Electronic Supplementary Information (ESI)

AIE-active Ir(III) complexes as Type-I dominant photosensitizers for

efficient photodynamic therapy

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1. Synthesis and characterization



Scheme S1. Synthetic routes of the complexes. (i) $IrCl_3 \cdot 3H_2O$, ethoxyethanol, water (3:1, v/v), reflux, 24 h; (ii) μ -chloro-bridged dimer, CH_2Cl_2 , ethanol, reflux, 12 h.

Synthesis of SFIrqa

IrCl₃·3H₂O (0.35 g, 1.00 mmol) and ligand SFPBI¹ (1.12 g, 2.20 mmol) in a solution of 2-ethoxyethanol and water were refluxed for about 24 h under Ar₂ atmosphere. The mixture was filtered after cooling to room temperature, followed by washing with water and ethanol, obtaining about 0.68 g µ-chloro-bridged dimer. Then, the ancillary ligand (0.05 g, 0.28 mmol) and the corresponding dimer (0.30 g, 0.11 mmol) in a mixed solvent of CH₂Cl₂ and ethanol were allowed to reflux overnight. The residue was purified by silica gel column chromatography with ethyl acetate: CH₂Cl₂ in 1:10 (v:v) ratio as the eluent, giving a red powder. Yield 70%. ¹H NMR (500 MHz, d_{δ} -DMSO, δ [ppm]): 8.67 (d, *J* = 9.0 Hz, 1H), 8.21 (d, *J* = 8.4 Hz, 1H), 8.09 (d, *J* = 7.8 Hz, 1H), 7.52 (t, *J* = 7.8 Hz, 1H), 7.42-7.46 (m, 4H), 7.35-7.39 (m, 4H), 7.29-7.34 (m, 3H), 7.21-7.27 (m, 6H), 7.14-7.19 (m, 4H), 6.99-7.12 (m, 8H), 6.74-6.77 (m, 3H), 6.69 (d, *J* = 7.8 Hz, 1H), 5.65 (s, 1H), 5.57 (s, 1H). ¹³C NMR (125 MHz, 1H), 5.69 (d, *J* = 8.4 Hz, 1H), 5.65 (s, 1H), 5.57 (s, 1H). ¹³C NMR (125 MHz, 1H), 5.69 (d, *J* = 8.4 Hz, 1H), 5.65 (s, 1H), 5.57 (s, 1H).

CDCl₃, δ [ppm]): 175.27, 163.83, 163.75, 155.13, 150.55, 149.89, 149.38, 149.21, 149.02, 148.85, 148.42, 148.03, 146.25, 142,74, 142.26, 142.17, 142.04, 141.78, 141.71, 141.59, 141.36, 141.07, 141.04, 139.92, 139.75, 138.54, 136.31, 135.88, 134.87, 134.84, 133.60, 133.51, 130.33, 130.02, 129.92, 129.68, 129.61, 129.56, 129.44, 129.43, 128.19, 127.94, 127.79, 127.70, 127.53, 127.34, 127.27, 127.24, 127.15, 127.07, 125.49, 124.70, 124.47, 124.32, 124.25, 124.06, 124.02, 123.74, 123.26, 123.16, 122.89, 121.33, 120.67, 120.47, 119.96, 119.76, 119.43, 119.30, 119.15, 116.99, 114.43, 110.64, 110.08. MS [m/z]: Calcd for C₈₆H₅₂IrN₅O₂: 1379.4, Found 1379.4 [M]⁺. Anal. calcd. For C₈₆H₅₂IrN₅O₂: C 74.87, H 3.80, N 5.08; Found: C 74.65, H 3.78, N 5.09.

Synthesis of SFIriqa

The synthesis method is the same as SFIrqa. Yield 73%. ¹H NMR (500 MHz, d_6 -DMSO, δ [ppm]): 9.95 (d, J = 8.4 Hz, 1H), 8.15 (d, J = 6.0 Hz, 1H), 8.06 (d, J = 8.4Hz, 1H), 7.87-7.92 (m, 7H), 7.81 (t, J = 7.8 Hz, 1H), 7.49-7.53 (m, 2H), 7.45 (d, J = 7.2 Hz, 1H), 7.29-7.39 (m, 10H), 7.22-7.26 (m, 3H), 7.11-7.19 (m, 9H), 6.98-7.09 (m, 5H), 6.79-6.84 (m, 2H), 6.68 (d, J = 7.8 Hz, 1H), 6.54-6.59 (m, 3H), 6.44-6.48 (m, 2H), 5.69 (s, 2H), 5.56 (s, 1H). ¹³C NMR (125 MHz, CDCl₃, δ [ppm]): 174.41, 164.17, 163.33, 151.31, 150.48, 149.99, 149.45, 149.29, 149.19, 149.15, 148.96, 148.87, 142.79, 142.37, 142.15, 142.12, 141.75, 141.56, 141.39, 141.13, 141.05, 139.81, 139.77, 136.58, 136.45, 135.98, 134.91, 134.03, 133.47, 131.32, 130.01, 129.93, 129.72, 129.62, 129.54, 129.46, 129.17, 128.79, 128.23, 127.96, 127.92, 127.81, 127.67, 127.66, 127.45, 127.41, 127.35, 127.25, 127.20, 127.10, 127.05, 126.46, 126.12, 125.69, 124.78, 124.74, 124.50, 124.19, 124.07, 124.01, 123.76, 123.64, 123.41, 122.71, 121.02, 120.74, 120.34, 119.82, 119.61, 119.41, 119.29, 119.15, 117.44, 114.29, 110.63, 110.03. MS [m/z]: Calcd for C₈₆H₅₂IrN₅O₂: 1379.4, Found 1379.4 [M]⁺. Anal. calcd. For C₈₆H₅₂IrN₅O₂: C 74.87, H 3.80, N 5.08; Found: C 74.72, H 3.92, N 4.90.

Synthesis of MFIrqa

The synthesis method is the same as **MFIriqa**. Yield 73%. ¹H NMR (500 MHz, d_{6} -DMSO, δ [ppm]): 8.73 (d, J = 8.4 Hz, 1H), 8.29 (d, J = 8.4 Hz, 1H), 8.11 (d, J = 7.8

Hz, 1H), 7.93-7.98 (m, 3H), 7.88-7.92 (m, 1H), 7.83-7.87 (m, 4H), 7.80 (d, J = 7.8 Hz, 1H), 7.72 (d, J = 7.8 Hz, 1H), 7.67 (d, J = 6.0 Hz, 1H), 7.59 (t, J = 7.8 Hz, 1H), 7.48 (d, J = 6.6 Hz, 1H), 7.42 (d, J = 7.2 Hz, 1H), 7.38 (d, J = 7.8 Hz, 1H), 7.28-7.32 (m, 2H), 7.24-7.28 (m, 2H), 7.14-7.23 (m, 6H), 7.11 (t, J = 7.8 Hz, 1H), 7.07 (d, J = 7.2 Hz, 1H), 6.93 (s, 1H), 6.80 (t, J = 7.8 Hz, 1H), 6.57 (d, J = 3.0 Hz, 2H), 6.52 (s, 1H), 5.68 (d, J = 8.4 Hz, 1H), 1.22 (s, 3H), 1.11 (s, 3H), 1.06 (s, 3H), 1.03 (s, 3H). ¹³C NMR (125 MHz, CDCl₃, δ [ppm]): 175.29, 164.23, 164.14, 155.22, 154.70, 154.59, 149.95, 148.12, 146.47, 145.96, 145.63, 140.79, 140.40, 140.04, 139.94, 139.10, 138.86, 138.47, 136.72, 136.29, 135.83, 135.80, 132.87, 132.83, 130.45, 130.40, 130.32, 130.18, 130.15, 130.11, 130.06, 129.72, 128.87, 128.65, 128.55, 128.14, 123.78, 123.20, 122.86, 122.41, 122.27, 120.27, 120.04, 119.87, 119.52, 116.98, 114.49, 110.65, 110.08. MS [m/z]: Calcd for C₆₆H₄₈IrN₅O₂: 1135.3, Found 1135.3 [M]⁺. Anal. calcd. For C₆₆H₄₈IrN₅O₂: C 69.82, H 4.26, N 6.17; Found: C 69.70, H 4.18, N 6.19.

Synthesis of MFIriqa

The synthesis method is the same as **MFIriqa**. Yield 68%. ¹H NMR (500 MHz, d_6 -DMSO, δ [ppm]): 10.02 (d, J = 9.0 Hz, 1H), 8.15 (d, J = 6.0 Hz, 1H), 8.04-8.08 (m, 2H), 7.95-7.97 (m, 2H), 7.84-7.93 (m, 9H), 7.73 (d, J = 7.8 Hz, 1H), 7.68 (d, J = 7.2 Hz, 1H), 7.40 (d, J = 7.2 Hz, 1H), 7.36-7.38 (m, 1H), 7.32-7.34 (m, 2H), 7.23-7.26 (m, 3H), 7.14-7.20 (m, 4H), 7.11 (d, J = 7.8 Hz, 1H), 7.02 (d, J = 7.2 Hz, 1H), 6.92 (s, 1H), 6.84-6.88 (m, 1H), 6.63 (s, 1H), 6.57 (s, 1H), 6.52 (s, 1H), 5.71 (d, J = 7.8 Hz, 1H), 1.19 (s, 3H), 1.10 (s, 3H), 1.03 (d, J = 11.4 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃, δ [ppm]): 173.44, 163.51, 162.73, 153.63, 153.58, 150.39, 149.10, 148.29, 145.21, 144.61, 141.38, 139.90, 139.82, 139.26, 138.93, 138.87, 138.17, 137.71, 135.82, 135.58, 135.38, 134.82, 132.35, 131.79, 130.22, 129.43, 129.34, 129.19, 129.11, 129.08, 129.05, 128.75, 128.10, 127.85, 127.76, 127.72, 127.55, 127.07, 126.38, 125.90, 125.54, 125.48, 125.38, 125.01, 124.58, 123.82, 133.51, 109.63, 109.01, 106.48. MS [m/z]: Calcd for C₆₆H₄₈IrN₅O₂: 1135.3, Found 1135.3 [M]⁺. Anal. calcd. For C₆₆H₄₈IrN₅O₂: C 69.82, H 4.26, N 6.17; Found: C 69.75, H 4.18, N 6.19.



Fig. S1 ¹H NMR spectrum of MFIriqa in d_6 -DMSO at room temperature.



Fig. S2 ¹³C NMR spectrum of MFIriqa in CDCl₃ at room temperature.



Fig. S3 High-resolution MALDI-TOF mass spectrum of MFIriqa.



Fig. S4 ¹H NMR spectrum of MFIrqa in d_6 -DMSO at room temperature.



Fig. S5 ¹³C NMR spectrum of MFIrqa in CDCl₃ at room temperature.



Fig. S6 MALDI-TOF mass spectrum of MFIrqa.



Fig. S7 ¹H NMR spectrum of SFIriqa in d_6 -DMSO at room temperature.



Fig. S8 ¹³C NMR spectrum of SFIriqa in CDCl₃ at room temperature.



Fig. S9 MALDI-TOF mass spectrum of SFIriqa.



Fig. S10 ¹H NMR spectrum of SFIrqa in d_6 -DMSO at room temperature.



Fig. S11 ¹³C NMR spectrum of SFIrqa in CDCl₃ at room temperature.



Fig. S12 MALDI-TOF mass spectrum of SFIrqa.

Name	MFIriqa	MFIrqa	SFIriqa
Identification code	CCDC 2212243	CCDC 2212244	CCDC 2212245
Formula	$\mathrm{C}_{66}\mathrm{H}_{48}\mathrm{IrN}_{5}\mathrm{O}_{2}$	$\mathrm{C_{66}H_{48}IrN_5O_2}$	$\mathrm{C}_{86}\mathrm{H}_{52}\mathrm{IrN}_{5}\mathrm{O}_{2}$
Formula weight	1135.29	1135.29	1379.52
Crystal system	monoclinic	monoclinic	orthorhombic
Space group	$P2_1/n$	$P2_1/n$	$P2_{1}2_{1}2_{1}$
	a 17.0548(8)	a 19.1462(11)	a 16.8482(8)
Cell Lengths (Å)	b 18.1638(8)	b 14.0770(8)	b 18.3470(9)
	c 19.8406(9)	c 22.2847(13)	c 22.5335(10)
	α 90	α 90	α 90
Cell Angles (°)	β 97.298(3)	β 104.354(3)	β 90
	γ 90	γ 90	γ 90
Cell Volume (Å3)	6096.4(5)	5818.7(6)	6965.4(6)
Ζ	4	4	4
Dcalcd.(g m-3)	1.237	5.021	1.316
F(000)	2288.0	7540.0	2784.0
Rint	0.0685	0.0427	0.0534
Goodness-of-fit on F2	1.034	0.824	1.020
R1a, wR2b [I>=2σ (I)]	0.0377, 0.0780	0.0285, 0.0961	0.0378, 0.0905
R1, wR2 [all data]	0.0533, 0.0834	0.0356, 0.1022	0.0446, 0.0949

Table S1 Crystal data and structure refinement of MFIriqa, MFIrqa and SFIriqa.

^a $R_1 = \Sigma ||Fo| - |Fc|| / \Sigma |Fo|$. ^b $wR_2 = |\Sigma w(|Fo|^2 - |Fc|^2)| / \Sigma |w(Fo^2)^2|^{1/2}$.

Name	SFIriqa	MFIrqa	MFIriqa
bond lengths (Å)	Ir1-O1 [2.152(5)]	Ir1-O1 [2.138(3)]	Ir1-O2 [2.145(3)]
	Ir1-N1 [2.043(5)]	Ir1-N1 [2.040(3)]	Ir1-N1 [2.129(3)]
	Ir1-N4 [2.156(5)]	Ir1-N2 [2.219(2)]	Ir1-N2 [2.041(3)]
	Ir1-N5 [2.059(5)]	Ir1-N3 [2.042(2)]	Ir1-N3 [2.033(3)]
	Ir1-C1 [2.006(7)]	Ir1-C1 [2.012(3)]	Ir1-C1 [2.021(3)]
	Ir1-C8 [2.023(6)]	Ir1-C13 [1.995(4)]	Ir1-C2 [1.994(4)]
	SFIriqa	MFIrqa	MFIriqa
bond angles (°)	O1-Ir1-N4	O1-Ir1-N2	O2-Ir1-N1
	[76.1(2)]	[75.95(9)]	[75.3(1)]
	N1-Ir1-C1	N1-Ir1-C13	N2-Ir1-C1
	[80.1(2)]	[79.9(1)]	[80.0(1)]
	N5-Ir1-C8	N3-Ir1-C1	N3-Ir1-C2
	[80.3(2)]	[80.2(1)]	[80.0(1)]
	O1-Ir1-C1	O1-Ir1-C13	O2-Ir1-C2
	[173.9(2)]	[175.3(1)]	[170.6(1)]
	N1-Ir1-N5	N1-Ir1-N3	N2-Ir1-N3
	[171.1(2)]	[171.9(1)]	[171.1(1)]
	N4-Ir1-C8	N2-Ir1-C1	N1-Ir1-C1
	[170.7(2)]	[167.1(1)]	[173.9(1)]

Table S2 Representative bond lengths and bond angles for MFIriqa, MFIrqa, andSFIriqa.

М	FIriqa	Μ	FIrqa	SI	FIriqa
Types of bond	distance (Å)	Types of bond	distance (Å)	Types of bond	distance (Å)
С-НС	2.778, 2.780, 2.840, 2.841, 2.851, 2.855	С-НО	2.561, 2.609	С-Нπ	2.775, 2.791, 2.793, 2.801, 2.816
С-НО	2.601, 2.680, 2.715	С-Нπ	2.792, 2.824	С-НО	2.541, 2.572
С-Нπ	2.791, 2.851			С-НС	2.835, 2.855
$\pi\pi$	3.375			С-НН	2.179

Table S3. The intermolecular interactions of MFIriqa, MFIrqa, and SFIriqa crystals.

Table S4 Calculated energy levels, oscillator strengths, and orbital transition analyses of lowest triplet state (T1) states for complexes MFIriqa, MFIrqa, SFIrqa, and SFIriqa.

Complex	State	eV	f	Assignment	Character
MFIriqa	T_1	1.83	0.00	H→L(96%)	³ MLCT/ ³ LLCT
MFIrqa	T_1	1.91	0.00	H→L(36%)	³ MLCT ³ LLCT / ³ LC
				H→L+1(44%)	³ MLCT ³ LLCT / ³ LC
SFIriqa	T_1	1.86	0.00	H→L(95%)	³ MLCT/ ³ LLCT
SFIrqa	T_1	1.92	0.00	H→L(37%)	³ MLCT ³ LLCT / ³ LC
				H→L+1(42%)	³ MLCT ³ LLCT / ³ LC

H stands for HOMO; L stands for LUMO.

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Fig. S13 PL spectra of MFIriqa, MFIrqa, SFIrqa, and SFIriqa in the film.



Fig. S14 Cyclic voltammograms of complexes MFIriqa, MFIrqa, SFIrqa, and SFIriqa in degassed CH_3CN solution with 0.1 M TBAPF₆ as electrolyte (scan rate = 100 mV/s)



Fig. S15 The intermolecular interactions of MFIriqa.



Fig. S16 The intermolecular interactions of MFIrqa.



Fig. S17 The intermolecular interactions of SFIriqa.



Fig. S18 PL spectra of (A) SFIrqa, (B) SFIriqa, (C) MFIrqa and (D) MFIriqa in DMSO/H₂O with different H₂O fractions (*fw*). Concentration: 0.1 μ M; Excitation wavelength: 460 nm.

2. ROS generation measurement

General ROS detection by DCFH-DA

The general ROS generation assay used 2,7-dichlorodihydrofluorescein (DCFH) as an indicator, which was converted from 2,7- dichlorodihydrofluorescein diacetate (DCFH-DA), responded with an aqueous solution of NaOH for 30 min at 4°C. The hydrolysate was then neutralized with ultrapure water to get the stock solution with a concentration of 40 μ M. Typically, DCFH-DA stock solution (working concentration: 5 μ M) and PBS were mixed with NPs solution, the mixtures were exposed to white light irradiation (30 mW cm⁻²), and the PL intensities at 525 nm (Excitation wavelength: 488 nm) were recorded at regular intervals in a PL spectrofluorometer.

Detection of singlet oxygen(¹O₂) by ABDA

The ¹O₂ generation was studied by using ABDA as an indicator as the absorbance of

ABDA decreases upon reaction with ${}^{1}O_{2}$. ABDA solution (working concentration: 20 μ M) was added to different AIE NPs (working concentration: 2 μ M), and the mixtures were exposed to white light irradiation (30 mW cm⁻²). After that, the absorbance of ABDA at 378 nm was recorded for different durations of white light irradiation to obtain the decay rate of the photosensitizing process.

Detection of •OH by HPF

The •OH generation measurements were conducted using hydroxyphenyl fluorescein (HPF) as the indicator. When •OH is generated in the solution, the HPF will be oxidized and fluoresce intensely green centered at 515 nm. HPF solution (working concentration: 5 μ M) and PBS were added to different AIE NPs (working concentration: 2 μ M), and the mixtures were exposed to white light irradiation (30 mW cm⁻²). The PL intensities at 515 nm (Excitation wavelength: 490 nm) were recorded at regular intervals in a PL spectrofluorometer.

3. Cellular study

Preparation of Nanoparticles

5 mg of **MFIriqa**, **MFIrqa**, **SFIriqa**, and **SFIrqa**, and 10 mg of DSPE-mPEG-2000 were respectively dissolved in 10 mL THF solution, which was poured into 10 mL ultrapure water under sonication (200 W). The dissolution obtained was further kept in sonication (200 W) for 15 min. Subsequently, the AIE NPs were obtained by filtration through a polyethersulfone (PES) syringe-driven filter ($0.45/0.22 \mu m$). To purify AIE NPs, they were washed with a 10 K centrifuge filter units (Millipore) under centrifugation at 8000 r.p.m. for 8 min. Finally, the four AIE NPs solutions were concentrated into 5 mL ultrapure water and stored at 4°C for further investigation.

Cell culture

Hela cells were cultured DMEM (containing 10% FBS and 1% Penicillin-Streptomycin solution) at 37 °C in a humidified incubator with 5% CO_2 .

Cell cytotoxicity evaluations

The cell viability of AIE NPs on Hela cancer cells was examined by CCK-8 assay. Briefly, Hela cells were seeded in 96-well microplates at a density of 5×10^3 cells/well in 100 µL of complete DMEM media and incubated at 37°C in a humidified incubator with 5% CO₂. Subsequently, the cells reached about 80% confluence, the culture medium was replaced with the fresh medium containing different concentrations (0, 50, 100, 150, and 200 µg/mL) of four NPs. After further incubation for 4 h, the cells were exposed to white light irradiation 30 mW cm⁻² for 30 min. Meanwhile, the **MFIriqa**, **MFIrqa**, **SFIriqa**, and **SFIrqa** NPs incubated cells without white light irradiation were also conducted for the dark cytotoxicity study. After an additional 24 h, each well was directly added 10 µL CCK-8 solution and incubated at 37 °C for 30 min. The absorbance of sample wells and control wells at 450 nm was also measured by a microplate reader.

Cell imaging

Before confocal microscopy imaging of cells with analytes, Hela cells in the exponential phase were plated on 35 nm glass bottom culture dishes for 24 h to reach around 80% confluency. After incubation with DMEM containing **MFIriqa**, **MFIrqa**, **SFIriqa**, and **SFIrqa** NPs solution (working concentration: 10 μ g/mL) for 4 h, the cells were further washed by PBS three times and imaged by CLSM in the channel mode at excitation of 488 nm. The emission filter was 650 - 750 nm.

Co-localization imaging

Before confocal microscopy imaging of cells with analytes, Hela cells in the exponential phase were plated on 35 nm glass bottom culture dishes for 24 h to reach around 80% confluency. Cells were first incubated with 1 mL DMEM containing **MFIriqa**, **MFIrqa**, **SFIriqa**, and **SFIrqa** NPs (working concentration: 10 µg/mL), 37 °C for 4 h. The DMEM was then removed, and the cells were further washed by PBS three times. Subsequently, cells were incubated with 1 mL DMEM containing BODIPY (1 µM, stock solution: 1 mM in DMSO), 37 °C for 30 min. The cells were

further washed by PBS three times and imaged by CLSM. For NPs, the excitation was 488 nm and the emission filter was 650 - 750 nm. For BODIPY, the excitation was 488 nm and the emission filter was 500 - 540 nm.

ROS generation measurement in vitro

The Hela cells were seeded in 6-well microplates at a density of 5×10^4 cells/well in 1 mL of complete DMEM media and incubated at 37 °C in a humidified incubator with 5% CO₂. Cells were first incubated with 1 mL DMEM containing **MFIriqa**, **MFIrqa**, **SFIriqa**, and **SFIrqa** NPs (working concentration: 200 µg/mL), 37°C for 4 h. The DMEM was then removed, and the cells were further washed by PBS three times. Then, the cells were incubated with 1 mL DMEM containing DCFH-DA (10 µM, stock solution: 10 mM in DMSO), 37 °C for 20 min, the cells were further washed by PBS three times and imaged by CLSM in the channel mode at excitation of 488 nm. The emission filter was 500 - 600 nm.



Fig. S19 Fluorescence intensity changes of (A) **SFIrqa**, (B) **SFIriqa**, (C) **MFIrqa** and (D) **MFIriqa** (2 μM) in DCFH-DA with increasing irradiation time upon white light (30 mW cm⁻²).



Fig. S20 Absorption of ABDA in ultrapure water of (A) SFIrqa, (B) SFIriqa, (C) MFIrqa and (D) MFIriqa (2 μ M) after exposure to white light irradiation of 30 mW cm⁻² with different time.



Fig. S21 Fluorescence intensity changes of HPF for •OH detection. (A) SFIrqa, (B) SFIriqa, (C) MFIrqa and (D) MFIriqa (2 μ M) after exposure to white light irradiation of 30 mW cm⁻² with different time.



Fig. S22 CLSM images of Hela with **MFIriqa**, **MFIrqa**, **SFIriqa**, and **SFIrqa** for 4 h. [AIE NPs 10µg/mL], Ex = 488 nm, Em = 650 - 750 nm.



Fig. S23 Viability of Hela cancer cells after treatment with a range of concentration with white light and without irradiation of 30 mW cm⁻² for 30 min for (A) SFIrqa, (B) SFIriqa, (C) MFIrqa and (D) MFIriqa for 4 h. The data were presented as mean \pm SD (n = 3).



Fig. S24 Intracellular ROS detection by CLSM after Hela cancer cells were incubated with DCFH-DA under white light irradiations (30 mW cm⁻², 10 min), Scale bar = 100 μ m.

List of acronyms

- CLSM = Confocal Laser Scanning Microscopy.
- CCK-8 = Cell Counting Kit-8.
- QY = Fluorescence quantum yield.
- ROS = Reactive oxygen species.
- PDT = Photodynamic therapy.
- AIE = Aggregation-induced emission.
- ACQ = Aggregation-caused quenching.

Cartesian coordinates of optimized T₁ structures

MFIriqa in T₁ state

Ir	0.14326000	-0.58249500	-0.73352000
Ν	1.75820200	-2.04521600	-1.03472100
0	0.49383400	-2.63947800	-4.33627400
0	-0.22685300	-1.40175100	-2.62560100

Ν	-1.28994100	-1.90012400	-0.01293400
Ν	1.49982800	0.84700000	-1.34623800
С	-1.48033600	0.61431600	-0.75306300
Ν	-3.41718300	-2.34175800	0.48721100
С	0.85468800	0.00680900	1.05862400
Ν	3.12235700	2.29437900	-0.87103000
С	1.74637700	-2.65702600	-2.28530300
С	2.75915000	-3.62789000	-2.63945200
С	3.78412600	-3.92936800	-1.67479900
С	3.74237400	-3.24633600	-0.41063800
Н	4.49725800	-3.44666900	0.34330800
С	2.73906900	-2.34055200	-0.15548100
Н	2.69038500	-1.82656800	0.79875000
С	4.78168500	-4.87162200	-1.98468100
Н	5.54595800	-5.08364800	-1.24001200
С	4.80178600	-5.52534900	-3.21788100
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