Supplemental Information

Design, synthesis and evaluation of liver-targeting fluorescent probe for detecting mercury ion

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Figures and captions

Fig.S1 Fluorescence response of probes toward different metal ions. The Hg^{2+} aqueous solution of 50 μ M was added into the probes solution of 5 μ M (10 mM stock solution dissolved in DMSO, then diluted using water). Fluorescence determination of Rho-Lac (A) and Rho-Glu (C) via fluorescence spectrophotometer. Fluorescence response of Rho-Lac (B) and Rho-Glu (D) using Ultra-Violet irradiation.



Fig. S2 Fluorescent signal alterations of probes to Hg^{2+} with other ions. Metal ions competition of Rho-Lac (A) and Rho-Glu (C). Data was analyzed using a two-way RMANOVA test (different metal ions × addition of Hg^{2+} interaction). Rho-Lac, F (1, 136) = 249099.5 and P < 0.0001, for whether to add Hg^{2+} ; Rho-Glu, F (1, 136) = 277322 and P < 0.0001, for whether to add Hg^{2+} . Anions interference of Rho-Lac (B) and Rho-Glu (D). Data was analyzed using a one-way ANOVA test. P > 0.05, compared with Rho-Lac+ Hg^{2+} (P₁=0.061252, P₂=0.673622, P₃= 0.062108, P₄=0.062898, P₅=0.058265, P₆= 0.05968, P₇= 0.057582, P₈=0.050567); P > 0.05, compared with Rho-Glu+ Hg^{2+} (P₁=0.49376, P₂=0.173098, P₃=0.052019, P₄=0.05616, P₅= 0.190145, P₆=0.065849, P₇=0.052146, P₈= 0.09182). Anions interference for Rho-Gal (E), Rho-Lac (F) and Rho-Glu (G) using Ultra-Violet irradiation.



Fig. S3 Fluorescence alternations of Rho-Lac (A) and Rho-Glu (B) in different pH. The 5 μ M probes solution and 10 μ M Hg²⁺ solution were prepared with different pH value using HAc-NaOAc buffer at pH value of 2, 3, 4, 5, 6, 7, 8, and 9 respectively.



Fig. S4 Linearity between Hg²⁺ concentration and fluorescence intensity. The 5 μ M Rho-Gal(A), Rho-Lac (B) and Rho-Glu (C) probes solution were added with Hg²⁺ solution of 0.01, 0.05, 0.1, 0.25, 0.5, 1.0, 2.5, 5 μ M respectively to detect the fluorescence response value.



Fig. S5 Job's plot of Rho-Lac (A) and Rho-Glu (B) probes-Hg²⁺. The Hg²⁺, probes solution (10 mM stock solution in DMSO) were prepared and diluted using water to keep the total concentration of probes and Hg²⁺ solution at 10 μ M.



Fig. S6 Fluorescent cycle changes of Rho-Gal (A), Rho-Lac (B) and Rho-Glu (C) to Hg^{2+} with addition of S²⁻. The probes stock solution of 10 mM were prepared with DMSO, then diluted to 5 μ M using water. The Hg^{2+} and S²⁻ solution of 50 μ M also was dissolved in water.



Fig. S7 Fluorescence stability of probes toward Hg^{2+} . Hg^{2+} solution with 50 μ M was added into the 5 μ M probes solution, and the fluorescence response was continuously detected at 1 min interval within 40 min.



Fig. S8 Fluorescence response of Rho-Gal to Hg^{2+} in dead cells (scale bar 10 µm). Two copies of HepG2 cells fixed for 15 min were prepared. One was treated with 20 µM Rho-Gal solution at room temperature for 20 min, and incubated with 200 µM Hg²⁺ solution at room temperature for 15 min, then stained with DAPI for 5 min. Another was incubated with 20 µM Rho-Gal solution at 37 ° C for 20 min, incubated with 200 µM Hg²⁺ solution at 37 ° C for 15 min, and stained with DAPI for 5 min. Then the cellular fluorescence imaging was acquired.



Fig. S9 Intracellular fluorescence manners of Rho-Gal to Hg^{2+} . The scale bar is 10 µm. Igreen and Iblue is the fluorescence intensity of Rho-Gal+Hg²⁺ and DAPI respectively. Cellular fluorescence imaging in quintuplicate was acquired and data was analyzed using a one-way ANOVA, P<0.0001. (A) (B) Rho-Gal incubated with HepG2 cells for different time. HepG2 cells were incubated in incubator with 20 µM Rho-Gal solution dissolved in DMEM for 5, 10, 20 and 30 min, respectively, then incubated with 200 µM Hg²⁺ solution dissolved in DMEM for 15 min, fixed for 15 min, and treated with DAPI for 5 min. Cellular fluorescence imaging in quintuplicate was acquired. (C) (D) Rho-Gal reacted with different concentration of Hg²⁺ in HepG2 cells. HepG2 cells were incubated with 20 µM Rho-Gal dissolved in DMEM in incubator for 15 min, then incubated with Hg²⁺ solution dissolved in DMEM for 15 min, then incubated with Hg²⁺ solution dissolved in DMEM for 15 min, then incubated with Hg²⁺ solution dissolved in DMEM for 15 min, then incubated with Hg²⁺ solution dissolved in DMEM for 15 min, then incubated with Hg²⁺ solution dissolved in DMEM for 15 min, then incubated with Hg²⁺ solution dissolved in DMEM with different concentration of 20, 40, 100 and 200 µM in incubator for 15 min, fixed for 15 min and treated with DAPI for 5 min.

Characterization of compounds



The compound was gray white solid, yield 35%. ¹H NMR (600 MHz, CDCl₃) δ 7.92 (dd, J = 5.6, 3.0 Hz, 1H), 7.46 (dd, J = 5.6, 3.1 Hz, 2H), 7.09 (dd, J = 5.2, 3.2 Hz, 1H), 6.51 (d, J = 8.9 Hz, 2H), 6.40 (d, J = 2.4 Hz, 2H), 6.31 (dd, J = 8.9, 2.4 Hz, 2H), 3.51 – 3.47 (m, 2H), 3.36 (q, J = 7.0 Hz, 8H), 3.32 – 3.29 (m, 2H), 1.19 (t, J = 7.1 Hz,12H). ¹³C NMR (151 MHz, CDCl₃) δ 170.10, 153.92, 153.28, 148.90, 132.69,

130.45, 128.51, 128.14, 123.81, 122.90, 108.24, 104.80, 97.80, 65.87, 62.68, 62.66, 44.65, 44.37, 12.61.



The compound was grey white solid, yield 86%. ¹H NMR (600 MHz, CDCl₃) δ 7.94 – 7.90 (m, 1H), 7.46 – 7.43 (m, 2H), 7.12 – 7.07 (m, 1H), 6.46 (d, *J* = 8.9 Hz, 2H), 6.40 (d, *J* = 2.6 Hz, 2H), 6.29 (dd, *J* = 8.9, 2.6 Hz, 2H), 3.91 (d, *J* = 2.4 Hz, 2H), 3.37 (dq, *J* = 14.2, 7.1 Hz, 10H), 3.24 (t, *J* = 7.0 Hz, 2H), 2.30 (t, *J* = 2.4 Hz, 1H), 1.18 (t, *J* = 7.1 Hz, 12H).



The compound was light red solid, yield 88%. ¹H NMR (600 MHz, CDCl₃) δ 7.90 – 7.86 (m, 1H), 7.77 (s, 1H), 7.42 – 7.38 (m, 2H), 7.07 – 7.03 (m, 1H), 6.42 (dd, *J* = 8.9, 4.9 Hz, 2H), 6.36 (d, *J* = 2.4 Hz, 2H), 6.29 – 6.22 (m, 2H), 5.78 (d, *J* = 9.3 Hz, 1H), 5.56 – 5.50 (m, 2H), 5.21 (dd, *J* = 10.3, 3.3 Hz, 1H), 4.40 – 4.35 (m, 2H), 4.20 – 4.17 (m, 1H), 4.12 – 4.07 (m, 2H), 3.38 (t, *J* = 7.2 Hz, 2H), 3.31 (q, *J* = 7.0 Hz, 8H), 3.18 (dt, *J* = 9.4, 7.1 Hz, 2H), 2.21 (s, 3H), 2.01 (s, 3H), 1.98 (s, 3H), 1.83 (s, 3H), 1.14 (t, *J* = 7.0 Hz, 12H).



The compound was light red solid, yield 84%. ¹H NMR (600 MHz, CDCl₃) δ 7.93 – 7.90 (m, 1H), 7.71 (s, 1H), 7.46 – 7.42 (m, 2H), 7.11 – 7.07 (m, 1H), 6.45 (dd, J = 8.8, 7.0 Hz, 2H), 6.39 (d, J = 1.7 Hz, 2H), 6.27 (dd, J = 8.9, 2.4 Hz, 2H), 5.78 (d, J = 8.8 Hz, 1H), 5.41 (ddd, J = 10.6, 9.4, 6.3 Hz, 3H), 5.15 (dd, J = 10.4, 7.9 Hz, 1H), 4.99 (dd, J = 10.4, 3.5 Hz, 1H), 4.54 (d, J = 7.9 Hz, 1H), 4.47 (dd, J = 12.1, 1.7 Hz, 1H), 4.38 (s, 2H), 4.14 (qd, J = 11.2, 6.6 Hz, 3H), 3.99 – 3.95 (m, 1H), 3.94 – 3.88 (m, 2H), 3.41 (t, J = 7.2 Hz, 2H), 3.35 (q, J = 7.0 Hz, 8H), 3.22 – 3.18 (m, 2H), 2.18 (s, 3H), 2.11 (s, 3H), 2.10 (s, 3H), 2.07 (s, 6H), 1.99 (s, 3H), 1.84 (s, 3H), 1.18 (td, J = 6.9, 1.0 Hz, 12H).



Rho-Gal, light red solid, yield 92%. ¹H NMR (600 MHz, CDCl₃) δ 8.00 (s, 1H), 7.81 (d, J = 6.7 Hz, 1H), 7.38 – 7.32 (m, 2H), 7.00 (d, J = 6.6 Hz, 1H), 6.39 – 6.34 (m, 4H), 6.26 – 6.21 (m, 2H), 5.57 (d, J = 9.0 Hz, 1H), 4.33 (t, J = 9.1 Hz, 1H), 4.26 (s, 2H), 4.18 (s, 1H), 3.87 – 3.77 (m, 4H), 3.28 (dd, J = 13.7, 6.6 Hz, 10H), 3.07 (t, J = 6.2Hz, 2H), 1.11 (t, J = 7.0 Hz, 12H). ¹³C NMR (151 MHz, CD₃OD) δ 168.85, 153.74, 153.35, 149.01, 144.74, 132.72, 130.54, 128.30, 128.09, 123.66, 122.26, 122.18, 108.21, 104.76, 97.66, 88.86, 78.54, 73.91, 70.05, 68.96, 66.90, 65.52, 63.26, 61.01, 43.99, 39.00, 11.51. HRMS: [M+H]⁺ calculated for C₃₉H₄₉N₆O₈, m/z 729.36064, measured m/z 729.36157. mp 165.3-167.9.



Rho-Lac, light red solid, yield 91%. ¹H NMR (600 MHz, CD₃OD) δ 8.05 (s, 1H), 7.88 – 7.84 (m, 1H), 7.52 – 7.48 (m, 2H), 7.03 – 7.00 (m, 1H), 6.43 (s, 2H), 6.34 (d, *J* = 0.9 Hz, 4H), 5.61 (d, *J* = 9.2 Hz, 1H), 4.43 (d, *J* = 7.7 Hz, 1H), 4.29 (s, 2H), 3.95 (t, *J* = 9.1 Hz, 1H), 3.89 (d, *J* = 2.9 Hz, 2H), 3.84 (d, *J* = 3.2 Hz, 1H), 3.80 (td, *J* = 11.1, 8.6 Hz, 2H), 3.73 (ddd, *J* = 12.8, 9.1, 3.1 Hz, 3H), 3.63 (dd, *J* = 7.5, 4.7 Hz, 1H), 3.59 (dd, *J* = 9.6, 7.8 Hz, 1H), 3.52 (dd, *J* = 9.7, 3.3 Hz, 1H), 3.36 (q, *J* = 7.0 Hz, 8H), 3.12 (t, *J* = 6.6 Hz, 2H), 1.14 (t, *J* = 7.0 Hz, 12H). ¹³C NMR (151 MHz, CD₃OD) δ 168.86, 153.74, 153.35, 149.02, 144.69, 132.73, 130.53, 128.29, 128.11, 123.66, 122.63, 122.18, 108.21, 104.74, 103.71, 97.65, 87.94, 78.26, 78.15, 75.75, 75.41, 73.43, 72.28, 71.16, 68.93, 66.93, 65.53, 63.25, 61.15, 60.14, 43.99, 38.99, 11.51. HRMS: [M+H]⁺ calculated for C₄₅H₅₉N₆O₁₃, m/z 891.41346, measured m/z 891.41516. mp 176.5-179.6.



Rho-Glu, light red solid, yield 83%. ¹H NMR (600 MHz, CD₃OD) δ 8.06 (s, 1H), 7.87 (dt, J = 5.9, 2.5 Hz, 1H), 7.54 – 7.49 (m, 2H), 7.04 – 7.02 (m, 1H), 6.45 (s, 2H), 6.39 – 6.34 (m, 4H), 5.59 (d, J = 9.2 Hz, 1H), 4.30 (s, 2H), 3.92 – 3.87 (m, 2H), 3.72 (dd, J = 12.2, 5.3 Hz, 1H), 3.61 – 3.55 (m, 2H), 3.55 – 3.50 (m, 1H), 3.38 (q, J = 7.0 Hz, 8H), 3.34 – 3.32 (m, 2H), 3.13 (t, J = 6.6 Hz, 2H), 1.16 (t, J = 7.0 Hz, 12H). ¹³C

NMR (151 MHz, CD₃OD) δ 168.85, 153.74, 153.35, 149.02, 144.65, 132.73, 130.54, 128.29 128.10, 123.66, 122.64, 122.17 108.21, 104.75, 97.65, 88.21, 79.73, 77.06, 72.65, 69.49, 66.92, 65.52, 63.25, 61.01, 43.99, 38.98, 11.51. HRMS: [M+H]⁺ calculated for C₃₉H₄₉N₆O₈, m/z 729.36064, measured m/z 729.36194. mp 164.6-167.5.







Fig. S11 Mass spectrum of Rho-Gal-Hg²⁺ complex



Fig. S13 Mass spectrum of Rho-Lac-Hg²⁺ complex



Fig. S14 Mass spectrum of Rho-Glu



Fig. S15 Mass spectrum of Rho-Glu-Hg²⁺ complex



Fig. S17 ¹H NMR of Rho-Lac



Fig. S19 ¹³C NMR of Rho-Gal



Fig. S21 ¹³C NMR of Rho-Glu