

Organic acid formation in the gas-phase ozonolysis of α,β -unsaturated ketones

- Supporting Information -

N. Illmann, I. Patroescu-Klotz, and P. Wiesen

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A. Quantification and supplementary yield plots

Table S1 Absolute and integrated absorption cross sections (base 10) used for the quantification via FTIR spectroscopy.

Species	Range / cm^{-1}	Integrated cross section / cm molecule^{-1}	Absolute cross section / $\text{cm}^2 \text{ molecule}^{-1}$	Reference
Acetaldehyde	1761		$(1.7 \pm 0.1) \times 10^{-19}$	b
Ethyl hydroperoxide	2992		$(8.1 \pm 0.4) \times 10^{-20}$	b
Ethyl vinyl ketone	1768–1660	$(8.2 \pm 0.6) \times 10^{-18}$		a
Formaldehyde	1820–1660	$(5.5 \pm 0.3) \times 10^{-18}$		b
Formic acid	1840–1704	$(2.6 \pm 0.5) \times 10^{-17}$		b
Formic anhydride	1822		$(2.6 \pm 0.4) \times 10^{-19}$	b
Formic propionic anhydride	1824–1762	$(2.6 \pm 0.3) \times 10^{-17}$		b
Propionic acid	1880–1710	$(1.6 \pm 0.1) \times 10^{-17}$		a

^a Determined within this work, ^b Wuppertal laboratory database.

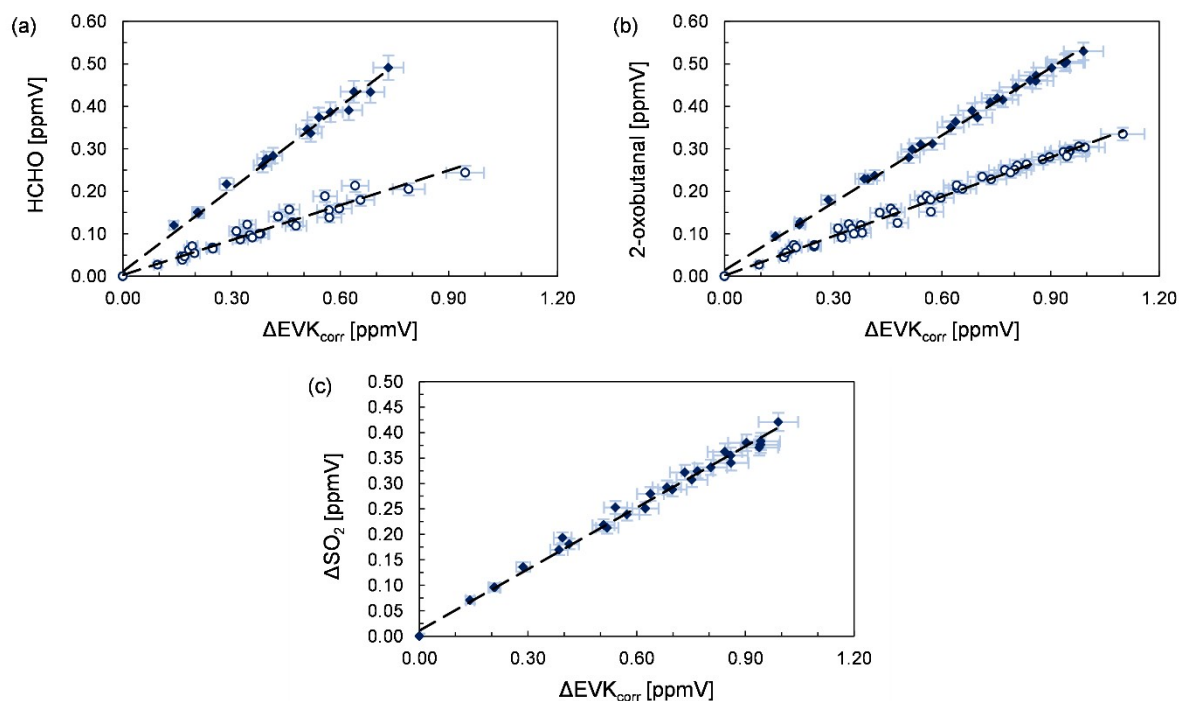


Fig. S1 Yield plots for (a) HCHO, and (b) 2-oxobutanal in the absence (open circles) and presence (filled diamonds) of SO_2 , and (c) the consumption of SO_2 vs. the consumption of EVK. The error bars represent the respective precision errors.

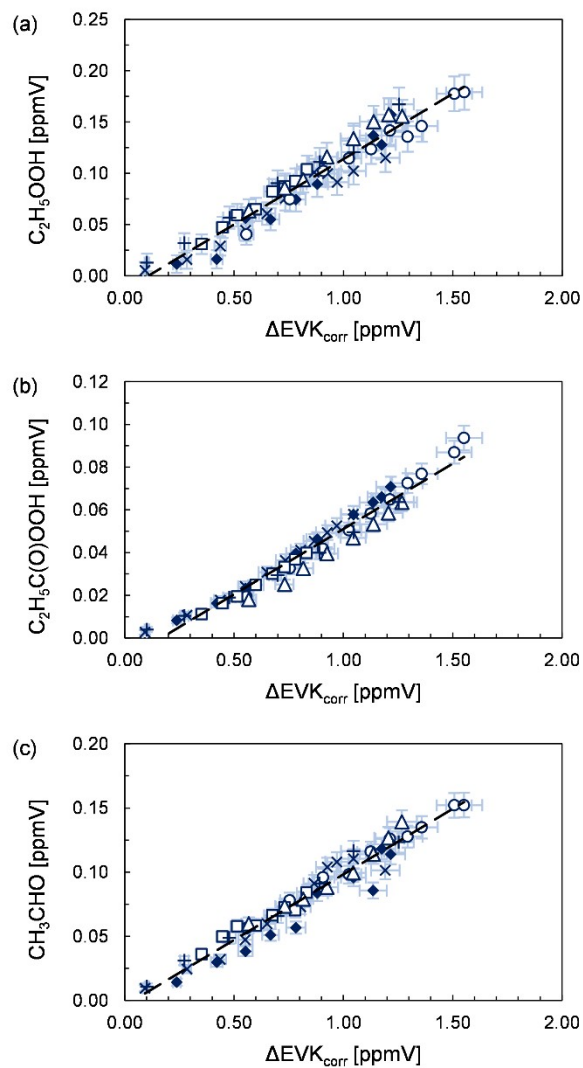


Fig. S2 Yield plots obtained for (a) ethyl hydroperoxide, (b) perpropionic acid, and (c) acetaldehyde, respectively, in the absence of SO_2 . Different experimental runs are denoted by different symbols. The error bars represent a precision error as a combination of a relative error plus the corresponding detection limit under the experimental conditions.

B. PTR-ToF-MS data analysis

In each experiment (in the absence of SO₂), the PTR-ToF-MS data show the formation of species resulting in signals at m/z 31, 45, 47, 57, 59, 75, 87 and 91. Representative time profiles for all signals are shown in Fig. S4. The assignment to species is summarized in Tab. S2.

The formation of HCHO is confirmed by the increase of the m/z 31 signal (Fig S3). 2-Oxobutanal formation is strongly indicated by the evolution of the m/z 87 signal (Fig. S3) corresponding to protonated 2-oxobutanal (C₄H₇O₂⁺, [M-H]⁺). A m/z 59 signal, following a similar evolution (Fig. S4), is observed. For the structural analogue oxoaldehyde methyl glyoxal, beside the [M-H]⁺ signal at m/z 73, fragment ions were reported at m/z 45 (C₂H₅O⁺), which corresponds to a loss of CO.¹ Accordingly, the m/z 59 signal is likely a fragment ion of 2-oxobutanal (C₃H₇O⁺). At the same time, methyl ketene, as suggested in the ozonolysis mechanism, might contribute to m/z 59.

The formation of formic acid is possibly supported by the evolution of the m/z 47 signal (CH₃O₂⁺). In separate control experiments we observed that acetic anhydride is found solely as m/z 61 signal (C₂H₅O₃⁺) suggesting anhydrides to appear exclusively as [M-H]⁺ of their corresponding acid. Given that both formic anhydride (FA) and formic propionic anhydride (FPA) are observed in the FTIR spectra, the presence of the m/z 47 signal is probably mainly a consequence of the anhydride formation.

The proton-transfer reaction of peracetic acid was shown to produce mainly (\approx 90 %) protonated acetic acid,² which hence interferes with acetic acid at m/z 61 (C₂H₅O₂⁺). A similar behaviour is likely for perpropionic acid, the expected dominant mass signal being m/z 75 (C₃H₇O₂⁺). Due to the formation of formic propionic acid in the reaction system, the increase of the m/z 75 signal is not an unequivocal proof for the peracid. However, although much less intense (by a factor of $\approx 10^2$), an increase is observed also for the m/z 91 ion, which could result from the protonated perpropionic acid.

The formation of ethyl hydroperoxide was clearly observed in the FTIR spectra. However, no signal was detected at m/z 63, corresponding to the protonated hydroperoxide (C₂H₇O₂⁺). The fragment ion (C₂H₅O⁺), built upon elimination of H₂O, interferes with acetaldehyde at m/z 45. Hence, although the evolution of the m/z 45 ion supports the FTIR data, a differentiation between the contribution of acetaldehyde and an ethyl hydroperoxide fragment is not possible.

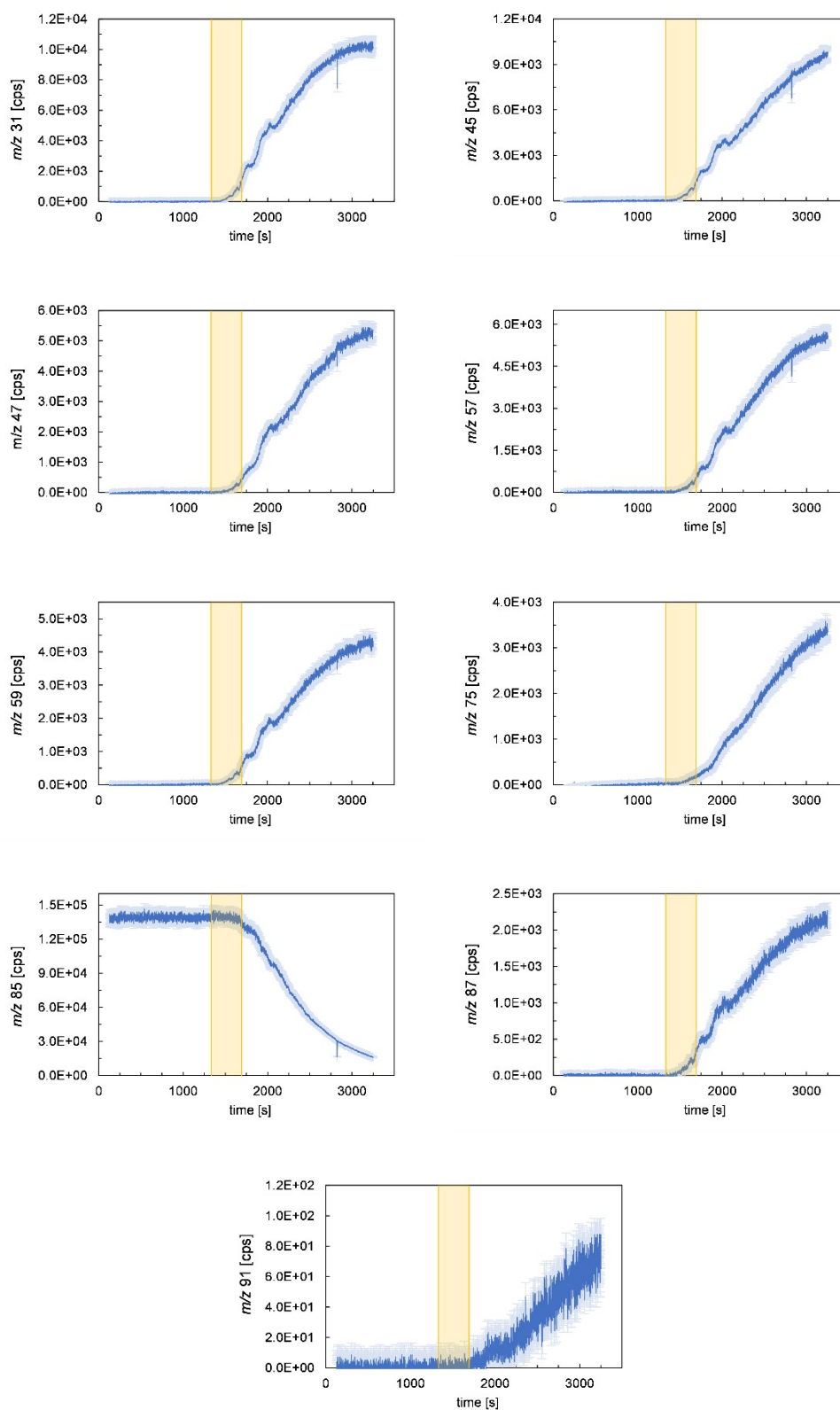


Fig. S3 Time profiles for m/z 31, 45, 47, 57, 59, 75, 85, 87, and 91 (dark blue) from a single EVK + O₃ experiment without sCI scavenger. The error bars (pale blue) represent the precision error. The pale yellow range marks the time of the ozone addition.

Table S2 Mass signals observed in the EVK + O₃ experiments, elemental composition and assigned species.

<i>m/z</i>	composition	Assigned species
31.018	CH ₃ O ⁺	HCHO
45.034	C ₂ H ₅ O ⁺	Acetaldehyde, Ethyl hydroperoxide (fragment)
47.013	CH ₃ O ₂ ⁺	Formic acid, Formic anhydride, Formic propionic anhydride
57.034	C ₃ H ₅ O ⁺	Methyl ketene or Propionyl
59.050	C ₃ H ₇ O ⁺	Propionaldehyde, 2-Oxobutanal (fragment)
75.045	C ₃ H ₇ O ₂ ⁺	Perpropionic acid, Formic propionic anhydride
87.045	C ₄ H ₇ O ₂ ⁺	2-Oxobutanal
91.040	C ₃ H ₇ O ₃ ⁺	Perpropionic acid

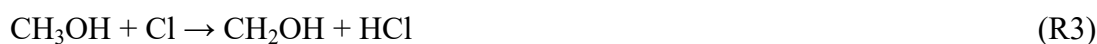
C. In situ generation and identification of peracids

1. Experimental

Infrared spectral features of peracetic acid (= peroxyacetic acid) and perpropionic acid (= peroxypropionic acid) were generated by the irradiation (I_{\max} at 360 nm) of acetaldehyde/methanol/Cl₂ and propionaldehyde/methanol/Cl₂ mixtures in synthetic air, respectively, in both the 480 L and 1080 L chamber. The current set-up of the 480 L chamber is described in detail elsewhere.³ Given that both the acetaldehyde + Cl and the propionaldehyde + Cl reaction proceed nearly exclusively via the abstraction of the aldehydic H atom, the respective RO₂ radicals are generated as follows:



In order to favour RO₂ + HO₂ reactions, the level of HO₂ is increased due to CH₃OH + Cl and the subsequent reaction sequence:



The [HO₂]/[RO₂] ratio can be adjusted by the initial [aldehyde]/[CH₃OH] ratio. Accordingly, the initial mixing ratios were set at 1.6–4.4 ppmV for acetaldehyde (Sigma Aldrich, > 99.5 %),

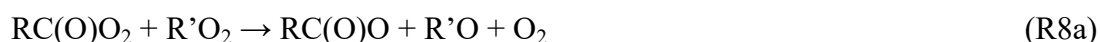
1.1–7.3 ppmV for propionaldehyde (Sigma Aldrich, 97 %), 1.9–16 ppmV for methanol (Sigma Aldrich, 99.9 %) and 1.9–21 ppmV for Cl₂ (Air Liquide, 99.8 %).

2. Results and discussion

Among RO₂ + HO₂ reactions the acetyl peroxy radical + HO₂ system is one of the most investigated reactions and reasonably well characterized.^{4,11} The reported studies performed in simulation chambers were conducted mainly by irradiating acetaldehyde/methanol/Cl₂ mixtures.^{6,7,10} Therefore, the in situ generation of peracetic acid was chosen to test the experimental set-up. All experiments were observed up to a maximum consumption of 60 – 70 % of the initial acetaldehyde. In order to promote RO₂ + HO₂ rather than RO₂ + RO₂ reactions, the initial aldehyde concentration was kept always lower than [CH₃OH]₀. In all experiments, significant amounts of hydrogen peroxide (H₂O₂) are formed. Since this can be rationalized through the HO₂ self-reaction, it indicates the HO₂ level to be near the upper experimental limit. In the case of acyl peroxy radicals, the RO₂ + HO₂ reaction was reported⁶ to proceed through 3 different channels:



Accordingly, both peracetic acid and acetic acid are formed in the CH₃C(O)H/CH₃OH/Cl₂ system. The alkoxy radical (R7c) eliminates CO₂ to yield a methyl radical, that is immediately converted into a methyl peroxy radical (CH₃O₂). The CH₃O₂ further chemistry evolves into methyl hydroperoxide and HCHO through the reaction with HO₂ and, possibly, into HCHO and CH₃OH through the CH₃O₂ self-reaction. However, if RO₂ + RO₂ plays a role, the cross reaction of the alkyl peroxy and acyl peroxy radical is about a factor of 10² faster and proceeds as follows¹²



In consequence, after the subtraction of the reaction sequence's products (acetic acid, O₃, CO₂, CH₃OOH, HCHO) as well as CH₃OH, HCHO, HC(O)OH, H₂O₂ and HCl, the residual

spectrum of such a reaction mixture should correspond to peracetic acid. Trace (b) of Fig. S4 shows a residual spectrum obtained from an irradiated $\text{CH}_3\text{C}(\text{O})\text{H}/\text{CH}_3\text{OH}/\text{Cl}_2$ mixture, which is in excellent agreement with a reference spectrum of peracetic acid from the Wuppertal laboratory database, presented in trace (a). The residual spectrum shows additional small curvatures centred at 1150 cm^{-1} and 1050 cm^{-1} . However, this can be partly explained by artefacts due to the subtraction of high amounts of methanol, whose absorption dominates at 1050 cm^{-1} . Additionally, a curvature of the baseline was observed in this region in some experiments, which might also be built artificially due to the subtraction process. Despite this, the experimental set-up was proven successful in obtaining the spectral features of peracids, as shown in Fig. S4.

The experiments irradiating $\text{C}_2\text{H}_5\text{C}(\text{O})\text{H}/\text{CH}_3\text{OH}/\text{Cl}_2$ mixtures were performed until a maximum consumption of 30–60 % of propionaldehyde. Similarly to the irradiation of mixtures containing acetaldehyde, H_2O_2 formation was also observed. This indicates a high HO_2 level in the experimental system, necessary to promote the $\text{RO}_2 + \text{HO}_2$ reactions. According to the reactions (R7a)–(R7c) perpropionic acid, propionic acid and $\text{C}_2\text{H}_5\text{C}(\text{O})\text{O}$ radicals are formed from the $\text{C}_2\text{H}_5\text{C}(\text{O})\text{O}_2 + \text{HO}_2$ reaction. The existence of all 3 reaction channels has been proven previously by Hasson et al.¹³ However, although FTIR spectroscopy has also been used in the Hasson et al. study,¹³ the peracid was quantified with HPLC and up to date no IR spectrum was reported for perpropionic acid.

As discussed above, in the presence of O_2 , the $\text{C}_2\text{H}_5\text{C}(\text{O})\text{O}$ radical eliminates CO_2 to form the ethyl peroxy radical. Similarly to methyl peroxy radical, the ethyl peroxy radical further chemistry yields ethyl hydroperoxide and possibly acetaldehyde. The residual spectrum assigned to perpropionic acid was obtained by subtraction of the spectral features of the following species: propionaldehyde, methanol, HCHO , formic acid, CO_2 , propionic acid, ethyl hydroperoxide, O_3 , acetaldehyde, peracetic acid, H_2O_2 , and HCl . The perpropionic acid spectrum is presented in Fig. S4, trace (c).

The spectrum contains characteristic absorption bands centred at 3303, 1760, 1450, 1180 and 880 cm^{-1} , which are nearly identical to the absorption features of peracetic acid, and were assigned on the basis of well acknowledged rules. The position of the absorption centred at 3303 cm^{-1} is identical to the peracetic acid spectrum. This absorption band is assigned to the strong OH stretching vibration, which is significantly shifted towards lower wavenumbers compared to free OH stretching vibrations of alcohols and acids, for instance. This is attributed to intramolecular H-bonding, which lowers the bond strength and subsequently the wavenumber of the absorption band. In the case of peracids, this results in the formation of

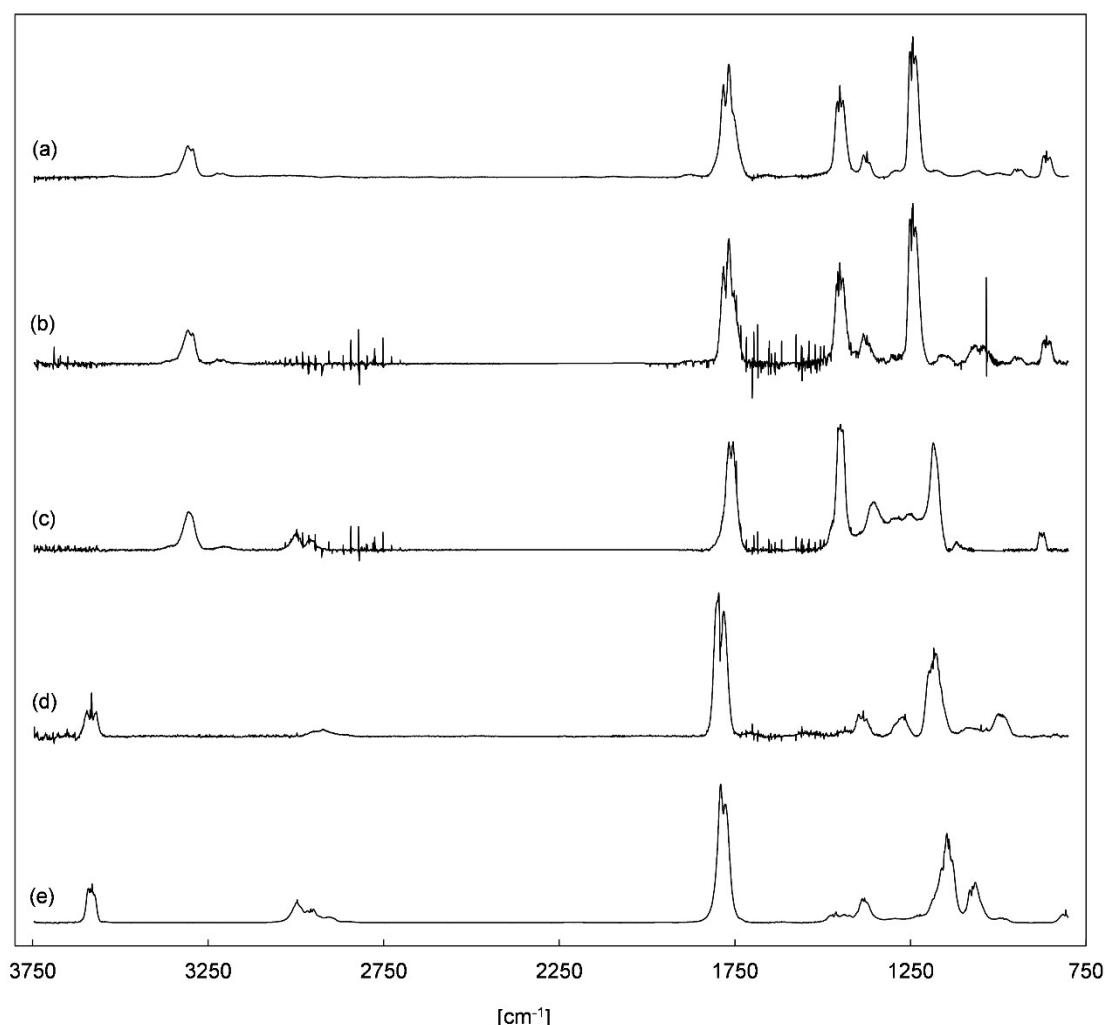


Fig. S4 (a) Reference spectrum of peracetic acid (peroxyacetic acid), (b) residual spectrum corresponding to peracetic acid generated in a $\text{CH}_3\text{C}(\text{O})\text{H}/\text{CH}_3\text{OH}/\text{Cl}$ mixture, (c) residual spectrum assigned to perpropionic acid generated in a $\text{C}_2\text{H}_5\text{C}(\text{O})\text{H}/\text{CH}_3\text{OH}/\text{Cl}$ mixture, (d) spectrum of acetic acid, and (e) spectrum of propionic acid. The spectra were straightened in the range $2400\text{--}1900\text{ cm}^{-1}$.

stable five-membered rings.¹⁴ This characteristic absorption is therefore an undoubted proof for the presence of a peracid. The residual spectrum contains an absorption band at 1450 cm^{-1} , again similar to the peracetic acid spectrum, which is assigned to the OH bending vibration. The absorption bands centred at 1760 cm^{-1} and 1180 cm^{-1} are attributed to the C=O and C-O stretching vibration. Both absorption features are shifted towards lower wavenumbers compared to the peracetic acid spectrum where a larger shift is observed for the C-O stretching vibration (from 1245 cm^{-1} to 1180 cm^{-1}). Trace (d) and trace (e) of Fig. S4 depict reference spectra of gas-phase acetic acid and propionic acid, those absorption bands show the same

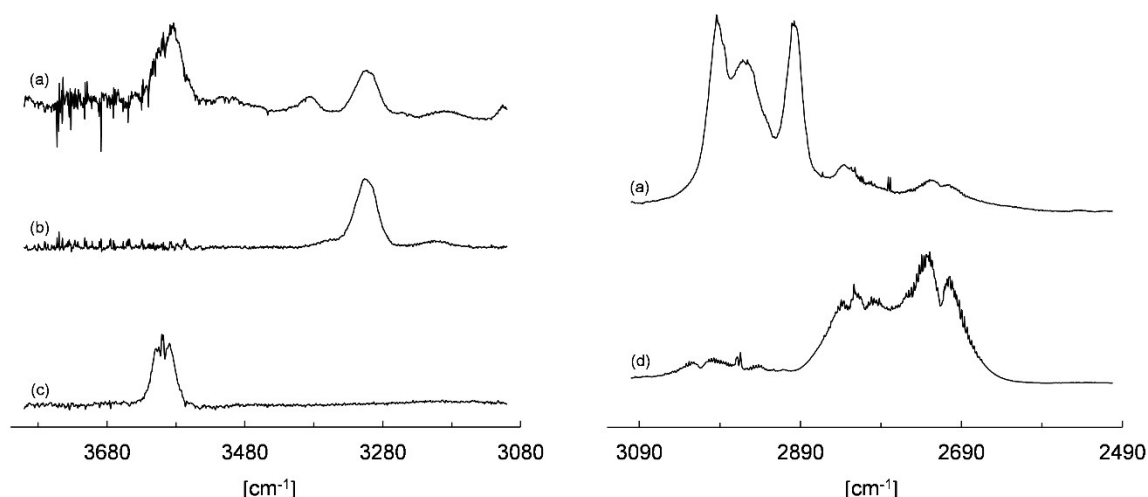


Fig. S5 (a) Spectrum of an EVK + O₃ experiment, collected after an EVK consumption of about 70 %, after subtraction of H₂O, EVK, HCHO, HC(O)OH, and 2-oxobutanal, and reference spectra of (b) perpropionic acid, (c) ethyl hydroperoxide, and (d) acetaldehyde, respectively.

behaviour for the C=O and C-O stretching vibration. Therefore, all these findings give confidence in attributing the absorption features of the residual spectrum to perpropionic acid.

Trace (a) of Fig. S5 shows residual absorption features from an evaluated ozonolysis experiment performed in the absence of SO₂. The spectrum was collected after an EVK consumption of about 70 %. H₂O, EVK, HCHO, HC(O)OH, and 2-oxobutanal were subtracted at this stage. Apparently, a product is formed with a clear absorption band centred on about 3303 cm⁻¹, which appears almost identical to the spectrum assigned to perpropionic acid (trace (b) of Fig. S5). In addition, the *m/z* 91 signal supports that perpropionic acid is formed, as discussed above. There is no obvious pathway forming acetyl peroxy radicals, which could finally result in the formation of peracetic acid. In addition, neither the PTR mass spectra nor other spectral ranges of the FTIR spectra support that peracetic acid is formed in the reaction system. All this are strong arguments that perpropionic acid is formed in the EVK + O₃ reaction.

D. Analysis of the RO₂ reactions

Perpropionic acid formation can solely be rationalized through the reaction of the propionyl peroxy radical (C₂H₅C(O)O₂) with HO₂. Up to now, this reaction has only been investigated by Hasson et al.¹³ who reported branching ratios of 0.35 ± 0.1 , 0.25 ± 0.1 , and 0.4 ± 0.1 for the pathways yielding perpropionic acid, propionic acid and ethyl peroxy radicals (C₂H₅O₂), respectively. The latter radical itself reacts with HO₂ to form ethyl hydroperoxide. The acetaldehyde formation can only proceed via RO₂ + RO₂ reactions. Both, the C₂H₅C(O)O₂ + C₂H₅O₂ reaction and the C₂H₅O₂ self-reaction produce eventually acetaldehyde. Since the rate coefficient of the self-reaction is 10² times smaller than that for the C₂H₅C(O)O₂ + C₂H₅O₂ reaction,^{15,16} the C₂H₅O₂ self-reaction is expectedly negligible in the present experiments (without SO₂). This is supported by the fact that neither FTIR spectra nor PTR mass spectra indicate the formation of ethanol, which is a specific product of the C₂H₅O₂ self-reaction.

The rate coefficient of the C₂H₅C(O)O₂ self-reaction is comparable to the RO₂ cross-reaction.¹⁵ Hence, the self-reaction increases the level of ethyl peroxy radicals in the experiments due to the conversion from C₂H₅C(O)O₂ into C₂H₅O₂ radicals. In the absence of SO₂, the acetaldehyde yield was found to be the same as the ethyl hydroperoxide yield within the assigned uncertainties. Therefore, about 50 % of the ethyl peroxy radicals react with HO₂, whereas the rest reacts with propionyl peroxy radicals.

About 20 % of C₂H₅C(O)O₂ + C₂H₅O₂ reaction results also in propionic acid, those formation was not observed in the absence of SO₂ due to the fast reaction of organic acids with sCI.

In order to prove if the temporal behaviour of acetaldehyde, ethyl hydroperoxide, propionic acid and perpropionic acid can be described solely through the known RO₂ reactions, the temporal evolution of the reaction products was simulated following the approach described recently in the literature.^{3,17} Table S3 summarizes the reaction sequences employed to describe the system. Model runs were performed assuming the EVK + O₃ reaction to form initially either C₂H₅C(O)O₂ or C₂H₅C(O)O₂ and C₂H₅O₂ radicals. Experimental and simulated time profiles for a representative experiment are shown in Fig. S6.

A major uncertainty of the simulations is [HO₂], given that the HO₂ concentration could not be determined experimentally. The value is adjusted mainly by reproducing concurrently the experimental time profiles of the peroxides. However, when only C₂H₅C(O)O₂ radicals are formed promptly from EVK + O₃, an estimation of the overall C₂H₅C(O)O₂ radical yield is given by the sum of the acetaldehyde, ethyl hydroperoxide and perpropionic acid yield. The

Table S3 Reaction sequence used for modelling the temporal behaviour of acetaldehyde, ethyl hydroperoxide, and perpropionic acid.

Reaction	Branching ratio	Rate coefficient ^a
(R9a) ^b CH ₂ =CHC(O)C ₂ H ₅ + O ₃ → C ₂ H ₅ C(O)O ₂		^c
(R9b) ^b CH ₂ =CHC(O)C ₂ H ₅ + O ₃ → C ₂ H ₅ O ₂		^c
(R10) C ₂ H ₅ C(O)O ₂ + HO ₂ → products		2.0 × 10 ⁻¹¹ ^d
(R10a) C ₂ H ₅ C(O)O ₂ + HO ₂ → C ₂ H ₅ C(O)OOH + O ₂	0.35 ^e	
(R10b) C ₂ H ₅ C(O)O ₂ + HO ₂ → C ₂ H ₅ C(O)OH + O ₃	0.25 ^e	
(R10c) C ₂ H ₅ C(O)O ₂ + HO ₂ (+ O ₂) → C ₂ H ₅ O ₂ + OH + O ₂ + CO ₂	0.40 ^e	
(R11) 2 C ₂ H ₅ C(O)O ₂ (+ O ₂) → 2 C ₂ H ₅ O ₂ + O ₂ + 2 CO ₂		1.7 × 10 ⁻¹¹ ^f
(R12) C ₂ H ₅ C(O)O ₂ + C ₂ H ₅ O ₂ → products		1.2 × 10 ⁻¹¹ ^f
(R12a) C ₂ H ₅ C(O)O ₂ + C ₂ H ₅ O ₂ (+ O ₂) → C ₂ H ₅ O ₂ + CO ₂ + CH ₃ C(O)H + HO ₂ + O ₂	0.8 ^f	
(R12b) C ₂ H ₅ C(O)O ₂ + C ₂ H ₅ O ₂ → C ₂ H ₅ C(O)OH + CH ₃ C(O)H + O ₂	0.2 ^f	
(R13) C ₂ H ₅ O ₂ + HO ₂ → C ₂ H ₅ OOH + O ₂		6.9 × 10 ⁻¹² ^f
(R14) 2 C ₂ H ₅ O ₂ → products		7.6 × 10 ⁻¹⁴ ^f
(R14a) 2 C ₂ H ₅ O ₂ (+ O ₂) → 2 CH ₃ C(O)H + 2 HO ₂ + O ₂	0.37 ^f	
(R14b) 2 C ₂ H ₅ O ₂ → CH ₃ C(O)H + C ₂ H ₅ OH + O ₂	0.63 ^f	

^a In [cm³ molecule⁻¹ s⁻¹], ^b either a prompt source of C₂H₅C(O)O₂ (R9a) or C₂H₅C(O)O₂ and C₂H₅O₂ radicals (R9a + R9b) was considered, ^c the radical source was included according to the experimental first-order loss of EVK, ^d the rate coefficient was assumed to be the same as the IUPAC recommended value for the CH₃C(O)O₂ + HO₂ reaction, ^e Hasson et al.²⁷, ^f IUPAC recommended values (2021).

temporal profiles of both acetaldehyde and ethyl hydroperoxide, whose quantification errors are much smaller than for perpropionic acid, were found to be very sensitive to the overall C₂H₅C(O)O₂ radical yield and [HO₂]. Though, adjusting both [HO₂] and the overall C₂H₅C(O)O₂ radical yield allows a simultaneous fit for all temporal profiles within the uncertainties of the experimentally determined product yields. On average, the experimental time profiles are reproduced using a C₂H₅C(O)O₂ radical yield of 0.39 and [HO₂] ≈ 2.3 × 10¹⁰ cm⁻³. Under these conditions propionic acid were formed with a yield of about 0.07, originating from C₂H₅C(O)O₂ + C₂H₅O₂ and C₂H₅C(O)O₂ + HO₂.

When a prompt source of C₂H₅O₂ radicals from the EVK + O₃ reaction is included, a match of all time profiles is achieved only for the lower accuracy range of the perpropionic acid data (Fig. S6). Accordingly, the overall C₂H₅C(O)O₂ radical yield is lower (about 0.30) than without considering a potentially prompt source of C₂H₅O₂ radicals, although in all model runs the C₂H₅O₂ source was found to be a factor of ≈ 5 smaller than the C₂H₅C(O)O₂ source. Under these conditions, the simulation generates a slightly lower propionic acid yield (0.05). For both scenarios, the simulations predict an ethanol level well below 1 ppbV at the end of each experiment. This, in turn, is below the detection limit of the instrument under the conditions used for the experiments.

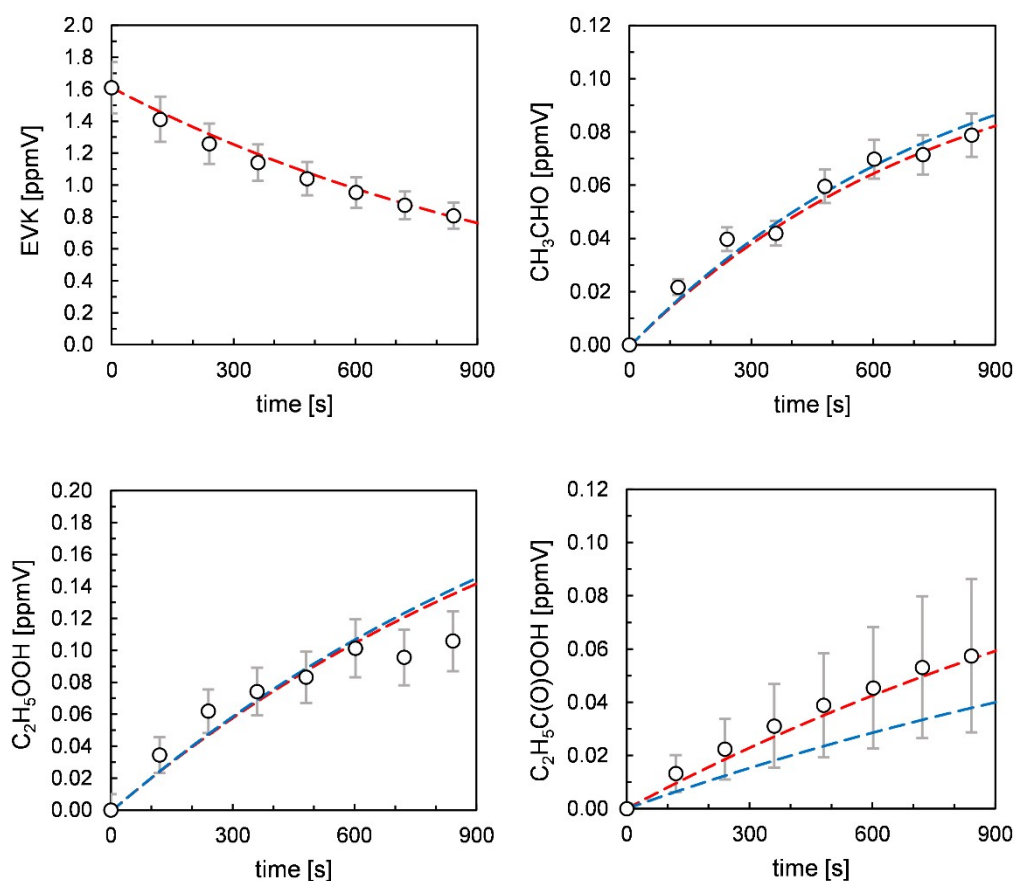


Fig. S6 Experimental (open circles) and simulated time profiles (dashed lines) of ethyl vinyl ketone (EVK), acetaldehyde (CH_3CHO), ethyl hydroperoxide ($\text{C}_2\text{H}_5\text{OOH}$), and perpropionic acid ($\text{C}_2\text{H}_5\text{C}(\text{O})\text{OOH}$) obtained from an experiment performed in the absence of SO_2 . The error bars represent the respective accuracy error. Since the addition of ozone cannot be simulated, $t = 0$ was set to the time, when the first FTIR spectrum was recorded after the ozone addition. Accordingly, the amount of products present in the first spectrum was subtracted from all data points. The simulations were performed assuming $[\text{HO}_2] = 2.7 \times 10^{10} \text{ cm}^{-3}$ and a $\text{C}_2\text{H}_5\text{C}(\text{O})\text{O}_2$ yield of 0.42 (scenario 1, red) or $[\text{HO}_2] = 2.2 \times 10^{10} \text{ cm}^{-3}$, a $\text{C}_2\text{H}_5\text{C}(\text{O})\text{O}_2$ yield of 0.32 and a $\text{C}_2\text{H}_5\text{O}_2$ yield of 0.07 (scenario 2, blue).

Although this modelling approach is rather uncertain, since the major criticism - that $[\text{HO}_2]$ is actually not known - remains, there is one important observation to be made. The fact that for this type of experiments the temporal evolution of acetaldehyde, ethyl hydroperoxide and perpropionic acid is reproduced with the same set of parameters, which are experimentally plausible, suggests that there is no reason to argue against their time profiles being properly described by the peroxy radical reaction sequence as listed in Table S3. At the same time it is not possible to conclude whether the ethyl peroxy radical evolves solely from the propionyl peroxy radical further chemistry or also promptly from the CI chemistry. Nevertheless, the present results hint at the propionyl peroxy radical being the dominant RO_2 .

Since in the presence of SO₂ no hydroperoxides could be quantified, this is an indication that the RO₂ further chemistry is dominated by RO₂ + RO₂ reactions in this type of experiment. Although all possible RO₂ + RO₂ reactions shift the reaction system towards acetaldehyde formation, the acetaldehyde yield is about a factor of 2 smaller in the presence of SO₂ than the sum of the acetaldehyde, ethyl hydroperoxide and perpropionic acid yield in the absence of SO₂. This indicates a much lower RO₂ level when the sCIs are scavenged with SO₂ and hence their origin from both chemically activated and stabilized CIs. However, in this set-up it is not possible to decipher precisely between the contribution of the C₂H₅O₂ self-reaction and the C₂H₅C(O)O₂ + C₂H₅O₂ reaction towards acetaldehyde formation. Since, contrary to ethyl hydroperoxide, the formation of perpropionic acid in the presence of SO₂ is at least suggested by the FTIR data, this might argue that the propionyl peroxy radical remains the dominant RO₂ radical. Further, the formation of ethanol, following the C₂H₅O₂ self-reaction, was not supported by the FTIR spectra in any of the experiments.

Due to the absence of quantifiable amounts of hydroperoxides, we were not able to adjust [HO₂] in the model as we did for the experiments without SO₂ added. However, we used the model to figure out the propionic acid/acetaldehyde ratio by variation of the RO₂ yield and [HO₂] considering the fact that an acetaldehyde yield of about 15 % was observed experimentally. Using the average RO₂ yields found for the experiments in the absence of SO₂, we obtained an upper limit of 0.5 for the propionic acid/acetaldehyde ratio considering the uncertainties on the acetaldehyde yield. Lowering the RO₂ yield in the simulation requires a lower HO₂ concentration in order to match the acetaldehyde yield. This, in turn, decreases the propionic acid/acetaldehyde ratio. At the same time, an increase in the C₂H₅O₂/C₂H₅C(O)O₂ ratio (scenario 2) lowers also the propionic acid/acetaldehyde ratio. Apparently, the simulations suggest the propionic acid/acetaldehyde ratio to be well below the experimentally observed ratio (1.1 ± 0.2) evidencing an additional source of propionic acid in the reaction system.

E. Error analysis

The experimental results suggest a 1:1 ratio for the decomposition of the primary ozonide. However, the sum of the primary carbonyls is well below 100 %. It appears that in the present investigation the POZ decomposition accounts solely for about 57 ± 14 % of the EVK + O₃ reaction. We did not find any hint for products suggesting pathways competitive to the POZ formation, e. g. the formation of an epoxide. At the same time, the increase of the HCHO + 2-oxobutanal yield when SO₂ is added to the system is larger than suggested from the SO₂

consumption. While a smaller increase appears plausible once the $s\text{CI} + \text{SO}_2$ reaction exhibits additional pathways (e. g. the formation of stable $s\text{CI-SO}_2$ adducts partitioning into the aerosol phase), the inverse case suggests rather quantification errors or secondary processes not considered in the analysis.

One might intuitively think that this indicates an overestimation of the EVK consumption. Though, even after re-analysis of the calibration data and additional control experiments there is no indication for an erroneous infrared cross section. A hypothetically secondary loss process might be the reaction of EVK with $s\text{CIs}$, lowering the product yields in the absence of SO_2 . At least for the structural analogue methyl vinyl ketone (MVK) it was shown that, under experimental conditions similar to the present study, the observed kinetics of $\text{MVK} + \text{O}_3$ was not affected by the reaction of the unsaturated ketone with CIs .¹⁸ It is therefore reasonable to assume that $\text{EVK} + \text{O}_3$ exhibits the same behaviour.

Our HCHO absorption cross section is in excellent agreement with literature data. The high linearity of the yield plots (Fig. S1) rules out an underestimation of the HCHO yield due to a potential wall loss. Alternatively, a conceivable loss process of HCHO might be its reaction with HO_2 (R15),^{19,20} which is known to become likely relevant under higher radical concentrations present in chamber experiments.



The HOCH_2O_2 further chemistry might evolve, for instance, into formic acid (HC(O)OH) formation via the RO_2 self-reaction or the reaction with HO_2 . In principle, the level of HO_2 is reasonably high mainly due to the conversion of OH into HO_2 by carbon monoxide.

When we simulate the temporal evolution of HCHO for a simplified reaction system observing the reactions (R15), (R16), $\text{HOCH}_2\text{O}_2 + \text{HOCH}_2\text{O}_2$ and $\text{HOCH}_2\text{O}_2 + \text{HO}_2$, it is found that HO_2 levels about a factor of 40–50 larger than concluded in Sect. D are necessary in order to observe a measurable influence on HCHO. Although the value obtained for $[\text{HO}_2]$ provides just a rough estimate, a factor of 40–50 is far beyond the uncertainties and experimentally implausible. The observed HCHO yield is therefore not expectedly affected by a secondary consumption of HCHO by HO_2 .

Assuming HCHO to be correct, the 2-oxobutanal yield would be underestimated by a factor of about 2.5 in order to match the 100 % primary carbonyl yield in the absence of SO_2 . A 20 % error, which we considered in our error calculation, appears reasonable due to the semi-

quantitative approach we used to quantify 2-oxobutanal. However, it is physically not plausible that the cross sections of the carbonyl absorptions of 2-oxobutanal and methyl glyoxal diverge by a factor of 2.5. At the same time, the linearity of the yield plots (Fig. S1) rules out a large influence of wall losses on the 2-oxobutanal yield.

Overall, we do not find a conclusive explanation for the lower than expected primary carbonyl yields in the absence of SO₂. Since quantification errors were ruled out, we can only imagine a secondary process consuming the primary carbonyls in the absence of SO₂. Further investigations are required to identify this process.

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