# **Supporting Information**

Water-soluble AIE-active fluorescent organic nanoparticles for ratiometric detection of SO<sub>2</sub> in mitochondria of living cells

#### **Experimental Section**

#### Materials and instruments

Unless otherwise stated, all chemicals and solvents are purchased as reagent grade and can be used without further purification. All reactions are carried out under magnetic stirring. Reactions are monitored by the analytical thin layer chromatography (TLC) on the silica F254 glass plate and displayed by UV light (254nm or 365nm) or immersed in EtOH-H<sub>2</sub>SO<sub>4</sub> (4%) before heating. Column chromatography was conducted over silica gel (mesh 200-300) or BioGel P - 2 fine resins (Bio-Rad, Hercules, CA). Nuclear magnetic resonance (<sup>1</sup>H and <sup>13</sup>C NMR) spectra were measured at room temperature with JEOL 's NMR (400 or 600 MHz) spectrometer. Mass spectra were recorded on a waters LCT Premier XEmass spectrometer or a Bruker MALDI-TOF mass spectrometer. Transmission electron microscope (TEM) images were recorded on FEI Talos F200S. Dynamic Light Scattering (DLS) measurements were performed on a Zeta potential analyzer (Brookhaven Instruments Corporation, America). All optical testing experiments were performed in PBS solution (10 mM, pH 7.4).

### UV-vis/fluorescence measurements

The probe TYDL was dissolved in water to prepare a 0.2 mM stock solution. The solutions of different analytes, including NaF(F<sup>-</sup>) NaCl(Cl<sup>-</sup>), KBr(Br<sup>-</sup>), KI(I<sup>-</sup>), NaAc(Ac<sup>-</sup>), K<sub>3</sub>PO<sub>4</sub>, (PO<sub>4</sub><sup>3-</sup>), Na<sub>2</sub>HPO<sub>4</sub>(HPO<sub>4</sub><sup>2-</sup>), K<sub>2</sub>CO<sub>3</sub>(CO<sub>3</sub><sup>2-</sup>), KHCO<sub>3</sub>(HCO<sub>3</sub><sup>-</sup>), Na<sub>2</sub>SO<sub>4</sub>(SO<sub>4</sub><sup>2-</sup>), NaClO<sub>4</sub>(ClO<sub>4</sub><sup>-</sup>), NaNO<sub>3</sub>(NO<sub>3</sub><sup>-</sup>), KSCN(SCN<sup>-</sup>), Na<sub>2</sub>S<sub>2</sub>O<sub>3</sub>(S<sub>2</sub>O<sub>3</sub><sup>2-</sup>), NaHS(HS<sup>-</sup>), GSH, Cys, BSA, Vc, NaHSO<sub>3</sub>(HSO<sub>3</sub><sup>-</sup>), TBACN(CN<sup>-</sup>) were prepared in phosphate buffer solution (PBS, 10 mM, pH 7.4).

#### Confocal imaging of living cells

HepG2 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% FBS, penicillin (100  $\mu$ g/mL), and streptomycin (100  $\mu$ g/mL) at 37 °C in a humidified incubator, and culture media were replaced with fresh media every day. The cells were further incubated with diverse concentration of probes in culture media at 37 °C and then washed 3 times with PBS buffer before cell fluorescence imaging experiments with confocal laser scanning microscopy. LO2 cells were cultured in Dulbecco's modified Eagle's medium (DMEM) supplemented with 20% FBS. Other processes were consistent with those of HepG2 cells.

# MTT assay

HepG2 cells were seeded in a 96-well plate with culture media. Each concentration of **TYDL** was incubated with cell for 24 h. After incubation, cells were incubated with cell culture media containing 0.5 mg/mL MTT (thiazolyl blue tetrazolium bromide) for 3 h and removed the media. Then the cells were dissolved in DMSO of 0.5 mL, the absorbance at 570 nm was measured. The cell viability(%) was calculated according to the following equation:

Cell viability =  $\frac{OD_{sample} - OD_{blank}}{OD_{control} - OD_{blank}} \times 100\%$ 

Synthesis of TYDL



Scheme S1. Synthetic route for probe TYDL

Compounds **3-8**<sup>17</sup> and **11**<sup>18</sup> were synthesized according to the procedures in the literatures.

Synthesis of Compound 12: Compound 12-1 (1.6 ml, 9.9mmol) and Compound 12-2 (8 ml, 9.9 mmol) were added to a 50 mL two-necked flask. The reaction mixture was heated to 85 °C overnight under reflux. After cooling to room temperature, precipitate were filtered and washed by hexane to afford the products. (3.0 g, yield 97%) <sup>1</sup>H NMR (600 MHz, Chloroform-d):  $\delta$  7.75 (dd, J = 7.34, 2.24 Hz, 1H), 7.59 (m, 3H), 4.77(q, J =7.48Hz), 3.16(s, 3H), 1.66(s, 6H), 1.63(t, J= 7.48Hz, 3H); <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  191.57, 181.55, 170.44, 170.39, 170.09, 169.72, 169.11, 148.54, 145.48, 143.63, 142.48, 141.14, 140.43, 136.36, 134.35, 132.73, 131.95, 130.98, 130.02, 129.69, 128.90, 124.80, 122.76, 119.96, 114.91, 101.29, 96.23, 72.86, 71.12, 70.73, 69.20, 66.70, 61.93, 60.85, 60.83, 53.52, 52.39, 52.39, 45.92, 45.92, 44.02, 29.76, 27.12, 22.76, 20.72, 19.80, 14.19; MS (ESI): m/z calcd for [C<sub>13</sub>H<sub>18</sub>N<sup>+</sup>]: 188.14; Found: 188.13.

Synthesis of Compound 9: To a 25 mL two-necked flask, compound 8 (50 mg, 0.0271 mmol) and compound 12 (34.1 mg, 0.1082 mmol) were added and dissolved in dry EtOH (4 mL). Pyridine (20 mL) was added and the mixture was heated under reflux for 8h. After cooling to room temperature, the solvent was removed under reduced pressure and the residue was purified by silica gel column using DCM / MeOH (50: 2, v / v) as an eluent to obtain compound 9 as an orange solid (24 mg, yield 36%). <sup>1</sup>H NMR (600 MHz, Chloroform-d): δ 8.12 (s, 2H), 7.87 (d, J = 38.4 Hz, 2H), 7.63 – 7.48 (m, 12H), 7.16 (dq, J = 17.5, 10.7, 9.5 Hz, 10H) ,5.41 (t, J = 9.5 Hz, 1H), 5.28 (s, 2H), 5.23 (s, 1H), 5.15 -5.09 (m, 2H), 5.03 (d, J = 7.5 Hz, 2H), 4.94 (s, 1H), 4.89 (dd, J = 9.2, 4.7 Hz, 2H), 4.86 - 4.81 (m, 1H), 4.72 (d, J = 10.0 Hz, 1H), 4.42 (d, J = 10.3 Hz, 5H), 4.13 – 3.94 (m, 10H), 3.87 (s, 2H), 3.84 – 3.79 (m, 2H), 3.75 – 3.63 (m, 5H), 3.57 – 3.52 (m, 1H), 3.03 (s, 6H), 2.10 – 2.04 (m, 10H), 2.02 (d, J = 5.4 Hz, 3H), 1.98 (s, 22H), 1.90 (d, J = 5.2 Hz, 7H), 1.85 – 1.69 (m, 14H), 1.54 (s, 4H); <sup>13</sup>C NMR (151 MHz, Chloroform-d) & 191.57, 181.55, 170.44, 170.39, 170.09, 169.72, 169.11, 148.54, 145.48, 143.63, 142.48, 141.14, 140.43, 136.36, 134.35, 132.73, 131.95, 130.98, 130.02, 129.69, 128.90, 124.80, 122.76, 119.96, 114.91, 101.29, 96.23, 72.86, 71.12, 70.73, 69.20, 66.70, 61.93, 60.85, 60.83, 53.52, 52.39, 52.39, 45.92, 45.92, 44.02, 29.76, 27.12, 22.76, 20.72, 19.80, 14.19; HRMS (ESI): m/z calcd for [C114H130N8O36]<sup>2+</sup>: 1093.9305; Found: 1093.9267.

**Synthesis of Compound 10 (TYDL):** To a 25mL eggplant shaped bottle, compound **9** (50 mg, 0.0204 mmol) in dry MeOH (8 mL) was added and 1 M NaOMe in MeOH was slowly added dropwise at room temperature. The resulting mixture was stirred for 1 hour at room temperature. The reaction pH was then adjusted with Amberlite IR-120 plus (H<sup>+</sup>) to pH 6 and the solvent was removed under reduced pressure. (27 mg, yield: 71%).<sup>1</sup>H NMR (600 MHz, DMSO-d6):  $\delta$  8.62 (d, J = 18.9 Hz, 2H), 8.05 (d, J = 7.3 Hz, 1H), 7.87 (d, J = 24.2 Hz, 1H), 7.73 (d, J = 8.0 Hz, 2H), 7.69 – 7.63 (m, 12H), 7.37 – 7.21 (m, 10H), 5.30 (s, 2H), 5.06 (s, 4H), 4.77 (s, 1H), 4.70 – 4.64 (m, 2H), 4.61 (s, 2H), 4.55 (s, 2H), 4.49 (s, 2H), 4.26 (s, 1H), 4.12 – 4.08 (m, 3H), 4.02 (s, 1H), 3.73 – 3.67 (m, 4H), 3.58 (s, 5H), 3.50 – 3.40 (m, 10H), 3.02 (s, 1H), 2.93 (s, 4H), 1.99 – 1.91 (m, 5H), 1.65 – 1.56 (m, 18H), 1.51 (d, J = 10.7 Hz, 3H), 1.45 – 1.42 (m, 2H). HRMS (ESI): m/z calcd for [C<sub>86</sub>H<sub>102</sub>N<sub>8</sub>O<sub>22</sub>]<sup>2+</sup>: 799.3549; Found: 799.3545.

Entry	<b>Probe Structure</b>	AIE active	Response mode	Water solubility	Can nanoparticles be formed?	Cell experiment	LOD	Ref.
1		Yes	Ratiometric	PBS buffer	Yes	Yes	75 nM	This work
2		Yes	Ratiometric	CH <sub>3</sub> CN: H <sub>2</sub> O=3:7 (v/v)	No	No	1.04 ppm (1×10 <sup>-5</sup> M)	[1]
3	×N C C C N So3	Yes	Ratiometric	PBS buffer/ DMSO=9/1	No	Yes	0.17 μΜ	[2]
4	но	Yes	Turn- on	DMF/H <sub>2</sub> O (v: v = 1:9)	No	No	38.4 nM	[3]
5		Yes	Turn- on	EtOH /PBS buffer = 4:6(v/v)	No	Yes	26 nM	[4]

Table S1. Comparison of TYDL with some other reported HSO<sub>3</sub><sup>-</sup> selective fluorescent probes.

6	$\begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & \\ & & & & \\ & &$	Yes	Turn-off	CH3OH/H2O=1/9 (v/v)	No	No	97.1nM 100.2nM 77.1nM	[5]
7	СНО	Yes	Turn-off	DMSO/HEPES =1/9	No	Yes	203.5 nM	[6]
8		Yes	Turn-off	THF/water =1/999 $(v/v)$	No	No	0.39 mM	[7]
9		No	Ratiometric	PBS buffer ,containing 30% DMF	No	Yes	0.38 mM	[8]
10		No	Ratiometric	PBS buffer containing 20% glycero	No	Yes	0.34 mM	[9]
11		No	Ratiometric	PBS buffer	No	Yes	0.34 mM	[10]
12		No	Ratiometric	DMSO/PBS buffer =4/6, pH = 7.4	No	Yes	15.1 nM	[11]

13	S N+	No	Ratiometric	HEPES buffer /DMF= 7:3	No	Yes	1.45 ×10 <sup>-7</sup> M	[12]
14		No	Ratiometric	EtOH/PBS buffer = 5:5	No	Yes	66 nM	[13]
15		No	Turn off	PBS containing 20% CH <sub>3</sub> CH <sub>2</sub> OH	No	Yes	6.29×10 <sup>-6</sup> M	[14]
16		No	Turn on	EtOH/Tris-HCl=1:9	No	Yes	0.14 mM	[15]
17	NO <sub>2</sub>	No	Turn off	DMSO/PBS buffer =1:9	No	No	8.7 μΜ	[16]



**Fig. S1.** Time-dependent fluorescence spectral change of probe **TYDL** (5  $\mu$ M) upon addition of HSO<sub>3</sub><sup>-</sup> (200  $\mu$ M) in pure water. (b) Fluorescence intensity ratio (*I*<sub>550</sub>/*I*<sub>697</sub>) of probe **TYDL** (5  $\mu$ M) upon addition of HSO<sub>3</sub><sup>-</sup> (200  $\mu$ M) in pure water.  $\lambda_{ex} = 300$  nm.



**Fig. S2.** a) Absorption spectral changes of **TYDL** (10  $\mu$ M) upon titration with HSO<sub>3</sub><sup>-</sup>(0-500  $\mu$ M) in pure water. b) the detection limit of **TYDL**.



**Fig. S3.** The effect of various pH values on the fluorescence ratio ( $I_{550}/I_{697}$ ) of **TYDL** (5  $\mu$ M) in the presence or absence of HSO<sub>3</sub><sup>-</sup> (200  $\mu$ M). Spectra were recorded after 10 min incubation of **TYDL** with HSO<sub>3</sub><sup>-</sup>.



Fig. S4. MS of Compound TYDL' [(TYDL-HSO<sub>3</sub>)<sup>2+</sup>].



Fig. S5. (a) Fluorescence spectra of different concentrations of TYDL in pure water. (b) The curve of fluorescence intensity at 697 nm versus the concentration of TYDL in pure water. The crosspoint corresponds to the CMC of TYDL.  $\lambda ex = 300$  nm



**Fig. S6.** Effects of different concentrations of **TYDL** on the viability of HepG2 cells. The results are the mean standard deviation of three separate measurements.



**Fig. S7.** Fluorescence images of **TYDL** responding to  $HSO_3^-$  in HepG2 cells. From left to right: HepG2 cells pretreated with TYDL (5  $\mu$ M) for 30 min, incubated with 200  $\mu$ M NaHSO<sub>3</sub> for 0, 30, 60, 90 min, and then imaged. From up to down: Red channel ( $\lambda_{em} = 600-750$  nm); Green channel ( $\lambda_{em} = 500-550$  nm); Bright-field; Merge.



**Fig. S8.** From up to down: HepG2 cells pretreated with **TYDL** (5  $\mu$ M) alone for 60 min, sequentially incubated with 200  $\mu$ M NaHSO<sub>3</sub> (60 min) and 5  $\mu$ M **TYDL** (60 min), and then imaged. From left to right: Red channel ( $\lambda_{em} = 600-750$  nm); Green channel ( $\lambda_{em} = 500-550$  nm); Bright-field; Merge.



**Fig. S9.** From up to down:LO2 cells pretreated with **TYDL** (5  $\mu$ M) alone for 60 min, sequentially incubated with 200  $\mu$ M NaHSO3 (60 min) and 5  $\mu$ M **TYDL** (60 min), and then imaged. From left to right: Red channel ( $\lambda_{em} = 600-750$  nm); Green channel ( $\lambda_{em} = 500-550$  nm); Bright-field; Merge.

#### <sup>1</sup>H NMR, <sup>13</sup>C NMR and HRMS Spectra of Compounds a=7.01811156174134060e-004, t0=-1.44510196286552570e-001 (DuoSpray ())



Fig. S11. <sup>1</sup>H NMR Spectrum of Compound 12



Fig. S13.MS Spectrum of Compound 9

# 





# Fig. S16.MS Spectrum of Compound 10



Fig. S17. <sup>1</sup>H NMR Spectrum of Compound 10

# References

- [1] T. Lin, X. Su, K. Wang, M. Li and S. Zhang, *Mater. Chem. Front.*, 2019, 3, 1052.
- [2] M. Lv, Y. Zhang, J. Fan, Y. Yang, S. Chen, G. Liang and S. Zhang, Analyst, 2020, 145, 7985.
- [3] X. Cai, J. Ye, Q. Zhou, Z. Yan and K. Li, *Microchem. J.*, 2020, 159, 105419.
- [4] G. Yin, Y. Gan, T. Yu, T. Niu, P, Yin, H. Chen, Y. Zhang, H. Li and S. Yao, *Talanta*, 2019, **191**, 428.
- [5] H. Yu, J. Zhi and J. Wang, J. Mater. Chem., 2021, 9, 3882.
- [6] B. Wang, M. Lv, W. Wu, Z. Xu, Y. Fan, L. Bian and Y. Wang, *J. Photoch. Photobio*. A, 2021, **411**, 113193.
- [7] B. Yang, X. Niu, Z. Huang, C. Zhao, Y. Liu and C. Ma, Tetrahedron, 2013, 69, 8250.
- [8] Y. Sun, J. Liu, J. Zhang, T. Yang and W. Guo, Chem. Commun., 2013, 49, 2637.
- [9] C. Sun, W. Cao, W. Zhang, L. Zhang, Y. Feng, M. Fang, G. Xu, Z. Shao, X. Yang and X. Meng, *Dyes Pigm.*, 2019, **171**, 107709.
- [10] Y. Zhang, L. Guan, H. Yu, Y. Yan, L. Du, Y. Liu, M. Sun, D. Huang and S. Wang, *Anal. Chem.*, 2016, **88**, 4426.
- [11] J. Lan, R. Zeng, Y. Ding, Y. Zhang, T. Zhang and T. Wu, Sens. Actuators B Chem., 2018, 268, 328.
- [12] U. Diwan, V. Kumar, R. Mishra, N. Rana, B. Koch, M. Singh, K. Upadhyay, Anal. Chim. Acta., 2016, **929**, 39.
- [13] M. Huang, L. Chen, J. Ning, W. Wu, X. He, J. Miao and B. Zhao, *Sens. Actuators B Chem.*, 2018, **261**, 196.
- [14] Q. Hu, R. Guo, L. Zhang, Q. Liu, S. Cai and W. Lin, Luminescence, 2021, 36,1006.
- [15] C. Wang, M. Yang, X. Deng and M. He, Chinese J. Inorg. Chem., 2020, 36, 762.
- [16] X. Li, D. Jin, Y. Du, Y. Liu, B. Wang and L. Chen, Anal. Methods, 2018, 10, 4695.
- [17] M. Hou, Y.-c. Liu, W. Zhou, J.-d. Zhang, F.-d. Yu, Y. Zhang, G.-j. Liu, G.-w. Xing, *Chem.-Asian J.*, 2021, **16**, 2014.

[18] Z.-w. Ning, S.-z. Wu, G.-j. Liu, Y.-m. Ji, L.-y. Jia, X.-x. Niu, R.-f. Ma, Y. Zhang, G.-w. Xing, *Chem.-Asian J.*, 2019, **14**, 2220.