Trans palmitoleic acid, a dairy fat biomarker, stimulates insulin secretion and activates G protein-coupled receptors with a different mechanism than *cis* isomer

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26 Supplemental Figure 1. Structures of GPR40 (A), GPR55 (B), GPR119 (C) and GPR120 (D) in 1-

palmitoyl-2-oleoyl-sn-glycero-3-phosphatidylcholine/1-palmitoyl-2-oleoyl-sn-glycero-3-

- phosphatidylglycerol/cholesterol membrane.



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Supplemental Figure 2. Acute and long-term effect of cis and trans isomers of palmitoleic on GSIS. MIN6 cells were treated with cPOA and tPOA (25 μM) for 1,5 h (A) and 24 h (B). EndoC-βH1 cells were treated with cPOA and tPOA (10 μ M) for 1,5 h (C) and 24 h (D). The bars represent the means \pm SEM from at least 3 independent experiments. ****p < 0.0001, ***p < 0.00, 1*p<0.05 vs 2 mM glucose

control; &&&&p < 0.0001, &&&p < 0.001, &&p < 0.01 vs 20 mM glucose control.





Simulation

Simulation



293 molecular dynamics simulations

²⁹² Supplemental Figure 7. Root mean square deviation of protein Cα atoms of each receptor throughout



Supplemental Figure 8. Density of systems during equilibration procedure for thermodynamic 296 integration



 $\begin{array}{c} 300\\ 301 \end{array}$ Supplemental Figure 9. System energy changes during cPOA-tPOA transformation. Total $\Delta\Delta G$ is 302 obtained through trapezoid integration of the area under each curve, followed by calculating the 303 difference between the complex state (complexed ligands, orange line) and protein-free state (free 304 ligands, blue line). Each point represents the value from three repetitions and standard deviation. As one 305 may notice, in each case, the energy of both systems was almost identical when the presence of tPOA 306 was slightly higher than cPOA. (A) depicts GPR40, (B) GPR55, (C) GPR119 and (D) GPR120 systems. 307