

Supplementary information

1. The preparation method of D-SPP

Dried powder of *Sargassum pallidum* was refluxed with 95% ethanol twice at 70 °C for 3 h to remove liposoluble compounds and impurities. The dried residue was extracted with distilled water with a solid to liquid ratio of 1:30 (w/v) at 90 °C for 3 h. After twice extraction, the combined extracts were concentrated, which was then subjected to deproteinization and decolorization according to the previous methods. Then, the resulting solution was precipitated (12 h, 4 °C) by adding dehydrated ethanol to a final concentration of 80% (v/v). The precipitate was centrifuged, collected and lyophilized to obtain crude polysaccharides (SPP). Previous reported method was utilized to prepare partially degraded *S. pallidum* polysaccharides (D-SPP). Namely, SPP aqueous solution was treated with an ultrasonic homogenizer (S-jia Lab Equipment Co., Ltd. Ningbo, China) at 500 W power for 1 h, then the solution was concentrated and lyophilized to obtained D-SPP powder. The molecule weight, chemical components and monosaccharide composition of D-SPP are listed in **Table S1** and **Fig. S1**.

Table S1 Chemical composition and structural properties of D-SPP

Items	D-SPP
Molecule weight (Da)	5.10×10^5
Chemical composition (% g/g)	
Total carbohydrates	$68.72 \pm 3.21\%$
Proteins	$6.54 \pm 0.61\%$
Monosaccharide (molar %)	
Fucose	$18.20 \pm 0.73\%$
Arabinose	$3.88 \pm 0.42\%$
Galactose	$25.51 \pm 1.18\%$
Glucose	$4.14 \pm 0.14\%$
Xylose	$5.67 \pm 0.17\%$
Mannose	$0.80 \pm 0.10\%$
Galacturonic acid	$29.94 \pm 1.47\%$
Glucuronic acid	$8.87 \pm 0.98\%$

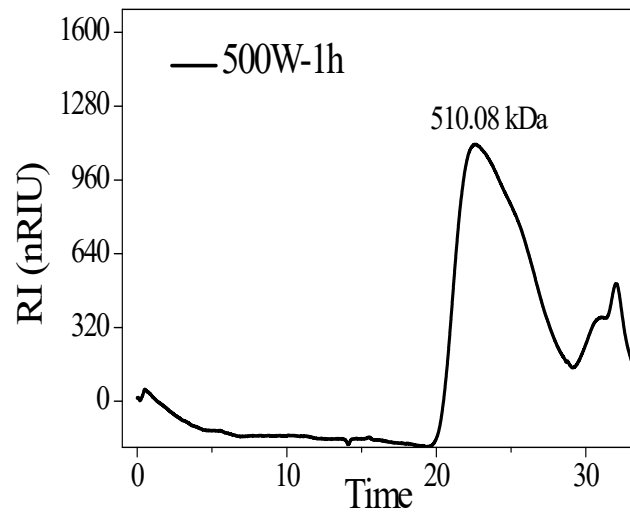


Fig. S1 The HPGPC profile of molecular weight distribution of D-SPP

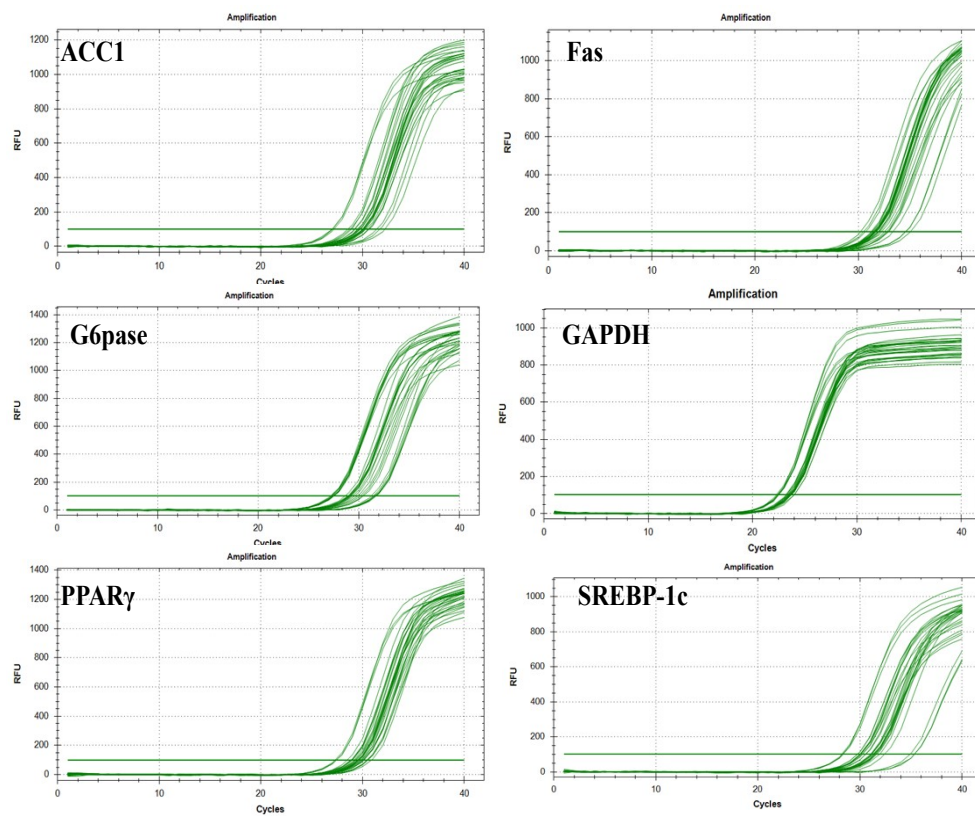


Fig. S2 Amplification curves of cDNA of lipogenesis from liver

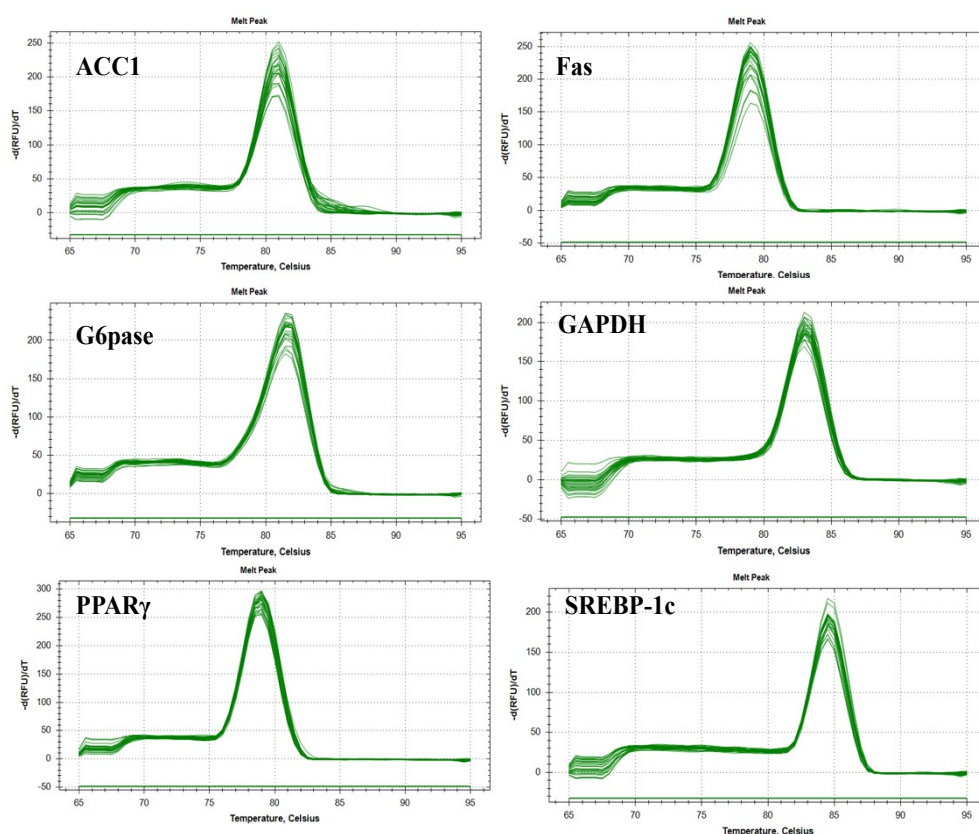


Fig. S3 Dissolution curves of cDNA of lipogenesis from liver

Table S2 Primer Sequences for Quantitative RT-PCR

Genes			Fragment length	A t (°C)
GAPDH	sense primer	CCTCGTCCCGTAGACAAAATG	133	60
	antisense primer	TGAGGTCAATGAAGGGGTCGT		
PPAR γ	sense primer	GACCACTCGCATTCCTTTGACA	155	60
	antisense primer	ATCGCACTTTGGTATTCTTGGA		
FAS	sense primer	TAGAACCTCCAGTCGTGAAACCA	150	60
	antisense primer	ATCTCATCTATCTTGCCCTCCTTG		
ACC1	sense primer	GCTAACCCAACTCAGCAAGACC	238	60
	antisense primer	CACCTGGTTGCTGTGGTAAAAA		
SREBP-1c	sense primer	GAAACCCGAAGTGGTGGAGAC	159	60
	antisense primer	TGGGCTTTGACCTGGCTATC		
G6pase	sense primer	ACACCGACTACTACAGCAACAGC	208	60

antisense primer AATCCCAACCACAAGATGACG
