Contents

1	General Synthetic Aspects	
2	Analytical Instruments and Details 3	
3	Chemicals	
4	Syntheses and Analytics	
	4.1 [EMIm] salts	
	4.2 [BMPL] salts	
	4.3 [BnPh ₃ P] salts	
5	NMR spectra 23	
	5.1 [EMIm] salts	
	5.2 [BMPL] salts	
	5.3 Phosphonium salts	
6	IR and Raman Spectra	
	6.1 [EMIm] salts	
	6.2 [BMPL] salts	
	6.3 Phosphonium salts	
7	Electrochemical Measurements101	
8	Viscosity and Density102	
9	Vogel-Fulcher-Tammann (VFT), Arrhenius, and Litovitz Fits103	
10 Diffusion NMR105		
11	11 Overview physical propterties of RTILs111	
12	12 Cytotoxicity	
13	Thermal Properties	
	13.1 [EMIm] salts	
	13.2 [BMPL] salts	
	13.3 [BnPh ₃ P] salts	
14	Quantum Chemical Calculations124	
15	Single Crystal X-Ray Diffraction125	

.6 Literature

1 General Synthetic Aspects

Reactions with air- and moisture-sensitive compounds were performed in round-bottom flasks or glass tubes equipped with valves with PTFE stems (Rettberg, Göttingen) under argon using Schlenk-line techniques.

2 Analytical Instruments and Details

¹H, ²H, ¹¹B, ¹³C, ¹⁹F and ³¹P spectra were recorded at 25 °C in (CD₃)₂CO on a Bruker Avance 500 NMR spectrometer, a Bruker Avance Neo I 600 or a Bruker Avance Neo 400 spectrometer. NMR signals were referenced against (CH₃)₄Si (¹H and ¹³C) with Ξ (¹³C) = 25.145020 MHz, (CD₃)₄Si with Ξ (²H) = 15.350609 MHz, BF₃·OEt₂ in CDCl₃ with Ξ (¹¹B) = 32.083974 MHz, CFCl₃ with Ξ (¹⁹F) = 94.094011 MHz, H₃PO₄ with Ξ (³¹P) = 40.480742 as external standard.^[1] ¹H and ¹³C chemical shifts were calibrated against the residual solvent signal and the solvent signal, respectively (δ (¹H): (CD₂H)(CD₃)CO 2.05 ppm; δ (¹³C): (CD₃)₂CO 206.26 and 29.84 ppm.^[2] IR spectra were recorded at room temperature with a Bruker Alpha FT IR spectrometer with an apodized resolution of 4 cm⁻¹ in the attenuated total reflection (ATR) mode in the region of 4000–400 cm⁻¹ using a setup with a diamond crystal. Raman spectra were recorded at room temperature with a mapodized resolution of 4 cm⁻¹ using the 1064 nm line of a Nd/YAG laser for excitation of the crystalline samples contained in melting point capillaries.

Elemental analyses (C, H, N) were performed either with a Euro EA3000 instrument (HEKA-Tech, Germany) or with an Elementar Vario MICRO cube instrument (Elementar Analysensysteme, Germany). Thermal analyses were performed with a DSC 204 F1 Phoenix (Netzsch) in the temperature range of -175 to 580 °C with a heating rate of 10 K·min⁻¹. Viscosities and densities were measured with a rolling-ball viscosimeter Lovis 2000 ME combined with a DMA 4100 M density meter (Anton Paar) at different angles and temperatures. The water content of the ILs was determined by Karl-Kischer titration with a Metrohm 831 KF Coulometer. The water content of the ILs was <50 ppm.

3 Chemicals

All standard chemicals were obtained from commercial sources and used without further purification. Solvents were dried according to standard protocols^[3] and stored under argon atmosphere in flasks with valves with PTFE stems (Rettberg, Göttingen).

 $K[B(CN)_4]^{[4]}$, $K[BF_3(CN)]^{[4]}$, $K[BF_2(CN)_2]^{[4]}$, $K[BF(CN)_3]^{[5]}$, $K[BFH(CN)_2]^{[6]}$, $K[BH_2(CN)_2]^{[7]}$, and $K[BH(CN)_2]^{[7]}$ were prepared according to known procedures.

4 Syntheses and Analytics

4.1 [EMIm] salts

4.1.1 [EMIm][BF₄]

[EMIm][BF₄] was purchased from Sigma-Aldrich.

¹H NMR (500.1 MHz, (CD₃)₂CO) $\delta = 8.91$ (s, 1H, NCHN), 7.73 (dd, 1H, ³J(¹H,¹H) \approx ⁴J(¹H,¹H) \approx 1.8 Hz, N–CH=CH–N), 7.65 (dd, 1H ³J(¹H,¹H) \approx ⁴J(¹H,¹H) \approx 1.8 Hz, N–CH=CH–N), 4.35 (q, 2H, ³J(¹H,¹H) = 7.3 Hz, CH₂CH₃), 4.00 (s, 3H, CH₃), 1.53 ppm (t, 3H, ³J(¹H,¹H) = 7.3 Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 137.2 (s, 1C, *C*H), 124.6 (s, 1C, *C*H), 122.9 (s, 1C, *C*H), 45.6 (s, 1C, *C*H₂CH₃), 36.4 (s, 1C, *C*H₃), 15.5 ppm (s, 1C, CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -0.9 ppm (s, 1B, *B*F₄).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -0.9 ppm (s, 1B, *B*F₄).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -150.88 (s, 4F, (~20%) ¹⁰BF₄) -150.94 ppm (q, 4F (~80%), ¹J(¹⁹F, ¹¹B) = 0.7 Hz, ¹¹BF₄).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -150.88 (s, 4F, (~20%) ¹⁰BF₄) -150.94 ppm (s, 4F (~80%), ¹¹BF₄).

Elemental analysis calcd (%) for C₆H₁₁BF₄N₂: C 36.40, H 5.60, N 14.15; found: C 35.65, H 5.86, N 14.51.

4.1.2 [EMIm][BF₃(CN)]

K[BF₃CN] (2.66 g, 20.0 mmol) was dissolved in water (25 ml) and a solution of [EMIm]Cl (3.0 g, 20.46 mmol) in water (4.3 mL) was added. The solution was then extracted with CH_2Cl_2 (5×4 mL) and the combined organic layers were dried with MgSO₄, filtered and the CH_2Cl_2 was removed in vacuum. The remaining ionic liquid was dissolved in CH_2Cl_2 (2.5 mL) and the solution was washed with deionized water (3×0.5 mL) and with bidistilled water (3×0.5 mL) and then dried in a fine vacuum.

Yield: 2.91 g, 14.20 mmol [EMIm][BF₃(CN)] (71%).

¹H NMR (500.1 MHz, (CD₃)₂CO) $\delta = 8.97$ (s, 1H, NCHN), 7.75 (dd, 1H, ³J(¹H,¹H) \approx ⁴J(¹H,¹H) \approx 1.8 Hz, N-CH=CH-N), 7.68 (dd, 1H ³J(¹H,¹H) \approx ⁴J(¹H,¹H) \approx 1.8 Hz, N-CH=CH-N), 4.39 (q, 2H, ³J(¹H,¹H) = 7.3 Hz, CH₂CH₃), 4.04 (s, 3H, CH₃), 1.56 ppm (t, 3H, ³J(¹H,¹H) = 7.3 Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 137.1 (s CH), 132.2 (m, BF₃(CN)) 124.7 (s, CH), 123.0 (s, CH), 45.7 (s, CH₂CH₃), 36.6 (s, CH₃), 15.6 ppm (s CH₂CH₃).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 137.1 (d, ¹J(¹³C,¹H) = 224.8 Hz, *C*H), 132.2 (q, ¹J(¹³C,¹¹B) = 88.7 Hz, BF₃(*C*N)) 124.7 (d, ¹J(¹³C,¹H) = 206.6 Hz, *C*H), 123.0 (d, ¹J(¹³C,¹H) = 209.9 Hz, *C*H), 45.7 (t, ¹J(¹³C,¹H) = 144.5 Hz, *C*H₂CH₃), 36.6 (q, ¹J(¹³C,¹H) = 143.8 Hz, *C*H₃), 15.6 ppm (qt, ¹J(¹³C,¹H) = 128.8 Hz, ²J(¹³C,¹H) = 3.8 Hz, CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -3.55 ppm (q, 1B, ¹*J*(¹⁹F, ¹¹B) = 27.0 Hz, *B*F₃(CN)).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -3.55 ppm (s, 1B, *B*F₃(CN)), ¹*J*(¹³C, ¹¹B) = 88.7 Hz, (¹³C-satelites).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -137.57 (spt, ¹J(¹⁹F,¹⁰B) = 9.1 Hz, ¹⁰BF₃(CN)), (-137.63 ppm (q, 3F, ¹J(¹⁹F,¹¹B) = 26.9 Hz, ¹¹BF₃(CN)).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -137.57 (spt, ¹J(¹⁹F, ¹⁰B) = 9.1 Hz, ¹⁰BF₃(CN)), -137.63 ppm (s, 3F, ¹¹BF₃(CN)).

IR (ATR): 2209 cm⁻¹ (ν (C \equiv N)).

Raman: 2211 cm⁻¹ (ν (C \equiv N)).

Elemental analysis calcd (%) for C₇H₁₁BF₃N₃: C 41.01, H 5.41, N 20.50; found: C 40.81, H 5.37, N 21.30.

4.1.3 [EMIm][BF₂(CN)₂]

Na[BF₂(CN)₂] (4.7 g, 37.96 mmol) was treated with [EMIm]Cl (5.9 g, 40.24 mmol) in water (15 mL). The suspension was then extracted with CH_2Cl_2 (60 mL) and the CH_2Cl_2 phase was washed with bidistilled water (5×1 mL). All volatiles were removed in a vacuum and the remaining ionic liquid was dried in a fine vacuum.

Yield: 4.9 g, 23.11 mmol [EMIm][BF₂(CN)₂] (61%).

¹H NMR (500.1 MHz, (CD₃)₂CO) $\delta = 8.94$ (s, 1H, NCHN), 7.71 (dd, 1H, ³J(¹H,¹H) $\approx {}^{4}J({}^{1}H,{}^{1}H) \approx 1.8$ Hz, N-CH=CH-N), 7.64 (dd, 1H ${}^{3}J({}^{1}H,{}^{1}H) \approx {}^{4}J({}^{1}H,{}^{1}H) \approx 1.8$ Hz, N-CH=CH-N), 4.37 (q, 2H, ${}^{3}J({}^{1}H,{}^{1}H) = 7.3$ Hz, CH₂CH₃), 4.03 (s, 3H, CH₃), 1.56 ppm (t, 3H, ${}^{3}J({}^{1}H,{}^{1}H) = 7.3$ Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.9 (s, *C*H), 130.5 (qt, ¹*J*(¹³C,¹¹B) = 80.7 Hz, ²*J*(¹⁹F,¹³C) = 50.4 Hz, BF₂(*C*N)₂), 124.6 (s, *C*H), 122.9 (s, *C*H), 45.6 (s, *C*H₂CH₃), 36.5 (s, *C*H₃), 15.4 ppm (s, CH₂CH₃).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.9 (d, ¹J(¹³C,¹H) = 221.1 Hz, *C*H), 130.5 (q, ¹J(¹³C,¹¹B) = 80.6 Hz, BF₂(*C*N)₂) 124.6 (d, ¹J(¹³C,¹H) = 206.2 Hz, *C*H), 122.9 (d, ¹J(¹³C,¹H) = 209.0 Hz, *C*H), 45.6 (t, ¹J(¹³C,¹H) = 144.4 Hz, *C*H₂CH₃), 36.5 (q, 1C, ¹J(¹³C,¹H) = 143.9 Hz, *C*H₃), 15.4 ppm (qt, 1C, ¹J(¹³C,¹H) = 128.8 Hz, ²J(¹³C,¹H) = 3.7 Hz, CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -7.3 ppm (t, 1B, ¹*J*(¹⁹F, ¹¹B) = 41.1 Hz, *B*F₂(CN)₂).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -7.3 ppm (s, 1B, BF₂(CN)₂).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -153.9 (spt, 1F (~20%), ¹*J*(¹⁹F,¹⁰B) = 13.8 Hz, ¹⁰B*F*₂(CN)₂), -154.0 ppm (q, 1F (~80%), ¹*J*(¹⁹F,¹¹B) = 41.2 Hz, ¹¹B*F*₂(CN)₂).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -153.9 (spt, 1F (~20%), ¹J(¹⁹F,¹⁰B) = 13.8 Hz, ¹⁰BF₂(CN)₂), -154.0 ppm (s, 1F (~80%), ¹¹BF₂(CN)₂).

IR (ATR): 2210 cm⁻¹ (ν (C \equiv N)).

Raman: 2212 cm⁻¹ (ν (C \equiv N)).

Elemental analysis calcd (%) for C₈H₁₁BF₂N₄: C 45.32, H 5.23, N 26.43; found: C 44.96, H 5.35, N 26.72.

4.1.4 [EMIm][BF(CN)₃]

 $K[BF(CN)_3]$ (75 g, 0.51 mol) was dissolved in water (150 mL) and [EMIm]Cl (76 g, 0.52 mol) and CH_2Cl_2 (100 mL) were added. The organic layer was separated and washed with deionized water (5×10 mL) and then with bidistilled water (5×5 mL). The CH_2Cl_2 was removed with a rotary evaporator and the remained ionic liquid was dried in a fine vacuum.

Yield: 110.1 g, 0.503 mol [EMIm][BF(CN)₃] (98%)

¹H NMR (500.1 MHz, (CD₃)₂CO) $\delta = 8.97$ (s, 1H, NCHN), 7.72 (dd, 1H, ³J(¹H,¹H) $\approx {}^{4}J({}^{1}H,{}^{1}H) \approx 1.6$ Hz, N–CH=CH–N), 7.65 (dd, 1H ${}^{3}J({}^{1}H,{}^{1}H) \approx {}^{4}J({}^{1}H,{}^{1}H) \approx 1.6$ Hz, N–CH=CH–N), 4.38 (q, 2H, ${}^{3}J({}^{1}H,{}^{1}H) = 7.3$ Hz, CH₂CH₃), 4.04 (s, 3H, CH₃), 1.57 ppm (t, 3H, ${}^{3}J({}^{1}H,{}^{1}H) = 7.3$ Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.9 (s, *C*H), 127.6 (qd, ¹*J*(¹³C, ¹¹B) = 74.7 Hz, ²*J*(¹⁹F, ¹³C) = 37.1 Hz, BF(*C*N)₃), 124.6 (s, *C*H), 122.9 (s, *C*H), 45.7 (s, *C*H₂CH₃), 36.5 (s, *C*H₃), 15.5 ppm (s, CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -17.8 ppm (d, 1B, ¹*J*(¹⁹F, ¹¹B) = 44.6 Hz, *B*F(CN)₃).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -17.8 ppm (s, 1B, *B*F(CN)₃).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -211.9 ppm (q, 1F, ¹J(¹⁹F, ¹¹B) = 44.6 Hz, BF(CN)₃).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -211.9 ppm (s, 1F, B*F*(CN)₃).

IR (ATR): 2214 cm⁻¹ (ν (C \equiv N)).

Raman: 2217 cm⁻¹ (ν (C \equiv N)).

Elemental analysis calcd (%) for C₉H₁₁BFN₅: C 49.35, H 5.06, N 31.98; found: C 48.85, H 5.07, N 32.05.

4.1.5 [EMIm][BFH(CN)₂]

 $K[BFH(CN)_2]$ (4.00 g, 28.0 mmol) was treated with a solution of [EMIm]Cl (5.05 g, 34.4 mmol) in water (15 mL). The mixture was stirred for 15 min and then extracted with CH_2Cl_2 (100 mL). The organic layer was washed with bidistilled water (5×3 mL) and all volatiles were removed in a vacuum. The remaining ionic liquid was dried in a fine vacuum, washed again with bidistilled water (5×1 mL) and finally dried in a fine vacuum.

Yield: 4.92 g, 25.36 mmol [EMIm][BFH(CN)₂] (77%).

¹H NMR (500.1 MHz, (CD₃)₂CO) $\delta = 8.94$ (s, 1H, NCHN), 7.72 (dd, 1H, ³J(¹H,¹H) \approx ⁴J(¹H,¹H) \approx 1.7 Hz, N–CH=CH–N), 7.65 (dd, 1H ³J(¹H,¹H) \approx ⁴J(¹H,¹H) \approx 1.7 Hz, N–CH=CH–N), 4.38 (q, 2H, ³J(¹H,¹H) = 7.3 Hz, CH₂CH₃), 4.03 (s, 3H, CH₃), 2.73 (qd, 1H,

 ${}^{1}J({}^{11}B,{}^{1}H) = 109.6 \text{ Hz}, {}^{2}J({}^{19}F,{}^{1}H) = 40.8 \text{ Hz}, \text{ BF}H(CN)_{2}), 1.56 \text{ ppm} (t, 3H, {}^{3}J({}^{1}H,{}^{1}H) = 7.3 \text{ Hz}, CH_{2}CH_{3}).$

¹H{¹¹B} NMR (500.1 MHz, (CD₃)₂CO) $\delta = 8.94$ (s, 1H, NCHN), 7.72 (dd, 1H, ³J(¹H, ¹H) \approx ⁴J(¹H, ¹H) \approx 1.7 Hz, N–CH=CH–N), 7.65 (dd, 1H ³J(¹H, ¹H) \approx ⁴J(¹H, ¹H) \approx 1.7 Hz, N–CH=CH–N), 4.38 (q, 2H, ³J(¹H, ¹H) = 7.3 Hz, CH₂CH₃), 4.03 (s, 3H, CH₃), 2.73 (d, 1H, ²J(¹⁹F, ¹H) = 40.8 Hz, BFH(CN)₂), 1.56 ppm (t, 3H, ³J(¹H, ¹H) = 7.3 Hz, CH₂CH₃).

¹H{¹⁹F} NMR (400.3 MHz, (CD₃)₂CO) $\delta = 8.94$ (s, 1H, NCHN), 7.72 (dd, 1H, ³J(¹H, ¹H) \approx ⁴J(¹H, ¹H) \approx 1.7 Hz, N–CH=CH–N), 7.65 (dd, 1H ³J(¹H, ¹H) \approx ⁴J(¹H, ¹H) \approx 1.7 Hz, N–CH=CH–N), 4.38 (q, 2H, ³J(¹H, ¹H) = 7.3 Hz, CH₂CH₃), 4.03 (s, 3H, CH₃), 2.73 (q, 1H, ¹J(¹¹B, ¹H) = 109.7 Hz, BFH(CN)₂), 1.56 ppm (t, 3H, ³J(¹H, ¹H) = 7.3 Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.9 (s, *C*H), 132.6 (qd, ¹J(¹³C,¹¹B) = 64.8 Hz, ²J(¹⁹F,¹³C) = 35.3 Hz, BFH(*C*N)₂), 124.6 (s, *C*H), 122.9 (s, *C*H), 45.6 (s, *C*H₂CH₃), 36.5 (s, *C*H₃), 15.5 ppm (s, CH₂CH₃).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.9 (d, ¹J(¹³C,¹H) = 222.2 Hz CH), 132.6 (q, ¹J(¹³C,¹¹B) = 64.8 Hz, BFH(CN)₂), 124.7 (d, ¹J(¹³C,¹H) = 206.0 Hz, CH), 123.0 (d, ¹J(¹³C,¹H) = 203.1 Hz, CH), 45.7 (t, ¹J(¹³C,¹H) = 144.3 Hz, CH₂CH₃), 36.6 (q, ¹J(¹³C,¹H) = 143.8 Hz, CH₃), 15.5 ppm (qt, ¹J(¹³C,¹H) = 128.8 Hz, ²J(¹³C,¹H) = 3.7 Hz, CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -14.2 ppm (dd, 1B, ¹J(¹¹B,¹H) = 109.8 Hz, ¹J(¹⁹F,¹¹B) = 56.0 Hz, *B*FH(CN)₂).

¹¹B{¹H} NMR (160.5 MHz, (CD₃)₂CO) δ = -14.2 ppm (d, 1B, ¹*J*(¹⁹F,¹¹B) = 56.0 Hz, *B*FH(CN)₂).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -14.2 ppm (d, 1B, ¹*J*(¹¹B, ¹H) = 109.8 Hz, *B*FH(CN)₂).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -233.9 ppm (qd, 1F, ¹J(¹⁹F,¹¹B) = 56.0 Hz, ²J(¹⁹F,¹H) = 40.8 Hz B*F*H(CN)₂).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -233.9 ppm (d, 1F, ²J(¹⁹F, ¹H) = 40.8 Hz BFH(CN)₂).

¹⁹F{¹H} NMR (377 MHz, (CD₃)₂CO) δ = -234.1 ppm (q, 1F, ¹J(¹⁹F, ¹¹B) = 56.0 Hz, BFH(CN)₂).

IR (ATR): 2375 (v(B−H), 2189 cm⁻¹ (v(C≡N)).

Raman: 2383 (v(B−H), 2201 cm⁻¹ (v(C≡N)).

Elemental analysis calcd (%) for C₈H₁₂BFN₄: C 49.52, H 6.23, N 28.88; found: C 49.84, H 5.96, N 28.82.

4.2 [BMPL] salts

4.2.1 [BMPL][BF₃(CN)]

 $K[BF_3CN]$ (1.33 g, 10.0 mmol) was dissolved in water (25 ml) and a solution of [BMPL]Cl (1.78 g, 10.03 mmol) was added. The solution was extracted with CH_2Cl_2 (5×4 mL) and the combined organic layers were dried with MgSO₄, filtered and the CH_2Cl_2 was removed in vacuum. The remaining ionic liquid was dried in a fine vacuum.

Yield: 1.21 g, 5.125 mmol [BMPL][BF₃(CN)] (51%).

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 3.75-3.65 (m, 4H, CH₂), 3.54-3.51 (m, 2H, CH₂), 3.23 (s, 3H, CH₃), 2.35-2.27 (m, 4H, CH₂), 1.93-1.86 (m, 2H, CH₂), 1.47-1.40 (m, 2H, CH₂CH₃), 0.98 ppm (t, 3H, ³J(¹H, ¹H) = 7.41 Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 132.2 (m, BF₃(*C*N)), 65.2 (t, ¹*J*(¹⁴N, ¹³C) = 3.1 Hz, NCH₂), 65.0 (t, ¹*J*(¹⁴N, ¹³C) = 3.0 Hz, NCH₂), 48.9 (t, ¹*J*(¹⁴N, ¹³C) = 4.1 Hz, NCH₃), 26.3 (s, *C*H₂), 22.3 (s, CH₂), 20.4 (t, *J*(¹⁴N, ¹³C) = 1.60 Hz, *C*H₂CH₃) 13.8 ppm (s, CH₂CH₃).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 132.2 (q, ¹J(¹³C, ¹¹B) = 89.5 Hz, BF₃(CN)), 65.2 (t, ¹J(¹³C, ¹H) = 145.0 Hz, NCH₂), 65.0 (t, ¹J(¹³C, ¹H) = 142.2 Hz, NCH₂), 48.9 (qm, ¹J(¹³C, ¹H) = 143.6 Hz, NCH₃), 26.3 (t, ¹J(¹³C, ¹H) = 129.3 Hz, CH₂), 22.3 (t, ¹J(¹³C, ¹H) = 135.5 Hz CH₂), 20.6 (t, ¹J(¹³C, ¹H) = 127.5 Hz, CH₂CH₃) 15.8 ppm (qm, 1C, ¹J(¹³C, ¹H) = 125.6 Hz, CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -3.56 ppm (q, 1B, ¹*J*(¹⁹F, ¹¹B) = 26.9 Hz, *B*F₃(CN)).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -3.56 ppm (s, 1B, *B*F₃(CN)), ¹*J*(¹³C, ¹¹B) = 89.6 Hz, (¹³C-satelites).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -137.68 (spt, ¹*J*(¹⁹F,¹⁰B) = 8.8 Hz, ¹⁰BF₃(CN)), -137.75 ppm (q, 3F, ¹*J*(¹⁹F,¹¹B) = 27.1 Hz, ¹¹BF₃(CN)).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -137.68 (spt, ¹J(¹⁹F,¹⁰B) = 8.8 Hz, ¹⁰BF₃(CN)), -137.75 ppm (s, 3F, ¹¹BF₃(CN)).

IR (ATR): 2207 cm⁻¹ (v(C≡N)).

Raman: 2209 cm⁻¹ (*v*(C≡N)).

Elemental analysis calcd (%) for C₁₀H₂₀BF₃N₂: C 50.87, H 8.54, N 11.87; found: C 50.70, H 8.36, N 12.70.

4.2.2 [BMPL][BF₂(CN)₂]

 $Na[BF_2(CN)_2]$ (3.2 g, 25.85 mmol) was treated with [BMPL]Cl (4.75 g, 26.7 mmol) in water (15 mL). The suspension was then extracted with CH_2Cl_2 (60 mL) and the CH_2Cl_2 phase was washed with bidistilled water (5×1 mL). All volatiles were removed in a vacuum and the resulting ionic liquid was dried in a fine vacuum.

Yield: 4.65 g, 19.13 mmol [BMPL][BF₂(CN)₂] (74%).

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 3.74-3.63 (m, 4H, CH₂), 3.54-3.48 (m, 2H, CH₂), 3.22 (s, 3H, CH₃), 2.36-2.27 (m, 4H, CH₂), 1.93-1.86 (m, 2H, CH₂), 1.47-1.39 (m, 2H, CH₂CH₃), 0.98 ppm (t, 3H, ³J(¹H, ¹H) = 7.41 Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 130.7 (qt, ¹J(¹³C, ¹¹B) = 80.6 Hz, ²J(¹⁹F, ¹³C) = 50.3 Hz, BF₂(CN)₂), 65.1 (t, ¹J(¹⁴N, ¹³C) = 3.3 Hz, NCH₂), 65.0 (t, ¹J(¹⁴N, ¹³C) = 3.0 Hz, NCH₂), 48.9 (t, ¹J(¹⁴N, ¹³C) = 4.0 Hz, NCH₃), 26.2 (s, CH₂), 22.3 (s, CH₂), 20.3 (t, J(¹⁴N, ¹³C) = 1.54 Hz, CH₂CH₃) 13.7 ppm (s, CH₂CH₃).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 130.7 (q, ¹J(¹³C, ¹¹B) = 80.6 Hz, ¹¹BF₂(*C*N)₂), 130.6 (spt, ¹J(¹³C, ¹⁰B) = 27.3 Hz, ¹⁰BF₂(*C*N)₂), 65.1 (t, ¹J(¹³C, ¹H) = 148.0 Hz, NCH₂), 64.9 (t, ¹J(¹³C, ¹H) = 143.3 Hz, NCH₂), 48.9 (qm, ¹J(¹³C, ¹H) = 143.8 Hz, NCH₃), 26.2 (t, ¹J(¹³C, ¹H) = 127.6 Hz, CH₂), 22.3 (t, ¹J(¹³C, ¹H) = 137.6 Hz CH₂), 20.3 (t, ¹J(¹³C, ¹H) = 127.5 Hz, CH₂CH₃) 13.7 ppm (qm, 1C, ¹J(¹³C, ¹H) = 125.4 Hz, CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -7.3 ppm (t, 1B, ¹*J*(¹⁹F, ¹¹B) = 41.2 Hz, *B*F₂(CN)₂).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -7.3 ppm (s, 1B, BF₂(CN)₂).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -153.8 ppm (spt, 1F, (~20%), ¹J(¹⁹F,¹⁰B) = 13.8 Hz, ¹⁰BF₂(CN)₂), -153.9 ppm (q, 1F (~80%), ¹J(¹⁹F,¹¹B) = 41.2 Hz, ¹¹BF₂(CN)₂).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -153.8 ppm (spt, 1F, (~20%), ¹J(¹⁹F,¹⁰B) = 13.8 Hz, ¹⁰BF₂(CN)₂), -153.9 ppm (s, 1F (~80%), ¹¹BF₂(CN)₂).

IR (ATR): 2209 cm⁻¹ (ν (C \equiv N)).

Raman: 2211 cm⁻¹ (*v*(C≡N)).

Elemental analysis calcd (%) for C₁₁H₂₀BF₂N₃: C 54.35, H 8.29, N 17.28; found: C 53.92, H 8.40, N 18.09.

4.2.3 [BMPL][BF(CN)₃]

 $K[BF(CN)_3]$ (75 g, 0.51 mol) was dissolved in water (150 mL) and [BMPL]Cl (92 g, 0.52 mol) and CH_2Cl_2 (100 mL) were added. The organic layer was separated and washed with deionized water (5×10 mL) and then with bidistilled water (5×5 mL). The CH_2Cl_2 was removed with a rotary evaporator and the remained ionic liquid was dried in a fine vacuum.

Yield: 123.8 g, 0.495 mol [BMPL][BF(CN)₃] (97%).

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 3.73-3.64 (m, 4H, CH₂), 3.53-3.49 (m, 2H, CH₂), 3.23 (s, 3H, CH₃), 2.34-2.27 (m, 4H, CH₂), 1.93-1.85 (m, 2H, CH₂), 1.47-1.39 (m, 2H, CH₂CH₃), 0.98 ppm (t, 3H, ³J(¹H, ¹H) = 7.41 Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 127.7 (qd, ¹J(¹³C,¹¹B) = 75.1 Hz, ²J(¹⁹F,¹³C) = 37.1 Hz, BF(*C*N)₃), 65.1 (t, 2C, ¹J(¹⁴N,¹³C) = 3.4 Hz, NCH₂), 65.0 (t, 1C, ¹J(¹⁴N,¹³C) = 3.0 Hz, NCH₂), 48.9 (t, 1C, ¹J(¹⁴N,¹³C) = 4.0 Hz, NCH₃), 26.2 (s, 1C, *C*H₂), 22.3 (s, 2C, CH₂), 20.3(t, 1C, *C*H₂CH₃) 13.7 ppm (s, 1C CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -17.8 ppm (d, 1B, ¹*J*(¹⁹F, ¹¹B) = 44.6 Hz, *B*F(CN)₃).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -17.8 ppm (s, 1B, *B*F(CN)₃).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -211.9 ppm (q, 1F, ¹J(¹⁹F, ¹¹B) = 44.7 Hz, BF(CN)₃).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -211.9 ppm (s, 1F, B*F*(CN)₃).

IR (ATR): 2212 cm⁻¹ (ν (C \equiv N)).

Raman: 2216 cm⁻¹ (v(C≡N)).

Elemental analysis calcd (%) for C₁₂H₂₀BFN₄: C 57.62, H 8.06, N 22.40; found: C 57.46, H 8.03, N 23.48.

4.2.4 [BMPL][BFH(CN)₂]

 $K[BFH(CN)_2]$ (3.00 g, 24.6 mmol) was dissolved in water (15 mL) and a solution of [BMPL]Cl (5.33 g, 30.0 mmol) in water (15 mL) was added. The mixture was extracted with CH_2Cl_2 (1×30 mL,5×15 mL) and the combined organic layers were concentrated to 25 mL with a rotary evaporator and washed with bidistilled water (5×2 mL). All volatiles were removed in vacuum and the remaining ionic liquid was dried in a fine vacuum.

Yield: 4.088 g, 18.490 mmol [BMPL][BFH(CN)₂] (75%).

¹H{¹¹B} NMR (500.1 MHz, (CD₃)₂CO) δ = 3.68-3.56 (m, 4H, CH₂), 3.46-3.41 (m, 2H, CH₂), 3.14 (s, 3H, CH₃), 2.73 (qd, 1H,¹J(¹¹B,¹H) = 110.0 Hz, ²J(¹⁹F,¹H) = 40.7 Hz, BFH(CN)₂) 2.30-2.23 (m, 4H, CH₂), 1.88-1.80 (m, 2H, CH₂), 1.45-1.37 (m, 2H, CH₂CH₃), 0.96 ppm (t, 3H, ³J(¹H,¹H) = 7.48 Hz, CH₂CH₃).

¹H{¹¹B} NMR (500.1 MHz, (CD₃)₂CO) δ = 3.68-3.56 (m, 4H, CH₂), 3.46-3.41 (m, 2H, CH₂), 3.14 (s, 3H, CH₃), 2.73 (d, 1H, ²J(¹⁹F, ¹H) = 40.7 Hz, BFH(CN)₂) 2.30-2.23 (m, 4H, CH₂), 1.88-1.80 (m, 2H, CH₂), 1.45-1.37 (m, 2H, CH₂CH₃), 0.96 ppm (t, 3H, ³J(¹H, ¹H) = 7.48 Hz, CH₂CH₃).

¹H{¹⁹F} NMR (400 MHz, (CD₃)₂CO) δ = 3.68-3.56 (m, 4H, CH₂), 3.46-3.41 (m, 2H, CH₂), 3.14 (s, 3H, CH₃), 2.73 (q, 1H, ¹J(¹¹B,¹H) = 110.0 Hz, BF*H*(CN)₂) 2.30-2.23 (m, 4H, CH₂), 1.88-1.80 (m, 2H, CH₂), 1.45-1.37 (m, 2H, CH₂CH₃), 0.96 ppm (t, 3H, ³J(¹H,¹H) = 7.48 Hz, CH₂CH₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 132.5 (qd, ¹J(¹³C,¹¹B) = 65.2 Hz, ²J(¹⁹F,¹³C) = 35.6 Hz, BFH(*C*N)₂), 64.8 (t, 2C, ¹J(¹⁴N,¹³C) = 3.2 Hz, NCH₂), 64.6 (t, 1C, ¹J(¹⁴N,¹³C) = 3.0 Hz, NCH₂), 48.7 (t, 1C, ¹J(¹⁴N,¹³C) = 4.1 Hz, NCH₃), 25.9 (s, 1C, *C*H₂), 22.0 (s, 2C, CH₂), 20.0 (t, 1C, *C*H₂CH₃) 13.6 ppm (s, 1C CH₂CH₃).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 132.5 (q, ¹J(¹³C, ¹¹B) = 65.2 Hz, BFH(*C*N)₂), 64.9 (t, 2C, ¹J(¹³C, ¹H) = 148.1 Hz, NCH₂), 64.7 (t, 1C, ¹J(¹³C, ¹H) = 143.8 Hz, NCH₂), 48.7 (qm, 1C, ¹J(¹³C, ¹H) = 143.9 Hz, NCH₃), 26.0 (t, 1C, ¹J(¹³C, ¹H) = 128.3 Hz, *C*H₂), 22.1 (t, 2C, ¹J(¹³C, ¹H) = 136.7 Hz CH₂), 20.1 (t, 1C, ¹J(¹³C, ¹H) = 127.0 Hz, *C*H₂CH₃) 13.6 ppm (qm, 1C, ¹J(¹³C, ¹H) = 125.4 Hz, CH₂CH₃).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -14.2 ppm (dd, 1B, ¹J(¹¹B,¹H) = 110.0 Hz, ¹J(¹⁹F,¹¹B) = 56.2 Hz, *B*FH(CN)₂).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -14.2 ppm (d, 1B, ¹J(¹¹B, ¹H) = 110.0 Hz, *B*FH(CN)₂).

¹¹B{¹H} NMR (160.5 MHz, (CD₃)₂CO) δ = -14.2 ppm (d, 1B, ¹*J*(¹⁹F,¹¹B) = 56.2 Hz, *B*FH(CN)₂).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -233.6 ppm (qd, 1F, ¹J(¹⁹F,¹¹B) = 56.0 Hz, ²J(¹⁹F,¹H) = 40.7 Hz, BFH(CN)₂).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -233.6 ppm (d, 1F, ²J(¹⁹F, ¹H) = 40.7 Hz, BFH(CN)₂). ¹⁹F{¹H} NMR (377 MHz, (CD₃)₂CO) δ = -233.6 ppm (q, 1F, ¹J(¹⁹F, ¹¹B) = 56.0 Hz, BFH(CN)₂). IR (ATR): 2371 (*v*(B−H), 2198 cm⁻¹ (*v*(C≡N)).

Raman: 2379 (v(B−H), 2200 cm⁻¹ (v(C≡N)).

Elemental analysis calcd (%) for C₁₁H₂₁BFN₃: C 58.69, H 9.40, N 18.67; found: C 58.46, H 9.33, N 19.66.

4.2.5 [BMIm][BH₂(CN)₂]

 $K[BH_2(CN)_2]$ (2.2 g, 21.2 mmol) was dissolved in water (5 mL) and a solution of [BMPL]Cl (5.84 g, 33.43 mmol) in water (15 mL) was added. The mixture was extracted with CH_2Cl_2 (5×5 mL) and the combined organic layers were washed with bidistilled water (5×3 mL). All volatiles were removed in a vacuum and the remaining ionic liquid was dried in a fine vacuum.

Yield: 2.87 g, 14.07 mmol [BMPL][BFH(CN)₂] (67%).

4.3 [BnPh₃P] salts

[BnPh₃P]Cl was dissolved in ethanol (20-30 mL) and an equimolar solution of the respective potassium or sodium borate in ethanol was added (10-15 mL). The mixture was stirred for 15 minutes. After adding water (150-200 mL) dropwise for 30 minutes, the resulting precipitate was filtered off and washed with water and diethyl ether. The colourless product was dried in vacuo.

4.3.1 $[BnPh_3P][BF_4]$

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.99–7.91 (m, 3H, P–Ph-*H_p*), 7.81–7.74 (m, 12H, P–Ph-*H_o*/*H_m*), 7.38–7.33 (m, 1H, P–CH₂–Ph-*H_p*), 7.28–7.24 (m, 2H, P–CH₂–Ph-*H_m*), 7.12–7.09 (m, 2H, P–CH₂–Ph-*H_o*), 5.09 ppm (d, 2H, ²*J*(³¹P, ¹H) = 15.0 Hz, P–CH₂–Ph).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴J(³¹P,¹³C) = 3.1 Hz, P-Ph-*C_p*), 135.2 (d, ³J(³¹P,¹³C) = 9.8 Hz, P-Ph-*C_o*), 132.0 (d, ³J(³¹P,¹³C) = 5.5 Hz, P-CH₂-Ph-*C_o*), 131.1 (d, ³J(³¹P,¹³C) = 12.6 Hz, P-Ph-*C_m*), 129.9 (d, ⁴J(³¹P,¹³C) = 3.3 Hz, P-CH₂-Ph-*C_m*), 129.6 (d, ⁵J(³¹P,¹³C) = 3.8 Hz, P-CH₂-Ph-*C_p*), 128.5 (d, ²J(³¹P,¹³C) = 8.5 Hz, P-CH₂-Ph-*C_i*), 118.8 (d, ¹J(³¹P,¹³C) = 86.1 Hz, P-Ph-*C_i*), 30.3 ppm (d, ¹J(³¹P,¹³C) = 53.1 Hz, P-CH₂-Ph).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -0.86 ppm (s, 1B, *B*F₄).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -0.86 ppm (s, 1B, *B*F₄).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -151.54 (s, 4F (~20%), ¹⁰BF₄) -151.59 ppm (s, 4F (~80%), ¹¹BF₄).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -151.54 (s, 4F (~20%), ¹⁰BF₄) -151.59 ppm (s, 4F (~80%), ¹¹BF₄).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 23.0 ppm (s, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 23.0 ppm (s, BzPh₃P).

Elemental analysis calcd (%) for C₂₅H₂₂BF₄P: C 68.21, H 5.04; found: C 68.15, H 5.04.

K[BF₄]: 300 mg, 2.38 mmol.

[BnPh₃P]Cl: 930 mg, 2.38 mmol.

yield: 710 mg, 1.61 mmol [BnPh₃P][BF₄] (68 %).

4.3.2 [BnPh₃P][BF₃(CN)]

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.98–7.94 (m, 3H, P–Ph- H_p), 7.81–7.74 (m, 12H, P–Ph- H_o/H_m), 7.38–7.34 (m, 1H, P–CH₂–Ph- H_p), 7.29–7.24 (m, 2H, P–CH₂–Ph- H_m), 7.13–7.09 (m, 2H, P–CH₂–Ph- H_o), 5.09 ppm (d, 2H, ²J(³¹P, ¹H) = 15.0 Hz, P–CH₂–Ph).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴J(³¹P,¹³C) = 3.1 Hz, P-Ph-*C_p*), 135.2 (d, ³J(³¹P,¹³C) = 9.7 Hz, P-Ph-*C_o*), 132.4 (m, BF₃(*C*N)), 132.0 (d, ³J(³¹P,¹³C) = 5.5 Hz, P-CH₂-Ph-*C_o*), 131.1 (d, ³J(³¹P,¹³C) = 12.6 Hz, P-Ph-*C_m*), 129.9 (d, ⁴J(³¹P,¹³C) = 3.2 Hz, P-CH₂-Ph-*C_m*), 129.6 (d, ⁵J(³¹P,¹³C) = 3.8 Hz, P-CH₂-Ph-*C_p*), 128.5 (d, ²J(³¹P,¹³C) = 8.5 Hz, P-CH₂-Ph-*C_i*), 118.8 (d, ¹J(³¹P,¹³C) = 86.1 Hz, P-Ph-*C_i*), 30.4 ppm (d, ¹J(³¹P,¹³C) = 48.4 Hz, P-CH₂-Ph).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (m, P–Ph-*C_p*), 135.2 (m, P–Ph-*C_o*), 132.4 (q, ¹*J*(¹³C,¹¹B) = 89.3 Hz, BF₃(*C*N)), 132.0 (m, P–CH₂–Ph-*C_o*), 131.1 (m, P–Ph-*C_m*), 129.9 (m, P–CH₂–Ph-*C_m*), 129.6 (m, P–CH₂–Ph-*C_p*), 128.5 (m, P–CH₂–Ph-*C_i*), 118.8 (m, P–Ph-*C_i*) 30.3 ppm (m, P–*C*H₂–Ph).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -3.47 ppm (t, 1B, ¹J(¹⁹F, ¹¹B) = 26.8 Hz, BF₃(CN)).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -3.47 ppm (s, 1B, BF₃(CN)).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -137.6 (spt, 3F (~20%), ¹J(¹⁹F,¹⁰B) = 8.7 Hz, ¹⁰BF₃(CN)), -137.7 ppm (q, 3F (~80%), ¹J(¹⁹F,¹¹B) = 26.8 Hz, ¹¹BF₃(CN)).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -137.6 (spt, 3F (~20%), ¹J(¹⁹F, ¹⁰B) = 8.7 Hz, ¹⁰BF₃(CN)), -137.7 ppm (s, 3F (~80%), ¹¹BF₃(CN)).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

IR (ATR): 2207 cm⁻¹ (ν (C \equiv N)).

Raman: 2209 cm⁻¹ (*v*(C≡N)).

Elemental analysis calcd (%) for C₂₆H₂₂BF₃NP: C 69.82, H 4.96, N 3.13; found: C 69.80, H 5.01, N 3.12.

K[BF₃(CN)]: 200 mg, 1.50 mmol.

[BnPh₃P]Cl: 590 mg, 1.50 mmol.

yield: 511 mg, 1.14 mmol [BnPh₃P][BF₃(CN)] (76 %).

4.3.3 $[BnPh_3P][BF_2(CN)_2]$

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.98–7.94 (m, 3H, P–Ph- H_p), 7.82–7.74 (m, 12H, P–Ph- H_o/H_m), 7.38–7.34 (m, 1H, P–CH₂–Ph- H_p), 7.29–7.24 (m, 2H, P–CH₂–Ph- H_m), 7.13–7.09 (m, 2H, P–CH₂–Ph- H_o), 5.08 ppm (d, 2H, ²J(³¹P, ¹H) = 15.0 Hz, P–CH₂–Ph).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴J(³¹P,¹³C) = 3.0 Hz, P-Ph-*C_p*), 135.1 (d, ³J(³¹P,¹³C) = 9.8 Hz, P-Ph-*C_o*), 130.8 (qd, BF₂(*C*N)₂), 131.9 (d, ³J(³¹P,¹³C) = 5.5 Hz, P-CH₂-Ph-*C_o*), 131.1 (d, ³J(³¹P,¹³C) = 12.6 Hz, P-Ph-*C_m*), 129.9 (d, ⁴J(³¹P,¹³C) = 3.2 Hz, P-CH₂-Ph-*C_m*), 129.6 (d, ⁵J(³¹P,¹³C) = 3.8 Hz, P-CH₂-Ph-*C_p*), 128.4 (d, ²J(³¹P,¹³C) = 8.5 Hz, P-CH₂-Ph-*C_i*), 118.7 (d, ¹J(³¹P,¹³C) = 86.1 Hz, P-Ph-*C_i*), 30.4 ppm (d, ¹J(³¹P,¹³C) = 48.4 Hz, P-CH₂-Ph).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (m, P-Ph-*C_p*), 135.2 (m, P-Ph-*C_o*), 135.1 (m, P-CH₂-Ph-*C_o*), 130.8 (q, ¹*J*(¹³C, ¹¹B) = 80.6 Hz, BF₂(*C*N)₂), 131.9 (m, P-CH₂-Ph-*C_o*), 131.1 (m, P-Ph-*C_m*), 129.9 (m, P-CH₂-Ph-*C_m*), 129.6 (m, P-CH₂-Ph-*C_p*), 128.4 (m, P-CH₂-Ph-*C_i*), 118.7 (m, P-Ph-*C_i*), 30.4 ppm (m, P-*C*H₂-Ph).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -7.2 ppm (t, 1B,¹J(¹⁹F,¹¹B) = 41.2 Hz) BF₂(CN)₂).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -7.2 ppm (s, 1B, BF₂(CN)₂).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -153.9 (spt, 2F (~20%), ¹*J*(¹⁹F,¹⁰B) = 13.8 Hz, ¹⁰BF₂(CN)₂), -154.0 ppm (q, 4F (~80%), ¹*J*(¹⁹F,¹¹B) = 41.2 Hz, ¹¹BF₂(CN)₂). ¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -153.9 (spt, 2F (~20%), ¹J(¹⁹F,¹⁰B) = 13.8 Hz, ¹⁰BF₂(CN)₂), -154.0 ppm (s, 4F (~80%), ¹¹BF₄).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

IR (ATR): 2208 cm⁻¹ (v(C≡N)).

Raman: 2211 cm⁻¹ (*v*(C≡N)).

Elemental analysis calcd (%) for C₂₇H₂₂BF₂N₂P: C 71.39, H 4.88, N 6.17; found: C 71.40, H 5.01, N 6.29.

Na[BF₂(CN)₂]: 2.40 g, 19.4 mmol.

[BnPh₃P]Cl: 8.0 g, 20.6 mmol.

yield: 8.43 g, 18.56 mmol [BnPh₃P][BF₂(CN)₂] (95.5 %).

4.3.4 [BnPh₃P][BF(CN)₃]

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.99–7.92 (m, 3H, P–Ph- H_p), 7.82–7.74 (m, 12H, P–Ph- H_o/H_m), 7.39–7.34 (m, 1H, P–CH₂–Ph- H_p), 7.30–7.25 (m, 2H, P–CH₂–Ph- H_m), 7.13–7.09 (m, 2H, P–CH₂–Ph- H_o), 5.09 ppm (d, 2H, ²J(³¹P, ¹H) = 15.0 Hz, P–CH₂–Ph).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴J(³¹P,¹³C) = 3.0 Hz, P-Ph-*C_p*), 135.2 (d, ³J(³¹P,¹³C) = 9.8 Hz, P-Ph-*C_o*), 131.9 (d, ³J(³¹P,¹³C) = 5.5 Hz, P-CH₂-Ph-*C_o*), 131.1 (d, ³J(³¹P,¹³C) = 12.6 Hz, P-Ph-*C_m*), 129.9 (d, ⁴J(³¹P,¹³C) = 3.3 Hz, P-CH₂-Ph-*C_m*), 129.6 (d, ⁵J(³¹P,¹³C) = 3.8 Hz, P-CH₂-Ph-*C_p*), 128.4 (d, ²J(³¹P,¹³C) = 8.5 Hz, P-CH₂-Ph-*C_i*), 127.9 (qd, ¹J(¹³C,¹¹B) = 74.6 Hz, ²J(¹⁹F,¹³C) = 37.4 Hz, BF(*C*N)₃), 118.7 (d, ¹J(³¹P,¹³C) = 86.1 Hz, P-Ph-*C_i*), 30.4 ppm (d, ¹J(³¹P,¹³C) = 48.5 Hz, P-CH₂-Ph).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (m, P-Ph-*C_p*), 135.2 (m, P-Ph-*C_o*), 132.0 (m, P-CH₂-Ph-*C_o*), 131.1 (m, P-Ph-*C_m*), 129.9 (m, P-CH₂-Ph-*C_m*), 129.6 (m, P-CH₂-Ph-*C_p*), 128.5 (m, P-CH₂-Ph-*C_i*), 127.9 (q, ¹J(¹³C, ¹¹B) = 74.7 Hz, BF(*C*N)₃), 118.7 (m, P-Ph-*C_i*), 30.4 ppm (m, P-CH₂-Ph).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -17.7 ppm (d, 1B, ¹*J*(¹⁹F, ¹¹B) = 44.5 Hz, *B*F(CN)₃).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -17.7 ppm (s, 1B, BF(CN)₃).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -211.8 (spt, 1F (~20%), ¹J(¹⁹F,¹⁰B) = 14.9 Hz, ¹⁰BF(CN)₃), -211.9 ppm (q, 1F (~80%), ¹J(¹⁹F,¹¹B) = 44.5 Hz, ¹¹BF(CN)₃).

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -211.8 (spt, 1F (~20%), ¹J(¹⁹F, ¹⁰B) = 14.9 Hz, ¹⁰BF(CN)₃) -211.9 ppm (s, 4F (~80%), ¹¹BF(CN)₃).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

IR (ATR): 2213 cm⁻¹ (ν(C≡N)).

Raman: 2217 cm⁻¹ (*v*(C≡N)).

Elemental analysis calcd (%) for C₂₈H₂₂BFN₃P: C 72.91, H 4.81, N 9.11; found: C 72.81, H 4.77, N 9.15.

K[BF(CN)₃]: 200 mg, 1.50 mmol.

[BnPh₃P]Cl: 590 mg, 1.50 mmol.

yield: 546 mg, 1.18 mmol [BnPh₃P][BF(CN)₃] (87 %).

4.3.5 $[BnPh_3P][B(CN)_4]$

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.99–7.93 (m, 3H, P–Ph- H_p), 7.82–7.74 (m, 12H, P–Ph- H_o/H_m), 7.39–7.34 (m, 1H, P–CH₂–Ph- H_p), 7.30–7.25 (m, 2H, P–CH₂–Ph- H_m), 7.13–7.09 (m, 2H, P–CH₂–Ph- H_o), 5.09 ppm (d, 2H, ²J(³¹P, ¹H) = 15.0 Hz, P–CH₂–Ph).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴J(³¹P,¹³C) = 3.0 Hz, P-Ph-*C_p*), 135.1 (d, ³J(³¹P,¹³C) = 9.8 Hz, P-Ph-*C_o*), 131.9 (d, ³J(³¹P,¹³C) = 5.5 Hz, P-CH₂-Ph-*C_o*), 131.1 (d, ³J(³¹P,¹³C) = 12.6 Hz, P-Ph-*C_m*), 129.9 (d, ⁴J(³¹P,¹³C) = 3.2 Hz, P-CH₂-Ph-*C_m*), 129.6 (d, ⁵J(³¹P,¹³C) = 3.8 Hz, P-CH₂-Ph-*C_p*), 128.4 (d, ²J(³¹P,¹³C) = 8.5 Hz, P-CH₂-Ph-*C_i*), 123.1 (q, ¹J(¹³C,¹¹B) = 70.8 Hz, B(CN)₄), 118.7 (d, ¹J(³¹P,¹³C) = 86.1 Hz, P-Ph-*C_i*), 30.4 ppm (d, ¹J(³¹P,¹³C) = 48.3 Hz, P-CH₂-Ph).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -38.5 ppm (s, 1B, *B*(CN)₄).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 25.2–21.1 ppm (m, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

IR (ATR): 2221 cm⁻¹ (ν (C \equiv N)).

Raman: 2224 cm⁻¹ (*v*(C≡N)).

Elemental analysis calcd (%) for C₂₉H₂₂BN₄P: C 74.38, H 4.74, N 11.96; found: C 74.36, H 4.75, N 12.01.

K[B(CN)₄]: 200 mg, 1.45 mmol.

[BnPh₃P]Cl: 565 mg, 1.45 mmol.

yield: 580 mg, 1.24 mmol [BnPh₃P][B(CN)₄] (85 %).

4.3.6 [BnPh₃P][BFH(CN)₂]

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.98–7.93 (m, 3H, P–Ph-*H_p*), 7.81–7.74 (m, 12H, P–Ph-*H_o*/*H_m*), 7.38–7.33 (m, 1H, PCH₂Ph-*H_p*), 7.28–7.24 (m, 2H, PCH₂Ph-*H_m*), 7.12–7.09 (m, 2H, PCH₂Ph-*H_o*), 5.08 (d, 2H, ²*J*(³¹P,¹H) = 15.0 Hz, P–C*H*₂–Ph), 2.82 ppm (qd, 1H, ¹*J*(¹¹B,¹H) = 109.3 Hz, ²*J*(¹⁹F,¹H) = 40.7 Hz, BF*H*(CN)₂).

¹H{¹¹B} NMR (500.1 MHz, (CD₃)₂CO) δ = 7.98–7.93 (m, 3H, P–Ph-*H_p*), 7.81–7.74 (m, 12H, P–Ph-*H_o*/*H_m*), 7.38–7.33 (m, 1H, PCH₂Ph-*H_p*), 7.28–7.24 (m, 2H, PCH₂Ph-*H_m*), 7.12–7.09 (m, 2H, PCH₂Ph-*H_o*), 5.08 (d, 2H, ²*J*(³¹P, ¹H) = 15.0 Hz, P–C*H*₂–Ph), 2.82 ppm (d, 1H, ²*J*(¹⁹F, ¹H) = 40.7 Hz, BF*H*(CN)₂).

¹H{¹⁹F} NMR (400 MHz, (CD₃)₂CO) δ = 7.98–7.93 (m, 3H, P–Ph-*H_p*), 7.81–7.74 (m, 12H, P–Ph-*H_o*/*H_m*), 7.38–7.33 (m, 1H, PCH₂Ph-*H_p*), 7.28–7.24 (m, 2H, PCH₂Ph-*H_m*), 7.12–7.09 (m, 2H, PCH₂Ph-*H_o*), 5.08 (d, 2H, ²*J*(³¹P,¹H) = 15.0 Hz, P–C*H*₂–Ph), 2.82 ppm (q, 1H, ¹*J*(¹¹B,¹H) = 109.3 Hz, BF*H*(CN)₂).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴J(³¹P,¹³C) = 3.0 Hz, P-Ph-*C_p*), 135.1 (d, ³J(³¹P,¹³C) = 9.7 Hz, P-Ph-*C_o*), 132.8 (qd, ¹J(¹³C,¹¹B) = 65.3 Hz, ²J(¹⁹F,¹³C) = 34.7 Hz, BFH(*C*N)₂), 131.9 (d, ³J(³¹P,¹³C) = 5.5 Hz, P-CH₂-Ph-*C_o*), 131.1 (d, ³J(³¹P,¹³C) = 12.6 Hz, P-Ph-*C_m*), 129.9 (d, ⁴J(³¹P,¹³C) = 3.2 Hz, P-CH₂-Ph-*C_m*), 129.5 (d, ⁵J(³¹P,¹³C) = 3.8 Hz, P-CH₂-Ph-*C_p*), 128.4 (d, ²J(³¹P,¹³C) = 8.5 Hz, P-CH₂-Ph-*C_i*), 118.6 (d, ¹J(³¹P,¹³C) = 86.1 Hz, P-Ph-*C_i*), 30.3 ppm (d, ¹J(³¹P,¹³C) = 48.0 Hz, P-CH₂-Ph).

¹³C{¹⁹F} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (m, P-Ph-*C_p*), 135.1 (m, P-Ph-*C_o*), 132.8 (q, ¹*J*(¹³C, ¹¹B) = 65.3 Hz, BFH(*C*N)₂), 131.9 (m, P-CH₂-Ph-*C_o*), 131.1 (m, P-Ph-*C_m*), 129.9 (m, P-CH₂-Ph-*C_m*), 129.5 (m, P-CH₂-Ph-*C_p*), 128.4 (m, P-CH₂-Ph-*C_i*), 118.6 (m, P-Ph-*C_i*), 30.3 ppm (m, P-CH₂-Ph).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) $\delta = -14.1 \text{ ppm}$ (dd, 1B, ¹J(¹¹B,¹H) = 109.4 Hz, ¹J(¹⁹F,¹¹B) = 56.0 Hz, BH₃(CN)).

¹¹B{¹H} NMR (160.5 MHz, (CD₃)₂CO) δ = -14.1 ppm (d, 1B, ¹J(¹⁹F, ¹¹B) = 56.0 Hz, BH₃(CN)).

¹¹B{¹⁹F} NMR (160.5 MHz, (CD₃)₂CO) δ = -14.1 ppm (d, 1B, ¹J(¹¹B, ¹H) = 109.4 Hz, BH₃(CN)).

¹⁹F NMR (470.5 MHz, (CD₃)₂CO) δ = -234.0 ppm (qd, 1F, ¹J(¹⁹F, ¹¹B) = 55.9 Hz,

 $^{2}J(^{19}F,^{1}H) = 40.7 \text{ Hz}, BFH(CN)_{2}).$

¹⁹F{¹¹B} NMR (470.5 MHz, (CD₃)₂CO) δ = -234.0 ppm (d, 1F, ²J(¹⁹F,¹H) = 40.7 Hz, BFH(CN)₂).

¹⁹F{¹H} NMR (377 MHz, (CD₃)₂CO) δ = -234.0 ppm (q, 1F, ¹J(¹⁹F, ¹¹B) = 55.9 Hz, BFH(CN)₂).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

IR (ATR): 2401 (v(B−H), 2201 cm⁻¹ (v(C≡N)).

Raman: 2403 (v(B−H), 2202 cm⁻¹ (v(C≡N)).

Elemental analysis calcd (%) for C₂₇H₂₃BFN₂P: C 74.33, H 5.31, N 6.42; found: C 74.51, H 5.41, N 6.53.

K[BFH(CN)₂]: 1.34 g, 11.0 mmol.

[BnPh₃P]Cl: 4.3 g, 11.0 mmol.

yield: 3.54 g, 8.37 mmol [BnPh₃P][BFH(CN)₂] (77 %).

4.3.7 [BnPh₃P][BH₃(CN)]

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.99–7.91 (m, 3H, P–Ph-*H_p*), 7.81–7.74 (m, 12H, P–Ph-*H_o*/*H_m*), 7.37–7.32 (m, 1H, PCH₂Ph-*H_p*), 7.28–7.23 (m, 2H, PCH₂Ph-*H_m*), 7.13–7.09 (m, 2H, PCH₂Ph-*H_o*), 5.12 (d, 2H, ²*J*(³¹P, ¹H) = 15.0 Hz, P–C*H*₂–Ph), 0.51 ppm (q, 3H, ¹*J*(¹¹B, ¹H) = 88.3 Hz, B*H*₃(CN)).

¹H{¹¹B} NMR (500.1 MHz, (CD₃)₂CO) δ = ¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.99–7.91 (m, 3H, P–Ph-*H*_p), 7.81–7.74 (m, 12H, P–Ph-*H*_o/*H*_m), 7.37–7.32 (m, 1H, P–CH₂–Ph-*H*_p), 7.28–7.23 (m, 2H, P–CH₂–Ph-*H*_m), 7.13–7.09 (m, 2H, P–CH₂–Ph-*H*_o), 5.12 (d, 2H, ²*J*(³¹P,¹H) = 15.0 Hz, P–CH₂–Ph), 0.51 ppm (s, 3H, B*H*₃(CN)).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴J(³¹P,¹³C) = 3.0 Hz, P-Ph-*C_p*), 135.2 (d, ³J(³¹P,¹³C) = 9.8 Hz, P-Ph-*C_o*), 132.0 (d, ³J(³¹P,¹³C) = 5.5 Hz, P-CH₂-Ph-*C_o*), 131.1 (d, ³J(³¹P,¹³C) = 12.6 Hz, P-Ph-*C_m*), 129.8 (d, ⁴J(³¹P,¹³C) = 3.3 Hz, P-CH₂-Ph-*C_m*), 129.5 (d, ⁵J(³¹P,¹³C) = 3.9 Hz, P-CH₂-Ph-*C_p*), 128.5 (d, ²J(³¹P,¹³C) = 8.5 Hz, P-CH₂-Ph-*C_i*), 118.8 (d, ¹J(³¹P,¹³C) = 86.0 Hz, P-Ph-*C_i*), 30.3 ppm (d, ¹J(³¹P,¹³C) = 54.1 Hz, P-CH₂-Ph).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -41.8 ppm (q, 1B, ¹J(¹¹B, ¹H) = 88.3 Hz, BH₃(CN)).

¹¹B{¹H} NMR (160.5 MHz, (CD₃)₂CO) δ = -41.8 ppm (s, 1B, BH₃(CN)).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 23.0 ppm (s, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 23.0 ppm (s, BzPh₃P).

IR (ATR): 2310 (ν (B–H), 2161 cm⁻¹ (ν (C \equiv N)).

Raman: 2334 (v(B−H), 2167 cm⁻¹ (v(C≡N)).

Elemental analysis calcd (%) for C₂₇H₂₄BN₂P: C 79.41, H 6.41, N 3.59; found: C 79.18, H 6.27, N 3.50.

K[BH₃(CN)]: 200 mg, 3.18 mmol.

[BnPh₃P]Cl: 1.24 g, 3.18 mmol.

yield: 846 mg, 2.15 mmol [BnPh₃P][BH₃(CN)] (68 %).

4.3.8 [BnPh₃P][BH₂(CN)₂]

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.99–7.92 (m, 3H, P–Ph-*H_p*), 7.82–7.74 (m, 12H, P–Ph-*H_o*/*H_m*), 7.38–7.33 (m, 1H, P–CH₂–Ph-*H_p*), 7.29–7.24 (m, 2H, P–CH₂–Ph-*H_m*), 7.13–7.09 (m, 2H, P–CH₂–Ph-*H_o*), 5.10 (d, 2H, ²*J*(³¹P,¹H) = 15.0 Hz, P–CH₂–Ph), 1.12 ppm (q, 2H, ¹*J*(¹¹B,¹H) = 93.7 Hz, B*H*₂(CN)₂).

¹H{¹¹B} NMR (500.1 MHz, (CD₃)₂CO) δ = 7.99–7.92 (m, 3H, P–Ph-*H_p*), 7.82–7.74 (m, 12H, P–Ph-*H_o*/*H_m*), 7.38–7.33 (m, 1H, P–CH₂–Ph-*H_p*), 7.29–7.24 (m, 2H, P–CH₂–Ph-*H_m*), 7.13–7.09 (m, 2H, P–CH₂–Ph-*H_o*), 5.10 (d, 2H, ²*J*(³¹P,¹H) = 15.0 Hz, P–C*H*₂–Ph), 1.12 ppm (s, 2H, B*H*₂(CN)₂). ¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴*J*(³¹P,¹³C) = 3.0 Hz, P–Ph-*C_p*), 135.1 (d, ³*J*(³¹P,¹³C) = 9.8 Hz, P–Ph-*C_o*), 134.6 (q, 2C, ¹*J*(¹³C,¹¹B) = 58.6 Hz, BH₂(CN)₂), 131.9 (d, ³*J*(³¹P,¹³C) = 5.5 Hz, P–CH₂–Ph-*C_o*), 131.1 (d, ³*J*(³¹P,¹³C) = 12.6 Hz, P–Ph-*C_m*), 129.9 (d, ⁴*J*(³¹P,¹³C) = 3.3 Hz, P–CH₂–Ph-*C_m*), 129.5 (d, ⁵*J*(³¹P,¹³C) = 3.8 Hz, P–CH₂–Ph-*C_p*), 128.4 (d, ${}^{2}J({}^{31}P,{}^{13}C) = 8.5 \text{ Hz}, P-CH_{2}-Ph-C_{i}), 118.7 (d, {}^{1}J({}^{31}P,{}^{13}C) = 86.1 \text{ Hz}, P-Ph-C_{i}), 30.3 \text{ ppm} (d, {}^{1}J({}^{31}P,{}^{13}C) = n.b., P-CH_{2}-Ph).$

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -41.6 ppm (t, 1B, ¹J(¹¹B, ¹H) = 93.7 Hz, BH₂(CN)₂).

¹¹B{¹H} NMR (160.5 MHz, (CD₃)₂CO) δ = -41.6 ppm (s, 1B, BH₂(CN)₂).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 23.0 ppm (s, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 23.0 ppm (s, BzPh₃P).

IR (ATR): 2385 (v(B−H), 2184 cm⁻¹ (v(C≡N)).

Raman: 2389 (*v*(B−H), 2191 cm⁻¹ (*v*(C≡N)).

Elemental analysis calcd (%) for C₂₇H₂₄BN₂P: C 77.53, H 5.78, N 6.70; found: C 77.65, H 5.79, N 6.70.

K[BH₂(CN)₂]: 200 mg, 1.92 mmol.

[BnPh₃P]Cl: 750 mg, 1.92 mmol.

yield: 717 mg, 1.71 mmol [BnPh₃P][BH₂(CN)₂] (89 %).

4.3.9 [BnPh₃P][BH(CN)₃]

¹H NMR (500.1 MHz, (CD₃)₂CO) δ = 7.98–7.93 (m, 3H, P–Ph-*H_p*), 7.82–7.73 (m, 12H, P–Ph-*H_o*/*H_m*), 7.38–7.34 (m, 1H, P–CH₂–Ph-*H_p*), 7.30–7.25 (m, 2H, P–CH₂–Ph-*H_m*), 7.11–7.09 (m, 2H, P–CH₂–Ph-*H_o*), 5.09 (d, 2H, ²*J*(³¹P,¹H) = 15.0 Hz, P–CH₂–Ph), 1.81 ppm (q, 1H, ¹*J*(¹¹B,¹H) = 96.7 Hz, B*H*(CN)₃).

¹H{¹¹B} NMR (500.1 MHz, (CD₃)₂CO) δ = 7.98–7.93 (m, 3H, P–Ph- H_p), 7.82–7.73 (m, 12H, P–Ph- H_o/H_m), 7.38–7.34 (m, 1H, P–CH₂–Ph- H_p), 7.30–7.25 (m, 2H, P–CH₂–Ph- H_m), 7.11–7.09 (m, 2H, P–CH₂–Ph- H_o), 5.09 (d, 2H, ²J(³¹P,¹H) = 15.0 Hz, P–CH₂–Ph), 1.81 ppm (s, 1H, BH(CN)₃).

¹³C{¹H} NMR (125.7 MHz, (CD₃)₂CO) δ = 136.2 (d, ⁴J(³¹P,¹³C) = 3.0 Hz, P-Ph-*C_p*), 135.1 (d, ³J(³¹P,¹³C) = 9.8 Hz, P-Ph-*C_o*), 131.9 (d, ³J(³¹P,¹³C) = 5.5 Hz, P-CH₂-Ph-*C_o*), 131.1 (d, ³J(³¹P,¹³C) = 12.6 Hz, P-Ph-*C_m*), 129.9 (d, ⁴J(³¹P,¹³C) = 3.3 Hz, P-CH₂-Ph-*C_m*), 129.6 (d, ⁵J(³¹P,¹³C) = 3.8 Hz, P-CH₂-Ph-*C_p*), 128.4 (d, ²J(³¹P,¹³C) = 8.5 Hz, P-CH₂-Ph-*C_i*), 128.0 (q, 3C, ¹J(¹³C,¹¹B) = 66.0 Hz, BH(*C*N)₃), 118.7 (d, ¹J(³¹P,¹³C) = 86.1 Hz, P-Ph-*C_i*), 30.4 ppm (d, ¹J(³¹P,¹³C) = 48.4 Hz, P-CH₂-Ph).

¹¹B NMR (160.5 MHz, (CD₃)₂CO) δ = -40.0 ppm (d, 1B, ¹*J*(¹¹B, ¹H) = 96.7 Hz, *B*H(CN)₃).

¹¹B{¹H} NMR (160.5 MHz, (CD₃)₂CO) δ = -40.0 ppm (s, 1B, BH(CN)₃).

³¹P NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

³¹P{¹H} NMR (202.5 MHz,(CD₃)₂CO) δ = 22.9 ppm (s, BzPh₃P).

IR (ATR): 2394 (v(B−H), 2213 cm⁻¹ (v(C≡N)).

Raman: 2397 (v(B−H), 2214 cm⁻¹ (v(C≡N)).

Elemental analysis calcd (%) for C₂₈H₂₃BN₃P: C 75.87, H 5.23, N 9.48; found: C 75.89, H 5.28, N 9.51.

K[BH(CN)₃]: 200 mg, 1.55 mmol.

[BnPh₃P]Cl: 603 mg, 1.55 mmol.

yield: 470 mg, 1.06 mmol [BnPh₃P][BH(CN)₃] (68 %).

5 NMR spectra

5.1 [EMIm] salts

5.1.1 [EMIm][BF₄]



Figure S1: ¹H NMR spectrum of [EMIm][BF₄].



Figure SS2: ¹³C{¹H} NMR spectrum of [EMIm][BF₄].



6

4

2

~

-2

-4

-6

0

 $-\delta$ (ppm)

Figure S4: ¹¹B{¹⁹F} NMR spectrum of [EMIm][BF₄].



≺−−− δ (ppm)

Figure S6: ¹⁹F{¹¹B} NMR spectrum of [EMIm][BF4].

5.1.2 [EMIm][BF₃(CN)]



Figure S7: ¹H NMR spectrum of [EMIm][BF₃(CN)].



Figure S8: ¹³C{¹H} NMR spectrum of [EMIm][BF₃(CN)].



Figure S8: ¹³C{¹⁹F} NMR spectrum of [EMIm][BF₃(CN)].



Figure S9: ¹¹B NMR spectrum of [EMIm][BF₃(CN)].



Figure S10: ¹¹B{¹⁹F} NMR spectrum of [EMIm][BF₃(CN)].



Figure S11: ¹⁹F NMR spectrum of [EMIm][BF₃(CN)].



Figure S12: ¹⁹F{¹¹B} NMR spectrum of [EMIm][BF₃(CN)].

5.1.3 [EMIm][BF₂(CN)₂]



Figure S13: ¹H NMR spectrum of [EMIm][BF₂(CN)₂].



Figure S14: ¹³C{¹H} NMR spectrum of [EMIm][BF₂(CN)₂].



Figure S15: ${}^{13}C{}^{19}F{}$ NMR spectrum of [EMIm][BF₂(CN)₂].



Figure S16: ¹¹B NMR spectrum of [EMIm][BF₂(CN)₂].



Figure S17: ¹¹B{¹⁹F} NMR spectrum of [EMIm][BF₂(CN)₂].



Figure S18: ¹⁹F NMR spectrum of [EMIm][BF₂(CN)₂].



Figure S19: ¹⁹F{¹¹B} NMR spectrum of [EMIm][BF₂(CN)₂].

5.1.4 [EMIm][BF(CN)₃]



Figure S21: ¹³C{¹H} NMR spectrum of [EMIm][BF(CN)₃].



Figure S23: ¹¹B{¹⁹F} NMR spectrum of [EMIm][BF(CN)₃].



Figure S24: ¹⁹F NMR spectrum of [EMIm][BF(CN)₃].



Figure S25: ${}^{19}F{}^{11}B$ NMR spectrum of [EMIm][BF(CN)₃].

5.1.5 [EMIm][BFH(CN)₂]



Figure S26: ¹H NMR spectrum of [EMIm][BFH(CN)₂].



Figure S27: ¹H{¹¹B} NMR spectrum of [EMIm][BFH(CN)₂].


Figure S28: ¹³C{¹H} NMR spectrum of [EMIm][BFH(CN)₂].



Figure S29: ¹³C{¹⁹F} NMR spectrum of [EMIm][BFH(CN)₂].



Figure S30: ¹¹B NMR spectrum of [EMIM][BFH(CN)₂].



Figure S31: ¹¹B{¹H} NMR spectrum of [EMIm][BFH(CN)²].



Figure S32: ¹¹B{¹⁹F} NMR spectrum of [EMIm][BFH(CN)₂].



Figure S33: ¹⁹F NMR spectrum of [EMIm][BFH(CN)₂].



Figure S34: ¹⁹F{¹¹B} NMR spectrum of [EMIm][BFH(CN)₂].

5.2 [BMPL] salts

5.2.1 [BMPL][BF₃(CN)]



Figure S35: ¹H NMR spectrum of [BMPL][BF₃(CN)].



Figure S36: 13C{¹H} NMR spectrum of [BMPL][BF₃(CN)].



Figure S37:¹³C{¹⁹F} NMR spectrum of [BMPL][BF₃(CN)].



Figure S38: ¹¹B NMR spectrum of [BMPL][BF₃(CN)].



Figure S40: ¹⁹F NMR spectrum of [BMPL][BF₃(CN)].



Figure S41: ¹⁹F{¹¹B} NMR spectrum of [BMPL][BF₃(CN)].

5.2.2 [BMPL][BF₂(CN)₂]



Figure S42: 1H NMR spectrum of [BMPL][BF₂(CN)₂].



Figure S43: ${}^{13}C{}^{1}H$ NMR spectrum of [BMPL][BF₂(CN)₂].



Figure S44: ¹³C{¹⁹F} NMR spectrum of [BMPL][BF₂(CN)₂].



Figure S45: ¹¹B NMR spectrum of [BMPL][BF₂(CN)₂].



Figure S46: ${}^{11}B{}^{19}F{}$ NMR spectrum of [BMPL][BF₂(CN)₂].



Figure S47: ¹⁹F NMR spectrum of [BMPL][BF₂(CN)₂].



Figure S48: ${}^{19}F{}^{11}B{}$ NMR spectrum of [BMPL][BF₂(CN)₂].

5.2.3 [BMPL][BF(CN)₃]



Figure S49: ¹H NMR spectrum of [BMPL][BF(CN)₃].



Figure S50: ¹³C{¹H} NMR spectrum of [BMPL][BF(CN)₃].



Figure S51: ¹¹B NMR spectrum of [BMPL][BF(CN)₃].



Figure S52: ¹¹B{¹⁹F} NMR spectrum of [BMPL][BF(CN)₃].



Figure S53: ¹⁹F NMR spectrum of [BMPL][BF(CN)₃].



Figure S54: ¹⁹F{¹¹B} NMR spectrum of [BMPL][BF(CN)₃].



Figure S55: ¹H NMR spectrum of [BMPL][BFH(CN)₂].



Figure S56: ¹H{¹¹B} NMR spectrum of [BMPL][BFH(CN)2].



Figure S57: ¹H{¹⁹F} NMR spectrum of [BMPL][BFH(CN)₂].



Figure S58: ${}^{13}C{}^{1}H$ NMR spectrum of [BMPL][BFH(CN)₂].



Figure S60: ¹¹B{¹H} NMR spectrum of [BMPL][BFH(CN)₂].



Figure S62: ¹⁹F NMR spectrum of [BMPL][BFH(CN)₂].



Figure S64: ${}^{19}F{}^{11}B$ NMR spectrum of [BMPL][BFH(CN)₂].

5.3 Phosphonium salts

5.3.1 [BnPh₃P][BF₄]



Figure S65: ¹H NMR spectrum of [BnPh₃P][BF₄].



Figure S66: ¹³C{¹H} NMR spectrum of [BnPh₃P][BF₄].



Figure S68: ¹¹B{¹H} NMR spectrum of [BnPh₃P][BF₄].



Figure S70: ¹⁹F{¹¹B} NMR spectrum of [BnPh₃P][BF₄].



Figure S71: ^{31}P NMR spectrum of [BnPh_3P][BF_4].



Figure S72: ${}^{31}P{}^{1}H$ NMR spectrum of [BnPh₃P][BF₄].



Figure S73: ¹H NMR spectrum of [BnPh₃P][BF₃(CN)].



Figure S74: ¹³C{¹H} NMR spectrum of [BnPh₃P][BF₃(CN)].



Figure S75: ¹³C{¹⁹F} NMR spectrum of [BnPh₃P][BF₃(CN)].



Figure S76: ¹¹B NMR spectrum of [BnPh₃P][BF₃(CN)].





Figure S78: $^{19}F\{^{11}B\}$ NMR spectrum of [BnPh_3P][BF_3(CN)].

5.3.3 $[BnPh_3P][BF_2(CN)_2]$



Figure S79: ¹H NMR spectrum of [BnPh₃P][BF₂(CN)₂].



Figure S80: ¹³C{¹H} NMR spectrum of [BnPh₃P][BF₂(CN)₂].



Figure S81: ¹³C{¹⁹F} NMR spectrum of [BnPh₃P][BF₂(CN)₂].



Figure S82: ¹¹B NMR spectrum of [BnPh₃P][BF₂(CN)₂].



Figure S84: ¹⁹F NMR spectrum of [BnPh₃P][BF₂(CN)₂].



Figure S86: ${}^{31}P$ NMR spectrum of [BnPh₃P][BF₂(CN)₂].



Figure S87: $^{31}P\{^{1}H\}$ NMR spectrum of [BnPh_3P][BF_2(CN)_2].





Figure S88: ¹H NMR spectrum of [BnPh₃P][BF(CN)₃].



Figure S90: ¹³C{¹⁹F} NMR spectrum of [BnPh₃P][BF(CN)₃].



Figure S91: ¹¹B NMR spectrum of [BnPh₃P][BF(CN)₃].



Figure S92: ¹¹B{¹⁹F} NMR spectrum of [BnPh₃P][BF(CN)₃].





Figure S94: $^{19}F\{^{11}B\}$ NMR spectrum of [BnPh_3P][BF(CN)_3].



Figure S95: ${}^{31}P$ NMR spectrum of [BnPh₃P][BF(CN)₃].



Figure S96: ${}^{31}P{}^{1}H$ NMR spectrum of [BnPh₃P][BF(CN)₃].

5.3.5 [BnPh₃P][B(CN)₄]



Figure S97: ¹H NMR spectrum of [BnPh₃P][B(CN)₄].



Figure S98: ¹³C{¹H} NMR spectrum of [BnPh₃P][B(CN)₄].




Figure S100: ³¹P NMR spectrum of [BnPh₃P][B(CN)₄].



Figure S101: ³¹P{¹H} NMR spectrum of [BnPh₃P][B(CN)₄].









Figure S103: ${}^{1}H{}^{11}B{}$ NMR spectrum of [BnPh₃P][BFH(CN)₂].



Figure S104: ¹H{¹⁹F} NMR spectrum of [BnPh₃P][BFH(CN)₂].



Figure S105: ${}^{13}C{}^{1}H$ NMR spectrum of [BnPh₃P][BFH(CN)₂].



Figure S106: ${}^{13}C{}^{19}F{}$ NMR spectrum of [BnPh3P][BFH(CN)₂].



Figure S108: ¹¹B{¹H} NMR spectrum of [BnPh₃P][BFH(CN)₂].



Figure S110: ¹⁹F NMR spectrum of [BnPh₃P][BFH(CN)₂].



Figure S112: $^{19}F\{^{11}B\}$ NMR spectrum of [BnPh_3P][BFH(CN)_2].



Figure S113: ³¹P NMR spectrum of [BnPh₃P][BFH(CN)₂].



Figure S114: ${}^{31}P{}^{1}H$ NMR spectrum of [BnPh₃P][BFH(CN)₂].

5.3.7 [BnPh₃P][BH₃(CN)]



Figure S116: ¹H{¹¹B} NMR spectrum of [BnPh₃P][BH₃(CN)].



Figure S118: ¹¹B NMR spectrum of [BnPh₃P][BH₃(CN)].



Figure S120: ${}^{31}P$ NMR spectrum of [BnPh₃P][BH₃(CN)].



Figure S121: ³¹P{¹H} NMR spectrum of [BnPh₃P][BH₃(CN)].





Figure S122: ¹H NMR spectrum of [BnPh₃P][BH₂(CN)₂].



Figure S124: ${}^{13}C{}^{1}H$ NMR spectrum of [BnPh₃P][BH₂(CN)₂].



Figure S125: ¹¹B NMR spectrum of [BnPh₃P][BH₂(CN)₂].



Figure S126:¹¹B{¹H} NMR spectrum of [BnPh₃P][BH₂(CN)₂].



Figure S127: ³¹P NMR spectrum of $[BnPh_3P][BH_2(CN)_2]$.



Figure S128: ${}^{31}P{}^{1}H$ NMR spectrum of [BnPh₃P][BH₂(CN)₂].

5.3.9 [BnPh₃P][BH(CN)₃]



Figure S130: ¹H{¹¹B} NMR spectrum of [BnPh₃P][BH(CN)₃].



Figure S132: ${}^{11}B{}^{1}H{}$ NMR spectrum of [BnPh₃P][BH(CN)₃].



Figure S133: ¹³C{¹H} NMR spectrum of [BnPh₃P][BH(CN)₃].



Figure S134: ${}^{31}P$ NMR spectrum of [BnPh₃P][BH(CN)₃].



Figure S135: ³¹P{¹H} NMR spectrum of [BnPh₃P][BH(CN)₃].

6 IR and Raman Spectra

6.1 [EMIm] salts



Figure S136: IR and Raman spectra of [EMIm][BF₄].



Figure S137: IR and Raman spectra of [EMIm][BF₃(CN)].



Figure S139: IR and Raman spectra of [EMIm][BF(CN)₃].



Figure S140: IR and Raman spectra of [BMPL][BF₃(CN)].



Figure S141: IR and Raman spectra of $[BMPL][BF_2(CN)_2]$.



 $A_{\rm IR}$

¥

Figure S143: IR and Raman spectra of [BMPL][BFH(CN)₂].

6.3 Phosphonium salts



Figure S144: IR and Raman spectra of [BnPh₃P][BF₄].



Figure S145: IR and Raman spectra of [BnPh₃P][BF₃(CN)].



Figure S147: IR and Raman spectra of $[BnPh_3P][BF(CN)_3]$.





Figure S149: IR and NMR spectra of $[BnPh_3P][BFH(CN)_2]$.



Figure S150: IR and Raman spectra of [BnPh₃P][BH₃(CN)].



Figure S151: IR and Raman spectra of [BnPh₃P][BH₂(CN)₂].



Figure S152: IR and Raman spectra of [BnPh₃P][BH(CN)₃]

7 Electrochemical Measurements

All electrochemical studies were performed with the neat ILs under an Ar atmosphere with a Metrohm PGSTAT30 potentiostat and a Microcell HC set-up with a Eurotherm temperature controller (rhd instruments). A 0.1 mL Pt-cell TSC-70 closed (rhd instruments) served as counter electrode and was equipped with a glassy carbon working electrode (surface area: $3.14 \cdot 10^{-2}$ cm²). Specific conductivities (σ) were determined at different temperatures (20, 40, 60, 70, 80 °C) by impedance spectroscopy from 500·10³ to 800 Hz. The cell constant was determined on a 1413 µS·cm⁻¹ conductivity solution HI 70031 (HANNA instruments).

Cyclic voltammetry was conducted at 20 °C with a scan rate of 50 mV·s⁻¹ using the same setup as employed for the conductivity measurements and an additional Ag/Ag⁺ micro reference electrode (acetonitrile, rhd instruments). Cathodic (E_c) and anodic (E_a) limits of stability have been assessed via linear fits below and above the onset of electrolyte decomposition and determination of the intersection point of both lines in analogy to a method described in the literature.^[19,20]

Т/К	293.1	313.1	333.1	343.1	353.1
[EMIm][B(CN) ₄]	12.1	20.7	31.0	36.6	42.4
[EMIm][BF(CN) ₃]	20.2	29.7	41.4	47.5	54.0
[EMIm][BF ₂ (CN) ₂]	25.0	35.0	45.9	51.7	58.2
[EMIm][BF ₃ (CN)]	22.9	n.d.	n.d.	n.d.	n.d.
[EMIm][BF ₄]	12.3	20.9	30.7	36.1	41.7
[EMIm][BH(CN) ₃]	20.1	30.6	42.5	49.0	55.7
[EMIm][BH ₂ (CN) ₂]	32.6	46.4	61.1	69.5	n.d.
[EMIm][BH ₃ (CN)]	26.6	40.2	54.2	62.8	n.d.
[EMIm][BFH(CN) ₂]	27.6	38.8	50.1	55.6	61.6
[BMPL][B(CN) ₄]	4.1	7.6	12.4	15.2	18.2
[BMPL][BF(CN) ₃]	7.7	12.1	17.4	20.2	23.0
[BMPL][BF ₂ (CN)2]	8.3	13.5	20.0	23.6	27.5
[BMPL][BF ₃ (CN)]	6.3	n.d.	n.d.	n.d.	n.d.
[BMPL][BFH(CN) ₂]	11.2	17.0	23.8	27.3	31.1

Table S1: Specific conductivity^a σ determined via impedance spectroscopy at different temperatures.

[a] Specific conductivity σ in mS·cm⁻¹. n.d.: not determined

8 Viscosity and Density

Table S2: Dynamic viscosity and density of ILs in the temperature range of 293–353 K at 60° angle.

Species	ρ / g·cm ⁻³								η / mPa·s							
<i>т /</i> к	293	303	313	323	333	343	353	293	303	313	323	333	343	353		
[EMIm][B(CN) ₄]	1.040	1.033	1.025	1.017	1.009	1.002	0.994	22.6	15.7	11.4	8.7	7.0	5.7	4.7		
[EMIm][BF(CN) ₃]	1.073	1.065	1.057	1.050	1.042	1.035	1.028	12.6	9.3	7.5	6.0	5.0	4.2	3.5		
[EMIm][BF ₂ (CN) ₂]	1.120	1.112	1.104	1.097	1.090	1.082	1.075	10.8	8.5	6.8	5.7	4.8	4.1	3.5		
[EMIm][BF ₃ (CN)]	1.191	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	14.6	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.		
[EMIm][BF ₄]	1.286	1.278	1.271	1.263	1.256	1.248	1.241	43.7	29.9	22.2	16.7	13.0	10.4	8.5		
[EMIm][BH(CN)₃]	1.013	1.006	0.999	0.992	0.985	0.979	0.972	12.3	9.2	7.2	5.8	4.7	3.9	3.4		
[EMIm][BH ₂ (CN) ₂]	0.981	0.975	0.969	0.969	0.957	0.950	0.945	10.2	7.9	6.4	5.3	4.4	3.8	3.3		
[EMIm][BH₃(CN)]	0.948	0.943	0.937	0.932	0.926	0.921	0.916	15.7	11.8	9.2	7.4	6.0	5.0	4.3		
[EMIm][BFH(CN) ₂]	1.056	1.049	1.042	1.035	1.028	1.022	1.015	10.6	8.3	6.7	5.5	4.6	3.9	3.4		
[BMPL][B(CN) ₄]	0.979	0.973	0.966	0.959	0.952	0.946	0.939	60.2	39.8	27.4	19.9	15.1	11.7	9.3		
[BMPL][BF(CN) ₃]	1.004	0.998	0.991	0.984	0.978	0.972	0.965	30.7	22.1	16.6	13.0	10.6	8.6	7.1		
[BMPL][BF ₂ (CN) ₂]	1.039	1.033	1.026	1.020	1.013	1.007	1.001	28.2	21.6	16.6	13.0	10.6	8.7	7.3		
[BMPL][BF ₃ (CN)]	1.089	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	57.7	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.		
[BMPL][BFH(CN) ₂]	0.987	0.981	0.975	0.969	0.963	0.957	0.952	25.2	19.1	14.8	11.7	9.5	7.9	6.6		
[BMIm][BH ₂ (CN) ₂]	0.962	0.956	0.950	0.944	0.938	0.932	0.926	18.3	13.3	10.1	8.0	6.4	5.3	4.5		

n.d.: not determined.

9 Vogel-Fulcher-Tammann (VFT), Arrhenius, and Litovitz Fits

The dynamic viscosity (η) and the specific conductivity (σ) at different temperatures (T) were fitted to the following equations (fitting parameters see below).

Vogel-Fulcher-Tammann (VFT):^[8]

$$\eta = \eta_0 \exp\left(\frac{k_\eta}{T - T_0}\right) \tag{1}$$

with η_0 = viscosity at infinite temperature; k_{η} = constant; $T_0 T_0$ = Vogel temperature.

$$\sigma = \sigma_0 \exp\left(\frac{-k_\sigma}{T - T_0'}\right) \tag{2}$$

With σ_0 = conductivity at infinite temperature; k_σ = constant.

Arrhenius:^[9]

$$\sigma = \sigma_0 \exp\left(\frac{-E_\sigma}{RT}\right) \tag{3}$$

with E_{σ} = activation energy of conductivity; R = universal gas constant.

Litovitz:^[10]

$$\sigma = \sigma_0 \exp\left(\frac{-B}{R T^3}\right) \tag{4}$$

with *B* = constant.

		VFT parameters $(\eta)^a$		VFT parameters (σ) with variable T_0' b			VFT parameters (σ) fixed T_0' ^c		Arrhenius parameters $(\eta)^d$		Arrhenius parameters (σ) ^e		Litovitz parameters $(\eta)^{f}$		Litovitz parameters (σ) ^g		
		η_0	kη	T ₀	σ_0	k_{σ}	T ₀ ʻ	σ_0	k_{σ}	η_{0}	En	σ_0	Eσ	η_0	В	σ_{0}	В
		[10 ⁻³ mPa⋅s]	[K]	[K]	[mS·cm⁻¹]	[K]	[K]	[mS·cm ^{−1}]	К	[10 ⁻³ mPa⋅s]	[kJ·mol⁻¹]	[mS·cm ^{−1}]	[kJ·mol⁻¹]	[10⁻³mPa⋅s]	[10 ⁻⁶ J·K ² ·mol ⁻¹]	[mS·cm ^{−1}]	[10 ⁻⁶ J·K ² ·mol ⁻¹]
[EMIm]	[BF ₄]	264 ± 76	658 ± 84	164 ± 9	467 ± 29	433 ± 19	174 ± 3			2 ± 0.4	24 ± 0.5	12975 ± 2042	17 ± 0.4	973 ± 2	796 ± 6	210 ± 2	592 ± 3
	$[BF_2(CN)_2]$	180 ± 50	657 ± 101	133 ± 14	3961 ± 433	1481 ± 37	0 ± 0	607 ± 50	514 ± 16	13 ± 1	16 ± 0.3	3961 ± 395	12 ± 0.3	709 ± 19	597 ± 6	189 ± 10	428 ± 16
	[BF(CN) ₃]	134 ± 56	720 ± 150	134±19	806±88	564 ± 41	139 ± 7			7 ± 1	18 ± 0.3	7967 ± 697	14 ± 0.2	709 ± 19	597 ± 6	228 ± 2	505 ± 3
	[BFH(CN) ₂]	138 ± 25	730 ± 68	125±9	354±24	334 ± 22	162 ± 5			11 ± 1	17 ± 0.2	2869 ± 257	11 ± 0.2	768 ± 14	550 ± 4	178 ± 1	391 ± 1
[BMPL]	[BF ₂ (CN) ₂]	44 ± 35	1440 ± 382	70 ± 33	680 ± 31	708 ± 18	132 ± 2			8 ± 1	20 ± 0.2	8455 ± 787	17 ± 0.3	1277 ± 59	651 ± 11	137 ± 2	590 ± 4
	[BF(CN) ₃]	318 ± 62	592 ± 58	163 ± 7	273 ± 27	479 ± 33	159 ± 6			4 ± 1	21 ± 0.5	4449 ± 538	15 ± 0.3	1041 ± 18	708 ± 4	101 ± 1	541 ± 2
	[BFH(CN) ₂]	88 ± 40	1084 ± 192	101 ± 19	289 ± 24	526 ± 22	145 ± 4			8±1	20 ± 0.2	4190 ± 391	14 ± 0.2	1167 ± 39	646 ± 8	122 ± 1	503 ± 2

Table S3: Fitting parameters for Vogel-Fulcher-Tammann (VFT), Arrhenius and Litovitz equations describing the behaviour of [EMIm]⁺ and [BMPL]⁺ salts of fluorocyanoborate anions in the temperature range of 293–353 K.

[a] $R^2 \ge 99.96\%$. [b] $R^2 \ge 99.99\%$. [c] T_0' was taken from the VFT(η) fit (4th column) and was used as an invariable constant ($T_0'=T_0$); $R^2 = 99.47$. [d] $R^2 \ge 99.86\%$. [e] $R^2 \ge 99.67\%$. [f] $R^2 \ge 99.93\%$.

[g] $R^2 ≥ 99.27\%$.

10 Diffusion NMR

All diffusion (DOSY) NMR measurements were performed on a 14.1 T Bruker Avance III Spectrometer with a BCU 2 cooling unit using a BBFO probe equipped with a z-gradient (max. gradient strength of 50 G cm⁻¹). The ionic liquids were filled into coaxial insert tubes with the insert being filled with DMSO-d₆. No sample rotation was employed.

Each sample was measured at three different temperatures, 293.1 K, 313.1 K and 333.1 K, which were calibrated prior to the measurement series. ¹H, ¹¹B and ¹⁹F 90° pulses were calibrated for each sample and temperature. Furthermore, diffusion times Δ and durations of the bipolar gradient pulse δ were estimated based on the known viscosities and existing data in literature^[11] and subsequently optimized for each sample prior to the experiment. The DOSY measurements were done using both the stimulated echo BPP-LED pulse (ledbpgp2s) sequence and the double stimulated echo pulse (dstebpgp3s) sequence. By comparison of the two datasets, convection effects could be excluded. Signals had decayed to below 10 % or, if possible, to below 5 % of the initial signal area. The analysis of the DOSY data was done using the Topspin T1/T2 module with the area option yielding the temperature dependent diffusion coefficients for each nucleus and thus for the different components of the ionic liquid.



Figure S153: Signal decay as function of gradient strength G for ${}^{1}H$ (left) and ${}^{19}F$ (right) DOSY NMR experiments for [EMIm][BF(CN)₃], [EMIm][BF₂(CN)₂] and [EMIm][BF₃(CN)].



Figure S154: Signal decay as function of gradient strength G for ¹H (left) and ¹⁹F (right) DOSY NMR experiments for [EMIm][BF4] and [EMIm][BFH(CN)₂].



 $\label{eq:BMPL} [B(CN)_4], [BMPL][BF(CN)_3] \mbox{ and } [BMPL][BF_2(CN)_2].$


Figure S156: Signal decay as function of gradient strength *G* for ¹H (left) and ¹⁹F (right) DOSY NMR experiments for [BMPL][BF₃(CN)] and [BMPL][BFH(CN)₂].

Т/К		293	3.1			31	13.1			33	3.1	
	D+		D⁻		D+		D⁻		D+		D⁻	
nucleus	¹ H	¹ H	¹⁹ F	¹¹ B	¹ H	¹ H	¹⁹ F	¹¹ B	¹ H	¹ H	¹⁹ F	¹¹ B
[EMIm][B(CN) ₄]	6.3			5.4	13.3			11.8	22.5			20.1
[EMIm][BF(CN) ₃]	10.26 ^b		9.62 ^b	n.d.	18.96 ^b		17.78 ^b	n.d.	29.53 ^b 29.30 ^c		27.69 ^b 27.73 ^c	27.76 ^b
[EMIm][BF ₂ (CN) ₂]	12.21 ^b		11.73 ^b	n.d.	21.13 ^b 21.31 ^c		[22.22] ^{bd} 20.42 ^c	n.d.	31.71 ^b 32.17 ^c		30.15 ^b 30.36 ^c	30.24 ^b 30.45 ^c
[EMIm][BF ₃ (CN)]	10.04 ^b		9.03 ^b	n.d.	17.50 ^b 17.58 ^c		15.93 ^b 16.07 ^c	n.d.	26.47 ^b 26.61 ^c		24.30 ^b 24.48 ^c	24.37 ^b 24.46 ^c
[EMIm][BF ₄]	3.86 ^b		2.95 ^b	n.d.	8.21 ^b 8.35 ^c		6.58 ^b 6.60 ^c	n.d.	14.13 ^b 14.22 ^c		11.64 ^b 11.63 ^c	11.67 ^b 11.56 ^c
[EMIm][BH(CN) ₃]	1.05 ^b	1.03 ^b			1.94 ^b	1.91 ^b			3.06 ^b 3.06 ^c	2.94 ^b 2.93 ^c		
[EMIm][BH ₂ (CN) ₂]	1.33 ^b	1.28 ^b			2.30 ^b	2.23 ^b			3.44 ^b 3.42 ^c	3.36 ^b 3.38 ^c		
[EMIm][BH ₃ (CN)]	0.94 ^b	0.89 ^b			1.73 ^b	1.66 ^b			2.71 ^b 2.70 ^c	2.63 ^b 2.66 ^c		
[EMIm][BFH(CN) ₂]	12.53 ^b	12.39 ^b	12.29 ^b	n.d.	21.70 ^b 21.80 ^c	21.25 ^b 21.25 ^c	21.25 ^b 21.30 ^c	21.34 ^b 21.48 ^c	32.64 ^b 33.05 ^c	31.93 ^b 32.06 ^c	31.79 ^b 31.81 ^c	31.85 ^b 32.00 ^c
[BMPL][B(CN) ₄]	1.96 ^b			2.21 ^b 2.23 ^c	4.81 ^b 4.84 ^c			5.40 ^b 5.43 ^c	9.01 ^b 9.05 ^c			10.04 ^b 10.00 ^c
[BMPL][BF(CN) ₃]	3.79 ^b		4.46 ^b	n.d.	7.75 ^b		9.41 ^b	n.d.	13.02 ^b 13.13 ^c		15.59 ^b 15.66 ^c	n.d.
[BMPL][BF ₂ (CN) ₂]	4.21 ^b		5.11 ^b	n.d.	8.01 ^b 8.04 ^c		9.71 ^b 9.78 ^c	n.d.	12.89 ^b 12.96 ^c		15.52 ^b 15.62 ^c	15.55 ^b 15.58 ^c
[BMPL][BF ₃ (CN)]	2.42 ^b		2.88 ^b	n.d.	5.01 ^b 5.01 ^c		5.99 ^b 6.04 ^c	n.d.	8.62 ^b 8.65 ^c		10.30 ^b 10.33 ^c	10.32 ^b 10.37 ^c
[BMPL][BFH(CN) ₂]	4.57 ^b	5.52 ^b	5.62 ^b	n.d	8.69 ^b 8.81 ^c	10.67 ^b 9.967 ^c	10.59 ^b 10.68 ^c	n.d.	14.97 ^b 14.07 ^c	16.98 ^b 17.31 ^c	17.02 ^b 17.04 ^c	17.03 ^b 17.20 ^c

Table S4: Diffusion coefficients^a D⁺ and D⁻ determined via DOSY NMR at different temperatures.

[a] Diffusion coefficients D⁺ and D⁻ in 10⁻¹¹m²·s⁻¹. [b] ledbpgp2s pulse sequence. [c] dstebpgp3s pulse sequence. [d] Unexpectedly high value; thus, omitted from discussion. n.d.: not determined.

11 Overview physical propterties of RTILs

Table S5: Selected physical properties^a of [EMIm]⁺ and [BMPL]⁺ RTILs in the temperature range of 293–333 K.

Т	salt	М	ρ	σ	D+	D⁻	$\Lambda_{\rm NMR}$	Λ_{imp}	1
К		g·mol⁻¹	g∙cm ⁻³	mS∙cm⁻¹	10 ⁻¹¹ m ² ·s ⁻¹	cm²⋅S mol ⁻¹	cm²·S·mol⁻¹	cm²·S·mol⁻¹	
	[EMIm][B(CN) ₄]	226.05	1.040	12.1	6.3	5.4	4.45	2.63	0.59
	[EMIm][BF(CN) ₃]	219.03	1.073	20.2	10.26	9.62	7.56	4.12	0.54
	[EMIm][BF ₂ (CN) ₂]	212.01	1.120	25.0	12.21	11.73	9.10	4.73	0.52
	[EMIm][BF ₃ (CN)]	204.99	1.191	22.9	10.04	9.03	7.25	3.94	0.54
	[EMIm][BF ₄]	197.97	1.286	12.3	3.86	2.95	2.59	1.89	0.73
2	[EMIm][BH(CN) ₃]	201.04	1.013	20.1	10.5	10.3	7.91	3.99	0.50
93.)	[EMIm][BH ₂ (CN) ₂]	176.03	0.981	32.6	13.3	12.8	9.93	5.85	0.59
17	[EMIm][BH ₃ (CN)]	151.02	0.948	26.6	9.4	8.9	6.96	4.24	0.61
	[EMIm][BFH(CN) ₂]	194.02	1.056	27.6	12.53	12.34 ^b	9.46	5.07	0.54
	[BMPL][B(CN) ₄]	257.15	0.979	4.1	1.96	2.21	1.59	1.08	0.68
	[BMPL][BF(CN)₃]	250.13	1.004	7.7	3.79	4.46	3.14	1.92	0.61
	[BMPL][BF ₂ (CN) ₂]	243.11	1.039	8.3	4.21	5.11	3.54	1.08 0.68 1.92 0.61 1.94 0.55 1.37 0.68	0.55
	[BMPL][BF ₃ (CN)]	236.09	1.089	6.3	2.42	2.88	2.02	1.37	2.63 0.59 4.12 0.54 4.73 0.52 3.94 0.54 1.89 0.73 3.99 0.50 5.85 0.59 4.24 0.61 5.07 0.54 1.08 0.68 1.92 0.61 1.94 0.55 1.37 0.68 2.56 0.66 4.57 0.51 6.15 0.47 6.72 0.45 n.d. n.d. 3.26 0.62 6.16 0.45 8.43 0.52 6.48 0.54
	[BMPL][BFH(CN) ₂]	225.12	0.987	11.2	4.57	5.57 ^b	3.86	2.56	0.66
	[EMIm][B(CN) ₄]	226.05	1.025	20.7	13.3	11.8	8.94	4.57	0.51
	[EMIm][BF(CN) ₃]	219.03	1.057	31.6	18.96	17.78	13.08	6.15	0.47
	[EMIm][BF ₂ (CN) ₂]	212.01	1.104	35.0	21.13	20.42 ^c	14.79	6.72	0.45
ω	[EMIm][BF ₃ (CN)]	204.99	n.d.	n.d.	17.5	15.93	11.90	n.d.	n.d.
13.1	[EMIm][BF ₄]	197.97	1.271	20.9	8.21	6.58	5.27	3.26	0.62
17	[EMIm][BH(CN) ₃]	201.04	0.999	30.6	19.4	19.1	13.71	6.16	0.45
	[EMIm][BH ₂ (CN) ₂]	176.03	0.969	46.4	23.0	22.3	16.13	8.43	0.52
	[EMIm][BH ₃ (CN)]	151.02	0.937	40.2	17.3	16.6	12.07	6.48	0.54
	[EMIm][BFH(CN) ₂]	194.02	1.042	38.8	21.7	21.28 ^b	15.30	7.23	0.47

	[BMPL][B(CN) ₄]	257.15	0.966	7.6	4.81	5.4	3.63	2.02	0.56
	[BMPL][BF(CN) ₃]	250.13	0.991	12.1	7.75	9.41	6.11	3.05	0.50
	[BMPL][BF ₂ (CN) ₂]	243.11	1.026	13.5	8.01	9.71	6.31	3.20	0.51
	[BMPL][BF ₃ (CN)]	236.09	n.d.	n.d.	5.01	5.99	3.92	n.d.	n.d.
	[BMPL][BFH(CN) ₂]	225.12	0.975	17	8.69	10.63 ^b	6.88	3.93	0.57
	[EMIm][B(CN) ₄]	226.05	1.009	31	22.5	20.1	14.26	6.95	0.49
	[EMIm][BF(CN)₃]	219.03	1.042	41.4	29.53	27.73 ^b	19.16	8.70	0.45
	[EMIm][BF ₂ (CN) ₂]	212.01	1.090	45.9	31.71	30.2 ^b	20.72	8.93	0.43
	[EMIm][BF₃(CN)]	204.99	n.d.	n.d.	26.47	24.34 ^b	17.00	n.d.	n.d.
	[EMIm][BF ₄]	197.97	1.256	30.7	14.13	11.66 ^b	8.63	4.84	0.56
	[EMIm][BH(CN) ₃]	201.04	0.985	44.5	30.6	29.4	20.08	8.67	0.43
3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3 3	[EMIm][BH ₂ (CN) ₂]	176.03	0.957	61.1	34.4	33.6	22.76	11.24	0.49
8.17	[EMIm][BH ₃ (CN)]	151.02	0.926	54.2	27.1	26.3	17.87	8.84	0.49
	[EMIm][BFH(CN) ₂]	194.02	1.028	50.1	32.64	31.86 ^b	21.58	9.45	0.44
	[BMPL][B(CN) ₄]	257.15	0.952	12.4	9.01	10.04	6.38	3.35	0.53
	[BMPL][BF(CN) ₃]	250.13	0.978	17.4	13.02	15.59	9.57	4.45	0.46
	[BMPL][BF ₂ (CN) ₂]	243.11	1.013	20	12.89	15.54 ^b	9.51	4.80	0.50
	[BMPL][BF ₃ (CN)]	236.09	n.d.	n.d.	8.62	10.31 ^b	6.33	n.d.	n.d.
	[BMPL][BFH(CN) ₂]	225.12	0.963	23.8	14.97	17.01 ^b	10.70	5.56	0.52

[a] molar mass M; density ρ ; specific conductivity σ measured via impedance spectroscopy; diffusion coefficients of cations (D⁺) and anion (D⁻) obtained with ledbpgp2 pulse sequence when not otherwise stated; molar conductivities calcd. by $\Lambda_{imp} = (D^++D^-)\cdot N_A\cdot k^{-1}\cdot T^{-1}$ and $\Lambda_{imp} = \sigma\cdot M\cdot \rho^{-1}$; ionicity $I = \Lambda_{imp}\cdot \Lambda_{NMR}^{-1}$. [b] mean values over all measured nuclei, see table S4 for details. [c] obtained with dstebpgp3s pulse sequence. n.d.: not determined.

12 Cytotoxicity

Cytotoxicity of selected [EMIm]⁺ and [BMPL]⁺ RTILs as well as cyano(fluoro)borate and cyano(hydrido)borate anions including the anion [BFH(CN)₂]⁻ (FHB) with sodium and potassium as counterion was conducted by formazan assay. The number of viable cells is determined by the cleavage of a tetrazolium salt into formazan by cellular enzymes. A decrease in the number of viable cells leads to a decrease in mitochondrial dehydrogenase and thus to lower formazan values. Cyano(fluoro)borate, cyano(hydrido)borate and FHB salts were dissolved in cell culture medium (DMEM + 10% FCS + 1% P/S) to a final concentration of 100 mM. EMIm- and BMPL-RTILs were dissolved in cell culture medium (DMEM + 10% FCS + 1% P/S) containing 2% DMSO to a final concentration of 100 mM. HEK 293 (4x104 cells/mL, 100 µL per well) and NIH3T3 (3x104 cells/mL, 100 µL per well) were seeded in a 96-well plate. Dilution series of each salt/RTIL ranging from 100 µM to 256 nM were prepared in growth medium and added (1:1 [V/V]) to the cells. A dilution series with DMSO was used as a solvent control for RTILs. Cells were incubated for 24 h at 37 °C and 5% CO₂, respectively. After incubation, 10 µL WST-1 reagent was added to each well and cells were incubated for up to four hours at 37 °C according to manufacturer instructions. Every 60 minutes, cells were analyzed and the absorbance of the soluble formazan product at 450 nm as well as background noise at 630 nm was determined using an Infinite M Plex microplate reader (Tecan Group Ltd., Männedorf, Switzerland). Relative cell viability (proliferation) with respect to untreated sample is expressed.



Figure S157: Cytotoxic evaluation using NIH3T3 (A,B) and HEK293 (C,D) cells of selected cyano(fluoro)borate and cyano(hydrido)borate anions including the anion [BFH(CN)₂] (FHB) with sodium and potassium as counterion.



Figure S158: Cytotoxic evaluation using HEK293 cells of selected EMIm- and BMPL-RTILs.

13 Thermal Properties

13.1 [EMIm] salts







Figure S160: DSC curve of [EMIm][BF₃(CN)].



Figure S161: DSC curve of [EMIm][BF₂(CN)₂].



Figure S162: DSC curve of [EMIm][BFH(CN)₂].

13.2 [BMPL] salts



Figure S163: DSC curve of [BMPL][BF₄].



Figure S164: DSC curve of [BMPL][BF₃(CN)].



Figure S165: DSC curve of [BMPL][BF₂(CN)₂].



Figure S166: DSC curve of [BMPL][BF(CN)₃].



Figure S167: DSC curve of [BMPL][BFH(CN)₂].



13.3 [BnPh₃P] salts

Figure S168: DSC curve of $[BnPh_3P][BF_4]$.



Figure S169: DSC curve of [BnPh₃P][BF₃(CN)].



Figure S170: DSC curve of [BnPh₃P][BF₃(CN)].



Figure S171: DSC curve of [BnPh₃P][BF₂(CN)₂].



Figure S172: DSC curve of [BnPh₃P][BF(CN)₃].



Figure S173: [BnPh₃P][B(CN)₄].



Figure S174: DSC curve of [BnPh₃P][BFH(CN)₂].



Figure S175: DSC curve of [BnPh₃P][BH₂(CN)₂].



Figure S176: DSC curve of [BnPh₃P][BH(CN)₃].

14 Quantum Chemical Calculations

Density functional calculations (DFT)^[12] were carried out using the B3LYP functional^[13] with the Gaussian16 program suite.^[14] Geometries were optimized, and energies were calculated with the 6-311++G(d,p) or def2-TZVPP basis sets. Diffuse functions were incorporated because improved energies are obtained for anions.^[15] All calculated structures represent true minima with no imaginary frequency on the respective hypersurface. Calculated energies are summarized in Tables S6a and S6b.

Species	Symmetry	<i>EE /</i> a.u.	<i>E</i> ₀ / a.u.	<i>H /</i> a.u.	G / a.u.
[BF ₄] ⁻	T _d	-424.679694	-424.666025	-424.660623	-424.691361
[BF₃(CN)] ⁻	C _{3v}	-417.636362	-417.616753	-417.610210	-417.645096
$[BF_2(CN)_2]^-$	<i>C</i> _{2v}	-410.588999	-410.563414	-410.555736	-410.593802
[BF(CN)₃] [−]	C _{3v}	-403.545131	-403.513328	-403.504586	-403.544877
[BFH(CN) ₂] [−]	Cs	-311.247313	-311.215304	-311.208446	-311.245063
[B(CN) ₄] ⁻	T _d	-396.509584	-396.471239	-396.461507	-396.502909
[BH₃(CN)]⁻	C _{3v}	-119.605662	-119.569409	-119.564645	-119.592633
[BH ₂ (CN) ₂] [−]	C _{2v}	-211.920506	-211.882496	-211.876420	-211.909823
[BH(CN)₃]⁻	C _{3v}	-304.221244	-304.182611	-304.174844	-304.212768

Table S6a: Calculated electronic energies (EE), energies with zero-point corrections (E₀), enthalpies (H), and free energies(G) at the B3LYP/6-311+G(d,p) level of theory.

Table S7b: Calculated energies with zero-point corrections (E), enthalpies (H),and free energies(G) at the B3LYP/def2-TZVPP level of theory.

Species	Symmetry	<i>E</i> [au]	<i>H</i> [au]	<i>G</i> [au]
BH ₃	D _{3h}	-26.595392	-26.594448	-26.615819
BF₃	D _{3h}	-324.698335	-324.697391	-324.726320
[BF ₄] ⁻	T _d	-424.715786	-424.714842	-424.745480
BF ₂ (CN)	<i>C</i> _{2<i>v</i>}	-317.620151	-317.619207	-317.652104
[BF₃(CN)] [–]	C _{3v}	-417.662672	-417.661728	-417.696568
BF(CN) ₂	<i>C</i> _{2<i>v</i>}	-310.538127	-310.537183	-310.573036
[BF ₂ (CN) ₂] ⁻	<i>C</i> _{2<i>v</i>}	-410.604844	-410.603900	-410.642603
B(CN)₃	D _{3h}	-303.462308	-303.461364	-303.499016
[BF(CN)₃] [−]	C _{3v}	-403.550137	-403.549193	-403.589490
BH ₂ (CN)	<i>C</i> _{2v}	-118.891448	-118.890503	-118.918544
[BFH₂(CN)] [−]	Cs	-218.924684	-218.923740	-218.955458
BH(CN) ₂	<i>C</i> _{2v}	-211.179866	-211.178921	-211.212376
[BFH(CN)₂] [−]	Cs	-311.243651	-311.242706	-311.279301
BFH(CN)	Cs	-218.253804	-218.252860	-218.284207
[BF₂H(CN)] [−]	Cs	-318.289004	-318.288060	-318.322085
[B(CN) ₄] ⁻	T _d	-396.503680	-396.502736	-396.544123
[BH(CN)₃] [_]	C _{3v}	-304.207085	-304.206141	-304.245093
$[BH_2(CN)_2]^-$	<i>C</i> _{2v}	-211.898844	-211.897899	-211.931289
[BH₃(CN)] [–]	<i>C</i> _{3v}	-119.577474	-119.576530	-119.604493
CN [−]	<i>C</i> ₃ <i>v</i>	-92.885370	-92.884426	-92.906762

15 Single Crystal X-Ray Diffraction

Single-crystals for $[BnPh_3P][BF_4]$, $[BnPh_3P][BF_3(CN)]$, $[BnPh_3P][BF_2(CN)_2]$, $[BnPh_3P][BF(CN)_3]$, $[BnPh_3P][B(CN)_4]$, $[BnPh_3P][BFH(CN)_2]$, $[BnPh_3P][BH_3(CN)]$, $[BnPh_3P][BH_2(CN)_2]$ and $[BnPh_3P][BH(CN)_3]$ were studied on a XtaLAB Synergy Dualflex, HyPix diffractometer using Cu-K_a radiation (micro-focus sealed X-ray tube, $\lambda = 1.54184$ Å).

All structures were solved by intrinsic phasing methods (SHELXT) and refinement is based on full-matrix least-squares calculations on F^2 (SHELXL).^[16] All non-hydrogen atoms were refined anisotropically. For CH idealized bond lengths and angles were used. Calculations were carried out using the ShelXle graphical interface.^[17] Molecular structure diagrams were drawn with the program Diamond 4.6.4.^[18] Crystallographic data have been deposited with the Cambridge Crystallographic Data Centre. Experimental details, crystal data, and CCDC numbers are collected in Table S4. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via <u>www.ccdc.cam.ac.uk/data_request/cif</u>

Compound	[BnPh ₃ P][BF ₄]	[BnPh ₃ P][BF ₃ (CN)]	[BnPh ₃ P][BF ₂ (CN) ₂]	[BnPh ₃ P][BF(CN) ₃]	[BnPh ₃ P][BFH(CN) ₂]
Chemical formula	$C_{25}H_{22}BF_4P$	C ₂₆ H ₂₂ BF ₃ NP	$C_{27}H_{22}BF_2N_2P$	C ₂₈ H ₂₂ BFN ₃ P	C ₂₇ H ₂₃ BFN ₂ P
M [g·mol⁻¹]	440.20	447.22	454.24	461.26	436.25
<i>T</i> [K]	100(2)	100(2)	100(2)	100(2)	100(2)
Crystal system	monoclinic	triclinic	monoclinic	orthorhombic	triclinic
Space group	P21/c	ΡĪ	P21/c	Pbca	ΡĪ
a [Å]	9.4958(1)	10.1902(1)	13.3421(1)	13.4477(1)	9.89780(10)
<i>b</i> [Å]	21.5696(2)	13.3195(1)	17.2505(2)	17.6586(2)	21.7519(2)
<i>c</i> [Å]	11.1131(1)	18.9127(2)	20.2011(2)	20.6374(2)	21.8961(3)
α [°]	90	70.363(1)	90	90	92.6690(10)
в [°]	107.745(1)	83.801(1)	93.974(1)	90	90.0760(10)
γ [°]	90	69.417(1)	90	90	92.7650(10)
Volume [ų]	2167.90(4)	2263.21(4)	4638.26(8)	4900.71(8)	4703.52(9)
Ζ	4	4	8	8	8
D _{calc} [Mg⋅m ⁻³]	1.349	1.313	1.301	1.250	1.232
μ [mm ⁻¹]	1.508	1.403	1.329	1.219	1.226
<i>F</i> (000) [e]	912	928	1888	1920	1824
Reflections collected	23570	93511	17182	27804	98260
Independent reflections, R(int)	4549, 0.0397	9509, 0.0537	17182, - ^[d]	5137, 0.0358	19628, 0.0464
Parameters / data restraints	280 / 0	577 / 0	596 / 0	307 / 0	1170/6
R1[<i>I</i> >2σ(<i>I</i>)] ^[a]	0.0355	0.0402	0.0351	0.0348	0.0664
wR2 (all) ^[b]	0.0903	0.1115	0.0931	0.0908	0.1749
GooF on F ^{2[c]}	1.051	1.036	0.742	1.048	1.096
Largest diff. peak / hole [e·Å⁻³]	0.288 / -0.362	0.535 -0.474	0.370 / -0.327	0.280/-0.413	1.193 / -0.474
CCDC no.	2216851	2216850	2216854	2216856	2216852

Table S8: Selected crystal data and details of the refinement of the crystal structures of [BnPh₃P][BF₄], [BnPh₃P][BF₃(CN)], [BnPh₃P][BF₂(CN)₂], [BnPh₃P][BF(CN)₃] and [BnPh₃P][BFH(CN)₂].

 $\begin{aligned} &[a] R1 = (\Sigma ||F_0| - |F_c||)/\Sigma |F_0|. \ [b] \ wR2 = [\Sigma w (F_0^2 - F_c^2)^2 / \Sigma w (F_0^2)^2]^{0.5}, \ weight \ scheme: \ w = [\sigma^2 F_0 + (aP)^2 + bP]^{-1}; \ P = [max (0, F_0^2 + 2F_c^2]/3 \ [BnPh_3P][BF_4]: \ a = 0.0389, b = 1.058; \\ &[BnPh_3P[BF_3(CN)]: \ a = 0.0694, b = 0.7314; \ [BnPh_3P][BF_2(CN)_2]: \ a = 0.0625, b = 3.4393; \ [BnPh_3P][BF(CN)_3]: \ a = 0.044, b = 1.8917; \ [BnPh_3P][BFH(CN)_2]: \ a = 0.045, b = 2.1091. \\ &[c] GooF: S = [\Sigma w (F_0^2 - F_c^2)^2 / (m - n)]^{0.5}; \ (m = reflections, n = variables). \ [d] \ twin. \end{aligned}$

Compound	[BnPh ₃ P][B(CN) ₄]	[BnPh ₃ P][BH ₃ (CN)]	$[BnPh_3P][BH_2(CN)_2]$	[BnPh ₃ P][BH(CN) ₃]
Chemical formula	$C_{29}H_{22}BN_4P$	C ₂₆ H ₂₅ BNP	C ₂₇ H ₂₄ BN ₂ P	C ₂₈ H ₂₃ BN ₃ P
M [g·mol⁻¹]	468.28	393.25	418.26	443.27
<i>Т</i> [K]	100(2)	100(2)	100(2)	95(2)
Crystal system	triclinic	triclinic	monoclinic	orthorhombic
Space group	ΡĪ	PĪ	<i>P</i> 2 ₁ /n	Pbca
a [Å]	8.9003(1)	9.9936(1)	9.5349(2)	13.4269(1)
b [Å]	11.6360(2)	13.0398(1)	14.5833(3)	17.4850(1)
<i>c</i> [Å]	13.5576(2)	18.8613(1)	16.4465(3)	20.6372(2)
α [°]	111.524(1)	70.179(1)	90	90
β [°]	90.565(1)	83.275(1)	98.440(2)	90
γ [°]	100.936(1)	71.674(1)	90	90
Volume [ų]	1277.58(3)	2194.86(4)	2262.12(8)	4844.98(7)
Ζ	2	4	4	8
D _{calc} [Mg·m ⁻³]	1.217	1.190	1.228	1.215
μ [mm ⁻¹]	1.133	1.175	1.187	1.151
<i>F</i> (000) [e]	488	832	880	1856
Reflections collected	26896	87588	24216	27306
Independent reflections, R(int)	5356, 0.0377	9218, 0.0434	4738, 0.0462	5067, 0.0366
Parameters / data restraints	316/0	547/0	288 / 0	302 / 0
R1[I>2σ(I)] ^[a]	0.0362	0.0402	0.0382	0.0358
wR2 (all) ^[b]	0.0941	0.1095	0.1021	0.0921
GooF on <i>F</i> ^{2[c]}	1.088	1.025	1.043	1.030
Largest diff. peak / hole [e·Å⁻³]	0.374 / -0.469	0.623 / -0.414	0.352 / -0.373	0.310/-0.461
CCDC no.	2216858	2216853	2216855	2216857

Table S9: Selected crystal data and details of the refinement of the crystal structures of [BnPh₃P][B(CN)₄], [BnPh₃P][BH₂(CN)₂], [BnPh₃P][BH₂(CN)₂], [BnPh₃P][BH₂(CN)₂],

 $[a] R1 = (\Sigma ||F_0| - |F_c||) / \Sigma |F_0|. [b] wR2 = [\Sigma w (F_0^2 - F_c^2)^2 / \Sigma w (F_0^2)^2]^{0.5}, \text{ weight scheme: } w = [\sigma^2 F_0 + (aP)^2 + bP]^{-1}; P = [max (0, F_0^2 + 2F_c^2) / 3 [BnPh_3P][B(CN)_4]: a = 0.044, b = 0.4619; [BnPh_3P[BH_3(CN)]: a = 0.0615, b = 0.9481; [BnPh_3P][BH_2(CN)_2]: a = 0.0514, b = 0.8377; [BnPh_3P][BH(CN)_3]: a = 0.045, b = 2.1091. [c] GooF: S = [\Sigma w (F_0^2 - F_c^2)^2 / (m - n)]^{0.5}; (m = reflections, n = variables).$

	Anion	[BF ₄] ⁻				[BF ₃ (CN)] ⁻			$[BF_2(CN)_2]^-$			[BF(CN) ₃] ⁻			[B(CN)₄] [−]		
							2										
		mean			mean			mean			mean			mean			
		value	range	calc.	value	range	calc.	value	range	calc.	value	range	calc.	value	range	calc.	
	<i>d</i> (B-C)	/	/	/	1.643	1.641(2) - 1.645(2)	1.63938	1.632	1.629(2) - 1.635(3)	1.62565	1.609	1.6121(19)	1.60928	1.595	1.5971(18)	1.59558	
[Å]	<i>d</i> (B-F)	1.393	1.4117(17)	1.41698	1.3833	1.3969(17)	1.41088	1.387	1.383(2) - 1.391(2)	1.41179	1.3962(1 5)	/	1.4194	/	/	/	
	<i>d</i> (B-H)	/	/	/	/	/	/	/	/	/	/	/	/	/	/	/	
	d(C≡N)	/	/	/	1.141	1.137(2) - 1.145(2)	1.15795	1.144	1.143(2) - 1.145(2)	1.15722	1.144	1.1426(18) - 1.1459(18)	1.15646	1.146	1.1456(18) - 1.1466(18)	1.15554	
	∢(B-C-N)	/	/	/	178.99	178.59(15) - 179.39(17)	179.9984	178.093	176.95(18) - 179.45(19) 107.43(14) -	179.74986	178.6	178.07 - 179.62 107 33(10) -	179.2416	178.2	176.98(14) - 178.69(14) 108 13(10) -	180.00	
	∢(NC-B-CN)	/	/	/	/	/ 107 99(11) -	/	107.78	108.13(14)	109.514	108.6	109.25(10)	108.8852	109.5	110.95(10)	109.471	
	∢(NC-B-F)	/	/	/	108.77	109.26(13)	108.90703	109.52	110.75(14)	109.1795	331.10	110.69(10)	110.0511	/	/	/	
0	∢(NC-B-H)	/	/	/	/	/	/	/	/	/	/	/	/	/	/	/	
	≮(F-B-F)	109.5	107.56(11) - 110.72(12)	109.47122	110.17	109.23(12) - 110.73(13)	110.02946	110.88	110.73(15) - 111.04(15)	110.59308	/	/	/	/	/	/	
	∢(H-B-H)	/	/	/	/	/	/	/	/	/	/	/	/	/	/	/	
	∢(F-B-H)	/	/	/	/	/	/	/	/	/	/	/	/	/	/	/	

Table S10: Experimental and calculated bonding parameters of [BF₄]⁻, [BF₃(CN)]⁻, [BF₂(CN)₂]⁻, [BF(CN)₃]⁻ and [B(CN)₄]⁻.

	Anion		[BFH(CN)₂] [−]			$[BH_3(CN)]^-$			[BH ₂ (CN) ₂] ⁻			[BH(CN) ₃] ⁻ range 1.5884(19) - 1.589(2) / / 1.1456(18) - 1.1482(18) 178.03(14) - 179.30(15) 109.25(11) - 110.53(11) / 108.7(9) - 109.4(9) / /	
											G		•
		mean value	range	calc.	mean value	range	calc.	mean value	range	calc.	mean value	range	calculated
	<i>d</i> (B-C)	1.615	1.605(5) - 1.625(5)	1.60957	1.587	1.587(2) - 1.586(2)	1.58968	1.589	1.588(2) - 1.590(2)	1.58889	1.589	1.5884(19) - 1.589(2)	1.5908
[Å]	<i>d</i> (B-F)	1.406	1.386(4) - 1.421(4)	1.43586	/	/	/	/	/	/	/	/	1
[,1]	<i>d</i> (B-H)	1.15	1.09(4) - 1.23(3)	1.21624	1.130	1.10(2) - 1.15(2)	1.22354	1.12	1.08(2) - 1.15(2)	1.21455	1.110(17)	/	1.20998
	d(C≡N)	1.141	1.138(4) - 1.146(4)	1.1586	1.151	1.148(2) - 1.153(2)	1.16376	1.147	1.146(2) - 1.148(2)	1.16007	1.147	1.1456(18) - 1.1482(18)	1.15741
	∢(B-C-N)	178.7	178.0(3) - 179.3(4)	177.33701	179.52	179.34(18) - 179.69(19)	180	178.39	177.97(16) - 178.80(16)	177.4882	178.51	178.03(14) - 179.30(15)	177.81037
	∢(NC-B-CN)	107.1	105.3(3) - 107.9(3)	109.56447	/	/	1	110.03(13)	/	111.8709	109.94	109.25(11) - 110.53(11)	110.95751
	∢(NC-B-F)	109.6	108.0(3) - 110.7(3)	110.09776	/	/	1	1	/	/	/	/	/
٥	∢(NC-B-H)	109.7	106.1(16) - 114.3(15)	107.74600	109.07	107.4(12) - 111.0(11)	108.9381	108.88	107.8(11) - 110.8(10)	108.8356	108.9	108.7(9) - 109.4(9)	107.94056
	≮(F-B-F)	/	/	/	/	/	/ 109.9990	/	/	/	/	/	/
	∢(H-B-H)	/	/	/	109.85	106.3(17) - 113.9(16)		111.3(15)	/	109.6052	/	/	/
	∢(F-B-H)	111.0	109.9(16) - 113.(2)	111.51972	/	/	1	/	/	/	/	/	/

Table S11: Experimental and calculated bonding parameters of [BFH(CN)₂]⁻, [BH₃(CN)]⁻, [BH₂(CN)₂]⁻ and [BH(CN)₃]⁻.

16 Literature

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