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Supporting information

C(sp³)–H Ritter amination by excitation of *in situ* generated Iodine (III) - BF₃ complexes

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1 General information

Starting materials and reagents were purchased from commercial suppliers (Sigma Aldrich, Alfa Aesar, Acros or Fluka) and were used without further purification. Solvents were used as p.a. grade. Reactions were monitored by analytic thin layer chromatography (TLC) using Fluka silica gel or NH2-Modified Silica Plates with a fluorescent indicator. Visualization of the developed TLC chromatogram was performed using 254 nm UV light source. Organic solutions were concentrated using Büchi rotary evaporator. Flash column chromatography was performed either by hand in filled Pasteour pipettes or on a Biotage[®] IsoleraTM Spektra. The columns were filled either with Silica gel (60-200 μ m) or basic alumina (50-200 μ m).

NMR spectroscopy

All NMR spectra were recorded at room temperature using a Bruker Avance 300 (300 MHz for ¹H, 75 MHz for ¹³C, 282 MHz for ¹⁹F) or a Bruker Avance 400 (400 MHz for ¹H, 101 MHz for ¹³C, 376 MHz for ¹⁹F) NMR spectrometer. All chemical shifts are reported in δ -scale as parts per million [ppm] (multiplicity, coupling constant J, number of protons), relative to the solvent residual peaks as the internal standard. Coupling constants J are given in Hertz [Hz]. Abbreviations used for signal multiplicity: ¹H-NMR: br = broad, s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, dt = doublet of triplets, and m = multiplet.

Gas chromatography

GC measurements were performed on a GC 7890 from Agilent Technologies system coupled to a FID. The system was equipped with a capillary column (HP-5ms UI, length 30 m, diam. 0.25 mm, film 0.25 μ m) and worked with H₂ as carrier gas. GC program: The initial temperature of the GC was set to 40 °C and kept for 1.5 minutes. Subsequently, the oven temperature was increased at a rate of 25 °C/min. until reaching 280 °C, which was maintained for 3 min. Then, temperature was further increased (42 °C/min) until reaching 300 °C and final temperature was hold for 5 minutes. Injector temperature was set to 280 °C and temperature of the detecting unit to 310 °C. A split ratio of 30:1 (split flow 42 mL/min) was applied, and the column flow was set to 1.4 mL/min. Data acquisition and evaluation was done with Agilent ChemStation Rev.C.01.04.

Mass spectrometry

High resolution mass spectrometry (HRMS) was performed at the Central Analytical Laboratory of the University of Regensburg. Mass spectra were measured on a Finnigan MAT 95, ThermoQuest Finnigan TSQ 7000, Finnigan MAT SSQ 710 A or Agilent Q-TOF 6540 UHD instrumenta and a Waters Acquity UPLC system equipped with Waters PDA, sample manager, sample organiser, column oven and Waters Xevo QTOF mass spectrometer.

UV-Vis

Absorption spectra were measured on an Agilent Cary 100 UV/Vis spectrometer in a 10 mm × 10 mm quartz cuvette at 25.0 °C under air atmosphere.

1.1 Photochemical setups

Photochemical reactions were performed in sealed reaction vials, placed approximately 2 cm above a 400 nm LED array and stirred under irradiation (**Figure S1**). The reaction temperature was controlled by a thermostated (25 °C) metal cooling block. The reactor setup is a custom-made device (University of Regensburg workshop) and is not a commercially available product.

A) regular 400 nm setup (optical power 100-120 mW/ LED spot)



B) Higher power 400 nm setup (optical power 600-800 mW/ LED spot)







Figure S1: Photochemical setups used in this work from different perspectives.

1.2 General experimental procedures

General procedure for preparative scale reactions

The reactions were set in 2 parallel vials, each containing 0.2 mmol of the substrate. A C(sp³)–H precursor (0.2 mmol, 1 eq.), 4-tertbutylbenzoic acid (17.8 mg, 0.1 mmol, 0.5 eq.) and Selectfluor (142 mg, 0.40 mmol, 2 eq.) were weighed into a crimp reaction vial. Then, 2 ml of a stock solution containing iodobenzene (6.7 μ l, 0.06 mmol, 0.3 eq.) and H₂O (3.6 μ l, 0.2 mmol, 1 eq.) in dry MeCN were added. A stirring bar was added, and the vial was capped. The reaction mixture was degassed by 4 nitrogen-vacuum cycles. Afterwards BF₃ (200 μ l of 16% BF₃ in MeCN, 0.4 mmol, 2 eq.) was added, and the vial was placed in a thermostated cooling block (25 °C) and irradiated through the plane bottom side of the vial by a 400 nm

LED for 16 hours (see **Figure S1**). After the completion, the two reaction batches were combined, and the solvent was removed *in vacuo*. The solid residue was dissolved in 2 ml H₂O and 4 ml EtOAc and transferred into a separatory funnel containing 2 ml of brine. The aqueous layer was washed two times with 5 ml of EtOAc. Combined organic phases were concentrated *in vacuo* to ca 0.5 ml solvent volume. The sample was filtered through a small basic alumina column (packed in a Pasteur pipette). The column was then washed with approximately 5 ml of EtOAc. In that way, we remove remaining hypervalent iodine species, 4-tertbutylbenzoic acid and polymeric side products. The obtained eluent always contained only the product and some remaining substrate. The solvent was removed in vacuo and the product mixture was redissolved in a minimal amount of DCM and applied on silica gel column (packed in a Pasteur pipette). The column was first washed with DCM (5 ml) to elute the remaining substrate, and then with EtOAc (5 ml) to elute the product. Removal of the solvent afforded analytical pure samples of the products. In some cases, we were able to skip the second (silica gel) column. The decision was based on the TLC of the eluent after the first column.

General procedure for screening and optimization studies:

Selectfluor (typically 71 mg, 0.2 mmol, 2 eq.) was weighed into a crimp reaction vial together with any other solid materials (different ligands, Lewis acids/bases, PIDA, PIFA, ...). Then, 1 ml of a stock solution containing, 4-fluorotoluene (11.0 μ l, 0.1 mmol, 1 eq.), iodobenzene (3.3 μ l, 0.03 mmol, 0.3 eq.) and H₂O (1.8 μ l, 0.1 mmol, 1 eq.) in dry MeCN were added. A stirring bar was added, and the vial was capped. The reaction mixture was degassed by 4 nitrogen-vacuum cycles. Afterwards, BF₃ (100 μ l of 16% BF₃ in MeCN, 0.2 mmol, 2 eq.) or any other liquids (Lewis acids) were added, and the vial was placed in a thermostated cooling block (25 °C) and irradiated through the plane bottom side of the vial by a 400 nm LED for 16 hours (see **Figure S1**). After the completion, the analysis was done following the typical procedure for 19F NMR yield determination (*vide infra*).

Note: The reaction is a bit sensitive (**Table S3**) to the amount of water in the reaction mixture. Stock solutions containing 4-fluorotoluene, H_2O and PhI were always used in order to have all the entries within the series (optimization table) comparable.

Procedure for NMR yield determination

The screening and optimization reactions were all performed on a 0.1 mmol scale following the abovedescribed general procedure. After the reaction completion, internal standard trifluorotoluene (12.3 μ l, 0.1 mmol) was added into the vial. The reaction vial was well shaken before we took out 0.4 ml of the reaction mixture and transferred it into a 2 ml conical bottom Eppendorf vial. Then 0.4 ml CDCl₃ was added to precipitate the remaining Selectfluor and quench the reaction. The vials were centrifuged, and the clean solution was submitted for ¹⁹F NMR analysis.

2 UV-Vis of hypervalent iodine (III)-BF₃ complex of 1i

The absorption increases upon addition of BF_3 to the hypervalent iodine reagent **1i** due to the formation of the complex **1i-BF_3**.¹



Figure S2: Spectral changes upon addition of BF_3 to 1i.

3 Optimization tables

3.1 Screening of different Lewis acids

Different acids have been screened for the C-H Ritter type amination. Most of the tested acids are ineffective supporting our previously disclosed findings about necessity of complexation of the hypervalent iodine reagent with $BF_{3.}^2$

Table S1: Screening of different Lewis acids.

F	ic L	Selecfluor (2 eq.) odobenzene (30 mol%) penzoic acid (50 mol%) ewis or Bronsted acid H ₂ O (1 eq.) MeCN 400 nm, 12 h 25 °C, N ₂	F	NHAc
	Entry	Acid/base	Yields [%]	
	Α	-	14	
	В	Cs ₂ CO ₃ (2 eq.)	0	
	С	TFA (2 eq.)	15	
	D	H ₂ SO ₄ (2 eq.)	48	
	E	HClO ₄ (70%, 2 eq.)	36	
	F	HFIP (2 eq.)	13	
	G	BF ₃ (16% in MeCN, 2 eq.)	77	
	Н	BF ₃ × 2H ₂ O (2 eq.)	21	
	I	B(OH)₃ (2 eq.)	18	
	J	HBF_4 (32% in water, 2 eq.)	26	
	К	LiBF ₄ (2 eq.)	13	
-	L	Zn(OTf) ₂ (0.5 eq.)	20	
_	М	Sc(OTf)₃ (0.5 eq.)	28	

0.1 mmol substrate, 30 mol% of the PhI, 50 mol% benzoic acid, 2 eq. Selectfluor, 2 eq. or 0.5 eq. of the indicated Lewis acid, 1 eq. H_2O , 1 ml MeCN (0.1 M) under N_2 atmosphere, 25 °C, 12 h. Yields were determined by ¹⁹F NMR using trifluorotoluene as internal standard.

3.2 Screening of different iodoarenes



Scheme S1: Screening of different iodoarenes, precursors of the *in situ* formed hypervalent iodine (III) reagent. ^a 0.1 mmol substrate, 30 mol% of the indicated ArI, 50 mol% benzoic acid, 2 eq. Selectfluor, 2 eq. BF₃ × MeCN, 1 eq. H₂O, 1 ml MeCN (0.1 M) under N₂ atmosphere, 25 °C, 12 h. Yields were determined by ¹⁹F NMR using trifluorotoluene as internal standard.

lodoarenes **10** and **11** underwent decomposition after the addition of BF₃ even without light irradiation.

3.3 Screening of different HAT catalysts



Scheme 2: Optimization of the ligand of the hypervalent iodine (III) reagent for hydrogen abstraction. ^a 0.1 mmol substrate, 30 mol% PhI, 50 mol% of the indicated HAT reagent, 2 eq. Selectfluor, 2 eq. BF₃ × MeCN, 1 eq. H₂O, 1 ml MeCN (0.1 M) under N₂ atmosphere, 25 °C, 12 h. Yields were determined by ¹⁹F NMR using trifluorotoluene as internal standard.

Table S2: Correlation of substituent electronic properties of the benzoic acid with the reaction yield
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Substituent	Hammet constant $\sigma_p{}^3$	Reaction yield	Note
-OMe	-0.27	12%	Decomposition
<i>-t-</i> Bu	-0.20	83%	
-Me	-0.17	81%	
-H	0.00	77%	
-F	0.06	76%	
1	0.18 or	41%	4-lodobenzoic gets oxidized under the
-1	0.88 for -I(OAc) ₂		reaction conditions.
-Cl	0.22	76%	
-Br	0.23	58%	
COOH	0.45	2.40/	Solubility issue. The iodine (III) reagent is
-000	0.45	۲4%	not dissolved even after addition of BF ₃ .
-CF ₃	0.54	58%	

3.4 Optimization of catalysts loadings and reagent quantities

Table S3:Effect of catalysts loadings and reagent quantities.



Entry	Selectfluor [eq.]	4-tert-butylbenzoic acid [mol%]	Phl [mol%]	BF₃ [eq.]	H ₂ O [eq.]	λ [nm]	Yield [%]
Α	1	50	30	2	1	400	57
В	1.3	50	30	2	1	400	69
С	1.5	50	30	2	1	400	77
D	2	50	30	2	1	400	83
E	2	30	30	2	1	400	71
F	2	10	30	2	1	400	32
G	2	50	20	2	1	400	77
Н	2	50	10	2	1	400	74
I	2	50	30	1	1	400	35
J	2	50	30	3	1	400	80
К	2	50	30	2	5	400	21
L	2	50	30	2	1	455	79
M	2	50	30	2	1	525	21

^a All the entries are deviations of the entry D: 0.1 mmol substrate, 30 mol% PhI, 50 mol% of the 4-*tert*-butylbenzoic acid, 2 eq. Selectfluor, 2 eq. BF₃ × MeCN, 1 eq. H₂O, 1 ml MeCN (0.1 M) under N₂ atmosphere, 25 °C, 12 h. Yields were determined by ¹⁹F NMR using trifluorotoluene as internal standard.

3.5 Control experiments with different iodine (III) sources and additives

Table S4: Control experiments with preformed iodine (III) reagent.



Entry	lodine (III) reagent	BF ₃	Additive	Yield [
Α	PhI(OAc) ₂ (2 eq.)	Yes	-	19
В	PhI(OAc) ₂ (2 eq.)	-	-	3
С	PhI(OC(O)CF ₃) ₂ (2 eq.)	Yes	-	8
D	PhI(OC(O)CF ₃) ₂ (2 eq.)	-	-	7
E	Selectfluor (2 eq.) + PhI (0.3 eq.)	Yes	-	77
F	Selectfluor (2 eq.) + PhI (0.3 eq.)	Yes	AcOH (4 eq.)	73
G	Selectfluor (2 eq.) + PhI (0.3 eq.)	Yes	TFA (4 eq.)	77

The approach by *in situ* generation of aryl iodine (III) reagent gives higher yield than the use of commercially available aryliodine (III) reagents. A possible reason could be competitive decarboxylation of (trifluroo)acetate ligands in the reaction conditions. This parasitic reaction leads to unproductive consumption of the hypervalent iodine (III) reagent.

4 Synthesis and characterization of starting materials, intermediates and products

4.1 Synthesis and characterization data of substrates and intermediates

Synthesis of hypervalent iodine reagent 1i



The reagent was prepared following adapted literature procedure.⁴ Phenyliodine (III) diacetate (PIDA, 482 mg, 1.50 mmol, 1.0 eq.) and 4-*tert*-butylbenzoic acid (547 mg, 3.08 mmol, 2.1 eq.) were dissolved in toluene (25 mL) and the toluene was removed with a rotary evaporator (50 °C) over 10 minutes. After its removal, toluene (25 mL) was added again and removed with the rotary evaporator. This process was repeated three times in total. After drying *in vacuo*, the remaining solid material was dissolved in CDCl₃ and submitted for ¹H- and ¹³C-NMR measurements. The reagent was used in the mechanistic experiments without any additional purification.

Iodobenzene bis(4-tert-butylbenzoate) (1i)



Yield	Quant, White powder
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 8.23 (d, <i>J</i> = 7.6 Hz, 2H), 7.89 (d, <i>J</i> = 8.4 Hz, 4H), 7.60 (t, <i>J</i> = 7.4
	Hz, 1H), 7.52 (t, <i>J</i> = 7.6 Hz, 2H), 7.39 (d, <i>J</i> = 8.4 Hz, 4H), 1.32 (s, 18H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 171.4, 156.1, 134.8, 131.6, 130.9, 130.0, 127.5, 125.2, 122.6,
	35.1, 31.2.



p-Cresol (2 mmol, 216 μ I) and triethylamine (6 mmol, 835 μ I) were dissolved in DCM (3 mI) in a 10 mI reaction vial and cooled down to 0 °C. Substituted benzoyl chloride (2 mmol) was dissolved in DCM (3 mI) in another vial and slowly added (ca. 5 min) to the reaction mixture in the first vial. After addition, the ice bath was removed, and the reaction mixture was stirred for 5 h at rt. After the reaction completion, the reaction mixture was transferred into a separatory funnel, diluted with DCM (20 mI), and quenched with HCl (10 mI; 0.1 M). The organic layer was washed again with a base NaOH (10 mI; 0.1 M) to remove any remaining benzoyl chloride or substrate, and then once again with HCl (10 mI; 0.1 M). The organic fraction was then dried over Na₂SO₄, filtered, and concentrated in vacuo. The products were analyzed by NMR spectroscopy and used without any additional purification.

p-tolyl 2,3,4,5,6-pentafluorobenzoate (10a)



Yield	77%, White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.24 (d, <i>J</i> = 8.3 Hz, 2H), 7.12 (d, <i>J</i> = 8.5 Hz, 2H), 2.38 (s, 3H).
¹³ C NMR	^{13}C NMR (101 MHz, CDCl ₃) (except for C ₆ F ₅) δ 157.7, 147.9, 136.6, 130.2, 120.9, 20.9.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl ₃) δ -137.8 − -137.9 (m), -148.0 (tt, <i>J</i> = 20.9, 4.8 Hz), -160.4 − - 160.6 (m).
HR-MS (EI)	(M) ⁺ : calc. 302.0361, found 302.0355



Yield	72%, colourless oil
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 8.65 (s, 2H), 8.14 (s, 1H), 7.26 (d, $J = 8.2$ Hz, 2H), 7.15 – 7.06 (m, 2H), 2.40 (s, 2H)
¹³ C NMR	(m, 2H), 2.40 (s, 3H). ¹³ C NMR (101 MHz, CDCl ₃) δ 162.8, 148.2, 136.3, 132.6 (q, <i>J</i> = 34.0 Hz), 132.0, 130.3 (bs),
	126.9 (p, J = 3.8 Hz), 122.8 (q, J = 273.0 Hz), 121.0, 20.9.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl ₃) δ -63.4.
HR-MS (EI)	(M) ⁺⁻ : calc. 348.0579, found 348.0574



The Suzuki cross-coupling reactions⁵ were performed by dissolving the 4-Methylphenylboronic acid (204 mg, 1.50 mmol, 1.5 eq.) and sodium bicarbonate (138 mg, 1.00 mmol, 1.0 eq.) as a base in a mixture of ethanol and water (1:1, 8 mL) inside a 10 mL crimp vial. To each vial, an aryl bromide and Pd/C as catalyst (31.8 mg corresponding to 0.03 mmol, 0.03 eq. of pure Pd) was added before it was closed, degassed and set under N₂ atmosphere. The reaction mixtures were stirred overnight at elevated temperature (16 h, 80 °C). After the completion, most of the ethanol was removed *in vacuo*, and the products were isolated by extraction (ethyl acetate/brine 1:1, 15 mL each). The product-containing organic phases were dried over sodium sulfate and after filtration, a spoon-full of flush silica gel (60M, Ø 0.04 – 0.036 mm) was added to prepare a dry load for a column chromatography. The dry load was transferred onto a silica gel column and the products were eluted with ethyl acetate as eluent (a single fraction collected). Products were analyzed by NMR spectroscopy.



48%, white solid
¹ H NMR (400 MHz, CDCl ₃) δ 8.08 (d, J = 8.4 Hz, 2H), 7.85 (d, J = 8.5 Hz, 2H), 7.55 (d, J = 8.1
Hz, 2H), 7.33 (d, <i>J</i> = 7.9 Hz, 2H), 2.44 (s, 3H).
^{13}C NMR (101 MHz, CDCl_3) δ 149.6, 139.7, 135.5, 131.3, 130.0, 129.1, 128.1, 127.4, 119.9
(q, <i>J</i> = 325.8 Hz), 21.3.
¹⁹ F NMR (376 MHz, CDCl₃) δ -78.9.

A similar compound **14a** is commercially available.

2,4'-dimethyl-5-nitro-1,1'-biphenyl (15a)



2
94%, orange oil
¹ H NMR (400 MHz, CDCl ₃) δ 8.12 – 8.05 (m, 2H), 7.44 – 7.36 (m, 1H), 7.27 (d, <i>J</i> = 8.2 Hz,
2H), 7.23 – 7.18 (m, 2H), 2.43 (s, 3H), 2.37 (s, 3H).
^{13}C NMR (101 MHz, CDCl_3) δ 146.3, 143.6, 143.1, 137.7, 136.7, 131.1, 129.2, 128.8, 124.7,
121.9, 21.2, 20.9.
(M) ⁺⁻ : calc. 227.0941, found 227.0945

Synthesis of 16a, 17a and 23a



The corresponding phenol (2 mmol) and triethylamine (6 mmol, 835 μ l) were dissolved in DCM (3 ml) in a 10 ml reaction vial and cooled down to 0 °C. Sulfonyl chloride (2 mmol) was dissolved in DCM (3 ml) in another vial and slowly added (ca. 5 min) to the reaction mixture in the first vial. After addition, the ice bath was removed, and the reaction mixture was stirred overnight (16 h) at rt. After the reaction completion, the reaction mixture was transferred into a separatory funnel, diluted with DCM (20 ml), and quenched with HCl (10 ml; 0.1 M). The organic layer was washed again with a base NaOH (10 ml; 0.1 M) to remove any remaining substrate, and then once again with HCl (10 ml; 0.1 M). The organic fraction was then dried over Na₂SO₄, filtered, and concentrated in vacuo. The products were analyzed by NMR spectroscopy and used without any additional purification.



p-tolyl 2,4,6-trimethylbenzenesulfonate (17a)



Yield	78%, White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.05 (d, <i>J</i> = 8.2 Hz, 2H), 6.96 (s, 2H), 6.86 – 6.81 (m, 2H), 2.55 (s, 6H), 2.32 (s, 3H), 2.29 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 147.3, 143.7, 140.5, 136.8, 131.7, 130.7, 130.1, 122.0, 22.8, 21.1, 20.9.
HR-MS (ESI)	(M+H) ⁺ : calc. 291.1049, found 291.1051

4-ethylphenyl 4-methylbenzenesulfonate (23a)



Yield83%, colorless oil¹H NMR¹H NMR (400 MHz, CDCl₃) δ 7.71 (d, J = 8.6 Hz, 2H), 7.30 (d, J = 8.0 Hz, 2H), 7.13 - 7.04 (m,
2H), 6.94 - 6.81 (m, 2H), 2.60 (q, J = 7.6 Hz, 2H), 2.45 (s, 3H), 1.20 (t, J = 7.6 Hz, 3H).¹³C NMR¹³C NMR (101 MHz, CDCl₃) δ 147.6, 145.2, 143.2, 132.6, 129.7, 128.9, 128.6, 122.1, 28.3,
21.7, 15.4.HR-MS (ESI)(M+H)*: calc. 277.0893 , found 277.0896

Synthesis of 18a



N,4-dimethylaniline (2 mmol, 242 mg) and triethylamine (6 mmol, 835 μ l) were dissolved in DCM (3 ml) in a 10 ml reaction vial and cooled down to 0 °C. Tosyl chloride (2 mmol, 380 mg) was dissolved in DCM (3 ml) in another vial and slowly added (ca. 5 min) to the reaction mixture in the first vial. After addition, the ice bath was removed, and the reaction mixture was stirred for 5 h at rt. After the reaction completion, the reaction mixture was transferred into a separatory funnel, diluted with DCM (20 ml), and quenched with HCl (10 ml; 0.1 M). The organic layer was washed again with a base NaOH (10 ml; 0.1 M) to remove any remaining tosyl chloride, and then once again with HCl (10 ml; 0.1 M). The organic fraction was then dried over Na₂SO₄, filtered, and concentrated in vacuo. The products were analyzed by NMR spectroscopy and used without any additional purification.

N,4-dimethyl-N-(p-tolyl)benzenesulfonamide (18a)



Yield	85%, slightly brownish oil
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.46 – 7.40 (m, 2H), 7.24 (d, <i>J</i> = 8.0 Hz, 2H), 7.09 (d, <i>J</i> = 8.1 Hz,
	2H), 6.99 – 6.93 (m, 2H), 3.13 (s, 3H), 2.42 (s, 3H), 2.33 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 143.5, 139.0, 137.3, 133.7, 129.5, 129.3, 128.0, 126.6, 38.2,
	21.6, 21.1.
HR-MS (ESI)	(M+H) ⁺ : calc. 276.1053, found 276.1052

Synthesis of 27a



The carboxylic acid **27aa** (1.68 g, 10.00 mmol, 1.0 eq.) was dissolved in methanol (50 mL) inside a 250 mL round-bottom flask and five drops of conc. H_2SO_4 were added as an acid catalyst. The reaction mixture was stirred overnight (~16 h, r.t.) and after the completion of the reaction, the methanol was removed by a rotary evaporator. The remaining oil was dissolved in ethyl acetate (20 mL) and washed twice with saturated brine (20 mL each). The organic phase was dried over Na_2SO_4 and after filtrating the drying agent off, the ethyl acetate was removed *in vacuo to obtain* a pure product **27a**.

N-(4-(trifluoromethoxy)benzyl)acetamide (27a)



Yield	81%, colourless oil
¹ H NMR	¹ H NMR (400 MHz, CDCl₃) δ 7.20 – 7.09 (m, 2H), 6.96 (t, <i>J</i> = 8.7 Hz, 2H), 3.66 (s, 3H), 2.92
	(t, J = 7.7 Hz, 2H), 2.61 (t, J = 7.7 Hz, 2H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 173.2, 161.5 (d, <i>J</i> = 244.0 Hz), 136.2 (d, <i>J</i> = 3.2 Hz), 129.7 (d, <i>J</i>
	= 7.8 Hz), 115.3 (d, J = 21.2 Hz), 51.7, 35.8, 30.1.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl ₃) δ -117.5.
HR-MS (EI)	(M) ⁺⁻ : calc. 182.0738, found 182.0741

Characterization of the products of photochemical reactions 4.2

N-(4-fluorobenzyl)acetamide (1b)⁶

	F 1b
Yield	81% (54 mg, 0.32 mmol), White solid
¹ H NMR	^{1}H NMR (400 MHz, CDCl_3) δ 7.25 – 7.13 (m, 2H), 7.07 – 6.89 (m, 2H), 6.33 (s, 1H), 4.32 (d, J
	= 5.8 Hz, 2H), 1.96 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.2, 162.1 (d, <i>J</i> = 245.5 Hz), 134.2 (d, <i>J</i> = 3.2 Hz), 129.4 (d, <i>J</i>
	= 8.1 Hz), 115.5 (d, <i>J</i> = 21.5 Hz), 42.9, 23.1.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl₃) δ -115.7.

NHAc

NHAc

N-(4-chlorobenzyl)acetamide (2b)⁶

	Cl 2b
Yield	78% (57 mg, 0.31 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.25 (d, <i>J</i> = 8.5 Hz, 2H), 7.15 (d, <i>J</i> = 8.4 Hz, 2H), 6.37 (s, 1H), 4.31 (d, <i>J</i> = 5.9 Hz, 2H), 1.96 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.3, 136.9, 133.2, 129.1, 128.8, 42.9, 23.1.

CI

N-(3-chlorobenzyl)acetamide (3b)⁷



Yield	43% (31 mg, 0.17 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.26 – 7.20 (m, 3H), 7.17 – 7.10 (m, 1H), 6.19 (s, 1H), 4.36 (d, <i>J</i>
	= 5.9 Hz, 2H), 2.00 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.2, 140.4, 134.5, 130.0, 127.8, 127.6, 125.9, 43.1, 23.2.

N-(4-bromobenzyl)acetamide (4b)⁶



Yield	73% (67 mg, 0.29 mmol). White solid
	¹ H NMR (400 MHz, CDCl ₃) δ 7.41 (d, J = 8.5 Hz, 2H), 7.11 (d, J = 8.5 Hz, 2H), 6.26 (s, 1H),
	4.31 (d, <i>J</i> = 5.9 Hz, 2H), 1.97 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.2, 137.4, 131.7, 129.5, 121.3, 43.0, 23.2.

N-benzylacetamide (5b)⁶



 Yield
 50% (30 mg, 0.20 mmol), White solid

 ¹H NMR
 ¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.17 (m, 5H), 6.04 (s, 1H), 4.39 (d, J = 5.7 Hz, 2H), 1.99 (s, 3H).

 ¹³C NMR
 ¹³C NMR (101 MHz, CDCl₃) δ 170.1, 138.3, 128.7, 127.9, 127.5, 43.7, 23.2.

*N-(4-(trifluoromethoxy)benzyl)acetamide (7b)*²



Yield	50% (47 mg, 0.20 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.28 (d, <i>J</i> = 8.7 Hz, 2H), 7.15 (d, <i>J</i> = 8.0 Hz, 2H), 6.14 (s, 1H),
	4.39 (d, <i>J</i> = 5.9 Hz, 2H), 1.99 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.2, 148.5 (d, <i>J</i> = 1.8 Hz), 137.2, 129.2, 121.7 (q, <i>J</i> = 258.6
	Hz), 121.2, 42.9, 23.2.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl ₃) δ -58.4.

methyl 4-(acetamidomethyl)benzoate (8b)



Yield	29% (24 mg, 0.12 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.96 (d, 2H), 7.31 (d, <i>J</i> = 8.5 Hz, 2H), 6.12 (s, 1H), 4.45 (d, <i>J</i> =
	5.9 Hz, 2H), 3.89 (s, 3H), 2.03 (s, 3H).
¹³ C NMR	^{13}C NMR (101 MHz, CDCl_3) δ 170.17, 166.9, 143.6, 123.0, 129.3, 127.5, 52.2, 43.3, 23.2.
HR-MS (EI)	(M) ⁺⁻ : calc. 207.0890, found 207.0891

methyl 3-(acetamidomethyl)benzoate (9b)



Yield	40% (33 mg, 0.16 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.97 – 7.84 (m, 2H), 7.49 – 7.42 (m, 1H), 7.37 (t, <i>J</i> = 7.9 Hz,
	1H), 6.16 (s, 1H), 4.44 (d, J = 5.9 Hz, 2H), 3.88 (s, 3H), 2.01 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.2, 166.9, 138.8, 132.4, 130.5, 128.8, 128.7, 52.2, 43.3,
	23.2.
HR-MS (EI)	(M) ⁺ : calc. 207.0890, found 207.0887

4-(acetamidomethyl)phenyl 2,3,4,5,6-pentafluorobenzoate (10b)



Yield	61% (88 mg, 0.24 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.33 (d, $J = 8.5$ Hz, 2H), 7.16 (d, $J = 8.7$ Hz, 2H), 6.37 (s, 1H), 4.40 (d, $J = 5.7$ Hz, 2H), 2.00 (s, 3H)
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃): (except for C ₆ F ₅) δ 170.4, 157.6, 149.2, 137.1, 129.2, 121.5,
10	43.0, 23.1.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl ₃) δ -137.6 – -137.8 (m), -147.5 (tt, <i>J</i> = 21.3, 5.4 Hz), -160.2 – - 160.4 (m).
HR-MS (ESI)	(M+H) ⁺ : calc. 360.0654 , found 360.0656

4-(acetamidomethyl)phenyl 3,5-bis(trifluoromethyl)benzoate (11b)



Yield	57% (92 mg, 0.23 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 8.61 (s, 2H), 8.13 (s, 1H), 7.35 (d, <i>J</i> = 8.5 Hz, 2H), 7.22 – 7.12
	(m, 2H), 6.45 (s, 1H), 4.42 (d, <i>J</i> = 5.8 Hz, 2H), 2.01 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.4, 162.7, 149.6, 136.9, 132.5 (q, <i>J</i> = 34.2 Hz), 131.7,
	130.3 (d, J = 3.1 Hz), 129.2, 127.0 (p, J = 3.6 Hz), 122.8 (q, J = 272.9 Hz), 121.6, 43.0, 23.1.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl ₃) δ -63.5.
HR-MS (ESI)	(M+H) ⁺ : calc. 406.0872 , found 406.0876



12b

12b was not obtained according to ¹H NMR. SM remained unreacted.



Yield	48% (35 mg, 0.1 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl₃) δ 8.06 (d, <i>J</i> = 8.4 Hz, 2H), 7.92 – 7.72 (m, 2H), 7.65 – 7.53 (m,
	2H), 7.42 (d, J = 8.2 Hz, 2H), 6.06 (s, 1H), 4.49 (d, J = 5.9 Hz, 2H), 2.05 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 149.1, 139.9, 137.5, 131.4, 128.6, 128.3, 127.8, 119.9 (q, <i>J</i> =
	325.8 Hz), 43.2, 23.3.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl ₃) δ -78.9.
HR-MS (ESI)	(M+H) ⁺ : calc. 358.0719 , found 358.0723
Experiment was perfo	ormed on a 0.2 mmol scale (only one reaction vial).

N-((2'-cyano-[1,1'-biphenyl]-4-yl)methyl)acetamide (14b)



Yield	81% (81 mg, 0.32 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.73 (dd, J = 7.8, 1.4 Hz, 1H), 7.62 (td, J = 7.7, 1.4 Hz, 1H), 7.50
	– 7.39 (m, 4H), 7.39 – 7.33 (m, 2H), 6.47 (s, 1H), 4.43 (d, J = 5.7 Hz, 2H), 1.99 (s, 3H).
¹³ C NMR	^{13}C NMR (101 MHz, CDCl_3) δ 170.4, 145.1, 139.1, 137.3, 133.7, 133.0, 130.1, 129.0, 128.2,
	127.7, 118.8, 111.1, 43.3, 23.1.
HR-MS (EI)	(M) ⁺⁻ : calc. 250.1101, found 250.1098

N-((2'-methyl-5'-nitro-[1,1'-biphenyl]-4-yl)methyl)acetamide (15b)



Yield	37% (42 mg, 0.15 mmol), White solid
¹ H NMR	1 H NMR (400 MHz, CDCl ₃) δ 8.15 – 7.97 (m, 2H), 7.45 – 7.33 (m, 3H), 7.28 – 7.21 (m, 2H),
	6.14 (s, 1H), 4.48 (d, J = 5.9 Hz, 2H), 2.33 (s, 3H), 2.05 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.2, 146.2, 143.6, 142.7, 138.8, 138.2, 131.2, 129.3, 127.9,
	124.6, 122.2, 43.4, 23.3, 20.8.
HR-MS (EI)	(M) ⁺ : calc. 284.1155, found 284.1157

4-(acetamidomethyl)phenyl 4-methylbenzenesulfonate (16b)²



Yield37% (47 mg, 0.15 mmol), White solid¹H NMR¹H NMR (400 MHz, CDCl₃) δ 7.69 (d, J = 8.4 Hz, 2H), 7.31 (d, J = 7.7 Hz, 2H), 7.18 (d, J = 8.7
Hz, 2H), 6.92 (d, J = 8.6 Hz, 2H), 5.92 (s, 1H), 4.37 (d, J = 5.9 Hz, 2H), 2.45 (s, 3H), 2.00 (s, 3H).¹³C NMR¹³C NMR (101 MHz, CDCl₃) δ 170.0, 148.8, 145.5, 137.5, 132.4, 129.9, 129.0, 128.5, 122.6, 42.9, 23.3, 21.8.

4-(acetamidomethyl)phenyl 2,4,6-trimethylbenzenesulfonate (17b - major)



Yield	38% (51 mg, 0.15 mmol), slightly brownish oil
¹ H NMR (major)	¹ H NMR (400 MHz, CDCl ₃) δ 7.15 (d, <i>J</i> = 8.6 Hz, 2H), 6.96 (s, 2H), 6.89 (d, <i>J</i> = 8.6 Hz, 2H),
	6.15 (s, 1H), 4.34 (d, J = 5.9 Hz, 2H), 2.53 (s, 6H), 2.37 – 2.26 (m, 4H), 1.98 (s, 3H).
¹³ C NMR (major)	^{13}C NMR (101 MHz, CDCl_3) δ 170.2, 148.7, 144.0, 140.4, 137.3, 131.9, 128.9, 122.4, 42.9,
	23.1, 22.7, 21.1.
HR-MS (ESI)	(M+H) ⁺ : calc. 348.1264 , found 348.1267 (major), 348.1265 (minor)
	(the isomers separate well in LC-QTOF instrument)

See chapter 4.3 for more detailed information about the identification of the minor isomer.

N-(4-((N,4-dimethylphenyl)sulfonamido)benzyl)acetamide (18b)

	Tos
	N ~ 18b
Yield	59% (78 mg, 0.23 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.39 (d, J = 8.3 Hz, 2H), 7.22 (d, J = 8.0 Hz, 2H), 7.17 (d, J = 8.4
	Hz, 2H), 7.01 (d, J = 8.3 Hz, 2H), 6.33 (s, 1H), 4.36 (d, J = 5.8 Hz, 2H), 3.09 (s, 3H), 2.39 (s,
	3H), 1.98 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 170.3, 143.8, 140.8, 137.7, 133.5, 129.5, 128.2, 127.9, 126.8,
	43.0, 38.1, 23.2, 21.6.
HR-MS (ESI)	(M+H) ⁺ : calc. 333.1267 , found 333.1268

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Yield	72% (52 mg, 0.29 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.26 (dd, <i>J</i> = 8.7, 5.2 Hz, 2H), 6.99 (t, <i>J</i> = 8.7 Hz, 2H), 6.19 (s,
	1H), 5.07 (p, <i>J</i> = 7.1 Hz, 1H), 1.97 (s, 3H), 1.45 (d, <i>J</i> = 6.9 Hz, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 169.4, 162.0 (d, J = 245.6 Hz), 139.0 (d, J = 3.2 Hz), 127.8 (d, J
	= 8.0 Hz), 115.4 (d, <i>J</i> = 21.4 Hz), 48.3, 23.2, 21.8.
¹⁹ F NMR	¹⁹ F NMR (377 MHz, CDCl₃) δ -115.9.

N-(1-(4-chlorophenyl)ethyl)acetamide (20b)⁸



77% (61 mg, 0.31 mmol), White solid
¹ H NMR (400 MHz, CDCl ₃) δ 7.30 – 7.24 (m, 2H), 7.24 – 7.19 (m, 2H), 6.26 (d, J = 7.7 Hz,
1H), 5.04 (p, <i>J</i> = 7.1 Hz, 1H), 1.95 (s, 3H), 1.43 (d, <i>J</i> = 7.0 Hz, 3H).
¹³ C NMR (101 MHz, CDCl ₃) δ 169.4, 142.0, 132.9, 128.7, 127.6, 48.2, 23.3, 21.8.

N-(1-(4-bromophenyl)ethyl)acetamide (21b)⁸



Yield	57% (55 mg, 0.23 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl₃) δ 7.50 – 7.39 (m, 2H), 7.22 – 7.14 (m, 2H), 5.75 (s, 1H), 5.07 (p, J
	= 7.1 Hz, 1H), 1.98 (s, 3H), 1.46 (d, J = 6.9 Hz, 3H).
¹³ C NMR	^{13}C NMR (101 MHz, CDCl_3) δ 169.2, 142.3, 131.8, 128.0, 121.2, 48.3, 23.4, 21.7.

*N-(1-phenylethyl)acetamide (22b)*²



Yield	58% (38 mg, 023 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.37 – 7.22 (m, 5H), 5.94 (s, 1H), 5.11 (p, <i>J</i> = 7.0 Hz, 1H), 1.96
	(s, 3H), 1.47 (d, <i>J</i> = 6.9 Hz, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 169.2, 143.2, 128.7, 127.4, 126.2, 48.8, 23.4, 21.8.

4-(1-acetamidoethyl)phenyl 4-methylbenzenesulfonate (23b)



Yield	87% (116 mg, 0.35 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.68 (dd, J = 8.3, 1.4 Hz, 2H), 7.30 (d, J = 8.3 Hz, 2H), 7.20 (d, J
	= 8.5 Hz, 2H), 6.89 (dd, J = 8.6, 2.0 Hz, 2H), 6.21 (s, 1H), 5.04 (p, J = 7.1 Hz, 1H), 2.43 (s,
	3H), 1.93 (d, <i>J</i> = 2.8 Hz, 3H), 1.40 (dd, <i>J</i> = 6.9, 2.2 Hz, 3H).
¹³ C NMR	^{13}C NMR (101 MHz, CDCl_3) δ 169.5, 148.6, 145.5, 142.4, 132.4, 129.9, 128.4, 127.5, 122.4,
	48.1, 23.2, 21.8, 21.7.
HR-MS (ESI)	(M+H) ⁺ : calc. 334.1108, found 334.1106

N-(1-(4-cyanophenyl)ethyl)acetamide (24b)⁹



Yield	60% (45 mg, 0.24 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.61 (d, <i>J</i> = 8.4 Hz, 2H), 7.41 (d, <i>J</i> = 8.1 Hz, 2H), 5.91 (s, 1H),
	5.11 (p, J = 7.1 Hz, 1H), 2.00 (s, 3H), 1.47 (d, J = 7.0 Hz, 3H).
¹³ C NMR	^{13}C NMR (101 MHz, CDCl ₃) δ 169.4, 148.9, 132.5, 126.9, 118.8, 111.1, 48.7, 23.3, 21.8.

ethyl 4-(1-acetamidoethyl)benzoate (25b)



Yield	69% (65 mg, 0.28 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.95 (d, <i>J</i> = 8.4 Hz, 2H), 7.33 (d, <i>J</i> = 8.2 Hz, 2H), 6.47 (d, <i>J</i> = 7.5
	Hz, 1H), 5.10 (p, J = 7.1 Hz, 1H), 4.33 (q, J = 7.1 Hz, 2H), 1.96 (s, 3H), 1.43 (d, J = 7.0 Hz,
	3H), 1.35 (t, <i>J</i> = 7.1 Hz, 4H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 169.5, 166.4, 148.6, 129.9, 129.4, 126.1, 70.0, 48.7, 23.2,
	21.8, 14.3.
HR-MS (EI)	(M) ⁺ : calc. 235.1203, found 235.1202

N-(1-(9,10-dioxo-9,10-dihydroanthracen-2-yl)ethyl)acetamide (26b)



Yield	62% (73 mg, 0.25 mmol), lightly yellowish solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 8.31 – 8.27 (m, 2H), 8.25 (d, J = 8.0 Hz, 1H), 8.21 (d, J = 1.6 Hz,
	1H), 7.87 – 7.75 (m, 2H), 7.73 (dd, J = 8.0, 1.7 Hz, 1H), 6.06 (d, J = 6.6 Hz, 1H), 5.23 (p, J =
	7.1 Hz, 1H), 2.06 (s, 3H), 1.55 (d, <i>J</i> = 7.0 Hz, 3H).
¹³ C NMR	^{13}C NMR (101 MHz, CDCl_3) δ 183.2, 182.8, 169.6, 150.4, 134.2, 134.1, 133.8, 133.5, 132.6,
	132.4, 127.9, 127.3, 124.1, 49.0, 23.4, 22.1.
HR-MS (EI)	(M) ⁺⁻ : calc. 293.1046, found 293.1041

methyl 3-acetamido-3-(4-fluorophenyl)propanoate (27b)



Yield	64% (61 mg, 0.26 mmol), White solid
¹ H NMR	1 H NMR (400 MHz, CDCl ₃) δ 7.29 – 7.18 (m, 2H), 7.01 – 6.87 (m, 3H), 5.43 – 5.29 (m, 1H),
	3.58 (s, 3H), 2.86 (dd, J = 15.8, 6.2 Hz, 1H), 2.76 (dd, J = 15.7, 6.2 Hz, 1H), 1.95 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 171.5, 169.5, 162.1 (d, J = 246.0 Hz), 136.5 (d, J = 3.2 Hz),
	128.0 (d, J = 8.1 Hz), 115.5 (d, J = 21.5 Hz), 51.9, 49.1, 39.9, 23.2.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl ₃) δ -115.4.
HR-MS (EI)	(M) ⁺⁻ : calc. 239.0952, found 239.0947

N-(bis(4-fluorophenyl)methyl)acetamide (28b)⁸



Yield	91% (95 mg, 0.36 mmol), White solid
¹ H NMR	¹ H NMR (400 MHz, CDCl ₃) δ 7.18 – 7.08 (m, 4H), 7.03 – 6.93 (m, 4H), 6.60 (d, J = 7.7 Hz,
	1H), 6.14 (d, <i>J</i> = 8.0 Hz, 1H), 1.96 (s, 3H).
¹³ C NMR	¹³ C NMR (101 MHz, CDCl ₃) δ 169.4, 162.1 (d, <i>J</i> = 246.5 Hz), 137.2 (d, <i>J</i> = 3.2 Hz), 129.1 (d, <i>J</i>
	= 8.1 Hz), 115.6 (d, <i>J</i> = 21.5 Hz), 55.7, 23.1.
¹⁹ F NMR	¹⁹ F NMR (376 MHz, CDCl₃) δ -115.3.

N-benzhydrylacetamide (29b)⁶



29b

 Yield
 62% (56 mg, 0.25 mmol), White solid

 ¹H NMR
 ¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.13 (m, 10H), 6.59 (d, J = 7.6 Hz, 1H), 6.23 (d, J = 8.1 Hz, 1H), 1.98 (s, 3H).

 ¹³C NMR
 ¹³C NMR (101 MHz, CDCl₃) δ 169.4, 141.6, 128.7, 127.6, 127.5, 57.1, 23.2.

4.3 Characterization of the minor isomer in reaction mixture 17b

The isomers cannot be separated on a silica TLC plate. The identification of the structure of the minor isomer was done by the ¹H NMR (**Figure S3** and **Figure S4**). The ortho isomer **17b'** shows 3 different benzylic signals, while isomer **17b''** is more symmetric and should show only 2 different benzylic protons. The ratio of integrals of the aliphatic NMR signals (1:1:1:1) additionally supports the structure of **17b'** as a minor isomer in the product mixture.



Figure S3: Identification of the minor isomer in the product mixture 17b – aliphatic region.

The assignments of the signals were done with the help of the ChemDraw NMR predictor. The only signal which is significantly different from the predicted value is signal of acetyl group of **17b'** at 1.72 ppm. The reason for the shift could be the proximity of magnetic field inducing π -systems.

The aromatic region of the ¹H NMR (**Figure S4**) shows quite complex pattern of signals which would fit better to the structure **17b'** than the symmetric molecule **17b''**.



Figure S4: Identification of the minor isomer in the product mixture 17b – aromatic region.

5 Copies of NMR spectra



Figure S5: ¹H NMR spectrum of compound **1i** (400 MHz in CDCl₃).



Figure S6: ¹³C NMR spectrum of compound 1i (101 MHz in CDCl₃).



Figure S7: ¹H NMR spectrum of compound **10a** (400 MHz in CDCl₃).



Figure S8: ¹³C NMR spectrum of compound **10a** (101 MHz in CDCl₃).

Mar09-2022.322.fid Narobe, RN-650-d,cdcl3 rau_sF19CPD CDCl3 {C:\Bruker\TopSpin3.0} AK_Koenig 57

137,83 137,84 137,84 137,89 147,89 147,89 147,89 148,1914,19 148,19 148,19 148,19 148,19 148,1914,19 148,19 148,19 148,1914,19 14



40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 f1 (ppm)

Figure S9: ¹⁹F NMR spectrum of compound **10a** (377 MHz in CDCl₃).



Figure S10: ¹H NMR spectrum of compound 11a (400 MHz in CDCl₃).



Figure S11: ¹³C NMR spectrum of compound 11a (101 MHz in CDCl₃).





40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 f1 (ppm)

Figure S12: ¹⁹F NMR spectrum of compound **11a** (377 MHz in CDCl₃).



Figure S14: $^{\rm 13}{\rm C}$ NMR spectrum of compound 13a (101 MHz in CDCl₃).



Figure S15: $^{19}\mathsf{F}$ NMR spectrum of compound 13a (377 MHz in CDCl₃).



Figure S16: ¹H NMR spectrum of compound 15a (400 MHz in CDCl₃).



Figure S17: ¹³C NMR spectrum of compound 15a (101 MHz in CDCl₃).



Figure S18: ¹H NMR spectrum of compound 16a (400 MHz in CDCl₃).



Figure S19: ¹³C NMR spectrum of compound 16a (101 MHz in CDCl₃).


Figure S20: ¹H NMR spectrum of compound **17a** (400 MHz in CDCl₃).



Figure S21: ¹³C NMR spectrum of compound **17a** (101 MHz in CDCl₃).



Figure S22: ¹H NMR spectrum of compound 18a (400 MHz in CDCl₃).



Figure S23: ¹³C NMR spectrum of compound 18a (101 MHz in CDCl₃).



Figure S24: ¹H NMR spectrum of compound 23a (400 MHz in CDCl₃).



Figure S25: ¹³C NMR spectrum of compound 23a (101 MHz in CDCl₃).



Figure S26: ¹H NMR spectrum of compound 27a (400 MHz in CDCl₃).



Figure S27: ¹³C NMR spectrum of compound 27a (101 MHz in CDCl₃).



27a

40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 fl (ppm)

Figure S28: 19 F NMR spectrum of compound 27a (377 MHz in CDCl₃).



Figure S29: ¹H NMR spectrum of compound 1b (400 MHz in CDCl₃).



Figure S30: ¹³C NMR spectrum of compound **1b** (101 MHz in CDCl₃).



40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 f1 (ppm)

Figure S31: ¹⁹F NMR spectrum of compound **1b** (377 MHz in CDCl₃).



Figure S32: ¹H NMR spectrum of compound 2b (400 MHz in CDCl₃).



Figure S33: ¹³C NMR spectrum of compound **2b** (101 MHz in CDCl₃).



Figure S34: ¹H NMR spectrum of compound 3b (400 MHz in CDCl₃).



Figure S35: ¹³C NMR spectrum of compound **3b** (101 MHz in CDCl₃).



Figure S36: ¹H NMR spectrum of compound 4b (400 MHz in CDCl₃).



Figure S37: ¹³C NMR spectrum of compound 4b (101 MHz in CDCl₃).



Figure S38: ¹H NMR spectrum of compound 5b (400 MHz in CDCl₃).



Figure S39: ¹³C NMR spectrum of compound **5b** (101 MHz in CDCl₃).



Figure S40: ¹H NMR spectrum of compound 6b (400 MHz in CDCl₃).



Figure S41: ¹³C NMR spectrum of compound **6b** (101 MHz in CDCl₃).



Figure S42: ¹H NMR spectrum of compound 7b (400 MHz in CDCl₃).



Figure S43: ¹³C NMR spectrum of compound 7b (101 MHz in CDCl₃).



40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 fl (ppm)

Figure S44: ¹⁹F NMR spectrum of compound **7b** (377 MHz in CDCl₃).



Figure S45: ¹H NMR spectrum of compound 8b (400 MHz in CDCl₃).



Figure S46: ¹³C NMR spectrum of compound 8b (101 MHz in CDCl₃).



Figure S47: ¹H NMR spectrum of compound 9b (400 MHz in CDCl₃).



Figure S48: ¹³C NMR spectrum of compound 9b (101 MHz in CDCl₃).



Figure S49: ¹H NMR spectrum of compound 10b (400 MHz in CDCl₃).



Figure S50: ¹³C NMR spectrum of compound **10b** (101 MHz in CDCl₃).



40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 fl (ppm)

Figure S51: ¹⁹F NMR spectrum of compound **10b** (377 MHz in CDCl₃).



Figure S52: ¹H NMR spectrum of compound **11b** (400 MHz in CDCl₃).



Figure S53: ¹³C NMR spectrum of compound **11b** (101 MHz in CDCl₃).



Figure S54: $^{19}\mathsf{F}$ NMR spectrum of compound 11b (377 MHz in CDCl₃).



Figure S55: ¹H NMR spectrum of compound **13b** (400 MHz in CDCl₃).



Figure S56: ¹³C NMR spectrum of compound **13b** (101 MHz in CDCl₃).



40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 f1 (ppm)

Figure S57: ¹⁹F NMR spectrum of compound **13b** (377 MHz in CDCl₃).



Figure S58: ¹H NMR spectrum of compound **14b** (400 MHz in CDCl₃).



Figure S59: ¹³C NMR spectrum of compound **14b** (101 MHz in CDCl₃).



Figure S60: ¹H NMR spectrum of compound 15b (400 MHz in CDCl₃).



Figure S61: ¹³C NMR spectrum of compound 15b (101 MHz in CDCl₃).



Figure S62: ¹H NMR spectrum of compound 16b (400 MHz in CDCl₃).



Figure S63: ¹³C NMR spectrum of compound 16b (101 MHz in CDCl₃).



Figure S64: ¹H NMR spectrum of compound **17b** (400 MHz in CDCl₃).



Figure S65: ¹³C NMR spectrum of compound **17b** (101 MHz in CDCl₃).



Figure S66: ¹H NMR spectrum of compound **18b** (400 MHz in CDCl₃).



Figure S67: ¹³C NMR spectrum of compound 18b (101 MHz in CDCl₃).



Figure S69: ¹³C NMR spectrum of compound **19b** (101 MHz in CDCl₃).



Figure S70: 19 F NMR spectrum of compound **19b** (377 MHz in CDCl₃).



Figure S71: ¹H NMR spectrum of compound 20b (400 MHz in CDCl₃).



Figure S72: ¹³C NMR spectrum of compound **20b** (101 MHz in CDCl₃).



Figure S73: ¹H NMR spectrum of compound **21b** (400 MHz in CDCl₃).



Figure S74: ¹³C NMR spectrum of compound **21b** (101 MHz in CDCl₃).



Figure S75: ¹H NMR spectrum of compound 22b (400 MHz in CDCl₃).



Figure S76: ¹³C NMR spectrum of compound **22b** (101 MHz in CDCl₃).



Figure S77: ¹H NMR spectrum of compound 23b (400 MHz in CDCl₃).



Figure S78: ¹³C NMR spectrum of compound 23b (101 MHz in CDCl₃).



Figure S79: ¹H NMR spectrum of compound 24b (400 MHz in CDCl₃).



Figure S80: ¹³C NMR spectrum of compound 24b (101 MHz in CDCl₃).



Figure S81: ¹H NMR spectrum of compound 25b (400 MHz in CDCl₃).



Figure S82: ¹³C NMR spectrum of compound 25b (101 MHz in CDCl₃).



Figure S83: ¹H NMR spectrum of compound 26b (400 MHz in CDCl₃).



Figure S84: ¹³C NMR spectrum of compound **26b** (101 MHz in CDCl₃).


Figure S85: ¹H NMR spectrum of compound 27b (400 MHz in CDCl₃).



Figure S86: ¹³C NMR spectrum of compound 27b (101 MHz in CDCl₃).



Figure S87: ¹⁹F NMR spectrum of compound **27b** (377 MHz in CDCl₃).



Figure S88: ¹H NMR spectrum of compound 28b (400 MHz in CDCl₃).



Figure S89: ¹³C NMR spectrum of compound 28b (101 MHz in CDCl₃).



40 30 20 10 0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 f1 (ppm)

Figure S90: ¹⁹F NMR spectrum of compound **28b** (377 MHz in CDCl₃).



Figure S91: ¹H NMR spectrum of compound 29b (400 MHz in CDCl₃).



Figure S92: ¹³C NMR spectrum of compound **29b** (101 MHz in CDCl₃).

6 Literature

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