# **Supporting Information**

# AIE-active TPA modified Schiff base for successive sensing of Cu<sup>2+</sup> and His via on-off-on method and its application in bioimaging

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#### 1. Materials and Apparatus

All reagents were purchased from Energy Chemical or Macklin Biochemical Co., Ltd., and were used directly as received. Tetrahydrofuran (THF) used for spectral determination was of HPLC grade, other reagents were of analytical grade. The solutions of metal ions were prepared from Al(ClO<sub>4</sub>)<sub>3</sub>·9H<sub>2</sub>O, CaCl<sub>2</sub>, Cd(ClO<sub>4</sub>)<sub>2</sub>·H<sub>2</sub>O,  $Co(ClO_4)_2 \cdot 6H_2O_7$  $CrCl_3 \cdot 6H_2O$ , FeCl<sub>3</sub>·6H<sub>2</sub>O, KCl, LiCl,  $MgCl_2 \cdot 6H_2O$ , Mn(ClO<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O, NaCl, Ni(ClO<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O, Zn(ClO<sub>4</sub>)<sub>2</sub>·6H<sub>2</sub>O, Pb(ClO<sub>4</sub>)<sub>2</sub>·3H<sub>2</sub>O, FeCl<sub>2</sub>,  $Hg_2(NO_3)_2 \cdot 2H_2O$ ,  $Cu(Ac)_2$ ,  $Cu(ClO_4)_2 \cdot 6H_2O$ , HgCl<sub>2</sub>,  $Cu(NO_3)_2 \cdot 3H_2O$ and CuCl<sub>2</sub>·2H<sub>2</sub>O. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were acquired on a Bruker Ascend 400 (400 MHz) spectrometer. FT-IR spectra were obtained on a Bruker INVENIO Fourier transform infrared spectrometer using KBr pellets. ESI-HRMS were acquired on a Thermo Fisher Q Exactive mass spectrometer with an ESI interface. Fluorescence were measured by a Thermo Scientific Lumina spectra fluorescence spectrophotometer, with a quartz cuvette (path length = 1 cm). UV-Vis absorption spectra were recorded on an UV-2600 spectrophotometer. pH values were determined on a PHS-3E pH meter. Dynamic light scattering (DLS) data were obtained using a Zetasizer Nano ZS90 laser particle size and zeta potential analyzer (Malvern). Scanning electron microscope (SEM) measurements were performed on a Hitachi SU8100 SEM. Confocal fluorescence imaging was carried out on a Leica TCS SP2 Laser Confocal Microscope.

#### 2. Preparation of compounds 2 and 3

N-(4-Nitrophenyl)-N-N-diphenylamine (2) and N-(4-aminophenyl)-N,N-diphenylamine (3) were prepared according to the reported literature<sup>1, 2</sup>. As shown in Scheme S1, compound 2 was synthesized by the reaction of diphenylamine with 4-fluoronitrobenzene in the presence of sodium hydride. The brown crude product was recrystallized in isopropanol to give pure compound 2 in a yield of 85% (1.2338 g). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  8.10 (d, J = 8.0 Hz, 2H), 7.49 (t, J = 8.0 Hz, 4H), 7.31 (m, 6H), 6.84 (d, J = 8.0 Hz, 2H) ppm. Compound 3 was synthesized by Pd/C-catalyzed reduction of compound 2. The raw product was recrystallized in ethanol to give pure compound 3 in a yield of 91% (0.2369 g). <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.23 (t, J = 8.0 Hz, 4H), 6.92 (m, 6H), 6.83 (d, J = 8.0 Hz, 2H), 6.60 (d, J = 8.0 Hz, 2H), 5.07 (s, 2H) ppm. The <sup>1</sup>H NMR data of compounds 2 and 3 are in agreement the published data<sup>1, 2</sup>.

#### 3. Theoretical calculation methods

All calculations were carried out with the Gaussian 09 software<sup>3</sup>. The PBE0 functional<sup>4</sup> and the def2-SVP basis sets<sup>5, 6</sup> were adopted for geometry optimization calculations. The DFT-D3 with BJ-damping<sup>7</sup> was applied to correct the weak interaction to improve the calculation accuracy. The SMD implicit solvation model<sup>8</sup> was used to account for the solvation effect of water. Orbital energy level analysis was performed by Multiwfn software<sup>9</sup>. The visualization of the orbitals for **HL** and **CuL**<sub>2</sub> complex were achieved using VMD<sup>10</sup> software. The excited states are calculated with linear response time-dependent DFT (TD-DFT) at the optimized ground state geometry. TD-DFT calculation was performed with PBE0 functional and the TZVP basis set<sup>11, 12</sup>.



Scheme S1 The synthetic route for compound 3.



Fig. S1 <sup>1</sup>H NMR spectra of compound 1 in DMSO-d<sub>6</sub>.



Fig. S2 (a) ESI-mass spectrum of compound 1 ( $C_{12}H_9NO_2$ ). (b) The simulation pattern of  $[H(C_{12}H_9NO_2)]^+$ .



Fig. S3 <sup>1</sup>H NMR spectra of HL in DMSO-d<sub>6</sub>.



Fig. S4 <sup>13</sup>C NMR spectra of HL in DMSO-d<sub>6</sub>.



Fig. S5 (a) ESI-mass spectrum of HL  $(C_{30}H_{23}N_3O)$ . (b) The simulation pattern of  $[H(C_{30}H_{23}N_3O)]^+$ .



Scheme S2 The ESIPT process in HL.



Fig. S6 Fluorescence emission spectrum of HL in the solid state,  $\lambda_{ex} = 465$  nm. Inset: Fluorescence photograph of HL in the solid state under 365 nm UV lamp.



Fig. S7 Determination of detection limit of HL for Cu<sup>2+</sup> by fluorescence titration experiment.



Fig. S8 Job's plot of Cu<sup>2+</sup> vs HL in THF/H<sub>2</sub>O (1/9, v/v) based on the absorbance at 343 nm.



Fig. S9 ESI-HRMS of HL upon the addition of 0.5 equiv. Cu<sup>2+</sup> in THF.



Fig. S10 Association constant calculations by plotting  $(A-A_0)^2/[(A_1-A_0)(A_1-A)]$  vs  $1/[Cu^{2+}]$  with linear fitting as well to find out the binding constant of  $CuL_2$ , where A = absorbance at 343 nm at any given concentration of  $Cu^{2+}$ ,  $A_0$  = absorbance maxima at 343 nm in presence of  $Cu^{2+}$ , and  $A_1$  = absorbance maxima at 343 nm in the absence of  $Cu^{2+}$ .



Fig. S11 ESI-HRMS of  $CuL_2$  upon the addition of 8.0 equiv. His in THF.



Fig. S12 UV-Vis (a) and fluorescence (b) spectra of HL (20  $\mu$ M) and HL (20  $\mu$ M)+4.0 equiv. His in THF/H<sub>2</sub>O (1:9, v/v).



Fig. S13 ESI-HRMS of HL+His in THF/H<sub>2</sub>O.



Fig. S14 <sup>1</sup>H NMR spectra of HL, His and HL+His in DMSO- $d_6/D_2O$ .



Fig. S15 Determination of detection limit of  $CuL_2$  for His by fluorescence titration experiment.



Fig. S16 Fluorescence spectra of HL (a), HL+Cu<sup>2+</sup> (b) and HL+Cu<sup>2+</sup>+His (c) in THF/H<sub>2</sub>O (1:9, v/v) with different pH values,  $\lambda_{ex}$ = 420 nm.



Scheme S3 The possible hydrolysis of HL.



Fig. S17 ESI-HRMS of HL in THF/H<sub>2</sub>O with pH values of 10 (a) and 13 (b).



Fig. S18 <sup>1</sup>H NMR of HL in DMSO- $d_6/D_2O$  with NaOD concentrations of 0.1 mM (pH 10) (a), 10 mM (pH 12) (b) and 0.1 M (pH 13) (c).

Refs.	Probe	Analyte	Response Mode	Detection Media	Emission Wavelength	Detection Limit	Test Strips	Cell Imaging	рН
13	S N-N H N-N	Cu <sup>2+</sup>	Turn-off	ACN/H <sub>2</sub> O, (7:3, v/v), pH=7.00	495 nm	$1.25 \times 10^{-8}$ M	_		3-8
14		Cu <sup>2+</sup>	Ratiometric	DMF/H <sub>2</sub> O, (2:8, v/v)	318 nm	$2.13 \times 10^{-9} M$	Yes		4-10
15	H <sub>3</sub> CO OH H <sub>3</sub> CO OCH <sub>3</sub>	Cu <sup>2+</sup>	on-off	H <sub>2</sub> O/THF (4:1, v/v)	525 nm	1.49 × 10 <sup>-7</sup> M	Yes		5-10
16		Cu <sup>2+</sup>	colorimetric	THF /EtOH	428 nm	$0.273 \times 10^{-6} \mathrm{M}$		Yes	
17	SH N <sup>+</sup>	Cu <sup>2+</sup>	turn-on	HEPES/EtOH, (v/v = 1/1), pH=7.4,	784 nm	$1.24 \times 10^{-7} \text{ M}$		Yes	4-9
18		Cu <sup>2+</sup>	on-off	DMSO	360 nm			Yes	
19	OH N, N, OH	Cu <sup>2+</sup>	on-off	THF/H <sub>2</sub> O (1:1, v/v)	598 nm	$2 \times 10^{-7} M$	Yes		

Table S1 Performance of HL compared with recent reported fluorescence probes for Cu<sup>2+</sup> and His.

20		Cu <sup>2+</sup>	on-off	EtOH/H <sub>2</sub> O (1:1, v/v), (HEPES=10 mM, pH=7.4)	417 nm	$8.85 \times 10^{-7} \mathrm{M}$	 Yes	7.4
21		Cu <sup>2+</sup>	on-off	THF/H <sub>2</sub> O (4:1, v/v),2.00 × 10 <sup>-3</sup> M Tris- HCl buffer, pH=7.00)	598 nm	$2.687 \times 10^{-7} M$	 	3-11
22		Cu <sup>2+</sup>	on-off	HEPES/CH <sub>3</sub> C N (0.5:9.5, v/v, 10 mM HEPES buffer, pH=7.4)	546 nm	$2.9 \times 10^{-10} \mathrm{M}$	 Yes	4-8
23		Cu <sup>2+</sup>	on-off	H <sub>2</sub> O	402 nm	$2.7 \times 10^{-9} M$	 Yes	4.85- 11.24
24		Cu <sup>2+</sup>	colorimetric	DMSO/H <sub>2</sub> O (7:3, v/v), 5 mM NaAc- HAc , pH=7.0	520 nm	$1.2 \times 10^{-7} M$	 _	
25	NH HN NH HN NH HN NH HN N NH HN N N N N	Cu <sup>2+</sup>	on-off	THF/H <sub>2</sub> O (4:1, v/v), pH = 7.0	598 nm	$2.36 \times 10^{-7} M$	 	2-11
26	HC HC NO <sub>2</sub> N <sup>-N</sup> NO <sub>2</sub> N <sup>-N</sup>	Cu <sup>2+,</sup> His	on-off-on	DMSO/H <sub>2</sub> O (1:1, v/v)	410 nm	$14 - 18 \times 10^{-7}$ M, $6 - 11 \times 10^{-7}$ M	 	6-7
27	$\begin{array}{c c} HO & & & \\ HO & H & \\ HO & H & \\ HO & & H \\ HN & & \\ HN & & \\ O & CI & \\ CI & \\ HN & & \\ O & CI & \\ HN & \\ HN & \\ O & CI & \\ HN & \\ H$	His	colorimetric	H <sub>2</sub> O (0.01 M HEPES buffer, pH=7.4)	443 nm	$8.5 \times 10^{-8}$ M, $1.9 \times 10^{-7}$ M	 	



Table S2 Crystal data and structure refinement for HL and ZnL<sub>2</sub>.

Identification code	HL	ZnL <sub>2</sub>
Empirical formula	C <sub>30</sub> H <sub>23</sub> N <sub>3</sub> O	$\mathrm{C}_{60}\mathrm{H}_{44}\mathrm{N}_{6}\mathrm{O}_{2}\mathrm{Zn}$
Formula weight	441.51	946.38
Crystal system	Monoclinic	Triclinic

Space group	$P2_1/n$	рĪ	
a/Å	15.6216(9)	11.9862(10)	
b/Å	9.9838(5)	14.4619(13)	
c/Å	16.1858(9)	15.5093(13)	
a/°	90	85.024(2)	
β/°	113.984(2)	72.211(2)	
γ/°	90	74.526(2)	
Volume/Å <sup>3</sup>	2306.4(2)	2467.1(4)	
Z	4	2	
$\rho_{calc}g/cm^3$	1.271	1.274	
μ/mm <sup>-1</sup>	0.078	0.548	
F(000)	928.0	984.0	
Index ranges	$-18 \le h \le 20, -13 \le k \le 13, -21 \le 1 \le 21$	$-14 \le h \le 9, -17 \le k \le 17, -18 \le l \le 18$	
Reflections collected	22181	18149	
Independent reflections	5607 [ $R_{int} = 0.0453, R_{sigma} = 0.0404$ ]	8416 [ $R_{int} = 0.0352$ , $R_{sigma} = 0.0473$ ]	
Data/restraints/parameters	5607/0/308	8416/87/677	
Goodness-of-fit on F <sup>2</sup>	1.045	1.055	
Final R indexes [I>=2σ (I)]	$R_1 = 0.0613, wR_2 = 0.1613$	$R_1 = 0.0482, wR_2 = 0.1218$	
Final R indexes [all data]	$R_1 = 0.0899, wR_2 = 0.1809$	$R_1 = 0.0615, wR_2 = 0.1339$	
Largest diff. peak/hole / e Å <sup>-3</sup>	0.21/-0.14	0.49/-0.37	
CCDC number	2211845	2211846	

 $R_1 = \Sigma(|F_o| - |F_c|)/|F_o|; \ wR_2 = \{\Sigma[(w|F^2_o| - |F^2_c|)^2/\Sigma w(F^2_o)^2]\}^{1/2}.$ 

# Table S3 Selected bond distances (Å) and angles (deg) for HL and $ZnL_2$ .

HL				
O1-C21	1.343(2)	C26-C30	1.383(2)	
N1-C19	1.275(2)	C26-C27	1.383(3)	
N1-C16	1.4177(19)	C12-C11	1.374(3)	
N2-C13	1.433(2)	C12-C7	1.380(3)	
N2-C6	1.398(2)	C21-C22	1.395(2)	
N2-C12	1.417(2)	C16-C17	1.372(2)	
C20-C25	1.386(2)	C16-C15	1.389(3)	
C20-C19	1.456(2)	C18-C17	1.385(2)	
C20-C21	1.401(2)	C11-C10	1.370(3)	
C24-C25	1.390(2)	C14-C15	1.383(2)	
C24-C26	1.478(2)	C23-C22	1.368(3)	
C24-C23	1.391(2)	C2-C3	1.379(3)	
C13-C18	1.373(3)	C3-C4	1.357(3)	
C13-C14	1.374(3)	C5-C4	1.373(3)	
N3-C29	1.323(3)	C10-C9	1.364(4)	
N3-C28	1.321(3)	C7-C8	1.378(3)	

C6-C1	1.379(3)	C30-C29	1.376(3)			
C6-C5	1.394(2)	C27-C28	1.374(3)			
C1-C2	1.377(3)	C9-C8	1.363(4)			
C19-N1-C16	121.95(15)	O1-C21-C20	121.95(15)			
C6-N2-C13	120.09(14)	O1-C21-C22	119.57(17)			
C6-N2-C12	122.44(14)	C22-C21-C20	118.48(17)			
C12-N2-C13	117.39(14)	C17-C16-N1	116.64(15)			
C25-C20-C19	119.17(15)	C17-C16-C15	118.33(15)			
C25-C20-C21	119.24(15)	C15-C16-N1	125.03(16)			
C21-C20-C19	121.59(16)	C13-C18-C17	120.48(18)			
C25-C24-C26	120.93(15)	C10-C11-C12	120.8(2)			
C25-C24-C23	116.83(16)	C13-C14-C15	120.62(18)			
C23-C24-C26	122.23(14)	C16-C17-C18	120.98(17)			
C18-C13-N2	120.18(17)	C22-C23-C24	121.96(15)			
C18-C13-C14	119.03(16)	C14-C15-C16	120.52(18)			
C14-C13-N2	120.77(17)	C1-C2-C3	120.6(2)			
C28-N3-C29	114.68(19)	C4-C3-C2	119.1(2)			
C1-C6-N2	120.83(16)	C4-C5-C6	120.6(2)			
C1-C6-C5	117.98(17)	C3-C4-C5	121.1(2)			
C5-C6-N2	121.19(17)	C9-C10-C11	120.3(2)			
C2-C1-C6	120.68(19)	C8-C7-C12	119.4(2)			
C20-C25-C24	122.63(16)	C23-C22-C21	120.85(17)			
C30-C26-C24	123.40(16)	C29-C30-C26	120.1(2)			
C30-C26-C27	114.91(18)	C28-C27-C26	120.56(19)			
C27-C26-C24	121.69(15)	C8-C9-C10	119.1(2)			
C11-C12-N2	119.48(18)	N3-C29-C30	125.0(2)			
C11-C12-C7	118.88(19)	N3-C28-C27	124.6(2)			
C7-C12-N2	121.64(18)	C9-C8-C7	121.3(2)			
N1-C19-C20	122.00(16)					
N1-Zn1	2.007(2)	O1-Zn1	1.919(2)			
N4-Zn1	2.018(2)	O2-Zn1	1.920(2)			
C16-N1-Zn1	121.97(19)	C19-N1-Zn1	119.1(2)			
C46-N4-Zn1	121.46(19)	C49-N4-Zn1	119.4(2)			
C25-O1-Zn1	124.9(2)	C51-O2-Zn1	124.77(19)			
N1-Zn1-N4	125.33(10)	01-Zn1-N1	97.47(10)			
O1-Zn1-N4	109.34(10)	O1-Zn1-O2	116.57(10)			
O2-Zn1-N1	112.68(9)	O2-Zn1-N4	96.75(9)			

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