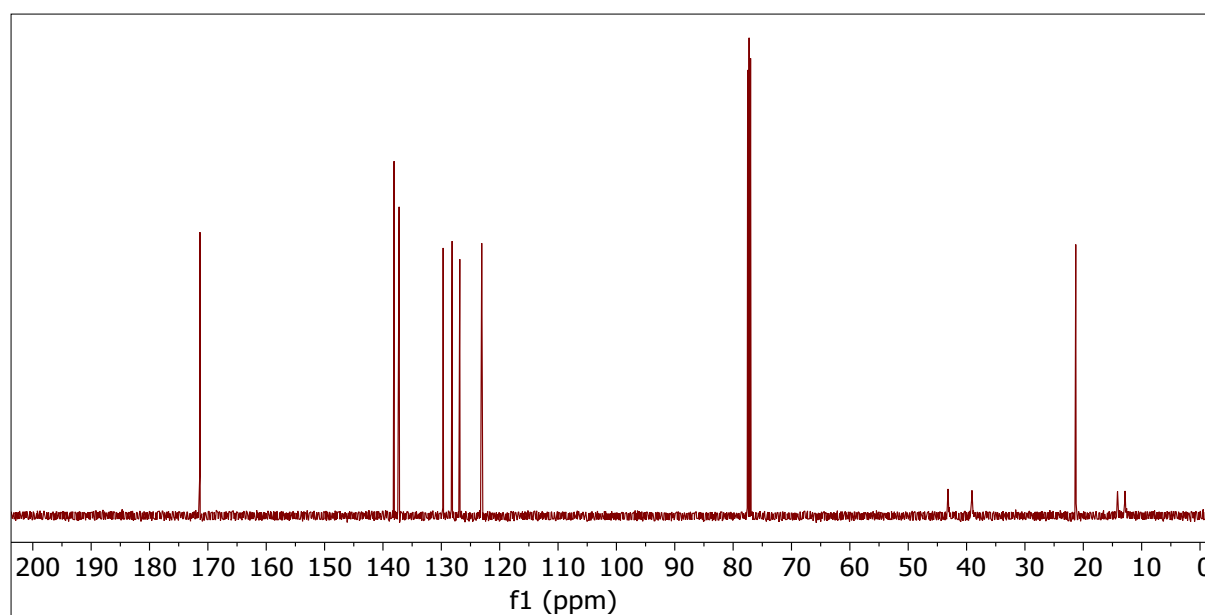
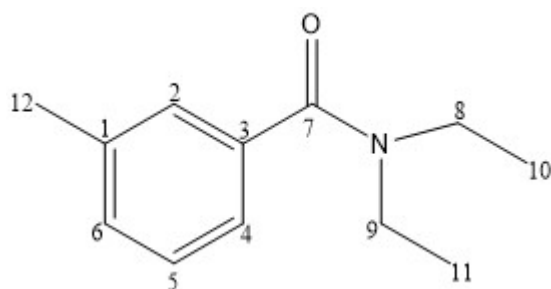


2.3 Figure S1: The standard deviations of the relative integrals of Camphor at different relaxation delays using inverse gated, SRT, and EXACT rec. Acquisition methods. The experimental time is the same using the inverse gated and the EXACT acquisition methods, however, it is double using the SRT acquisition method.

3 DEET experimental results

3.1 Table 3: DEET ^{13}C numbers and assignment in ppm

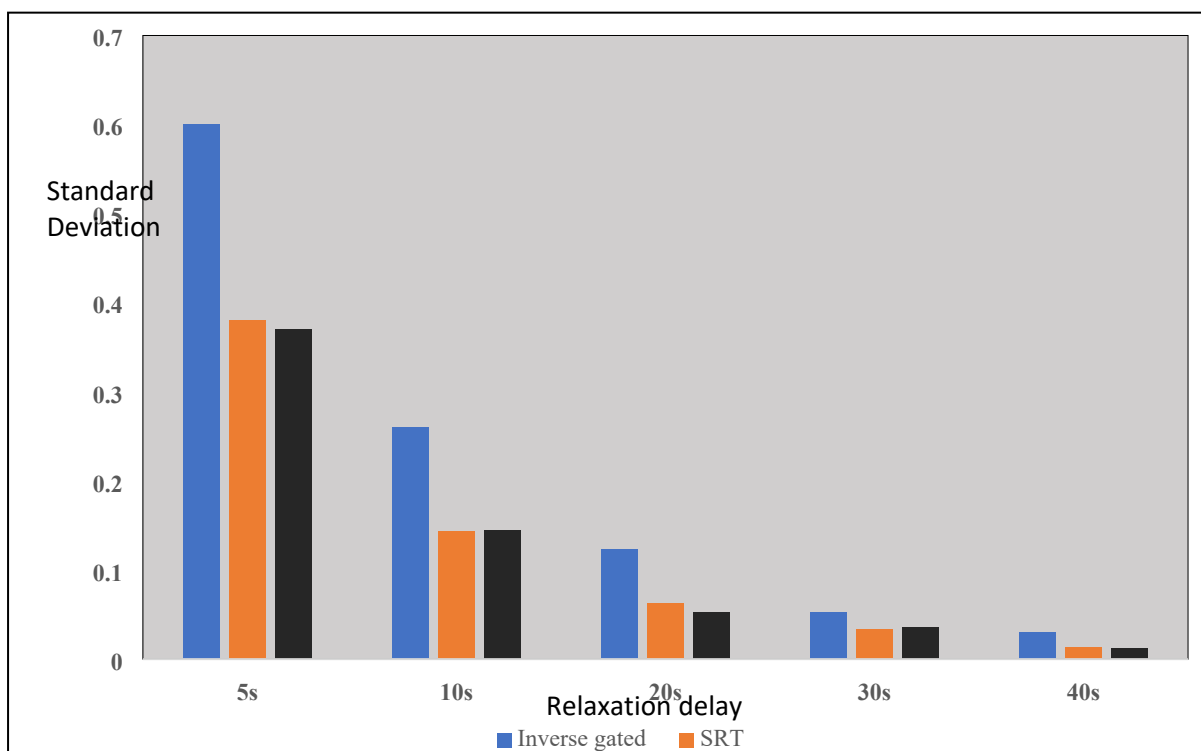
Carbon	^{13}C Assignment (ppm)
C1	138.16
C2	126.87
C3	137.23
C4	123.10
C5	128.17
C6	129.72
C7	171.43
C8	39.12
C9	43.23
C10	12.89
C11	14.18
C12	21.34



3.2 Figure S2: DEET ^{13}C NMR spectrum

3.3 Table 4: Integrals and standard deviations of DEET sample at different relaxation delays using inverse gated (IG), SRT, and EXACT acquisition methods.

Inverse Gated Acquisition (DEET)					
	rd =5s	rd =10s	rd =20s	rd =30s	rd =40s
C1	1.40	1.46	1.29	1.15	1.04
C2	2.40	1.73	1.25	1.03	1.01
C3	1.36	1.33	1.20	1.06	1.03
C4	2.32	1.73	1.25	1.06	0.95
C5	2.64	1.72	1.28	1.00	1.01
C6	2.43	1.64	1.17	0.98	0.97
C7	1.00	1.00	1.00	1.00	1.00
C12	1.93	1.70	1.43	1.05	0.97
	0.610	0.262	0.122	0.053	0.031
Semi-Real Time Acquisition (DEET)					
	rd =5s	rd =10s	rd =20s	rd =30s	rd =40
C1	1.28	1.16	1.05	1.06	1.02
C2	1.91	1.21	1.05	1.09	0.98
C3	1.25	1.04	0.99	1.06	1.00
C4	1.97	1.15	0.89	1.02	1.00
C5	1.96	1.33	0.99	1.07	1.02
C6	1.81	1.17	0.97	1.08	1.01
C7	1.00	1.00	1.00	1.00	1.00
C12	1.80	1.44	1.10	1.09	1.00
	0.383	0.143	0.063	0.0327	0.0130
EXACT Acquisition 50% sampling density (DEET)					
	rd =5s	rd =10s	rd =20s	rd =30s	rd=40
C1	1.25	1.16	1.06	1.04	1.00
C2	1.89	1.33	1.15	1.08	1.00
C3	1.20	1.09	1.05	1.03	1.02
C4	1.90	1.26	1.03	1.07	1.01
C5	1.90	1.35	1.08	1.08	1.00
C6	1.76	1.22	1.04	1.07	1.02
C7	1.00	1.00	1.00	1.00	1.00
C12	1.86	1.44	1.14	1.12	1.03
	0.377	0.144	0.052	0.036	0.0119

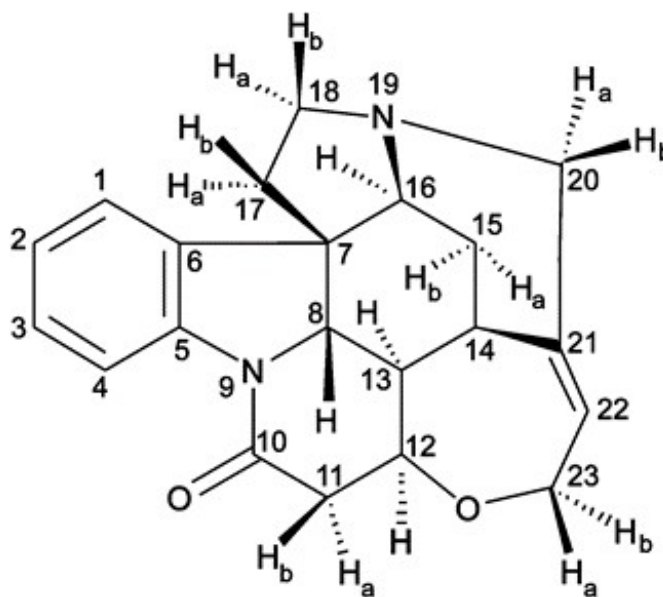


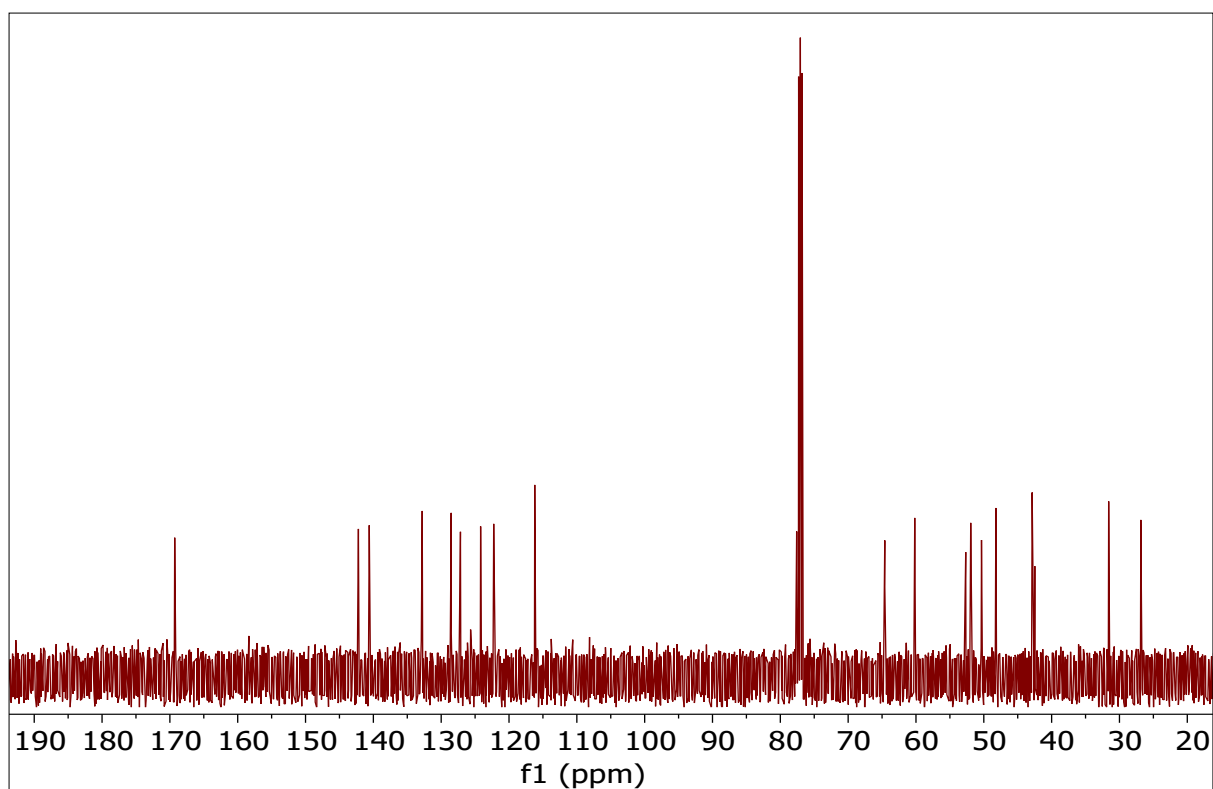
3.4 Figure S3: The standard deviations of the relative integrals of DEET at different relaxation delays using inverse gated, SRT, and EXACT rec. Acquisition methods. The experimental time is the same using the inverse gated and the EXACT acquisition methods, however, it is double using the SRT acquisition method.

4 Strychnine experimental results

4.1 Table 5: Strychnine ^{13}C numbers and assignment in ppm

^{13}C	^{13}C Assignment
C1	122.19
C2	124.12
C3	128.48
C4	116.22
C5	142.27
C6	132.8
C7	51.99
C8	60.15
C10	169.27
C11	42.47
C12	77.64
C13	48.28
C14	31.68
C15	26.9
C16	60.22
C17	42.9
C18	50.34
C20	52.71
C21	140.63
C22	127.08
C23	64.61





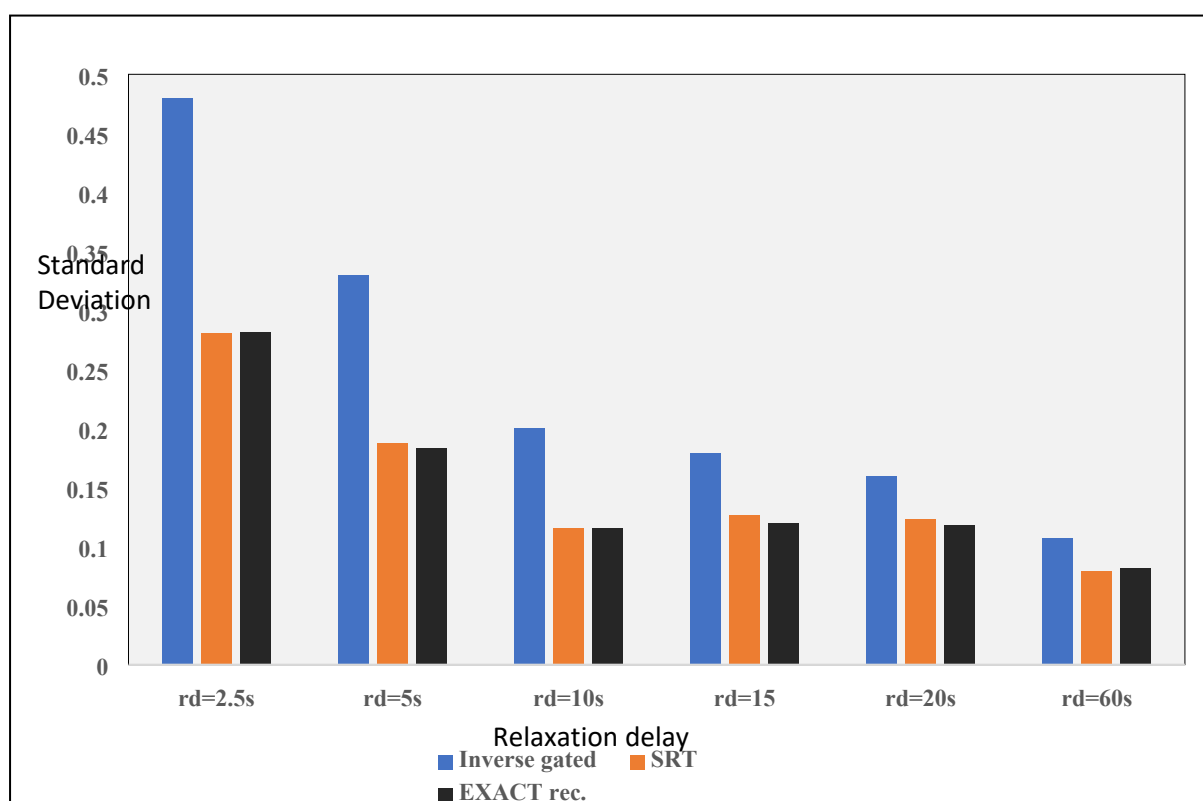
4.2 Figure S4: Strychnine ^{13}C spectrum

4.3 Table 6: Integrals and standard deviations of strychnine sample at different relaxation delays using inverse gated (IG), SRT, and EXACT acquisition methods.

Inverse Gated Acquisition (Strychnine)						
	rd=2.5s	rd=5s	rd=10s	rd=15s	rd=20 IG	rd=60 IG
C1	1.85	1.78	1.05	0.88	0.63	0.82
C2	2.01	1.74	1.08	0.84	0.75	0.64
C3	2.06	1.72	1.42	1.03	0.71	0.66
C4	2.29	2.04	1.02	0.87	0.76	0.75
C5	1.20	1.17	0.8	1.08	1.04	0.93
C6	0.93	1.14	0.73	0.89	0.75	0.93
C7	1.03	1.77	0.63	0.66	0.76	0.77
C8	2.67	1.42	0.94	1.32	1.19	0.92
C10	1.00	1.00	1.00	1.00	1.00	1.00
C11	1.57	1.51	0.84	1.08	0.76	0.83
C12	1.95	1.27	0.74	0.75	1.21	0.90
C13	2.32	1.87	1.12	1.02	0.88	0.92
C14	1.69	1.92	0.79	1.04	0.94	0.88

C15	1.11	2.04	0.96	0.95	0.77	0.80
C16	2.18	2.13	0.93	1.08	1.03	0.98
C17	1.92	1.57	1.01	0.71	0.87	0.79
C18	1.62	1.66	0.93	0.87	0.72	0.64
C20	1.16	1.3	0.82	0.69	0.94	0.74
C21	1.60	2.06	0.93	1.23	0.93	0.90
C22	1.88	1.91	0.89	0.85	1.06	0.77
C23	1.76	1.82	0.78	1.21	0.71	0.75
	0.487	0.333	0.170	0.179	0.164	0.107
Semi-Real Time Acquisition (Strychnine)						
	rd=2.5	rd=5s	rd=10s	rd=15s	rd=20s	rd=60
C1	1.40	1.3	1.19	0.93	1.04	1.10
C2	1.68	1.46	0.9	0.87	0.96	1.05
C3	1.49	1.18	0.89	0.86	0.85	0.89
C4	1.65	1.36	0.87	0.86	1.06	0.97
C5	0.99	1.05	1.07	0.81	1.01	0.85
C6	0.82	0.93	0.9	0.83	0.85	1.06
C7	0.98	1.54	0.95	1.09	0.93	1.05
C8	1.71	1.13	1.02	1.06	0.98	1.08
C10	1.00	1.00	1.00	1.00	1.00	1.00
C11	1.48	1.05	0.99	0.83	0.86	0.91
C12	1.51	1.36	0.99	1.23	1.04	1.04
C13	1.49	1.17	1.12	0.96	1	1.01
C14	1.36	1.37	0.72	0.85	0.81	1.06
C15	1.05	1.68	0.84	0.85	0.69	0.97
C16	1.55	1.10	1.00	1.18	1.17	1.11
C17	1.05	1.26	0.96	0.94	1.09	1.07
C18	1.02	1.05	0.97	0.78	0.91	0.99
C20	1.09	1.14	0.74	1.05	1.11	0.92
C21	0.9	1.16	0.83	0.8	1.07	1.08
C22	1.28	1.11	0.96	1.04	1.05	0.85
C23	1.18	1.29	0.81	0.94	0.76	1.01
	0.281	0.188	0.116	0.127	0.123	0.079
EXACT Acquisition 50% sampling density (Strychnine)						
	rd=2.5	rd=5s	rd=10s	rd=15s	rd=20s	rd=60
C1	1.41	1.36	1.32	1.12	1.17	1.08
C2	1.57	1.60	1.11	1.00	1.15	0.90
C3	1.48	1.43	1.06	0.97	1.26	0.91

C4	1.58	1.46	0.91	1.13	1.20	0.96
C5	0.93	1.19	1.10	0.92	1.07	0.83
C6	0.79	1.03	1.00	0.96	0.9	1.09
C7	1.03	1.04	1.16	1.25	0.92	0.99
C8	1.69	1.25	1.09	1.23	1.03	1.05
C10	1.00	1.00	1.00	1.00	1.00	1.00
C11	1.46	1.34	1.13	0.96	0.86	0.91
C12	1.64	1.63	1.14	1.22	1.11	1.11
C13	1.54	1.64	1.35	1.06	1.09	0.97
C14	1.43	1.3	1.08	1.03	0.84	1.09
C15	1.07	1.12	1.09	0.96	0.94	1.02
C16	1.62	1.35	1.27	1.23	0.99	1.03
C17	1.10	1.17	1.06	1.19	0.87	1.06
C18	1.03	1.24	1.14	1.06	1.02	0.96
C20	1.05	1.22	1.01	1.20	0.97	0.92
C21	0.92	1.35	0.89	0.82	1.13	1.05
C22	1.25	1.27	1.14	1.10	0.94	0.85
C23	1.19	1.33	1.00	1.13	1.09	0.91
	0.282	0.183	0.116	0.120	0.118	0.082



4.4 Figure S5: The standard deviations of the relative integrals of Strychnine at different relaxation delays using inverse gated, SRT, and EXACT rec. Acquisition methods. The experimental time is the same using the inverse gated and the EXACT acquisition methods, however, it is double using the SRT acquisition method.

5 Strychnine, Camphor, DEET mixture results

5.1 Table 7: Integrals average and standard deviation of Strychnine, Camphor, and DEET mixture using SRT acquisition method with 40s relaxation delay (rd).

SRT, rd=40s					
Strychnine		Camphor		DEET	
C1	0.12	C1	1.19	C1	1.73
C2	0.13	C3	1.02	C2	1.77
C3	0.2	C4	1.00	C3	1.74
C4	0.16	C5	0.95	C4	1.76
C5	0.15	C6	1.01	C5	1.71
C6	0.19	C7	1.00	C6	1.73
C7	0.11	C8	1.00	C7	1.73
C8	0.18	C9	0.99	C12	1.77
C10	0.17	C10	1.00		
C11	0.18				
C13	0.15				
C14	0.16				
C15	0.17				
C16	0.17				
C17	0.27				
C18	0.12				
C20	0.14				
C21	0.15				
C22	0.16				
C23	0.13				
integrals average	0.160		1.017		1.742
std.	0.035		0.067		0.0218

5.2 Table 8: Integrals, Integrals average and standard deviation of Strychnine, Camphor, and DEET mixture using EXACT (with reconstruction) acquisition method with 40s relaxation delay (rd).

EXACT rd=40s					
Strychnine		Camphor		DEET	
C1	0.12	C1	1.26	C1	1.69
C2	0.15	C3	1.00	C2	1.72
C3	0.22	C4	0.96	C3	1.69
C4	0.14	C5	0.87	C4	1.73
C5	0.14	C6	0.99	C5	1.71
C6	0.18	C7	1.00	C6	1.69
C7	0.13	C8	0.94	C7	1.72
C8	0.16	C9	0.95	C12	1.75
C10	0.16	C10	0.96		

C11	0.14				
C13	0.15				
C14	0.14				
C15	0.17				
C16	0.18				
C17	0.26				
C18	0.12				
C20	0.15				
C21	0.14				
C22	0.19				
C23	0.13				
integrals average	0.158		0.992		1.712
std.	0.034		0.108		0.021

5.3 Table 9: Integrals, Integrals average and standard deviation of Strychnine, Camphor, and DEET mixture using inverse gated (IG) acquisition method with 60s relaxation delay (rd)

IG, rd=60s					
Strychnine		Camphor		DEET	
C1	0.12	C1	1.16	C1	1.81
C2	0.15	C3	1.01	C2	1.65
C3	0.28	C4	0.95	C3	1.80
C4	0.14	C5	0.83	C4	1.77
C5	0.17	C6	1.00	C5	1.61
C6	0.17	C7	1.00	C6	1.67
C7	0.17	C8	0.87	C7	1.78
C8	0.13	C9	1.02	C12	1.70
C10	0.16	C10	1.01		
C11	0.16				
C13	0.13				
C14	0.16				
C15	0.15				
C16	0.15				
C17	0.25				
C18	0.12				
C20	0.12				
C21	0.13				
C22	0.22				
C23	0.11				
integrals average	0.159		0.983		1.723
std.	0.044		0.094		0.071

Strychnine C12 and Camphor C2 could not be integrated due to signal overlap.

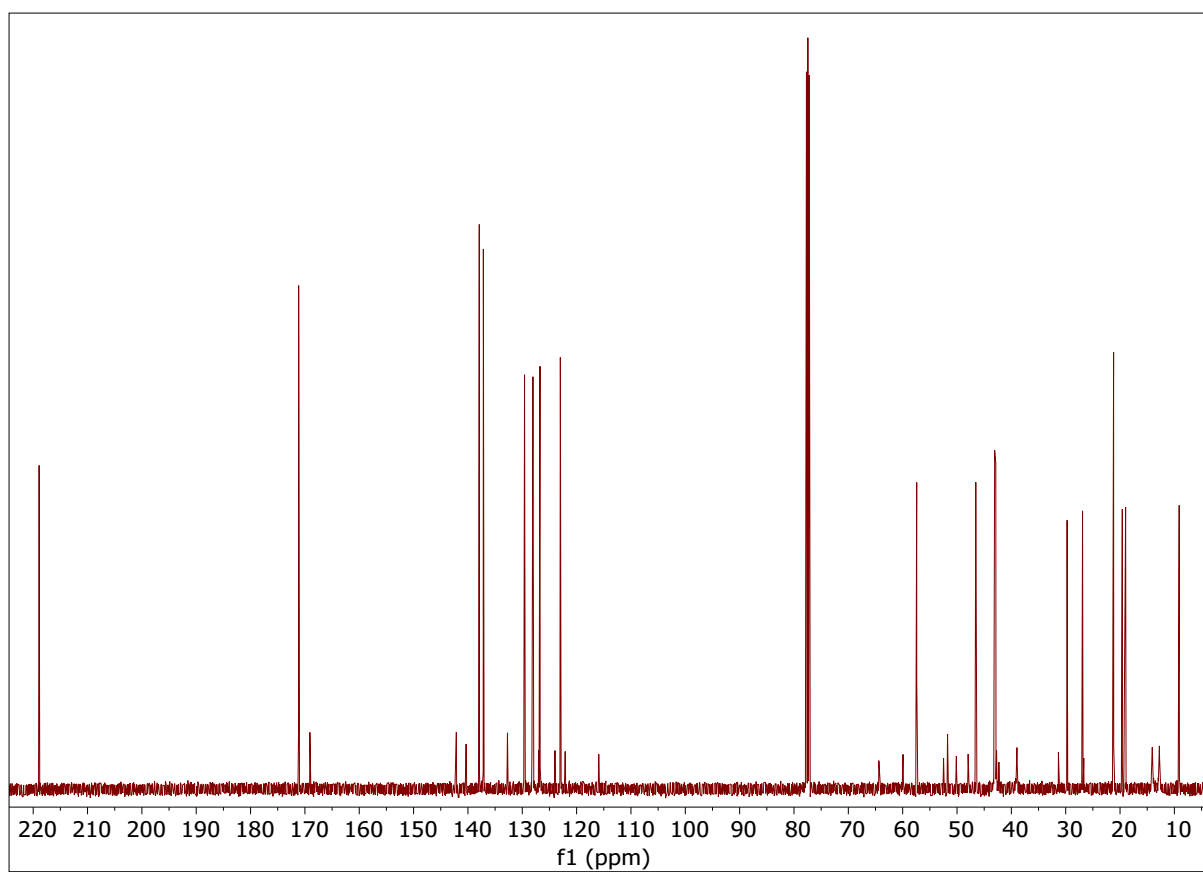


Figure 5.4 S6: Strychnine, Camphor, DEET mixture ^{13}C spectrum.

6 Varian-format pulse sequence for SRT and EXACT regular sampling

```
#include <standard.h>

void pulsequence()
{

Double

chunkpoints = getval("chunkpoints"), /*number of datapoints collected*/
cycles = getval("cycles"), /*number of cycles*/
srt = getval("srt"); /*for srt=0, FID starts with chunk, for srt=1, the FID strats with gap*/

initval(cycles,v20);
/* equilibrium period */
status(A);
hsdelay(d1);

/* --- tau delay --- */
status(B);

/* --- observe period --- */
pulse(pw,oph);

/*--Acquisition start--*/

/*Preloop acquisition for srt = 0*/

if(srt==0)
{status(C);
decpower(42); /*Decpower statement for the WALTZ-16 status C*/
```

```

rcvtron();
acquire (chunkpoints, 1/sw);
rcvroff();

status(B);
decpower(62);          /*Decpower statement for the 180 pulse status B*/
delay(((0.5*chunkpoints/(2*sw))-pwx-rof1);
decrpulse(pwx*2, 1,rof1,rof2);
delay(((0.5*chunkpoints/(2*sw))-pwx-rof2-alfa);
}

/* Preloop acquistion for srt = 1 */

if(srt==1)
{
status(C);
decpower(42);
if (srt==1) {acquire (1,1/sw);
rcvroff();

status(B);
decpower(62);
delay(((0.5*chunkpoints/(2*sw))-pwx-0.5/sw-rof1);
decrpulse(pwx*2,1,rof1,rof2);
delay(((0.5*chunkpoints/(2*sw))-pwx-rof2-alfa);}

```

```

starthardloop(v20);
    //decr(v20);
    {

status(C);
decpower(42);
rcvtron();
acquire(chunkpoints,1/sw);
rcvroff();

status(B);
decpower(62);
delay((0.5*chunkpoints/(2*sw))-pwx-rof1);    /*Delays by same length as chunk*/
decrpulse(pwx*2,3,rof1,rof2);
delay((0.5*chunkpoints/(2*sw))-pwx-rof2-alfa);

status(C);
decpower(42);
rcvtron();
acquire(chunkpoints,1/sw);
rcvroff();

status(B);
decpower(62);
delay((0.5*chunkpoints/(2*sw))-pwx-rof1);
decrpulse(pwx*2,1,rof1,rof2);
delay((0.5*chunkpoints/(2*sw))-pwx-rof2-alfa);

endhardloop();

```

```
incr(v20);
```

```
}
```