Supporting Information For:

# Supramolecular Chiral Sensing by Supramolecular Helical Polymers

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## General

All solvents were commercial reagent grade and were used without further purification except where noted. Dry 1,2dichloroethane and dry NEt<sub>3</sub> was obtained by distillation over CaH<sub>2</sub> and KOH, respectively. <sup>1</sup>H,<sup>13</sup>C, and 2D NMR (DQF-COSY, NOESY, and HSQC) spectra were recorded on a JEOL ECA-600 spectrometer, and chemical shifts were reported on the delta scale in ppm relative to residual pyridine ( $\delta$  = 8.74 and 150.4 for <sup>1</sup>H and <sup>13</sup>C, respectively). Diffusion-ordered <sup>1</sup>H NMR spectroscopic (DOSY) measurement was carried out on a JEOL ECA-500 spectrometer. Melting points (M.p.) were measured with a Yanagimoto micro melting point apparatus. Infrared (IR) spectra were recorded on a JASCO FT/IR-4600 spectrometer with ZeSe ATR accessory. Preparative separations were performed by silica gel gravity column chromatography (Silica Gel 60 N (spherical, neutral)). Preparative medium pressure liquid chromatography (MPLC) separations were carried out with a YAMAZEN smart flash EPCLC AL-580S using a preparative Bio-Beads (S-X1) column. Recycling preparative GPC-HPLC separations were carried out on JAI LaboACE LC-5060 using preparative JAIGEL-2.5HH, 2HH columns in series. UV/vis absorption spectra were recorded on a JASCO V-760 spectrometer. Circular dichroism (CD) spectra were recorded on a JASCO J-1500 spectrometer. Morphological evaluation of the samples was performed using atomic force microscopy (AFM) using a Bruker Multimode 8HR under ambient conditions with a Bruker cantilever model NCHV in the tapping mode. NanoScope Analysis 2.0 was used for the AFM image analysis. **1**<sup>[1]</sup> and 2-octyldecanol<sup>[2]</sup> were synthesized according to reported methods.

#### Method

Determination of an affinity constant ( $K_a$ ) corresponding to the intermolecular interactions between bis(porphyrin) moieties of **2**.<sup>[3]</sup>

A hyperbolic curve was obtained by plotting the concentrations of **2** against the extinction coefficients ( $\varepsilon$ ) ( $\lambda$  = 435 nm). Curve fitting analysis was performed using Igor pro program based on an isodesmic model. The fitting functions are given by eq. S1,

$$\varepsilon(C) = \frac{2K_aC + 1 - \sqrt{4K_aC + 1}}{2K_a^2C^2}(\varepsilon 1 - \varepsilon a) + \varepsilon a \tag{S1}$$

where C,  $K_a$ ,  $\varepsilon_1$ , and  $\varepsilon_a$  denote the total concentration of **2**, the affinity constant, the  $\varepsilon$  of the monomer, and the  $\varepsilon$  of the aggregate species, respectively.

#### Determination of diffusion coefficient (D) of the molecular species in the solution of 2.<sup>[4]</sup>

Chloroform-*d* solutions of **2** were placed in an NMR sample tube (3 mm $\Phi$ ). The pulse-field gradient diffusion NMR spectra were recorded using a bipolar pulse pair stimulated echo pulse sequence on a JEOL JNM-ECA500 spectrometer with a three mm inverse H3X/FG probe at 24 °C. The resulting DOSY data were analyzed using a MestReNova program to obtain the diffusion coefficient values (*D*) shown in Fig. 2b. The signal decay of the selected protons was fit to fitting functions given by eq. S2 (Fig. S5), where *I* and *I*<sub>0</sub> denote the NMR signal intensities in the presence and absence of gradient pulses, respectively. *D* is the diffusion coefficient value.

$$I = I_0 e^{-xD} \tag{S2}$$

The x denotes  $[-\gamma^2 g^2 \delta^2 (\Delta - \delta/3)]$ , where  $\gamma$ , g,  $\delta$ , and  $\Delta$  indicate the gyromagnetic ratio, gradient strength, its duration, and separation between the edges of the gradient pulses, respectively.



1) KOH, H<sub>2</sub>O, 1,4-dioxane

2) 2-octyldecanol, dry NEt<sub>3</sub>, diphenylphosphoryl azide, dry 1,2-dichloroethane



Scheme S1. Synthesis of 2.

**bis(porphyrin) dimer 2:** To a solution of **1** (61.2 mg, 20.6  $\mu$ mol) in 1,4-dioxane (9.8 mL) and water (0.82mL) was added potassium hydroxide (23.0 mg, 410  $\mu$ mol). The mixture was stirred for 2 h at 100 °C under a nitrogen atmosphere. The reaction mixture was acidified with 1M hydrochloric acid to pH 4. After concentrating the organic layer to 1/2 of its original volume, the resulting precipitate was filtered off, washed with water and MeOH, and dried over under the reduced pressure. The resulting solid was dissolved in dry 1,2-dichloroethane (2.7 mL) in a pressure tube, and dry triethylamine (50  $\mu$ L), diphenylphosphoryl azide (11.5 mg, 41.8  $\mu$ mol), and 2-octyldecanol (159 mg, 588  $\mu$ mol) were added at room temperature. The mixture was stirred for 20 h at 90 °C under a nitrogen atmosphere and diluted with CHCl<sub>3</sub>. The organic layer was washed with aqueous NaHCO<sub>3</sub>, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated under the reduced pressure. MPLC (chloroform, eluent), subsequent silica-gel column chromatography (4% MeCN in toluene, eluent), and GPC gave the desired product **2** (25.0 mg, 35%, for 2 steps) as a purple solid.

M.p.: > 300 °C. <sup>1</sup>H NMR (pyridine- $d_5$ , 600 MHz):  $\delta$  11.39 (s, 2H), 11.27 (s, 4H), 9.25 (d, 4H, J = 5.0 Hz), 9.13 (d, 4H, J = 4.8 Hz), 9.11 (d, 4H, J = 4.8 Hz), 9.08 (d, 4H, J = 5.0 Hz), 8.98 (m, 8H), 8.87 (m, 8H), 8.52 (*overlapped*, 4H), 8.49 (*overlapped*, 8H), 8.43 (*overlapped*, 4H), 8.32 (*overlapped*, 10H), 8.16 (*overlapped*, 12H), 8.03 (m, 4H), 7.69–7.81 (*overlapped*, 16H), 7.63 (m, 4H), 7.55 (m, 8H), 4.52 (d, 4H, J = 6.3 Hz), 1.90 (m, 2H), 1.22–1.57 (m, 56H), 0.90 (t, 12H, J = 6.5 Hz), -2.51 (s, 4H), -2.55 (s, 4H). <sup>13</sup>C{<sup>1</sup>H} NMR (chloroform- $d_1$ , 100 MHz):  $\delta$  163.3, 163.2, 155.8, 144.5, 143.3, 143.2, 142.9, 142.8, 141.3, 139.5, 137.9, 137.8, 136.9, 135.4, 135.3, 132.1, 131.7, 128.7, 128.7, 128.3, 127.9, 127.7, 126.1, 122.1, 121.7, 121.5, 121.4, 121.1, 121.0, 120.5, 120.0, 118.0, 83.6, 76.6, 68.3, 38.8, 32.7, 32.2, 30.9, 30.4, 30.1, 27.7, 23.5, 14.8. HRMS (ESI<sup>+</sup>) calcd. for C<sub>232</sub>H<sub>201</sub>O<sub>8</sub>N<sub>24</sub> *m*/*z* 1150.2014 [M+3H]<sup>3+</sup>, found *m*/*z* 1150.2024.



**Figure S1.** <sup>1</sup>H (600 MHz) and <sup>13</sup>C{<sup>1</sup>H} (150 MHz) spectra of **2** in pyridine- $d_5$ . The expanded views of the spectra and the full spectra are shown in panels (a), (c) and (b), (d), respectively. Asterisk (\*) denotes residual chlorofom.



Figure S2. DQF-COSY spectrum of 2 in pyridine-d<sub>5</sub>. Asterisk (\*) denotes residual chlorofom.



Figure S3. NOESY spectrum of 2 in pyridine- $d_5$ . Asterisk (\*) denotes residual chlorofom.



**Figure S4.** HSQC spectrum of **2** in pyridine- $d_5$ . CH<sub>3</sub> and CH carbons are phased up (red), and CH<sub>2</sub> carbons are phased down (blue). Asterisk (\*) denotes residual chlorofom.



**Figure S5.** Plot of <sup>1</sup>H NMR signal intensities obtained from the diffusion experiments for **2** in chloroform-*d* at the concentrations of (a) 30, (b) 20, (c) 15, (d) 10, (e)8.0, (f) 5.0, (g) 2.0, and (h) 1.0 mmol L<sup>-1</sup> and their fitting curves (red lines).



Figure S6. HRMS (ESI<sup>+</sup>) spectrum of 2 (top panel) and its expanded view (bottom panel).



Figure S7. UV/vis spectral absorption changes seen for 2 in (+)-limonene at 90 °C. The concentrations are (a-t) 0.51, 1.0, 1.5, 2.0, 2.5, 3.5, 4.4, 5.3, 6.3, 7.2, 8.0, 8.9, 9.7, 10.6, 11.4, 12.2, 13.0, 14.0, 15.0,  $15.8 \times 10^{-8}$  mol L<sup>-1</sup>. The blue and red lines indicate spectra of 2 at the concentrations of  $0.51 \times 10^{-8}$  mol L<sup>-1</sup> and  $15.8 \times 10^{-8}$  mol L<sup>-1</sup>, respectively.



Figure S8. AFM image of the cast films of 2 on mica prepared from its (+)-limonene solution.



**Figure S9.** Valuable temperature ECD spectra of **2** ( $2.5 \times 10^{-5} \text{ mol } L^{-1}$ ) in (a) (–)- $\alpha$ -pinene and (b) (–)- $\beta$ -pinene. The temperatures are (a-j) 100, 90, 80, 70, 60, 50, 40, 30, 20, 10, 0 °C. The red and blue lines denote the spectra observed at 100 °C and 0 °C, respectively.



**Figure S10.** ECD spectra of **2** ( $2.5 \times 10^{-5} \text{ mol } \text{L}^{-1}$ ) in the mixture of (+)- and (-)- $\alpha$ -pinene with various ee values. The ee values are 100, 90, 80, 70, 60, 50, 40, 30, 20, 10, 0, -10, -20, -30, -40, -50, -60, -70, -80, -90, -100%, where ee = ([(+)- $\alpha$ -pinene] - [(-)- $\alpha$ -pinene]) / ([(+)- $\alpha$ -pinene] + [(-)- $\alpha$ -pinene])}.



**Figure S11.**  $\Delta \varepsilon$  values of **2** (2.5 × 10<sup>-5</sup> mol L<sup>-1</sup>) in the mixture of (+)- and (-)- $\alpha$ -pinene with various ee values and the linear regression lines for the ee vs  $\Delta \varepsilon$  values observed at (a) 422 nm and (b) 437 nm. The x and y indicate ee and  $\Delta \varepsilon$  values, respectively. Enantiomeric excess (ee) = {([(+)- $\alpha$ -pinene] – [(-)- $\alpha$ -pinene]) / ([(+)- $\alpha$ -pinene] + [(-)- $\alpha$ -pinene])}.

## References

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