Supporting Information For:

## Supramolecular Chiral Sensing by Supramolecular Helical Polymers

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## Contents

General ..... S2
Method ..... S3
Scheme S1 Synthesis of 2 ..... S4
Figure $\mathbf{S} 1{ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR spectra of 2 . ..... S5
Figure S2-S4 2D NMR spectra of $\mathbf{2}$. ..... S6-S8
Figure S5. Plot of ${ }^{1} \mathrm{H}$ NMR signal intensities obtained from the diffusion experiments for $\mathbf{2}$ ..... S9
Figure S6. HRMS spectrum of 2 ..... S10
Figure S7. UV/vis spectra of 2 ..... S10
Figure S8. AFM image of 2 ..... S11
Figure S9. VT ECD spectra of 2 ..... S11
Figure S10. ECD spectra of $\mathbf{2}$ in the mixture of $(+)$ - and $(-)$ - $\alpha$-pinene with various ee values. ..... S12
Figure S11. Plot of $\Delta \varepsilon$ values of $\mathbf{2}$ in the mixture of $(+)$ - and $(-)$ - $\alpha$-pinene with various ee values ..... S13
References ..... S14

## General

All solvents were commercial reagent grade and were used without further purification except where noted. Dry 1,2dichloroethane and dry $\mathrm{NEt}_{3}$ was obtained by distillation over $\mathrm{CaH}_{2}$ and KOH , respectively. ${ }^{1} \mathrm{H},{ }^{13} \mathrm{C}$, and 2 D NMR (DQFCOSY, NOESY, and HSQC) spectra were recorded on a JEOL ECA-600 spectrometer, and chemical shifts were reported on the delta scale in ppm relative to residual pyridine ( $\delta=8.74$ and 150.4 for ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$, respectively). Diffusion-ordered ${ }^{1} \mathrm{H}$ NMR spectroscopic (DOSY) measurement was carried out on a JEOL ECA-500 spectrometer. Melting points (M.p.) were measured with a Yanagimoto micro melting point apparatus. Infrared (IR) spectra were recorded on a JASCO FT/IR-4600 spectrometer with ZeSe ATR accessory. Preparative separations were performed by silica gel gravity column chromatography (Silica Gel 60 N (spherical, neutral)). Preparative medium pressure liquid chromatography (MPLC) separations were carried out with a YAMAZEN smart flash EPCLC AL-580S using a preparative Bio-Beads (S-X1) column. Recycling preparative GPC-HPLC separations were carried out on JAI LaboACE LC-5060 using preparative JAIGEL-2.5HH, 2HH columns in series. UV/vis absorption spectra were recorded on a JASCO V-760 spectrometer. Circular dichroism (CD) spectra were recorded on a JASCO J-1500 spectrometer. Morphological evaluation of the samples was performed using atomic force microscopy (AFM) using a Bruker Multimode 8HR under ambient conditions with a Bruker cantilever model NCHV in the tapping mode. NanoScope Analysis 2.0 was used for the AFM image analysis. $\mathbf{1}^{[1]}$ and 2-octyldecanol ${ }^{[2]}$ were synthesized according to reported methods.

## Method

Determination of an affinity constant $\left(K_{a}\right)$ corresponding to the intermolecular interactions between bis (porphyrin) moieties of 2 . ${ }^{[3]}$

A hyperbolic curve was obtained by plotting the concentrations of 2 against the extinction coefficients $(\varepsilon)(\lambda=435 \mathrm{~nm})$. Curve fitting analysis was performed using Igor pro program based on an isodesmic model. The fitting functions are given by eq. S1,

$$
\begin{equation*}
\varepsilon(C)=\frac{2 K_{a} C+1-\sqrt{4 K_{a} C+1}}{2 K_{a}^{2} C^{2}}(\varepsilon 1-\varepsilon a)+\varepsilon a \tag{S1}
\end{equation*}
$$

where $C, K_{\mathrm{a}}, \varepsilon_{1}$, and $\varepsilon_{\mathrm{a}}$ denote the total concentration of 2 , the affinity constant, the $\varepsilon$ of the monomer, and the $\varepsilon$ of the aggregate species, respectively.

Determination of diffusion coefficient (D) of the molecular species in the solution of 2. ${ }^{[4]}$
Chloroform- $d$ solutions of 2 were placed in an NMR sample tube ( $3 \mathrm{~mm} \Phi$ ). The pulse-field gradient diffusion NMR spectra were recorded using a bipolar pulse pair stimulated echo pulse sequence on a JEOL JNM-ECA500 spectrometer with a three mm inverse $\mathrm{H} 3 \mathrm{X} / \mathrm{FG}$ probe at $24^{\circ} \mathrm{C}$. The resulting DOSY data were analyzed using a MestReNova program to obtain the diffusion coefficient values $(D)$ shown in Fig. 2b. The signal decay of the selected protons was fit to fitting functions given by eq. S2 (Fig. S5), where $I$ and $I_{0}$ denote the NMR signal intensities in the presence and absence of gradient pulses, respectively. $D$ is the diffusion coefficient value.

$$
\begin{equation*}
I=I_{0} e^{-x D} \tag{S2}
\end{equation*}
$$

The x denotes $\left[-\gamma^{2} g^{2} \delta^{2}(\Delta-\delta / 3)\right]$, where $\gamma, g$, $\delta$, and $\Delta$ indicate the gyromagnetic ratio, gradient strength, its duration, and separation between the edges of the gradient pulses, respectively.


Scheme S1. Synthesis of 2.
bis(porphyrin) dimer 2: To a solution of $\mathbf{1}(61.2 \mathrm{mg}, 20.6 \mu \mathrm{~mol})$ in 1,4 -dioxane $(9.8 \mathrm{~mL})$ and water $(0.82 \mathrm{~mL})$ was added potassium hydroxide ( $23.0 \mathrm{mg}, 410 \mu \mathrm{~mol}$ ). The mixture was stirred for 2 h at $100^{\circ} \mathrm{C}$ under a nitrogen atmosphere. The reaction mixture was acidified with 1 M hydrochloric acid to pH 4 . After concentrating the organic layer to $1 / 2$ of its original volume, the resulting precipitate was filtered off, washed with water and MeOH , and dried over under the reduced pressure. The resulting solid was dissolved in dry 1,2 -dichloroethane ( 2.7 mL ) in a pressure tube, and dry triethylamine ( $50 \mu \mathrm{~L}$ ), diphenylphosphoryl azide ( $11.5 \mathrm{mg}, 41.8 \mu \mathrm{~mol}$ ), and 2-octyldecanol ( $159 \mathrm{mg}, 588 \mu \mathrm{~mol}$ ) were added at room temperature. The mixture was stirred for 20 h at $90^{\circ} \mathrm{C}$ under a nitrogen atmosphere and diluted with $\mathrm{CHCl}_{3}$. The organic layer was washed with aqueous $\mathrm{NaHCO}_{3}$, dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, and concentrated under the reduced pressure. MPLC (chloroform, eluent), subsequent silica-gel column chromatography ( $4 \% \mathrm{MeCN}$ in toluene, eluent), and GPC gave the desired product $\mathbf{2}$ ( 25.0 mg , $35 \%$, for 2 steps) as a purple solid.
M.p.: > $300^{\circ} \mathrm{C}$. ${ }^{1} \mathrm{H}$ NMR (pyridine- $d_{5}, 600 \mathrm{MHz}$ ): $\delta 11.39(\mathrm{~s}, 2 \mathrm{H}), 11.27(\mathrm{~s}, 4 \mathrm{H}), 9.25(\mathrm{~d}, 4 \mathrm{H}, J=5.0 \mathrm{~Hz}), 9.13(\mathrm{~d}, 4 \mathrm{H}, J=$ $4.8 \mathrm{~Hz}), 9.11(\mathrm{~d}, 4 \mathrm{H}, J=4.8 \mathrm{~Hz}), 9.08(\mathrm{~d}, 4 \mathrm{H}, J=5.0 \mathrm{~Hz}), 8.98(\mathrm{~m}, 8 \mathrm{H}), 8.87(\mathrm{~m}, 8 \mathrm{H}), 8.52$ (overlapped, 4H), 8.49 (overlapped, 8H), 8.43 (overlapped, 4H), 8.32 (overlapped, 10H), 8.16 (overlapped, 12H), 8.03 (m, 4H), 7.69-7.81 (overlapped, 16H), $7.63(\mathrm{~m}, 4 \mathrm{H}), 7.55(\mathrm{~m}, 8 \mathrm{H}), 4.52(\mathrm{~d}, 4 \mathrm{H}, J=6.3 \mathrm{~Hz}), 1.90(\mathrm{~m}, 2 \mathrm{H}), 1.22-1.57(\mathrm{~m}, 56 \mathrm{H}), 0.90(\mathrm{t}, 12 \mathrm{H}, J=6.5 \mathrm{~Hz}),-2.51(\mathrm{~s}$, $4 \mathrm{H}),-2.55(\mathrm{~s}, 4 \mathrm{H}) .{ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR (chloroform $-d_{1}, 100 \mathrm{MHz}$ ): $\delta 163.3,163.2,155.8,144.5,143.3,143.2,142.9,142.8,141.3$, $139.5,137.9,137.8,136.9,135.4,135.3,132.1,131.7,128.7,128.7,128.3,127.9,127.7,126.1,122.1,121.7,121.5,121.4$, 121.1, 121.0, 120.5, 120.0, 118.0, 83.6, 76.6, 68.3, 38.8, 32.7, 32.2, 30.9, 30.4, 30.1, 27.7, 23.5, 14.8. HRMS (ESI ${ }^{+}$) calcd. for $\mathrm{C}_{232} \mathrm{H}_{201} \mathrm{O}_{8} \mathrm{~N}_{24} \mathrm{~m} / \mathrm{z} 1150.2014[\mathrm{M}+3 \mathrm{H}]^{3+}$, found $m / z$ 1150.2024.


| 165 | 160 | 155 | 150 | 145 | 140 <br> $\delta(\mathrm{ppm})$ | 135 | 130 | 125 |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |



Figure S1. ${ }^{1} \mathrm{H}(600 \mathrm{MHz})$ and ${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}(150 \mathrm{MHz})$ spectra of $\mathbf{2}$ in pyridine- $d_{5}$. The expanded views of the spectra and the full spectra are shown in panels (a), (c) and (b), (d), respectively. Asterisk (*) denotes residual chlorofom.


Figure S2. DQF-COSY spectrum of $\mathbf{2}$ in pyridine- $d_{5}$. Asterisk $\left(^{*}\right)$ denotes residual chlorofom.


Figure S3. NOESY spectrum of $\mathbf{2}$ in pyridine- $d_{5}$. Asterisk (*) denotes residual chlorofom.


Figure S4. HSQC spectrum of $\mathbf{2}$ in pyridine- $d_{5} . \mathrm{CH}_{3}$ and CH carbons are phased up (red), and $\mathrm{CH}_{2}$ carbons are phased down (blue). Asterisk (*) denotes residual chlorofom.


Figure S5. Plot of ${ }^{1} \mathrm{H}$ NMR signal intensities obtained from the diffusion experiments for $\mathbf{2}$ in chloroform- $d$ at the concentrations of (a) 30 , (b) 20 , (c) 15 , (d) 10 , (e) 8.0 , (f) 5.0 , (g) 2.0 , and (h) $1.0 \mathrm{mmol} \mathrm{L}^{-1}$ and their fitting curves (red lines).


Figure S6. HRMS (ESI ${ }^{+}$) spectrum of 2 (top panel) and its expanded view (bottom panel).


Figure S7. UV/vis spectral absorption changes seen for 2 in $(+)$-limonene at $90^{\circ} \mathrm{C}$. The concentrations are (a-t) $0.51,1.0$, $1.5,2.0,2.5,3.5,4.4,5.3,6.3,7.2,8.0,8.9,9.7,10.6,11.4,12.2,13.0,14.0,15.0,15.8 \times 10^{-8} \mathrm{~mol} \mathrm{~L}^{-1}$. The blue and red lines indicate spectra of 2 at the concentrations of $0.51 \times 10^{-8} \mathrm{~mol} \mathrm{~L}^{-1}$ and $15.8 \times 10^{-8} \mathrm{~mol} \mathrm{~L}^{-1}$, respectively.


Figure S8. AFM image of the cast films of $\mathbf{2}$ on mica prepared from its (+)-limonene solution.


Figure S9. Valuable temperature ECD spectra of $2\left(2.5 \times 10^{-5} \mathrm{~mol} \mathrm{~L}^{-1}\right)$ in (a) (-)- $\alpha$-pinene and (b) (-)- $\beta$-pinene. The temperatures are $(\mathrm{a}-\mathrm{j}) 100,90,80,70,60,50,40,30,20,10,0^{\circ} \mathrm{C}$. The red and blue lines denote the spectra observed at $100^{\circ} \mathrm{C}$ and $0^{\circ} \mathrm{C}$, respectively.


Figure S10. ECD spectra of $\mathbf{2}\left(2.5 \times 10^{-5} \mathrm{~mol} \mathrm{~L}^{-1}\right)$ in the mixture of $(+)$ - and $(-)-\alpha-$ pinene with various ee values. The ee values are $100,90,80,70,60,50,40,30,20,10,0,-10,-20,-30,-40,-50,-60,-70,-80,-90,-100 \%$, where ee $=([(+)-\alpha-$ pinene $]-[(-)-\alpha-$ pinene $]) /([(+)-\alpha-$ pinene $]+[(-)-\alpha-$ pinene $])\}$.


| 422 nm |  |
| ---: | ---: |
| ee | $\Delta \varepsilon$ |
| 100 | 37.2927433 |
| 80 | 28.5187582 |
| 60 | 20.7358768 |
| 40 | 15.0092076 |
| 20 | 6.76389828 |
| 0 | 0.62267451 |
| -20 | -6.9165056 |
| -40 | -14.000052 |
| -60 | -22.297289 |
| -80 | -29.040231 |
| -100 | -36.354081 |



| 437 nm |  |
| ---: | ---: |
| ee | $\Delta \varepsilon$ |
| 100 | -37.903137 |
| 80 | -32.181483 |
| 60 | -25.824892 |
| 40 | -18.178104 |
| 20 | -11.68455 |
| 0 | -2.6949081 |
| -20 | 5.81970634 |
| -40 | 15.2501915 |
| -60 | 23.7771043 |
| -80 | 32.7335909 |
| -100 | 41.4838001 |

Figure S11. $\Delta \varepsilon$ values of $2\left(2.5 \times 10^{-5} \mathrm{~mol} \mathrm{~L}^{-1}\right)$ in the mixture of $(+)$ - and $(-)$ - $\alpha$-pinene with various ee values and the linear regression lines for the ee vs $\Delta \varepsilon$ values observed at (a) 422 nm and (b) 437 nm . The x and y indicate ee and $\Delta \varepsilon$ values, respectively. Enantiomeric excess $(\mathrm{ee})=\{([(+)$ - $\alpha$-pinene $]-[(-)$ - $\alpha$-pinene $]) /([(+)$ - $\alpha$-pinene $]+[(-)$ - $\alpha$-pinene $])\}$.

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