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Supporting Information (SI)

For

**Molecular mechanisms of humic acid-enhanced formation of the
ordered secondary structure of a conserved catalytic domain in
phytase**

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24 **Table S1.** Assignments of Amide I and Amide III band positions for protein or25 peptide secondary structures with respective to IR and Raman spectra.¹⁻⁴

Structures	Amide I (cm ⁻¹)	Amide III (cm ⁻¹)
α -helix	1648-1657	1260-1310
Radom coil	1642-1660	1230-1260
β -sheet	1623-1641	1220-1265

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28 **Table S2.** ΔG_b and corresponding fitting parameters of -COOH, NH₃⁺ and CH₃ from29 DFS data (x_t , distance to transition between bound and unbound states; f_{eq} ,30 equilibrium force). All values are represented as mean \pm standard deviation.

Solutions	Surfaces	Functional groups	k (N/m)	Fitting parameters		$-\Delta G_b$ (KJ/mol)
				x_t (Å)	f_{eq} (pN)	
without phytate	mica	COOH	0.1012	0.82 \pm 0.066	230.53 \pm 14.20	17.42 \pm 1.98
		NH ₃ ⁺	0.1078	0.90 \pm 0.052	165.19 \pm 12.04	14.56 \pm 1.65
		CH ₃	0.0965	0.61 \pm 0.042	82.68 \pm 7.49	6.45 \pm 0.83
	HA	COOH	0.0921	0.90 \pm 0.088	189.04 \pm 12.46	16.17 \pm 1.75
		NH ₃ ⁺	0.1105	0.91 \pm 0.034	251.98 \pm 15.89	21.76 \pm 2.62
		CH ₃	0.0791	0.64 \pm 0.055	90.45 \pm 7.84	6.55 \pm 0.96
with phytate	mica	COOH	0.0872	0.74 \pm 0.048	239.20 \pm 19.14	16.78 \pm 1.76
		NH ₃ ⁺	0.0932	0.72 \pm 0.052	157.61 \pm 8.90	12.13 \pm 1.53
		CH ₃	0.0784	0.75 \pm 0.053	84.12 \pm 6.04	7.32 \pm 0.78
	HA	COOH	0.0694	0.73 \pm 0.064	193.87 \pm 13.19	14.45 \pm 1.86
		NH ₃ ⁺	0.0943	0.85 \pm 0.047	238.84 \pm 16.24	19.10 \pm 1.54
		CH ₃	0.0847	0.62 \pm 0.067	87.28 \pm 7.94	6.93 \pm 0.82

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32 **Table S3.** ΔG_b and corresponding fitting parameters of ACD, ACD-1 and ACD-233 from DFS data (x_t , distance to transition between bound and unbound states; f_{eq} ,

34 equilibrium force). All values are represented as mean \pm standard deviation.

Solutions	Surfaces	Active domains	k (N/m)	Fitting parameters		$-\Delta G_b$ (KJ/mol)
				x_t (Å)	f_{eq} (pN)	
without phytate	mica	ACD	0.0915	0.54 \pm 0.044	95.52 \pm 6.07	6.14 \pm 0.55
		ACD-1	0.0786	0.60 \pm 0.045	86.17 \pm 7.28	6.13 \pm 0.88
		ACD-2	0.0623	0.63 \pm 0.032	90.14 \pm 6.47	6.23 \pm 0.67
	HA	ACD	0.0831	0.77 \pm 0.086	203.59 \pm 14.3	15.71 \pm 1.2
		ACD-1	0.0905	0.59 \pm 0.038	91.49 \pm 8.49	6.38 \pm 0.51
		ACD-2	0.0718	0.64 \pm 0.057	204.78 \pm 14.2	14.8 \pm 1.57
with phytate	mica	ACD	0.0859	0.66 \pm 0.047	167.31 \pm 9.65	11.64 \pm 0.63
		ACD-1	0.0672	0.71 \pm 0.052	87.04 \pm 9.12	7.20 \pm 0.60
		ACD-2	0.0826	0.53 \pm 0.062	87.54 \pm 6.89	6.37 \pm 0.72
	HA	ACD	0.0799	0.88 \pm 0.054	231.26 \pm 13.14	19.29 \pm 1.94
		ACD-1	0.1043	0.59 \pm 0.057	94.74 \pm 8.16	6.58 \pm 0.96
		ACD-2	0.0619	0.74 \pm 0.057	96.45 \pm 12.2	6.7 \pm 0.88

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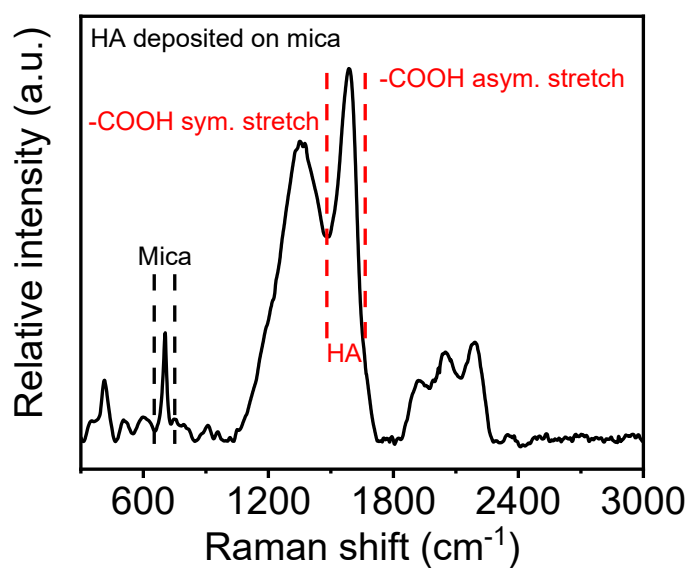
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37 **Full-length phytase (FLP):**

38	MSDMKSGNIS	RHGVRAPPFL	SLLIPLTPQS	AFAQSEPELK
39	LESVVIVSMK	AILITKATQL	MQDVTPDAWP	TWPVKLGWLT
40	PRGGELIAYL	GHYQRQRLVA	DGLLTKKGCP	QPGQVAIID
41	VDERTRKTGE	AFAAGLAPDC	AISVHTQADT	SSPDPLFNPL
42	KTGVCQLDNA	NVTDAILSRA	GGSIADFTGH	RQTAFRELER
43	VLNFPQSNLC	LNREKQDESC	SLTQALPSEL	KVSADNVSLT
44	GAVSLASMLT	EIFLLQ HD QG	MPEPGGEGSP	IHTSGTPC

45 **Fig. S1.** Amino acid sequences of FLP (Origin: EC 3.1.3.26, *Escherichia coli*)

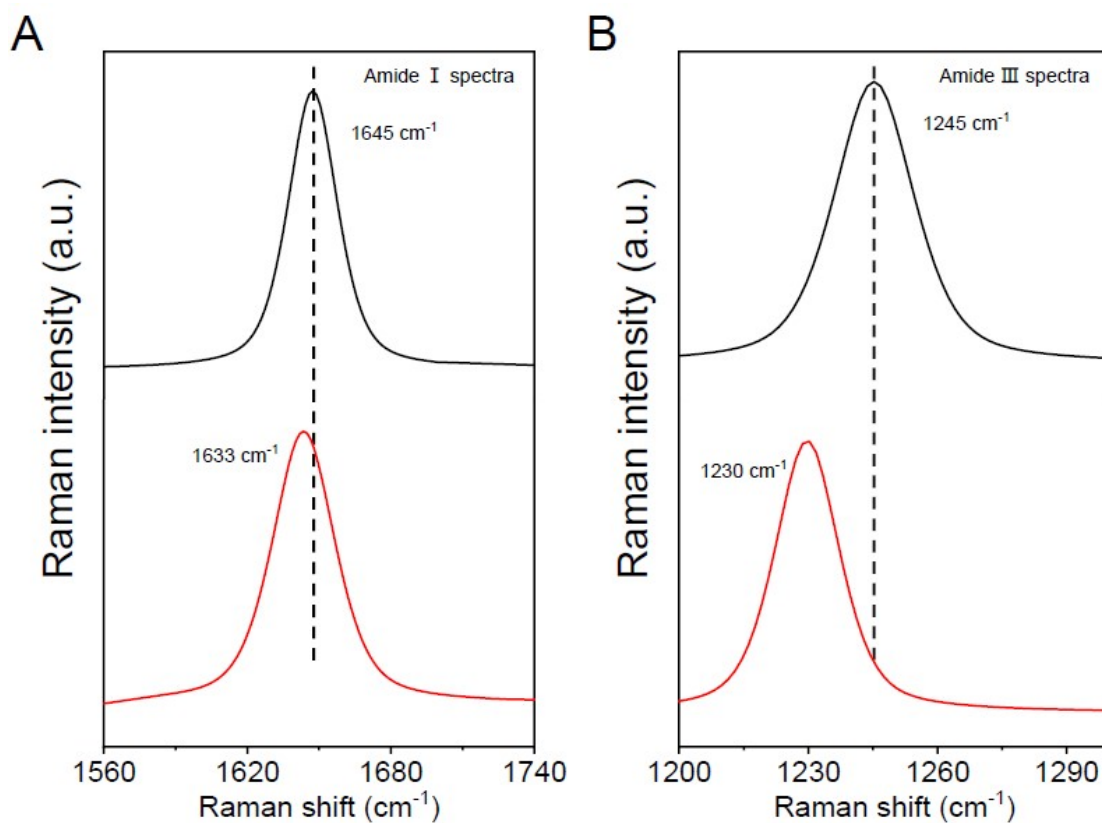
46 containing N-terminal (RHGVRAP, ACD-1) and C-terminal (HD) active domains.



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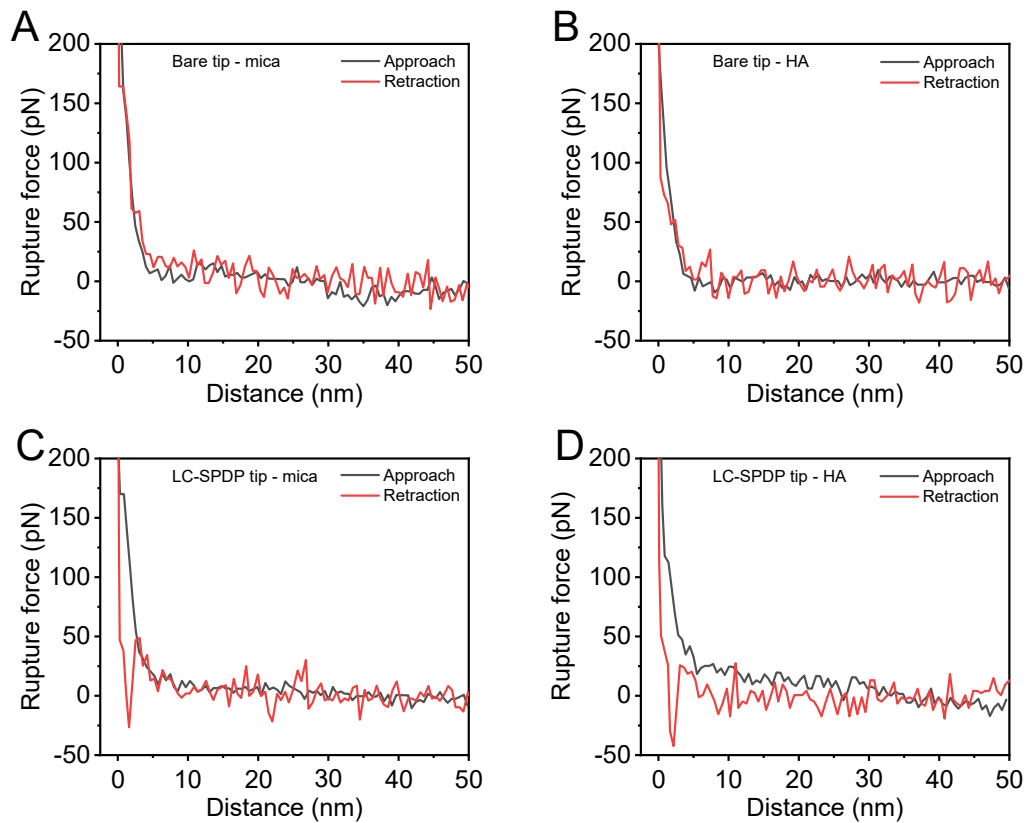
48 **Fig. S2.** Raman characteristic vibrations of mica and HA selected to conduct Raman
 49 mapping.

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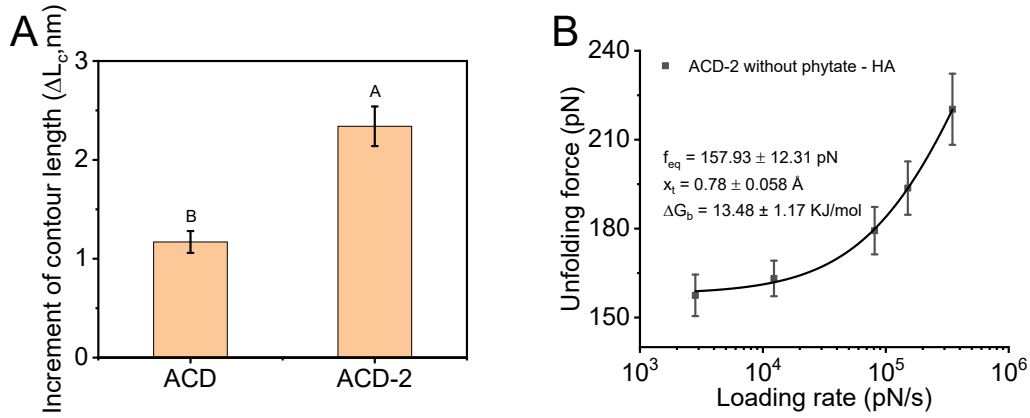
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52 **Fig. S3.** Secondary structures of ACD in the region of (A) Amide I and (B) Amide III
53 before (black lines) and after (red lines) treatment with HA.



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55 **Fig. S4.** Representative F-D curves between the Au-coated bare tip with (A) mica and
56 (B) HA surface. Representative F-D curves between the LC-SPDP-functionalized tip
57 with (C) mica and (D) HA surface. These typical F-D curves as key controls
58 collectively excluded the interference from the interaction between Au-coated or LC-
59 SPDP-functionalized tips and the two surfaces compared to functionalized tips of
60 ACD(-1/2) as well as $-\text{CH}_3-$, $-\text{COOH}-$, $-\text{NH}_3-$ terminal organic molecules.



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62 **Fig. S5.** (A) Analyses of ΔL_c of ACD and ACD-2 separated from HA deposited on

63 mica surface. Different uppercase letters indicate significant difference at $P < 0.01$,

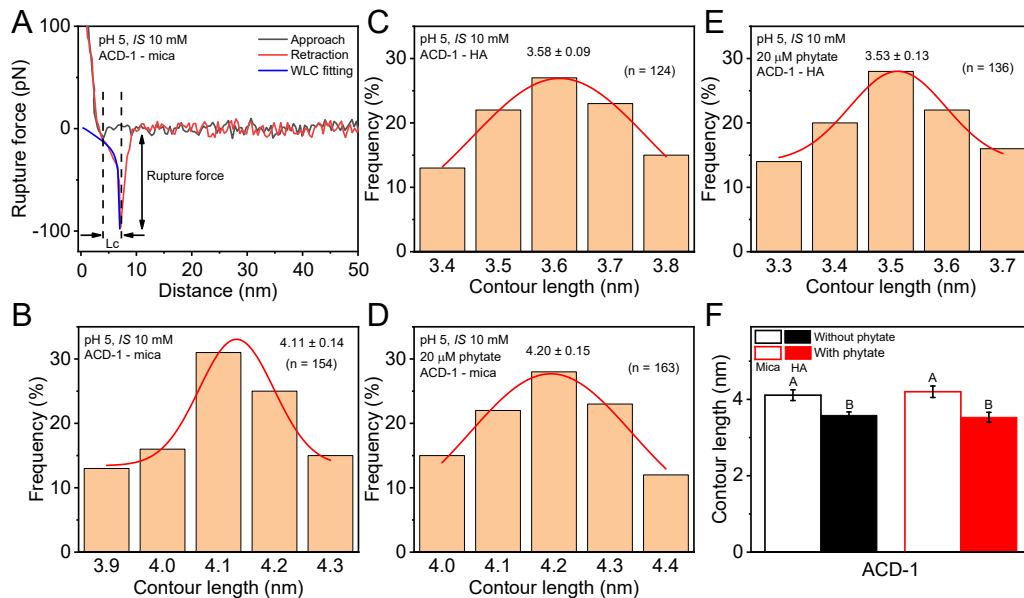
64 which was analyzed by SPSS software. (B) A plot of mean unfolding force of the first

65 force-peak occurred in double-force curves versus loading rates, determining f_{eq} and

66 ΔG_b for allowing to break H-bonding as ACD-2 is retracted from HA. The derived f_{eq}

67 and ΔG_b are presented as mean \pm standard deviations ($n = 3$).

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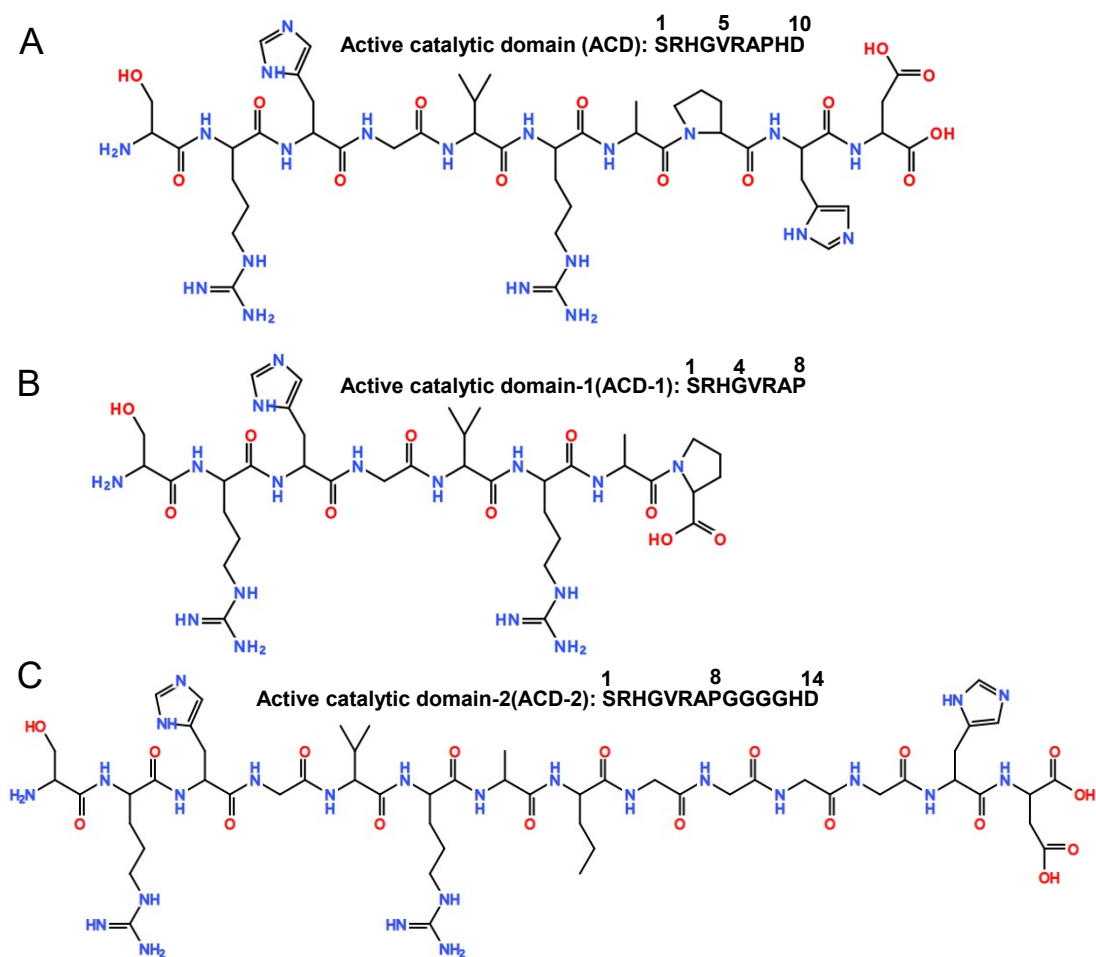


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70 **Fig. S6.** (A) Representative F-D curve between ACD-1 and mica fitted by the WLC

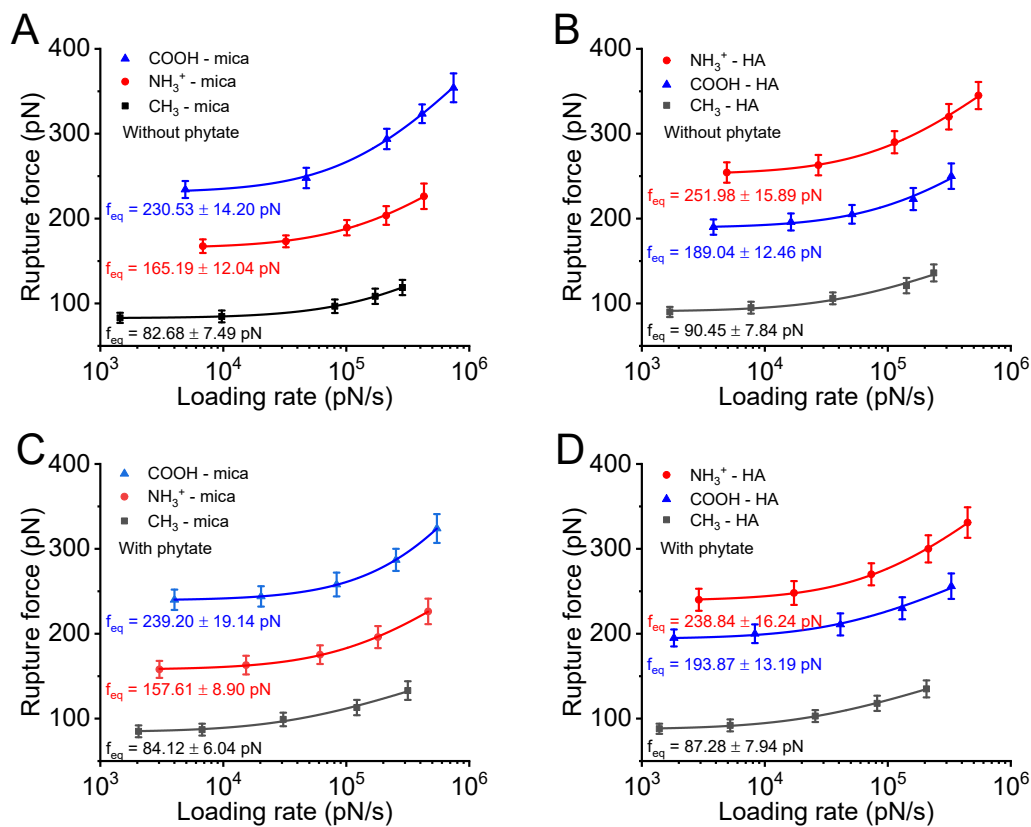
71 model. The contour length (L_c) obtained from F-D curves through WLC fitting during
72 detaching ACD-1 from (B, D) mica and (C, E) mica with adsorbed HA in solutions
73 (pH 5 and $IS = 10$ mM NaCl) in the absence and presence of $20 \mu\text{M}$ phytate. (F)
74 Analysis of L_c of ACD-1 separated from mica or mica with adsorbed HA. Different
75 uppercase letters indicate significant difference at $P < 0.01$, which was analyzed by
76 SPSS software.

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79 **Fig. S7.** Chemical structures of (A) ACD, (B) ACD-1 and (C) ACD-2 showing
80 representative chemistries containing CH_3 , NH_3^+ and COOH .



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82 **Fig. S8.** Plots of mean rupture forces versus loading rates of COOH-, NH₃⁺- or CH₃-

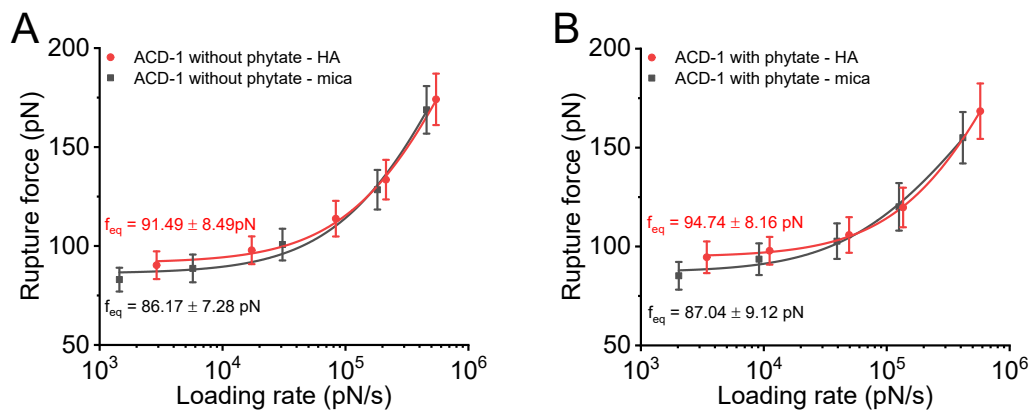
83 terminal model organic molecule at the mica/HA interface in the (A, B) absence and

84 (C, D) presence of 20 μM phytate in solutions (pH 5 and 1S 10 mM NaCl).

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89 **Fig. S9.** Plots of mean rupture forces versus loading rates of ACD-1 at the mica/HA
 90 interface in the (A) absence and (B) presence of 20 μM phytate in solutions (pH 5 and
 91 I_S 10 mM).

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