## 1 Dual-channel Lanthanide-doped Nanoprobe for Reliable Multi-signal Ratiometric Detection

- 2 of H<sub>2</sub>S in Whole Blood
- 3
- 4 Method

5 Materials & Characterization. All lanthanide oxides were purchased from STREM Chemicals, Inc. USA. Lanthanide chlorides were prepared by dissolving the corresponding metal oxide in HCl 6 7 solution at elevated temperature and then evaporating the water completely under reduced pressure. 8 All other chemicals were purchased from Alfa Aesar Chemical Co. Ltd. All chemical reagents were 9 of analytical grade and were used directly without further purification. Deionized water was used 10 throughout. The nanoprobe morphology and element distribution were determined by a Tecnai 11 G<sup>2</sup>F30 high-resolution transmission electron microscopy. The powder X-ray diffraction pattern was measured with a Brucker D8 advance X-ray diffractometer from 10° to 70° (Cu K $\alpha$  radiation,  $\lambda =$ 12 13 1.54 Å). Dynamic light scattering (DLS) and zeta potential experiments were carried out on an 14 ALV-5000 spectrometer goniometry equipped with an ALV/LSE-5004 light scattering electronic and multiple tau digital correlator and a JDS Uniphase He-Ne laser (632.8 nm) with an output power 15 of 22 mW. The hydrodiameter distribution was measured at 25 °C with a detection angle of 90°. 16 17 Inductively coupled mass spectroscopy (ICP-MS) analysis was performed on Agilent 7500ce ICP-18 MS. UV-vis-NIR absorption spectra are obtained on a Shimadzu UV-3600 UV-vis-NIR 19 spectrophotometer.

Preparation of Ln-CuMOF. The NaErF4@NaLuF4 nanophosphors were prepared and modified 20 with polyvinylpyrrolidone according to the previous reports 1, 2. In a typical experiment, a mixture 21 of 1 mmol ErCl<sub>3</sub>, 15 mL OA, and 15 mL ODE are added into a 100 mL three-necked flask. Under 22 23 the vacuum, the mixture is heated to 160 °C to form a clear solution, and then cooled to room 24 temperature. After the solution cooling down, 0.025 mmol NaOH (0.1 g) and 0.04 mmol NH<sub>4</sub>F (0.1481 g) are added into the flask directly and stirred for 30 minutes. The solution is slowly heated 25 26 with gently stirred, degassed at 100 °C, and then heated to 300 °C and maintained for 1 hour under 27 the Argon atmosphere. After the solution is cooled naturally, the NaErF<sub>4</sub> nanoparticles are separated via centrifugation (10000 rpm) and washed with ethanol/cyclohexane (1:1 v/v) three times. The 28 29 NaLuF<sub>4</sub> shell was coated by the same procedure by replacing the lanthanide chloride with LuCl<sub>3</sub>. The as-obtained NaErF<sub>4</sub>@NaLuF<sub>4</sub> were redispersed in 10 mL chloroform (0.5 mg mL<sup>-1</sup>). A solution 30 of polyvinylpyrrolidone (Mw = 30,000) in 15 mL chloroform (0.1 g mL<sup>-1</sup>) was added into the 31 NaErF<sub>4</sub>@NaLuF<sub>4</sub> suspension and the mixture was stirred at 40°C for 24 hours. The 32 polyvinylpyrrolidone-modified NaErF<sub>4</sub>@NaLuF<sub>4</sub> were collected by centrifugation at 9,000 rpm for 33 34 10 min. The polyvinylpyrrolidone-modified NaErF<sub>4</sub>@NaLuF<sub>4</sub> was cleaned with ethanol three times. 35 Then, 10 mg polyvinylpyrrolidone-modified NaErF<sub>4</sub>@NaLuF<sub>4</sub> was dispersed in 15 mL CuCl<sub>2</sub> solution (1.5 mM) and ultrasonically treated for 5 min. The dispersion was added to a 15 mL 36 dimethylformamide solution of benzene-1,3,5-tricarboxylate (6 mM). The solution was degassed, 37 slowly heated to 100 °C with gently stirring, and maintained for one hour under the argon 38 atmosphere. After the solution was cooled naturally, the Ln-CuMOF were separated by 39 40 centrifugation (12000 rpm) and washed with ethanol and water three times, respectively. The Ln-41 CuMOF was stored in water at 4 °C.

42  $H_2S$  detection. The preparation and quantification of standard  $H_2S$  solution followed the previous

43 reports <sup>3</sup>. Due to the neutral nature of serum, the  $H_2S$  would not only dissolve but ionize to be  $S^{2-}$ 

44 and HS<sup>-</sup>. To simulate the existence of H<sub>2</sub>S in physiological conditions, H<sub>2</sub>S standard solution was

1 prepared by adjusting the pH of freshly formulated  $Na_2S$  solution to neutral (pH = 7.0) with diluted

 $2 \quad hydrochloric acid solution in medical saline. All H_2S standard solutions were hermetically stored at$ 

3 4 °C and used within 60 min after formulation. The  $H_2S$  standard solutions were pre-quantified by

 $4 \quad using the standard methylene blue colorimetric protocol to ensure the accuracy of H_2S \ concentration$ 

5 for further study. Additional hemin and human serum albumin were added to the final solution to

6 simulate the whole blood. To set up the standard curve for  $H_2S$  detection, 0.5 mL standard  $H_2S$ 

7 solution was mixed with 0.5 mL dispersion of Ln-CuMOF (5 mg mL<sup>-1</sup>) and incubated at 37  $^{\circ}$ C under

gently shaking for 15 min. After that, the temperature and fluorescence signals were collected under
 9 808 nm laser irradiation (1.5 W cm<sup>-2</sup>) with the infrared camera and fluorescence spectroscopy,

10 respectively.

11 Ethical statement. The establishment of acute pancreatitis mice was performed by Beijing Vital

12 River Laboratory Animal Technology Co., Ltd. following the Guidelines for Care and Use of

13 Laboratory Animals and approved by the Animal Ethics Committee of the Vital River Institutional

14 Animal Care and Use Committee.

15 **Serodiagnostic study.** The blood of mice was collected from the femoral vein and sealed with the 16 addition of heparin before tests. Serum samples were collected by separating the blood by

17 centrifugation and used for comparison. Twenty AP mice and twenty normal mice were used for

18 receiver-operating characteristic analysis. For the quantification of  $H_2S$  in biological fluid samples,

19 0.5 mL standard sample was mixed with 0.5 mL dispersion of Ln-CuMOF (5 mg mL<sup>-1</sup>) and

20 incubated at 37 °C under gently shaking for 15 min. After that, the temperature and fluorescence

21 signals were collected under 808 nm laser irradiation (1.5 W cm<sup>-2</sup>) with the infrared camera and

22 fluorescence spectroscopy, respectively. The receiver-operating characteristic analysis was

23 performed according to previous reports <sup>3</sup>.

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2 Figure S1. XRD patterns of the separated product from the reaction of Ln-CuMOF with H<sub>2</sub>S. The

3 referenced standard card is 06-0464, corresponding to the hexagonal CuS.



2 Figure S2. Relative fluorescence intensity a) and hydrodiameter b) of Ln-CuMOF over a storage

3 period of seven days.



2 Figure S3. UV-vis-NIR spectrum of Ln-CuMOF after reaction with H<sub>2</sub>S.
3



2 Figure S4. The receiver-operating characteristic curve presents the assay's probability of correctly

- 3 distinguishing between normal and AP cases based on the plasma H<sub>2</sub>S concentration in the whole
- 4 blood samples determined by the typical colorimetric method.

**Table R1.** ICP-MS results of Ln-CuMOF and separated product after reaction with  $H_2S$ .

	Cu (ng mL <sup>-1</sup> )	S (ng mL <sup>-1</sup> )	Cu:S (mol/mol)
Ln-CuMOF	$124.3\pm18.6$	not detected	-
Separated product	$323.6\pm24.3$	$157.4\pm15.1$	1.037