Electronic Supplementary Information

Six-Electron Organic Redoxmers for Aqueous Redox Flow Batteries

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S1. Experimental

S1.1 Materials

Hexaketocyclohexane hydrate (99%), 4-diaminobenzenesulfonic acid, and 4-hydroxy-2nitroaniline (98%) were purchased from AmBeed. 2-[2-(2-chloroethoxy)ethoxy]ethanol (96%), palladium on activated carbon (10% Pd), and all the solvents were purchased from Fisher. These chemicals were used as received.

S1.2 Characterizations

¹H and ¹³C NMR spectra were collected on an AVANCE III HD 400 MHz spectrometer (Bruker, Germany). The radical state of **TPz-1** was determined by continuous wave X band EPR spectroscopy using a Bruker ESP-300 EPR spectrometer.

S1.3 Electrochemical measurement

Cyclic voltammetry (CV) was performed using a CHI760D potentiostat (CH Instruments) with a three-electrode configuration that consisted of a glassy carbon working electrode (3 mm diameter), a carbon felt counter electrode and an Ag/AgCl reference electrode. The samples were 3 mM redoxmer in 1 M NaOH electrolyte. The CV data were obtained at a

series of specified scan rates (Figure S7a and S8a), which were used to calculate the kinetic parameters of redoxmers described in Section S3.

S1.4 Flow cell test

The 5 mM flow cells were assembled with stacked layers of ELAT carbon cloth electrodes (Nuvant) sandwiching a Nafion 212 membrane with an active area of 10 cm^2 . The electrolytes were pumped through the carbon cloth electrodes using a peristaltic pump at a flow rate of 20 mLmin⁻¹. The flow cell tests were conducted at room temperature on a Neware CT-4008 battery tester in an argon filled glove box with the O₂ level less than 1 ppm. The anolyte was 5 mM **TPZ-2** in 1 M NaOH (10 mL) and the catholyte was 0.2 M K₄Fe(CN)₆ in 1 M NaOH (15 mL).

A different cell was used to test the 1 mM **TPz-2** flow cells, consisting of 2.5mm-thick GFD graphite felt electrodes (SGL Carbon, Germany) with an active area of 2.4 cm². The anolyte was 1 mM **TPZ-2** in 1 M NaOH (10 mL) and the catholyte was 0.04 M K₄Fe(CN)₆/0.02 M K₃Fe(CN)₆ (15 mL), both in 1 M NaOH.

S2. Synthesis of Redoxmers

1) Synthesis of TPz-1

Hexaketocyclohexane (2 mmol, 0.37 g) and 3,4-diaminobenzenesulfonic acid (6.6 mmol, 1.24 g) were dissolved in a mixture solvent of glacial acetic acid/ethanol (1:1 v/v, totally 50 mL) in a nitrogen atmosphere. The solution was maintained at 118 °C for 16 hours with vigorous stirring. The resulting dark green precipitate was filtered, washed with acetone and ethanol, and dried in vacuum to yield a dark green solid (0.98 g, 80%). ¹H NMR (400 MHz, D₂O): δ (ppm) 7.83 (s, 3H), 7.63 (d, 3H), 7.58 (d, 3H).

2) Synthesis of TPz-2

a) Synthesis of the modified diaminobenzene



To a solution of 4-hydroxy-2-nitroaniline (25 mmol, 3.85 g) in anhydrous DMF (60 mL) were added K₂CO₃ (62.5 mmol, 8.63 g) and 2-[2-(2-chloroethoxy)ethoxy]ethanol (30 mmol, 5.04 g) under a nitrogen atmosphere. The reaction mixture was stirred under reflux at 130 °C overnight. After cooling down, deionized water (100 mL) and CH₂Cl₂ (100 mL) were added to the reaction. The organic phase was separated and washed with a large amount of deionized water, dried over anhydrous magnesium sulfate, and concentrated under reduced pressure. The crude residue was purified using silica gel column chromatography with ethyl acetate/n-hexane (1:1 v/v) as an eluent to give an oily red compound, 4-tri(ethylene glycol)-2-nitroaniline (6.00 g, 84%). ¹H NMR (400 MHz, CDCl₃): δ (ppm) 7.56~7.57 (d, 1H), 7.10~7.11 (d, 1H), 6.75~6.78 (d, 1H), 5.93 (s, 2H), 4.10~4.13 (t, 2H), 3.85~3.88 (t, 2H), 3.73~3.74 (t, 6H), 3.63 (t, 2H).

The 4-tri(ethylene glycol)-2-nitroaniline (8 mmol, 2.3 g) was dissolved in MeOH (80 mL) and mixed with 10% Pd-C (0.5 g) under a nitrogen atmosphere. A hydrogen balloon was connected to the reaction mixture and vigorous stirring was continued overnight at room temperature. The catalyst was filtered through Celite and the solvent was evaporated to yield a brown solid, 4-tri(ethylene glycol)-1,2-diaminobenzene (1.96 g, 96%), which was used immediately in the next step without further purification due to its air sensitivity.

b) Synthesis of TPz-2

Synthesis of **TPz-2** followed a similar procedure for **TPz-1**. Hexaketocyclohexane (2 mmol, 0.372 g) and 4-tri(ethylene glycol)-1,2-diaminobenzene (6.6 mmol, 1.68 g) were dissolved in a mixture solvent of glacial acetic acid/ethanol (1:1 v/v, totally 50 mL) in a nitrogen atmosphere. The solution was maintained at 118 °C for 16 hours with vigorous stirring. The resulting dark green precipitate was filtered and purified by silica gel column chromatography with CH₂Cl₂/MeOH/Et₃N (100:10:1 v/v/v) as an eluent to give a dark green solid (1.18 g, 71%). ¹H NMR (400 MHz, CDCl₃): δ (ppm) 8.36 (d, 3H), 7.74 (s, 3H), 7.62(d, 3H), 4.40 (t, 6H), 4.03 (t, 6H), 3.83~3.81 (m, 18H), 3.68 (t, 6H). ¹³C NMR (400 MHz, CDCl₃) δ : 161.57, 161.36, 145.19, 145.54, 144.94, 144.89, 143.56, 143.28, 143.09, 142.85, 141.34, 141.18, 141.09, 140.93, 140.72, 140.21, 139.91, 131.55, 131.42, 126.80, 126.75, 126.54, 126.49, 107.40, 72.59, 70.95, 70.44, 69.39, 68.26, 61.76.

3) Synthesis of DHPS



2,5-Dihydroxy-1,4-benzoquinone (1.4 g, 10 mmol) was added to 50 mL water in a roundbottomed flask pre-heated to 100 °C. After stirring for 5 minutes, solid 3,4diaminobenzenesulfonic acid (1.89 g, 10 mmol) was slowly added in a few portions. The reaction mixture was refluxed overnight and then cooled down to room temperature under vigorous stirring. 30 ml of acetone was added to dilute the reaction mixture, yielding a gold solid. The solid was filtered, washed with water and acetone, and dried under vacuum for several days to give the product (2.82 g, 97%). NMR (400 MHz, DMSO): δ (ppm) 8.23 (s, 1H), 8.07(d, 1H), 7.98 (d, 1H), 7.30 (s, 1H), 7.29 (d, 1H).



Figure S1. ¹H NMR spectrum of **TPz-1** in D_2O . The asterisk represents the NMR solvent (H₂O).



Figure S2. ¹H NMR spectrum of 4-tri(ethylene glycol)-2-nitroaniline in CDCl₃. The asterisk represents the NMR solvent (CHCl₃).



Figure S3. ¹H NMR spectrum of 4-tri(ethylene glycol)-1,2-diaminobenzene. The asterisk represents the NMR solvent (CHCl₃).



Figure S4. ¹H NMR spectrum of **TPz-2** in CDCl₃. The asterisk represents the NMR solvent (CHCl₃).



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 Chemical shift (ppm)

Figure S5. ¹³C NMR spectrum of **TPz-2** in CDCl₃. The asterisk represents the NMR solvent (CHCl₃).



Figure S6. ¹H NMR spectrum of **DHPS** in DMSO. The asterisk represents the NMR solvent (DMSO).

S3. Kinetic Parameter Determination

Calculations of the diffusion coefficients (*D*) and charge transfer rate constants (k^0) followed a reported method.¹ Normally, Randles-Sevcik Eq. 1 is used to calculate *D* for a reversible redox reaction. Here, "reversible" is determined by the oxidation-reduction peak separation $\Delta E_p \leq (57/n) \text{ mV.}^2$

$$i_p = 0.4463nFAC(\frac{nFvD}{RT})^{\frac{1}{2}}$$
 (1)

where i_p is the peak current, *n* is the number of transferred electrons, *F* is the Faraday constant (96485 C mol⁻¹), *A* is the surface area of the electrode ($A = 0.0707 \text{ cm}^2$), *C* is the bulk concentration of redoxmers (3 mM), *v* is the scan rate and *T* is the room temperature (298 K). *D* is obtained from the slope of the linearly fitted $i_p - v^{1/2}$ plot, that is $2.69 \times 10^5 n^{3/2} ACD^{1/2}$.

For an irreversible redox reaction (determined by $\Delta E_p > (200/n)$ mV, a correction factor α was introduced to obtain Eq. 2:

$$i_p = 0.4463nFAC\left(\frac{n\alpha FvD}{RT}\right)^{\frac{1}{2}}$$
(2)

Here, α is defined by Eq. 3:

$$\left|E_{p}-E_{p/2}\right| = \left(\frac{48}{\alpha n}\right) mV \tag{3}$$

where E_p and $E_{p/2}$ are the potentials at the peak current and half of the peak current, respectively. For **DHPS**, α =0.51. For **TPz-2**, α =0.52 for first redox peak and α =0.56 for second redox peak. Using Eq. 2, the *D*s for the anodic and cathodic peaks (D_0 and D_R) were calculated (Figure S7b and S8b). *D* is the average value of D_0 and D_R .

Based on the CV curves at different scan rates, the straight relationship of $\ln(i_p)$ and the overpotential $(E_p - E^{0'})$, expressed in Eq. 4, was used to determine $k^{0:1}$

$$\ln(i_p) = \ln(0.227nFAk^0 C^*) - \left(\frac{\alpha F}{RT}\right) (E_P - E^{0'})$$
(4)

Here, $E^{0'}$ is the formal potential and is determined from Eq. 5:³

$$E^{0'} = \frac{\sum_{i=1}^{j} \frac{E_{pa_i} + E_{pc_i}}{2}}{j}$$
(5)

where *j* is the total number of CV scans, E_{pa} and E_{pc} represent the anodic and cathodic peak potentials, respectively. The linearly fitted plot of $\ln(i_p)$ vs $(E_p - E^{0'})$ gives an intercept that is a function of k^0 , as shown in Figure S7c, d and S8c, d.



Figure S7. Kinetic performance of 3 mM **DHPS** in 1 M NaOH: (a) CV curves at different scan rates; (b) linearly fitted $i_p - v^{1/2}$ plot; (c,d) linearly fitted $\ln(i_p) - (E_p - E^0)$ plot for both anodic and cathodic peaks.



Figure S8. Kinetic performance of 3 mM **TPz-2** in 1 M NaOH (first redox peak): (a) CV curves at different scan rates; (b) linearly fitted $i_p - v^{1/2}$ plot; (c,d) linearly fitted $\ln(i_p) - (E_p - E^0)$ plot for both anodic and cathodic peaks.

S4. Supporting Figures for Flow Cells



Figure S9. Rate capability of a 5 mM **DHPS** flow cell under the same conditions as **TPz-2** based flow cells.



Figure S10. Self-discharge open-circuit voltage curve of the fully charged 1 mM **TPz-2** flow cell shown in Figure 1b.

References

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