

1 Supplemental Information

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3 Understanding the Impact of Different Source Water Types on the Biofilm Characteristics and
4 Microbial Communities of Manganese Removing Biofilters

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15 Additional Materials and Methods

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17 Water Quality Analysis

18 TOC was measured with a TOC analyzer (TOC-VCSH TOC Analyzer, Shimadzu) following

19 Method 5310 B.¹ True colour was measured using the Platinum-Cobalt Standard Method on a

20 HACH DR5000 following Method 8025.¹ Alkalinity was measured using a T50 auto titrator

21 (Mettler Toledo) following Method 2320.¹ Phosphate and ammonia were measured using a HACH

22 DR5000 following Method 8048 and the salicylate method (#8155) respectively.¹ To quantify

23 dissolved metals, samples were filtered through a 0.45 µm cellulose nitrate membrane. Filtered and

24 unfiltered samples were acidified with trace metals nitric acid to a pH of < 2 and analysed by

25 inductively coupled plasma mass spectrometry (ICP-MS, Thermofisher X Series II) following

26 Standard Method 3125 to determine the concentration of total and dissolved metals respectively.²⁻⁴

27 Cellular ATP was quantified in duplicate for 50 mL aliquots of raw water and filter effluent water
28 using commercial test kits for aqueous samples (Quench Gone Aqueous, LuminUltra Technologies
29 Ltd.). Raw water and filter effluent water were also filtered through a sterile 0.22 um nitrocellulose
30 membrane and genomic DNA was extracted from the filters using a commercial test kit (QIAGEN
31 DNeasy Power Water Kit, QIAGEN). All analyses were initiated within 24 hrs of sample
32 collection.

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34 Filter Media Analysis

35 Mn and Fe on the surface of filter media was analysed following EPA Method 3050B.⁵ Filter media
36 was dried at 105°C for approximately 72 hrs before being digested. Particles were removed from
37 solution by decanting, centrifugation, and filtration (0.45 µm). Mn and Fe in the filtered digestate
38 were quantified by ICP-MS following Standard Method 3125.² Total ATP was quantified in
39 duplicate using a commercial test kit for biofilm samples (Deposit and Surface Analysis,
40 LuminUltra Technologies Ltd.) and EPS was extracted in triplicate following the method of.⁶ The
41 protein fraction of EPS was quantified in duplicate using a commercial test kit (Pierce BCA Protein
42 Assay Kit, Thermo Scientific) and the carbohydrate fraction was quantified in duplicate following
43 the method of DuBois et al., (1956) as modified by Keithley and Kirisits (2018).^{6,7} HPC
44 enumeration was performed following Method 9215.¹ Genomic DNA was extracted using a
45 commercial kit for biofilm samples (DNeasy Power Biofilm Kit, QIAGEN). Wet filter media was
46 weighed in triplicate then dried at 105°C for 72 hrs and weighed again to determine a dry weight
47 correction factor. The dry weight correction factor was applied to the measurement of metals, tATP,
48 protein, and carbohydrate on filter media.

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50 Additional Data

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52 Table S1. Duration, in seconds, for each stage of the backwash cycle at the M-GW/SW, Shediac,
53 and W-GW biofiltration plants.

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Backwash Cycle	M-GW/SW	S-GW	W-GW
Level Down (s)	120	400	600
Air Scour (s)	120	120	120
Air + Water (s)	200	180	240
Rinse (s)	1100	180	300
Pre-Filtration (s)	780	500	720

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57 Table S2. Average (\pm S.D.) accumulation of Mn and Fe on filter media taken from the M-GW/SW,
58 S-GW, and W-GW biofiltration plants before and after backwashing. Filter media mass has been
59 corrected to dry weight.

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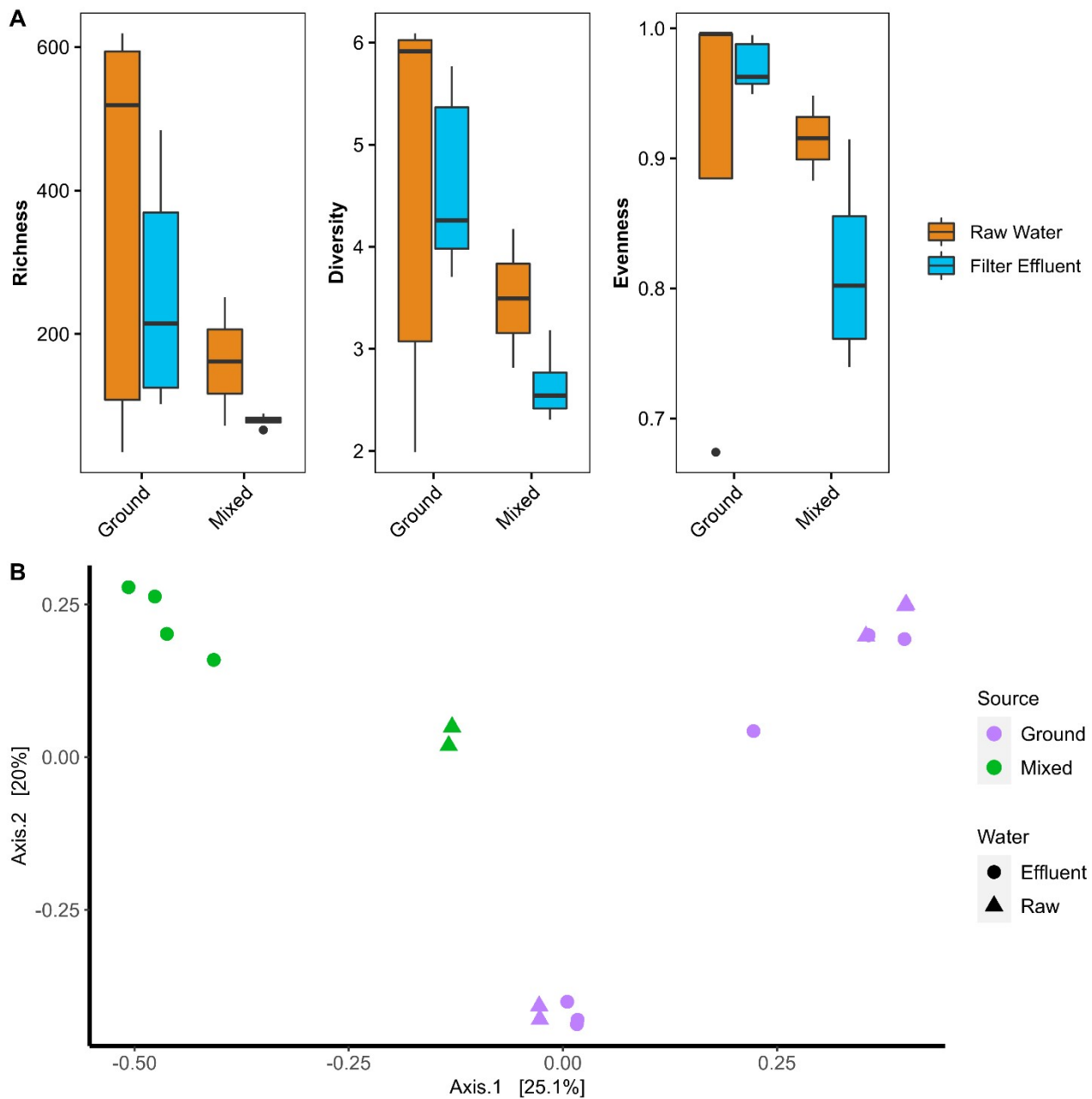
	Sample Location	M-GW/SW	S-GW	W-GW
Mn (mg/g dw)	Pre-Backwash	55 \pm 13	55 \pm 7	76 \pm 26
	Post-Backwash	44 \pm 7	52 \pm 7	112 \pm 38
Fe (mg/g dw)	Pre-Backwash	18 \pm 14	52 \pm 11	0.4 \pm 0.2
	Post-Backwash	12 \pm 6	50 \pm 17	0.8 \pm 1.0

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66 Figure S1. (A) Richness (observed ASVs), diversity (Shannon's Index), and evenness (Simpson's

67 Index) of communities within groundwater and mixed-water biofilters. Richness ($p < 0.05$),

68 diversity ($p < 0.05$), and evenness ($p < 0.05$) were all significantly greater the mixed-water

69 community than in the groundwater communities (Wilcoxon rank sum test with a Benjamini-

70 Hochberg correction for FDR). The horizontal line within the box indicates the median, the upper

71 and lower bounds of the box indicate the third and first quartiles, respectively, and the whiskers

72 extend to the largest value no greater than 1.5 times the interquartile range. Values beyond the

73 whiskers are outliers. (B) PCoA ordination of Bray-Curtis dissimilarity matrix of microbial

74 communities in raw and effluent water samples from groundwater and mixed-water biofilters. The
75 groundwater community was significantly different ($p < 0.001$) from the mixed-water community
76 (Adonis test, 999 permutations).

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