

*Supplementary Note: Noncompliant participants.*

Ninety-two percent (143 out of 155) of sequenced microbiome samples were from participants compliant with the prune treatment. Alpha and beta diversity, as well as differential taxa, were compared among all samples including noncompliant participants post-intervention (58 Control, 58 50g prune, 39 100g prune).

*Demographics.* For the 155 participants whose fecal microbiomes were sequenced, the average age was 62.3 years (range: 55-75) at baseline, and 98% (n=152) were Caucasian. Participants were mostly healthy or overweight (78 healthy, 52 overweight, 25 obese), with an average BMI of 25.6 kg/m<sup>2</sup> (range: 18.5-39.2).

*Alpha Diversity.* Post intervention, Pielou's Evenness was significant ( $p=0.049$ ), and Shannon trended significant ( $p=0.077$ ) between groups. The 50g prune group had slightly lower alpha diversity than the control and 100g prune groups (Pielou's Evenness, significant,  $q<0.098$ ). These values represent slightly less significance than among compliant individuals only.

*Beta Diversity.* As in compliant individuals only, differences in the microbiomes of control, 50g prune, and 100g prune groups were observed in all four beta diversity metrics tested (Bray-Curtis, weighted Unifrac, significant,  $p<0.03$ ; Jaccard, unweighted Unifrac, trend,  $p<0.07$ ). PermDISP was not significant for any metric ( $p>0.31$ ), so these differences are due to true differences in the means of groups. Bray-Curtis detected significant differences between control and the 50g prune group ( $p=0.036$ ), as well as between control and the 100g prune group ( $p=0.048$ ). The difference between 50g and 100g prunes was a trend ( $q=0.069$ ). For weighted Unifrac, all three comparisons trended significant ( $q=0.076$ ). Jaccard found trends comparing the 100g prune group to the other two groups ( $q=0.0975$ ). On the whole, these results indicate slightly less significance than among compliant individuals only.

*Differential Taxa.* As before, no taxa were significant by ANCOM. However, LEfSe identified 52 taxa as differentially abundant at LDA>2, slightly less than among compliant samples only (75). The same taxa were identified as highly differentially abundant (Lachnospiraceae, LDA=4.5; *Blautia*, 4.4; *Anaerostipes*, 4.2; Oscillospirales, 3.9).

*Phenolics.* Post-intervention, hippuric acid was the only metabolite to be differentially excreted in urine among the 155 participants (Kruskal-Wallis, p=0.001). Hippuric acid was significantly increased in the 50g and 100g prune groups compared to control (p<0.01). These results indicate less significance than among compliant individuals only.

Table S1. Phenolic composition of supplied prunes in year 1 and year 3 of the study. STD is the standard deviation. DW is the dry weight of the prunes.

<b>Phenolic acids and flavonoids</b>				
mg/g DW	Year 1		Year 3	
	Average	STD	Average	STD
Ferulic acid	1.44	0.05	2.42	0.04
Caffeic acid	8.08	0.96	9.27	0.24
p coumaric acid	5.4	0.05	8.77	0.48
protocatechuic acid	3.21	0.11	1.63	0.026
quercetin	0.01	0	0.05	0.01
quercetin-3-glucoside	0.16	0.03	0.26	0.01

<b>Chlorogenic acid derivatives</b>				
mg/g DW	Year 1		Year 3	
	Average	STD	Average	STD
3-Caffeoylquinic acid	59.9	3.7	64.2	5.1
5-Caffeoylquinic acid	4.81	1.2	5.03	0.53
4-Caffeoylquinic acid	68.3	16.5	66.4	3.3
3-Feruleoylquinic acid	5.1	0.32	5.31	0.08
4-Feruleoylquinic acid	3.88	1	4.48	0.49
5-Feruleoylquinic acid	3.53	0.48	3.35	0.23

**Table S2.** Differential taxa among compliant participants post-intervention by prune treatment. Taxa presented are LDA>2. ‘Group’ represents the treatment group in which specific taxa was most abundant for that comparison.

id	LEfSe: Overall			LEfSe: Control vs. 50g			LEfSe: Control vs. 100g			LEfSe: 50g Prune vs. 100g		
	Group	LDA	p	Group	LDA	Group	LDA	Group	LDA	Group	LDA	Group
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales	Control	4.31	1.21E-02	Control	4.34					100g Prune	3.77	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Oscillospiraceae	Control	3.99	1.16E-03	Control	3.98					50g Prune	3.88	
d_Bacteria_p_Firmicutes_c_Bacilli	Control	3.94	1.28E-03							100g Prune	3.24	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Peptostreptococcales_Tissierellales	Control	3.86	1.13E-02	Control	3.88					100g Prune	3.17	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Peptostreptococcales-Tissierellales_f_Peptostreptococcaceae	Control	3.83	2.10E-02	Control	3.85					100g Prune	3.58	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Oscillospiraceae_g_UCG-002	Control	3.67	2.00E-02	Control	3.65					100g Prune	3.18	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Christensenellales	Control	3.64	4.31E-02							100g Prune	3.18	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Christensenellales_f_Christensenellaceae	Control	3.64	4.31E-02							100g Prune	3.18	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Oscillospiraceae_g_UCG-002	Control	3.60	2.61E-02	Control	3.58					100g Prune	3.41	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Peptostreptococcales-Tissierellales_f_Peptostreptococcaceae_g_Romboutsia	Control	3.56	4.90E-02	Control	3.56					100g Prune	2.99	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Peptostreptococcales-Tissierellales_f_Peptostreptococcaceae_g_Romboutsia_	Control	3.56	4.90E-02	Control	3.56					100g Prune	2.99	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Clostridiiales	Control	3.55	1.07E-02	Control	3.54							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Clostridiaceae	Control	3.55	1.07E-02	Control	3.54							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Clostridiaceae_f_Clostridiaceae_g_Clostridium_sensu_stricto_1	Control	3.49	5.89E-03	Control	3.46					Control	3.38	50g Prune 2.92
d_Bacteria_p_Firmicutes_c_Clostridia_o_Clostridiaceae_f_Clostridiaceae_g_Clostridium_sensu_stricto_1_	Control	3.39	2.25E-02							100g Prune	2.92	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Ruminococcaceae_g_Incertae_Sedis_	Control	3.26	9.81E-03	Control	3.21					Control	3.19	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Oscillospiraceae_g_NKA214_group	Control	3.25	1.52E-03	Control	3.24					100g Prune	2.91	
d_Bacteria_p_Firmicutes_c_Bacilli_o_Erysipelotrichales	Control	3.23	9.55E-03	Control	3.24							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Oscillospiraceae_g_UCG-005	Control	3.23	1.18E-02	Control	3.23							
d_Bacteria_p_Firmicutes_c_Bacilli_o_Erysipelotrichales_f_Erysipelotrichaceae	Control	3.23	2.08E-02	Control	3.22	Control	3.16					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Oscillospiraceae_g_Uncultured	Control	3.03	4.87E-03	Control	3.02	Control	3.00					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_[Ruminococcus]_torques_group_s_Ruminococcus_torques	Control	2.90	1.57E-02	Control	3.02	Control	2.79					
d_Bacteria_p_Verrucomicrobota_c_Verrucomicrobota_o_Verrucomicrobiales	Control	2.80	3.20E-02	Control	2.54							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Monoglobales_f_Monoglobales_e_g_Monogobus_	Control	2.79	2.74E-02			Control	2.69					
d_Bacteria_p_Verrucomicrobota_c_Verrucomicrobota_o_Verrucomicrobia	Control	2.76	3.20E-02	Control	2.75							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Eisenbergiella	Control	2.73	2.32E-02	Control	2.69							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Eisenbergiella_s_uncultured_organism	Control	2.68	1.42E-02	Control	2.54	Control	2.39					
d_Bacteria_p_Verrucomicrobota_c_Verrucomicrobota_o_Verrucomicrobiales_f_Akkermansiaceae	Control	2.66	8.33E-03	Control	2.53	Control	2.41					
d_Bacteria_p_Verrucomicrobota_c_Verrucomicrobota_o_Verrucomicrobiales_f_Akkermansiaceae_g_Akkermansia	Control	2.59	3.20E-02	Control	2.73							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Oscillospiraceae_g_UCG-005_s_gut_metagenome	Control	2.58	3.20E-02	Control	2.63							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Uncultured_g_Uncultured	Control	2.53	4.46E-02	Control	2.60							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Uncultured_g_Uncultured	Control	2.51	2.07E-02	Control	2.74							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Uncultured	Control	2.50	2.07E-02	Control	2.72							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Uncultured	Control	2.40	3.63E-02			Control	2.35					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Ruminococcaceae_g_Incertae_Sedis_s_Uncultured_bacterium	Control	2.34	2.86E-02	Control	2.33	Control	2.37					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Ruminococcaceae_g_UCB1819	Control	2.34	2.86E-02	Control	2.33	Control	2.37					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Ruminococcaceae_g_UCB1819_s_Uncultured_organism	Control	2.34	2.86E-02	Control	2.33	Control	2.37					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Oscillospiraceae_g_NKA214_group_s_Uncultured_rumen	Control	2.32	3.19E-02	Control	2.29					100g Prune	2.17	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Lachnospiraceae	50g Prune	4.57	5.71E-03	50g Prune	4.58					50g Prune	4.46	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospirales	50g Prune	4.57	5.52E-03	50g Prune	4.58					50g Prune	4.46	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Blaatia	50g Prune	4.38	7.49E-03	50g Prune	4.33					50g Prune	4.37	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Blaatia_	50g Prune	4.36	7.82E-03	50g Prune	4.34					50g Prune	4.36	
d_Bacteria_p_Firmicutes_c_Bacilli_o_Lactobacillales	50g Prune	3.93	2.77E-03			Control	3.86	50g Prune 3.90				
d_Bacteria_p_Firmicutes_c_Bacilli_o_Lactobacillales_f_Streptococcaceae_g_Streptococcus	50g Prune	3.91	2.36E-03			Control	3.85	50g Prune 3.88				
d_Bacteria_p_Firmicutes_c_Bacilli_o_Lactobacillales_f_Streptococcaceae_g_Streptococcus_	50g Prune	3.91	2.45E-03			Control	3.85	50g Prune 3.88				
d_Bacteria_p_Firmicutes_c_Bacilli_o_Lactobacillales_f_Streptococcaceae	50g Prune	3.91	4.16E-03			Control	3.85	50g Prune 3.88				
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_[Eubacterium]_hallii_group	50g Prune	3.68	4.34E-02	50g Prune	3.66							
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_[Eubacterium]_hallii_group_	50g Prune	3.68	4.34E-02	50g Prune	3.66							
d_Bacteria_p_Firmicutes_c_Bacilli_o_Lactobacillales_f_Carnobacteriaceae_g_Granulicatella_	50g Prune	3.00	2.25E-02							50g Prune	2.49	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Ruminococcaceae_g_Faecalibacterium_s_metagenome	50g Prune	2.90	8.73E-03	50g Prune	2.83	100g Prune	2.68					
d_Bacteria_p_Firmicutes_c_Bacilli_o_Lactobacillales_f_Carnobacteriaceae	50g Prune	2.87	2.25E-02							50g Prune	2.49	
d_Bacteria_p_Firmicutes_c_Bacilli_o_Lactobacillales_f_Carnobacteriaceae_g_Granulicatella	50g Prune	2.76	2.25E-02							50g Prune	2.60	
d_Bacteria_p_Actinobacteria_c_Actinobacteria_o_Actinomycetales	50g Prune	2.68	4.09E-02			Control	3.10	50g Prune 2.17				
d_Bacteria_p_Actinobacteria_c_Actinobacteria_o_Actinomycetales_f_Actinomycetaceae_g_Actinomyces	50g Prune	2.67	4.09E-02			Control	3.12	50g Prune 2.30				
d_Bacteria_p_Proteobacteria_c_Gammaproteobacteria_o_Burkholderiales_f_Suttermellaceae_g_Parasutterella_s_uncultured_organism	50g Prune	2.48	4.09E-02	50g Prune	2.31	100g Prune	2.32			50g Prune	2.26	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Lachnospiridium_s_glycyrhizinilyticum	50g Prune	2.11	1.75E-02									
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Aerobiceroides	100g Prune	4.27	1.04E-04	50g Prune	4.10	100g Prune	4.30					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Aerobiceroides_g_Aerobiceroides	100g Prune	4.27	1.15E-04	50g Prune	4.10	100g Prune	4.30					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Aerobiceroides_s_Aerobiceroides	100g Prune	3.66	3.82E-03	50g Prune	3.33	100g Prune	3.64					
d_Bacteria_p_Firmicutes_c_Clostridia_o_Coriobacteriales_f_Eggerthellaceae	100g Prune	3.58	3.30E-02	50g Prune	3.16	100g Prune	3.47					
d_Bacteria_p_Proteobacteria_c_Gammaproteobacteria_o_Enterobacteriales	100g Prune	3.58	3.30E-02	50g Prune	3.16	100g Prune	3.47					
d_Bacteria_p_Bacteroidota_c_Bacteroidia_o_Bacteroidales_f_Tannerellaceae_g_Parabacteroides_merdae	100g Prune	3.48	5.80E-03							100g Prune	3.43	
d_Bacteria_p_Bacteroidota_c_Bacteroidia_o_Bacteroidales_f_Tannerellaceae	100g Prune	3.46	9.42E-03	Control	3.25					100g Prune	3.41	
d_Bacteria_p_Bacteroidota_c_Bacteroidia_o_Bacteroidales_f_Tannerellaceae_g_Parabacteroides	100g Prune	3.46	9.42E-03	Control	3.25					100g Prune	3.41	
d_Bacteria_p_Bacteroidota_c_Bacteroidia_o_Bacteroidales_f_Bacteroidaceae_g_Bacteroides_o_vatus	100g Prune	3.22	2.64E-03							100g Prune	2.91	
d_Bacteria_p_Bacteroidota_c_Bacteroidia_o_Bacteroidales_f_Rikenellaceae_g_Alistipes_shahii	100g Prune	3.05	1.74E-02							100g Prune	2.92	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Coprococcus_s_unidentified	100g Prune	2.98	3.02E-02	Control	2.78					100g Prune	2.90	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Lachnospirales_s_uncultured_bacterium	100g Prune	2.87	1.08E-02							100g Prune	2.84	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Moreyella	100g Prune	2.56	2.07E-02							100g Prune	2.19	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Lachnospirales_f_Lachnospiraceae_g_Moreyella_s_human_gut	100g Prune	2.53	2.07E-02							100g Prune	2.12	
d_Bacteria_p_Firmicutes_c_Bacteroidota_c_Bacteroidia_o_Bacteroidales_f_Barnesiellaceae_g_Coprobacter	100g Prune	2.44	1.02E-02							100g Prune	2.42	
d_Bacteria_p_Desulfovibriota_c_Desulfovibrioia_o_Desulfovibrionales_f_Desulfovibrionaceae_g_Desulfovibrion_piger	100g Prune	2.37	1.25E-02							100g Prune	2.47	
d_Bacteria_p_Firmicutes_c_Clostridia_o_Oscillosporales_f_Ruminococcaceae_g_Ruminococcus_s_gut_metagenome	100g Prune	2.24	1.20E-02							100g Prune	2.37	

Figure S1. CONSORT flow diagram for this study. The number of subjects that participated at each step are depicted. Reasons for dropout or exclusion from analysis are summarized.

Figure S2. Alpha Rarefaction. Each curve represents one sample. This plot diagrams the number of species retained as a function of sequencing depth. Choosing a depth where most samples have plateaued will maximize the number of features retained and ensure diversity metrics reflect the actual diversity. A sequencing depth of 11,940 reads was chosen, which maximizes retained features minimizing excluded samples.

Figure S3. Significant correlations between log counts of taxa and phenolics using data both pre- and post-intervention. Data are for compliant and noncompliant subjects who had detectable metabolite. (A) 3-Hydroxyhippuric acid vs. *Oscillospiraceae* UCG-005 (Bonferroni-corrected  $p=0.02$ , Spearman's  $r=-0.33$ ), (B) 3-hydroxyhippuric acid vs. *Hydrogenoanaerobacterium* ( $p=0.08$ ,  $r=-0.31$ ), (C) Total urinary phenolics vs. *Blautia* metagenome ( $p=0.03$ ,  $r=-0.24$ ), (D) Total urinary phenolics vs. uncultured *Lachnospiraceae* ( $p=0.05$ ,  $r=0.24$ ). Blue lines are a linear model and a shaded 95% confidence interval. Note that for plots (A) and (B), an outlier with 800  $\mu\text{M}$  3-hydroxyhippuric acid and 0 counts of both taxa was removed from the figure to improve readability.

Figure S4. Correlation between *Lachnospiraceae* UCG-001 and (A) plasma IL-1B (Bonferroni-corrected  $p=0.002$ , Spearman's  $r=-0.29$ ) or (B) plasma IL-6 (Bonferroni-corrected  $p=0.09$ , Spearman's  $r=-0.25$ ). The blue line is a linear model and shaded indicates 95% confidence interval.  $n=308$ .