

Supporting information

Copper(II) complex enhanced chemodynamic therapy through GSH depletion and autophagy flow blockade

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Materials and experimental procedures

Materials

Bax [E63], Bcl-2 [E17], DDIT-3 [9C8] (CHOP) antibodies were purchased from Abcam (Cambridge, UK). Phospho-eIF2 α (Ser51) (D9G8) antibody was purchased from Cell Signaling Technology (CA, USA). Phospho-PERK (Thr982) antibody was purchased from Invitrogen (CA, USA).

Experimental procedures

•OH determination by EPR

Equal molar copper(II) complex and GSH were dissolved in DMF and distilled water, respectively. And they were mixed and reacted for 5 min at r. t.. H₂O₂ and DMPO were added to the mixture. Subsequently, the mixture was diluted with distilled water, and H₂O₂ and DMPO were added to detect •OH by Bruker A300 Spectrometer.

Cytotoxic activity

The cells were incubated for 48 h after dosing. MTT was added for 4-6 h. The upper liquid was poured out, DMSO was added, shaken for 5 min, and the absorbance at 570 and 490 nm was read. The growth inhibition was calculated as: $(1 - OD_{\text{test}}/OD_{\text{control}}) \times 100\%$.

Intracellular GSH/GSSG ratio detection

T24 cells were treated with C2 (18 μ M) for 48 h. Cells were collected and washed. The remaining steps of GSH and GSSG content detection were carried out according to the manufacturer's instructions. Both GSH and GSSG test kits were purchased from Solarbio (China). Finally, GSH/GSSG ratio was obtained by dividing GSH by GSSG.

Cellular Ca²⁺ detection

T24 cells were treated with C2 (12, 15, 18 μM), respectively. After 24 hours, cells were stained with Ca²⁺ fluorescent dye Fura-3AM for 30 minutes. Subsequently, the treated cells were further washed with PBS, and the changes of cellular Ca²⁺ were immediately detected by flow cytometry.

Caspase-3/9 activity determination

T24 cells were treated with C2 (12, 15, 18 μM), respectively. After 24 hours, cells were collected and stained with FITC-DEVD-FMK (caspase-3) and FITC-LEHD-FMK (caspase-9) at 37 °C for at least 30 minutes. And flow cytometry was used to analyze the cells with activated caspase.

Structural characterization of all compounds

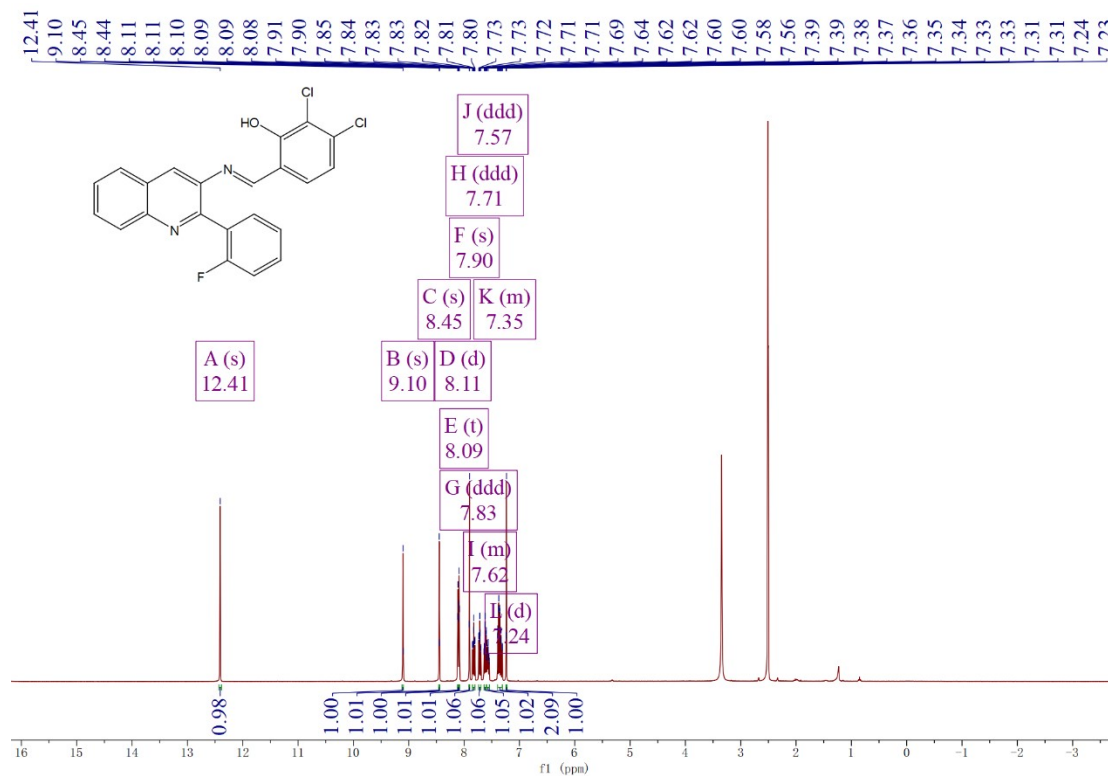


Figure S1. ¹H NMR (400MHZ, DMSO-*d*₆) spectrum of L¹

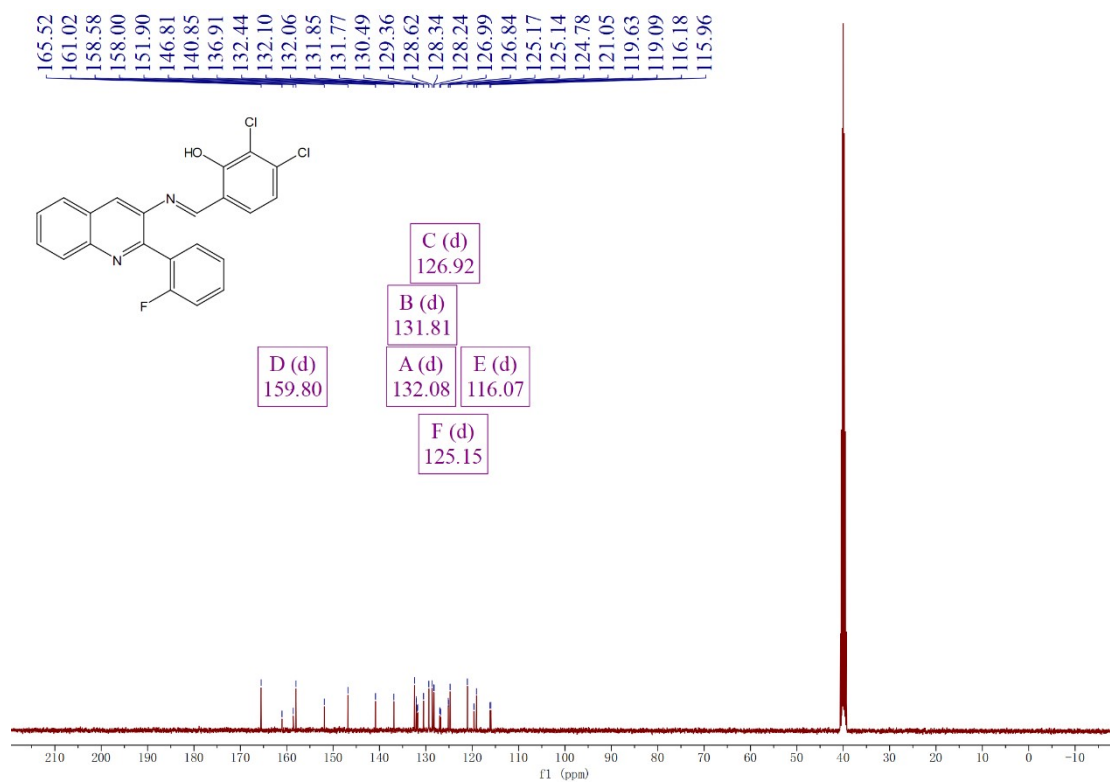


Figure S2. ¹³C NMR (100MHZ, DMSO-*d*₆) spectrum of **L¹**

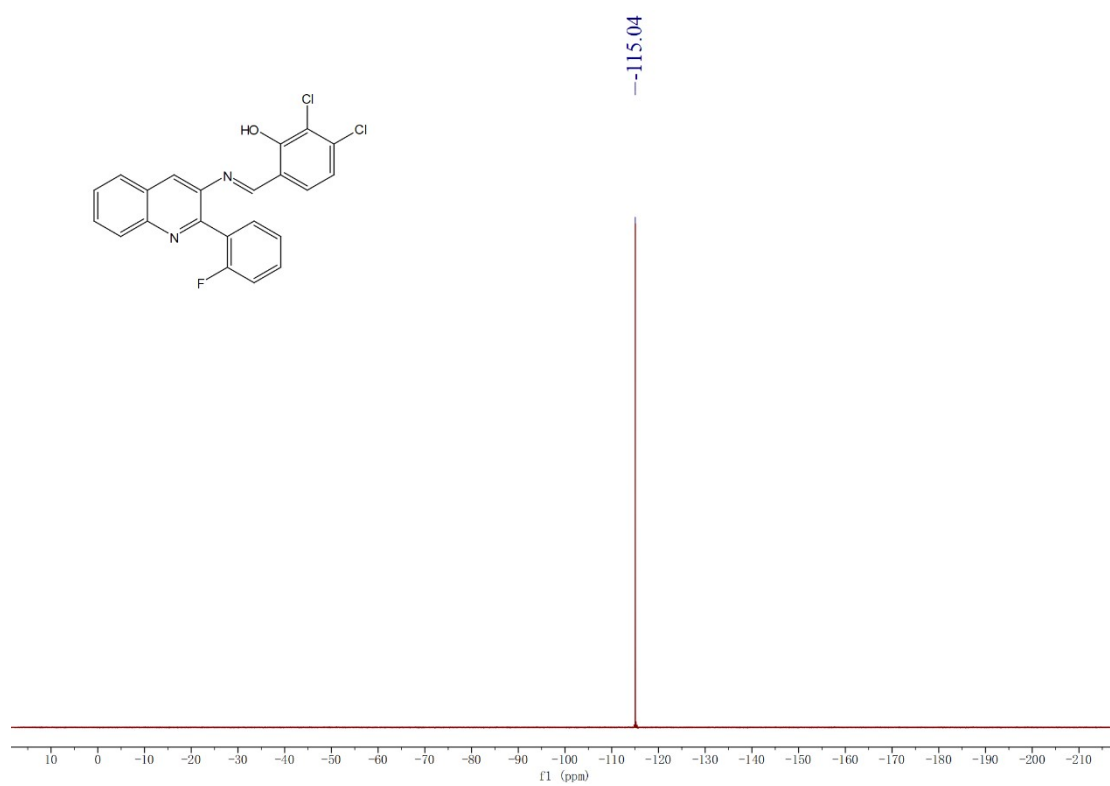


Figure S3. ¹⁹F NMR (376MHZ, DMSO-*d*₆) spectrum of **L¹**

20201013-SWY-L26-3.4-2Cl-7 #24 RT: 0.08 AV: 1 NL: 1.48E6
T: FTMS - p ESI Full ms [200.00-1000.00]

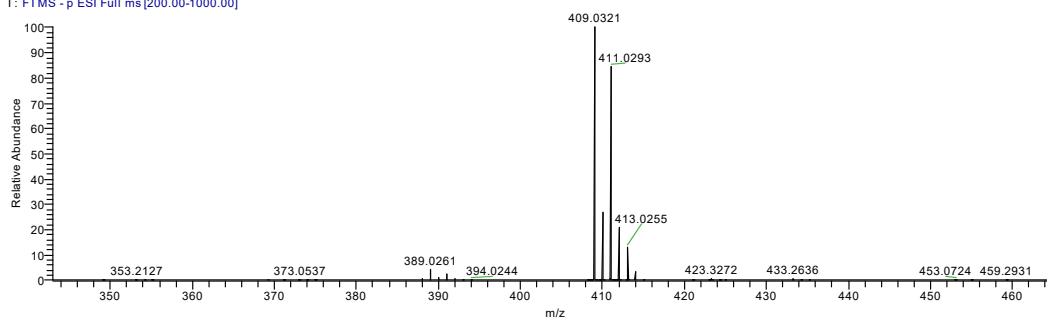


Figure S4. HRMS spectrum of L¹

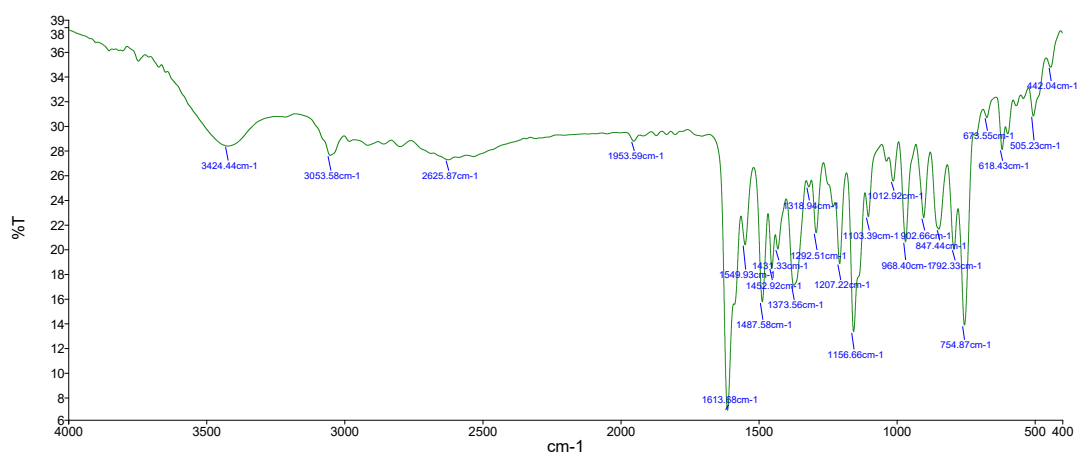


Figure S5. FT-IR spectrum of L¹

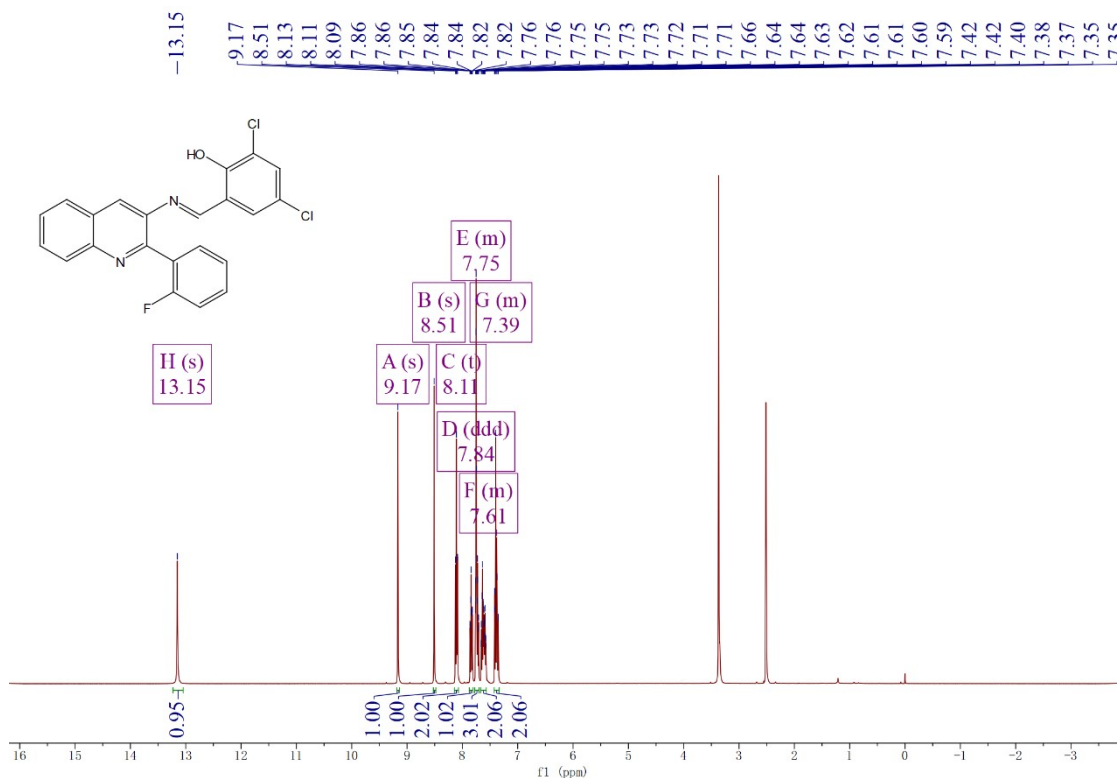


Figure S6. ¹H NMR (400MHZ, DMSO-*d*₆) spectrum of L²

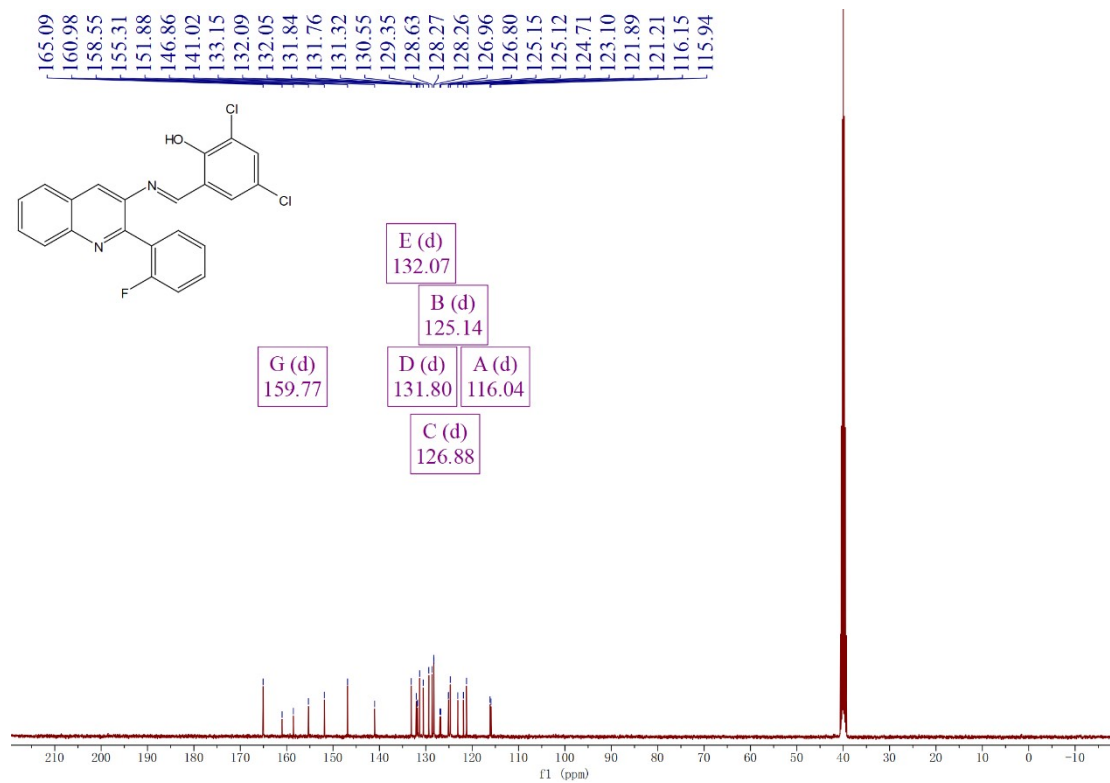
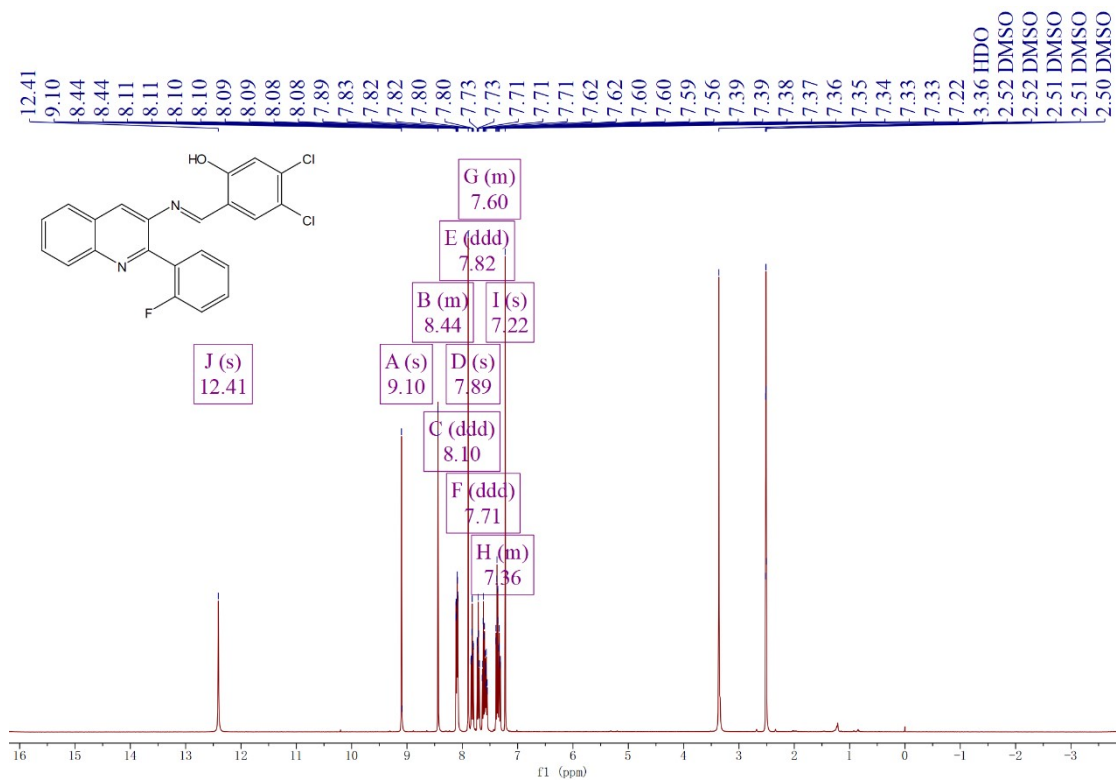
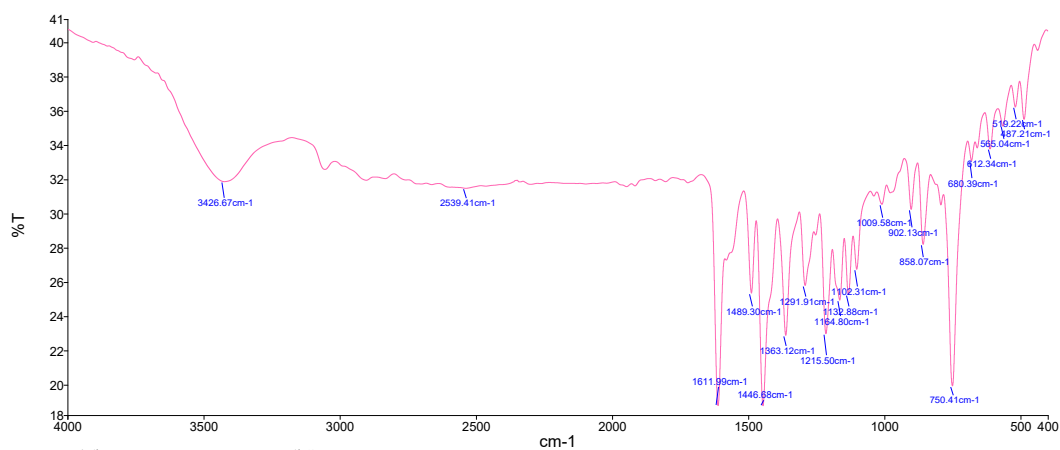
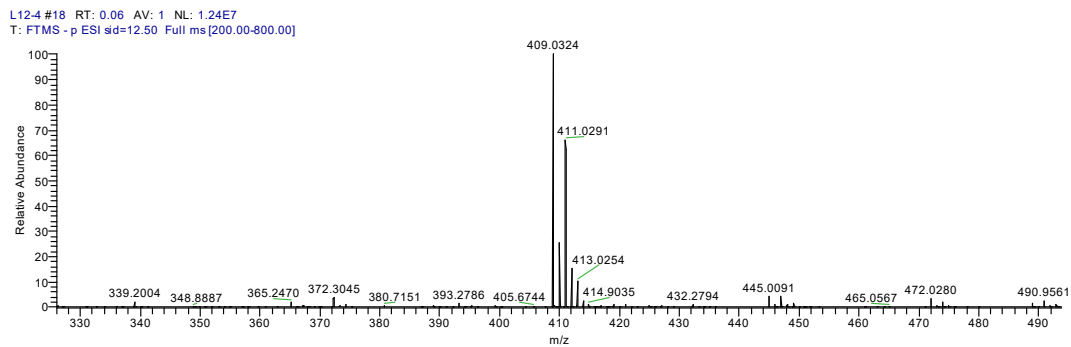


Figure S7. ¹³C NMR (100MHz, DMSO-*d*₆) spectrum of **L²**



Figure S8. ¹⁹F NMR (376MHz, DMSO-*d*₆) spectrum of **L²**



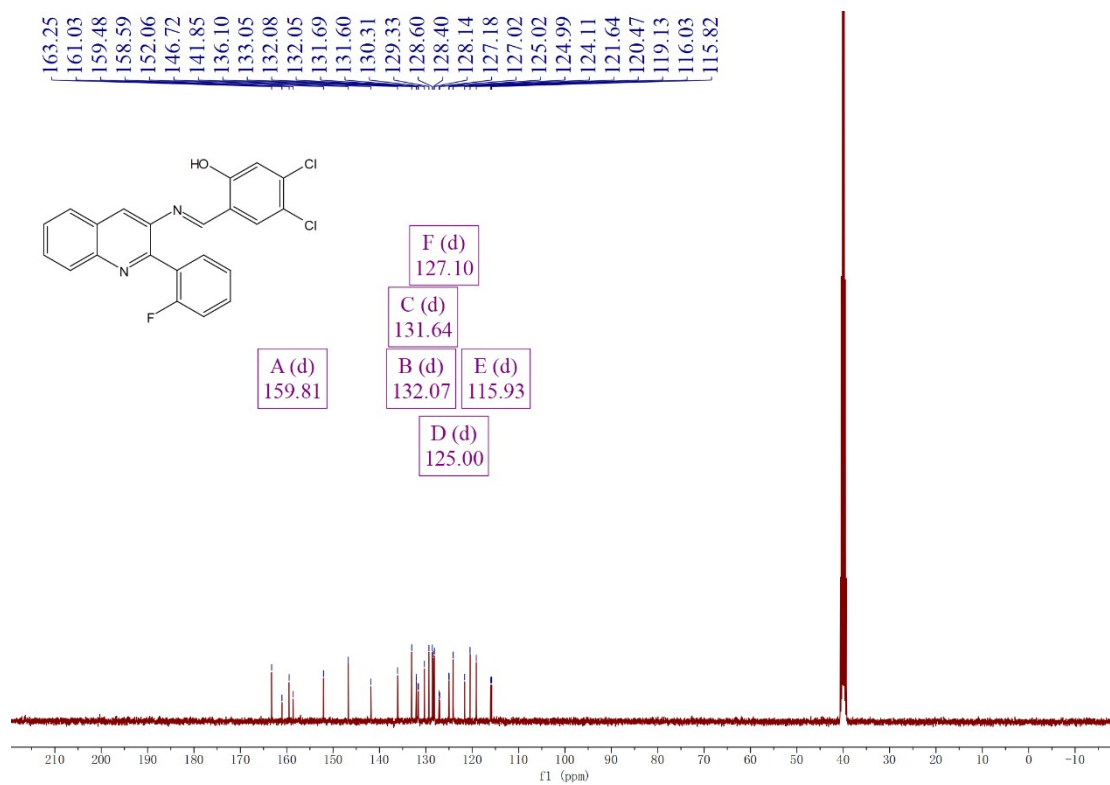


Figure S12. ^{13}C NMR (100MHZ, $\text{DMSO-}d_6$) spectrum of L^3



Figure S13. ^{19}F NMR (376MHZ, $\text{DMSO-}d_6$) spectrum of L^3

20201013-SWY-L25-4.5-2Cl-5 #97 RT: 0.33 AV: 1 NL: 3.33E6
T: FTMS - p ESI Full ms [200.00-1000.00]

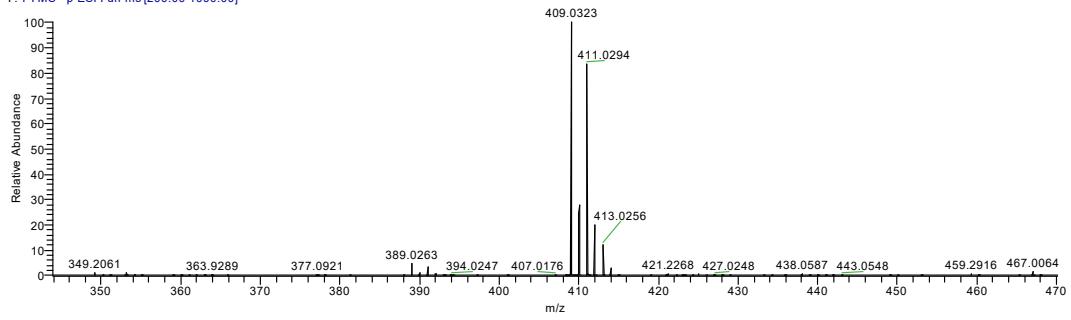


Figure S14. HRMS spectrum of L³

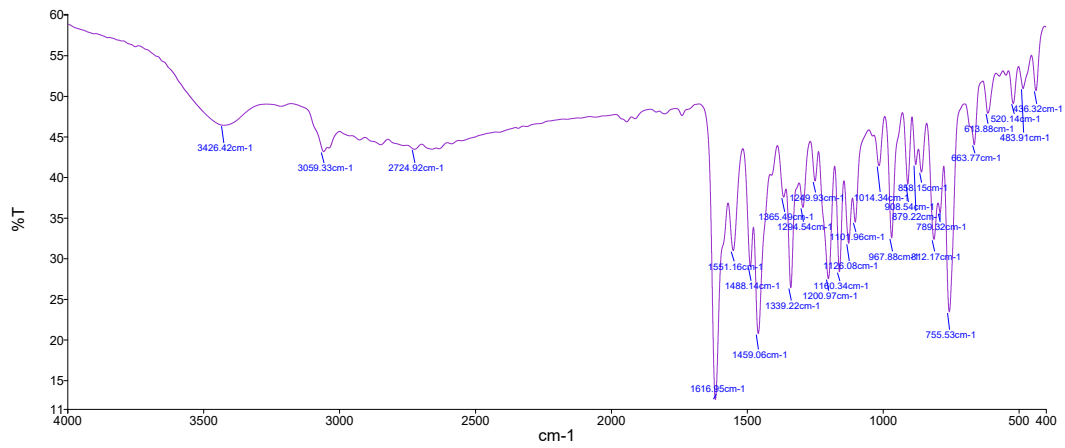


Figure S15. FT-IR spectrum of L³

20201113-SWY-L26-3.4-2Cl-Cu-1 #47-48 RT: 0.16-0.17 AV: 2 NL: 3.50E6
T: FTMS - p ESI Full ms [200.00-1200.00]

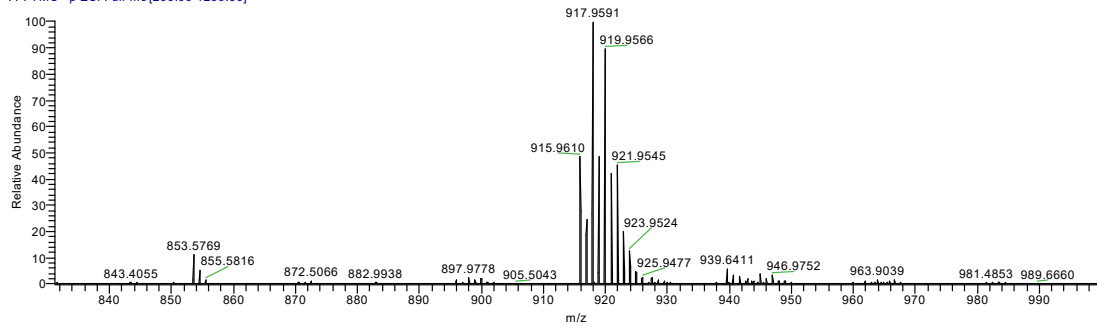


Figure S16. HRMS spectrum of C1

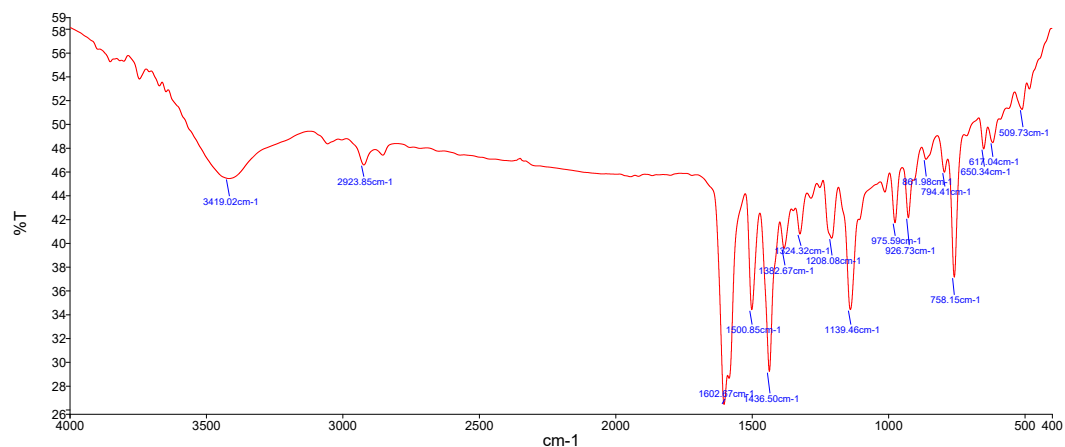


Figure S17. FT-IR spectrum of C1

L12(3.5-Cl)Cu #105 RT: 0.36 AV: 1 NL: 1.33E6
 T: FTMS + p ESI sd=12.50 Full ms [710.00-1000.00]

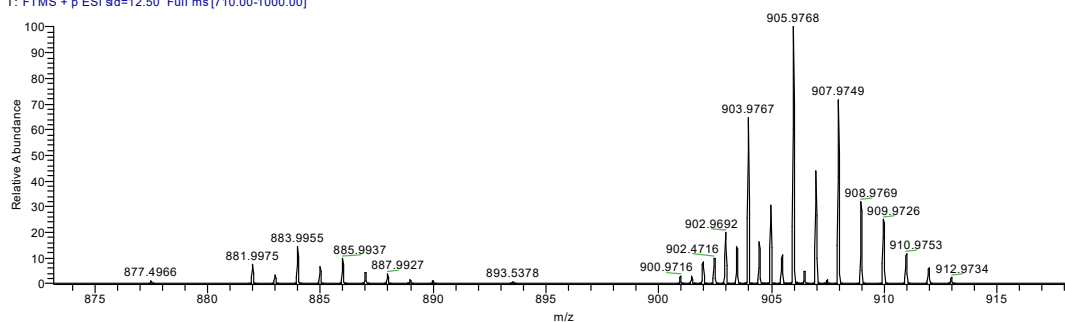


Figure S18. HRMS spectrum of C2

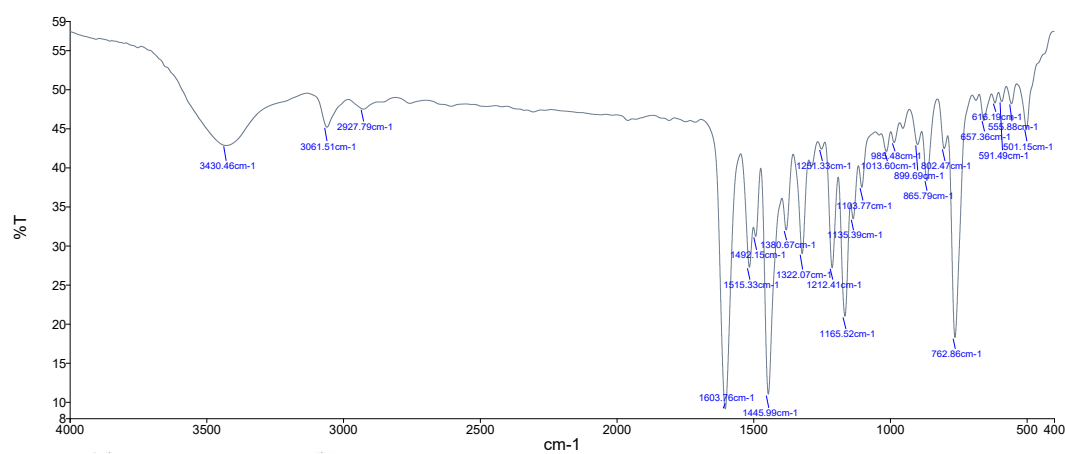


Figure S19. FT-IR spectrum of C2

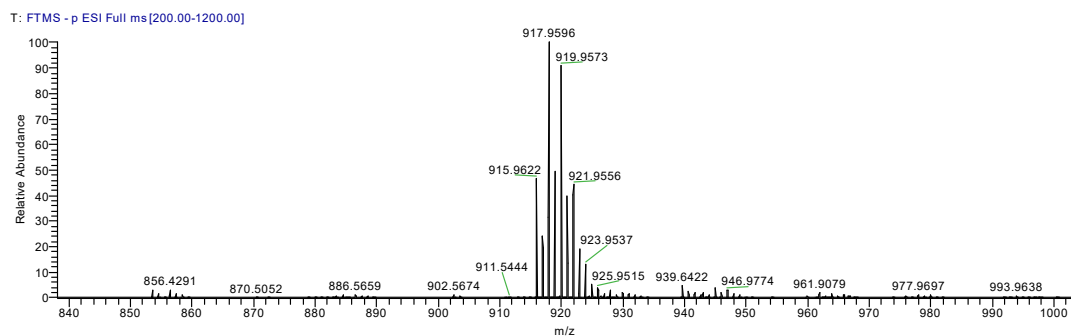


Figure S20. HRMS spectrum of C3

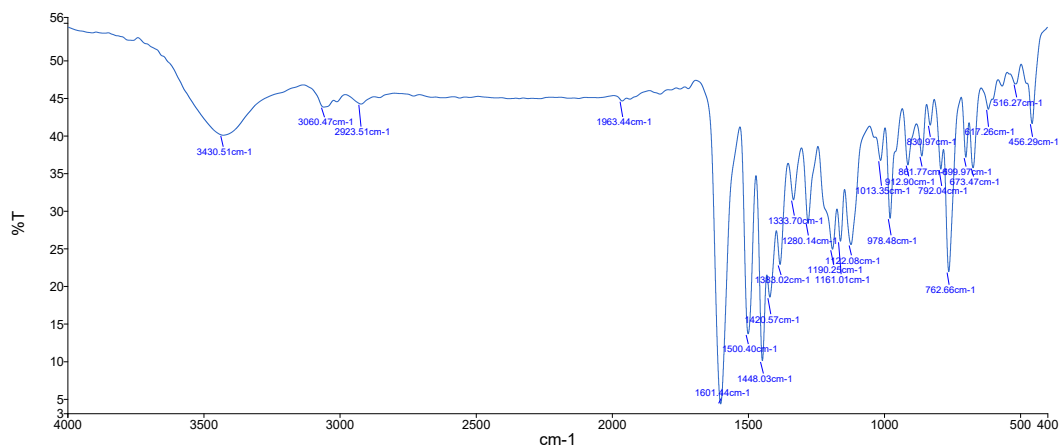


Figure S21. FT-IR spectrum of C3

UV-Vis spectra of C1–C3

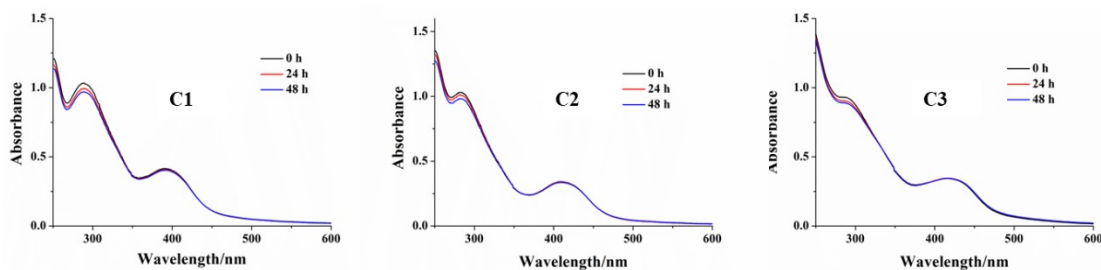


Figure S22. UV-Vis spectra of C1, C2 and C3 in PBS containing 1% DMF

HPLC trace of C1–C3

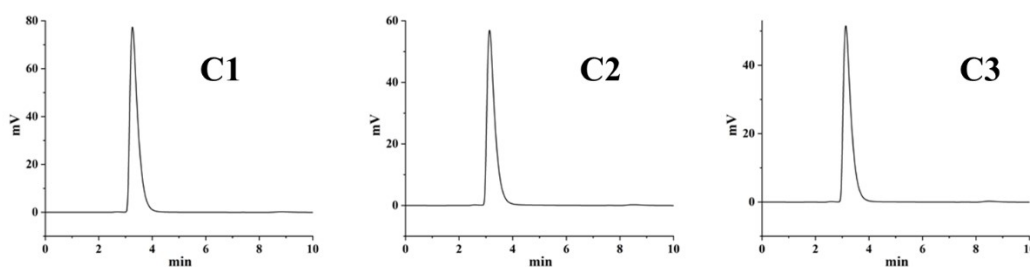


Figure S23. HPLC trace of C1, C2 and C3 in CH₃OH containing 1% DMF

^1H NMR spectra of the reactions of C1–C3 with GSH

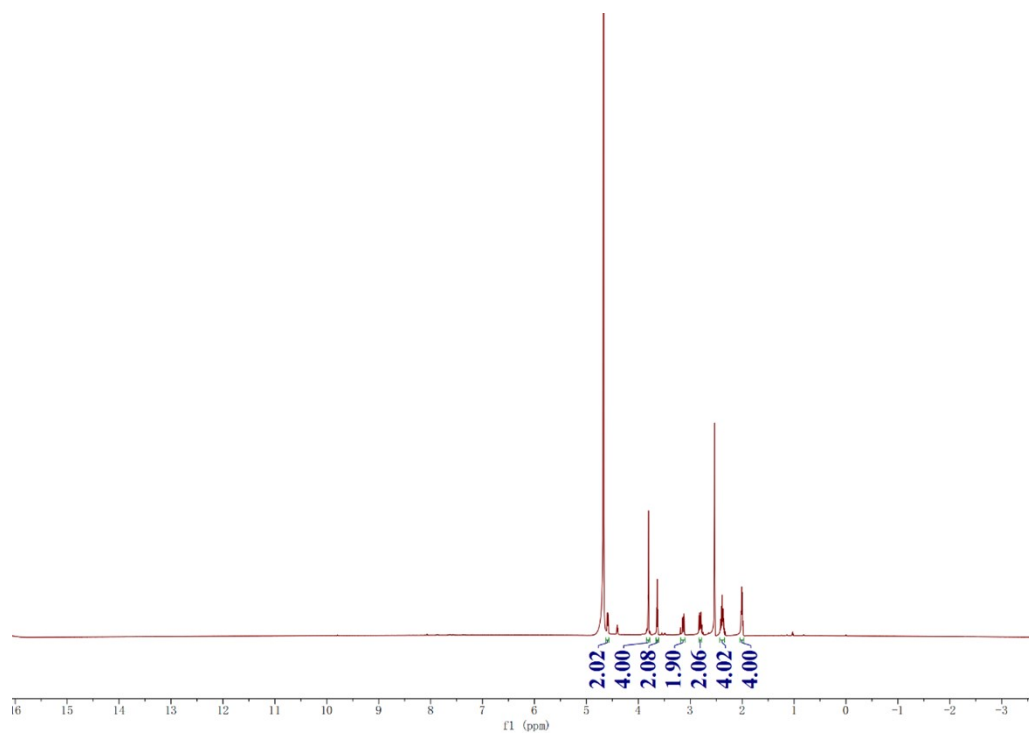


Figure S24. ^1H NMR (600MHz) spectrum of the reaction of C1 with GSH (1:5 proportion of $\text{DMSO-}d_6$ and D_2O).

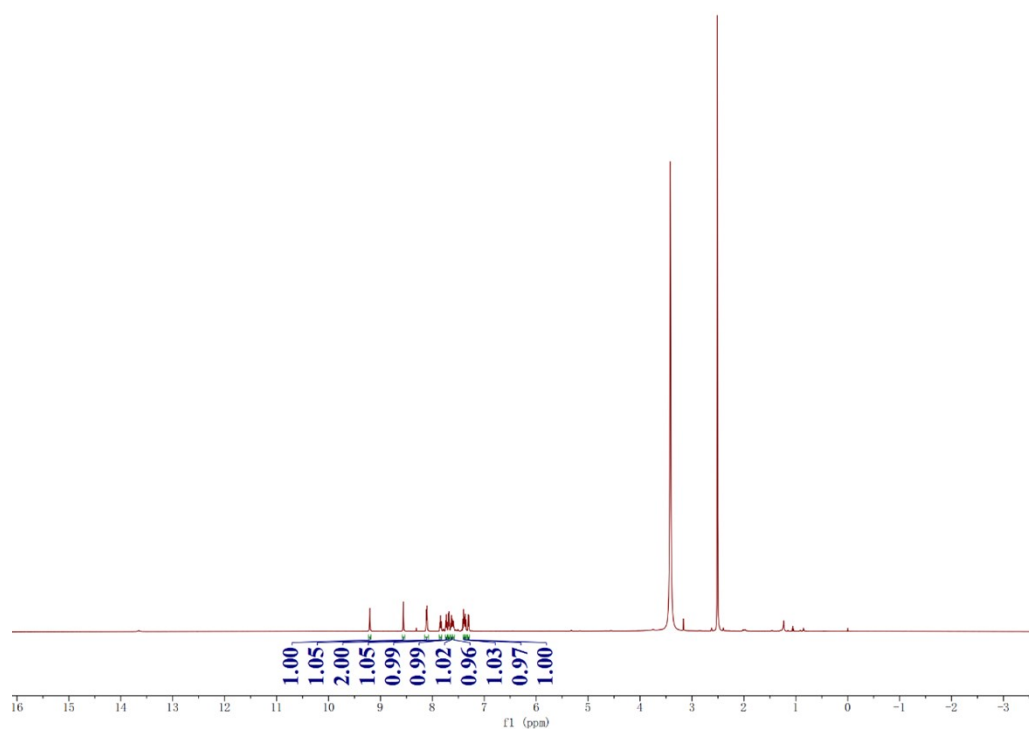


Figure S25. ^1H NMR (600MHz) spectrum of the reaction of C1 with GSH (60:1 proportion of $\text{DMSO-}d_6$ and D_2O).

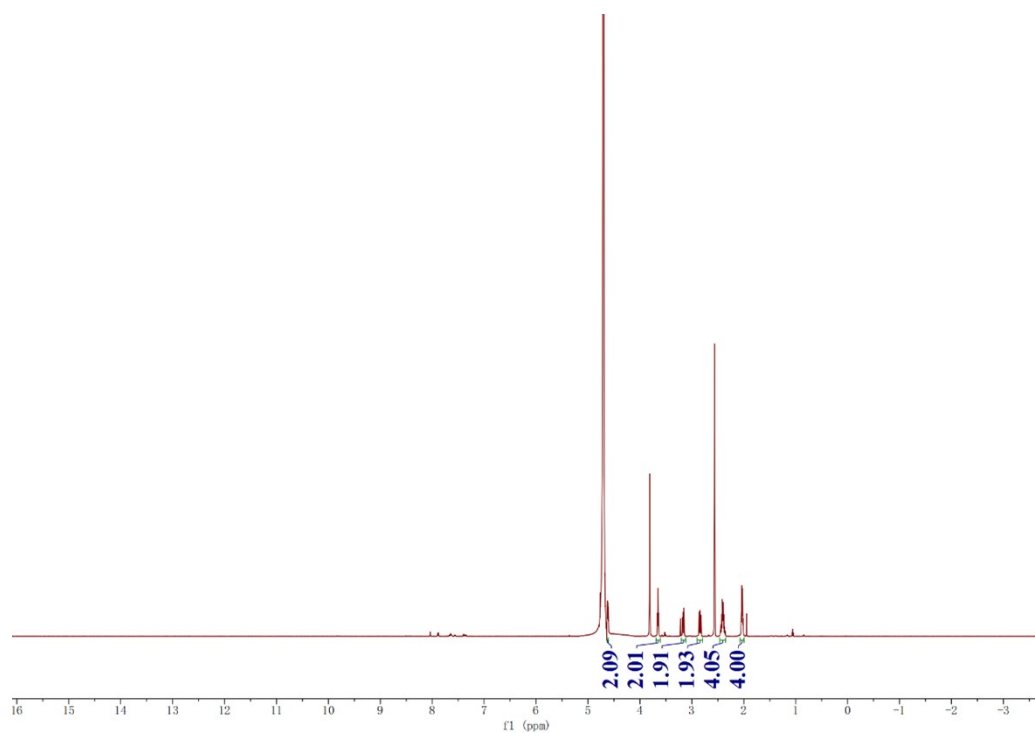


Figure S26. ¹H NMR (600MHz) spectrum of the reaction of **C2** with GSH (1:5 proportion of DMSO-*d*₆ and D₂O).

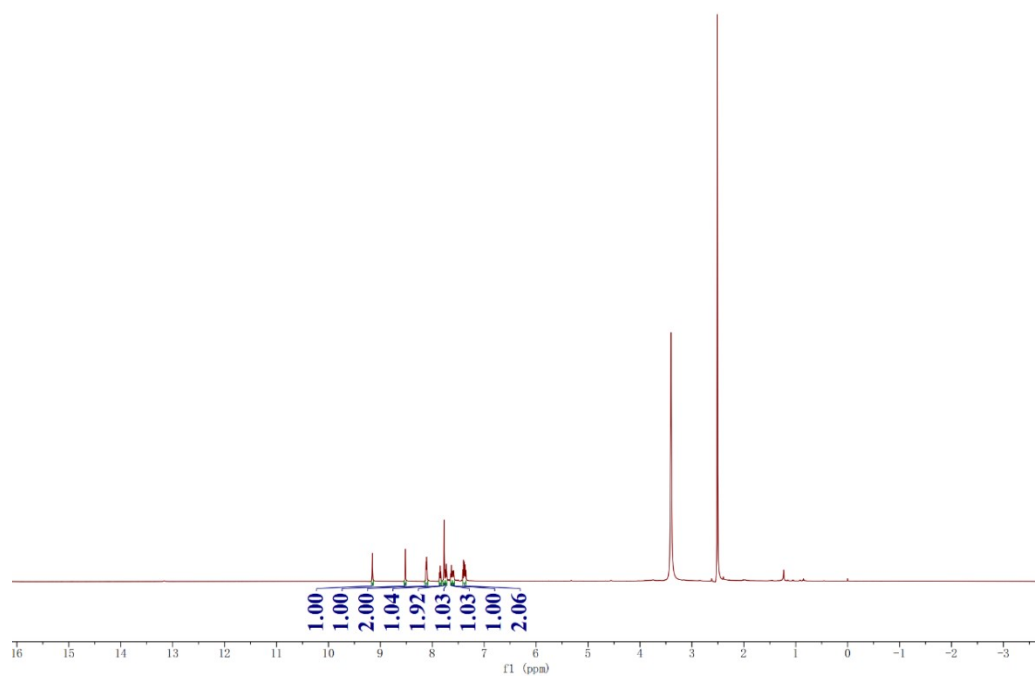


Figure S27. ¹H NMR (600MHz) spectrum of the reaction of **C2** with GSH (60:1 proportion of DMSO-*d*₆ and D₂O).

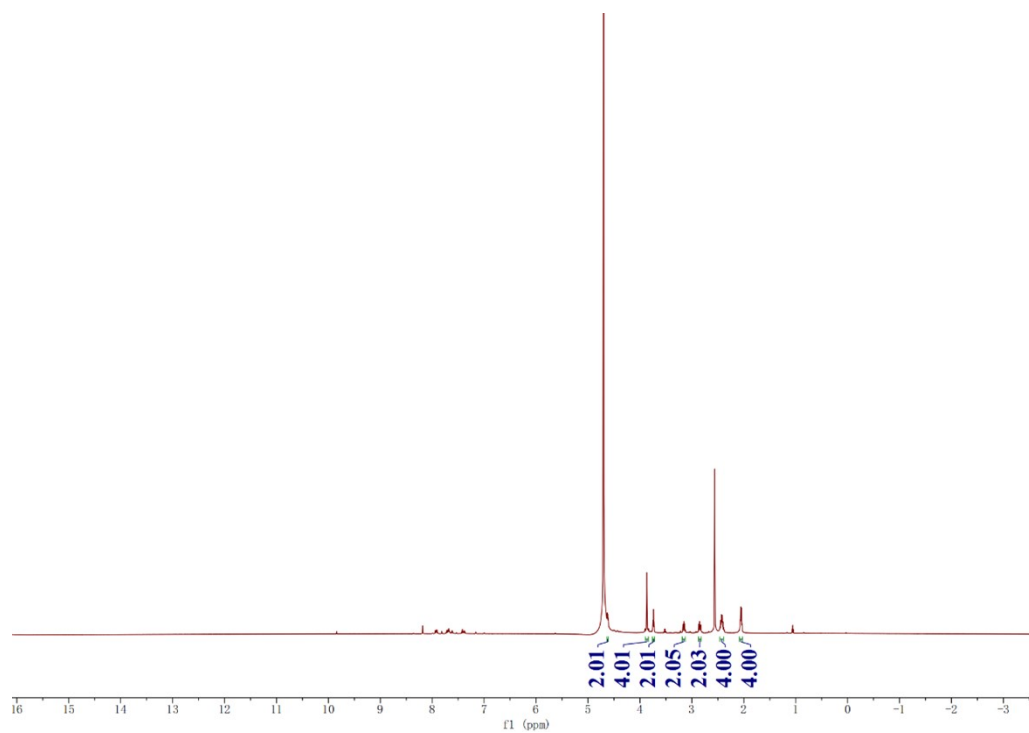


Figure S28. ¹H NMR (600MHz) spectrum of the reaction of **C3** with GSH (1:5 proportion of DMSO-*d*₆ and D₂O).

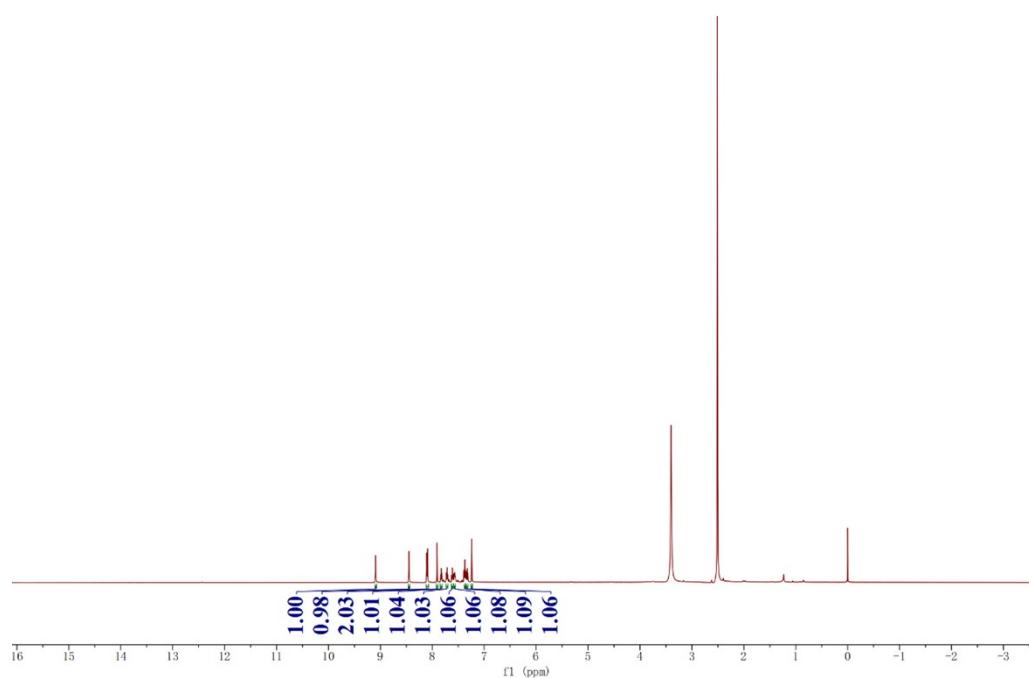


Figure S29. ¹H NMR (600MHz) spectrum of the reaction of **C3** with GSH (60:1 proportion of DMSO-*d*₆ and D₂O).

Crystal data of C1–C3

Table S1. A summary of crystallographic data of **C1**, **C2** and **C3**

	C1	C2	C3
Empirical formula	C ₄₄ H ₂₄ Cl ₄ CuF ₂ N ₄ O ₂	C ₄₄ H ₂₄ Cl ₄ CuF ₂ N ₄ O ₂	C ₄₆ H ₃₂ Cl ₄ CuF ₂ N ₄ O ₂
Formula weight	884.01	884.01	948.09
Crystal system	Monoclinic	Monoclinic	Monoclinic
Space group	<i>P2₁/n</i>	<i>C2/c</i>	<i>C2/c</i>
a/Å	10.0633 (1)	19.6802 (6)	24.3709 (2)
b/Å	19.9246 (1)	9.9152 (2)	11.5828 (1)
c/Å	10.1296 (1)	20.2125 (6)	15.5789 (1)
α/°	90	90	90
β/°	110.210 (1)	94.951 (3)	102.622 (1)
γ/°	90	90	90
Volume/Å ³	1906.01 (3)	3929.39 (19)	4291.38 (6)
Z	2	4	4
ρ _{calc} /cm ³	1.540	1.494	1.467
μ/mm ⁻¹	3.85	0.88	3.49
F(000)	894	1788	1932
CCDC	2115672	2115673	2115674

Table S2 Selected bond lengths[Å] and angles [°] of **C1**, **C2** and **C3**

	C1	C2	C3
Cu1—O1 ⁱ	1.9112 (11)	1.8933 (13)	1.9003 (11)
Cu1—O1	1.9113 (11)	1.8933 (13)	1.9002 (11)
Cu1—N1 ⁱ	1.9692 (13)	1.9941 (16)	1.9904 (13)
Cu1—N1	1.9692 (13)	1.9941 (16)	1.9903 (13)
O1 ⁱ —Cu1—O1	180.0	180.0	88.53 (7)
O1—Cu1—N1 ⁱ	89.19 (5)	91.15 (6)	149.69 (6)
O1—Cu1—N1	90.81 (5)	88.85 (6)	93.19 (5)
O1 ⁱ —Cu1—N1 ⁱ	90.81 (5)	88.85 (6)	93.18 (5)
O1 ⁱ —Cu1—N1	89.18 (5)	91.15 (6)	149.69 (6)
N1 ⁱ —Cu1—N1	180.0	180.0	100.18 (8)