Circularly polarized luminescence from Tb(III) interacting with chiral polyether macrocycles

Supporting Information

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Table of Contents

1	Ge	neral Information and Materials	S4
	1.1	CSP-HPLC	S4
	1.2	Optical properties	S4
	1.3	¹ H-NMR	S5
2	Syr	nthesis and characterization of organic compounds	S6
	2.1	Synthesis of unsaturated ester macrocycle 2	S6
	2.2	Synthesis of ligands	S6
	2.3	Resolution of ligand 1a ⁶	S7
	2.4	Key chiroptical properties of ligand 1a and complexes	S8
3	Qu	alitative test of potential ligands	S9
4	Absorbance and fluorescence spectra and titrations		
	4.1	Procedure	S10
	4.2	Ligand 1a and Tb(III)	S10
	4.3	Titration of ligand 1a with Tb(III)	S11
	4.4	Titration of ligand 1a with Ba(II)	S12
	4.5	Titration of ligand 1b with Tb(III)	S12
	4.6	Titration of ligand 1c with Tb(III)	S13
	4.7	Titration of ligand 1d with Tb(III)	S13
5	ECD and CPL spectra		S14
	5.1	Procedure	S14
	5.2	Ligand 1a and Tb(III) - ECD	S15
	5.3	Ligand 1a and Ba(II) - ECD	S16
	5.4	Ligand 1a , Tb(III) and Ba(II) - g _{abs}	S17
6	¹ H-	NMR titrations	S18
	6.1	Procedure	S18
	6.2	¹ H-NMR spectra	S18
7	Sol	id state structure and crystallographic data	S19
	7.1	Procedure	S19
	7.2	Data complex $[1e \cdot La \cdot (H_2O)_2](ClO_4)_3$	S19
8	Lur	ninescence lifetime measurement	S21
	8.1	Procedure	S21

8	.2	Time trace	S21
9	Refe	erences	S22

1 General Information and Materials

1.1 CSP-HPLC

Enantiomers of ligand **1a** were resolved by chiral stationary phase HPLC on an Agilent 1260 Infinity II apparatus (quaternary pump, auto sampler, column thermostat and diode array detector) using a semi-preparative CHIRALPAK[®] IG column (250 x 10 mm, 5 mic). Mixtures of HPLC grade CH₂Cl₂ and MeOH (99:1, with 0.1% diethanolamine as additive) were used as mobile phase.

1.2 Optical properties

Optical properties were recorded in analytical grade solvent (acetonitrile). UV-Vis absorption spectra were recorded on a JASCO V-650 spectrophotometer at 20 °C. Electronic circular dichroism (ECD) spectra were recorded on a Jasco J-815 spectropolarimeter at 20 °C in a 1 cm cuvette.

Fluorescence spectra were measured using a Varian Cary 50 Eclipse spectrophotometer. All fluorescence spectra were corrected for the wavelength-dependent sensitivity of the detection. Fluorescence quantum yields ϕ were measured in diluted solutions (at least 5 different concentrations for each sample) with an optical density lower than 0.1 using the following equation:

$$\frac{\Phi_{\chi}}{\Phi_{r}} = \left(\frac{A_{R}(\lambda)}{A_{\chi}(\lambda)}\right) \left(\frac{n_{\chi}^{2}}{n_{r}^{2}}\right) \left(\frac{D_{\chi}}{D_{r}}\right)$$

where A is the absorbance at the excitation wavelength (λ), n the refractive index and D the integrated intensity. "r" and "x" stand for reference and sample respectively. The fluorescence quantum yields were measured in acetonitrile relative to 9,10-diphenylanthracene (ϕ = 93% in cyclohexane). Excitation of reference and sample was performed at the same wavelength.

Circularly polarized luminescence (CPL) spectra were recorded with the home-made spectrofluoropolarimeter previously described.¹ The samples were excited with a 254 nm fluorescent mercury lamp, using a 90° geometry between excitation and detection.

 $Ba(ClO_4)_2$ and $Tb(OTf)_3$ salts used for titration experiments were purchased from commercial sources and used without purification.

Lifetimes were determined using the phosphorescence mode of a Fluorolog 3 spectrophotometer (Horiba Jobin Yvon) in which the lamp of the instrument is flashed. Excitation was performed at 305 nm (1 nm slit) and detection with a visible photomultiplier

tube (220-850 nm, R928P, Hamamatsu) at 545 nm (3 nm slit) at 545 nm, with an initial time gate of 50 μ s.

1.3 ¹H-NMR

The ¹H NMR spectra were recorded in deuterated CDCl₃ using an Agilent Inova 600 (¹H: 600 MHz). ¹H NMR chemical shifts are given in ppm relative to Me₄Si using solvent resonances as internal standards (CD₃CN δ = 1.94 ppm). Data were reported as follows: chemical shift (δ) in ppm, multiplicity (s = singulet, d = doublet, t = triplet, dd = doublet of doublet, q = quartet and m = multiplet), coupling constant (Hz) and integration.

2 Synthesis and characterization of organic compounds

2.1 Synthesis of unsaturated ester macrocycle 2

Unsaturated ester macrocycle **2** was synthetized according to previously reported procedure from the literature²:



2.2 Synthesis of ligands

Ligand **1a**,³ **1b**,³ **1c**,⁴ **1d**,³ **S1**,³ **S2**⁴ and **S3**⁵ were synthesized according to the previously reported procedure. See Figure S1:



Figure S1. Synthesis of ligands.

2.3 Resolution of ligand **1a**⁶

Compound **1a** were resolved by chiral stationary phase HPLC using a semi-preparative CHIRALPAK[®] IG column using a mixture of CH₂Cl₂-MeOH (99:1, with 0.1% diethanolamine as additive) as mobile phase at 20 °C. It is worth mentioning that it is necessary to remove traces of diethanolamine present in the separated compounds. The residue was thus dissolved in CH₂Cl₂, the organic phase was washed three times with H₂O, dried over anhydrous Na₂SO₄, filtered and concentrated under vacuum to afford the pure products.

In the Figure **S2** is shown the HPLC traces of ligand **1a** on analytical CHIRALPAK[®] IG column (left, test run) and on semi preparative CHIRALPAK[®] IG column (right, run for resolution) with CH_2Cl_2 -MeOH (99:1, 0.1% diethanolamine) as mobile phase.



2 Vial with 10 mg DCM : MeOH : DEA = 99 : 1 : 0.1 Inj 250 μL Flow 5 mL/min

Figure S2. HPLC traces of the racemic mixture on analytical CHIRALPAK[®] IG column (left) and semi preparative (right).

In the Figure **S3** is shown the HPLC traces of ligand **1a** on analytical CHIRALPAK[®] IG column of the separated enantiomers: 1^{st} eluted enantiomer on the left and 2^{nd} eluted enantiomer on the right with CH₂Cl₂-MeOH (99:1, 0.1% diethanolamine) as mobile phase.

Analytical IG column

Analytical IG column

1st eluted enantiomer (99% ee)

2nd eluted enantiomer (94% *ee*)



Figure S3. HPLC traces of the separated enantiomers. Left: 1st eluted enantiomer. Right: 2nd eluted enantiomer.

2.4 Key chiroptical properties of ligand **1a** and complexes

ECD description for 1st eluted enantiomer of **1a** in acetonitrile, λ/nm ($\Delta\epsilon/M^{-1}\text{cm}^{-1}$): 279(-2.4), 240 (+1.0).

ECD description for 1st eluted enantiomer of **1a** complexed to Tb(III) in acetonitrile, λ /nm ($\Delta \epsilon$ /M⁻¹cm⁻¹): 274 (+7.7), 226 (–20). *See section 5 for further precision.*

ECD description for 1st eluted enantiomer of **1a** complexed to Ba(II) in acetonitrile, λ /nm ($\Delta \epsilon$ /M⁻¹cm⁻¹): 274 (+4.4), 236 (-28.1). See section 5 for further precision.

3 Qualitative test of potential ligands

For the qualitative test, three solutions in three different vials (1 mL) were prepared and their emission was compared under UV irradiation (366 nm excitation wavelength). In the first one (reference 1), only the macrocycle of interest (<1 mg) is dissolved in acetonitrile. In the second one (reference 2), only terbium triflate (tip of a spatula) was dissolved in acetonitrile. In the third one, a mixture of the macrocycle of interest (<1 mg) and terbium triflate (tip of a spatula, excess) were dissolved in acetonitrile. In the reference 1 (1st vial), only the fluorescence of the macrocycle can be observed when visible. In the reference 2, no emission of the terbium salt was observed at this wavelength, but for the third vial (macrocycle/Tb mixture) resulted in the characteristic green terbium emission (see below). Combination of ligand **1a** and terbium presents the most efficient luminescence and were selected for this study.



Figure S4. Ligand tested with terbium(III) and qualitative results of the mixture (picture under 366 nm irradiation).

4 Absorbance and fluorescence spectra and titrations

4.1 Procedure

In a typical experiment, UV-Vis absorbance and fluorescence spectra of a solution of interest compound (ca. $0.5 \cdot 10^{-6}$ M) in acetonitrile were recorded in a 1 cm cell. For the complexation experiments, an excess of Tb(OTf)₃ (or Ba(ClO₄)₂) or an aliquot of a Tb(OTf)₃ (or Ba(ClO₄)₂) solution in acetonitrile (ca. $2 \cdot 10^{-3}$ M) was added to the solution and the UV-Vis absorbance and fluorescence spectra were recorded again.



4.2 Ligand **1a** and Tb(III)

Figure S5. Absorbance (red and blue lines) and fluorescence (pink and green lines) spectra of ligand 1a without (red and pink lines) or with 3.0 equivalents of Tb(III) (blue and green lines).

4.3 Titration of ligand **1a** with Tb(III)

In a typical experiment, a known aliquot of a Tb(OTf)₃ solution in acetonitrile (ca. $2 \cdot 10^{-3}$ M) was added to a solution of the ligand (ca. $0.5 \cdot 10^{-6}$ M) in acetonitrile. Spectra were recorded from 0 equivalent of Tb(III) to 4.0 equivalents.



Figure S6. Titrations spectra in absorbance of ligand **1a** with Tb(III): 0 to 4.0 equivalents. Left: absorbance spectra. Right: evolution of absorbance as function of the equivalents added at different relevant wavelength.



Figure S7. Titrations spectra in emission of ligand **1a** with Tb(III): 0 to 4.0 equivalents. Left: normalized fluorescence spectra. Right: evolution of fluorescence as function of the equivalents added at different relevant wavelength.

4.4 Titration of ligand **1a** with Ba(II)

In a typical experiment, a known aliquot of a $Ba(CIO_4)_2$ solution in acetonitrile (ca. $4 \cdot 10^{-3}$ M) was added to a solution of the ligand (ca. $0.5 \cdot 10^{-6}$ M) in acetonitrile. Spectra were recorded from 0 equivalent of Ba(II) to 4.0 equivalents.



Figure S8. Titrations spectra in absorbance of ligand **1a** with Ba(II): 0 to 4.0 equivalents. Left: absorbance spectra. Right: evolution of absorbance as function of the equivalents added at 274 nm.



4.5 Titration of ligand **1b** with Tb(III)

Figure S9. Titrations spectra of ligand **1b** with Tb(III): 0 to 4.0 equivalents. Left: absorbance spectra. Right: normalized fluorescence spectra.

4.6 Titration of ligand **1c** with Tb(III)



Figure S10. Titrations spectra of ligand 1c with Tb(III): 0 to 5.0 equivalents. Left: absorbance spectra. Right: normalized fluorescence spectra.



4.7 Titration of ligand **1d** with Tb(III)

Figure S11. Titrations spectra of ligand **1d** with Tb(III): 0 to 4.0 equivalents. Left: absorbance spectra. Right: normalized fluorescence spectra.

5 ECD and CPL spectra

5.1 Procedure

For ECD:

In a typical experiment, the ECD spectrum of a solution of enantiopure ligand (ca. $0.5 \cdot 10^{-5}$ M) in acetonitrile was recorded in a 1 cm cell at 20 °C. For the complexation experiments, 3.0 equivalents of Tb(OTf)₃ or Ba(ClO₄)₂ (ca. $2 \cdot 10^{-3}$ M stock solutions in acetonitrile) were added to the ligand solution and the ECD spectrum was recorded again.

The change in intensity in ECD is quantified using $\delta\Delta\epsilon$, which is the difference in normalized ECD intensity in presence and absence of tested metal ions:

$$\delta \Delta \varepsilon = |\Delta \varepsilon (\text{cation}) - \Delta \varepsilon (\text{without})|$$
 (S1)

For CPL:

The CPL spectrum of a solution containing the enantiopure ligand (ca. $2 \cdot 10^{-5}$ M) in acetonitrile and 3.0 equivalents of Tb(OTf)₃ (from a ca. 10^{-3} M stock solution in acetonitrile) was recorded in a 1 cm cell.

The circular polarization degree of the emission is quantifying using the luminescence dissymmetry factor g_{lum} defined by equation S2 where I_L and I_R correspond to left and right circularly polarized component of the emission respectively:

$$g_{lum} = 2 \; rac{\mathrm{I_L} - \mathrm{I_R}}{\mathrm{I_L} + \mathrm{I_R}}$$
 (S2)

5.2 Ligand 1a and Tb(III) - ECD



Figure S12. ECD (top) and absorbance (bottom) spectra ligand 1a (red) and [1a·Tb]³⁺ complex (green). En1 and En2 corresponds to the 1st and 2nd eluted enantiomers on CHIRALPAK[®] IG column and a mixture of CH₂Cl₂-MeOH (99:1, 0.1% diethanolamine) as mobile phase.

5.3 Ligand 1a and Ba(II) - ECD



Figure S13. ECD (top) and absorbance (bottom) spectra ligand 1a (red) and [1a·Ba]²⁺ complex (blue). En1 and En2 corresponds to the 1st and 2nd eluted enantiomers on CHIRALPAK[®] IG column and a mixture of CH₂Cl₂-MeOH (99:1, 0.1% diethanolamine) as mobile phase.

5.4 Ligand 1a, Tb(III) and Ba(II) - gabs



Figure S14. g_{abs} spectra of ligand **1a** (red), $[\mathbf{1a} \cdot \text{Tb}]^{3+}$ (green) and $[\mathbf{1a} \cdot \text{Ba}]^{2+}$ (blue) complexes. En1 and En2 corresponds to the 1st and 2nd eluted enantiomers on CHIRALPAK[®] IG column and a mixture of CH₂Cl₂-MeOH (99:1, 0.1% diethanolamine) as mobile phase.

6 ¹H-NMR titrations

6.1 Procedure

To an NMR tube, $LuCl_3$ was added (0, 0.5, 1 and 2 equivalents) as a solid. Just before the measurement 0.5 mL of a 15 mM solution of **1a** was added and the tube was shaken, and the ¹H spectrum was recorded.



Figure S15, ¹H NMR titration of 1a with LuCl₃ (0, 0.5, 1, 2 equivalents) c = 15 mM, 600 MHz

7 Solid state structure and crystallographic data

7.1 Procedure

About 5 mg of **1a** were dissolved in 2 mL of MeCN and a large excess of $La(ClO_4)_3$ was added. The solution was filtered and the solvent allowed to slowly evaporate at room temperature over the course of one week.

7.2 Data complex [1e·La·(H₂O)₂](ClO₄)₃



Figure S16. Crystal data and structure refinement for [1e·La·(H₂O)₂](ClO₄)₃

CCDC number		2189306		
Empirical formula		C56 H58 Cl3 La N6 O22		
Formula weight		1412.34		
Temperature		120.00(11) K		
Crystal system		Monoclinic		
Space group		P21/c		
Unit cell dimensions		a = 23.5523(2) Å	α = 90°	
		b = 11.12589(12) Å	$\beta = 99.5672(10)$ °	
		c = 23.3538(2) Å	γ = 90°	
Volume		6034.52(11) ų		
Z		4		
Density (calculated)		1.555 g/cm ³		
F(000)		2880.0		
Crystal size		$0.591 \times 0.136 \times 0.036 \text{ mm}^3$		
20 range for data collection		7.614 to 149.28°		
Index ranges		-29 ≤ h ≤ 29, -13 ≤ k ≤ 13, -29 ≤ l ≤ 29		
Reflections collected		19051		
Independent reflections		19051 [Rint = ?, Rsigma = 0.0100]		
Data/restraints/parameters		19051/3/809		
Goodness-of-fit on F ²		1.034		
Final R indexes [I>=2 σ (I)]		R1 = 0.0439, wR2 = 0	.1169	
Final R indexes [all data]	R	1 = 0.0465, wR2 = 0.1192		
Largest diff. peak/hole		1.19/-1.26 e Å ⁻³		

8 Luminescence lifetime measurement

8.1 Procedure

A 10^{-5} M solution of **1a** (S1) and a 10^{-3} M solution of Tb(OTf)₃ (S2) both in MeCN or MeCN-d3 were prepared. The samples were prepared by mixing the 2 solutions directly in a cuvette.

The lifetime of complex **1a** in acetonitrile was determined using the phosphorescence mode of a Fluorolog 3 spectrophotometer (Horiba Jobin Yvon) in which the lamp of the instrument is flashed. Excitation was performed at 305 nm (1 nm slit) and detection with a visible photomultiplier tube (220-850 nm, R928P, Hamamatsu) at 545 nm (3 nm slit) at 545 nm, with an initial time gate of 50 μ s.



8.2 Time trace

Figure S17. Luminescence decay of [**1a**·Tb][ClO₄]₃ at 545 nm upon 305 nm excitation in MeCN and MeCN-d3 solutions. Solid lines are exponential fits to the data points (open circles).

9 References

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