

Supplementary information

Supplementary Table S1. Composition of the experimental diets (% w/w)

| Ingredients | Control | EtOH | EtOH+L-RR | EtOH+M-RR | EtOH+H-RR |
|---------------|---------|------|-----------|-----------|-----------|
| AIN93M diet | 100 | 100 | 98 | 96 | 92 |
| Red raspberry | - | - | 2 | 4 | 8 |
| Total | 100 | 100 | 100 | 100 | 100 |

Supplementary Table S2. Estimated values of various nutrients of AIN93M standard feed

| Ingredients | AIN93M |
|-------------------------------------------------|--------------|
| Energy | 3601 Kcal/kg |
| Total calculated digestible energy from protein | 14.1% |
| Total calculated digestible energy from lipids | 10% |
| Water content | 6.8% |
| Fat | 4% |
| Carbohydrates | 72.7% |
| Protein | 12.5% |
| Ash | 3.89% |
| Amino acid content | |
| Alanine | 3.3g |
| Arginine | 4.5g |
| Aspartic acid | 8g |
| Glutamate | 25.5g |
| Glycine | 2.3g |
| Lysine | 9.2g |
| Methionine | 3.3g |
| Cystine | 2.4g |
| Tryptophan | 1.6g |

| | |
|---------------|-------|
| Proline | 14.3g |
| Serine | 6.7g |
| Histidine | 3.3g |
| Leucine | 10.9g |
| Isoleucine | 5.9g |
| Phenylalanine | 6.2g |
| Tyrosine | 6.6g |
| Threonine | 4.7g |
| Valine | 7g |

Mineral content

| | |
|------------------|---------|
| Calcium | 5000ppm |
| Phosphorus | 3000ppm |
| Potassium | 3600ppm |
| Sodium | 1033ppm |
| Magnesium | 511ppm |
| Iron | 45ppm |
| Zinc | 35ppm |
| Copper | 6ppm |
| Iodine | 0.2ppm |
| Chromium | 1ppm |
| Inorganic Sulfur | 300ppm |
| Chlorine | 1613ppm |

Vitamin content

| | |
|------------------|-----------|
| Vitamin A | 4IU/g |
| Vitamin D | 1IU/g |
| Vitamin E | 0.075IU/g |
| Vitamin K | 0.86 ppm |
| Thiamine, B1 | 5 ppm |
| Riboflavin | 6 ppm |
| Niacin | 30 ppm |
| Pantothenic acid | 15 ppm |
| Vitamin B6 | 6 ppm |
| Choline | 1000 ppm |
| Folic acid | 2ppm |
| Biotin | 0.2ppm |
| Vitamin B12 | 25ppb |

Product code: LAD3001M.

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Method for the determination of red raspberry polyphenols composition

Red raspberry extract was analyzed by high-performance liquid chromatography (HPLC) based on targeted and untargeted method. The analytical conditions were as follows: HPLC column, shim-pack GISS C18 (pore size 1.9 μm , length 2.1 \times 100 mm); the solvent system was solvent A (aqueous 0.04% acetic acid) and solvent B (acetonitrile (0.04% acetic acid)). The gradient program was 0 min, 5% B; 12.0 min, 95% B; 13.2 min, 95% B; 13.3 min, 5% B; 15.0 min, 5% B, with a flow rate of 0.4 mL/min. The temperature was 40 °C and the injection volume was 2 μL . Raspberry extract was performed in the Full-Scan mode by Q Exactive Focus Orbitrap LC-MS/MS (Thermo Scientific, USA) Using Compound Discoverer 3.3 software to analyze the raw data. The ESI source operation parameters were as follows: nebulizing gas flow, 3 L/min; heating gas flow, 10 L/min; interface temperature, 500 °C; DL temperature, 250 °C; heat block temperature, 400 °C; drying gas flow, 10 L/min.

Identification of compounds

The identification was carried out by comparison of the accurate m/z values, the retention time (RT), and the fragmentation patterns with those obtained from available standards analyzed in the same conditions. For high-quality (S/N>10) metabolic signals, we compared the MS2 spectral information of metabolic signals with the database by using Compound Discover 3.3 software and annotated these metabolic signals in batches. Then we identified metabolic signals that did not match the information in the database by querying the MS2 spectral data taken from the literature or to search the databases (e.g., METLIN, MassBank).

Supplementary Table S3. Primers used in this study

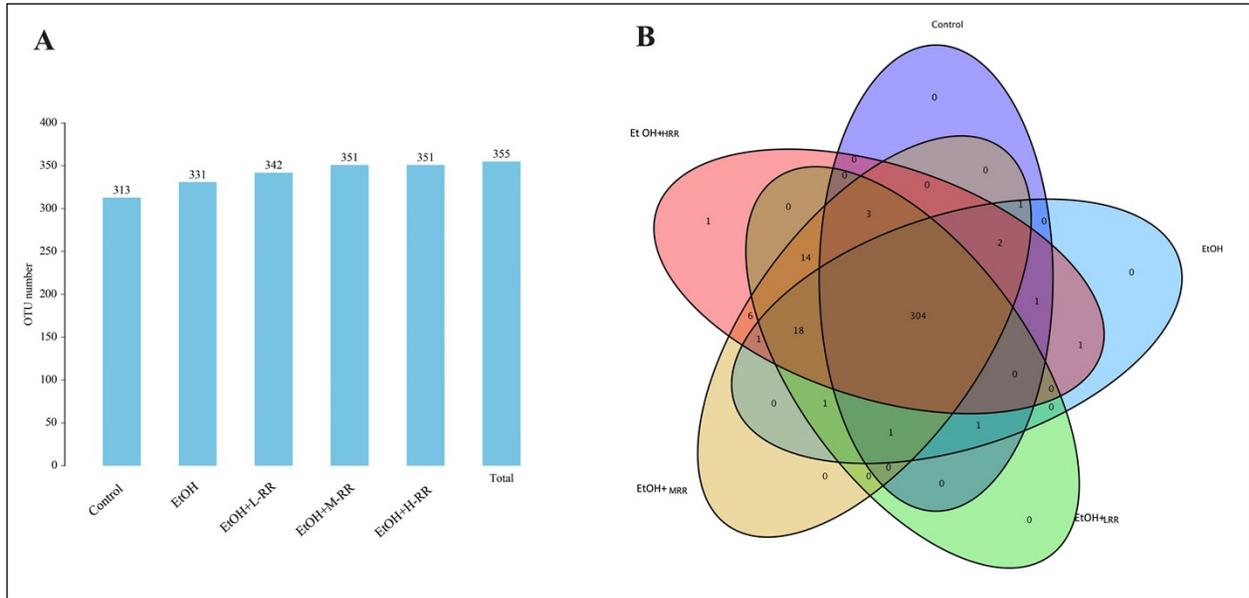
| Gene name | Primer sequence | Gene ID/Reference |
|---------------------------------|--------------------------------------------------------|-------------------|
| <i>ZO-1</i> | F: CGGAACTATGACCATCGCCTAC R: CTTCGGGATGTTGTCTGGAGTC | NM-001163574.1 |
| <i>Occludin</i> | F: TTCCACACTTGCTTGGGACAGA R: TCCGCCATAGCCATAGCCATAG | NM-0088756.2 |
| <i>Claudin-1</i> | F: GGGCTGATCGCAATCTTTGTGT R: CCACTAATGTCGCCAGACCTGA | NM-016674.4 |
| <i>Claudin-4</i> | F: GGCTGAGCGATGGCGTCTAT R: CGATGTTGCTGCCGATGAAGG | NM-009903.2 |
| <i>E-cadherin</i> | F: GCCATCGCCTACACCATCGT R: GCAGCCTGAACCACCAGAGT | NM-009864.3 |
| <i>CD14</i> | F: GAGCGTGTGCTTGGCTTGTT R: CCGTAAGCCGCTTTAAGGACA | 1 |
| <i>TLR4</i> | F: GCATGGATCAGAACTCAGCAAA R: CTCCACAGCCACCAGATTCTC | 1 |
| <i>NF-κB</i> | F: GCTGCCAAAGAAGGACACGACA R: GGCAGGCTATTGCTCATCACAG | NM_008689 |
| <i>CYP 2E1</i> | F: TGGGGAAACAGGGTAATGAG R: CTGGCCTTTGGTCTTTTTGA | 2 |
| <i>β-actin</i> | F: GGCTGTATTCCCTCCATCG R: CCAGTTGGTAACAATGCCATG | NM_007393.5 |

F, forward; R, reverse

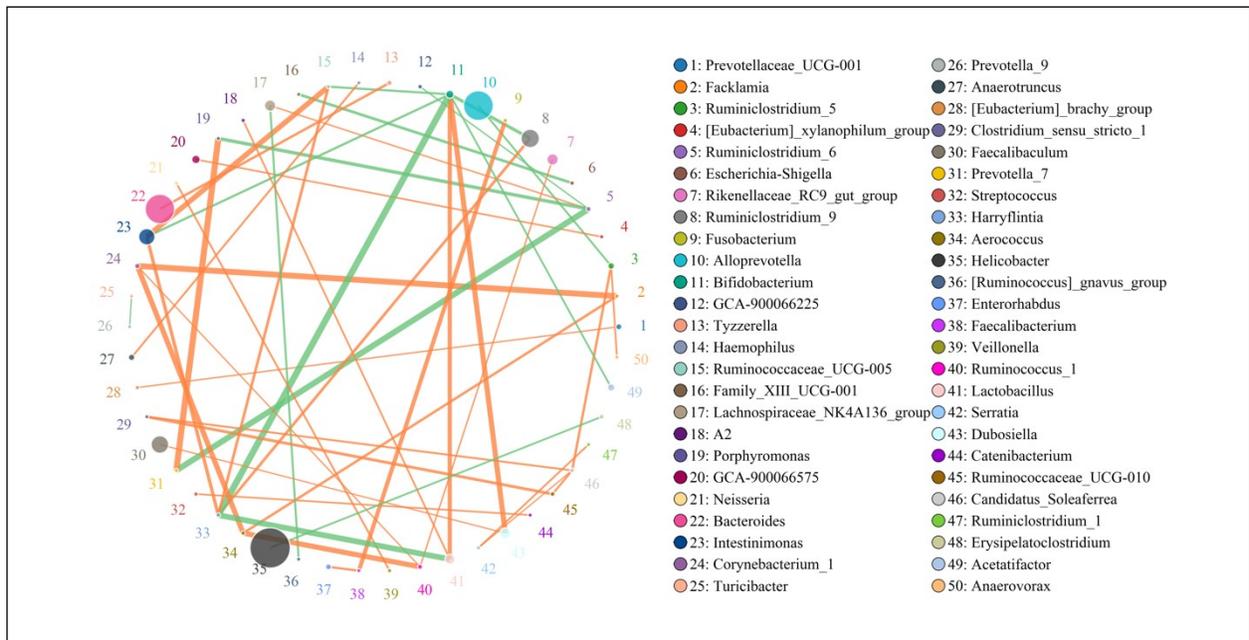
Primers were designed using BLAST of NCBI (USA) and synthesized by TSINGKE Biotech Co., Ltd. (Wuhan, China).

Supplementary Table S4. Antibodies and their dilution ratio used in this study

| Detection antibody | Company | Item No. | Dilution with TBST |
|----------------------------------------------------|--------------------------------------------------|-----------------|-------------------------------|
| Mouse monoclonal antibody β -actin (42KD) | Wuhan Boster Biological Engineering Co., Ltd. | BM0627 | 1:500 |
| Rabbit anti ZO-1 (240KD) | Boster | PB9234 | 1:1000 |
| Rabbit anti Occludin (59KD) | Affinity | DF7504 | 1:2000 |
| Mouse anti E-cadherin (120KD) | Wuhan Sanying Biotechnology Co., Ltd. | 60335-1-Ig | 1:3000 |
| Rabbit anti Claudin1(23KD) | Affiniy | AF0127 | 1:2000 |
| Rabbit anti Claudin4 (22KD) | Wuhan Sanying Biotechnology Co., Ltd. | 16195-1-AP | 1:1000 |
| Rabbit anti MLC2 (19KD) | Wuhan Sanying Biotechnology Co., Ltd. | 10906-1-AP | 1:2000 |
| Mouse anti p-MLC2 (18KD) | CST | 3675S | 1:1000 |
| Rabbit anti MLCK (211KD) | Affinity | AF5314 | 1:1000 |
| Rabbit anti NF κ B (65KD) | Invitrogen | PA5-17264 | 1:1000 |
| Rabbit anti p-NF κ B (60KD) | Affinity | AF2006 | 1:1000 |
| HRP-labeled goat anti-mouse secondary antibody | Boster | BA1051 | 1:10000 |
| HRP-labeled goat anti-rabbit secondary antibody | Boster | BA1054 | 1:10000 |



Supplementary Fig. 1. (A) Operational taxonomic unit (OTU) number in cecal samples of mice in the five groups after six weeks of treatment. (B) Venn diagram showing microbial common and unique features among mice from the five experimental groups.



Supplementary Fig. 2. Cecal content species network at genus level of mice in the five experimental groups.

Circles represent species and the size of the circle corresponds to the abundance; The correlation between two species is represented by the edges, the thickness of the edge represents the strength of the correlation, and the color of the line: orange represents the positive correlation, while green represents green the negative correlation.

References:

- 1 J. Xiao, R. Zhang, Y. Wu, C. Wu, X. Jia, L. Dong, L. Liu, Y. Chen, Y. Bai and M. Zhang, Rice Bran Phenolic Extract Protects against Alcoholic Liver Injury in Mice by Alleviating Intestinal Microbiota Dysbiosis, Barrier Dysfunction, and Liver Inflammation Mediated by the Endotoxin-TLR4-NF- κ B Pathway, *J. Agric. Food Chem.*, 2020, **68**, 1237–1247.
- 2 Y. Ren, Y. Ding, F. Meng, L. Jiang, H. Li, J. Huang, P. Yu and Z. Qiu, Quantification of CYP2E1 in rat liver by UPLC-MS/MS-based targeted proteomics assay: a novel approach for enzyme activity assessment, *Anal. Bioanal. Chem.*, 2020, **412**, 5409–5418.