Supporting Information for

# Site-selective methylene C-H oxidation of an alkyl diamine enabled by supramolecular recognition using a bioinspired manganese catalyst

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# **1.** Materials and Methods

## 1.1. Materials

Reagents and solvents used were of commercially available reagent quality unless stated otherwise. Solvents were purchased from SDS, Aldrich, Scharlab and Fluorochem. Sigma-Aldrich HPLC-grade acetonitrile was employed for oxidation reactions.

# 1.2. Instrumentation

<sup>1</sup>H-NMR and <sup>13</sup>C-NMR were performed with a Bruker Ultrashield ASCEND Nanobay spectrometer. NMR titrations and NOESY experiments were performed with a Bruker Ultrashield ADVANCE III400. Spectra were referenced to the residual solvent peaks. Mass spectrometry was performed by electrospray ionization in a Bruker Esquire 6000 Ion trap mass spectrometer using acetonitrile as mobile phase. Samples were introduced into the mass spectrometer ion source by direct infusion through a syringe pump. Chromatographic analysis of the reaction was performed on an Agilent GC-7820-A (Agilent HP-5) column and a flame ionization detector. GC-MS analysis were performed on an Agilent J&W) interfaced with an Agilent 5975X mass spectrometer. NH<sub>3</sub> was used as the ionization gas.

# 2. Synthesis of the catalysts

The complexes  $[Mn(OTf)_2(pdp)]$ ,  $[Mn(OTf)_2(^{CR}pdp)]$ ,  $[Zn(OTf)_2(pdp)]^3$  and  $[Zn(OTf)_2(^{CR}pdp)]^4$  were prepared according to the reported procedures.

### 3. Synthesis of the substrate (S1)



1,14-tetradecandiamine was prepared following a procedure inspired by J. Pinnavaia and co-workers<sup>5</sup> and B. Paulus, C. A. Schalley and co-workers.<sup>6</sup> 1,16-hexadecandioic acid (305.0 mg, 1.07 mmol, 1 equiv.) were solved in 20 mL of CHCl<sub>3</sub> (0.06M). Under N<sub>2</sub> atmosphere, 4.5 mL of concentrated H<sub>2</sub>SO<sub>4</sub> (8.56 mmol, 8 equiv.) were slowly added and the mixture was heated to 55°C. Then 500.0 mg of NaN<sub>3</sub> (8.13 mmol, 7.6 equiv.) were added in small portions during 2 hours under vigorous stirring. The reaction mixture was stirred for two additional hours. The crude was cooled to room temperature and moved to an extraction funnel and the acidic phase was collected in an H<sub>2</sub>O/ice vessel to make the product precipitate. The solid was filtered and solved in 2M NaOH until pH = 11. The aqueous phase was extracted with CHCl<sub>3</sub> (5 x 50 mL) and the combined organic phases were dried over anhydrous MgSO<sub>4</sub>, filtered and the solvent was removed under vacuum to afford the diamine as a white solid (155.0 mg, 0.680 mmol, 66% yield). The product was used in the next step without further purification. Spectral data match those previously reported.<sup>7</sup> <sup>1</sup>H-NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$ , ppm: 2.68 (t, *J* = 7.0 Hz, 4H), 1.47 (br, 4H), 1.45-1.41 (m, 4H), 1.30-1.24 (m, 20H). <sup>13</sup>C-NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$ , ppm: 42.3, 33.7, 29.6, 29.6, 29.5, 26.9.



1,14-tetradecandiamimium tetrafluoroborate **(S1)** was prepared following a procedure inspired by Olivo *et al.*<sup>2</sup> 1,14-tetradecandiamine (155.0 mg, 0.680 mmol, 1 equiv.) was solved in 2 mL of CH<sub>2</sub>Cl<sub>2</sub> (0.5 M) and cooled to 0°C. After 5 minutes of stirring, 570 µL of a 2.63 M CH<sub>2</sub>Cl<sub>2</sub> solution of HBF<sub>4</sub>·Et<sub>2</sub>O (1.496 mmol, 2.2 equiv.) were added dropwise. The mixture was stirred for 30 minutes at 0°C. The solvent was removed and the solid was washed with diethyl ether (8 mL) under stirring for 5 minutes. The supernatant was removed and the washing process was repeated twice and the solid obtained was dried under vacuum. Recrystallization with CH<sub>3</sub>CN affords **S1** as a crystalline white solid (202.0 mg, 0.500 mmol, 74% yield). <sup>1</sup>H-NMR (400 MHz, CD<sub>3</sub>CN)  $\delta$ , ppm: 6.10 (t, *J* = 51.1 Hz, 6H), 2.98-2.88 (m, 4H), 1.67-1.52 (m, 4H), 1.42-1.18 (m, 20H). <sup>13</sup>C-NMR (100 MHz, CD<sub>3</sub>CN)  $\delta$ , ppm: 40.3, 29.4, 29.3, 29.1, 28.7, 26.6, 25.8.





Figure S2. <sup>13</sup>C-NMR spectrum (100 MHz, CD<sub>3</sub>CN) of S1.

# 4. NMR titration

#### 4.1. General procedure

0.5 mL of a 1 mM CD<sub>3</sub>CN catalyst solution of were introduced in an NMR tube and analyzed by <sup>1</sup>H-NMR. Then, portions of a CD<sub>3</sub>CN solution containing substrate (5 mM) and catalyst (1 mM) were periodically added and analyzed after each addition. When 1 equiv. of substrate was added, a NOESY experiment was conducted (mixing time d<sub>8</sub>=0.8 seconds). After the acquisition the titration was finished following the same procedure.

# 4.2. Titration of (*S*,*S*)-Zn(<sup>CR</sup>pdp) with S1



<b>S1</b> (pure)			6			<u>к I / I / I / I / I / I / I / I / I / I </u>
2.51 equiv.			MAR. A MA			
2.34 equiv.			MA. A M	A	L.	
2.18 equiv.	111	1	MA. A M		L	
2.03 equiv.		1	MA. A M		L	
1.91 equiv.			MA. A M			
1.76 equiv.	111	1	MA. A M		L	Wh.
1.69 equiv.	111	1	MA AM		L	wh.
1.61 equiv.		1	MA AM		L	- Main I
1.47 equiv.	111		MA. A M	n. Ma	L	mil.
1.30 equiv.		1	MI AM	n. Man	Lhe	Mila
1.13 equiv.		1	Mr. A.A.	n. Man	L La	Mullin
1.02 equiv.	L h	1	MI. A Mu	n . Allen	L	Alla
0.87 equiv.	I A K	1	MI AM	a. Am	uli	Autor 1
0.64 equiv.			Mr. AM	n. m	ula	Aut I
0.56 equiv.	L A L	1	MA. AM	a. /a.	ula	M.A.
0.49 equiv.		1	MA. A.M.		1.	M.M.
0.22 equiv.	1 A J	1	MA AM		yh	-Uha
0 equiv.		1				
0.5 10.0 9.5 9.0 8.5	8.0 7.5 7.0 6.5	6.0 5.5 5.0 4	.5 4.0 3.5	3.0 2.5	2.0	1.5 1.0 0.5 0.0

Figure S3. Titration of (S,S)-Zn(<sup>CR</sup>pdp) (1 mM) with S1 in CD<sub>3</sub>CN at 25°C.



**Figure S4.** Downfield shift of the crown signals during the titration of (S,S)-Zn(<sup>CR</sup>pdp) (1 mM) with S1 in CD<sub>3</sub>CN at 25°C (enlargement of crown region).



**Figure S5.** Titration curve of the above NMR experiments (Figure S3). A reliable binding constant could not be extracted, but anyway, this curve is consistent with the formation of 1:1 adducts at millimolar concentrations.



Figure S6. COSY spectrum of a 1:1 mixture of (S,S)-Zn(<sup>CR</sup>pdp) : S1 in CD<sub>3</sub>CN (1 mM) at 25°C.



Figure S7. NOESY spectrum of a 1:1 mixture of (S,S)-Zn(<sup>CR</sup>pdp) : S1 in CD<sub>3</sub>CN (1 mM) at 25°C (mixing time d<sub>8</sub>=0.8s).



# 4.3. Control experiment using (S,S)-Zn(pdp)

**Figure S8.** Titration of *(S,S)*-Zn(pdp) (1 mM) with S1 in  $CD_3CN$  at 25°C. <u>Comments</u>: Note that the signals of the complex and S1 do not show any shift, indicating lack of interaction between the Zn complex and the protonated diamine.



**Figure S9.** NOESY spectrum of a 1:1 mixture of **(S,S)-Zn(pdp)** : **S1** in CD<sub>3</sub>CN (1 mM) at 25°C (mixing time  $d_8$ =0.8s). There are no intermolecular correlations, indicating no binding between the complex and the substrate.

#### 5. Catalytic oxidation of S1

#### 5.1. General procedures



General procedure for the oxidation of S1. A CH<sub>3</sub>CN solution (200  $\mu$ L, 0.1 M) of S1 (7.8 mg, 0.0193 mmol, 1 equiv.) and C1 (0.25 mg, 0.193  $\mu$ mol, 1 mol%) was prepared in a 10 mL vial equipped with a stirring bar. 22 equiv. of AcOH (25  $\mu$ L, 0.418 mmol) were then added to the solution. The resulting mixture was cooled to 0°C in an ice bath. 100  $\mu$ L of H<sub>2</sub>O<sub>2</sub> (from a 1.9 M solution in CH<sub>3</sub>CN diluted from 50% in water, 0.193 mmol, 10 equiv.) were directly added by syringe pump over 30 minutes. At this point, the reaction was quenched with 2 mL of 2-propanol and the solvent was removed under reduced pressure. A known amount of 1,3,5-trimethoxybenzene (around 0.5 equiv.) were added in the vial as solid, the mixture was solved in CD<sub>3</sub>CN and analyzed by <sup>1</sup>H-NMR to determine the conversion and yield of the reaction.



**Derivatization of the reaction.**<sup>8</sup> The contents of the NMR tube were evaporated under vacuum and the crude was solve in 2 mL of  $CH_2Cl_2$ . 70 µL of  $Et_3N$  (0.475 mmol, 25 equiv.) and 10 µL of pivaloyl chloride (0.0475 mmol, 2.5 equiv.) were added. The mixture was stirred at room temperature overnight and washed with a saturated solution of NaHCO<sub>3</sub> (2 mL) and 2M HCl (2 mL). The organic phase was filtered through an anhydrous MgSO<sub>4</sub> plug and directly injected in the GC to determine the selectivity of the reaction.



**Jones' oxidation.**<sup>9</sup> After derivatization, the solvent was removed under reduced pressure and the crude obtained was solved in 2 mL of acetone. 200  $\mu$ L of Jones' reagent were added at 0°C and the mixture was stirred overnight at room temperature. The reaction crude is filtered through a Celite<sup>©</sup> plug and directly injected in the GC to determine the selectivity of the reaction.

# 5.2. GC-MS fragmentation of the products













# 6. References

- (1) Ottenbacher, R. V; Samsonenko, D. G.; Talsi, E. P.; Bryliakov, K. P. *Org. Lett.* **2012**, *14*, 4310.
- (2) Olivo, G.; Farinelli, G.; Barbieri, A.; Lanzalunga, O.; Di Stefano, S.; Costas, M. Angew. Chem. Int. Ed. **2017**, *56*, 16347.
- (3) Vicens, L.; Olivo, G.; Costas, M. Angew. Chem. Int. Ed. 2022, 61, e202114932.
- (4) Olivo, G.; Capocasa, G.; Ticconi, B.; Lanzalunga, O.; Di Stefano, S.; Costas, M. Angew. Chem. Int. Ed. **2020**, *59*, 12703.
- (5) Tanev, P. T.; Liang, Y.; Pinnavaia, T. J. J. Am. Chem. Soc. **1997**, *119*, 8616.
- (6) von Krbek, L. K. S.; Achazi, A. J.; Schoder, S.; Gaedke, M.; Biberger, T.; Paulus, B.; Schalley, C. A. *Chem. Eur. J.* **2017**, *23*, 2877.
- (7) Pingen, D.; Schwaderer, J. B.; Walter, J.; Wen, J.; Murray, G.; Vogt, D.; Mecking, S. *ChemCatChem* **2018**, *10*, 3027.
- (8) Milan, M.; Carboni, G.; Salamone, M.; Costas, M.; Bietti, M. ACS Catal. 2017, 7, 5903.
- (9) Eisenbraun, E. J. Org. Synth. **1965**, 45, 28.