

**Supporting information**

## **Cobalt-catalysed CH-alkylation of indoles with alcohols by borrowing hydrogen methodology**

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## **S1. Materials and methods**

All indoles and alcohols were obtained commercially from various chemical companies. Unless otherwise stated all reagents were used directly without purification. Cobalt(II) nitrate hexahydrate (cat no. 239267-100G; ≥98%), zinc nitrate hexahydrate (cat no. 228737-500G; 98%) and silica suspension (Silica LUDOX® HS-40 colloidal silica, cat no. 420816-1L; 40 wt. % suspension in H<sub>2</sub>O) were purchased from Sigma Aldrich. 2,6-Diaminopyridine (cat no. A12295 -100G) was purchased from Alfa Aesar. The pyrolysis experiments were carried out in Dekema Austromat 624 oven.

XRD powder pattern were recorded on a Panalytical X'Pert diffractometer equipped with a Xcelerator detector using automatic divergence slits and Cu  $\text{K}\alpha_1/\alpha_2$  radiation (40 kV, 40 mA;  $\lambda = 0.15406 \text{ nm}$ ,  $0.154443 \text{ nm}$ ). Cu beta-radiation was excluded using a nickel filter foil. The measurements were performed in  $0.0167^\circ$  steps and 100 s of data collecting time per step. The samples were mounted on silicon zero background holders. The obtained intensities were converted from automatic to fixed divergence slits ( $0.25^\circ$ ) for further analysis. Peak positions and profile were fitted with Pseudo-Voigt function using the High Score Plus software package (Panalytical). Phase identification was done by using the PDF-2 database of the International Center of Diffraction Data (ICDD).

Scanning Transmission Electron Microscopy (STEM) was performed with a probe aberration corrected JEM-ARM 200F (JEOL) equipped high angle annular dark field (HAADF) and annular bright field (ABF) detectors and energy dispersive x-ray spectroscopy (EDXS), Dry60SGD (JEOL), for chemical analysis. Attached to the microscope is an electron energy loss spectrometer (EELS), Enfinium ER (Gatan). The specimen was dry deposited onto a Cu grid (mesh 300) covered by a holey carbon film.

The XPS (X-ray Photoelectron Spectroscopy) measurements were performed on an ESCALAB 220iXL (Thermo Fisher Scientific) with monochromated Al K $\alpha$  radiation ( $E = 1486.6 \text{ eV}$ ). Samples are prepared on a stainless-steel holder with conductive double-sided adhesive carbon tape. The electron binding energies were obtained without charge compensation leading to a binding energy of 284.7 eV of the main C 1s core level. For quantitative analysis the peaks were deconvoluted with Gaussian-Lorentzian curves using the software Unifit 2021. The peak areas were normalized by the transmission function of the spectrometer and the element specific sensitivity factor of Scofield.

GC and GC-MS analysis were performed on Agilent 6890N instrument. GC conversion and yields were determined by GC-FID, HP6890 chromatograph with FID detector, column HP530 m x 250 mm x 0.25  $\mu\text{m}$ . NMR spectra were recorded using Bruker 300 Fourier, Bruker AV 300 and Bruker AV 400 spectrometers. Chemical shifts are reported in ppm relative to the deuterated solvent. Coupling constants are expressed in Hertz (Hz). The following abbreviations are used: s = singlet, d = doublet, t = triplet and m = multiple. The residual solvent signals were used as references for <sup>1</sup>H and <sup>13</sup>C NMR spectra (CDCl<sub>3</sub>:  $\delta\text{H} = 7.26 \text{ ppm}$ ,  $\delta\text{C} = 77.12 \text{ ppm}$ ; DMSO-d<sub>6</sub>:  $\delta\text{H} = 2.50 \text{ ppm}$ ,  $\delta\text{C} = 39.52 \text{ ppm}$ ).

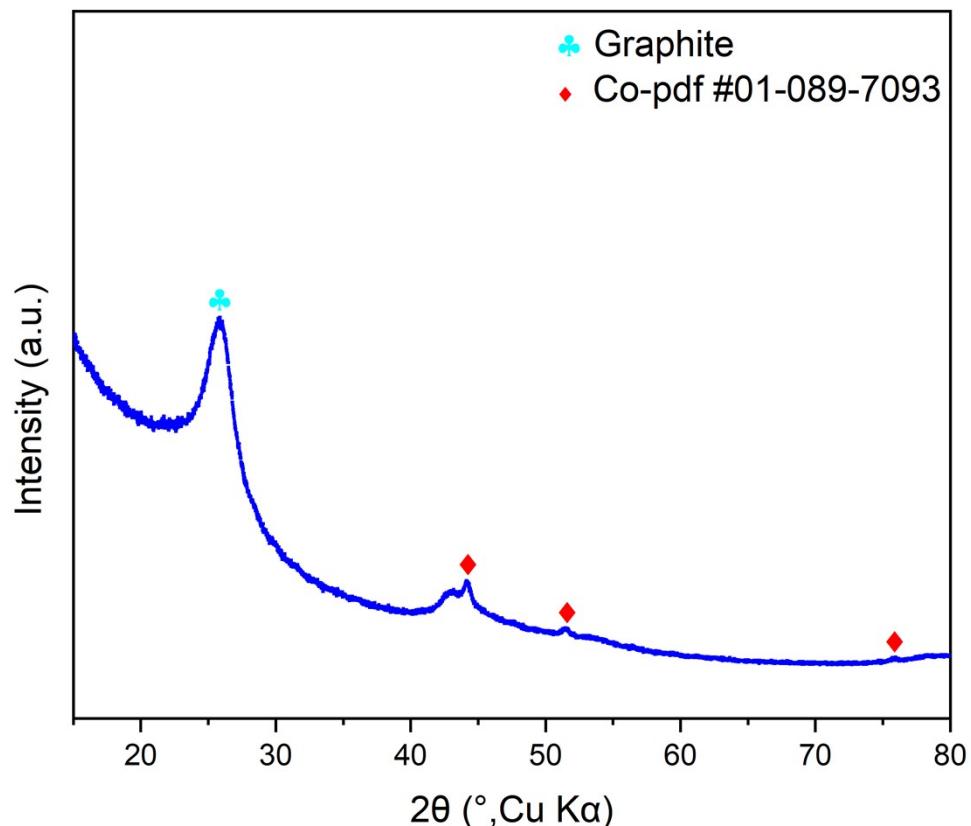
All catalytic experiments were carried out in ACS pressure tubes.

## **S2 Procedure for the preparation of catalysts**

In a 100 mL oven-dried single-necked round-bottom flask, 1.20 g colloidal silica aqueous solution (Ludox HS-40, 40 wt. % suspension in H<sub>2</sub>O) was dissolved in 30 mL deionized water and stirred for 30 minutes at room temperature. To the above solution, Co(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O (291.03 mg, 2,6-diaminopyridine (DAP; 1636.95 mg) and Zn(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O ( 297.49 mg) were added and continued stirring for 20 hours at room temperature. Then, the solvent of the reaction mixture was removed by freeze dryer. The obtained solid material was ground to a fine powder, transferred to a ceramic crucible, and placed in a pyrosis oven. Afterward, the furnace was heated to the defined temperature (700 °C, 800 °C, and 900 °C) for 3 h at the heating rate of 5 °C/min under argon atmosphere. After the pyrolysis, the oven was cooled down to room temperature and the material was removed from the oven. Next, the obtained samples were etched in 5 M NH<sub>4</sub>HF<sub>2</sub> aqueous solution at RT for 24 h to remove the SiO<sub>2</sub> template and larger particles. Finally, the resulting catalytic material was filtered and washed subsequently with deionized water and ethanol for three times and finally dried under vacuum overnight.

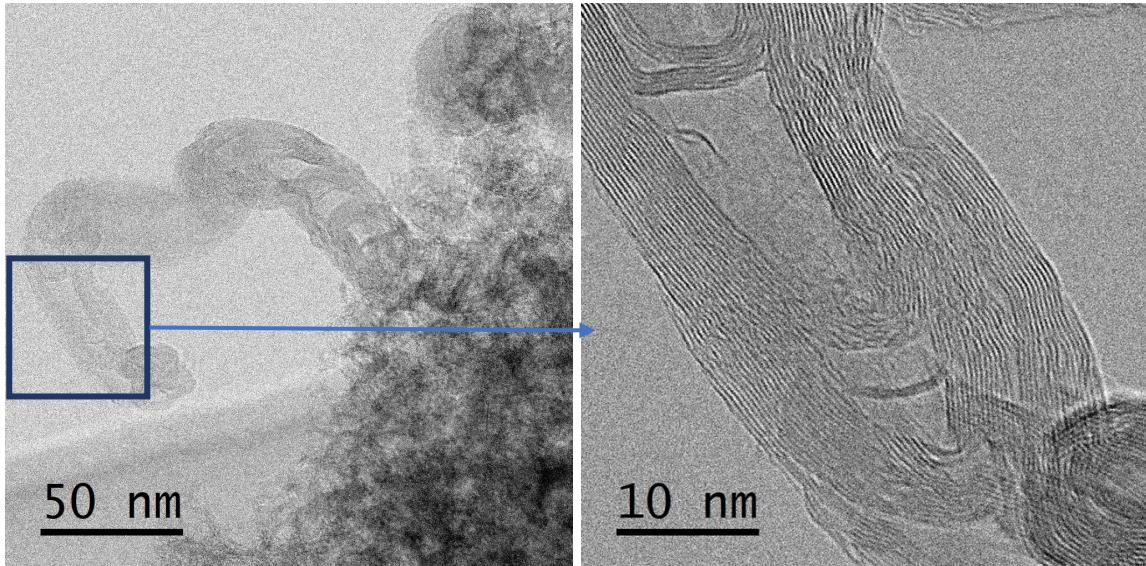
## **S3 Characterization of catalysts**

### XRD patterns

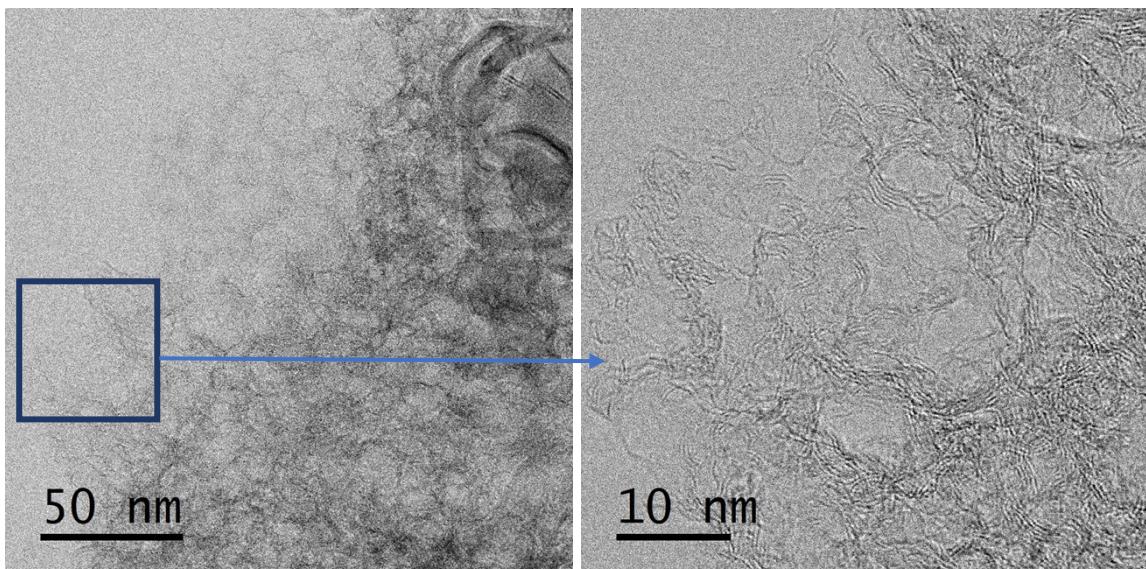


**Fig. S1.** XRD pattern of Co@NC-900 catalyst.

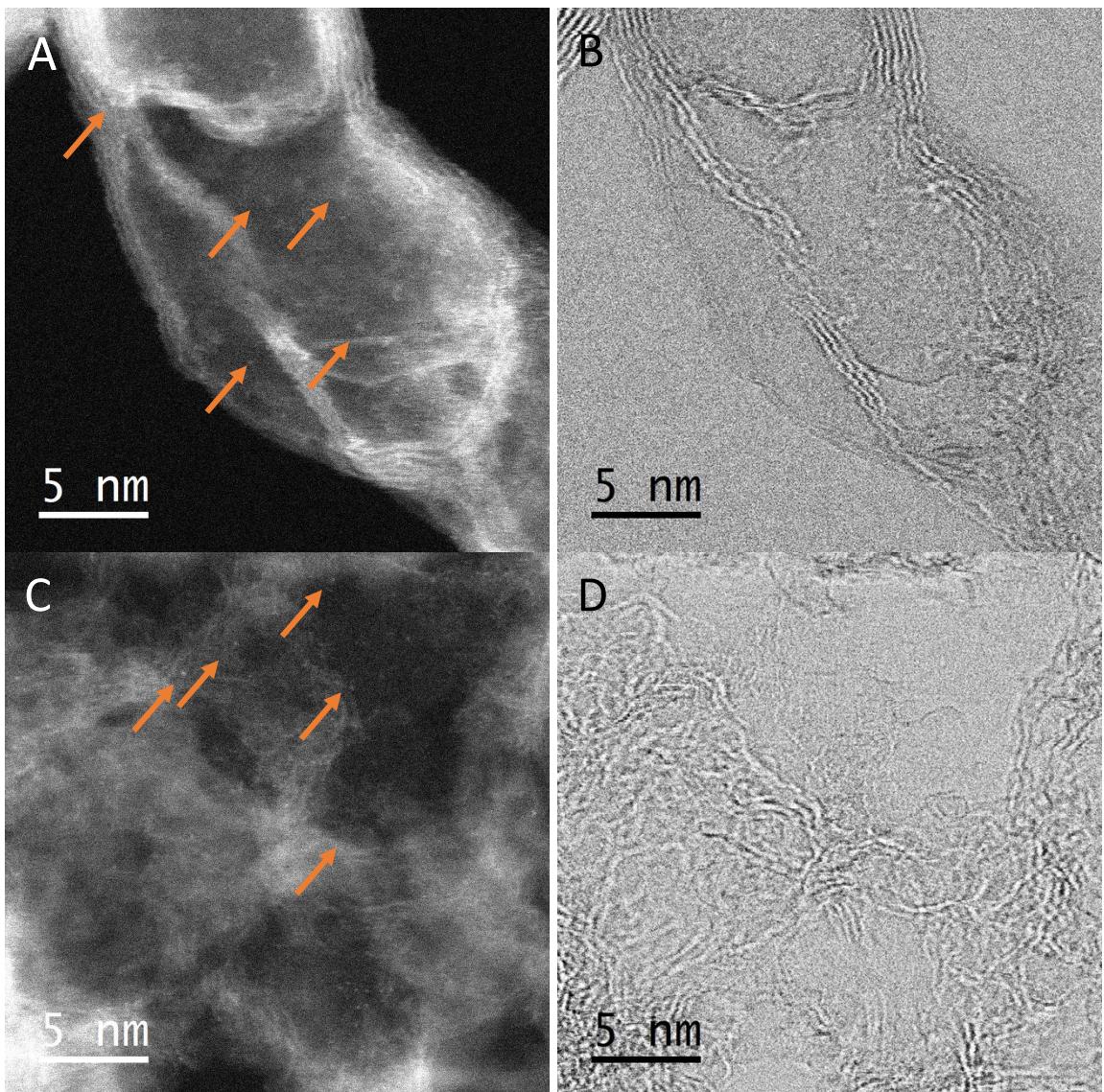
STEM images



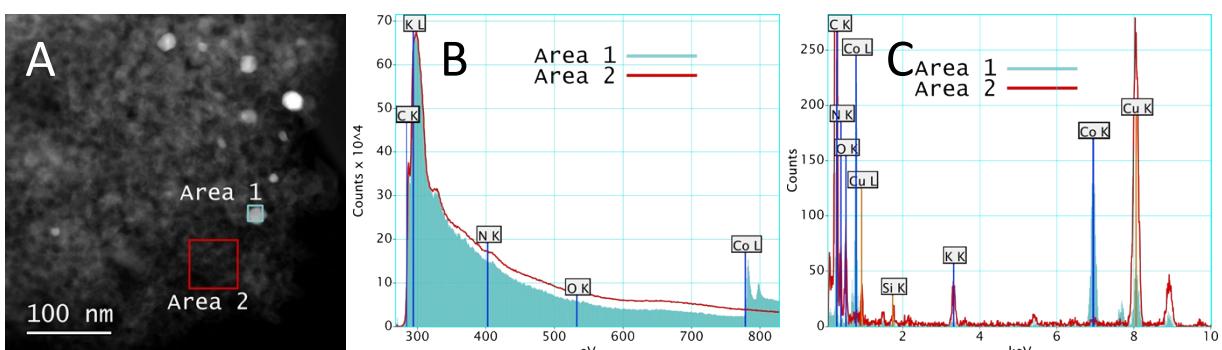
**Fig. S2.** ABF-STEM images of Co@NC-900 catalyst showing highly ordered graphitic carbon in the form of a carbon nano tube attached to a disordered carbon phase.



**Fig. S3.** ABF-STEM images of Co@NC-900 catalyst showing the disordered carbon phase in more detail.



**Fig. S4.** HAADF-STEM images (A and C) of ordered carbon (A) and disordered carbon (C) with bright dots (some marked by arrows) potentially representing K atoms remaining on the surface and corresponding ABF-STEM images (B and D) highlighting the carbon structure.



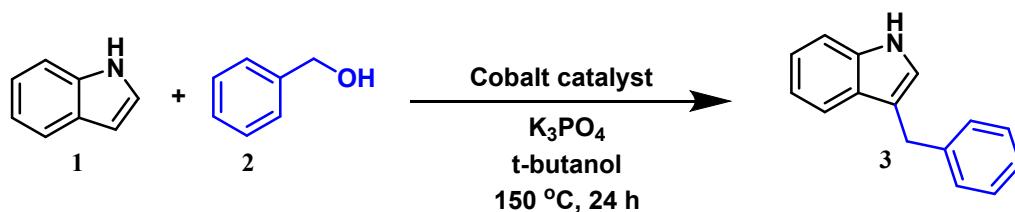
**Fig. S5.** ADF-STEM image (A) marked with reference frames from where the electron energy loss spectra (B) and the energy dispersive x-ray spectra (C) have been extracted from the spectrum image dataset. EDXS nicely shows the presence of K in areas where there are no Co particles. As there is also no Co signal of potentially dispersed Co atoms in area 2 this indicates the bright dots in S4 might stem from dispersed K atoms. EELS also indicates the metallic state of the Co particle in area 1 by the almost complete absence of an oxygen signal.

## **S4. General procedure for the C3-alkylation of indoles with alcohols**

### **S4.1 C3-Alkylation of indoles with alcohols**

In a 20 mL pressure tube fitted with magnetic stirring bar, 0.5 mmol of indole and 1 mmol alcohol, 50 mg catalyst (Co@NC-900) and 0.5 mmol K<sub>3</sub>PO<sub>4</sub> were added. Then, 2 mL tert-butanol was added, and the pressure tube was flushed with argon 3 times and fitted with screw cap. The pressure tube containing reaction mixture was placed in aluminum block and reaction was carried out under stirred condition at 150 °C for desired time. After the completion of the reaction, the pressure tube was cooled to room temperature. Then, the samples were removed from pressure tube, and the solid catalyst was filtered off and washed thoroughly with ethyl acetate. The reaction products were analyzed by GC-MS. The corresponding C3-alkylated indoles were purified by column chromatography (silica; pentene-ethyl acetate mixture) and characterized by NMR spectral analysis. Following procedure is applied for determining the conversions and yields by GC: after completion of the reaction, n-hexadecane (50 µL) as standard was added to the reaction pressure tube and the reaction products were diluted with ethyl acetate followed by filtration using plug of silica and then analyzed by GC.

**Table S1.** C3-Alkylation of indole with benzyl alcohol: Evaluation of Co catalysts. <sup>[a]</sup>

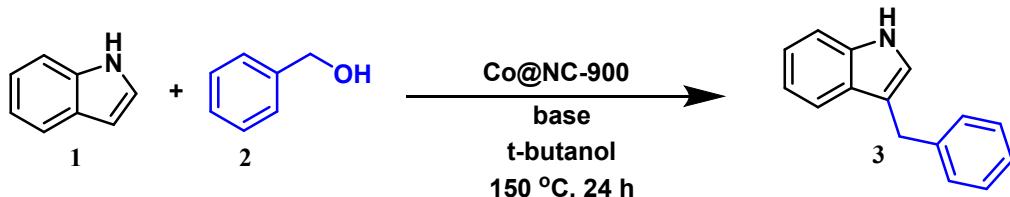


Entry	Catalyst	Conversion of 1 (%)	Yield of 3 (%)
1	Co@NC-SiO <sub>2</sub> -900	40	39
2	Co@NC-900	>99	97
3	Co@NC-700	80	77
4	Co@NC-800	88	85
5	Co@NC-900 (Prepared without Zn(NO <sub>3</sub> ) <sub>2</sub> )	74	70
6	Co-NPs <sup>[b]</sup>	59	56
7	Co(NO <sub>3</sub> ) <sub>2</sub> -Zn(NO <sub>3</sub> ) <sub>2</sub> -SiO <sub>2</sub>	<2	<1
8	Co(NO <sub>3</sub> ) <sub>2</sub> -Zn(NO <sub>3</sub> ) <sub>2</sub> -DAP	<2	<1
9	Co(NO <sub>3</sub> ) <sub>2</sub> -DAP	<2	<1
10	Without catalyst	<2	<1
11 <sup>c</sup>	Co@NC-900	60	56
12 <sup>d</sup>	Co@NC-900	91	90

13	NC-900	<2	<1
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[<sup>a</sup>] Reaction conditions: 0.5 mmol indole, 1 mmol benzyl alcohol, 50 mg catalyst (1.26 mol% Co), 0.5 mmol K<sub>3</sub>PO<sub>4</sub> (1 equiv.), 2 mL t-butanol, 150 °C, 24 h, conversions and yields are based on indole and determined by GC using n-hexadecane standard. [<sup>b</sup>] Co-NPs were prepared by the pyrolysis of Co(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O, Zn(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O and colloidal silica at 900 °C and then silica was removed. [<sup>c</sup>] 130 °C. [<sup>d</sup>] 140 °C.

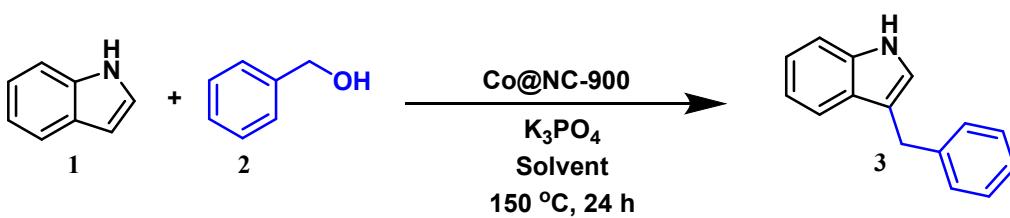
**Table S2.** C3-Alkylation of indole with benzyl alcohol: Testing of different bases.



Entry	Base	Conv. (%)	Yield 3 (%)
1	K <sub>2</sub> CO <sub>3</sub> (1 equiv.)	62%	55%
2	KOH (1 equiv.)	90%	87%
3	t-BuOK (1 equiv.)	>99%	96%
4	K <sub>3</sub> PO <sub>4</sub> (1 equiv.)	>99%	94%
5	K <sub>3</sub> PO <sub>4</sub> (0.7 equiv.)	71%	67%
6	K <sub>3</sub> PO <sub>4</sub> (0.5 equiv.)	49%	38%
7	K <sub>3</sub> PO <sub>4</sub> (1.5 equiv.)	>99%	95%

Reaction conditions: 0.5 mmol indole, 1 mmol benzyl alcohol, 50 mg catalyst (1.26 mol% Co), 2 mL t-butanol, 150 °C, 24 h. Conversions and yields are based on indole and determined by GC using n-hexadecane standard.

**Table S3.** C3-Alkylation of indole with benzyl alcohol: Testing of different solvents.

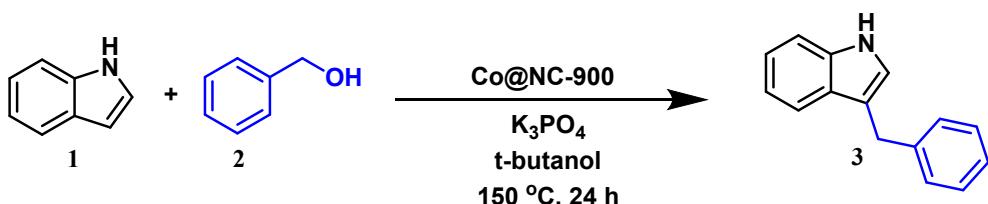


Entry	Solvent	Conv. (%)	Yield 3 (%)
1	THF	65%	61%
2	toluene	84%	80%
3	dioxane	73%	70%
4	t-butanol	>99%	94%

5	isopropanol	85%	82%
6	tert-amyl alcohol	86%	82%

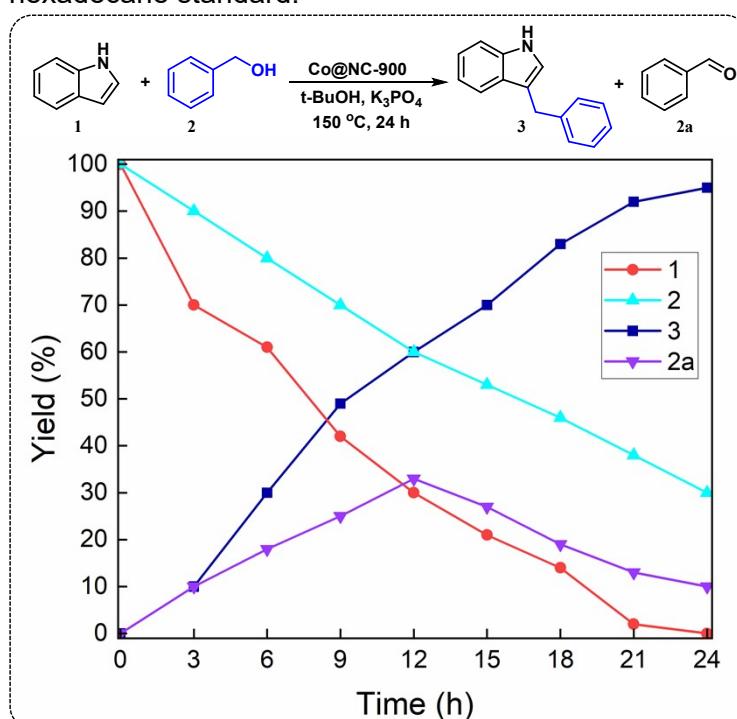
Reaction conditions: 0.5 mmol indole, 1 mmol benzyl alcohol, 50 mg catalyst (1.26 mol% Co), 2 mL t-butanol, 150 °C, 24 h. Conversions and yields are based on indole and determined by GC using n-hexadecane standard.

**Table S4.** C3-Alkylation of indole with benzyl alcohol: Testing of different alcohol amount.



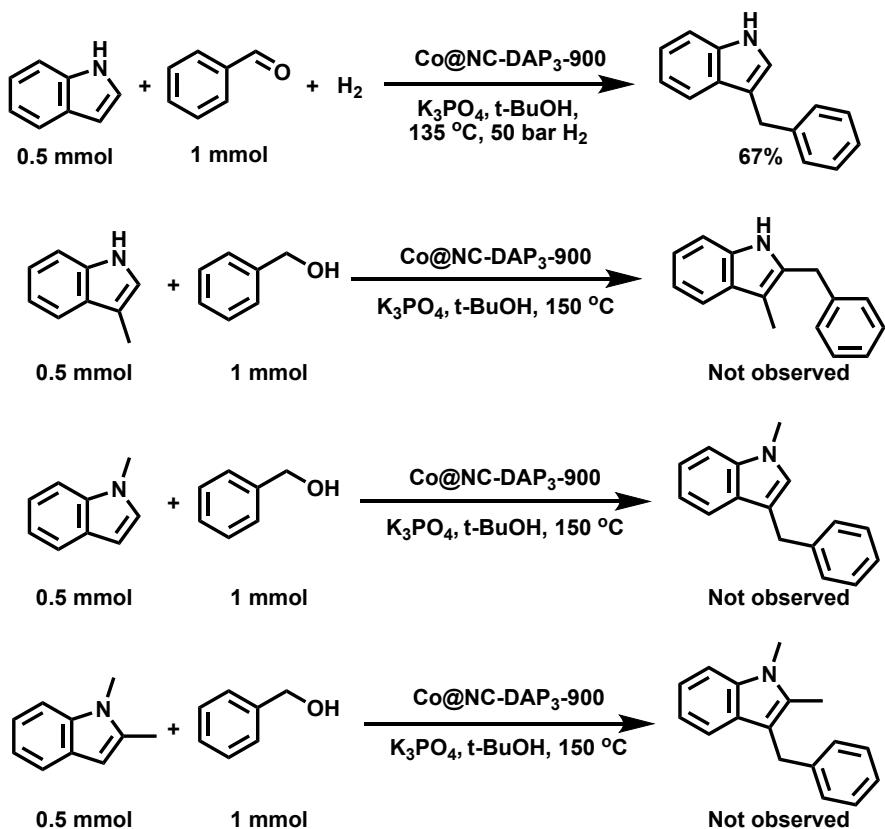
Entry	Alcohol amount	Conv. (%)	Yield 3 (%)
1	0.5 mmol (1 equiv.)	71%	68%
2	0.75 mmol (1.5 equiv.)	80%	77%
3	1 mmol (2 equiv.)	>99%	94%

Reaction conditions: 0.5 mmol indole, 1 mmol benzyl alcohol, 50 mg catalyst (1.26 mol% Co), 2 mL t-butanol, 150 °C, 24 h. Conversions and yields are based on indole and determined by GC using n-hexadecane standard.



**Fig. S6. C3-alkylation of indole with benzyl alcohol: reaction progress with time.**

Reaction conditions: 0.5 mmol indole, 1 mmol benzyl alcohol, 50 mg catalyst (1.26 mol% Co), 2 mL t-butanol, 150 °C, 24 h. Conversions and yields are based on indole and determined by GC using n-hexadecane standard.



**Scheme S1.** Control reactions.

#### S4.2 Methylation of indoles

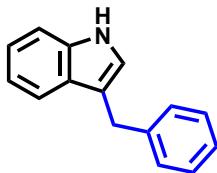
The magnetic stirring bar, 0.5 mmol indole, 70 mg Co@NC-900 and 1 mmol t-BuOK were transferred to 20 mL pressure tube. Then 1 mL methanol and 1 mL t-butanol were added to the above mixture. Then, the pressure tube was flushed with argon for 3 times and closed with screw cap. The pressure tube containing reaction mixtures were placed into aluminum block and allowed to progress at 160 °C for desired time. After the completion of the reaction, the pressure tube was cooled to room temperature. Then, the samples were removed from pressure tube, and the solid catalyst was filtered off and washed thoroughly with ethyl acetate. The reaction products were analyzed by GC-MS. The corresponding C3-alkylated products were purified by column chromatography (silica; pentene-ethyl acetate mixture) and characterized by NMR and GC-MS analysis.

#### S5 Catalyst recycling

The magnetic stirring bar, 1 mmol indole and 2 mmol benzyl alcohol, 100 mg Co@NC-900 and 1 mmol K<sub>3</sub>PO<sub>4</sub> were transferred to 20 mL pressure tube and 3 mL tert-butanol was added. The pressure tube was flushed with argon for 3 times and closed with screw cap. Then, it was placed into an aluminum block and heated to 150 °C for 24 h. After the completion of the reaction, the pressure tube was cooled down to room temperature. To the reaction products, 100 µL n-hexadecane as standard was added. The catalyst was then separated by centrifugation and the centrifugate containing reaction products was subjected to GC analysis. The separated catalyst was

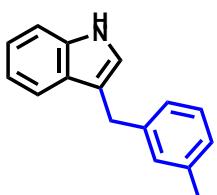
then washed with water, methanol, and ethyl acetate, dried under vacuum and used without further purification or reactivation for the next run.

### S6 NMR data



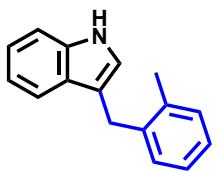
$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  7.92 (bs, 1H), 7.54 (dq,  $J$  = 7.8, 0.9 Hz, 1H), 7.41 – 7.16 (m, 7H), 7.10 (ddd,  $J$  = 8.0, 7.0, 1.1 Hz, 1H), 6.92 (dt,  $J$  = 2.2, 1.0 Hz, 1H), 4.14 (s, 2H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  141.23, 136.47, 128.72, 128.36, 127.49, 125.90, 122.35, 122.07, 119.39, 119.19, 115.87, 111.09, 31.62.



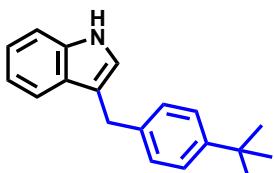
$^1\text{H}$  NMR (300 MHz, Chloroform-*d*)  $\delta$  7.64 (bs, 1H), 7.41 (dd,  $J$  = 7.8, 1.1 Hz, 1H), 7.17 (dt,  $J$  = 8.1, 1.0 Hz, 1H), 7.08 – 6.94 (m, 5H), 6.88 (d,  $J$  = 7.4 Hz, 1H), 6.70 (dt,  $J$  = 2.2, 1.0 Hz, 1H), 3.94 (s, 2H), 2.17 (s, 3H).

$^{13}\text{C}$  NMR (75 MHz, Chloroform-*d*)  $\delta$  141.27, 137.96, 136.49, 129.57, 128.32, 126.74, 125.83, 122.43, 122.07, 122.04, 119.41, 119.24, 115.94, 111.16, 111.13, 102.62, 31.59, 21.53.



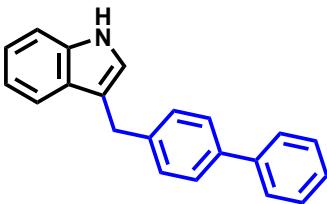
$^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.90 (bs, 1H), 7.67 – 7.61 (m, 1H), 7.41 (dt,  $J$  = 8.1, 0.9 Hz, 1H), 7.28 – 7.17 (m, 6H), 6.76 (dt,  $J$  = 2.2, 1.1 Hz, 1H), 4.14 (d,  $J$  = 1.1 Hz, 2H), 2.40 (s, 3H).

$^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  139.13, 136.52, 136.48, 130.17, 129.45, 127.59, 126.23, 125.99, 122.45, 122.08, 119.37, 119.10, 115.20, 111.13, 29.29, 19.56.

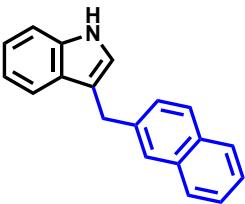


$^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.77 (bs, 1H), 7.52 – 7.47 (m, 1H), 7.29 – 7.21 (m, 3H), 7.18 – 7.13 (m, 2H), 7.12 – 7.09 (m, 1H), 7.02 (ddd,  $J$  = 8.0, 7.0, 1.1 Hz, 1H), 6.82 (dt,  $J$  = 2.0, 0.9 Hz, 1H), 4.02 (d,  $J$  = 0.9 Hz, 2H), 1.24 (s, 9H).

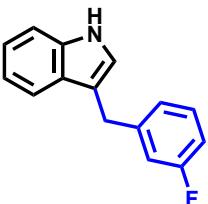
$^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  148.67, 138.23, 136.47, 128.34, 127.59, 125.27, 122.35, 122.04, 119.36, 119.24, 116.03, 111.11, 34.41, 31.49, 31.03.



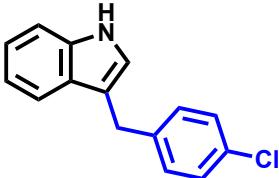
<sup>1</sup>H NMR (300 MHz, DMSO) δ 10.87 (s, 1H), 7.58 (dd, *J* = 18.7, 7.5 Hz, 4H), 7.50 – 7.27 (m, 7H), 7.21 (d, *J* = 2.1 Hz, 1H), 7.10 – 7.00 (m, 1H), 6.94 (t, *J* = 7.4 Hz, 1H), 4.08 (s, 2H).  
<sup>13</sup>C NMR (75 MHz, DMSO) δ 141.64, 140.58, 138.03, 136.88, 129.43, 129.33, 127.58, 127.44, 126.97, 126.94, 123.65, 121.41, 119.01, 118.76, 114.15, 111.87, 31.11.



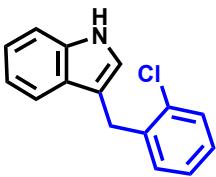
<sup>1</sup>H NMR (400 MHz, DMSO-d6) δ 10.91 (s, 1H), 7.86 – 7.77 (m, 4H), 7.50 – 7.34 (m, 5H), 7.23 (d, *J* = 2.4 Hz, 1H), 7.06 (ddd, *J* = 8.1, 6.9, 1.2 Hz, 1H), 6.93 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 4.22 (s, 2H).  
<sup>13</sup>C NMR (101 MHz, DMSO-d6) δ 139.94, 136.94, 133.63, 132.03, 128.09, 128.05, 127.93, 127.79, 127.52, 126.54, 126.41, 125.63, 123.78, 121.44, 119.02, 118.78, 114.13, 111.91, 31.79.



<sup>1</sup>H NMR (300 MHz, Chloroform-d) δ 7.87 (bs, 1H), 7.46 (ddt, *J* = 7.9, 1.3, 0.7 Hz, 1H), 7.31 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.21 – 7.13 (m, 2H), 7.10 – 6.99 (m, 2H), 6.99 – 6.77 (m, 3H), 4.07 (t, *J* = 0.7 Hz, 2H).  
<sup>13</sup>C NMR (75 MHz, Chloroform-d) δ 163.03 (d, *J* = 245.1 Hz), 143.95 (d, *J* = 6.9 Hz), 136.48, 129.72 (d, *J* = 8.3 Hz), 127.33, 124.32 (d, *J* = 2.8 Hz), 122.47, 122.22, 119.53, 119.09, 115.68, 115.40, 114.98, 112.83 (d, *J* = 21.1 Hz), 111.20, 31.39 (d, *J* = 1.7 Hz).  
<sup>19</sup>F NMR (376 MHz, Chloroform-d) δ -113.76 (td, *J* = 9.1, 5.7 Hz).

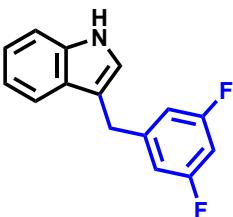


<sup>1</sup>H NMR (300 MHz, Chloroform-d) δ 7.83 (s, 1H), 7.36 (ddt, *J* = 7.9, 1.4, 0.8 Hz, 1H), 7.14 – 7.05 (m, 5H), 6.97 (ddd, *J* = 8.0, 7.0, 1.1 Hz, 1H), 6.79 (dt, *J* = 2.2, 1.0 Hz, 1H), 3.99 – 3.92 (m, 2H).  
<sup>13</sup>C NMR (75 MHz, Chloroform-d) δ 139.70, 136.48, 131.61, 130.03, 128.72, 128.44, 127.28, 122.38, 122.22, 119.50, 119.07, 115.28, 111.17, 31.02.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.97 (bs, 1H), 7.57 (dq, *J* = 7.9, 0.9 Hz, 1H), 7.42 – 7.35 (m, 2H), 7.24 – 7.19 (m, 2H), 7.17 – 7.09 (m, 3H), 6.95 (dt, *J* = 2.1, 1.0 Hz, 1H), 4.23 (t, *J* = 0.8 Hz, 2H).

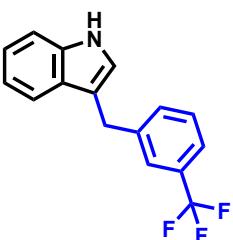
<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 138.75, 136.39, 134.02, 130.58, 129.37, 127.43, 127.38, 126.75, 122.76, 122.13, 119.49, 119.13, 114.02, 111.15, 29.09.



<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.02 (bs, 1H), 7.47 (ddt, *J* = 7.9, 1.4, 0.8 Hz, 1H), 7.38 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.22 (ddd, *J* = 8.2, 7.1, 1.3 Hz, 1H), 7.10 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 6.99 (dd, *J* = 2.3, 1.1 Hz, 1H), 6.90 – 6.70 (m, 2H), 6.63 (tdd, *J* = 9.0, 2.6, 2.1 Hz, 1H), 4.11 – 4.09 (m, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 164.77, 161.49, 136.48, 127.16, 122.54, 122.34, 119.66, 118.93, 114.09, 111.52, 111.42, 111.22, 111.19, 101.73, 101.40, 101.06, 31.39.

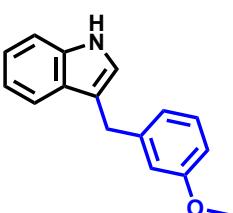
<sup>19</sup>F NMR (282 MHz, CDCl<sub>3</sub>) δ -110.73 (t, *J* = 8.3 Hz).



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.86 (bs, 1H), 7.48 (d, *J* = 2.2 Hz, 1H), 7.44 – 7.32 (m, 3H), 7.31 – 7.21 (m, 2H), 7.18 – 7.06 (m, 1H), 7.06 – 6.95 (m, 1H), 6.91 – 6.72 (m, 1H), 4.08 (s, 2H).

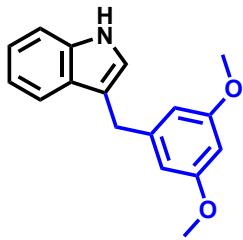
<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 142.17, 136.48, 132.08, 130.62 (q, *J* = 31.9 Hz), 128.78, 127.24, 125.68, 125.39 (q, *J* = 3.9 Hz), 122.88 (q, *J* = 3.8 Hz), 122.49, 122.29, 119.60, 118.96, 114.83, 111.22, 31.47.

<sup>19</sup>F NMR (376 MHz, Chloroform-*d*) δ -62.39 (d, *J* = 2.6 Hz).



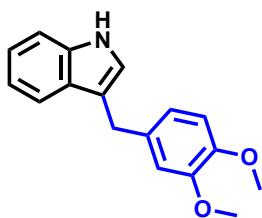
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.81 (bs, 1H), 7.44 (ddt, *J* = 7.8, 1.4, 0.7 Hz, 1H), 7.23 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.13 – 7.06 (m, 2H), 7.02 – 6.96 (m, 1H), 6.83 – 6.74 (m, 3H), 6.65 (dddd, *J* = 8.2, 2.7, 1.0, 0.5 Hz, 1H), 4.00 (s, 2H), 3.66 (s, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 159.69, 142.96, 136.47, 129.33, 127.49, 122.41, 122.06, 121.24, 119.40, 119.16, 115.60, 114.59, 111.20, 111.12, 55.17, 31.68.



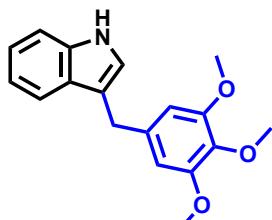
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.96 (bs, 1H), 7.65 – 7.51 (m, 1H), 7.35 (dt, *J* = 8.1, 0.9 Hz, 1H), 7.25 – 7.16 (m, 1H), 7.11 (ddd, *J* = 8.0, 7.0, 1.1 Hz, 1H), 6.93 (dt, *J* = 2.1, 1.0 Hz, 1H), 6.50 (dt, *J* = 2.3, 0.6 Hz, 2H), 6.35 (t, *J* = 2.3 Hz, 1H), 4.09 – 4.07 (m, 2H), 3.77 (s, 6H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 160.78, 143.77, 136.46, 127.49, 122.42, 122.05, 119.40, 119.13, 115.40, 111.11, 106.95, 97.88, 55.29, 31.92.



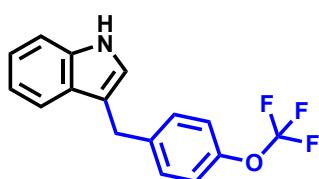
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.99 (bs, 1H), 7.54 (ddt, *J* = 7.9, 1.4, 0.7 Hz, 1H), 7.36 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.23 – 7.16 (m, 1H), 7.09 (ddd, *J* = 8.1, 7.0, 1.1 Hz, 1H), 6.92 – 6.77 (m, 4H), 4.09 – 4.07 (m, 2H), 3.86 (s, 3H), 3.82 (s, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 148.84, 147.24, 136.50, 133.82, 127.44, 122.26, 122.06, 120.59, 119.35, 119.14, 116.15, 112.11, 111.17, 111.10, 55.94, 55.82, 31.24.



<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.02 (bs, 1H), 7.56 (ddt, *J* = 7.8, 1.4, 0.7 Hz, 1H), 7.37 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.20 (dddd, *J* = 8.1, 7.1, 1.3, 0.4 Hz, 1H), 7.13 – 7.07 (m, 1H), 6.93 (dd, *J* = 2.3, 1.1 Hz, 1H), 6.53 (d, *J* = 0.6 Hz, 2H), 4.07 – 4.06 (m, 2H), 3.83 (s, 3H), 3.80 (s, 6H).

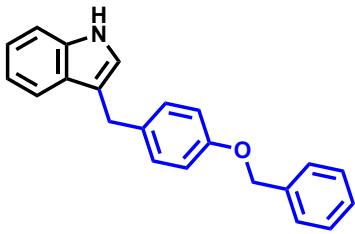
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 153.12, 136.93, 136.48, 127.43, 122.32, 122.11, 119.40, 119.07, 115.72, 111.12, 105.75, 60.88, 56.07, 32.00.



<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.97 (bs, 1H), 7.50 (ddt, *J* = 7.9, 1.5, 0.8 Hz, 1H), 7.38 (dt, *J* = 8.2, 0.9 Hz, 1H), 7.31 (dt, *J* = 1.5, 0.8 Hz, 1H), 7.30 – 7.28 (m, 1H), 7.22 (ddd, *J* = 8.2, 7.0, 1.3 Hz, 1H), 7.17 – 7.07 (m, 3H), 6.94 (dt, *J* = 2.2, 1.0 Hz, 1H), 4.13 (q, *J* = 0.8 Hz, 2H).

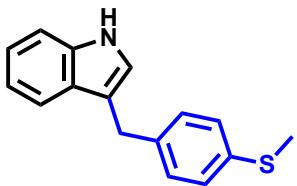
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 147.48, 139.99, 136.49, 129.85, 127.28, 122.42, 122.25, 120.90, 119.53, 119.04, 118.85, 115.19, 111.19, 30.94.

<sup>19</sup>F NMR (282 MHz, Chloroform-*d*) δ -57.87.



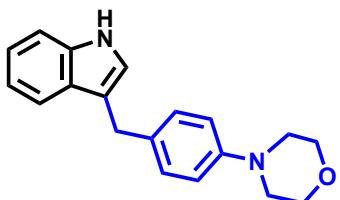
<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.93 (bs, 1H), 7.53 (dq, *J* = 7.9, 0.9 Hz, 1H), 7.45 – 7.30 (m, 6H), 7.23 – 7.17 (m, 3H), 7.09 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 6.94 – 6.88 (m, 3H), 5.04 (s, 2H), 4.07 (s, 2H).

<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 157.10, 137.25, 136.49, 133.62, 129.62, 128.58, 127.91, 127.51, 127.45, 122.23, 122.04, 119.35, 119.20, 116.26, 114.73, 111.07, 70.08, 30.73.



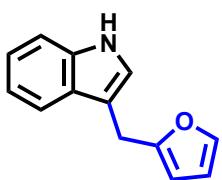
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.99 (bs, 1H), 7.57 (ddt, *J* = 7.9, 1.4, 0.7 Hz, 1H), 7.39 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.29 – 7.22 (m, 5H), 7.15 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 6.94 (dd, *J* = 2.2, 1.1 Hz, 1H), 4.15 – 4.11 (m, 2H), 2.51 (s, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 138.45, 136.50, 135.37, 129.72, 129.27, 127.41, 127.15, 123.76, 122.40, 122.11, 119.41, 119.15, 115.63, 111.17, 31.11, 16.34.



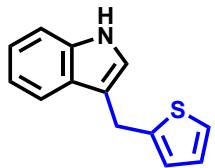
<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.96 (bs, 1H), 7.52 (dq, *J* = 7.9, 0.9 Hz, 1H), 7.35 (dt, *J* = 8.2, 0.9 Hz, 1H), 7.25 – 7.14 (m, 3H), 7.07 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 6.91 (dd, *J* = 2.3, 1.1 Hz, 3H), 4.07 – 4.03 (m, 2H), 3.88 (s, 4H), 3.14 (t, *J* = 4.8 Hz, 4H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 143.75, 136.50, 129.50, 129.24, 127.45, 122.26, 122.02, 119.31, 119.18, 116.37, 116.11, 111.08, 66.73, 50.21, 30.72.



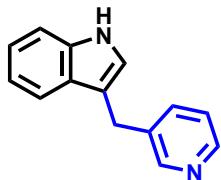
<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.92 (bs, 1H), 7.63 – 7.59 (m, 1H), 7.39 – 7.33 (m, 2H), 7.23 (ddd, *J* = 8.2, 7.0, 1.2 Hz, 1H), 7.15 (ddd, *J* = 8.0, 7.0, 1.1 Hz, 1H), 7.04 (dt, *J* = 2.2, 1.0 Hz, 1H), 6.32 (dd, *J* = 3.2, 1.9 Hz, 1H), 6.06 (dq, *J* = 3.0, 0.9 Hz, 1H), 4.16 (d, *J* = 1.1 Hz, 2H).

<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 154.93, 141.11, 136.34, 127.27, 122.38, 122.14, 119.49, 119.10, 112.64, 111.17, 110.32, 105.72, 24.50.



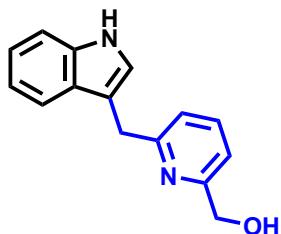
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.85 (bs, 1H), 7.49 (ddt, *J* = 7.8, 1.4, 0.8 Hz, 1H), 7.30 – 7.25 (m, 1H), 7.18 – 7.08 (m, 1H), 7.07 – 6.99 (m, 2H), 6.94 (dt, *J* = 2.1, 1.0 Hz, 1H), 6.88 – 6.78 (m, 2H), 4.24 (t, *J* = 1.0 Hz, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 144.69, 136.39, 127.13, 126.73, 124.75, 123.44, 122.28, 122.19, 119.50, 119.07, 115.31, 111.17, 25.97.



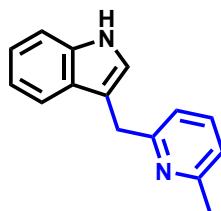
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.63 – 8.43 (m, 2H), 8.37 (bs, 1H), 7.53 (dddt, *J* = 24.7, 7.9, 1.4, 0.8 Hz, 2H), 7.36 (dt, *J* = 8.1, 0.9 Hz, 1H), 7.24 – 7.16 (m, 2H), 7.09 (ddd, *J* = 8.0, 7.0, 1.1 Hz, 1H), 6.93 (dt, *J* = 2.2, 1.0 Hz, 1H), 4.12 (s, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 149.83, 147.19, 136.82, 136.56, 136.41, 127.12, 123.45, 122.55, 122.25, 119.55, 118.85, 114.35, 111.28, 28.88.



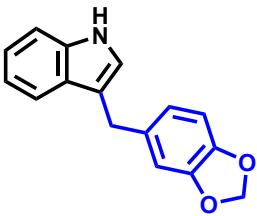
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.42 (bs, 1H), 7.59 – 7.45 (m, 2H), 7.32 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.23 – 7.01 (m, 4H), 6.97 (d, *J* = 2.3 Hz, 1H), 4.77 (s, 2H), 4.61 (bs, 1H), 4.30 (s, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 160.29, 158.18, 137.36, 136.47, 127.43, 122.92, 122.03, 121.40, 119.41, 119.08, 118.01, 113.40, 111.32, 63.94, 34.14.



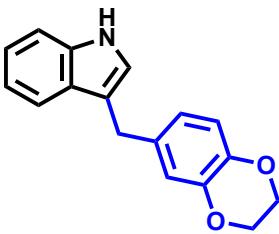
<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 8.25 (bs, 1H), 7.53 (dt, *J* = 7.9, 0.9 Hz, 1H), 7.42 (t, *J* = 7.7 Hz, 1H), 7.35 (dd, *J* = 8.1, 0.9 Hz, 1H), 7.18 (ddd, *J* = 8.2, 7.0, 1.1 Hz, 1H), 7.13 – 6.85 (m, 4H), 4.30 (s, 2H), 2.59 (s, 3H).

<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 160.54, 157.48, 136.96, 136.46, 127.52, 122.83, 122.03, 120.72, 119.73, 119.39, 119.26, 113.78, 111.13, 34.31, 24.40.



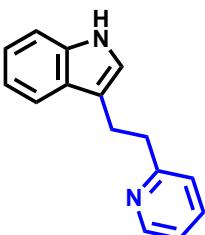
<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.89 (bs, 1H), 7.59 – 7.55 (m, 1H), 7.36 (dt, *J* = 8.1, 0.9 Hz, 1H), 7.24 (ddd, *J* = 8.2, 7.0, 1.2 Hz, 1H), 7.14 (ddd, *J* = 8.0, 7.0, 1.1 Hz, 1H), 6.92 (dt, *J* = 2.2, 1.0 Hz, 1H), 6.84 – 6.76 (m, 3H), 5.93 (s, 2H), 4.08 (s, 2H).

<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 147.64, 145.73, 136.52, 135.23, 127.40, 122.33, 122.12, 121.44, 119.44, 119.20, 115.94, 111.17, 109.31, 108.13, 100.82, 31.38.



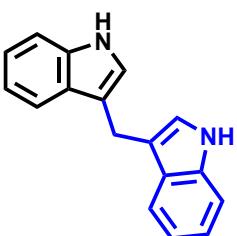
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.94 (bs, 1H), 7.54 (ddt, *J* = 7.8, 1.3, 0.7 Hz, 1H), 7.38 – 7.32 (m, 1H), 7.19 (dddd, *J* = 8.2, 7.1, 1.3, 0.4 Hz, 1H), 7.09 (ddd, *J* = 8.1, 7.0, 1.1 Hz, 1H), 6.93 (dt, *J* = 2.2, 1.0 Hz, 1H), 6.83 – 6.76 (m, 3H), 4.22 (s, 4H), 4.02 (t, *J* = 0.9 Hz, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 143.29, 141.70, 136.48, 134.67, 127.44, 122.25, 122.02, 121.59, 119.35, 119.18, 117.33, 116.99, 115.94, 111.07, 64.41, 64.34, 30.85.



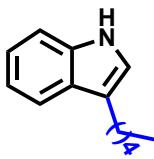
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.67 – 8.54 (m, 1H), 7.61 (ddd, *J* = 7.8, 1.3, 0.8 Hz, 1H), 7.50 (td, *J* = 7.7, 1.8 Hz, 1H), 7.41 – 7.31 (m, 1H), 7.24 – 7.01 (m, 3H), 7.01 – 6.79 (m, 2H), 6.41 (dd, *J* = 3.2, 0.9 Hz, 1H), 4.58 (t, *J* = 7.1 Hz, 2H), 3.29 (t, *J* = 7.1 Hz, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 158.27, 149.24, 136.80, 135.79, 128.59, 127.86, 123.85, 121.85, 121.44, 120.95, 119.29, 109.31, 101.12, 46.10, 38.72.



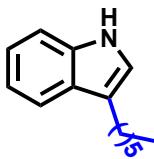
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.88 (bs, 2H), 7.64 (ddt, *J* = 7.8, 1.4, 0.8 Hz, 2H), 7.36 (ddd, *J* = 8.1, 0.8 Hz, 2H), 7.24 – 7.16 (m, 2H), 7.10 (ddd, *J* = 7.8, 7.0, 1.1 Hz, 2H), 6.96 – 6.89 (m, 2H), 4.25 (t, *J* = 1.0 Hz, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 136.48, 127.59, 122.22, 121.90, 119.24, 119.19, 115.70, 111.06, 21.22.



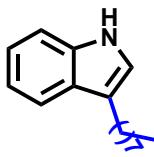
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.89 (bs, 1H), 7.62 (ddt, *J* = 7.8, 1.3, 0.6 Hz, 1H), 7.40 – 7.31 (m, 1H), 7.23 – 7.07 (m, 2H), 6.98 (dt, *J* = 2.1, 1.0 Hz, 1H), 2.80 – 2.71 (m, 2H), 1.80 – 1.66 (m, 2H), 1.49 – 1.27 (m, 4H), 0.96 – 0.83 (m, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 148.43, 136.35, 124.59, 121.82, 120.98, 119.04, 117.26, 111.01, 31.88, 29.87, 25.13, 22.61, 14.13.



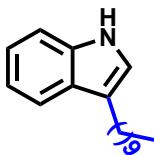
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.88 (bs, 1H), 7.68 – 7.55 (m, 1H), 7.36 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.25 – 7.03 (m, 2H), 7.01 – 6.93 (m, 1H), 2.89 – 2.62 (m, 2H), 1.83 – 1.64 (m, 2H), 1.45 – 1.25 (m, 6H), 0.97 – 0.82 (m, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 136.35, 128.25, 127.66, 121.82, 120.98, 119.04, 117.26, 111.01, 31.81, 30.16, 29.37, 25.18, 22.72, 14.16.



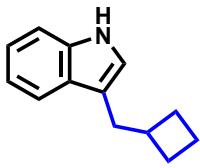
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.79 (bs, 1H), 7.54 (ddt, *J* = 7.7, 1.4, 0.7 Hz, 1H), 7.32 – 7.22 (m, 1H), 7.14 – 7.00 (m, 2H), 6.89 (dt, *J* = 2.2, 1.0 Hz, 1H), 2.67 (ddt, *J* = 8.6, 7.0, 0.8 Hz, 2H), 1.71 – 1.55 (m, 2H), 1.44 – 0.90 (m, 10H), 0.84 – 0.77 (m, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 136.37, 127.67, 121.82, 120.97, 119.04, 117.27, 111.00, 31.81, 30.16, 29.70, 29.55, 29.36, 25.18, 22.71, 14.14.

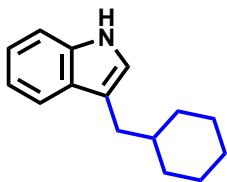


<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.82 (bs, 1H), 7.67 (dq, *J* = 7.7, 0.9 Hz, 1H), 7.37 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.29 – 7.11 (m, 2H), 6.97 (dd, *J* = 2.3, 1.1 Hz, 1H), 2.94 – 2.70 (m, 2H), 1.88 – 1.66 (m, 2H), 1.51 – 1.26 (m, 14H), 1.08 – 0.80 (m, 3H).

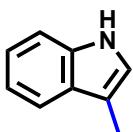
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 136.39, 127.70, 121.85, 121.05, 119.08, 117.24, 111.08, 32.01, 30.26, 29.77, 29.75, 29.66, 29.45, 25.23, 22.78, 14.21.



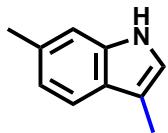
<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.89 (bs, 1H), 7.61 (d, *J* = 7.8 Hz, 1H), 7.35 (d, *J* = 8.0 Hz, 1H), 7.24 – 7.04 (m, 2H), 6.94 (d, *J* = 2.3 Hz, 1H), 2.85 (d, *J* = 7.4 Hz, 2H), 2.72 (dt, *J* = 15.2, 7.7 Hz, 1H), 2.18 – 2.01 (m, 2H), 1.88 (dddd, *J* = 12.3, 10.6, 4.8, 3.3 Hz, 2H), 1.81 – 1.71 (m, 2H).  
<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 136.25, 127.82, 121.81, 121.13, 119.07, 119.03, 115.56, 110.97, 36.27, 32.31, 28.45, 18.29.



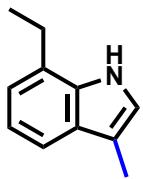
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.90 (bs, 1H), 7.61 (ddt, *J* = 7.7, 1.5, 0.7 Hz, 1H), 7.41 – 7.30 (m, 1H), 7.19 (ddd, *J* = 8.2, 7.0, 1.3 Hz, 1H), 7.15 – 7.07 (m, 1H), 6.95 (dd, *J* = 2.1, 1.0 Hz, 1H), 2.64 (dd, *J* = 6.9, 0.8 Hz, 2H), 1.82 – 1.56 (m, 7H), 1.21 – 1.15 (m, 2H), 1.05 – 0.91 (m, 2H).  
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 136.27, 128.08, 121.88, 121.70, 119.24, 119.01, 115.53, 110.98, 38.80, 33.57, 33.13, 26.66, 26.40.



<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.83 (bs, 1H), 7.62 (ddd, *J* = 7.6, 1.4, 0.8 Hz, 1H), 7.41 – 7.32 (m, 1H), 7.27 – 7.12 (m, 2H), 6.97 (s, 1H), 2.37 (d, *J* = 1.1 Hz, 3H).  
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 136.30, 128.32, 121.90, 121.60, 119.15, 118.86, 111.76, 110.97, 9.69.

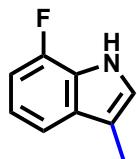


<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.69 (bs, 1H), 7.50 (d, *J* = 8.0 Hz, 1H), 7.15 – 7.13 (m, 1H), 7.00 (ddt, *J* = 7.6, 1.5, 0.6 Hz, 1H), 6.90 (dq, *J* = 2.2, 1.1 Hz, 1H), 2.51 (s, 3H), 2.36 (d, *J* = 1.1 Hz, 3H).  
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 136.76, 131.65, 126.22, 120.95, 120.92, 118.53, 111.56, 110.97, 21.76, 9.77.



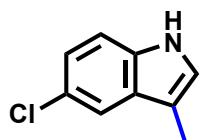
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.81 (bs, 1H), 7.49 (ddq, *J* = 7.7, 1.2, 0.6 Hz, 1H), 6.98 (tq, *J* = 2.5, 1.1 Hz, 1H), 2.88 (qt, *J* = 7.5, 0.6 Hz, 2H), 2.38 (d, *J* = 1.1 Hz, 3H), 1.40 (t, *J* = 7.6 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 135.13, 128.07, 126.39, 121.24, 120.47, 119.49, 116.64, 112.22, 24.06, 13.91, 9.85.



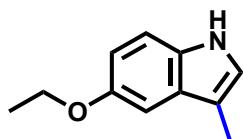
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.04 (bs, 1H), 7.34 (dq, *J* = 7.9, 0.8 Hz, 1H), 7.08 – 6.96 (m, 2H), 6.90 (dd, *J* = 11.2, 7.8, 0.9, 0.4 Hz, 1H), 2.34 (d, *J* = 1.1 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 149.55 (d, *J* = 243.4 Hz), 132.11, 122.25, 119.33 (d, *J* = 6.1 Hz), 114.62 (d, *J* = 3.4 Hz), 112.62, 106.83, 106.62, 9.74.



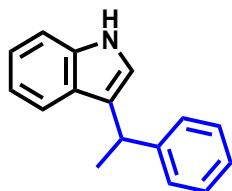
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.86 (bs, 1H), 7.57 (dt, *J* = 2.0, 0.7 Hz, 1H), 7.25 (dd, *J* = 8.6, 0.6 Hz, 1H), 7.16 (ddd, *J* = 8.6, 2.0, 0.4 Hz, 1H), 7.04 – 6.93 (m, 1H), 2.31 (d, *J* = 1.1 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 134.60, 129.46, 124.89, 123.07, 122.14, 118.44, 111.98, 111.58, 9.57.



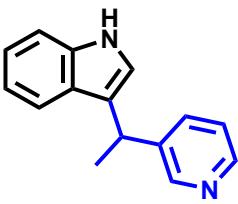
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.78 (bs, 1H), 7.23 (dd, *J* = 8.8, 0.6 Hz, 1H), 7.05 (d, *J* = 2.4 Hz, 1H), 7.00 – 6.82 (m, 2H), 4.13 (q, *J* = 7.0 Hz, 2H), 2.32 (d, *J* = 1.1 Hz, 3H), 1.47 (t, *J* = 7.0 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 153.11, 131.50, 128.68, 122.48, 112.64, 111.66, 111.39, 101.96, 64.33, 15.15, 9.76.



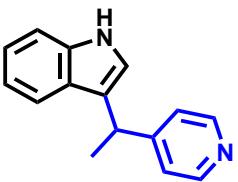
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.86 (bs, 1H), 7.63 (ddt, *J* = 7.7, 1.6, 0.8 Hz, 1H), 7.38 – 7.30 (m, 2H), 7.24 – 7.09 (m, 5H), 6.97 – 6.95 (m, 1H), 6.53 (ddd, *J* = 3.1, 2.1, 1.0 Hz, 1H), 4.34 (qd, *J* = 7.2, 1.0 Hz, 1H), 1.68 (d, *J* = 7.1 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 146.85, 136.65, 128.35, 127.49, 125.96, 124.17, 122.00, 121.12, 120.77, 119.75, 119.24, 111.06, 102.65, 36.98, 22.45.



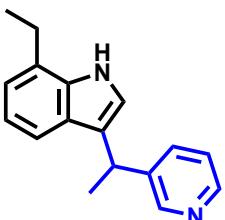
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.62 (s, 1H), 8.44 (d, *J* = 4.7 Hz, 1H), 8.24 (s, 1H), 7.56 (dt, *J* = 7.9, 1.8 Hz, 1H), 7.42 – 7.28 (m, 2H), 7.24 – 7.11 (m, 2H), 7.09 – 6.93 (m, 2H), 4.41 (q, *J* = 7.3 Hz, 1H), 1.73 (d, *J* = 7.2 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 148.90, 147.07, 136.70, 135.26, 126.47, 123.55, 122.23, 121.27, 119.96, 119.44, 119.36, 111.22, 34.55, 22.11.



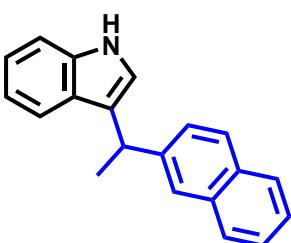
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.56 – 8.40 (m, 2H), 8.17 (bs, 1H), 7.37 (dt, *J* = 8.2, 0.9 Hz, 1H), 7.29 (ddt, *J* = 8.0, 1.5, 0.8 Hz, 1H), 7.25 – 7.12 (m, 3H), 7.09 – 6.97 (m, 2H), 4.36 (q, *J* = 7.2 Hz, 1H), 1.71 (d, *J* = 7.2 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 149.37, 123.01, 122.32, 121.36, 119.51, 119.31, 111.23, 36.45, 21.56.



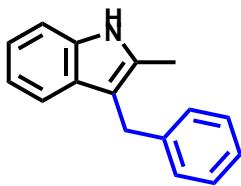
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.62 (d, *J* = 2.3 Hz, 1H), 8.44 (dd, *J* = 4.9, 1.7 Hz, 1H), 8.11 (bs, 1H), 7.61 (dddd, *J* = 7.9, 2.2, 1.6, 0.6 Hz, 1H), 7.26 – 7.13 (m, 2H), 7.09 – 6.91 (m, 3H), 4.42 (q, *J* = 7.2 Hz, 1H), 2.86 (q, *J* = 7.6 Hz, 2H), 1.73 (d, *J* = 7.2 Hz, 3H), 1.36 (t, *J* = 7.6 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 156.86, 148.29, 146.40, 138.47, 135.88, 126.61, 123.70, 120.86, 120.83, 120.28, 119.83, 117.04, 34.64, 23.99, 22.07, 13.79.



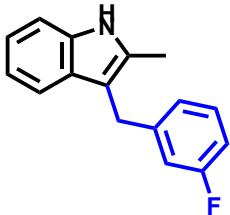
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.98 (bs, 1H), 7.82 – 7.71 (m, 4H), 7.47 – 7.32 (m, 5H), 7.15 (ddd, *J* = 8.2, 7.0, 1.2 Hz, 1H), 7.07 – 6.93 (m, 2H), 4.55 (q, *J* = 7.1 Hz, 1H), 1.80 (d, *J* = 7.1 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 144.32, 136.68, 133.65, 132.22, 127.91, 127.72, 127.58, 126.95, 126.56, 125.76, 125.30, 125.16, 122.01, 121.38, 121.22, 119.73, 119.27, 111.01, 37.09, 22.30.



<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.80 (bs, 1H), 7.43 (ddt, *J* = 7.7, 1.5, 0.8 Hz, 1H), 7.33 – 7.25 (m, 5H), 7.21 – 7.11 (m, 2H), 7.10 – 7.05 (m, 1H), 4.11 (s, 2H), 2.41 (s, 3H).

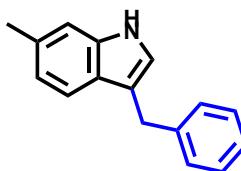
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 141.68, 135.31, 131.66, 128.93, 128.30, 127.18, 125.68, 121.02, 119.27, 118.40, 110.58, 110.15, 30.12, 11.83.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.64 (bs, 1H), 7.20 (dd, *J* = 7.8, 1.0 Hz, 1H), 7.12 (dt, *J* = 8.0, 0.9 Hz, 1H), 7.03 (td, *J* = 7.9, 6.0 Hz, 1H), 6.95 (ddd, *J* = 8.2, 7.1, 1.3 Hz, 1H), 6.90 – 6.83 (m, 2H), 6.74 – 6.64 (m, 2H), 3.90 (s, 2H), 2.22 (s, 3H).

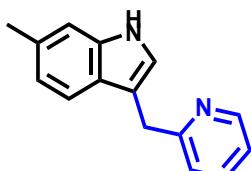
<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 163.03 (d, *J* = 245.1 Hz), 144.38 (d, *J* = 6.9 Hz), 135.31, 131.80, 129.62 (d, *J* = 8.2 Hz), 128.73, 123.89 (d, *J* = 2.7 Hz), 121.17, 119.40, 118.22, 115.08 (d, *J* = 21.3 Hz), 112.58 (d, *J* = 21.1 Hz), 110.22, 109.89, 29.86 (d, *J* = 1.8 Hz), 11.79.

<sup>19</sup>F NMR (282 MHz, Chloroform-*d*) δ -113.88 (ddd, *J* = 10.1, 8.8, 6.0 Hz).



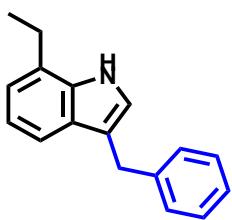
<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.72 (bs, 1H), 7.38 (d, *J* = 8.1 Hz, 1H), 7.30 – 7.15 (m, 5H), 7.11 (dt, *J* = 1.5, 0.8 Hz, 1H), 6.91 (dd, *J* = 8.2, 1.4 Hz, 1H), 6.80 (dt, *J* = 2.1, 1.0 Hz, 1H), 4.09 (d, *J* = 1.0 Hz, 2H), 2.45 (s, 3H).

<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 141.38, 136.96, 131.87, 128.71, 128.35, 125.87, 125.40, 121.75, 121.16, 118.87, 115.64, 111.09, 31.69, 21.76.



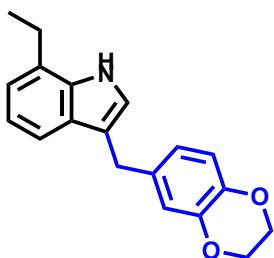
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.56 (ddd, *J* = 4.9, 1.9, 0.9 Hz, 1H), 8.08 (bs, 1H), 7.54 (td, *J* = 7.7, 1.9 Hz, 1H), 7.43 – 7.33 (m, 1H), 7.22 – 7.05 (m, 3H), 7.05 – 6.97 (m, 1H), 6.91 (ddd, *J* = 8.1, 1.5, 0.6 Hz, 1H), 4.30 (d, *J* = 0.8 Hz, 2H), 2.44 (t, *J* = 0.7 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 161.27, 148.79, 136.95, 136.74, 131.91, 125.28, 122.88, 122.12, 121.22, 121.14, 118.82, 113.52, 111.13, 34.44, 21.70.



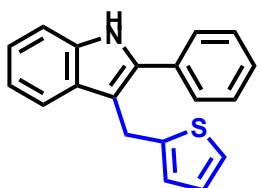
<sup>1</sup>H NMR (300 MHz, Chloroform-d) δ 7.88 (bs, 1H), 7.55 – 7.12 (m, 6H), 7.09 – 6.99 (m, 2H), 6.89 (dt, *J* = 2.2, 1.0 Hz, 1H), 4.10 (d, *J* = 1.0 Hz, 2H), 2.84 (qd, *J* = 7.6, 0.8 Hz, 2H), 1.36 (td, *J* = 7.6, 4.3 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-d) δ 141.30, 135.31, 128.73, 128.33, 127.23, 126.44, 125.86, 121.94, 120.60, 119.69, 116.94, 116.35, 31.74, 24.02, 13.83.



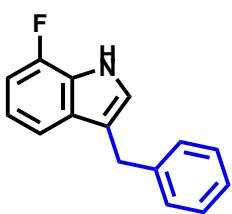
<sup>1</sup>H NMR (300 MHz, Chloroform-d) δ 7.91 (bs, 1H), 7.45 (dd, *J* = 7.2, 2.0 Hz, 1H), 7.19 – 6.99 (m, 2H), 6.99 – 6.72 (m, 4H), 4.24 (s, 4H), 4.05 (q, *J* = 1.0 Hz, 2H), 2.87 (q, *J* = 7.6 Hz, 2H), 1.40 (td, *J* = 7.6, 1.1 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-d) δ 143.33, 141.72, 135.35, 134.83, 127.23, 126.49, 121.94, 121.66, 120.58, 119.70, 117.40, 117.02, 116.97, 116.40, 64.44, 64.37, 31.02, 24.04, 13.91.



<sup>1</sup>H NMR (300 MHz, Chloroform-d) δ 8.22 – 8.02 (m, 1H), 7.60 – 7.52 (m, 3H), 7.50 – 7.37 (m, 4H), 7.22 (ddd, *J* = 8.1, 7.1, 1.2 Hz, 1H), 7.15 – 7.08 (m, 2H), 6.91 (dd, *J* = 5.1, 3.4 Hz, 1H), 6.83 (dq, *J* = 3.5, 1.2 Hz, 1H), 4.40 (d, *J* = 1.2 Hz, 2H).

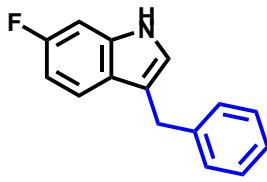
<sup>13</sup>C NMR (75 MHz, Chloroform-d) δ 145.37, 135.93, 132.70, 128.97, 127.96, 127.94, 126.79, 124.39, 123.30, 122.52, 119.92, 119.46, 111.22, 110.82, 91.81, 25.34.



<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.09 (bs, 1H), 7.35 – 7.27 (m, 5H), 7.26 – 7.20 (m, 1H), 7.01 (tdd, *J* = 7.9, 4.8, 0.9 Hz, 1H), 6.97 – 6.89 (m, 2H), 4.13 (s, 2H).

<sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 149.59 (d, *J* = 243.7 Hz), 140.87, 131.22 (d, *J* = 5.1 Hz), 128.69, 128.45, 126.07, 124.78 (d, *J* = 13.1 Hz), 123.04, 119.66 (d, *J* = 6.1 Hz), 116.71 (d, *J* = 2.3 Hz), 114.99 (d, *J* = 3.5 Hz), 106.93 (d, *J* = 16.1 Hz), 31.63.

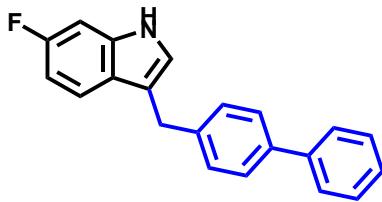
<sup>19</sup>F NMR (376 MHz, Chloroform-d) δ -135.43 – -135.51 (m).



<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.89 (bs, 1H), 7.38 (ddt, *J* = 8.6, 5.3, 0.6 Hz, 1H), 7.32 – 7.14 (m, 5H), 7.01 (ddd, *J* = 9.7, 2.3, 0.5 Hz, 1H), 6.90 – 6.75 (m, 2H), 4.08 (d, *J* = 1.0 Hz, 2H).

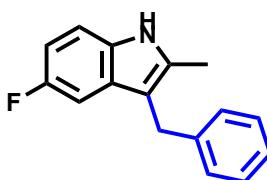
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 160.06 (d, *J* = 237.5 Hz), 140.92, 136.29, 128.65, 128.40, 126.02, 122.56 (d, *J* = 3.6 Hz), 119.90 (d, *J* = 10.2 Hz), 115.96, 108.14 (d, *J* = 24.5 Hz), 97.38 (d, *J* = 26.0 Hz), 31.59.

<sup>19</sup>F NMR (282 MHz, Chloroform-*d*) δ -121.26, -121.29 (d, *J* = 4.1 Hz), -121.33 (d, *J* = 4.1 Hz), -121.35, -121.38.



<sup>1</sup>H NMR (300 MHz, DMSO-*d*<sub>6</sub>) δ 10.96 (bs, 1H), 7.65 – 7.53 (m, 4H), 7.47 – 7.40 (m, 3H), 7.39 – 7.29 (m, 3H), 7.24 – 7.19 (m, 1H), 7.13 (ddd, *J* = 10.2, 2.4, 0.5 Hz, 1H), 6.81 (ddd, *J* = 9.8, 8.7, 2.4 Hz, 1H), 4.07 (s, 2H).

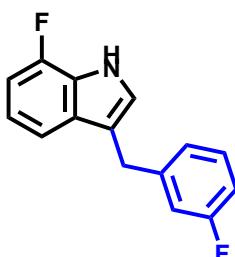
<sup>13</sup>C NMR (75 MHz, DMSO-*d*<sub>6</sub>) δ 159.32 (d, *J* = 233.8 Hz), 141.39, 140.56, 138.11, 136.80, 136.63, 129.38 (d, *J* = 7.4 Hz), 127.60, 126.98 (d, *J* = 4.5 Hz), 124.33, 124.26 (d, *J* = 3.6 Hz), 119.98 (d, *J* = 10.2 Hz), 114.44, 107.41, 107.09, 97.82 (d, *J* = 25.4 Hz), 31.00.



<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.77 (bs, 1H), 7.27 – 7.14 (m, 6H), 7.02 (dd, *J* = 9.8, 2.5 Hz, 1H), 6.83 (ddd, *J* = 9.4, 8.7, 2.5 Hz, 1H), 4.02 (s, 2H), 2.39 (s, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 157.81 (d, *J* = 233.6 Hz), 141.22, 133.66, 131.73, 128.38, 128.21, 125.84, 110.59 (d, *J* = 9.7 Hz), 109.19, 108.84, 103.47 (d, *J* = 23.5 Hz), 30.13, 11.95.

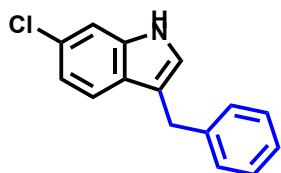
<sup>19</sup>F NMR (282 MHz, Chloroform-*d*) δ -125.02 (td, *J* = 9.5, 4.4 Hz).



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 8.15 (bs, 1H), 7.29 – 7.21 (m, 2H), 7.07 (ddd, *J* = 7.6, 1.7, 0.9 Hz, 1H), 7.04 – 6.85 (m, 5H), 4.11 (t, *J* = 0.7 Hz, 2H).

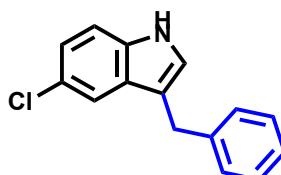
<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 163.02 (d, *J* = 245.2 Hz), 149.59 (d, *J* = 243.7 Hz), 143.51 (d, *J* = 7.1 Hz), 131.02 (d, *J* = 5.1 Hz), 129.78 (d, *J* = 8.4 Hz), 124.80 (d, *J* = 13.0 Hz), 124.25 (d, *J* = 2.8 Hz), 123.11, 119.79 (d, *J* = 6.1 Hz), 115.84 (d, *J* = 2.2 Hz), 115.60, 115.39, 114.85 (d, *J* = 3.5 Hz), 112.96 (d, *J* = 21.1 Hz), 107.05 (d, *J* = 16.1 Hz), 31.35.

<sup>19</sup>F NMR (376 MHz, Chloroform-*d*) δ -113.56 – -113.74 (m), -135.41 (dt, *J* = 11.1, 4.1 Hz).



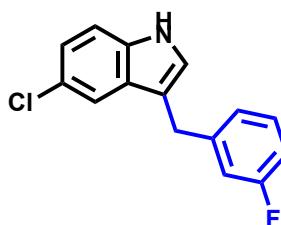
<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.95 (bs, 1H), 7.39 (dt, *J* = 8.4, 0.6 Hz, 1H), 7.35 (dd, *J* = 1.8, 0.6 Hz, 1H), 7.30 – 7.22 (m, 5H), 7.03 (dd, *J* = 8.4, 1.8 Hz, 1H), 6.92 (dt, *J* = 2.2, 1.0 Hz, 1H), 4.08 (dt, *J* = 1.0, 0.6 Hz, 2H).

<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 140.82, 136.80, 128.62, 128.41, 128.03, 126.09, 126.04, 122.96, 120.14, 120.07, 116.02, 111.01, 31.49.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.93 (bs, 1H), 7.54 (dq, *J* = 7.9, 0.9 Hz, 1H), 7.37 (dt, *J* = 8.2, 0.9 Hz, 1H), 7.31 – 7.28 (m, 3H), 7.23 – 7.17 (m, 2H), 7.09 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H), 6.92 (dt, *J* = 2.3, 1.0 Hz, 1H), 4.13 (d, *J* = 1.0 Hz, 2H).

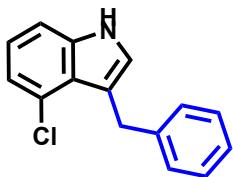
<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 141.22, 136.46, 128.71, 128.34, 127.48, 125.89, 122.33, 122.07, 119.38, 119.18, 115.87, 111.08, 31.61.



<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 8.00 (bs, 1H), 7.43 (dt, *J* = 2.0, 0.7 Hz, 1H), 7.28 – 7.21 (m, 2H), 7.13 (ddd, *J* = 8.6, 2.0, 0.4 Hz, 1H), 7.03 (ddq, *J* = 7.6, 1.6, 0.8 Hz, 1H), 6.98 – 6.95 (m, 1H), 6.95 – 6.84 (m, 2H), 4.05 (p, *J* = 0.7 Hz, 2H).

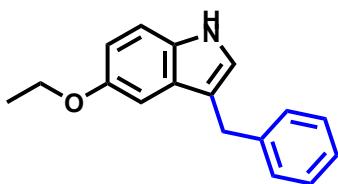
<sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>) δ 163.03 (d, *J* = 245.7 Hz), 143.41, 134.80, 129.81 (d, *J* = 8.2 Hz), 128.43, 125.30, 124.20 (d, *J* = 2.8 Hz), 123.83, 122.55, 118.54, 115.56, 115.28, 114.80, 113.13, 112.86, 112.16, 31.14 (d, *J* = 1.7 Hz).

<sup>19</sup>F NMR (282 MHz, Chloroform-*d*) δ -113.61 (d, *J* = 4.0 Hz), -113.62 – -113.66 (m), -113.67.



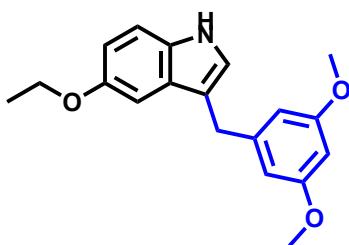
<sup>1</sup>H NMR (300 MHz, Chloroform-d) δ 7.95 (bs, 1H), 7.53 (ddt, *J* = 7.9, 1.3, 0.7 Hz, 1H), 7.39 – 7.34 (m, 1H), 7.32 – 7.26 (m, 3H), 7.23 – 7.17 (m, 2H), 7.12 – 7.05 (m, 1H), 6.92 (dt, *J* = 2.2, 0.9 Hz, 1H), 4.45 – 3.92 (m, 2H).

<sup>13</sup>C NMR (75 MHz, Chloroform-d) δ 141.21, 136.47, 128.70, 128.33, 125.88, 122.32, 122.06, 119.37, 119.17, 115.88, 111.06, 31.61.



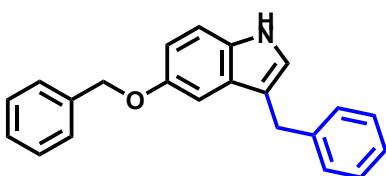
<sup>1</sup>H NMR (300 MHz, Chloroform-d) δ 7.85 (bs, 1H), 7.42 – 7.15 (m, 6H), 7.03 (dt, *J* = 2.5, 0.6 Hz, 1H), 6.95 – 6.86 (m, 2H), 4.15 – 4.13 (m, 2H), 4.09 (q, *J* = 7.0 Hz, 2H), 1.47 (t, *J* = 7.0 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-d) δ 153.16, 141.26, 131.70, 128.75, 128.41, 127.95, 125.94, 123.27, 115.44, 112.73, 111.83, 102.29, 64.28, 31.67, 15.11.



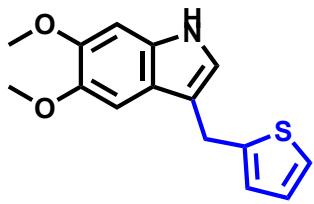
<sup>1</sup>H NMR (300 MHz, Chloroform-d) δ 7.92 (bs, 1H), 7.24 (dd, *J* = 8.7, 0.6 Hz, 1H), 7.03 (dd, *J* = 2.5, 0.6 Hz, 1H), 6.97 – 6.81 (m, 2H), 6.49 (dt, *J* = 2.3, 0.6 Hz, 2H), 6.34 (t, *J* = 2.3 Hz, 1H), 4.13 – 4.03 (m, 4H), 3.78 (s, 6H), 1.45 (t, *J* = 7.0 Hz, 3H).

<sup>13</sup>C NMR (75 MHz, Chloroform-d) δ 160.78, 153.16, 143.72, 131.65, 127.92, 123.25, 115.02, 112.71, 111.76, 106.91, 106.39, 102.19, 97.88, 64.26, 55.28, 31.90, 15.08.



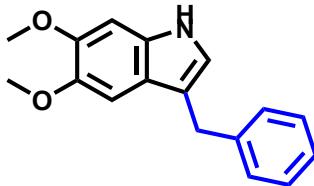
<sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.70 (bs, 1H), 7.41 – 7.32 (m, 2H), 7.32 – 7.25 (m, 2H), 7.25 – 7.15 (m, 5H), 7.14 – 7.09 (m, 2H), 6.96 (dd, *J* = 2.5, 0.6 Hz, 1H), 6.88 – 6.79 (m, 1H), 6.78 – 6.73 (m, 1H), 4.96 (s, 2H), 3.98 (d, *J* = 0.9 Hz, 2H).

<sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 153.14, 141.20, 137.72, 131.83, 128.73, 128.56, 128.41, 127.89, 127.83, 127.71, 125.94, 123.30, 115.56, 112.91, 111.84, 102.78, 71.00, 31.67.



<sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.94 (bs, 1H), 7.13 (dd, *J* = 5.1, 1.2 Hz, 1H), 6.98 (s, 1H), 6.96 – 6.86 (m, 3H), 6.83 (s, 1H), 4.28 (d, *J* = 1.0 Hz, 2H), 3.89 (s, 3H), 3.88 (s, 3H).

<sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 147.13, 144.88, 144.83, 130.69, 126.73, 124.68, 123.42, 120.91, 119.93, 114.97, 100.83, 94.70, 56.39, 56.18, 26.11.

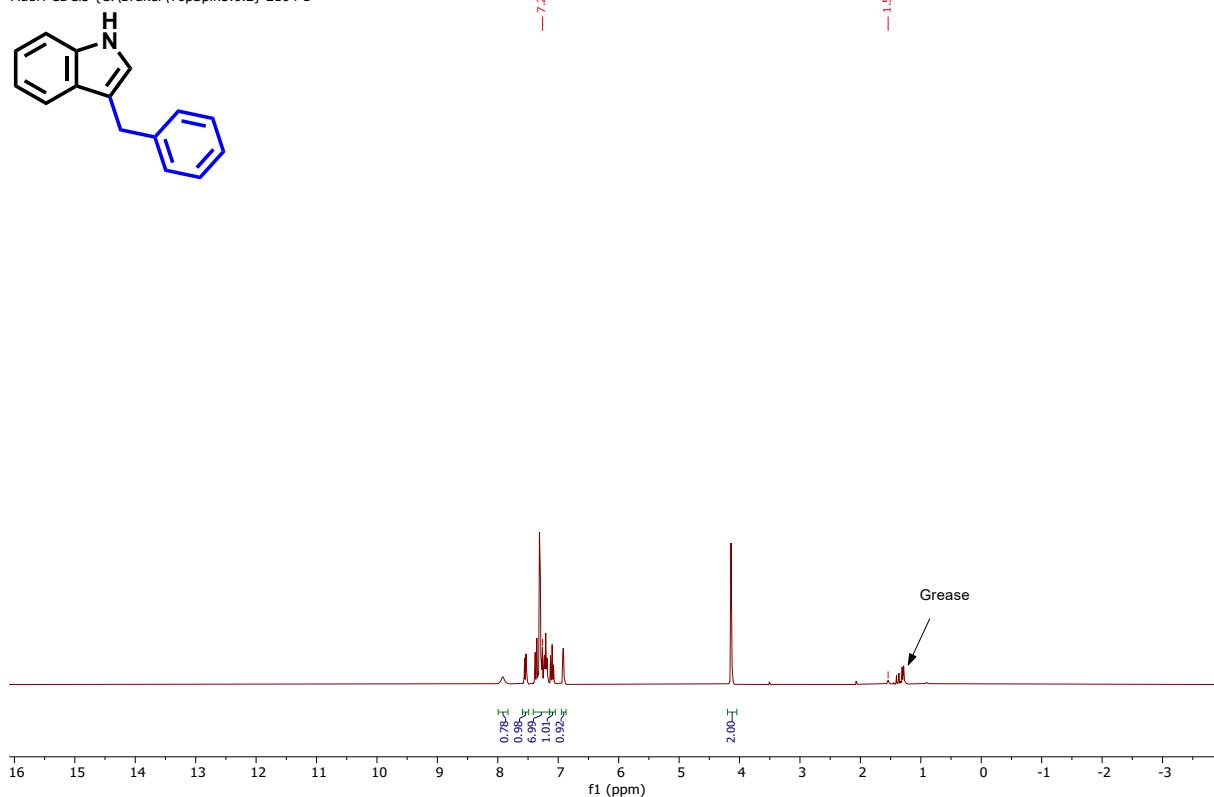


<sup>1</sup>H NMR (300 MHz, Chloroform-*d*) δ 7.81 (bs, 1H), 7.29 – 7.08 (m, 5H), 6.86 (s, 1H), 6.76 (s, 1H), 6.71 (dt, *J* = 2.1, 1.0 Hz, 1H), 4.02 (d, *J* = 0.9 Hz, 2H), 3.81 (s, 3H), 3.80 (s, 3H).

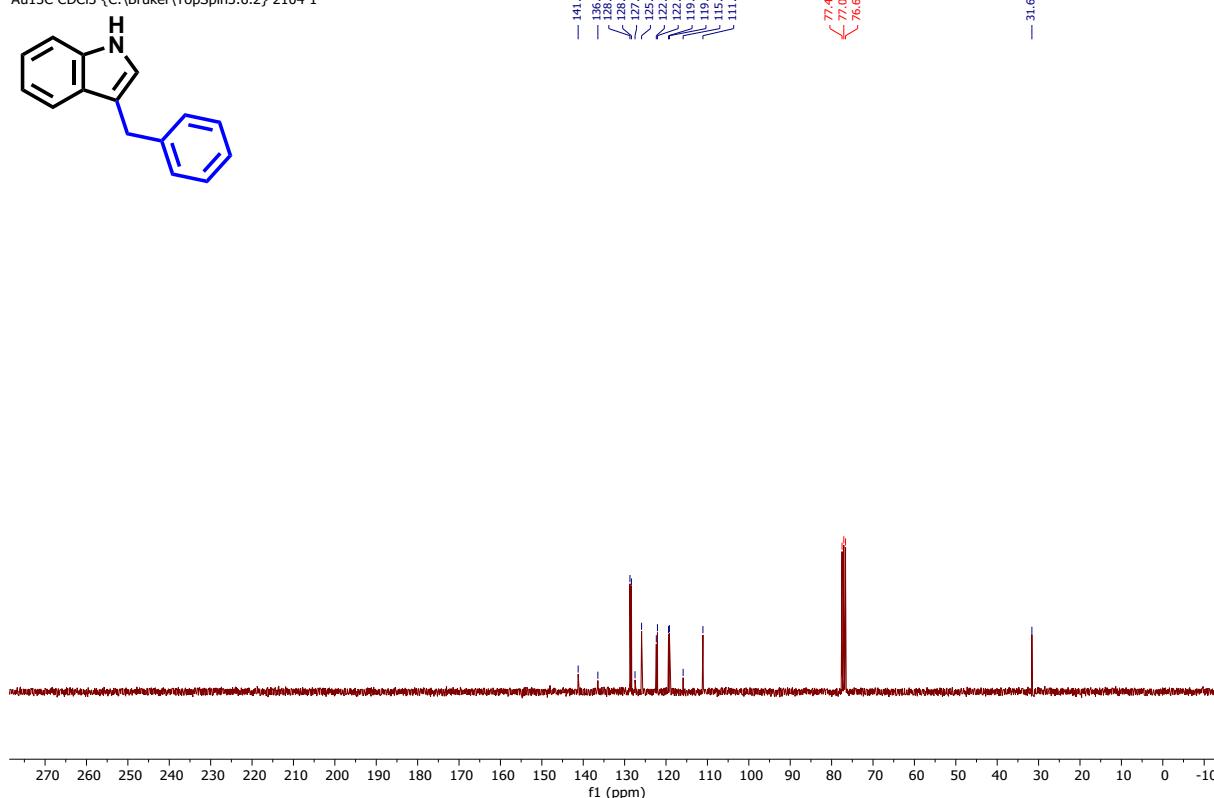
<sup>13</sup>C NMR (75 MHz, Chloroform-*d*) δ 147.07, 144.81, 141.27, 130.75, 128.71, 128.37, 125.92, 121.02, 120.30, 115.52, 100.97, 94.66, 56.38, 56.20, 31.76.

## S7 NMR spectra

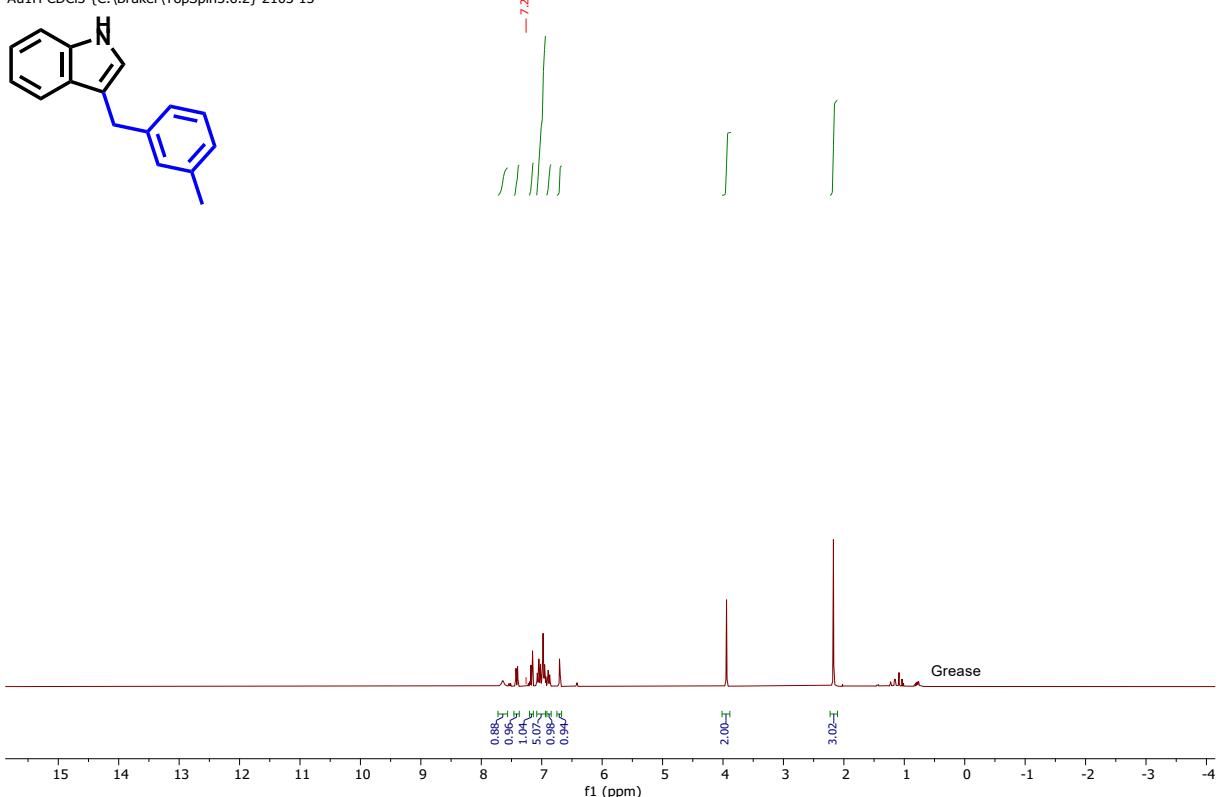
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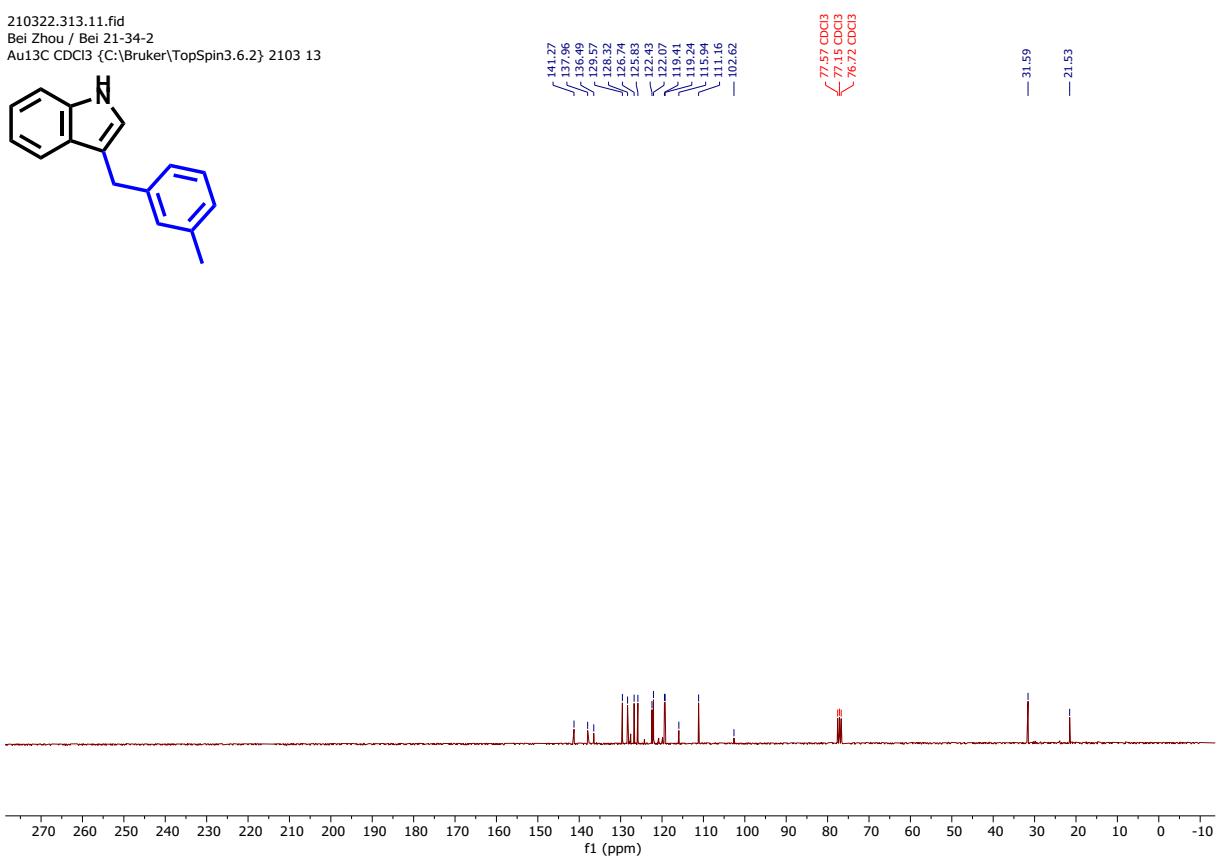
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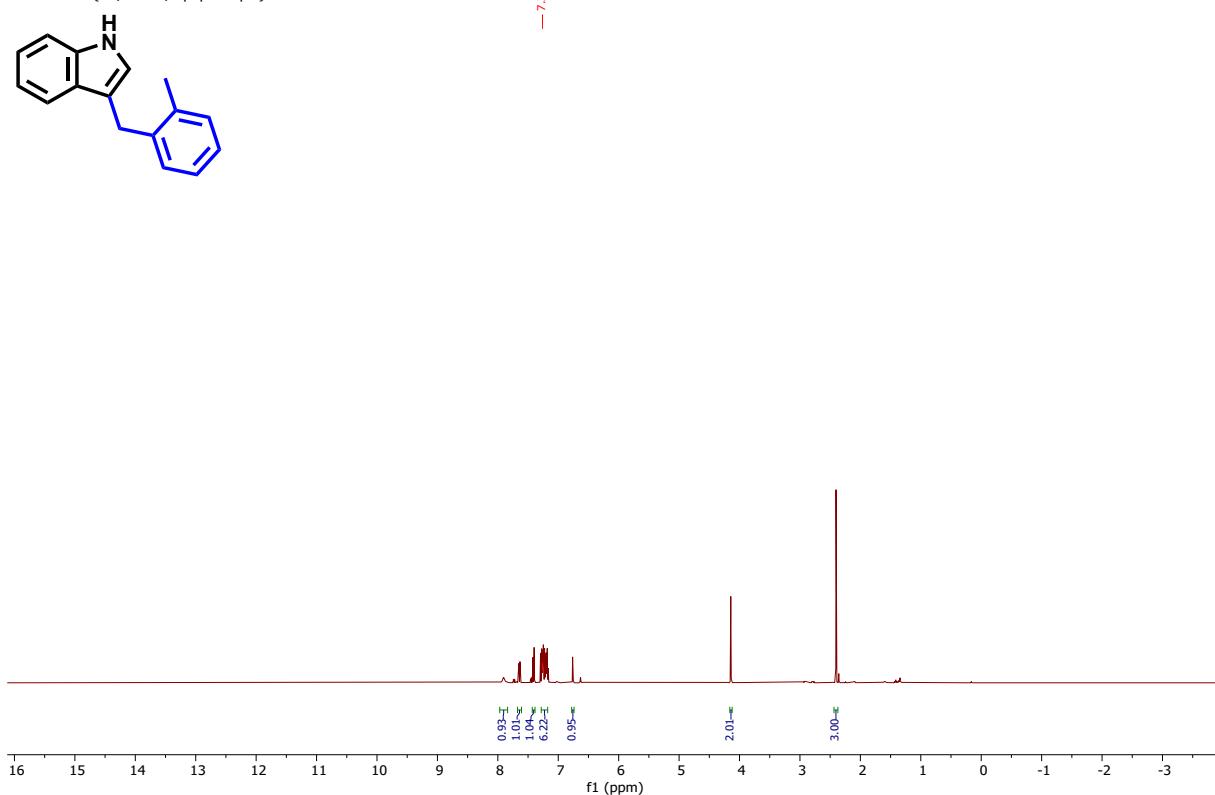
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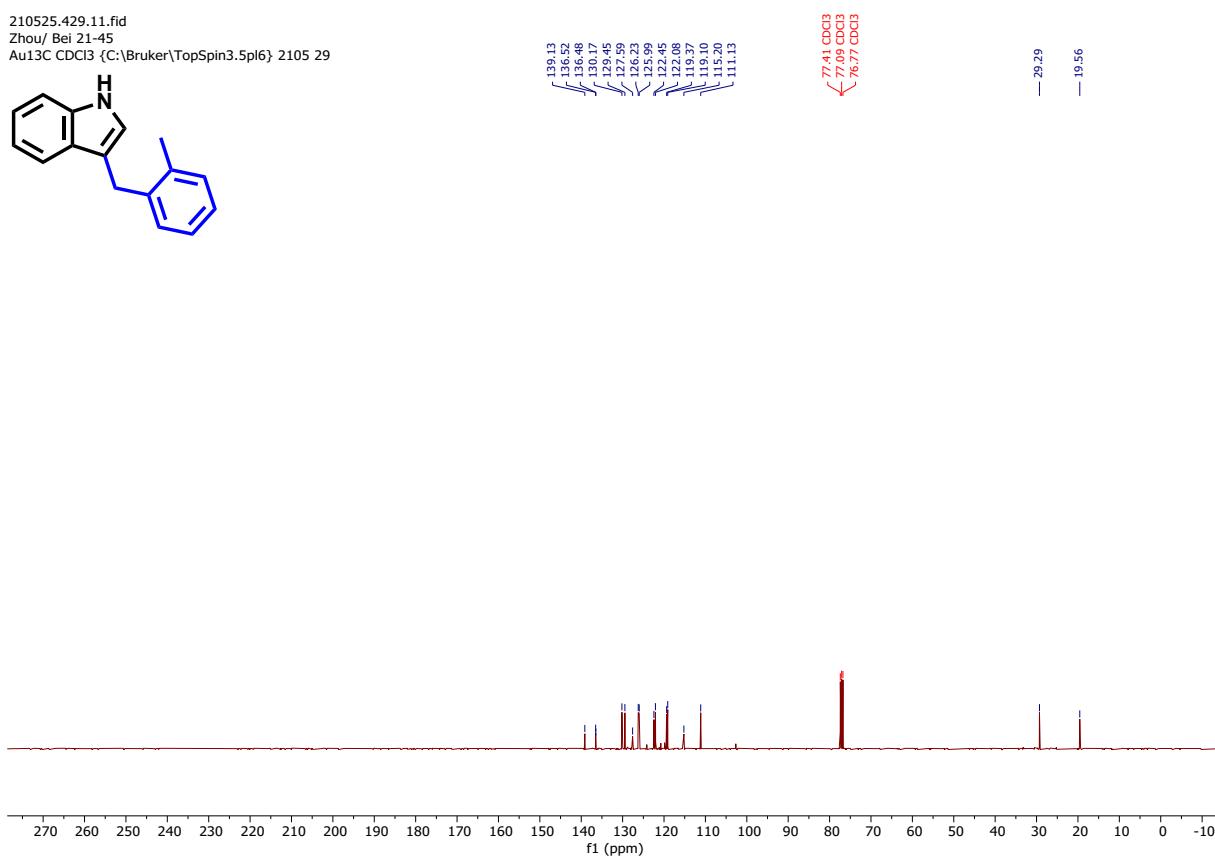
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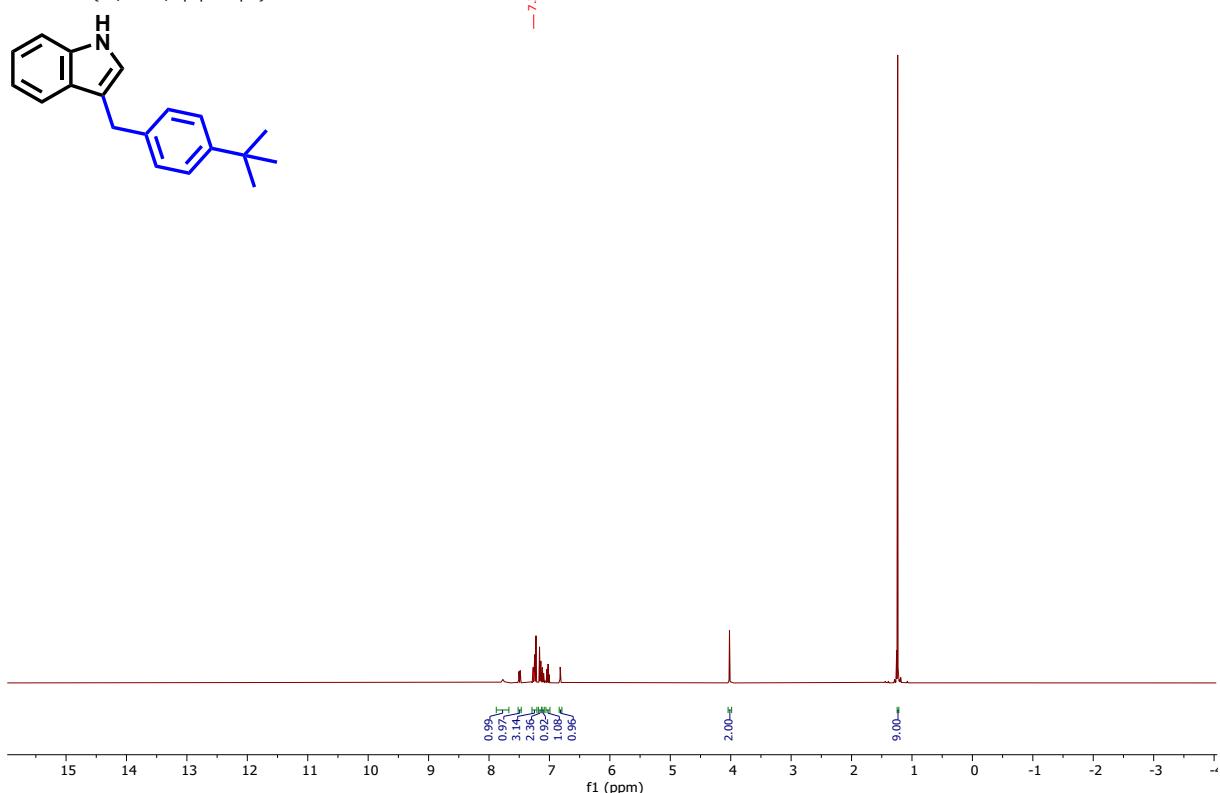
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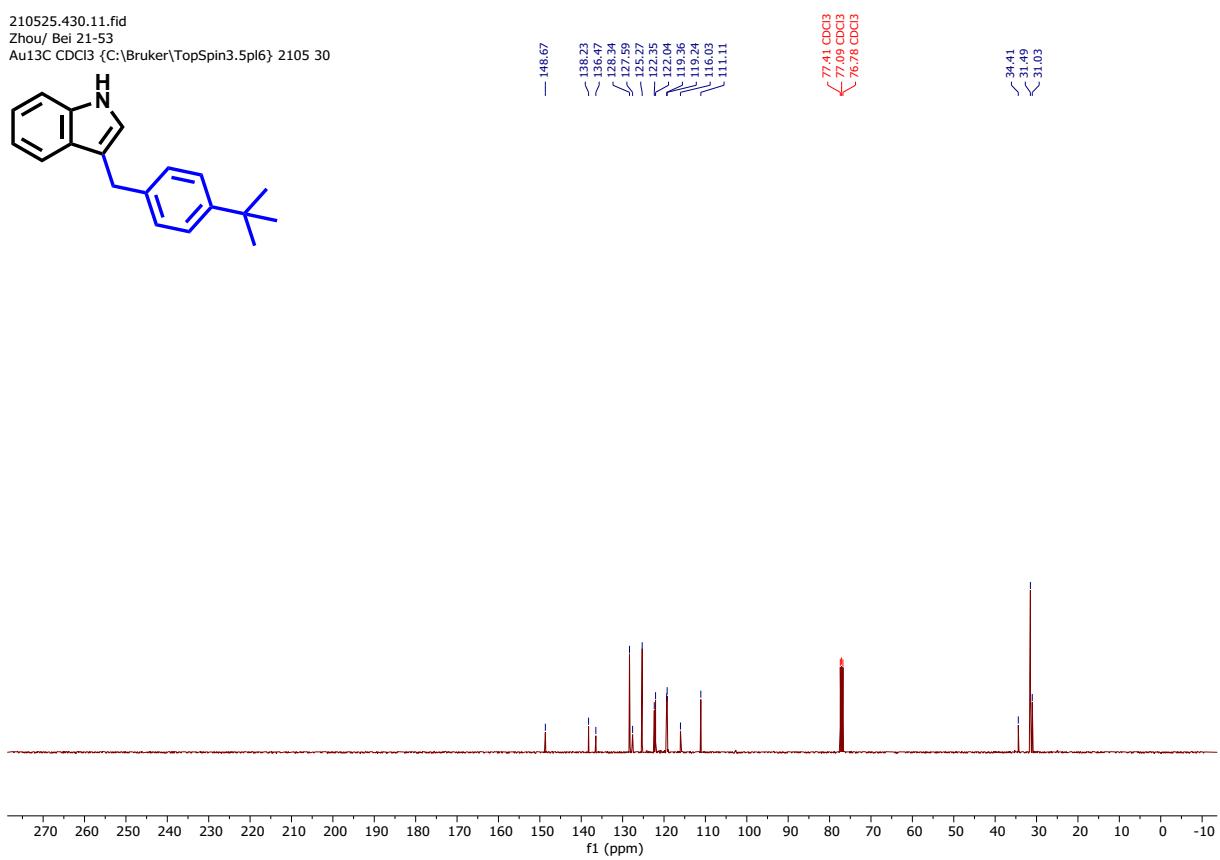
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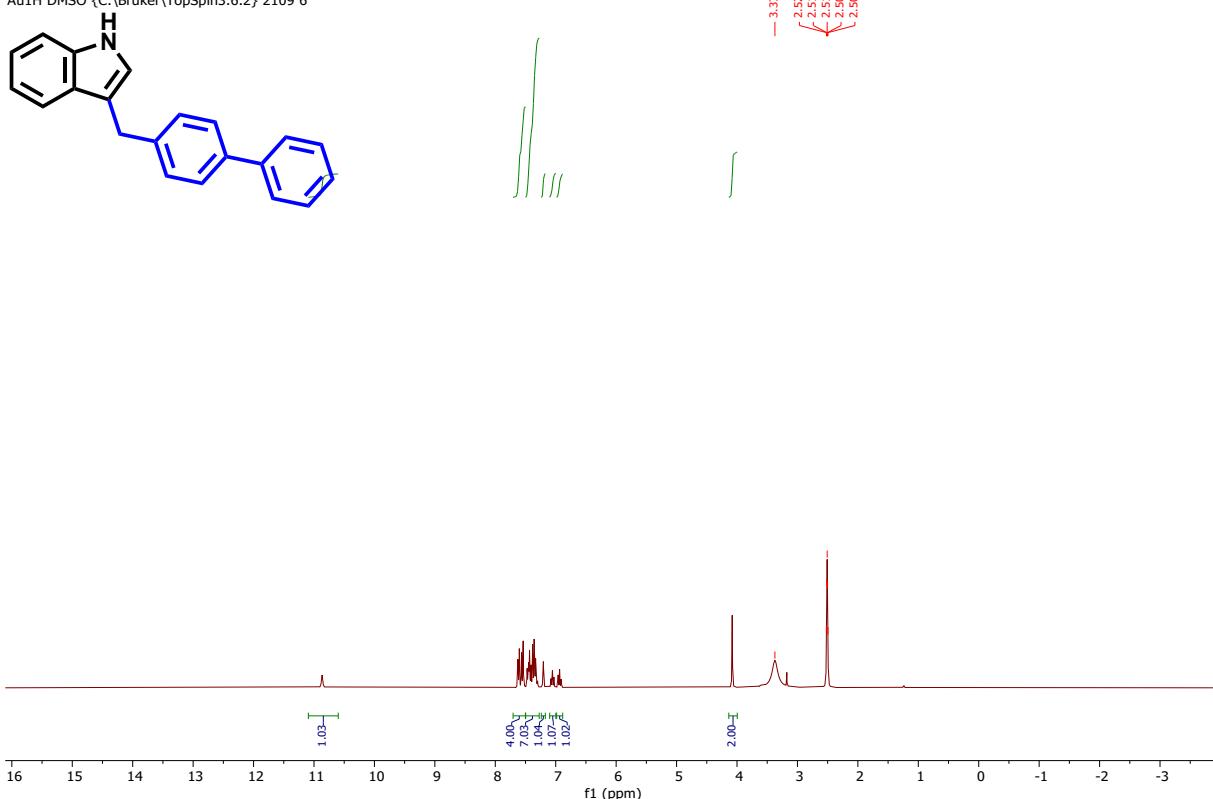
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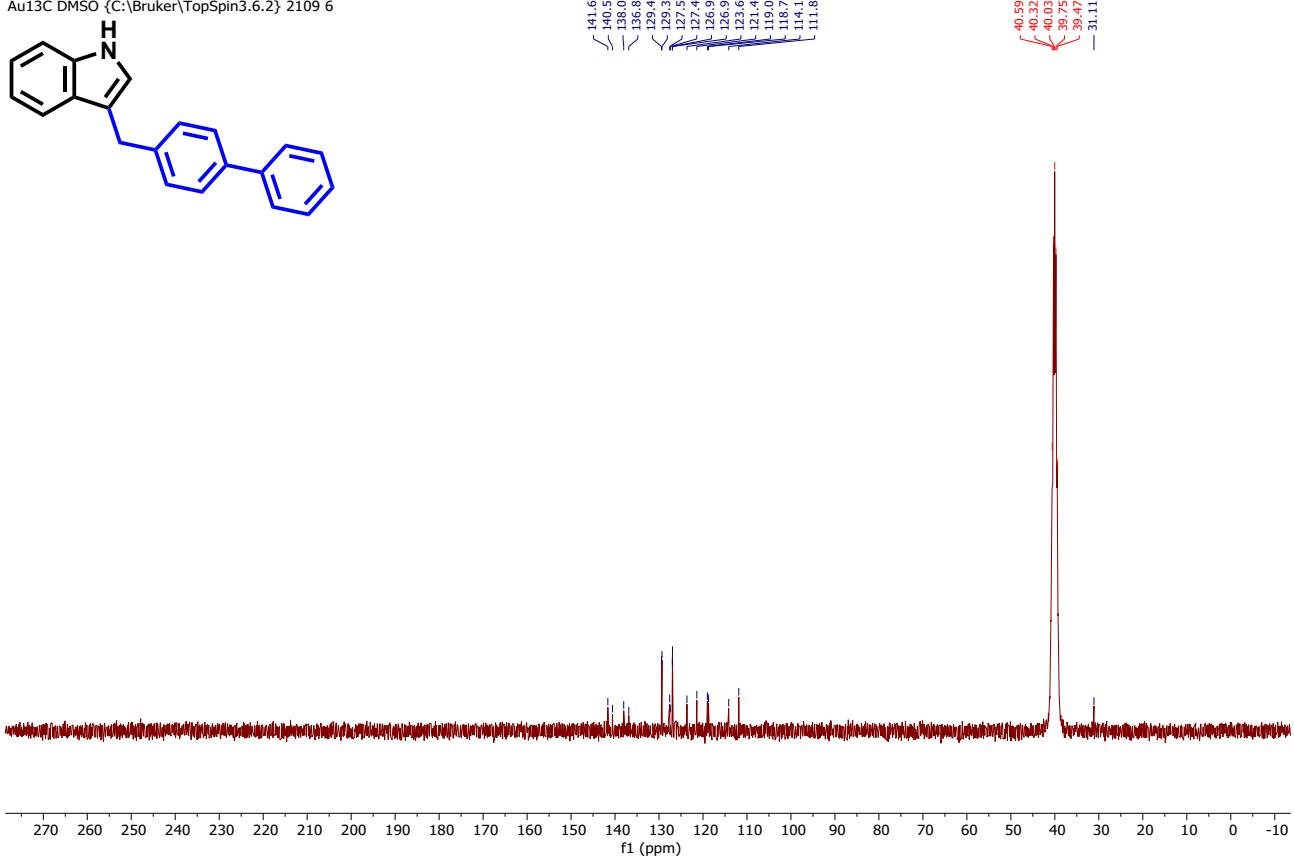
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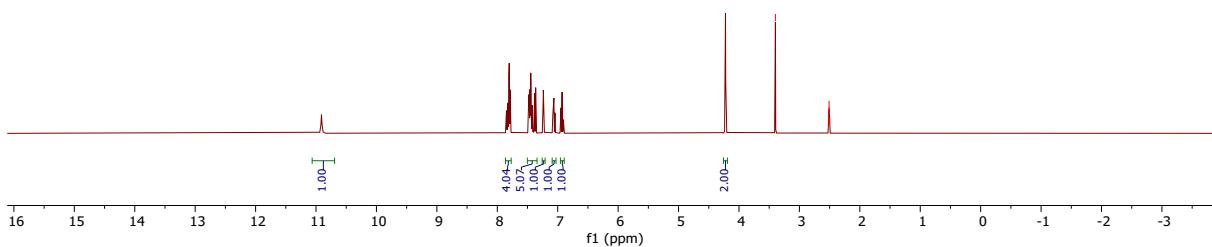
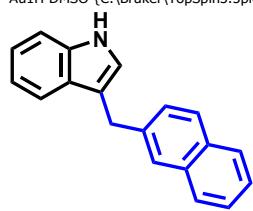
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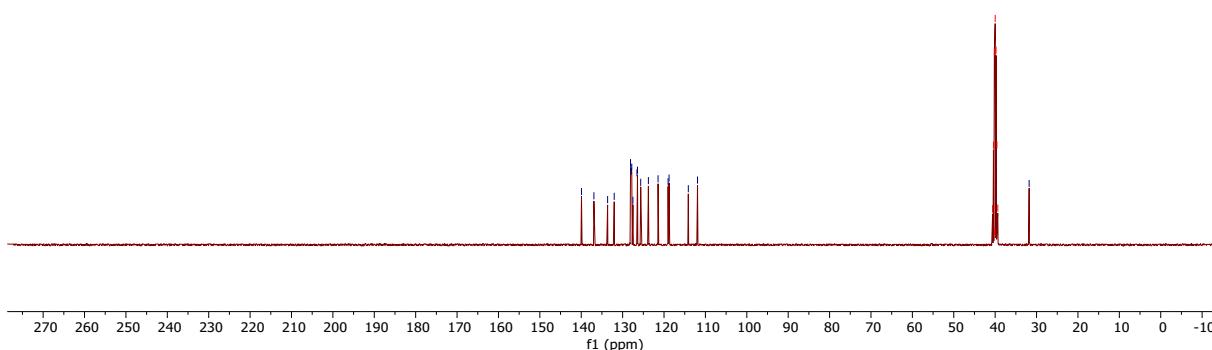
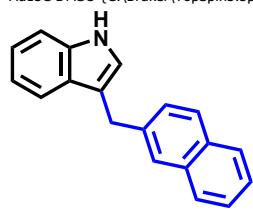
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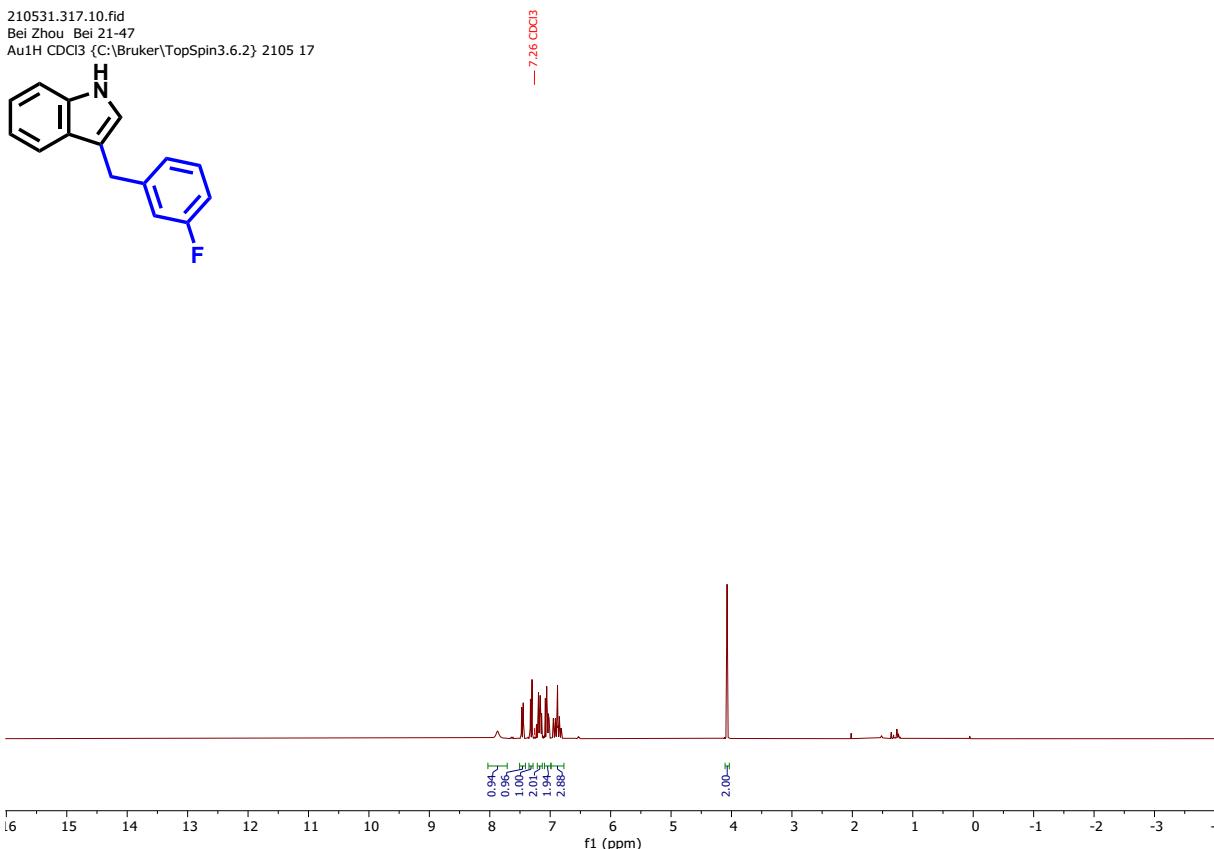
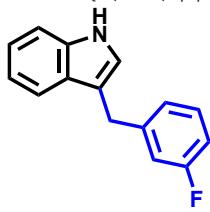
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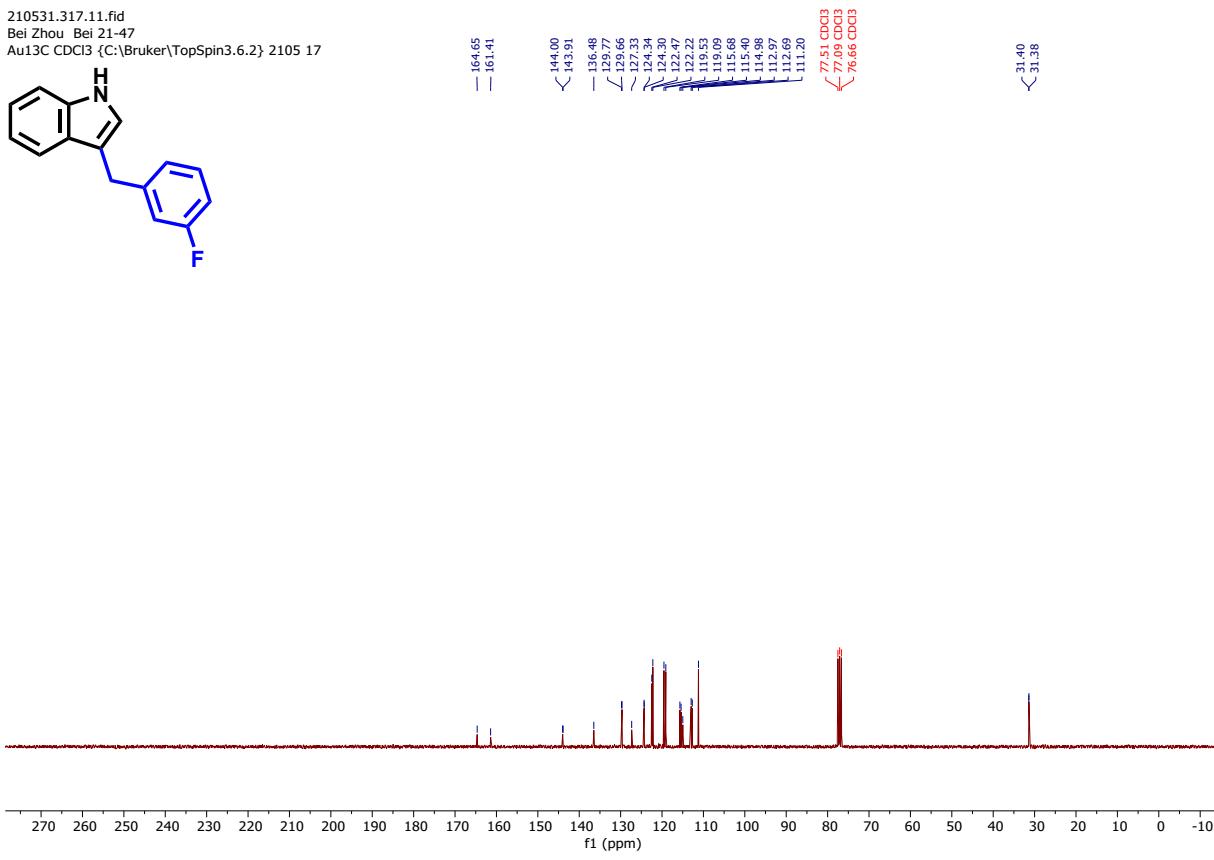
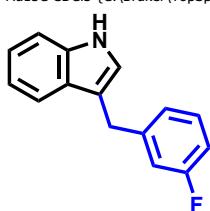
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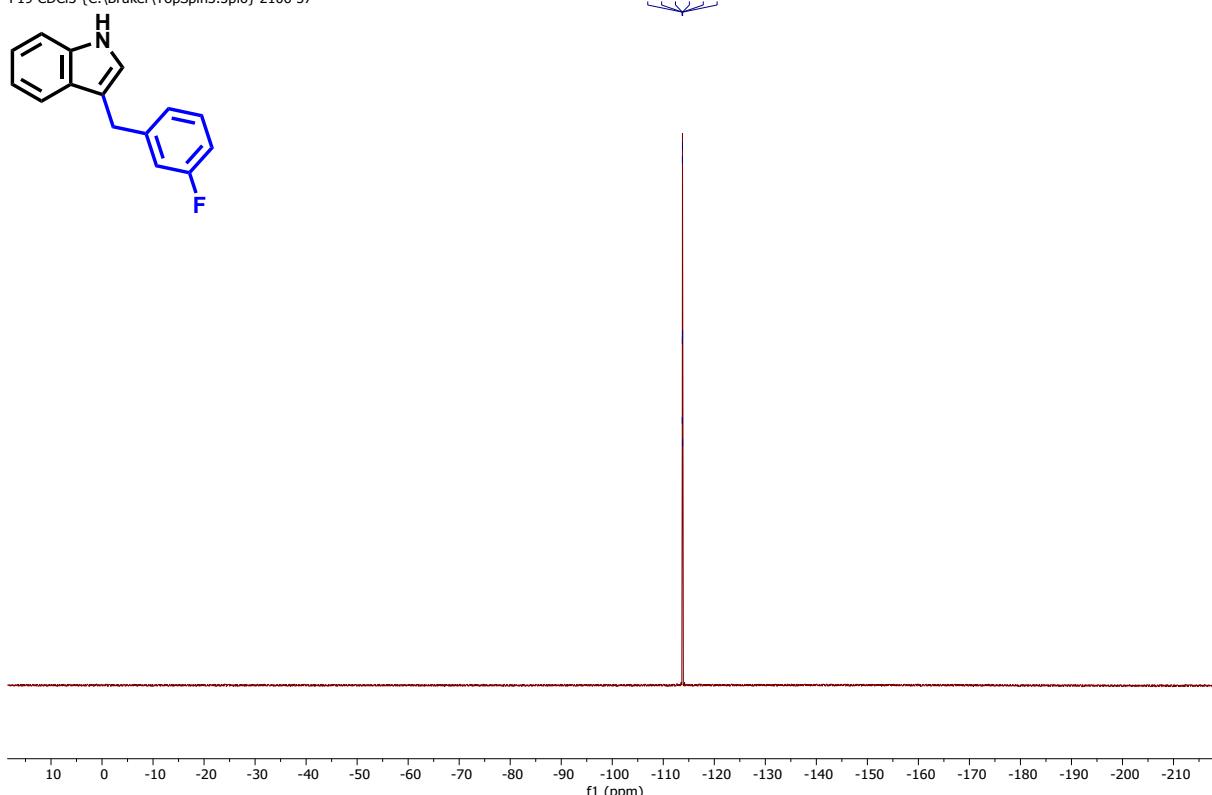
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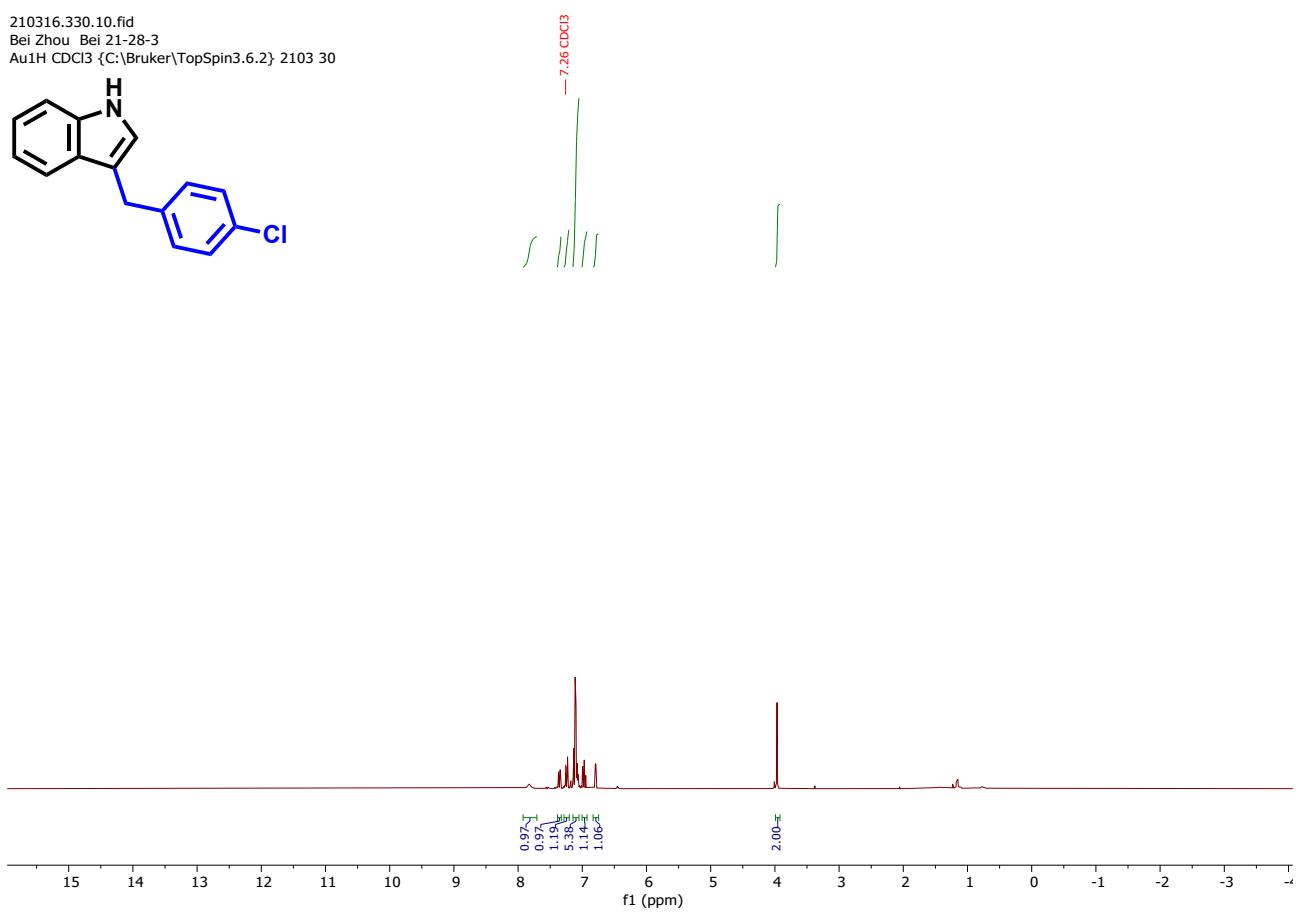
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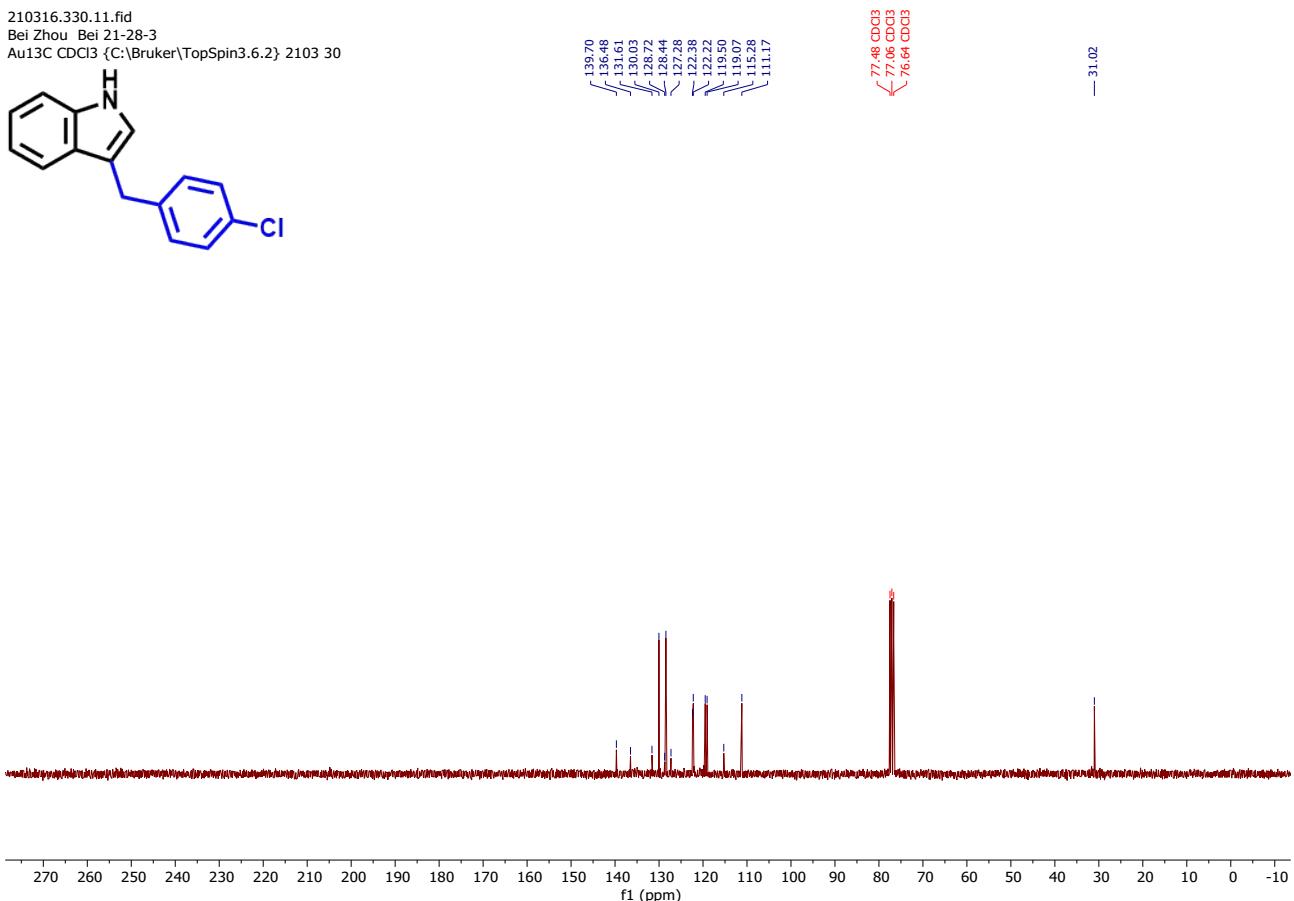
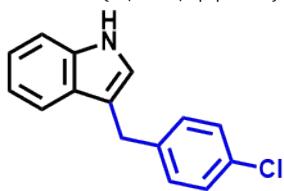
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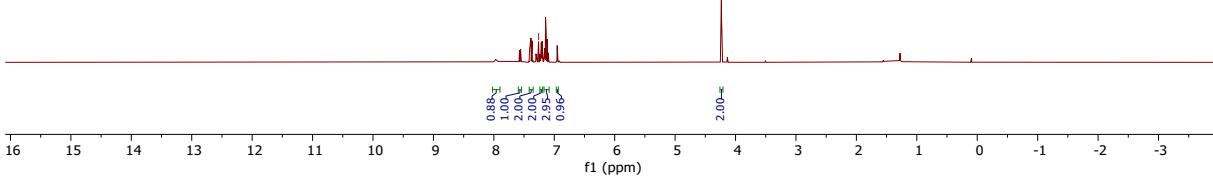
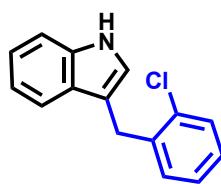
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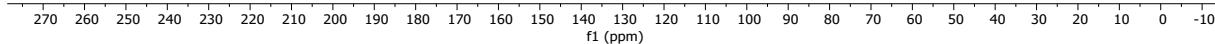
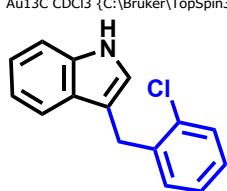
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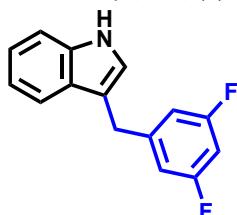
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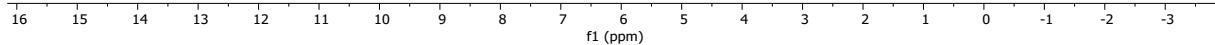
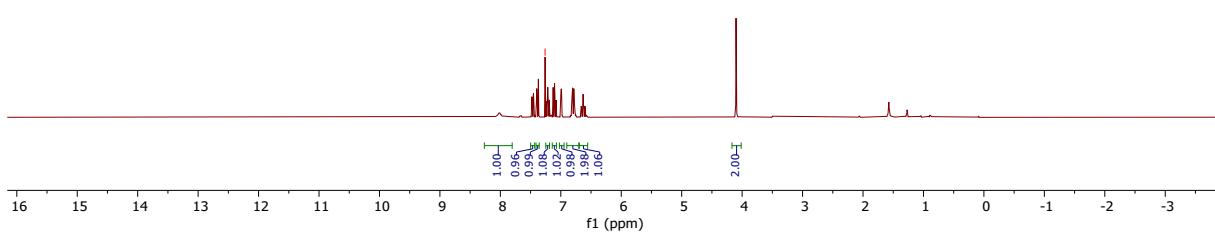
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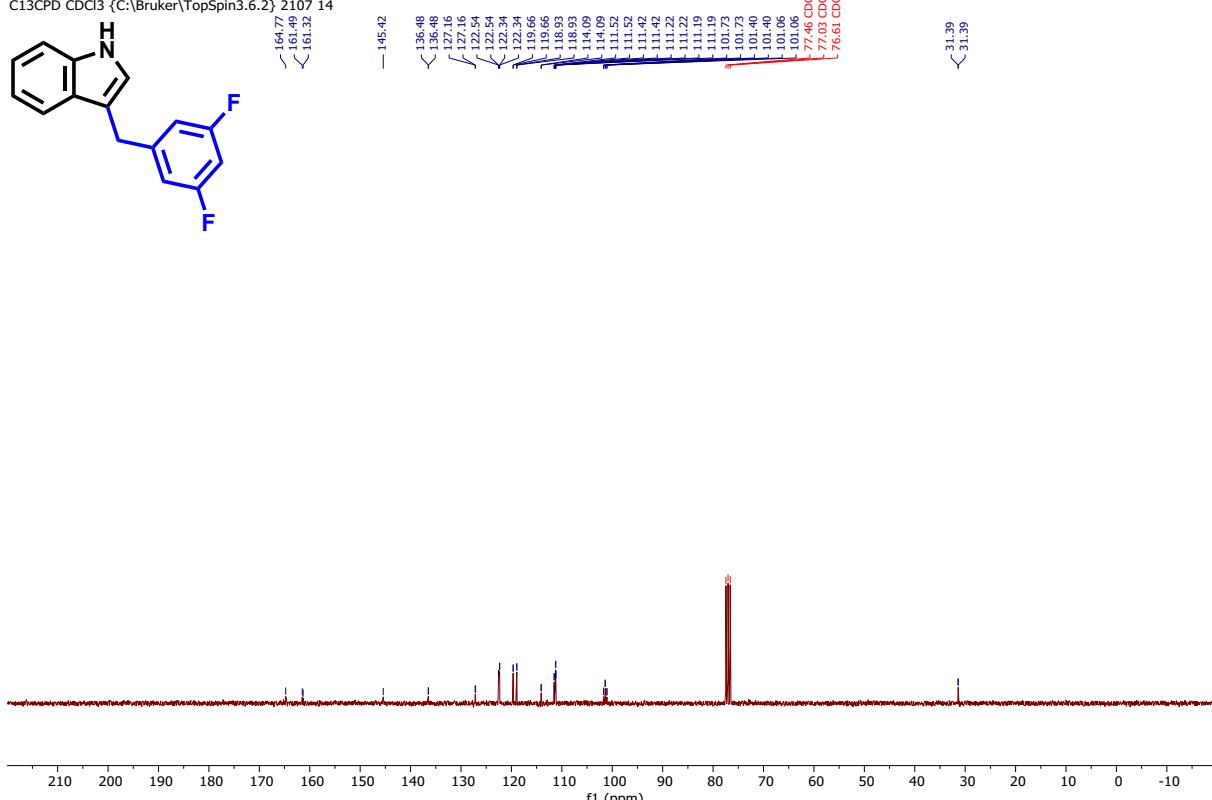
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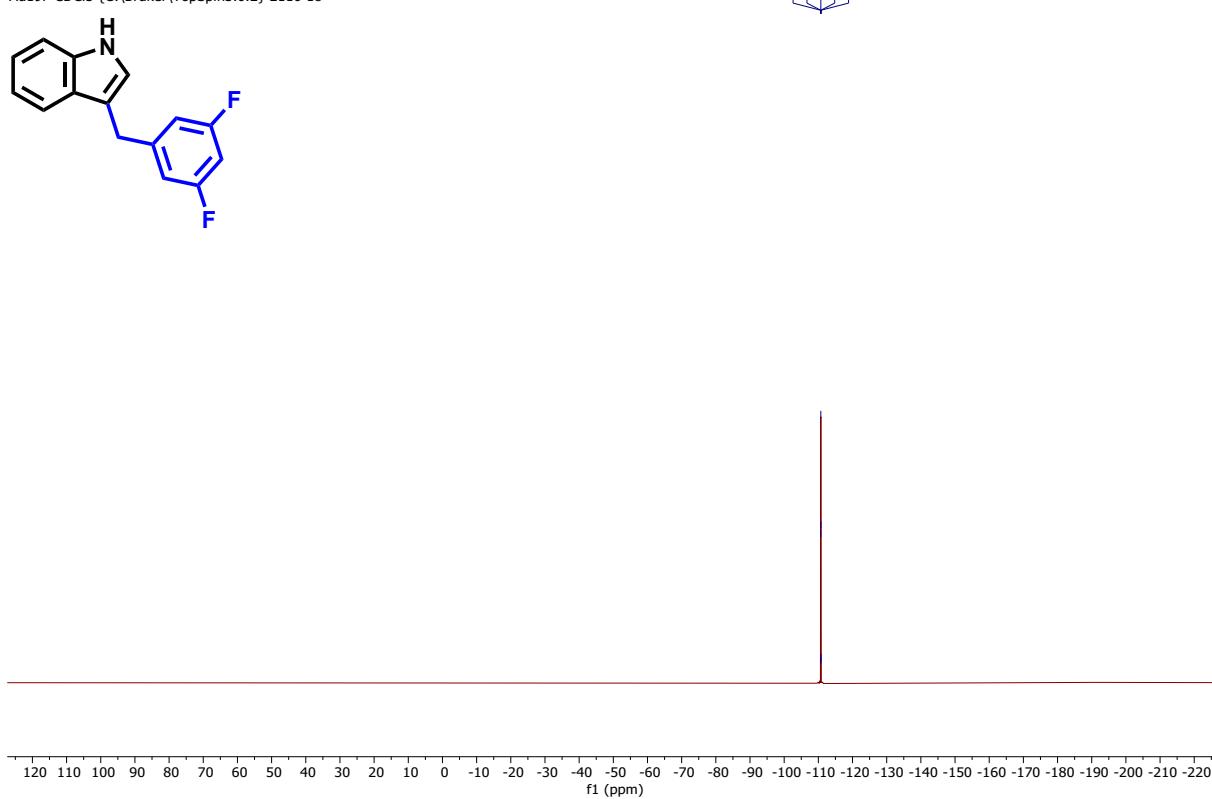
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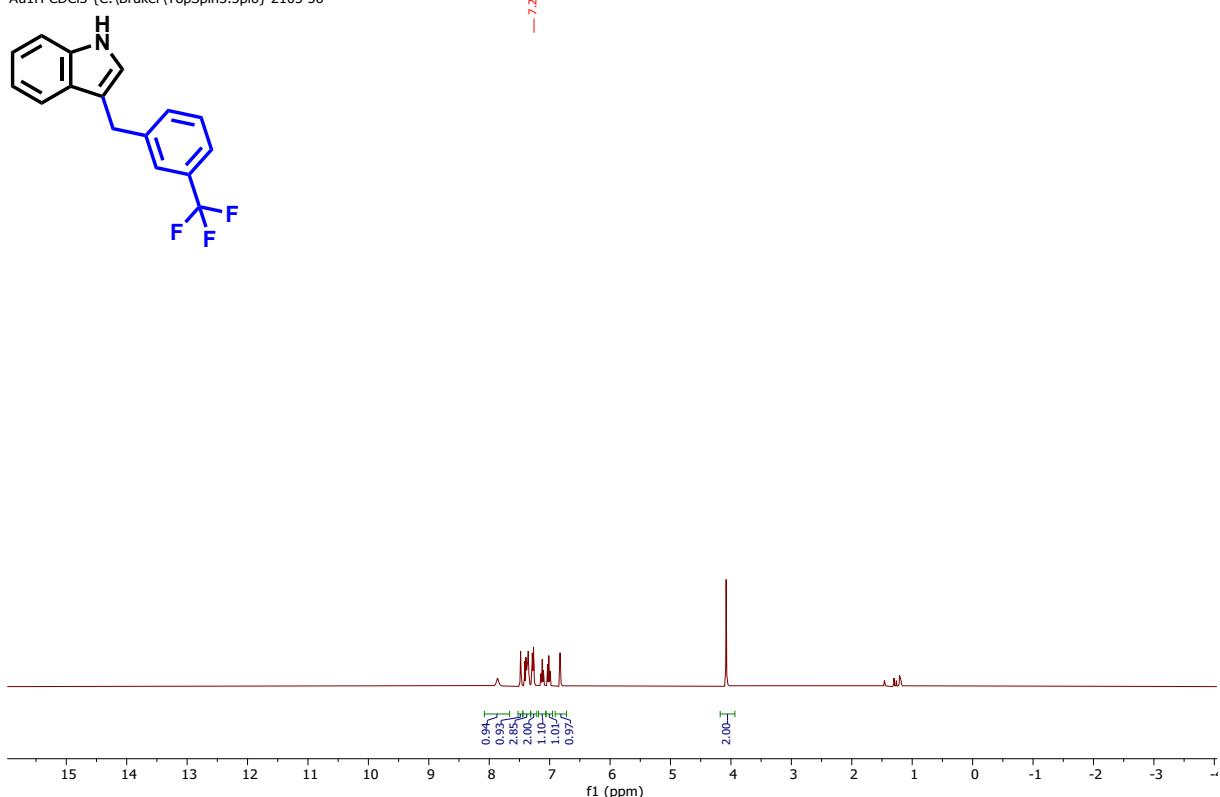
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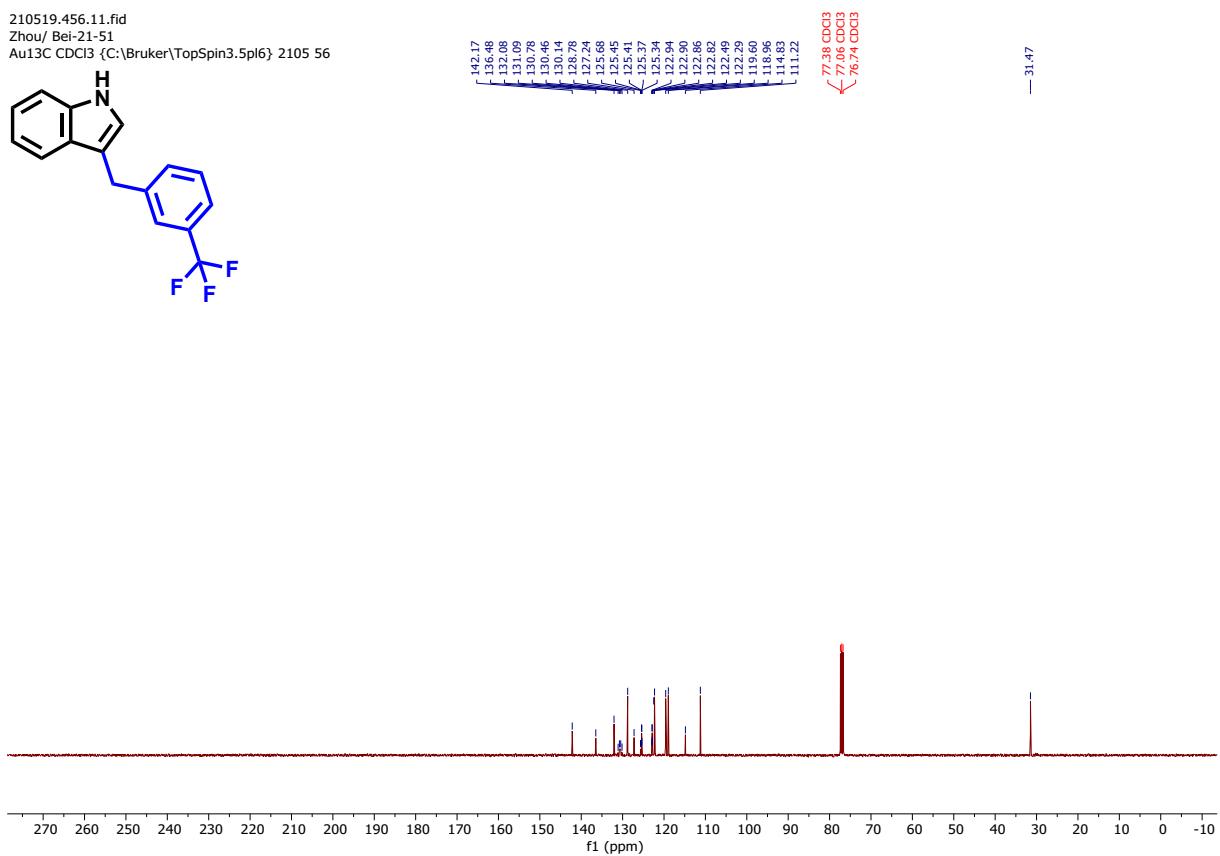
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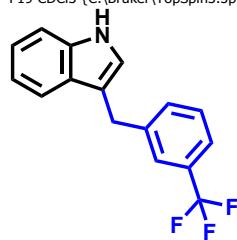
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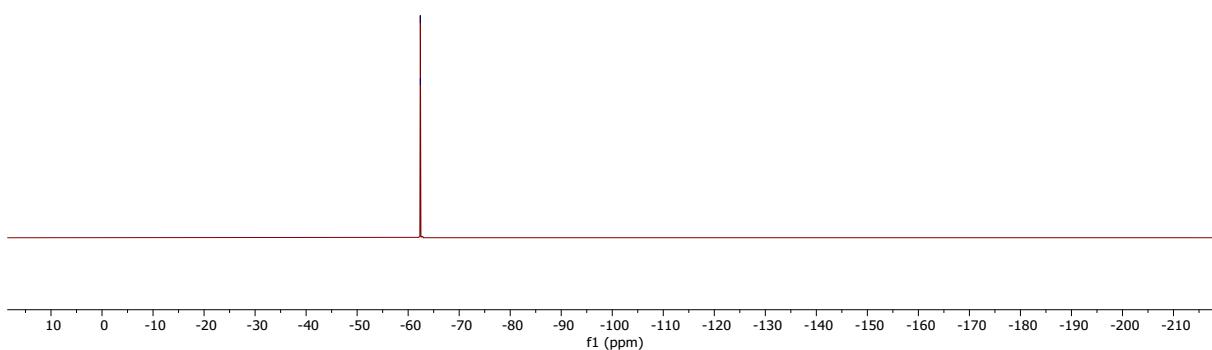
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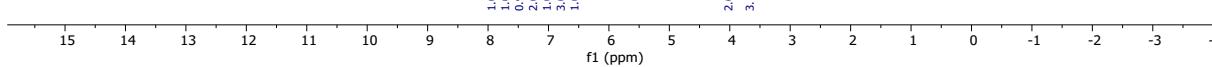
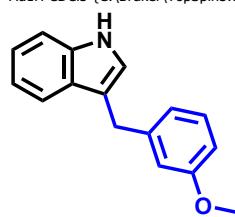
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-62.38  
-62.39

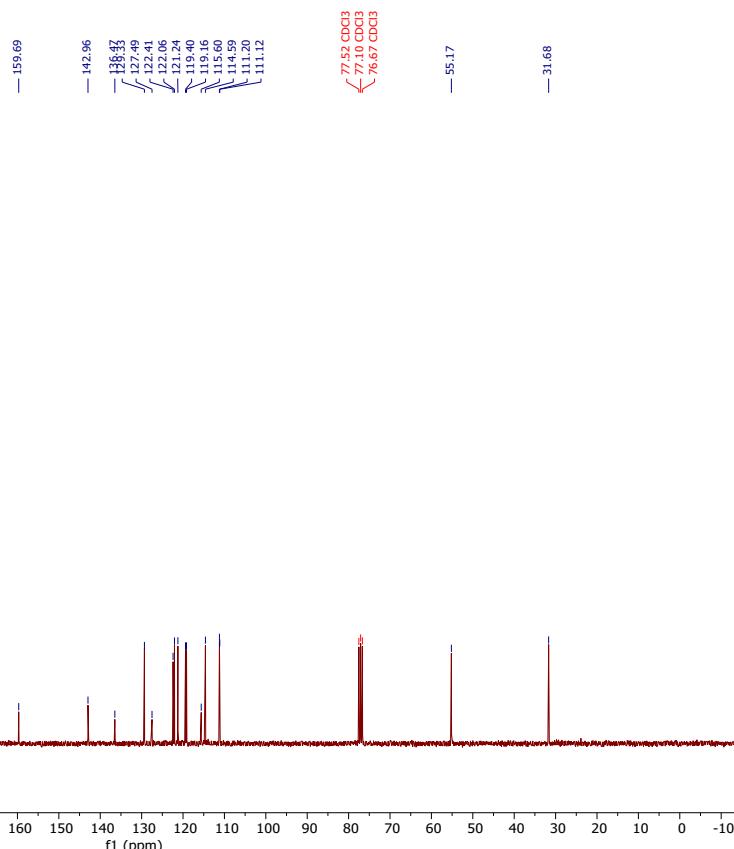
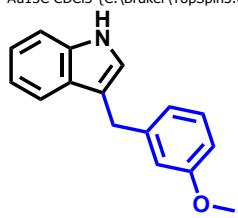


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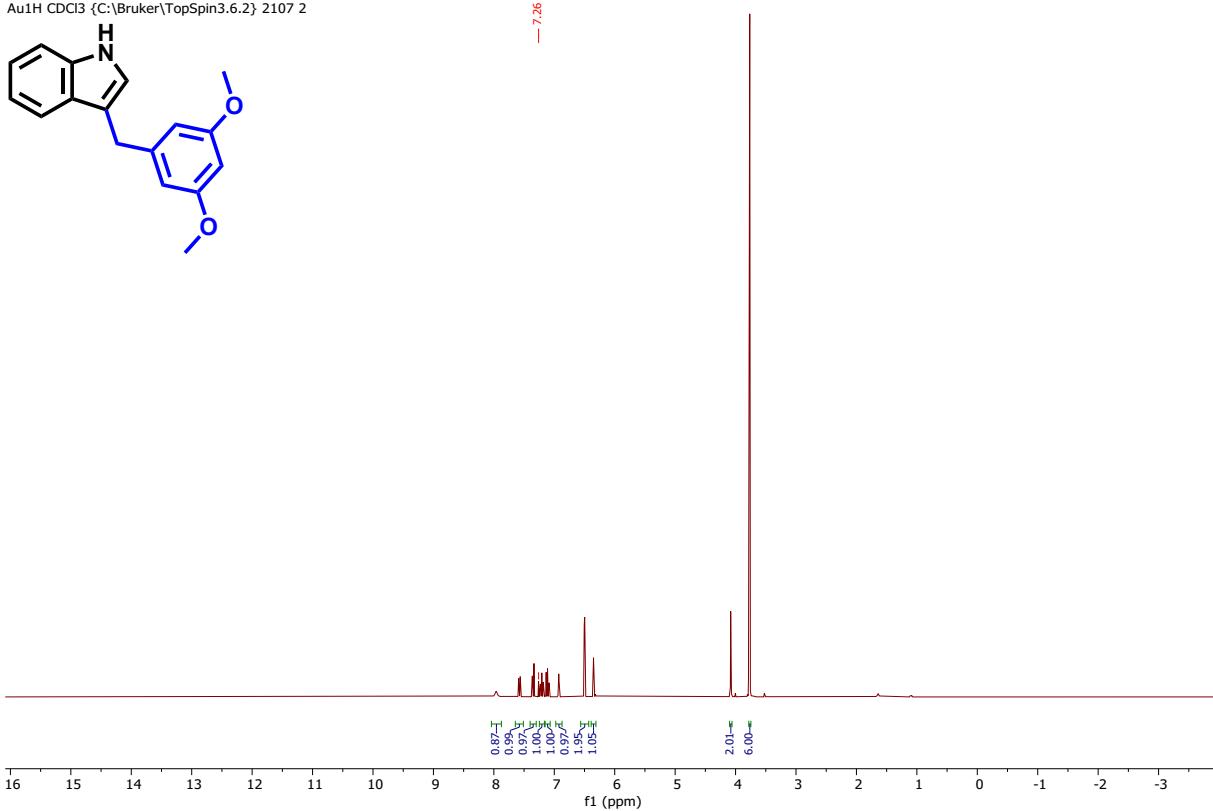
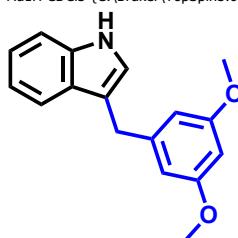


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2.09 3.11

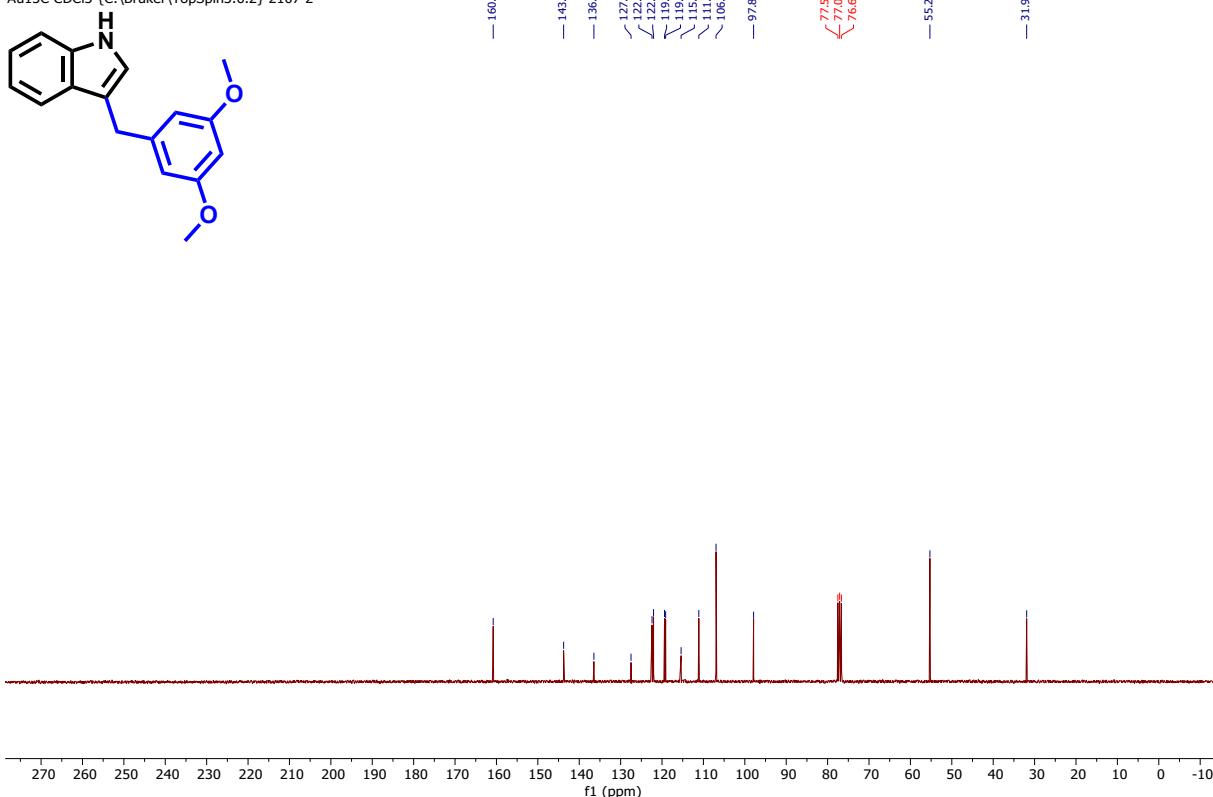
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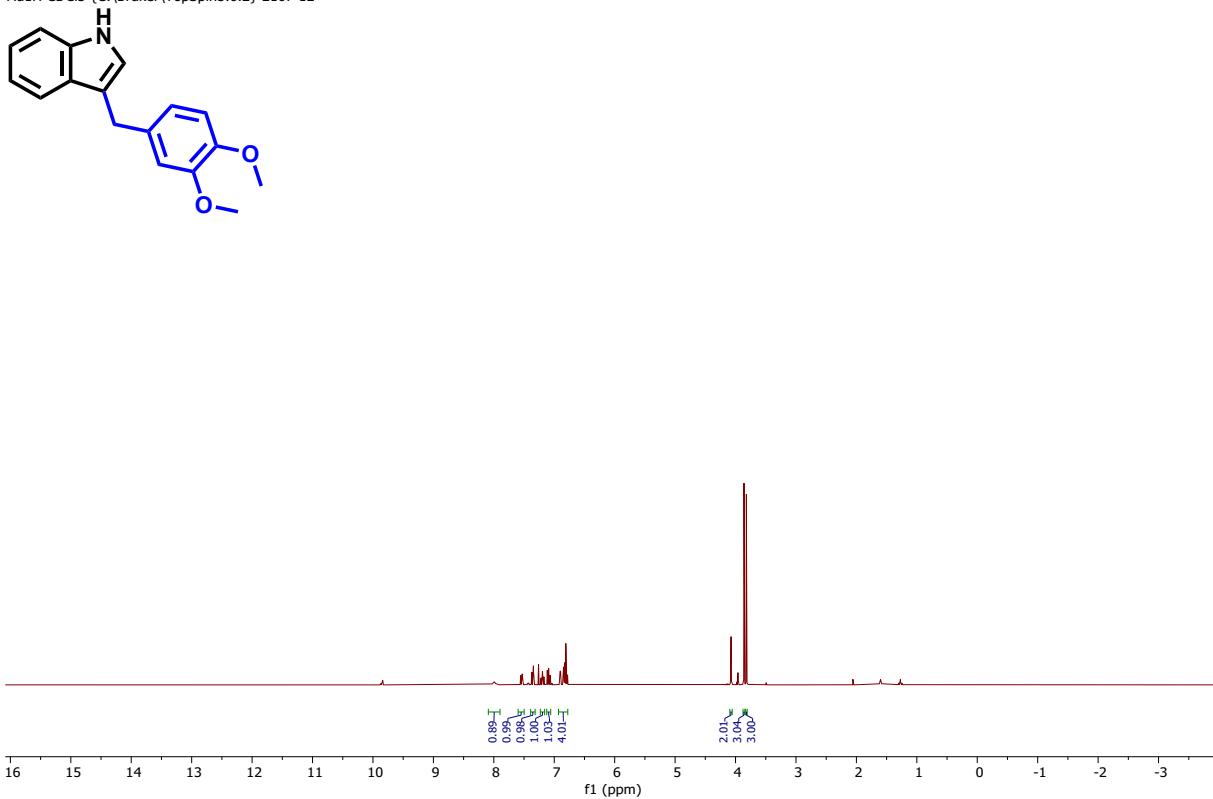
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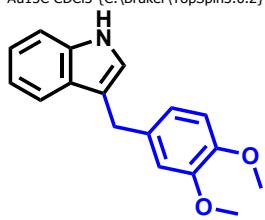
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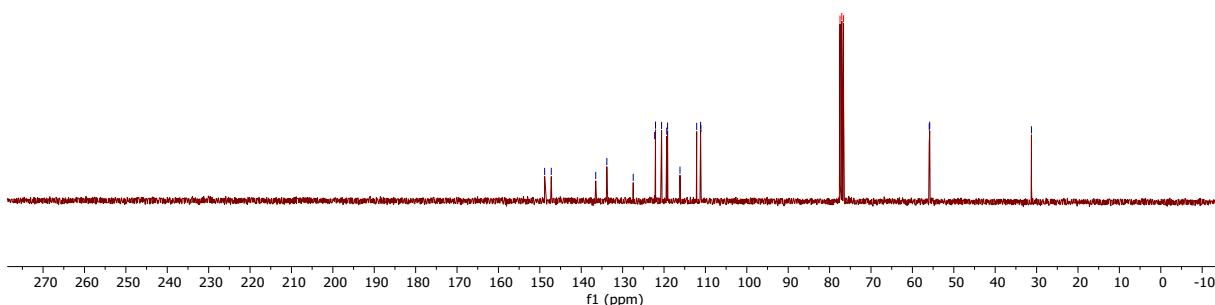
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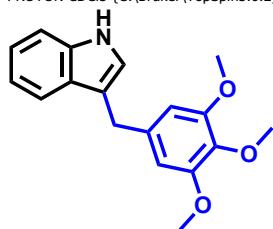
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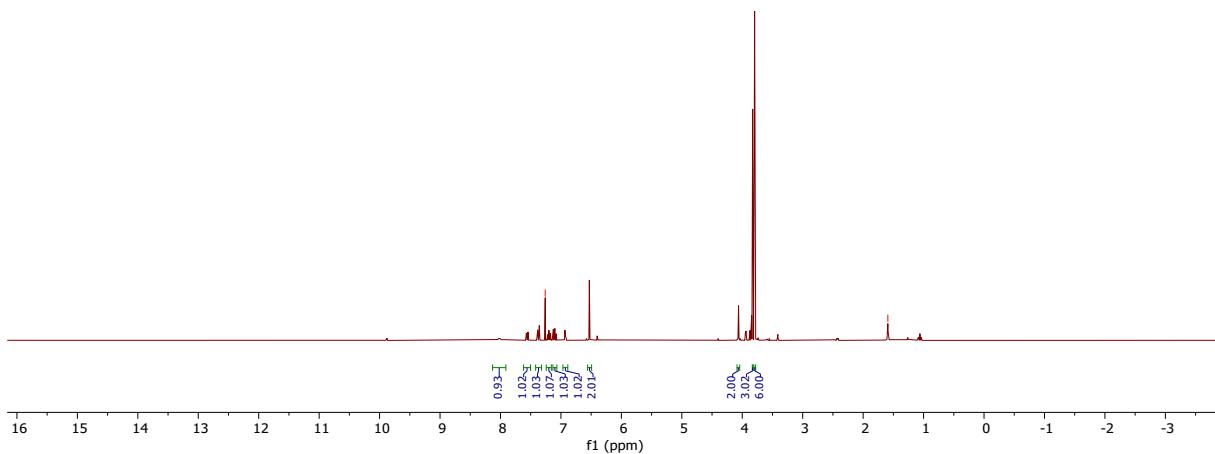
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147.24  
136.50  
133.82  
127.44  
122.26  
122.06  
120.59  
119.35  
119.14  
116.15  
112.11  
111.17  
111.10  
77.48 CDCl<sub>3</sub>  
77.06 CDCl<sub>3</sub>  
76.64 CDCl<sub>3</sub>  
55.94  
55.82  
-31.24



210809.f328.10.fid  
Bei Zhou Bei 21-99-1  
PROTON CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 28



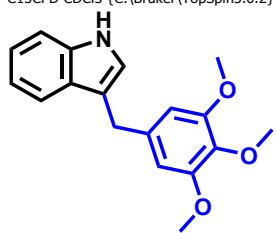
-7.26 CDCl<sub>3</sub>  
-1.59 H<sub>2</sub>O



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Bei Zhou Bei 21-99-1

C13CPD CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 28



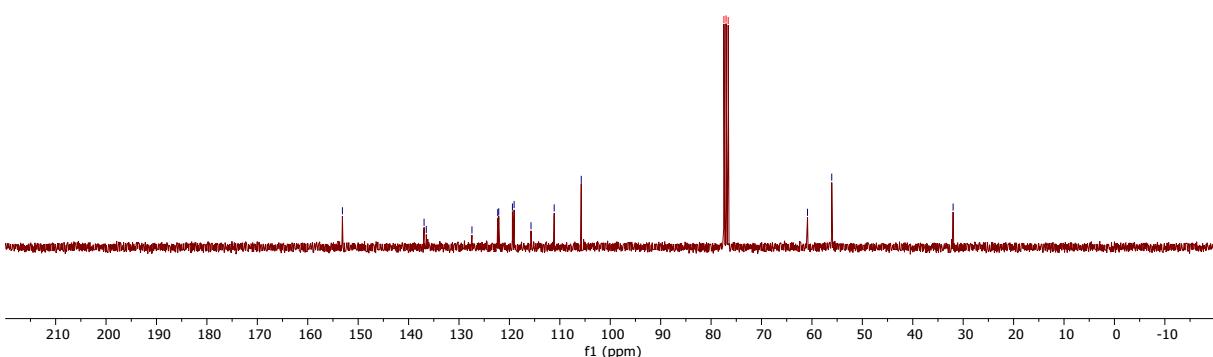
- 153.12

< 136.48  
> 127.43  
< 122.32  
< 122.11  
< 119.40  
< 119.07  
< 115.72  
> 111.12  
> 105.75

7745 CDCl<sub>3</sub>  
7703 CDCl<sub>3</sub>  
7661 CDCl<sub>3</sub>

- 60.88  
— 56.07

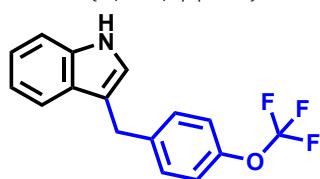
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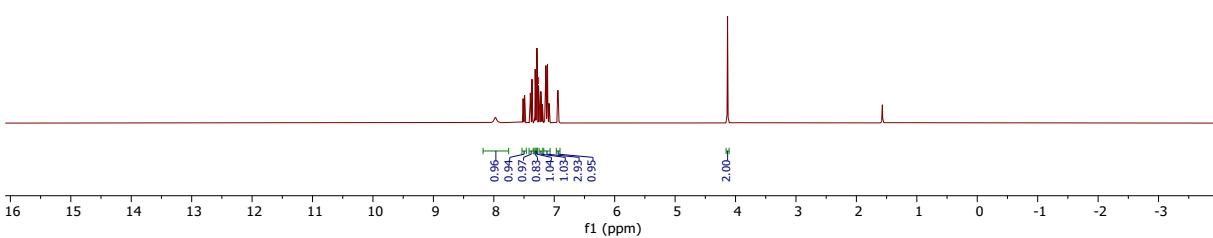
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Bei Zhou Bei 21-94-3

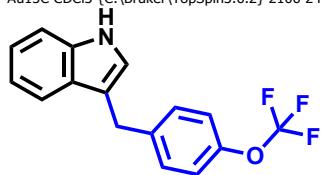
Au1H CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2106 24



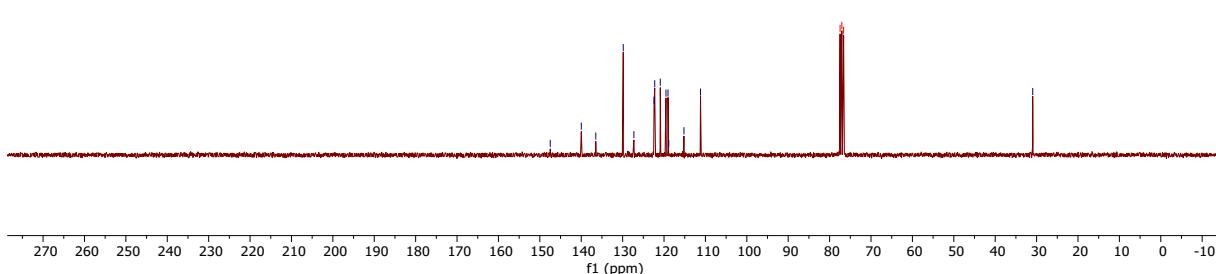
- 7.26 CDCl<sub>3</sub>



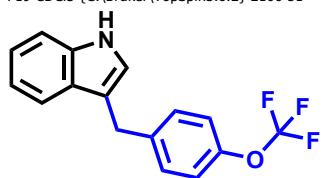
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Au13C CDCl3 {C:\Bruker\TopSpin3.6.2} 2106 24



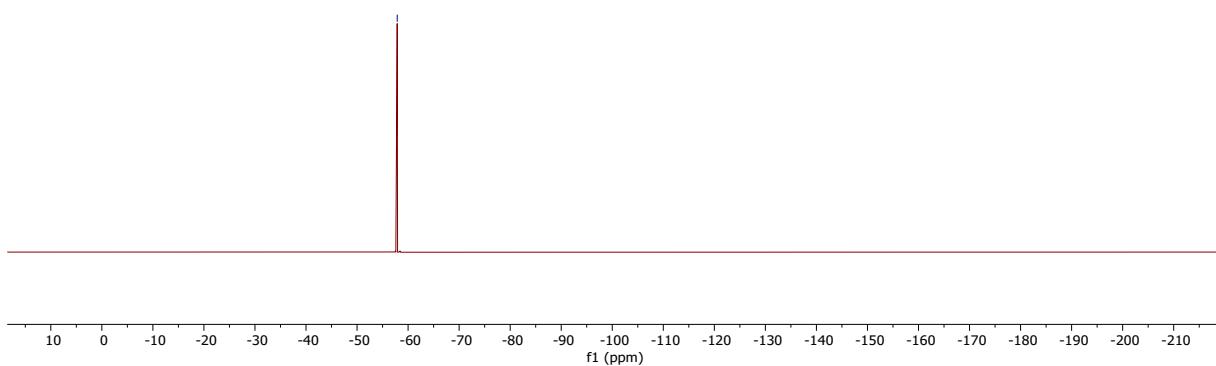
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-139.99  
-136.49  
-129.85  
-127.28  
-122.42  
-122.25  
-120.90  
-119.53  
-119.04  
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-115.19  
-111.19  
-77.47 CDCl3  
-77.05 CDCl3  
-76.62 CDCl3  
-30.94



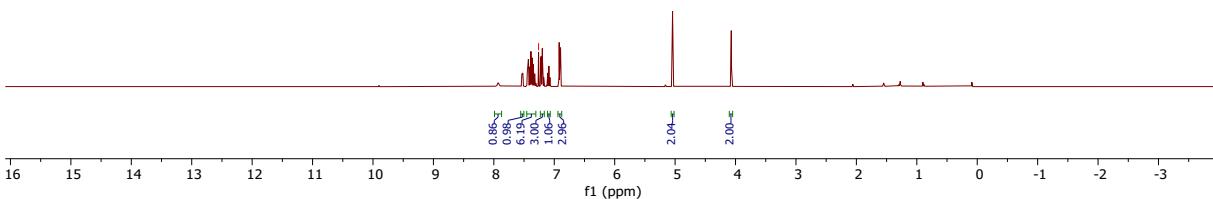
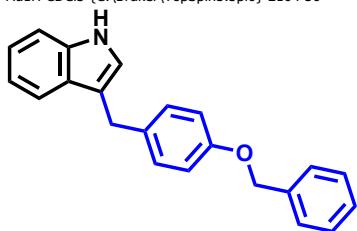
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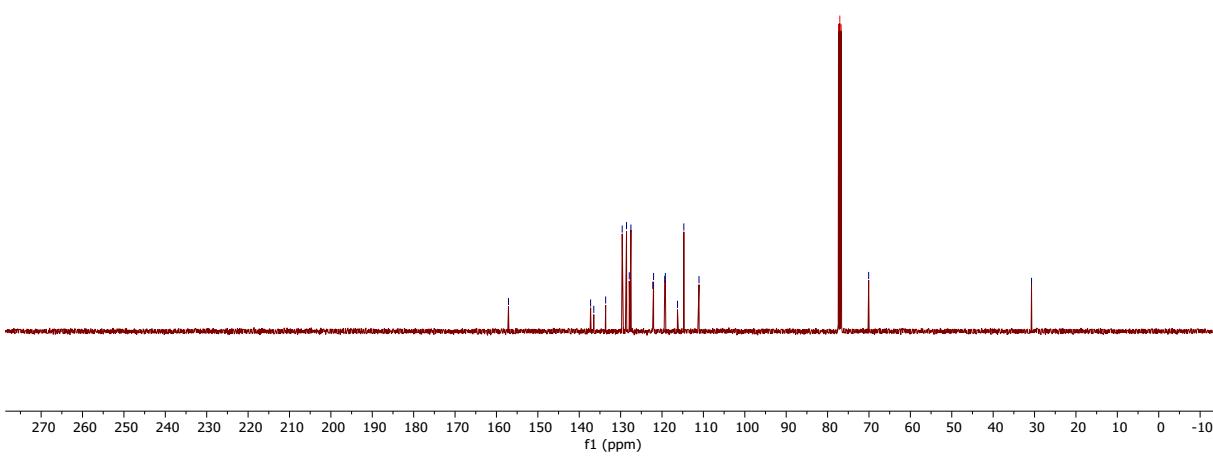
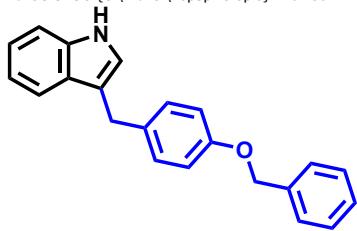
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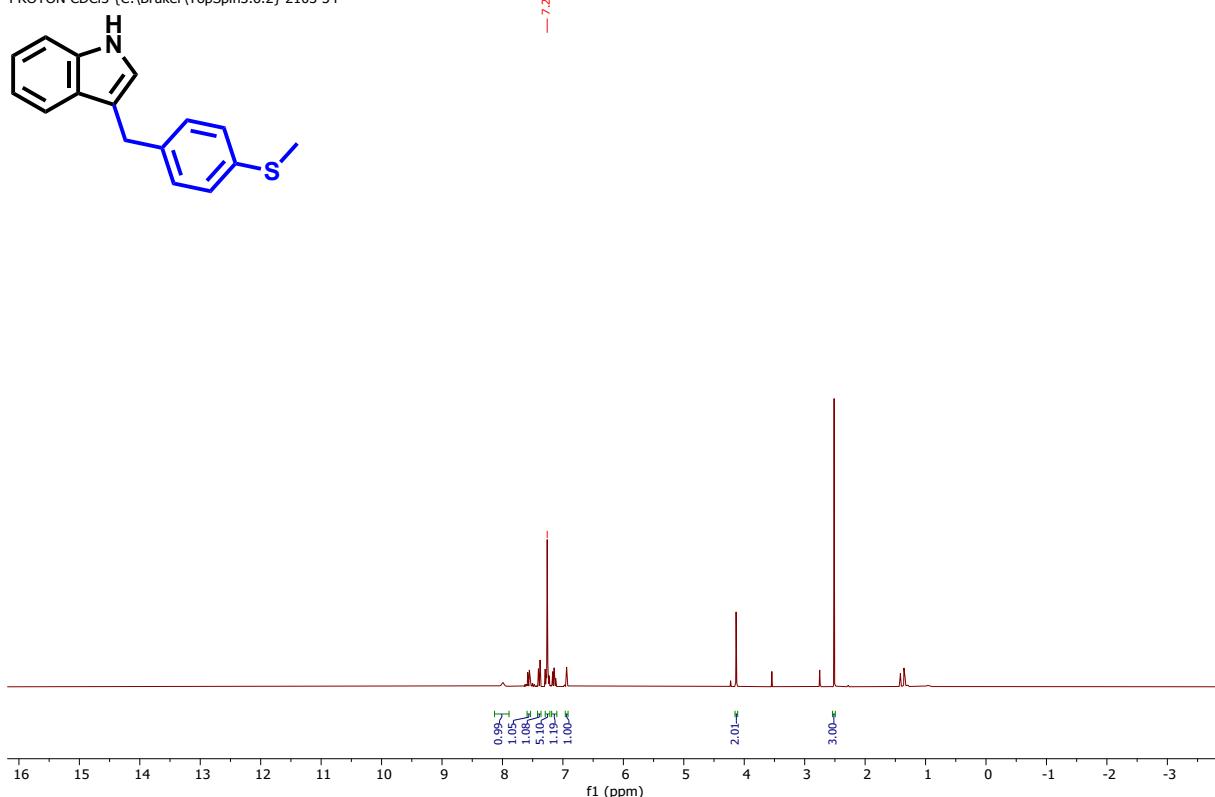
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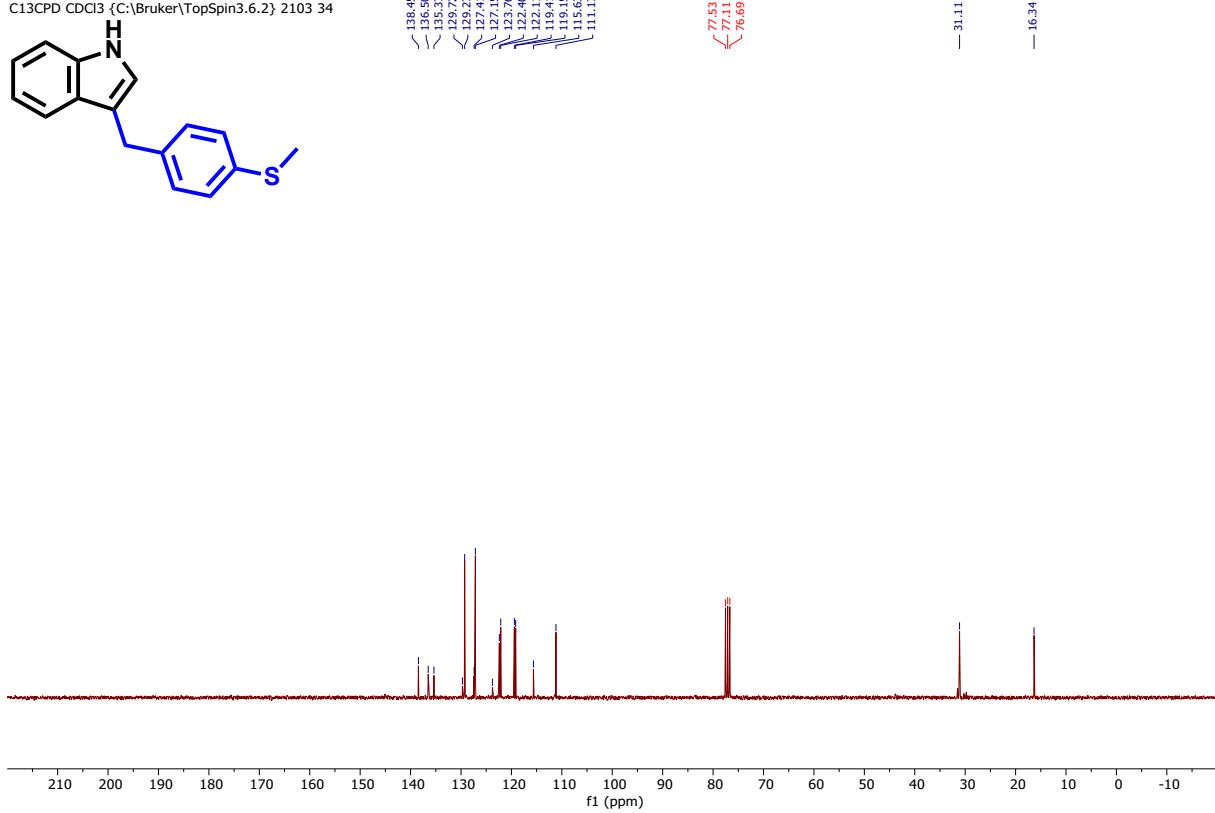
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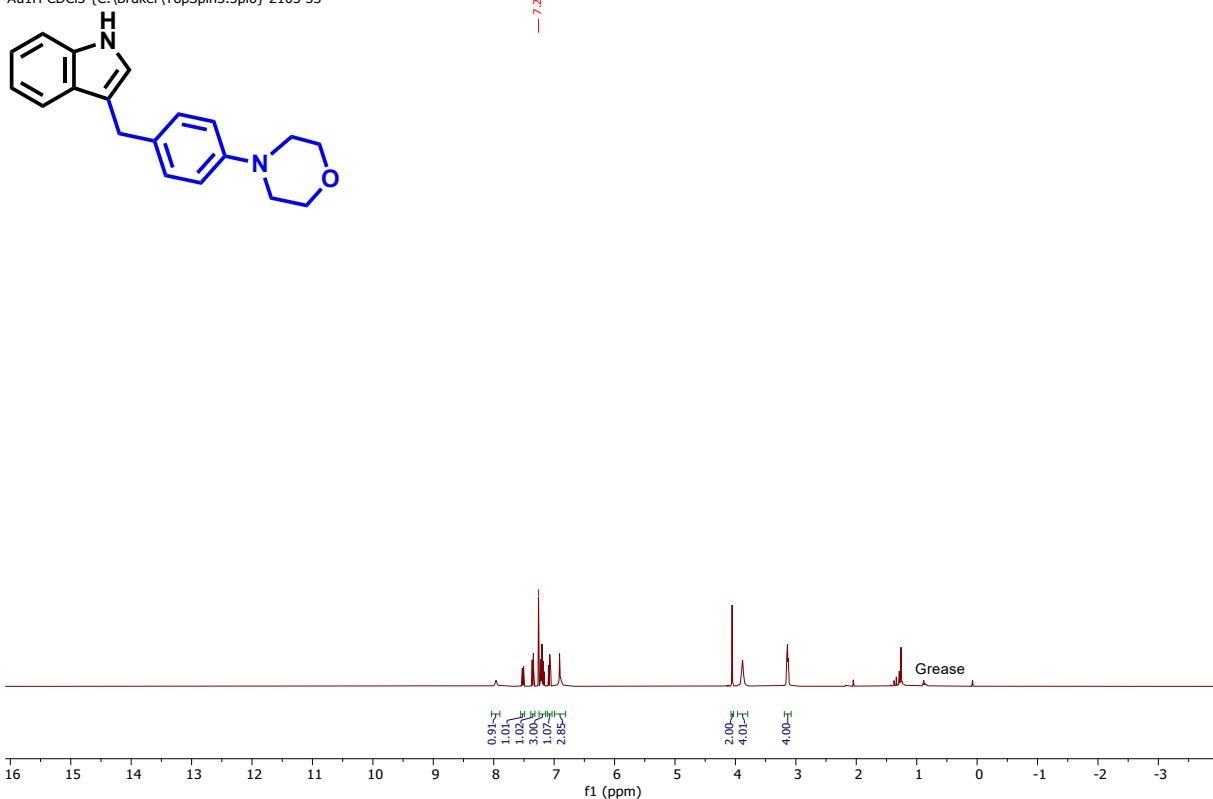
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PROTON CDCl3 {C:\Bruker\TopSpin3.6.2} 2103 34



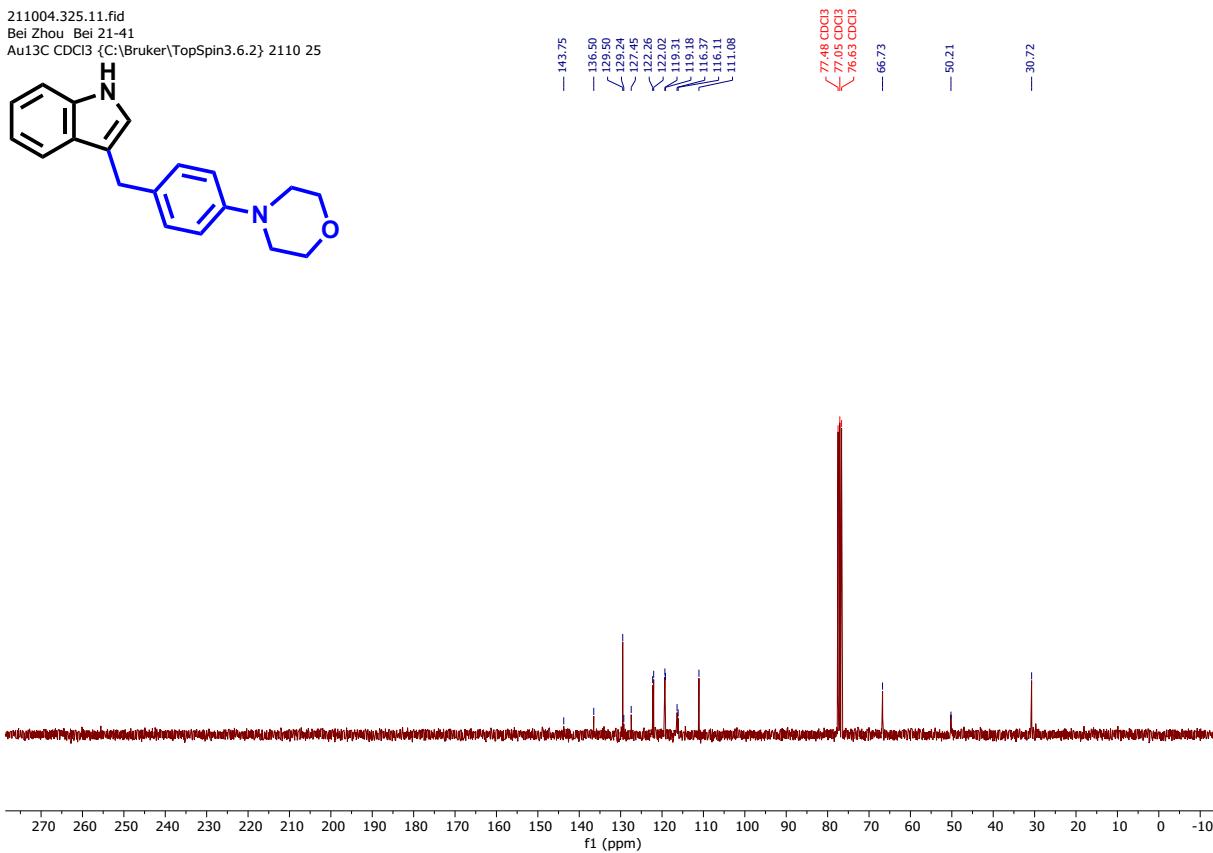
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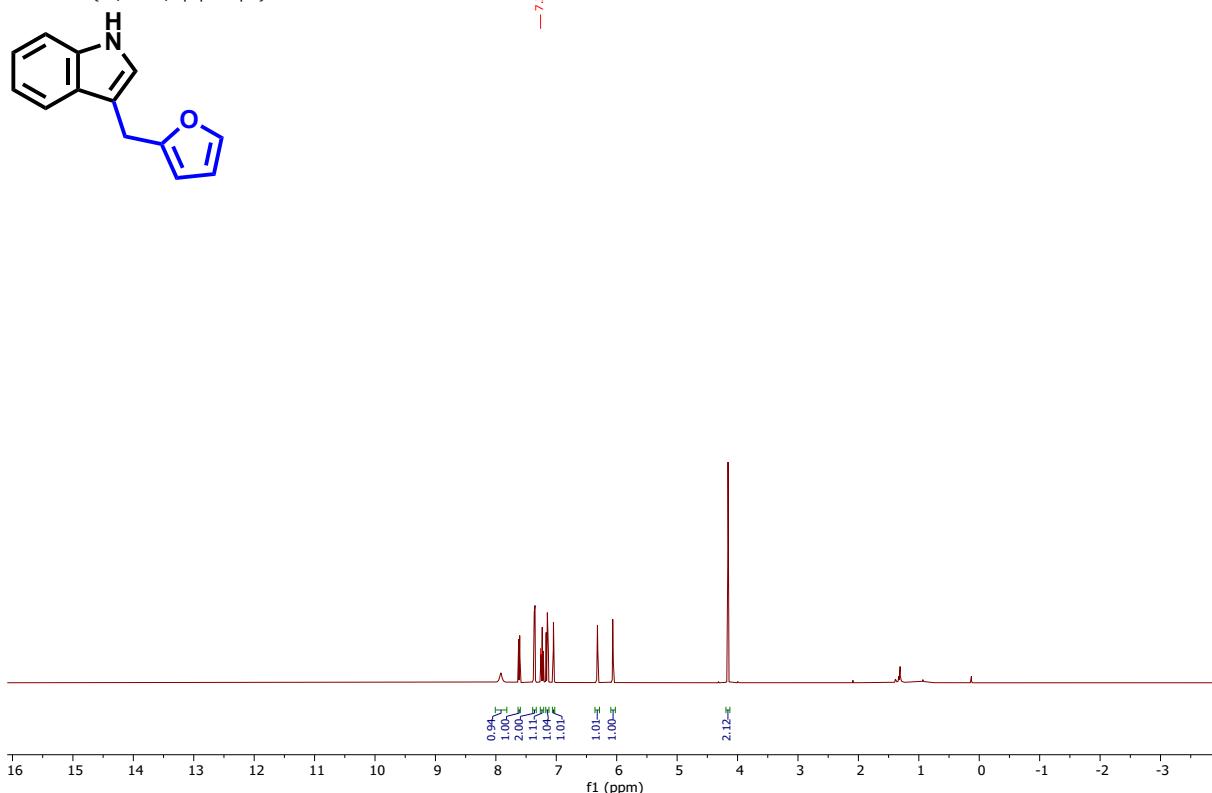
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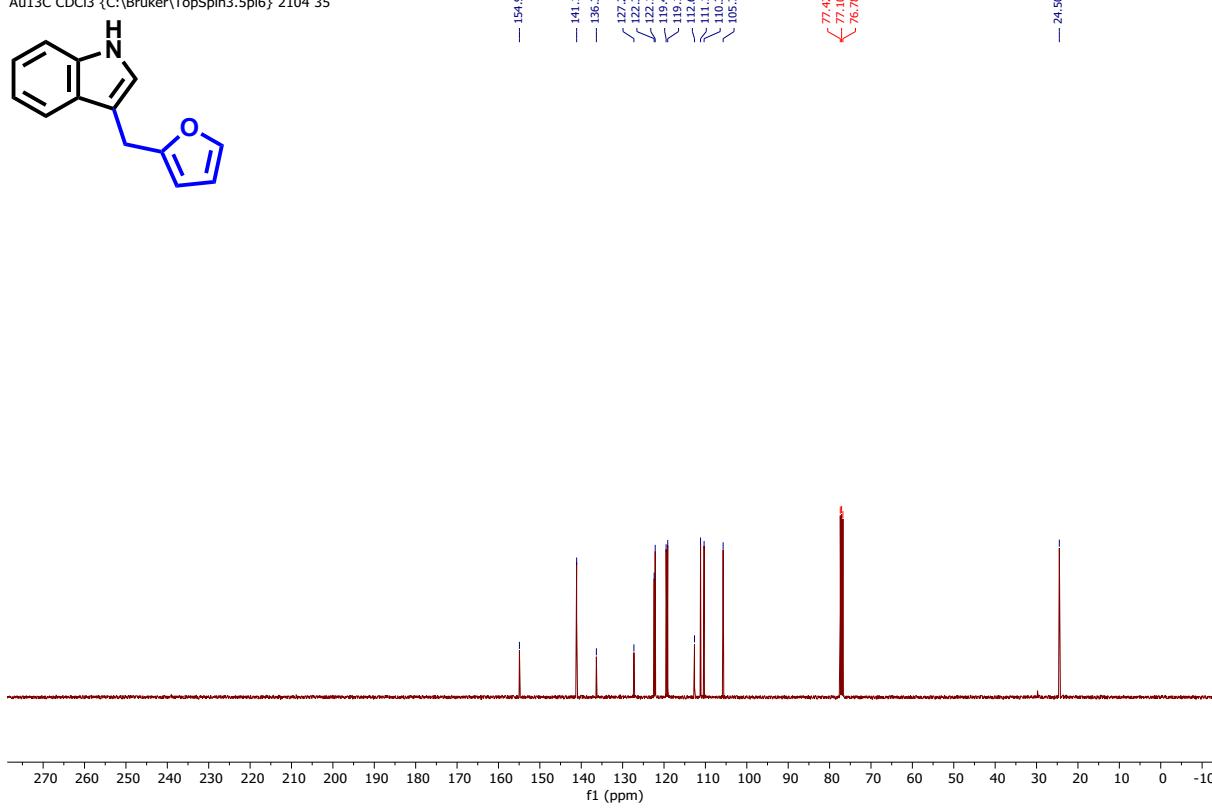
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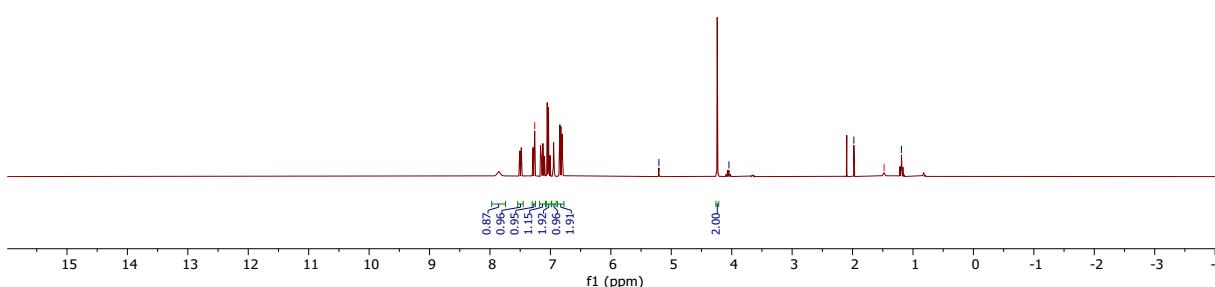
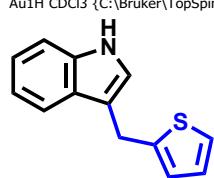
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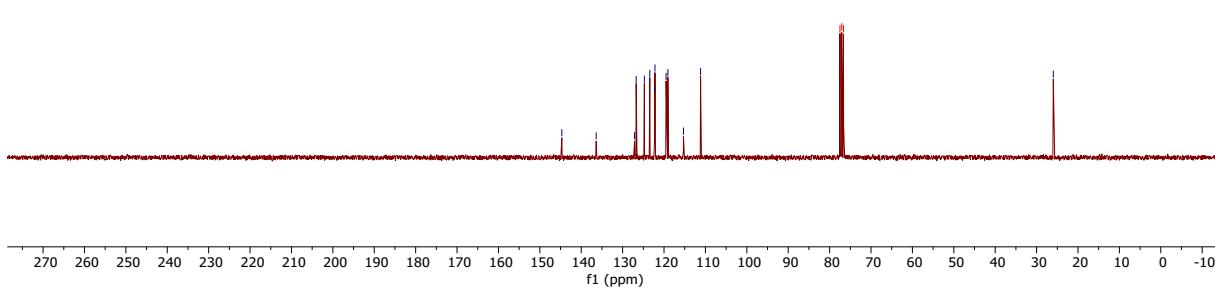
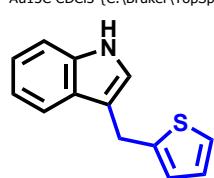
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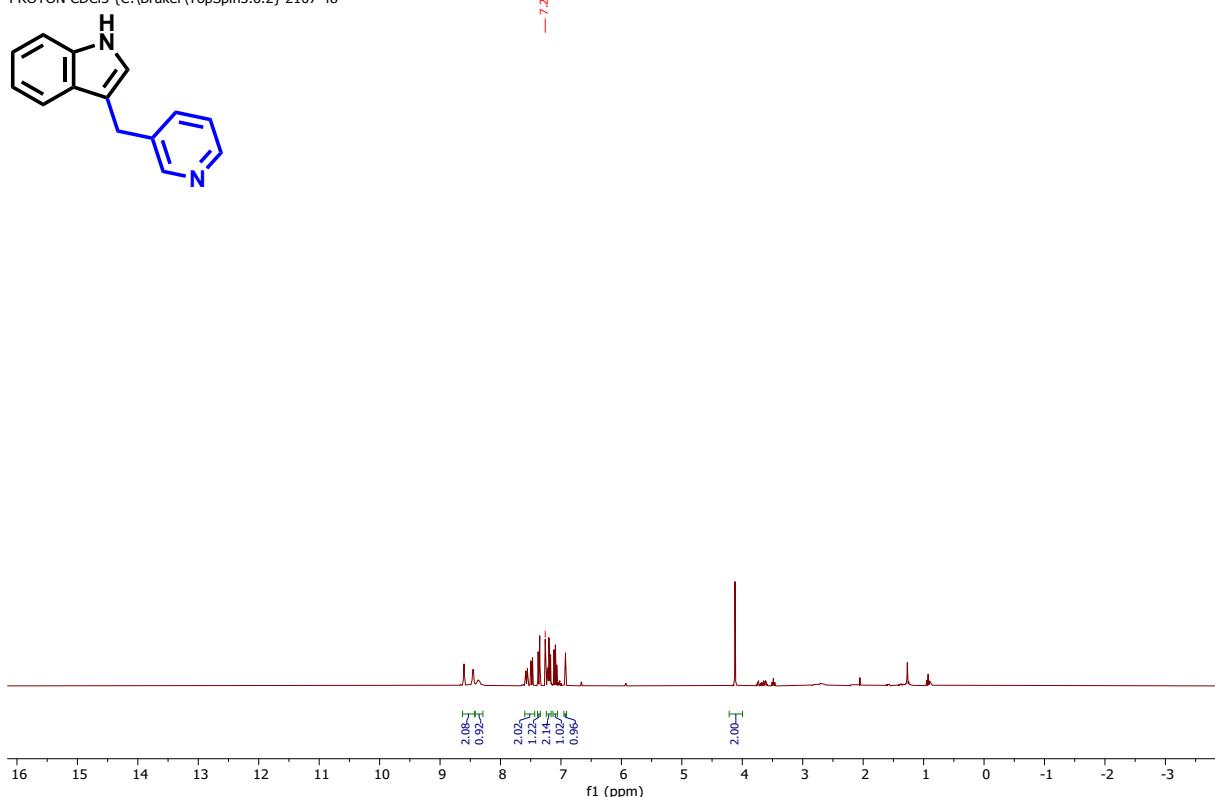
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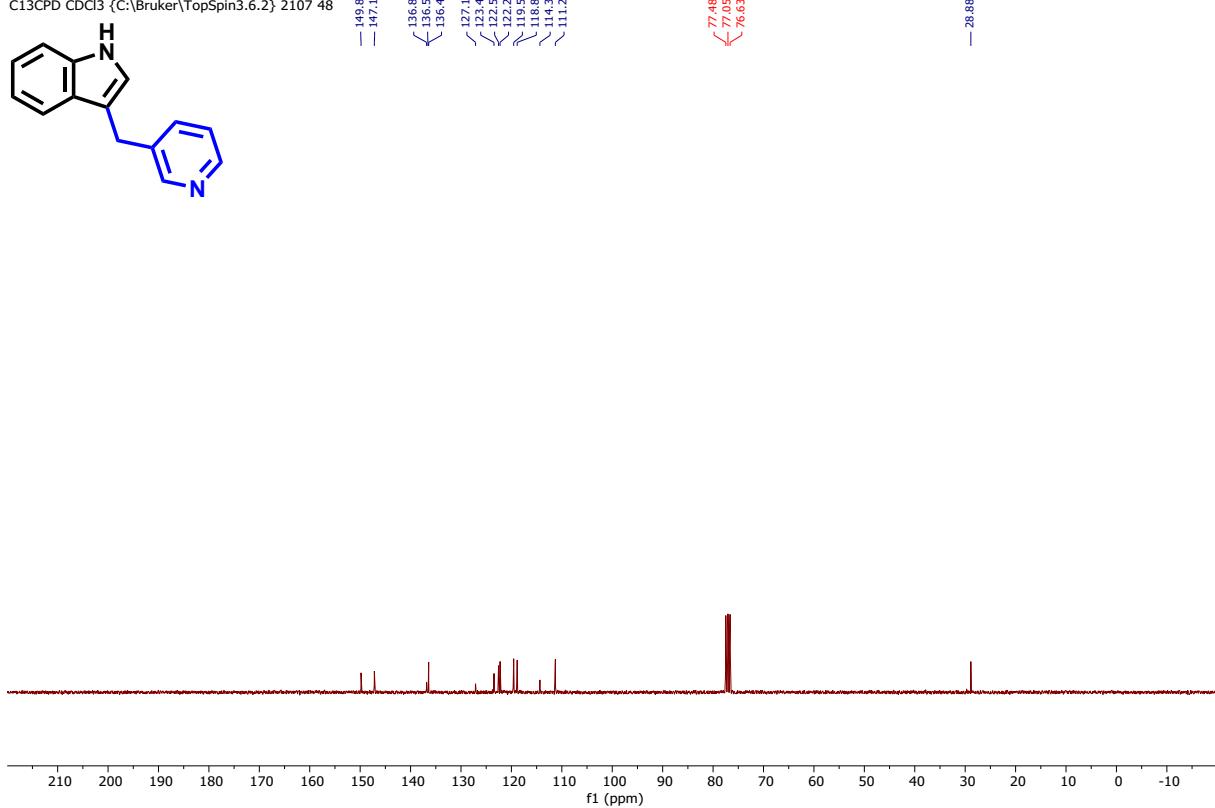
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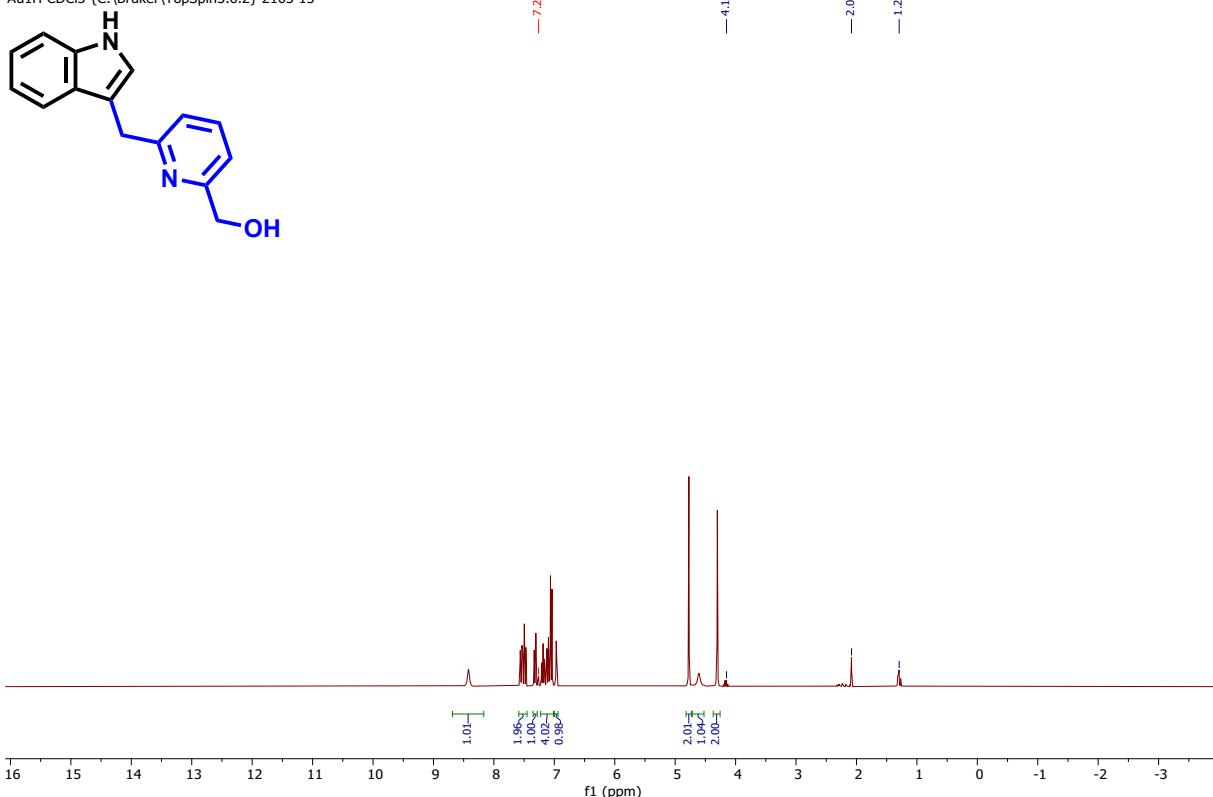
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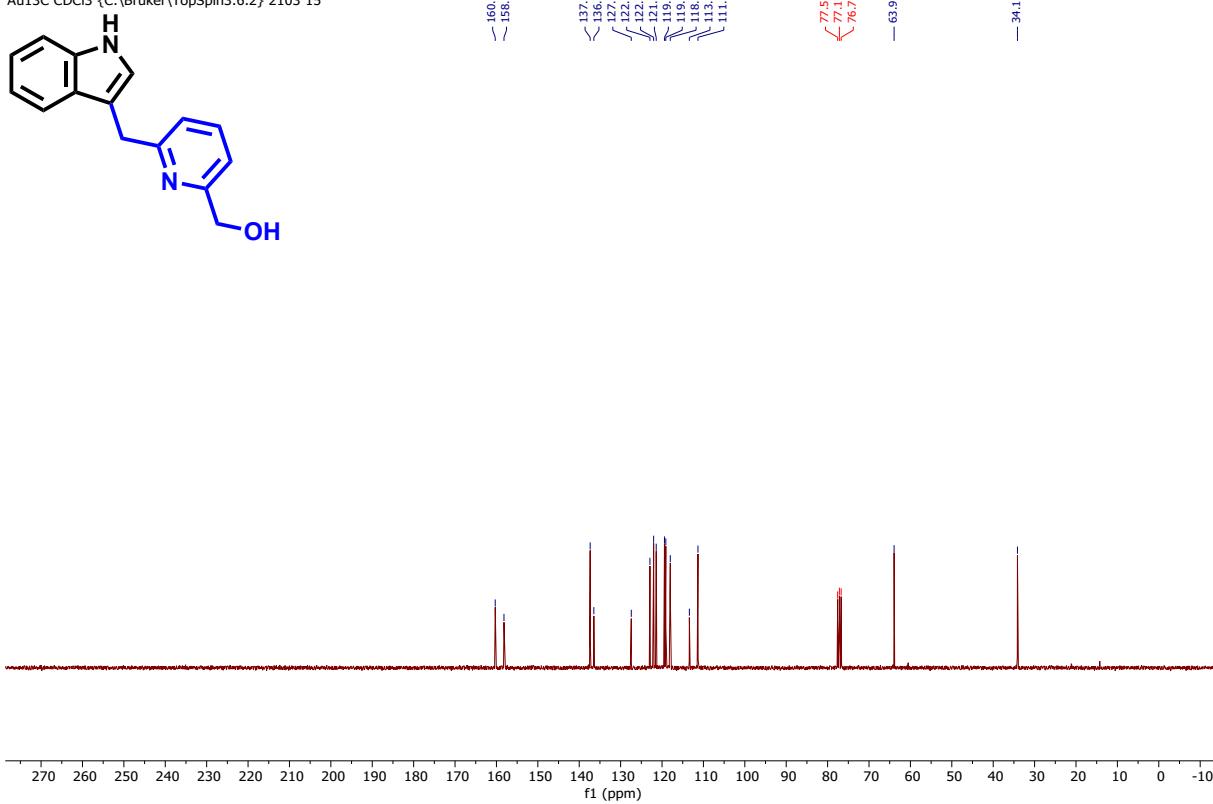
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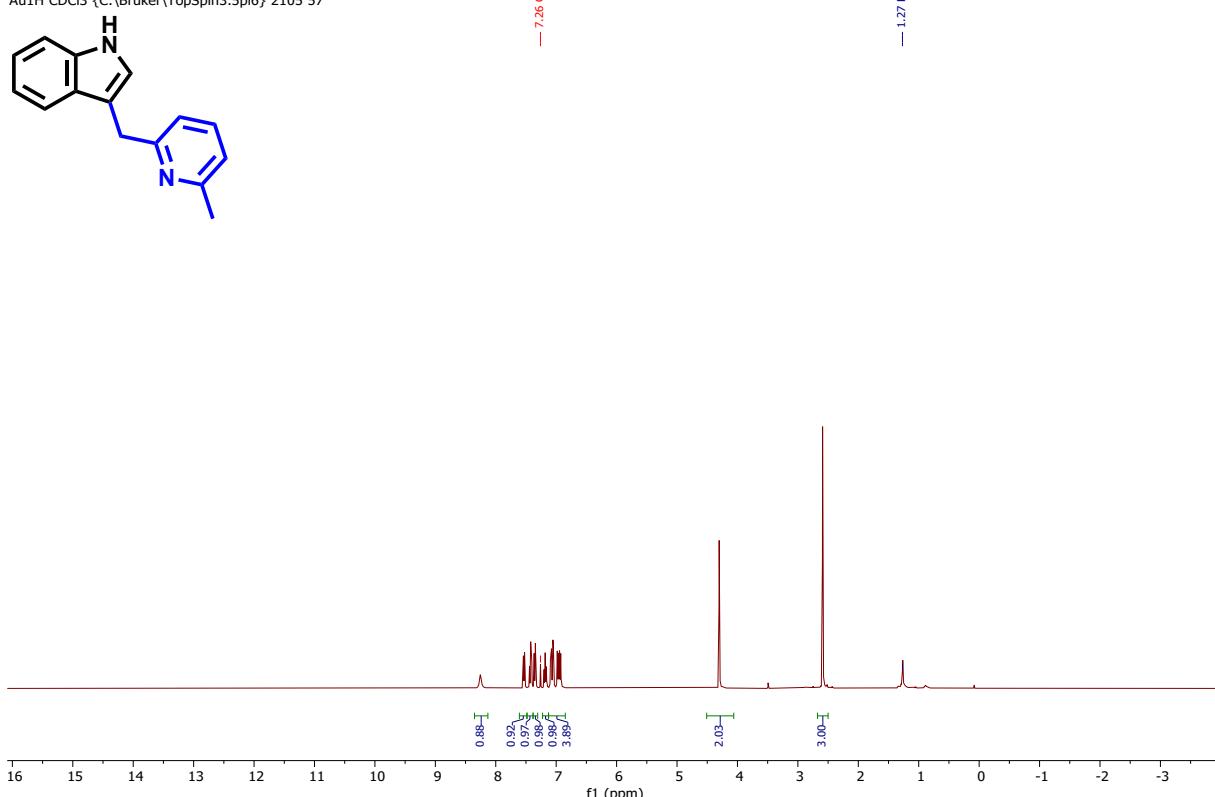
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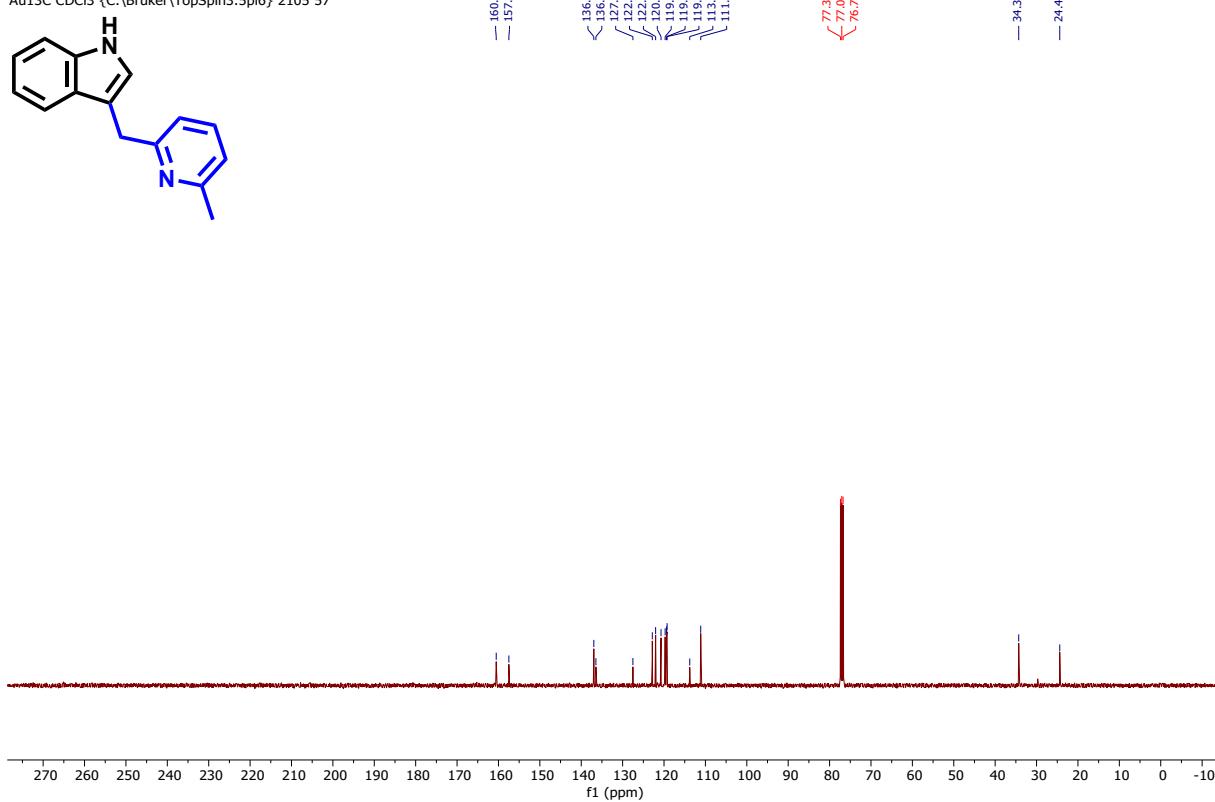
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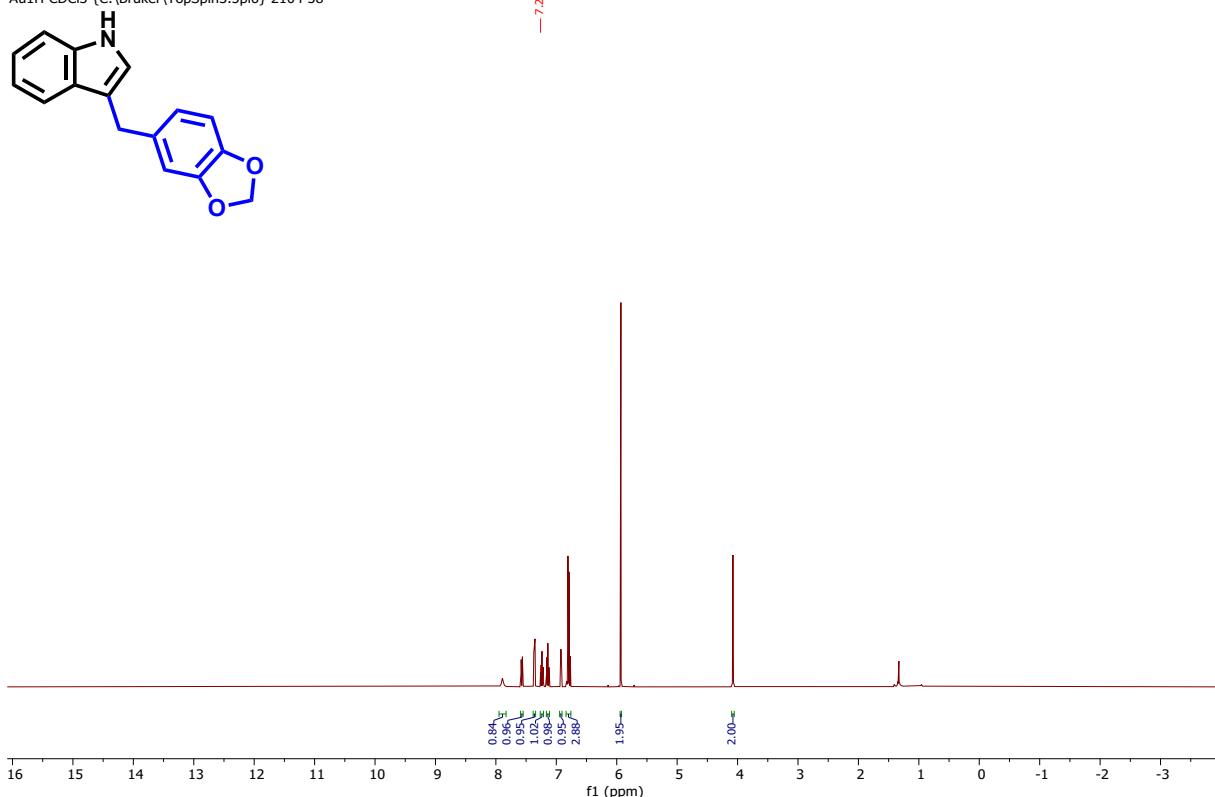
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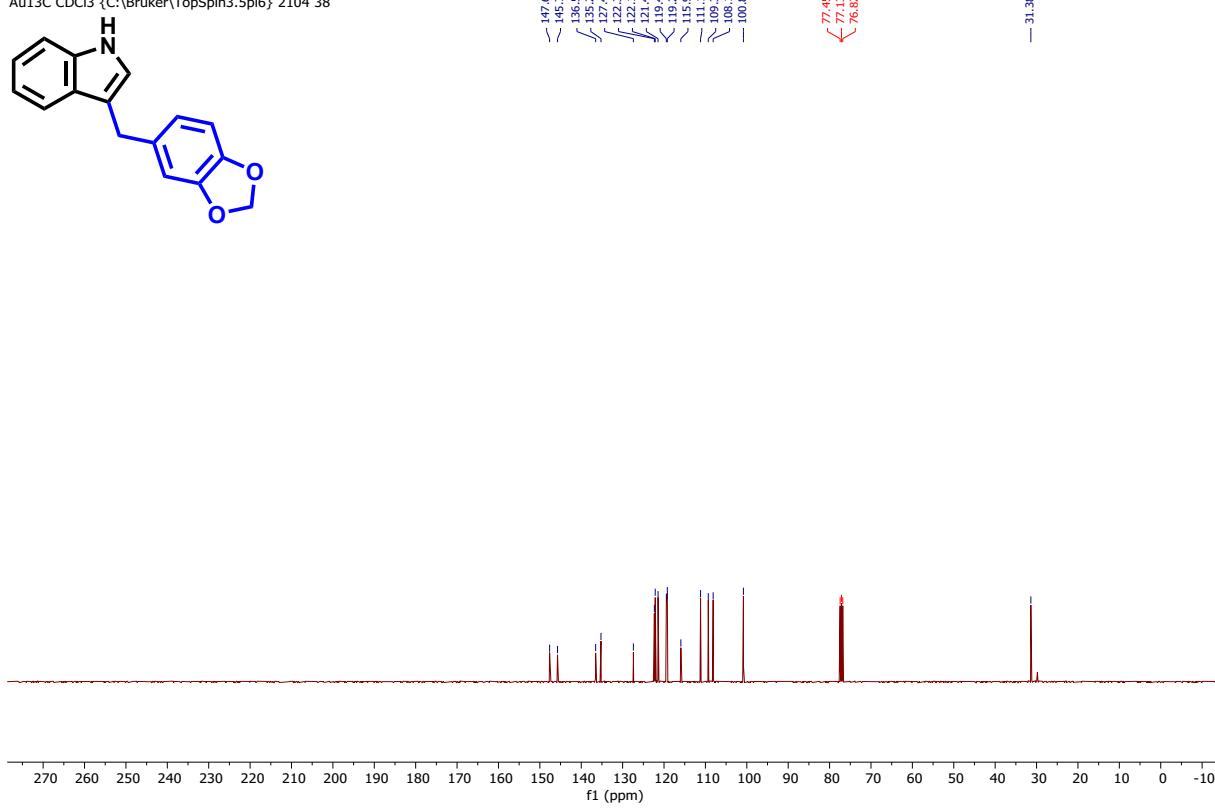
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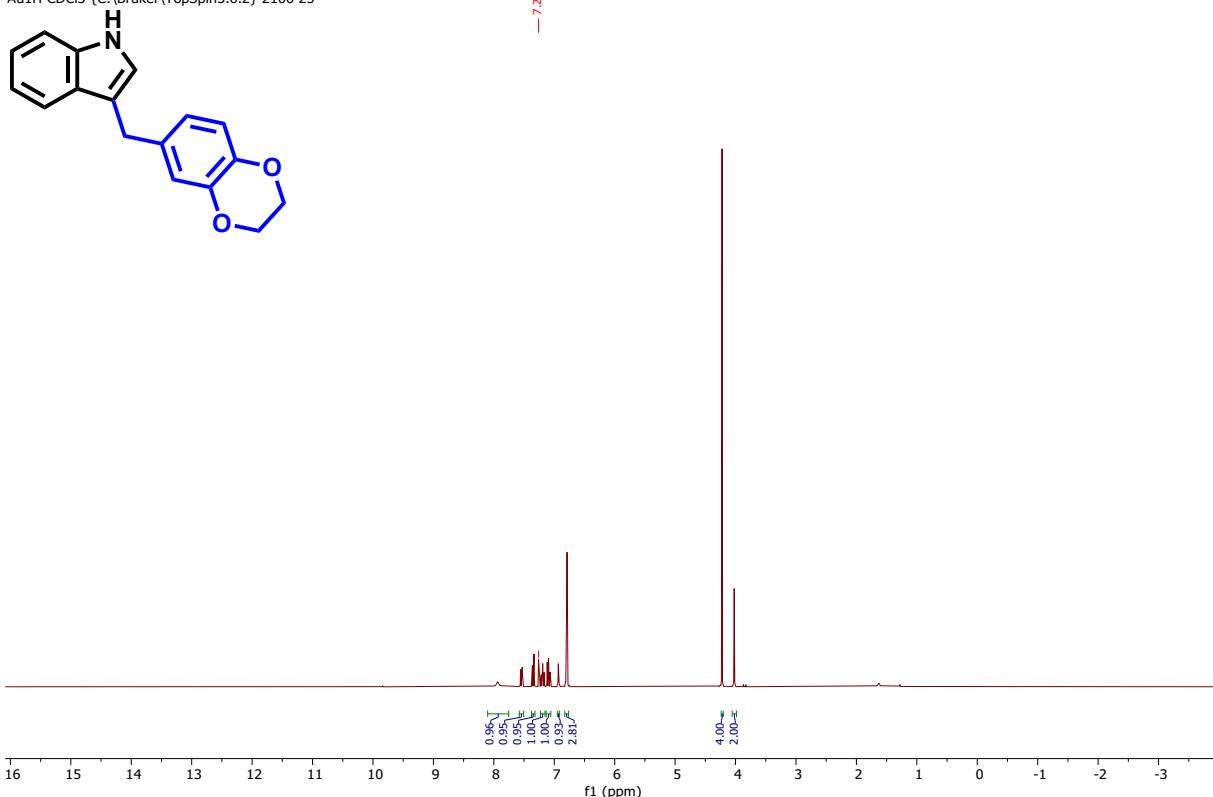
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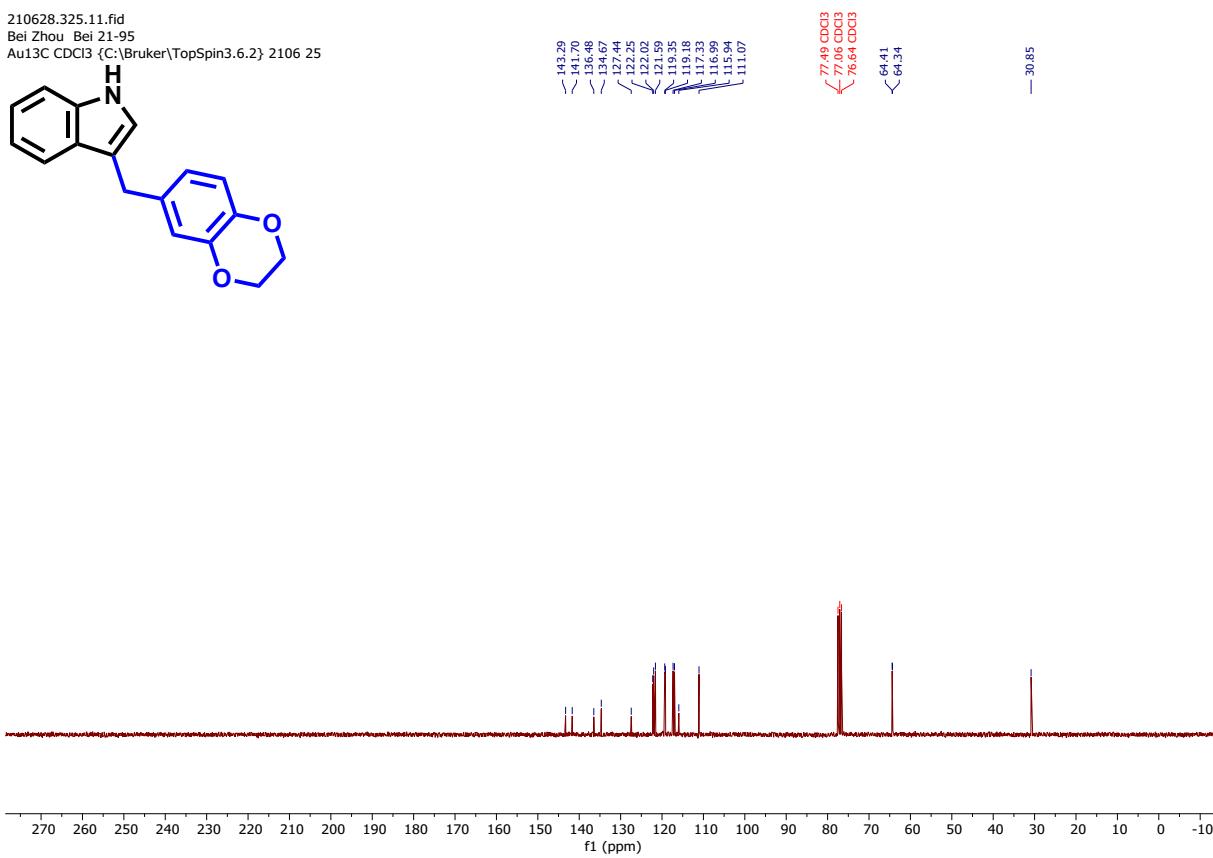
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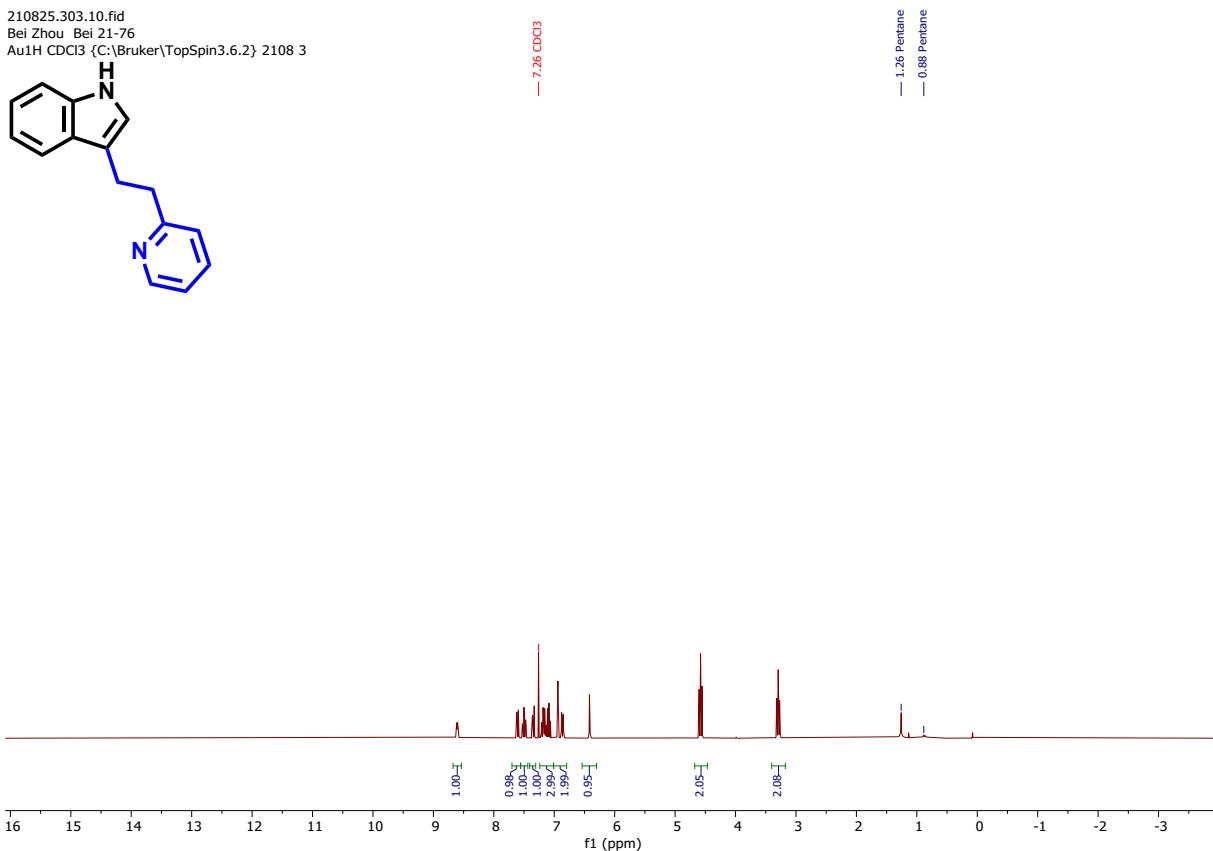
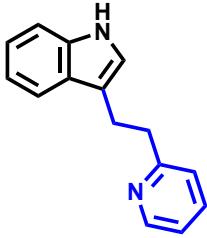
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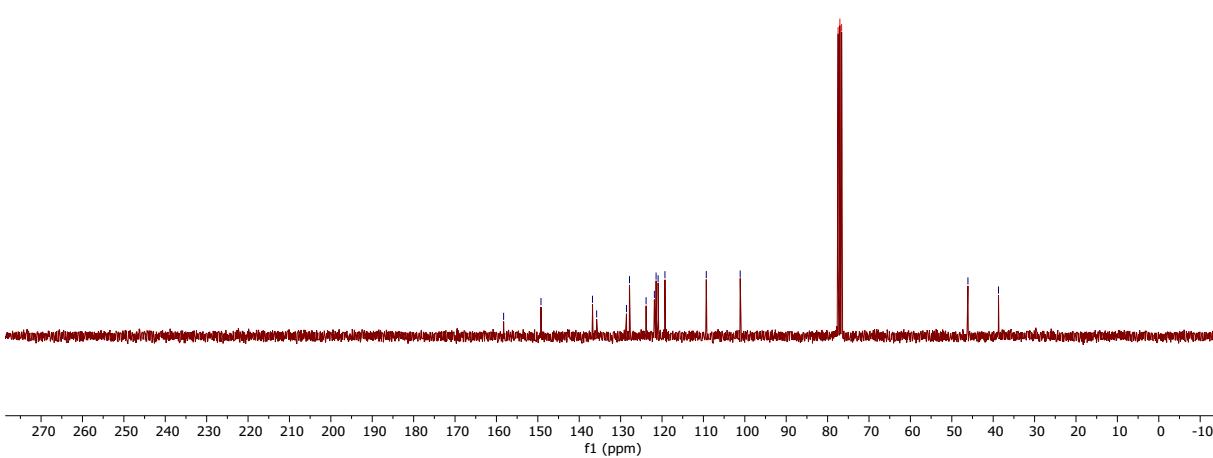
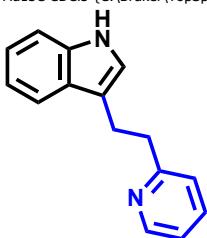
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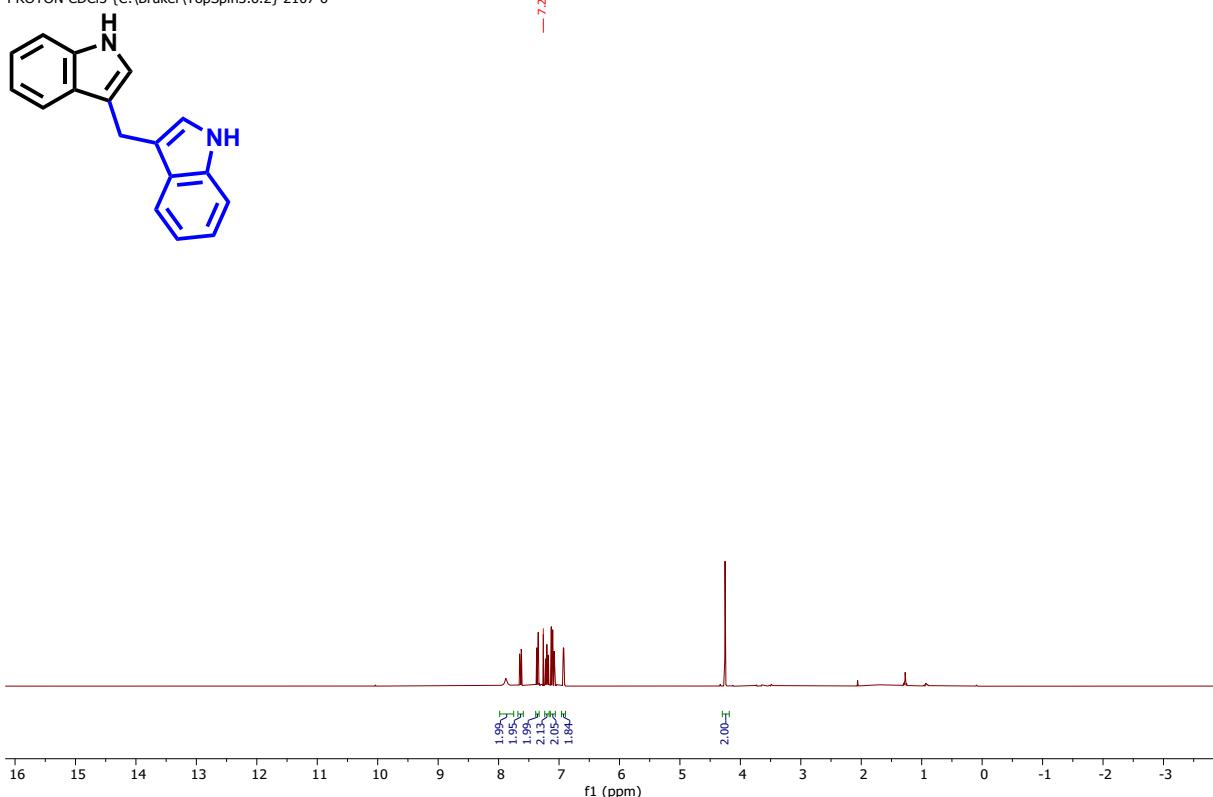
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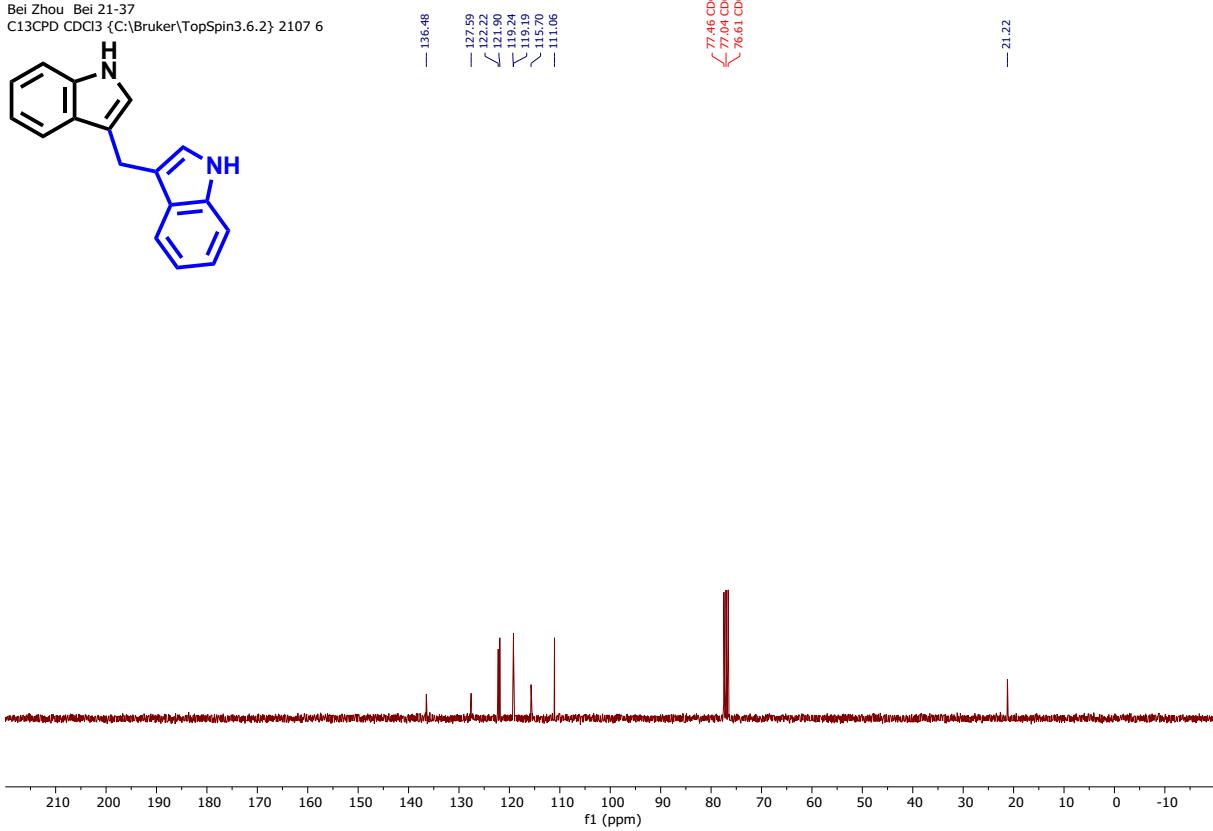
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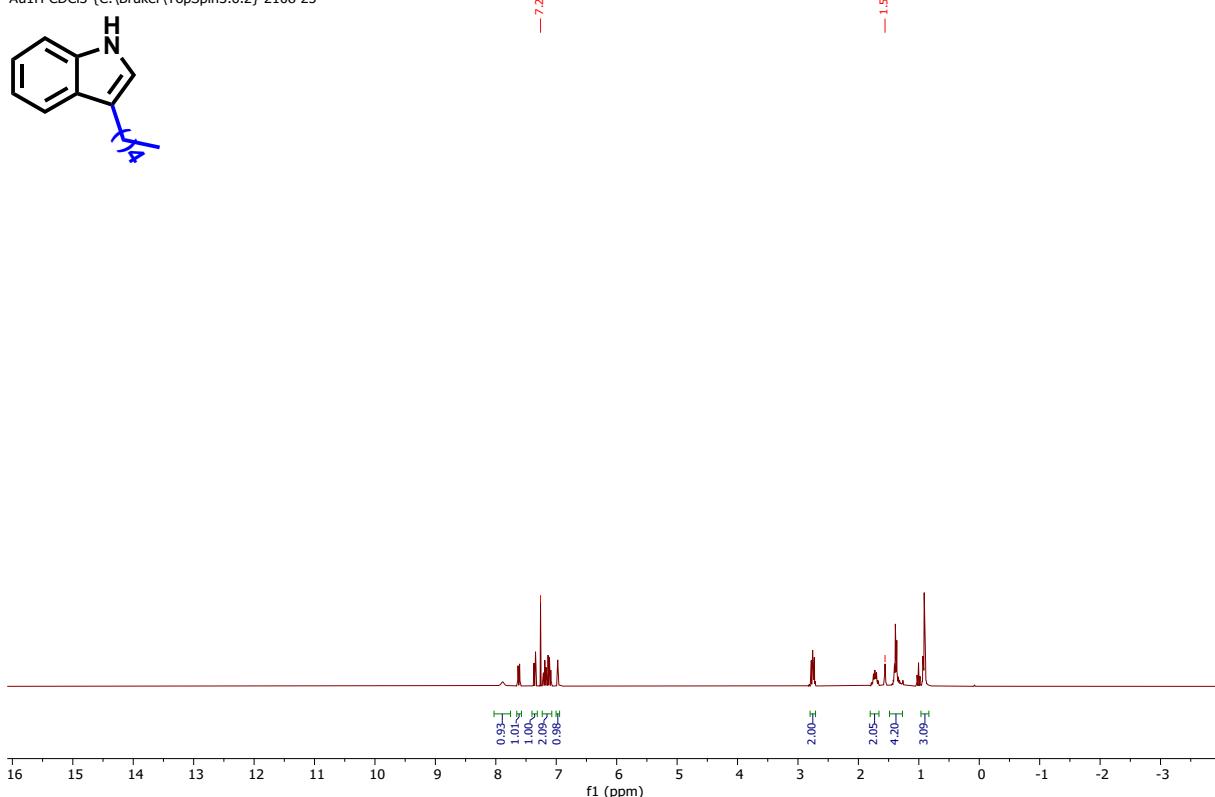
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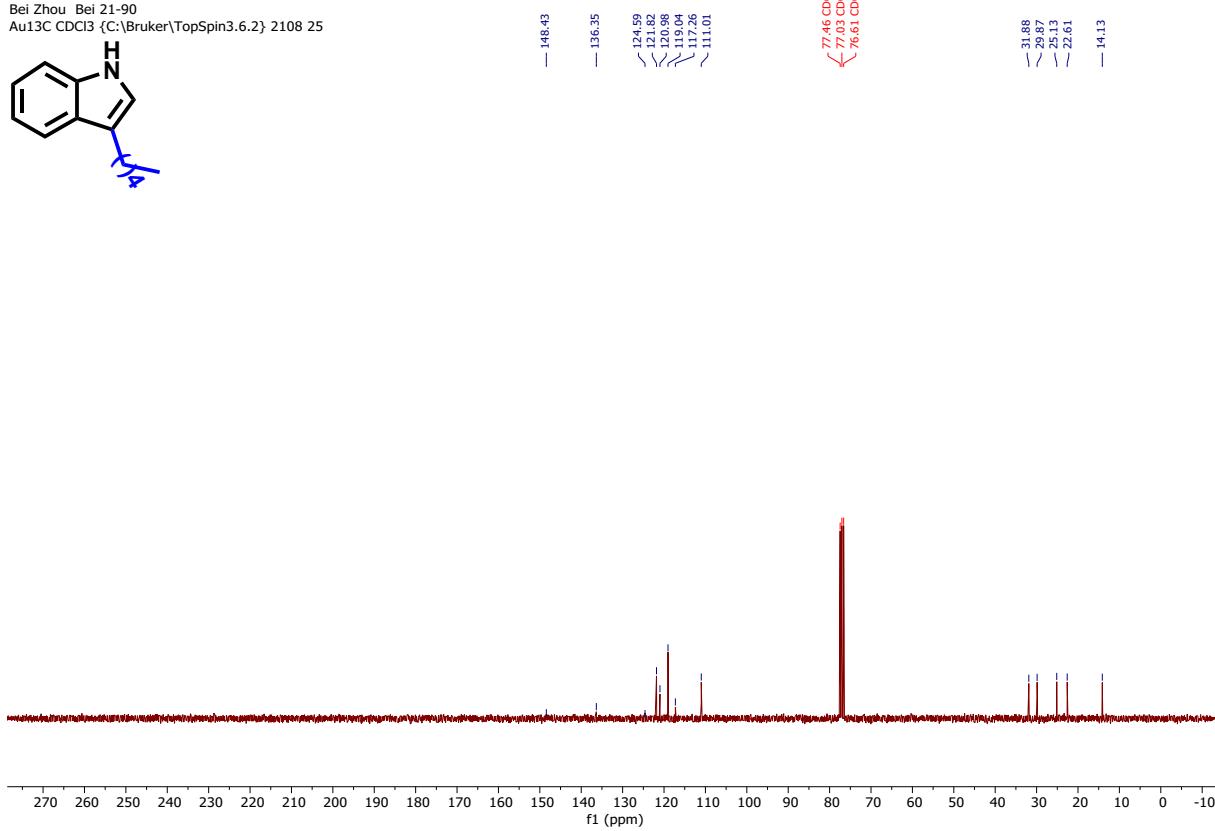
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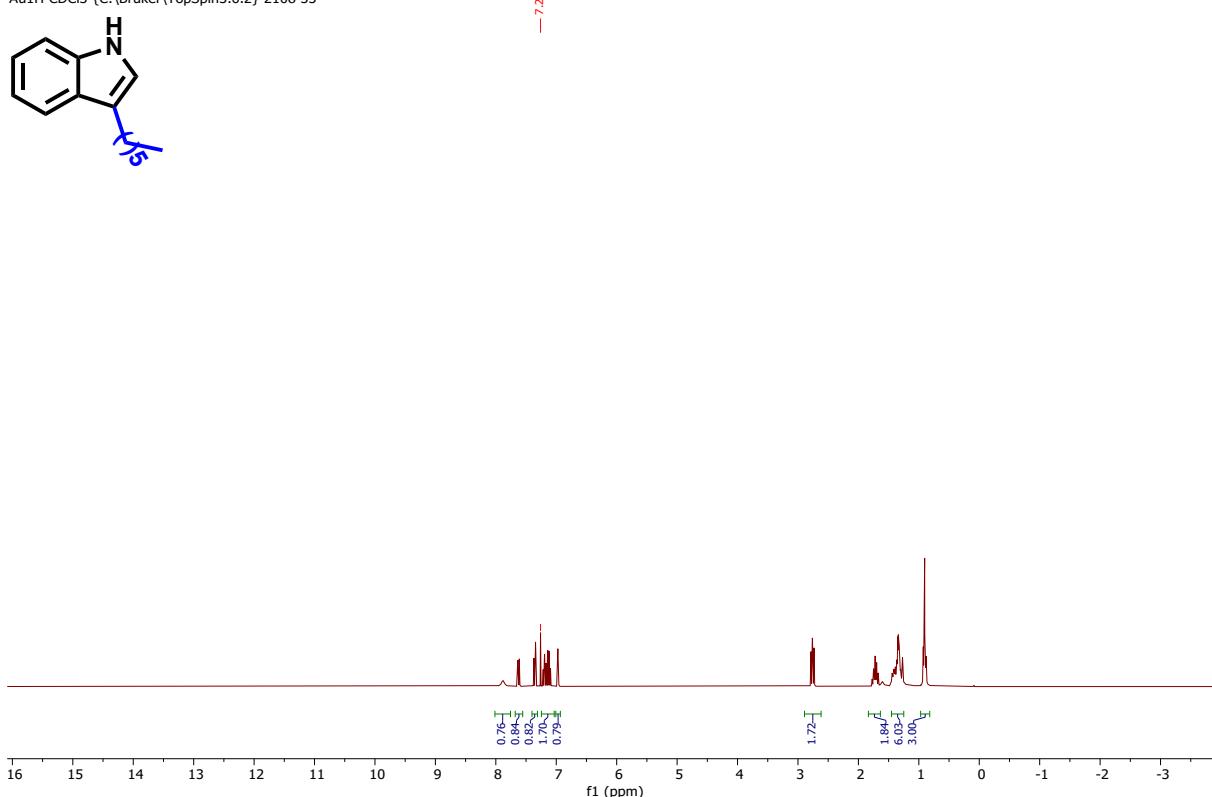
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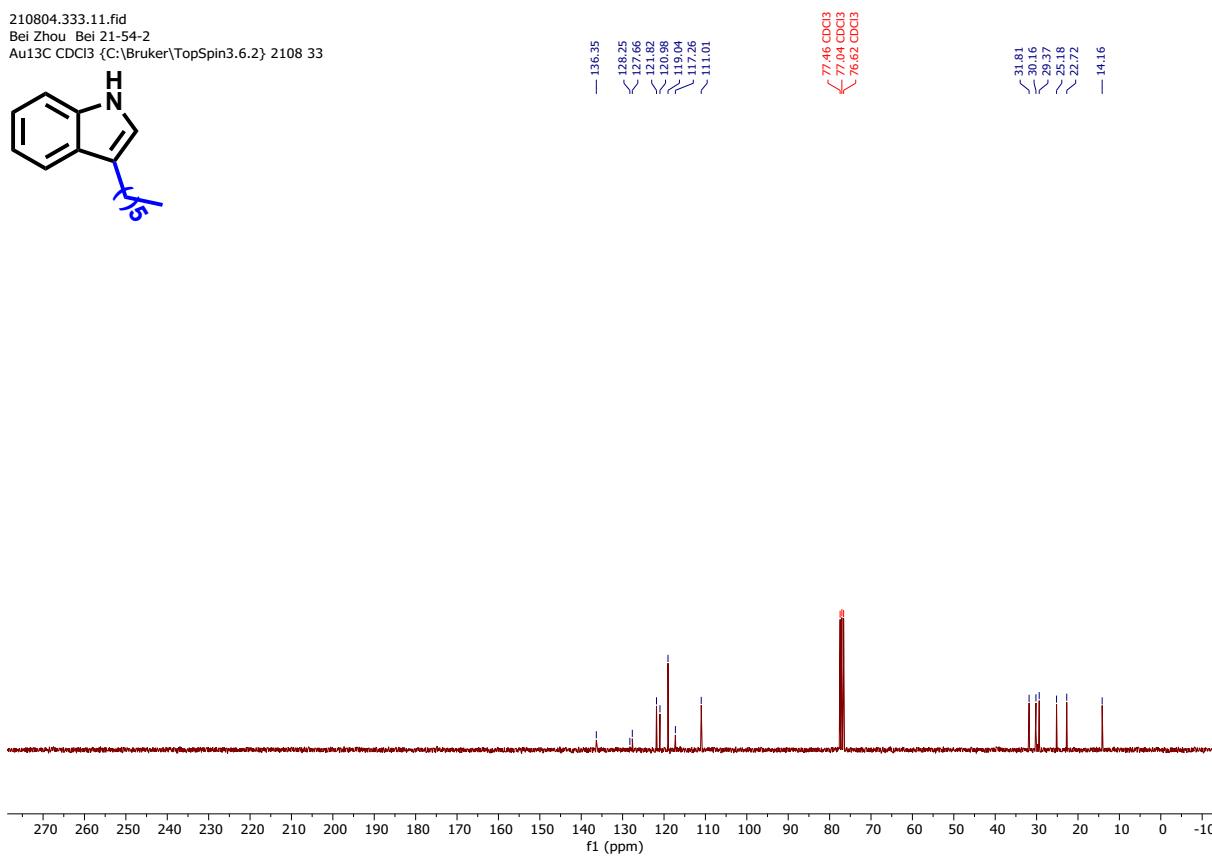
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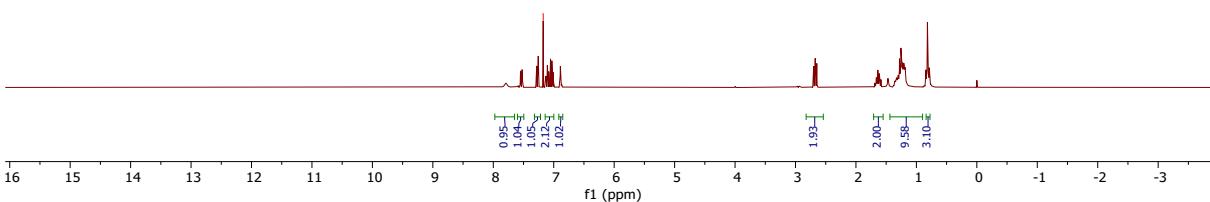
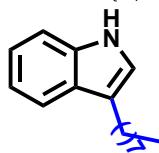
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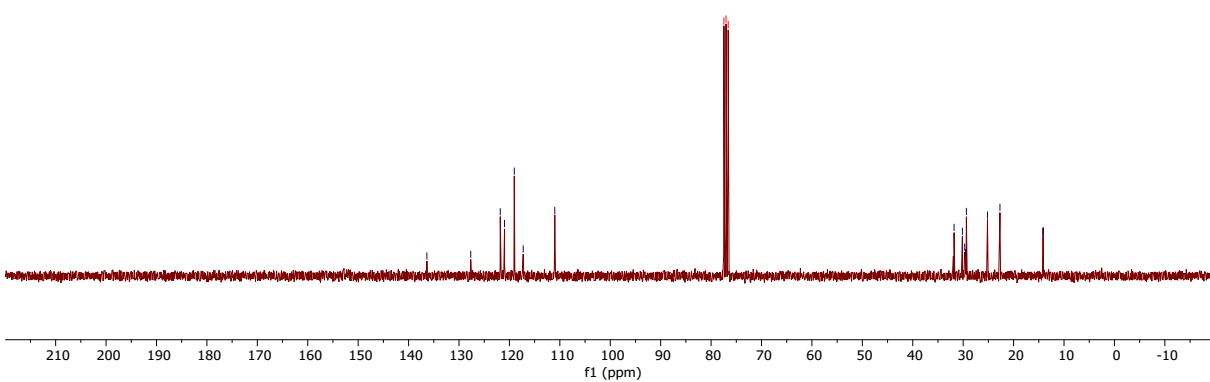
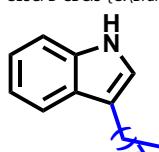
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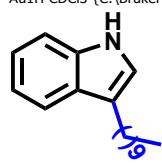
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PROTON CDCl3 {C:\Bruker\TopSpin3.6.2} 2108 21



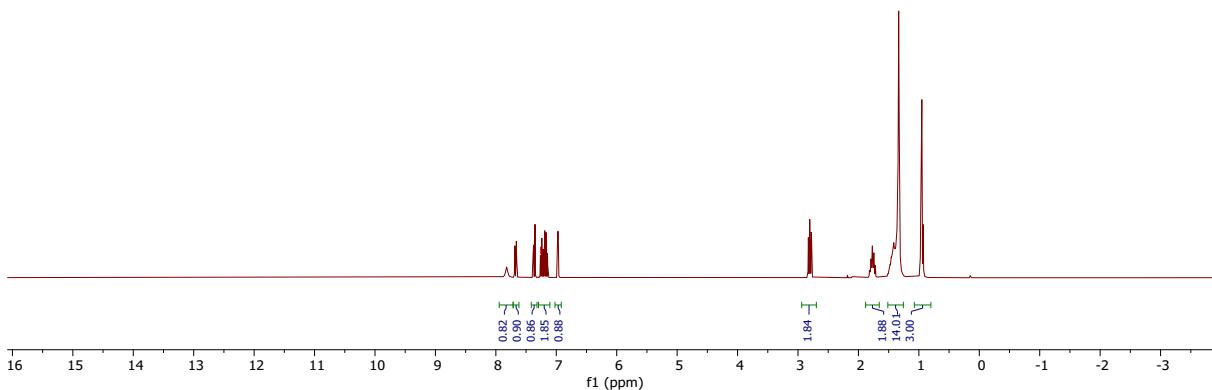
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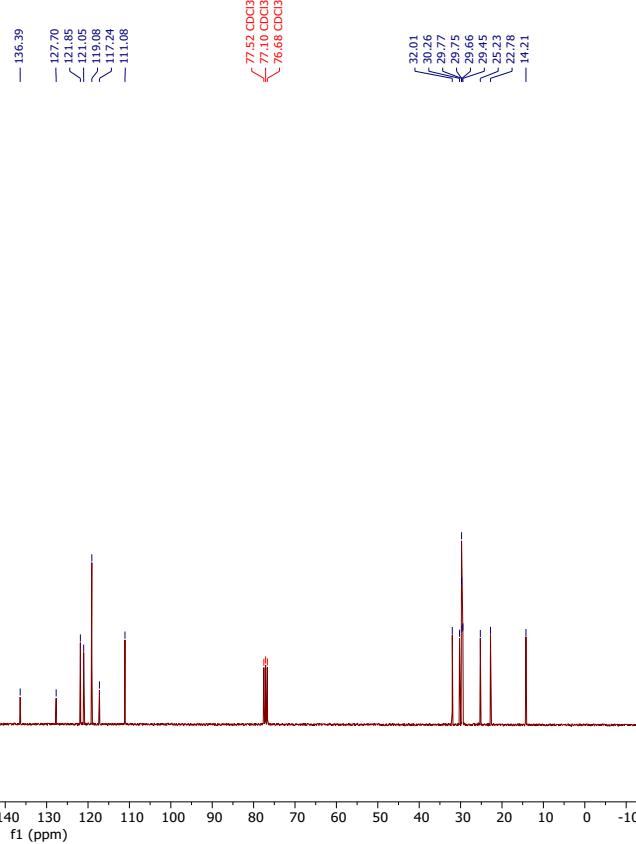
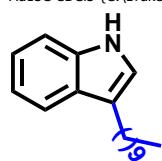
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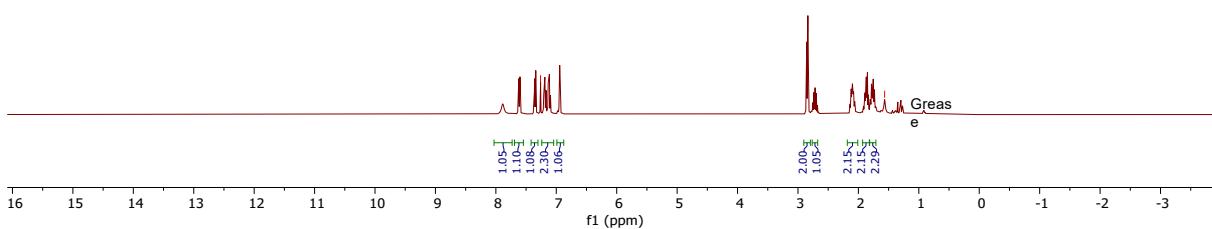
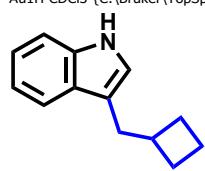
— 7.26 CDCl<sub>3</sub>



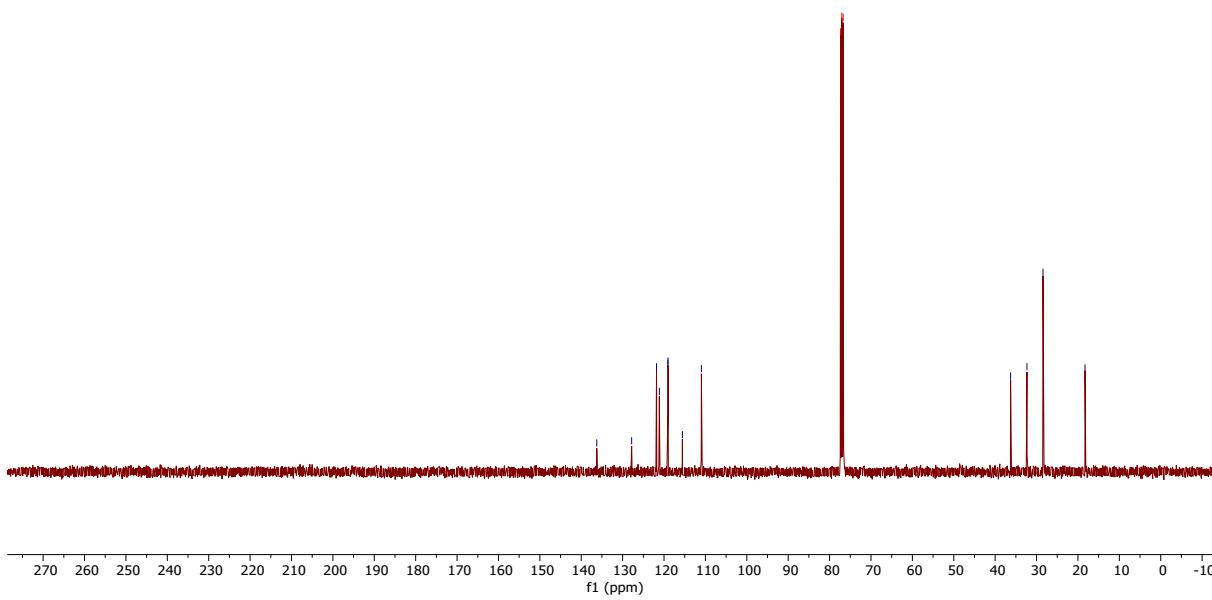
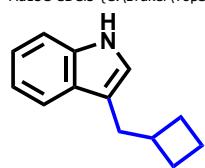
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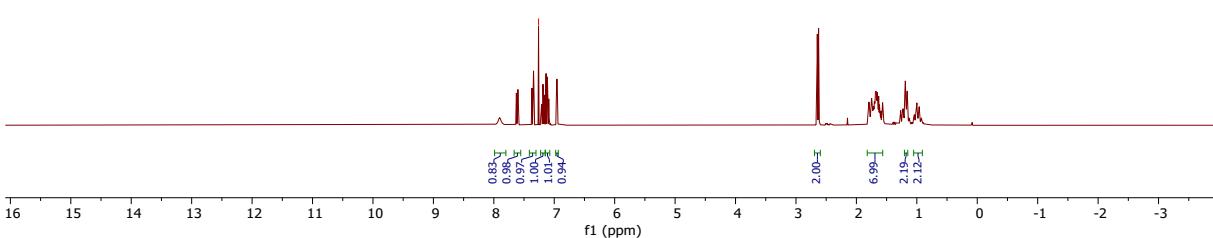
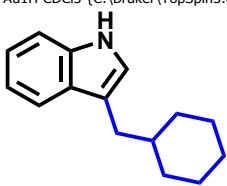
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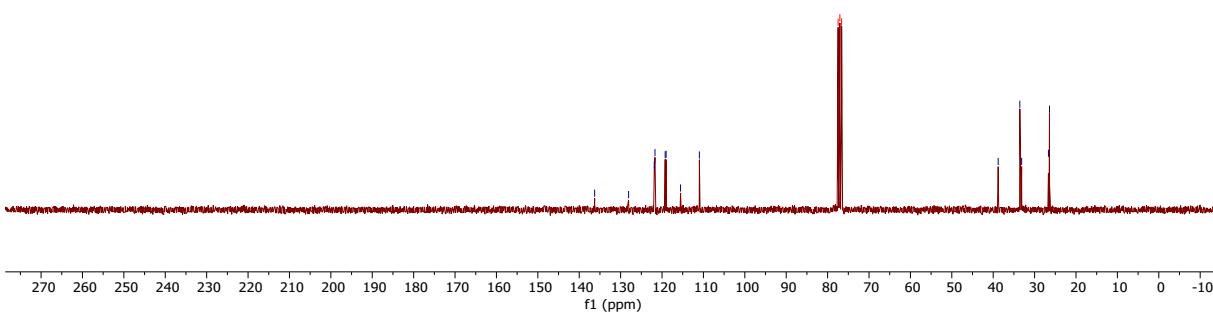
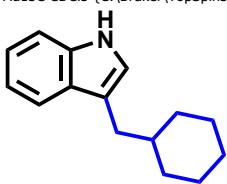
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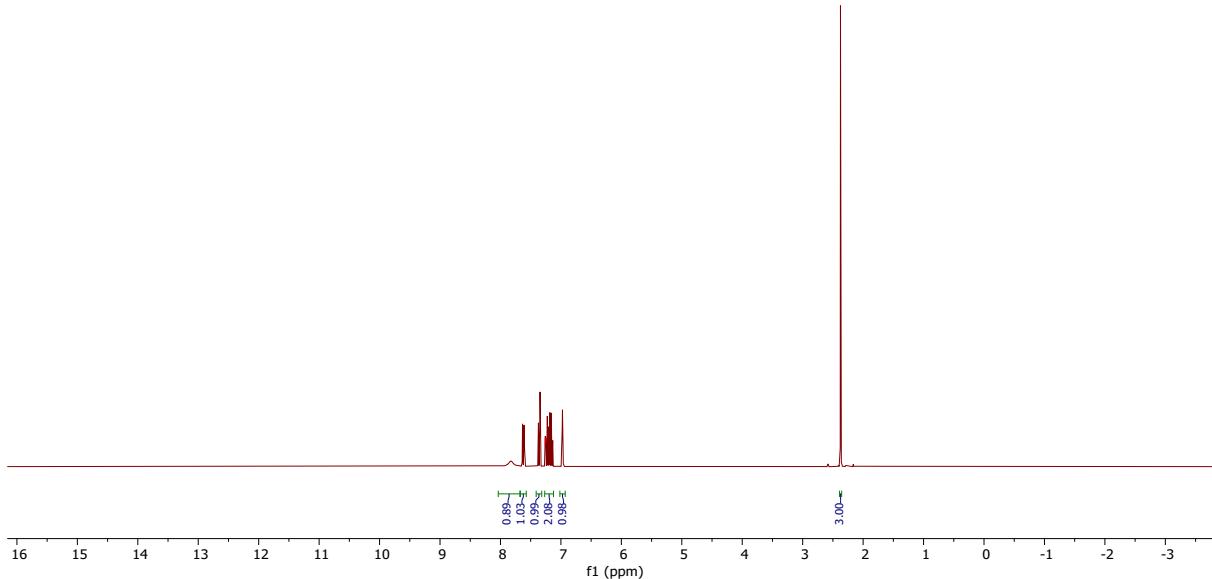
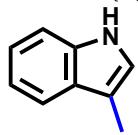
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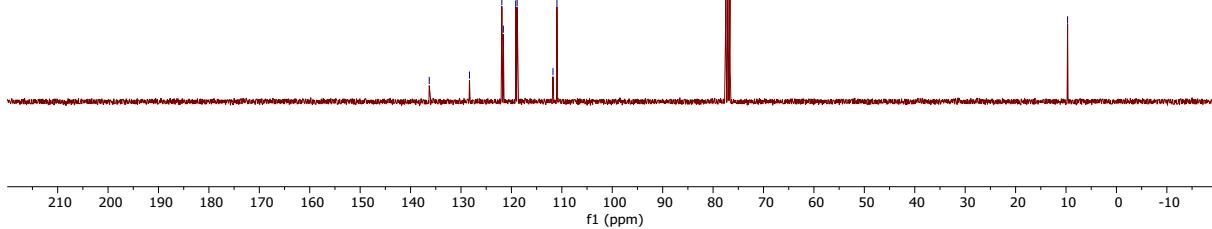
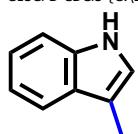
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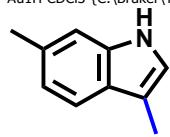
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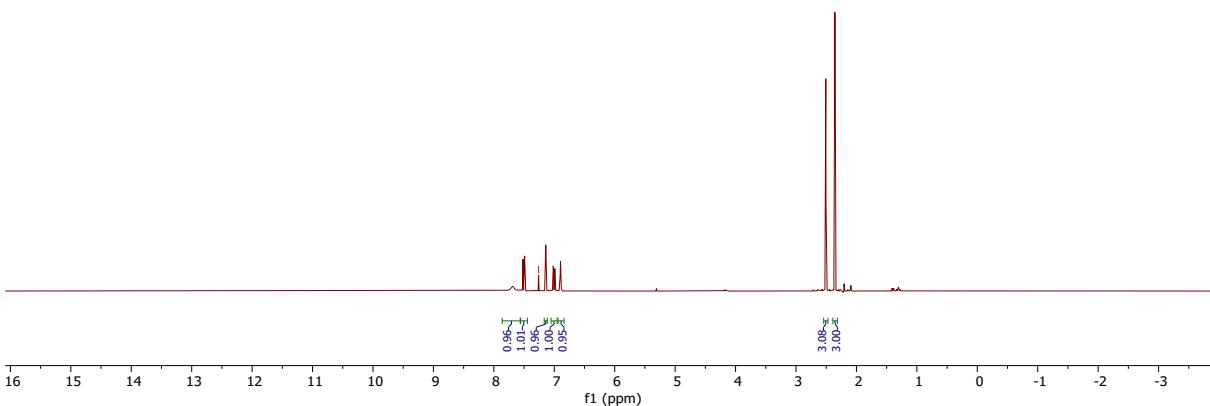
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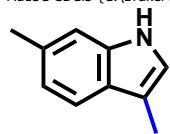
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Au1H CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 1



— 7.26 CDCl<sub>3</sub>



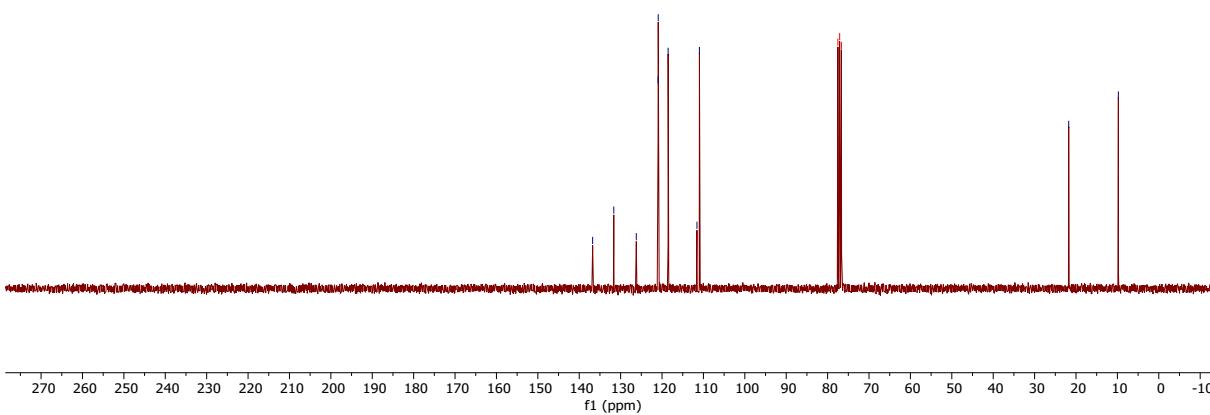
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Au13C CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 1



— 136.76  
— 131.65  
— 128.22  
✓ — 120.95  
✓ — 120.92  
— 118.53  
— 111.56  
— 110.97

— 77.51 CDCl<sub>3</sub>  
— 77.09 CDCl<sub>3</sub>  
— 76.87 CDCl<sub>3</sub>

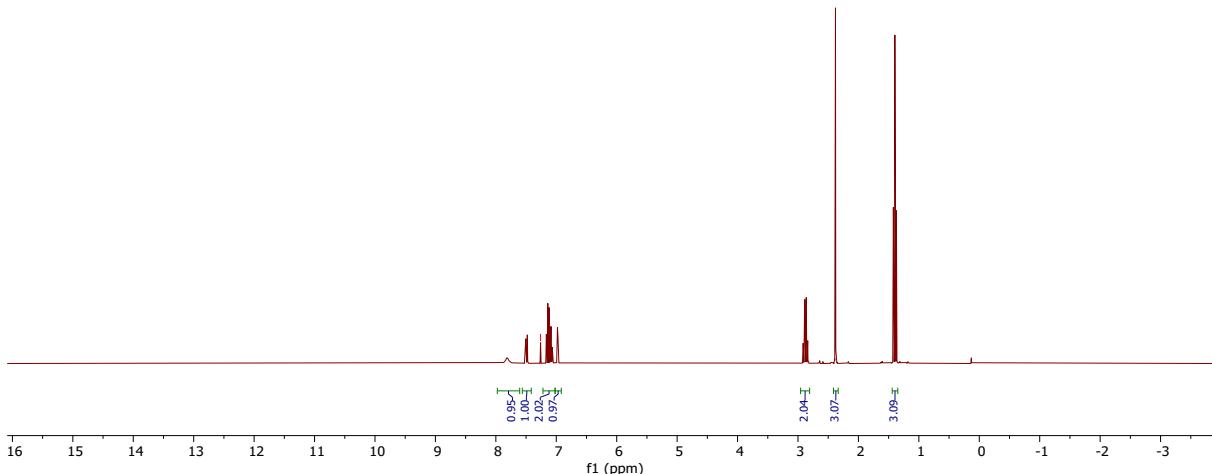
— 21.76  
— 9.77



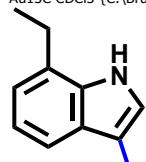
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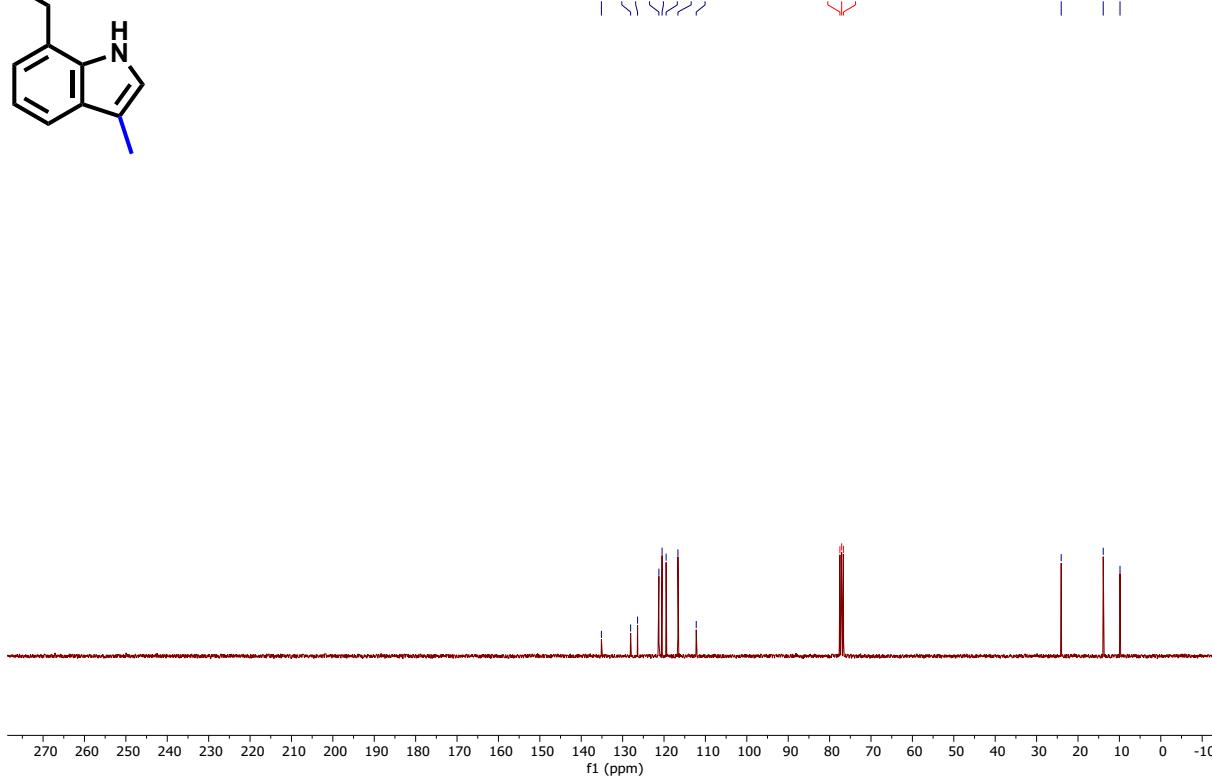
- 7.26 CDCl<sub>3</sub>



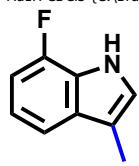
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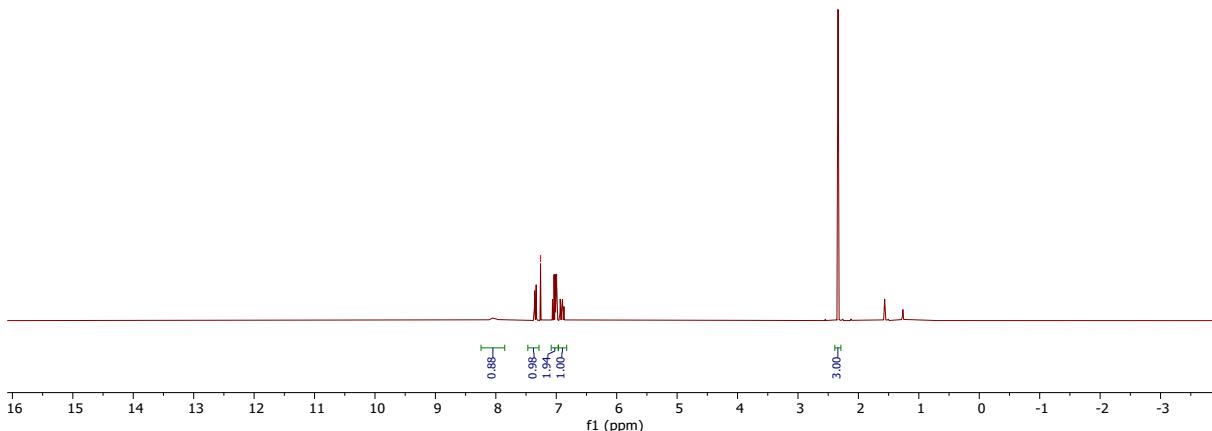
— 77.51 CDCl<sub>3</sub>  
— 77.09 CDCl<sub>3</sub>  
— 76.66 CDCl<sub>3</sub>  
— 24.06  
— 13.91  
— 9.85



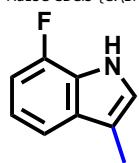
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— 7.26 CDCl<sub>3</sub>



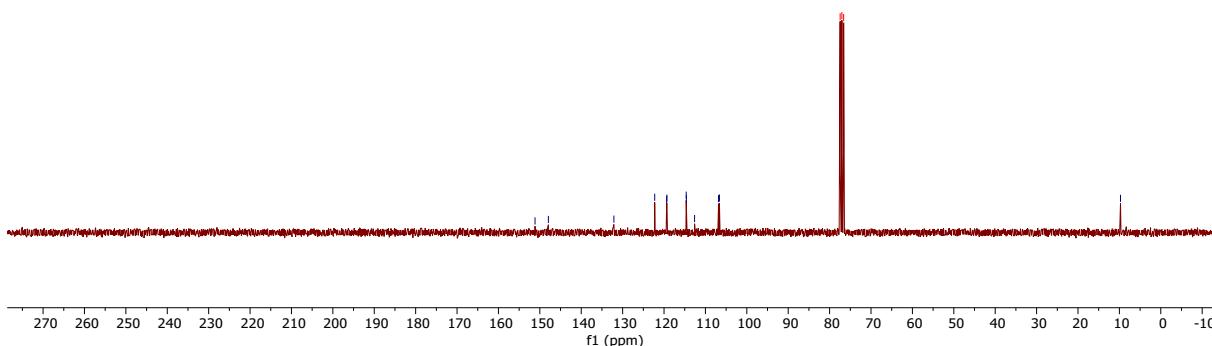
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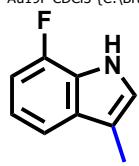
— 151.16  
— 147.94  
— 132.11  
— 122.25  
— 119.38  
— 119.29  
— 114.64  
— 114.60  
— 112.62  
— 108.85  
— 108.62

— 77.46 CDCl<sub>3</sub>  
— 77.03 CDCl<sub>3</sub>  
— 76.61 CDCl<sub>3</sub>

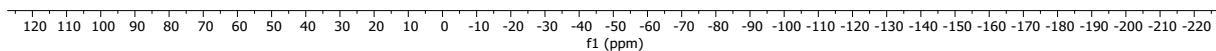
— 9.74



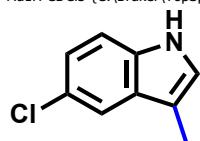
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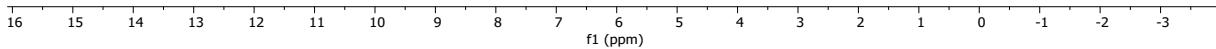
-135.81  
-135.83  
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-135.87



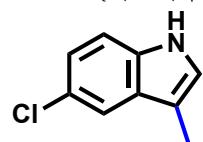
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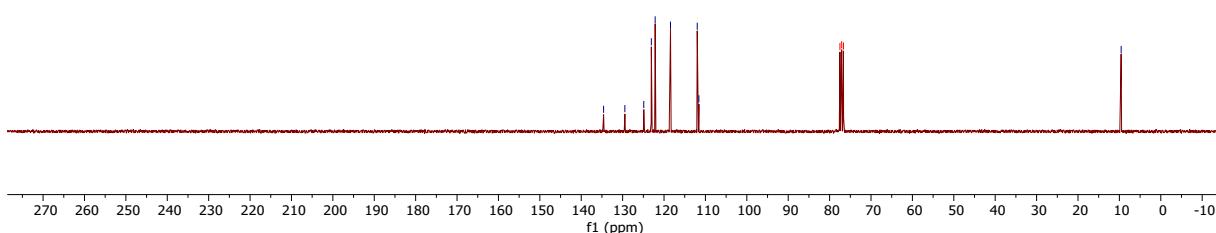
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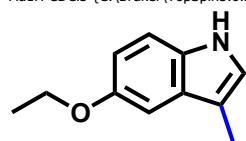
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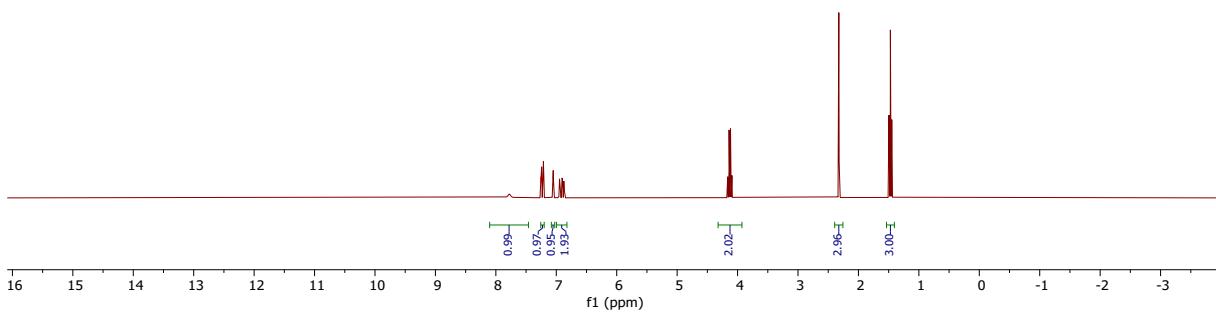
-134.60  
-129.46  
-124.89  
-123.07  
-122.14  
-118.44  
-111.98  
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-77.51 CDCl<sub>3</sub>  
-77.08 CDCl<sub>3</sub>  
-76.66 CDCl<sub>3</sub>  
-9.57



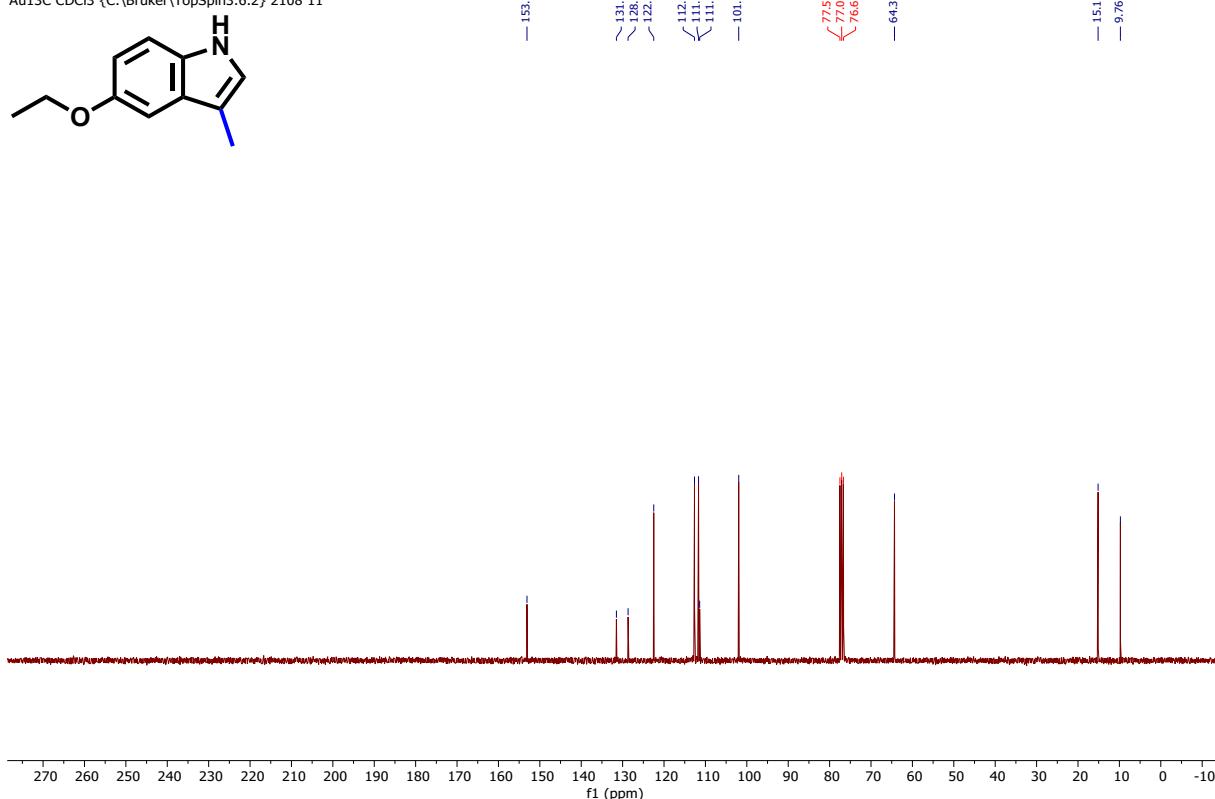
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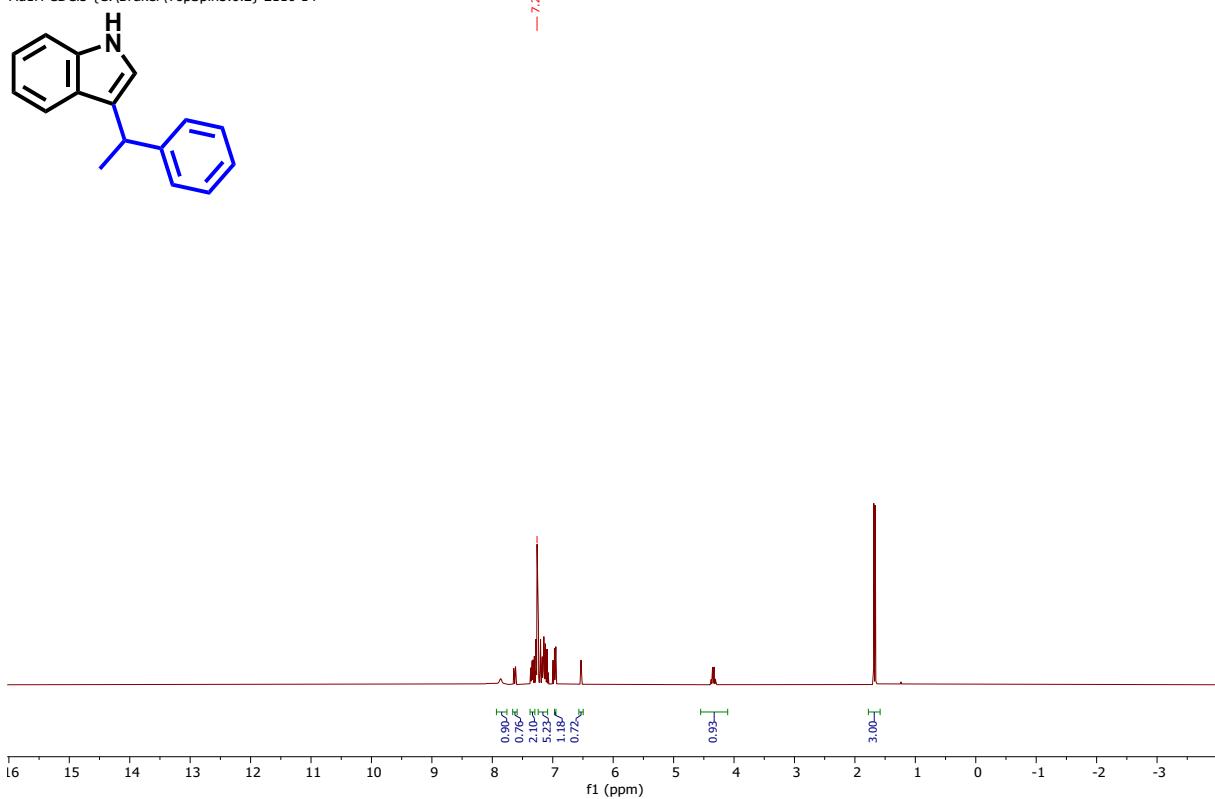
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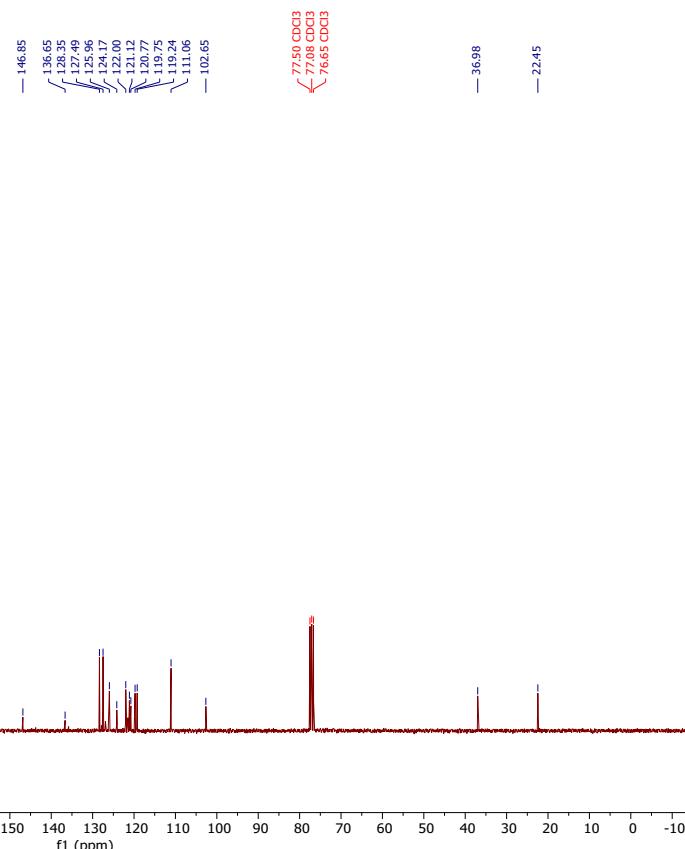
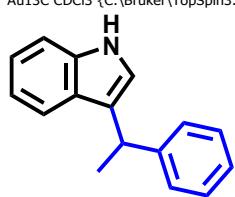
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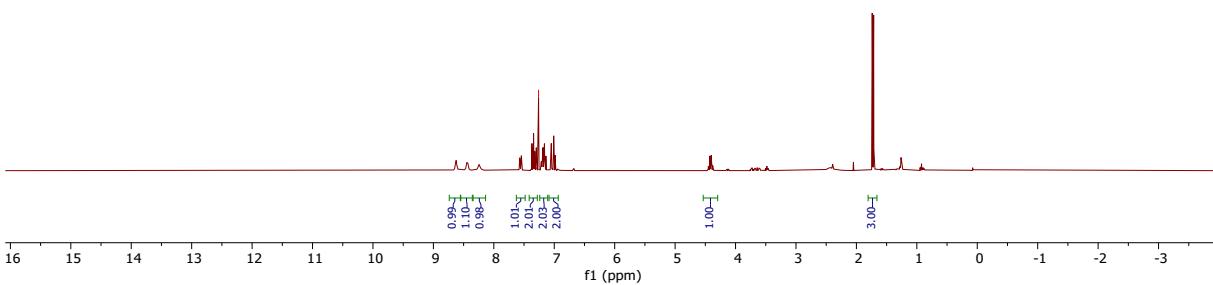
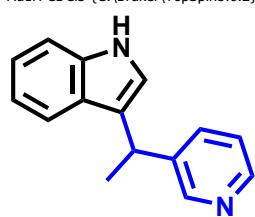
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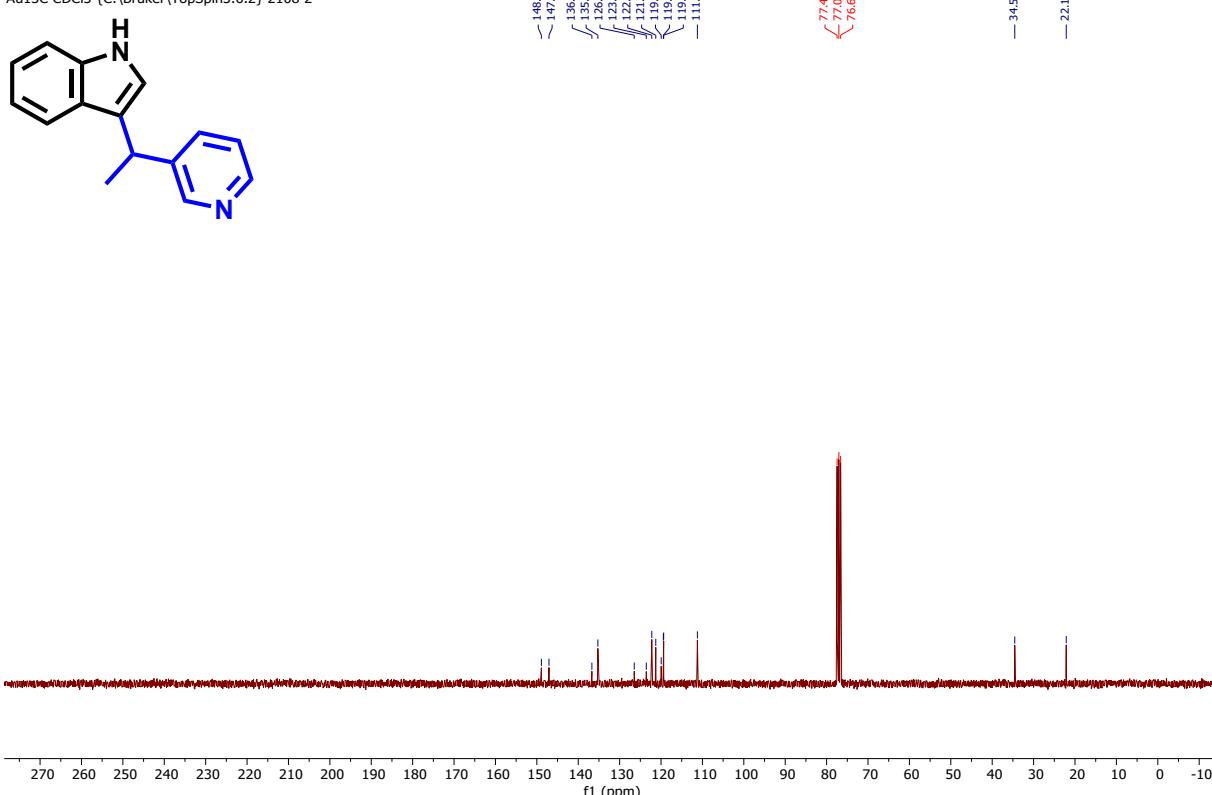
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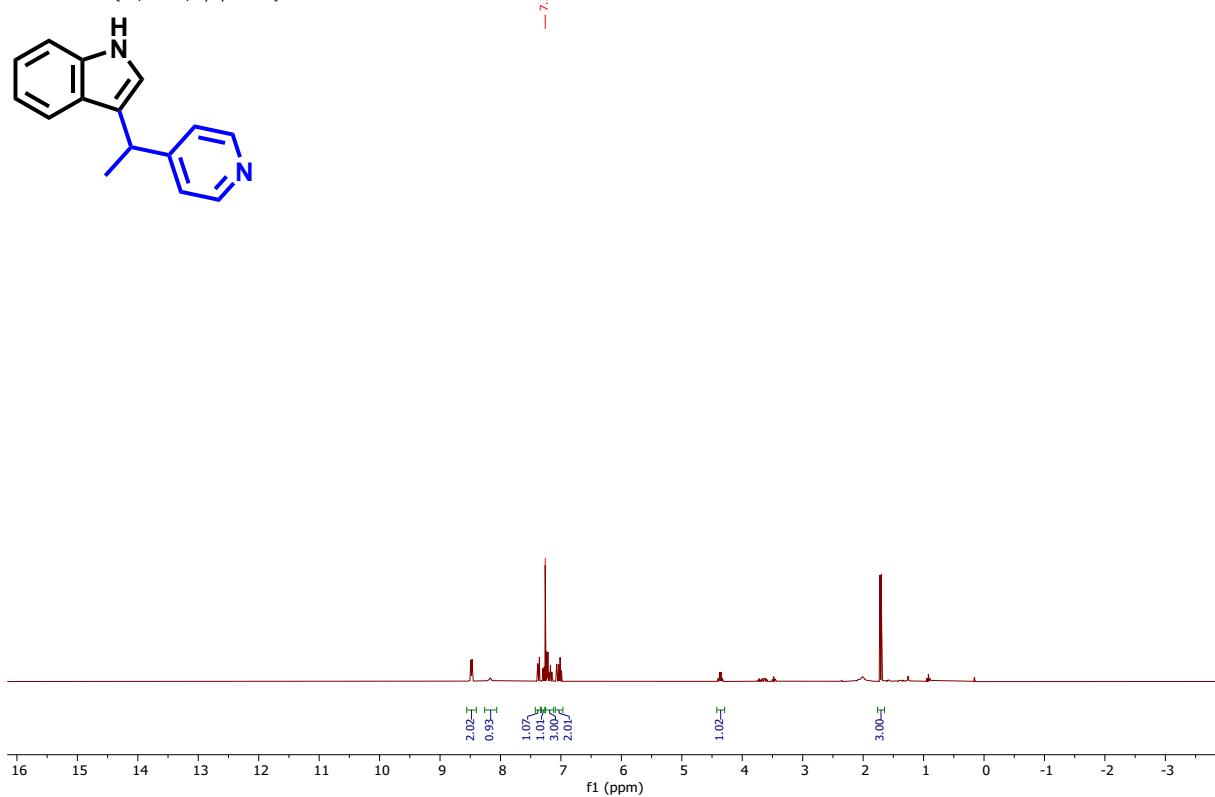
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Au1H CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 2



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Bei Zhou Bei 21-6-1  
Au13C CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 2



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PROTON CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 23

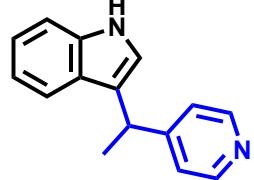


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Bei Zhou Bei 21-7-1

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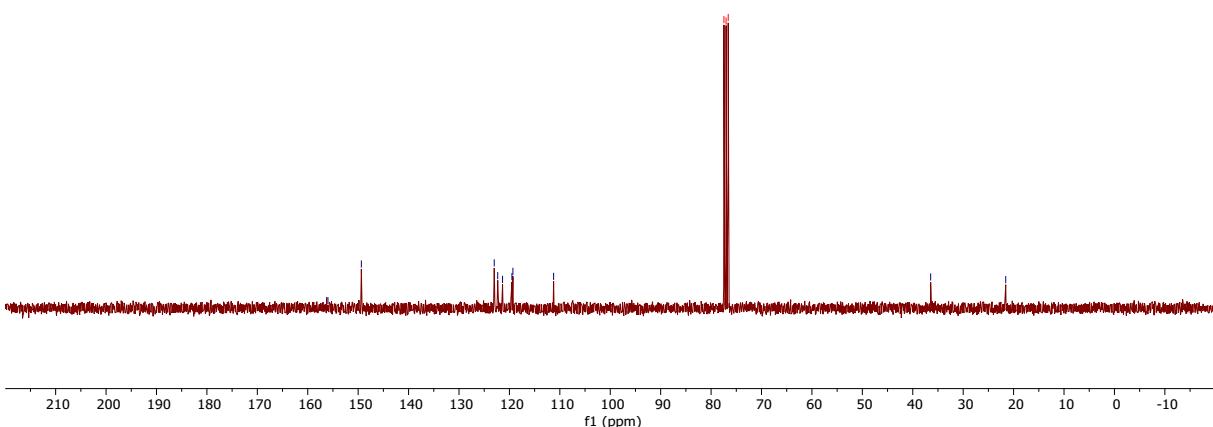
— 153.9 — 149.37



— 123.01 — 122.32 — 121.36 — 119.51 — 119.31 — 111.23

— 77.45 CDCl<sub>3</sub> — 77.03 CDCl<sub>3</sub> — 76.60 CDCl<sub>3</sub>

— 36.45 — 21.56

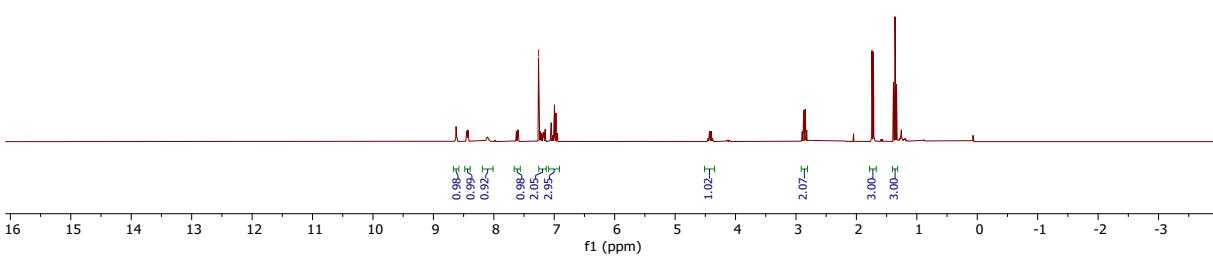
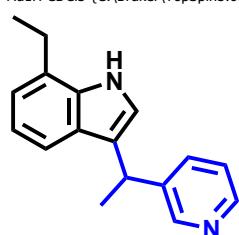


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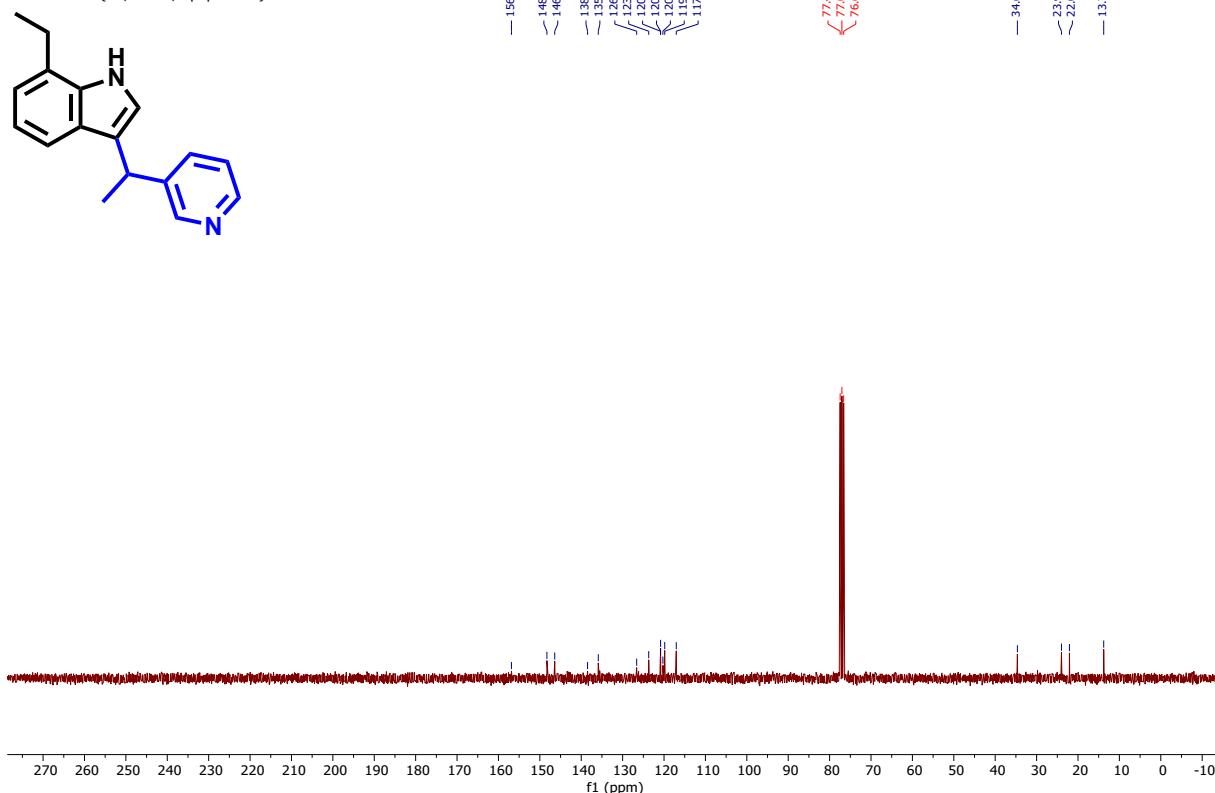
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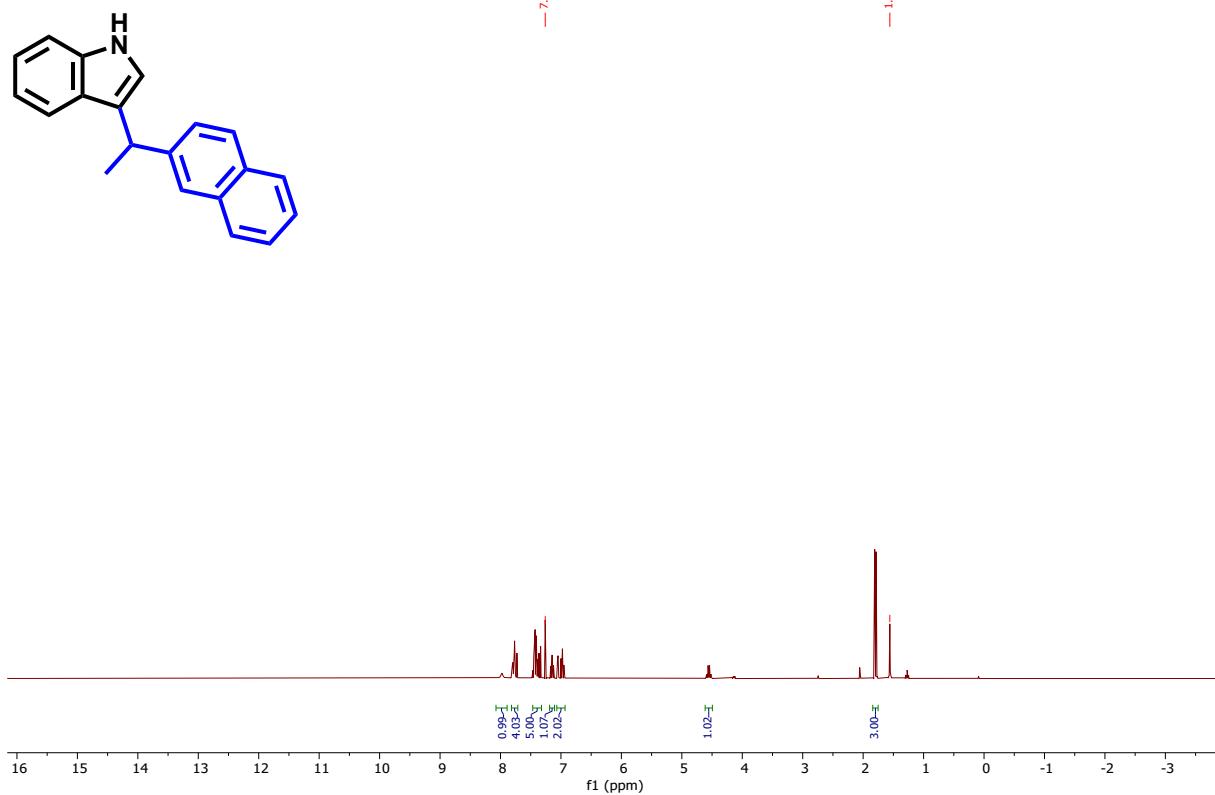
— 7.36 CDCl<sub>3</sub>



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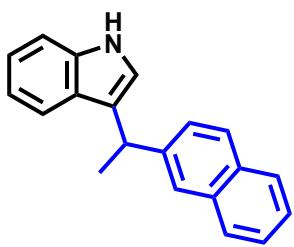
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Bei Zhou 21-1-2

C13CPD CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2107



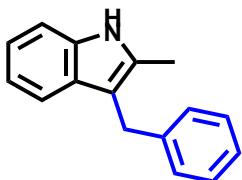
— 37.69

— 22.30

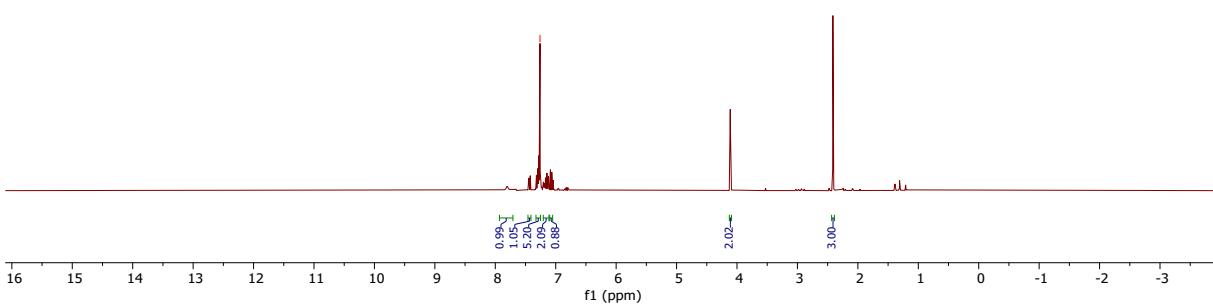
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Bei Zhou Bei 21-112

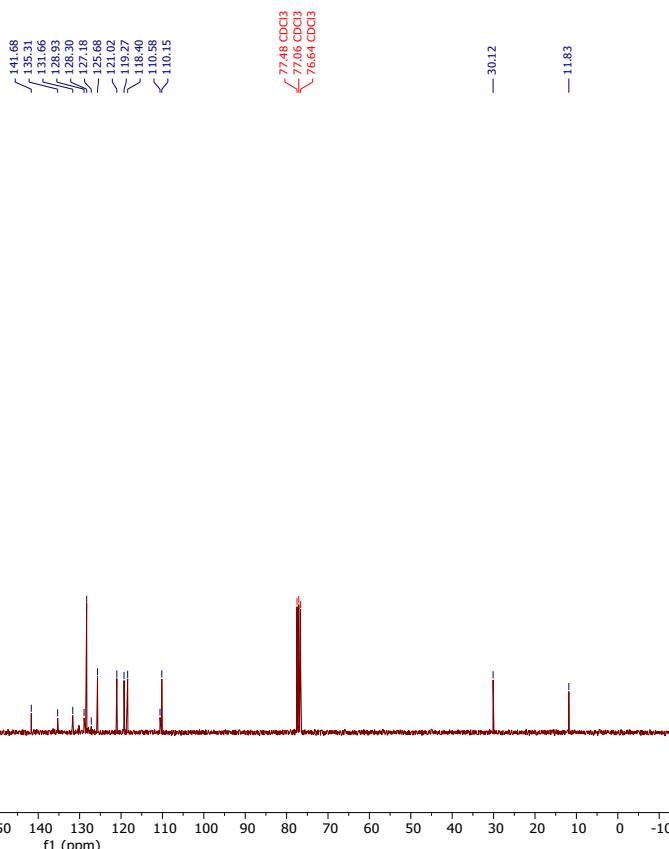
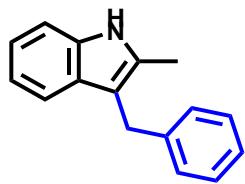
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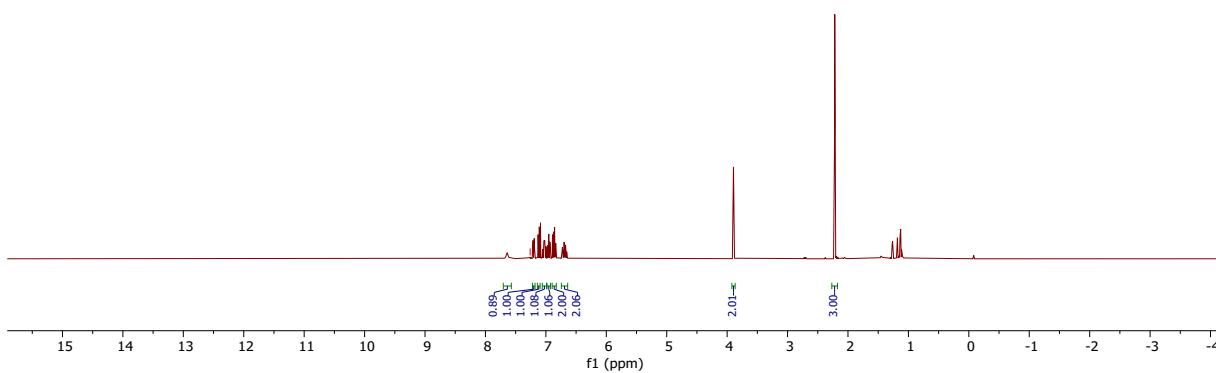
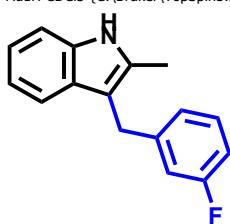
— 7.26 CDCl<sub>3</sub>



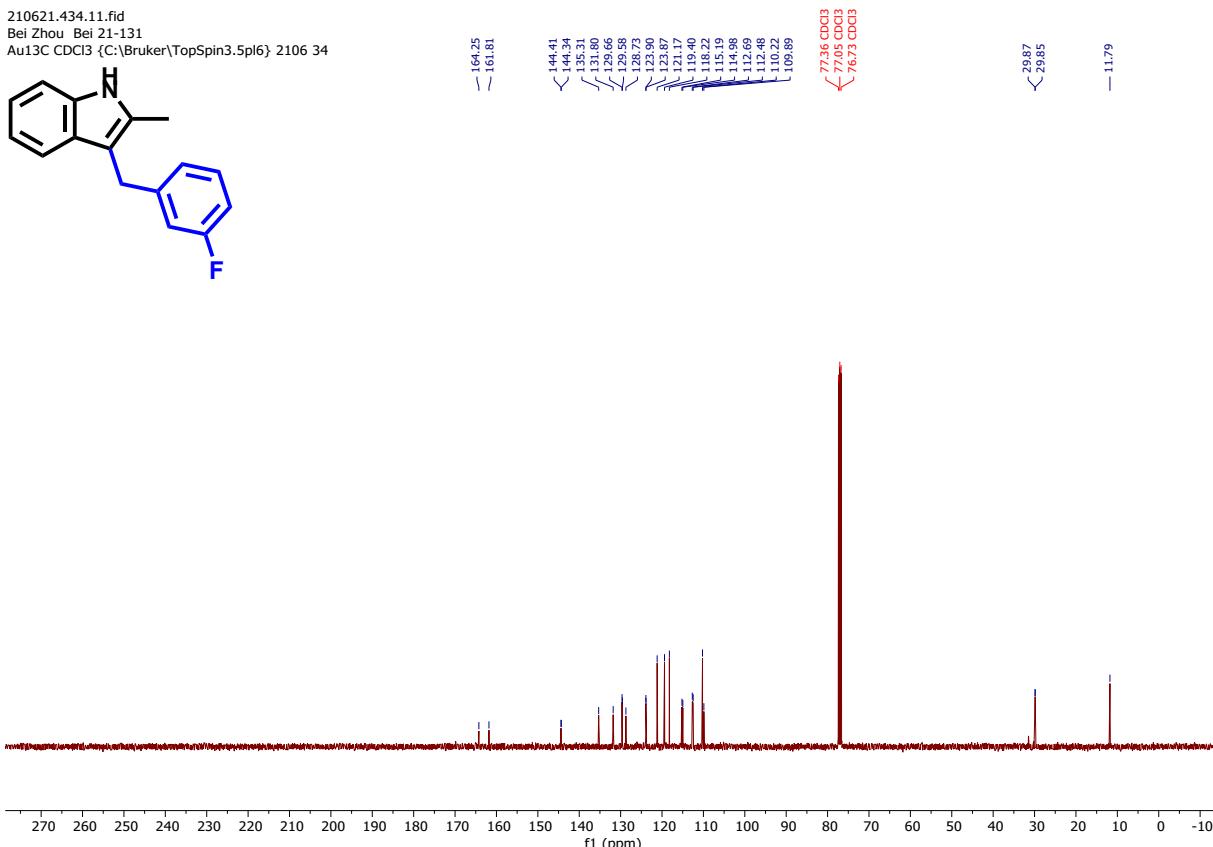
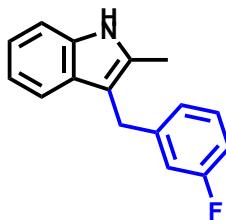
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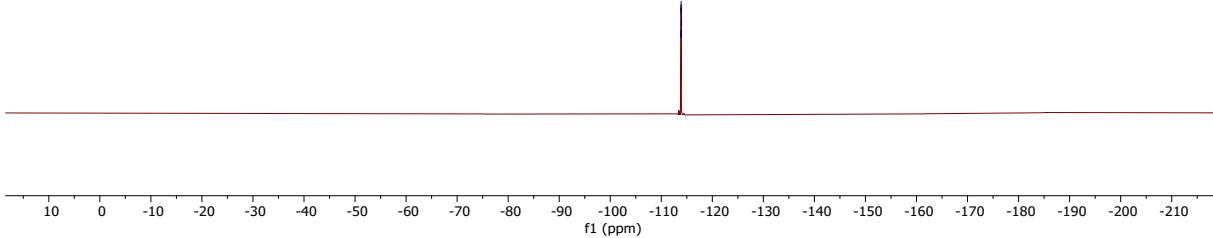
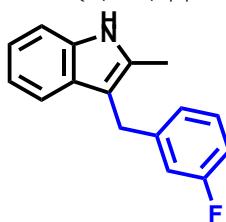
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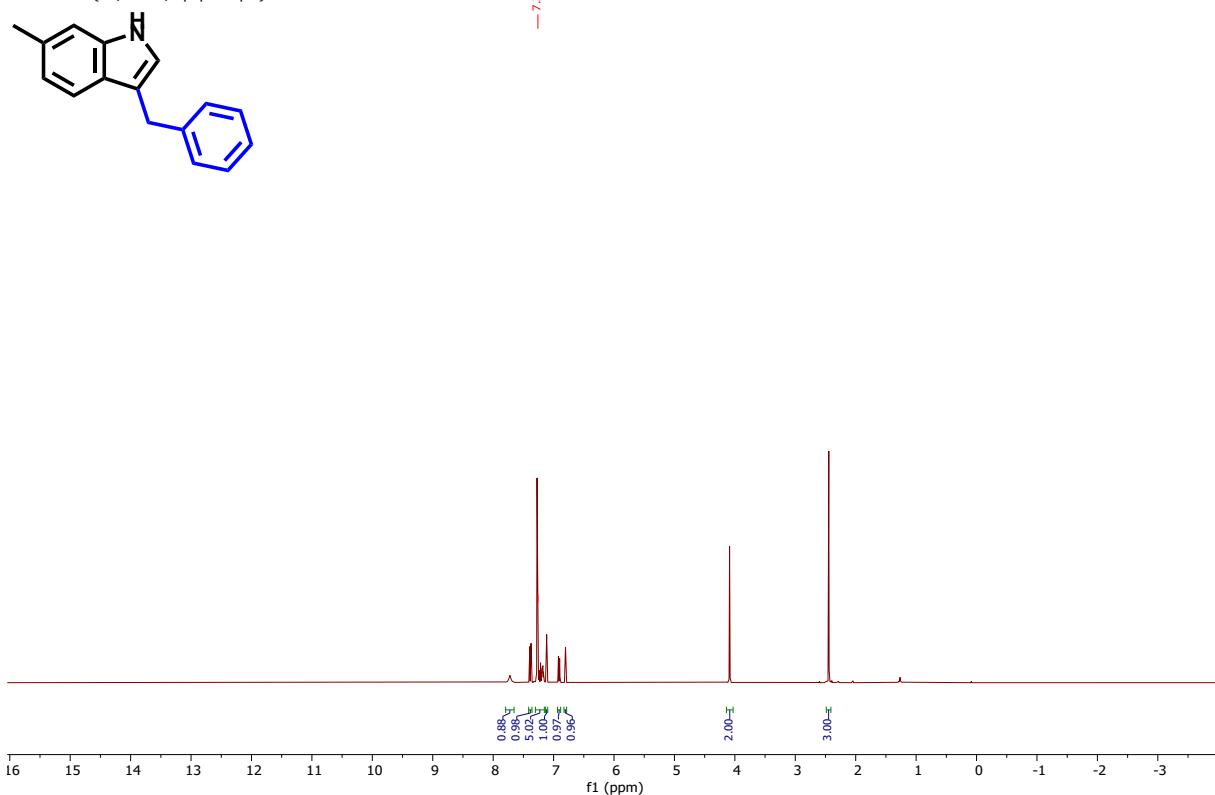
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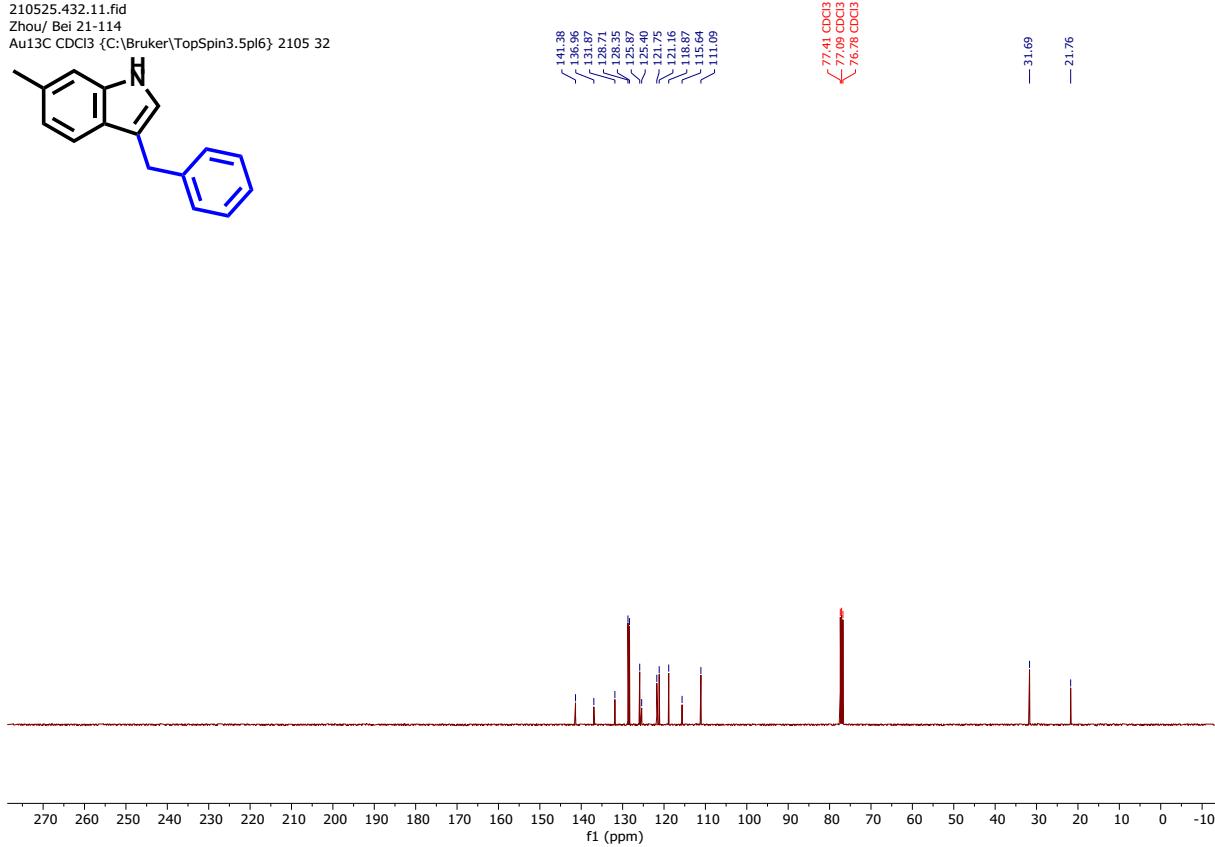
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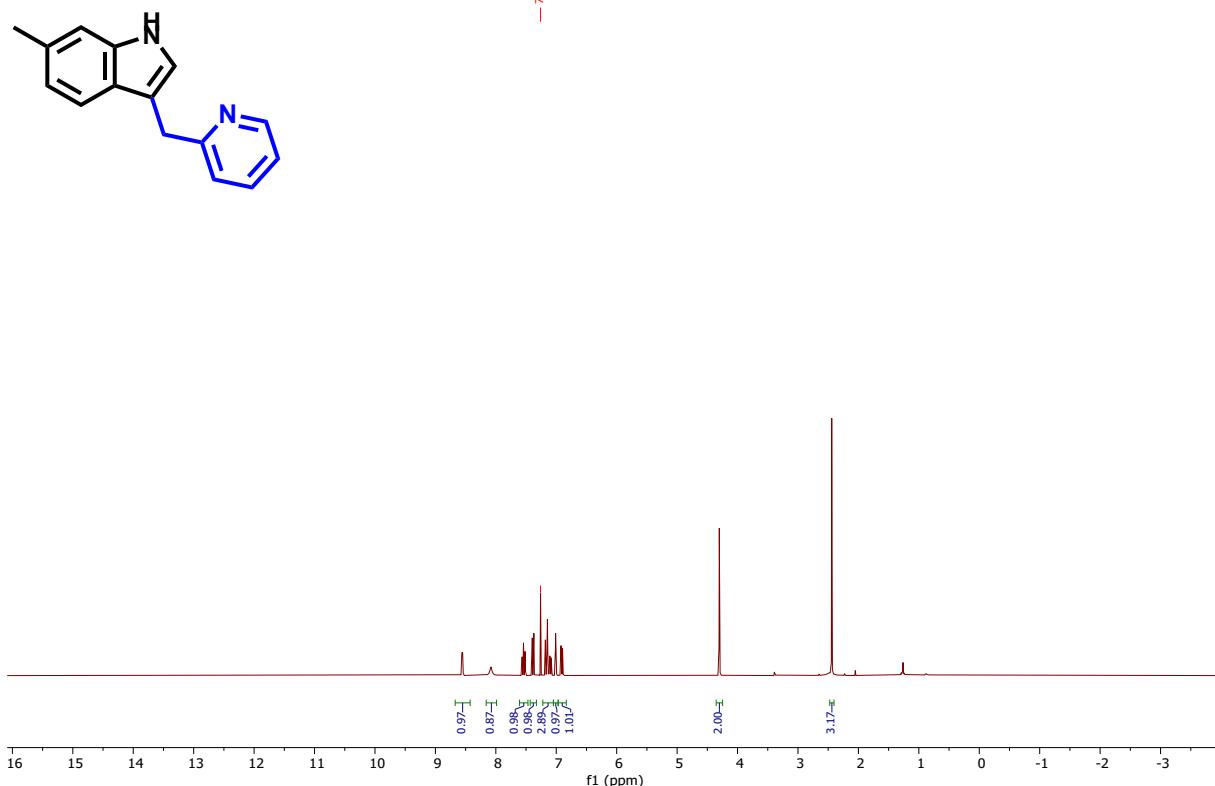
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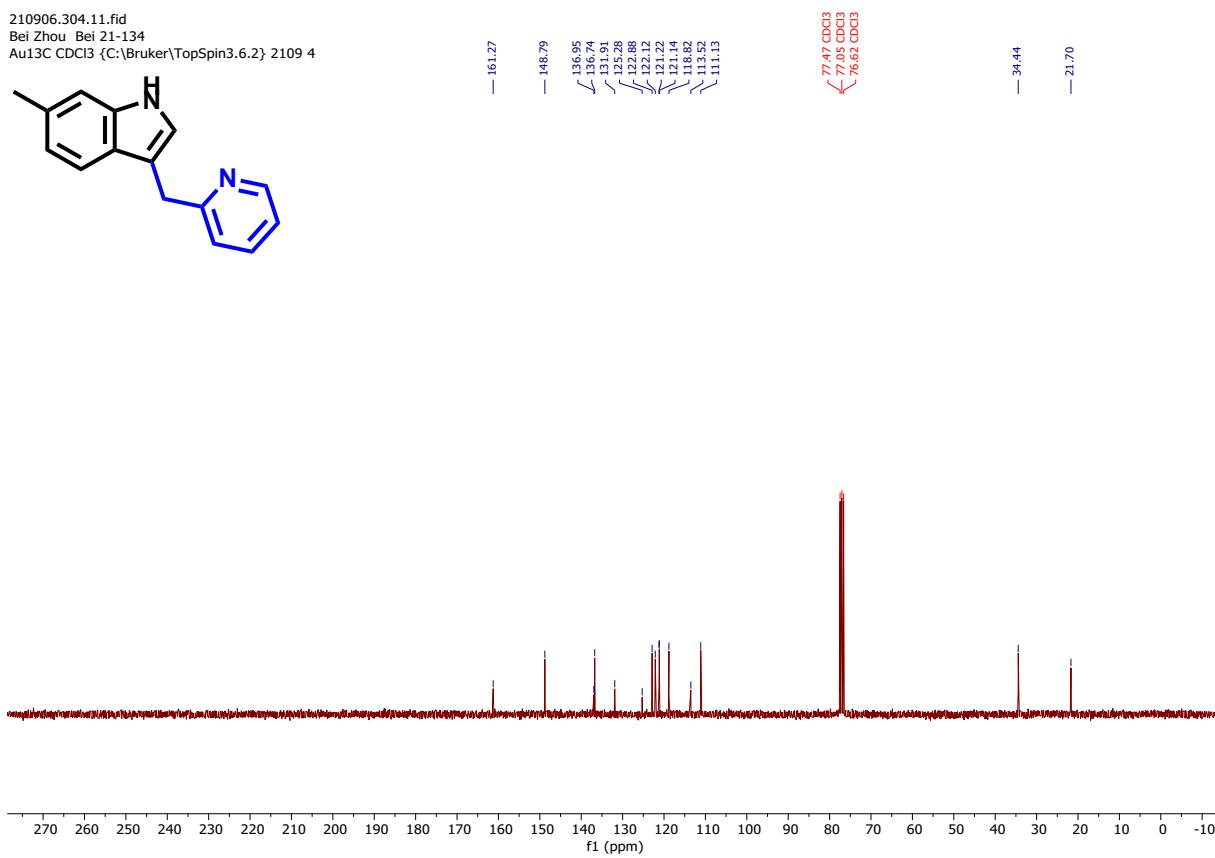
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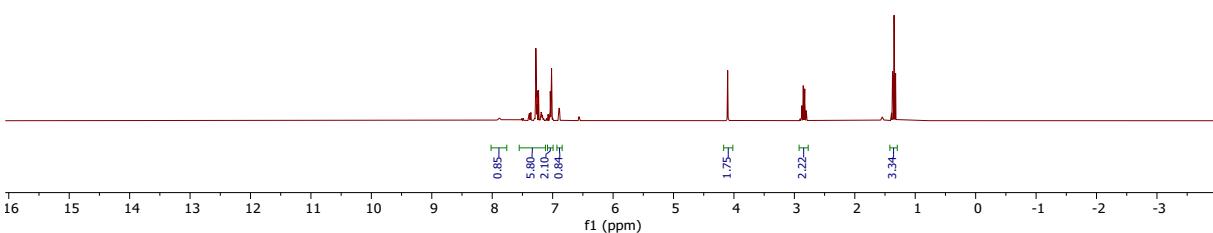
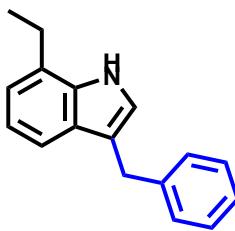
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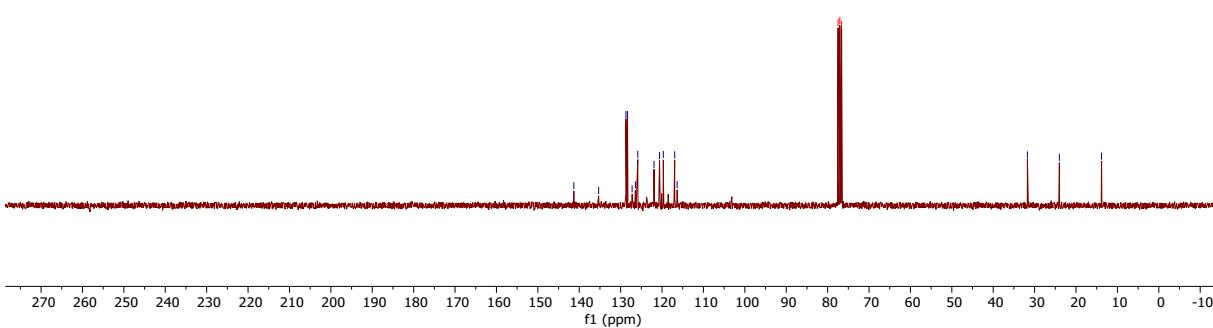
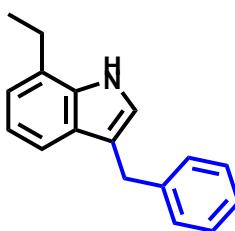
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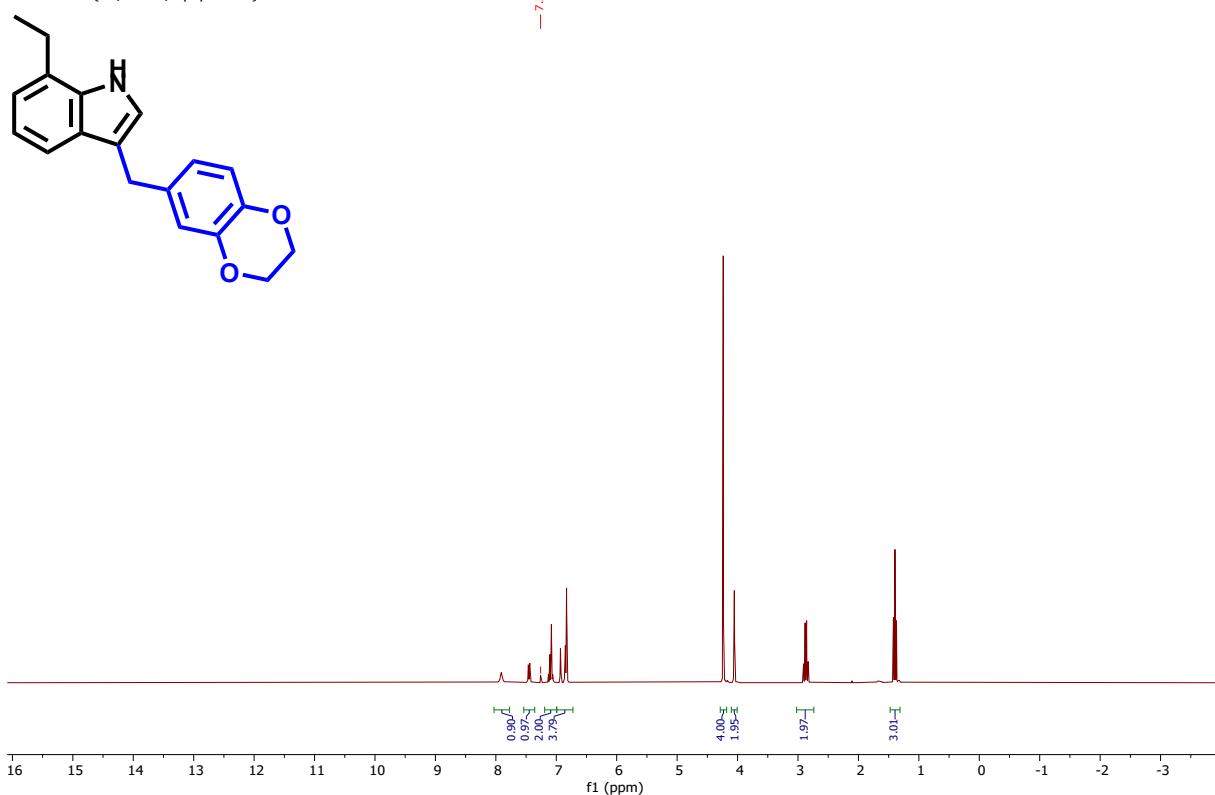
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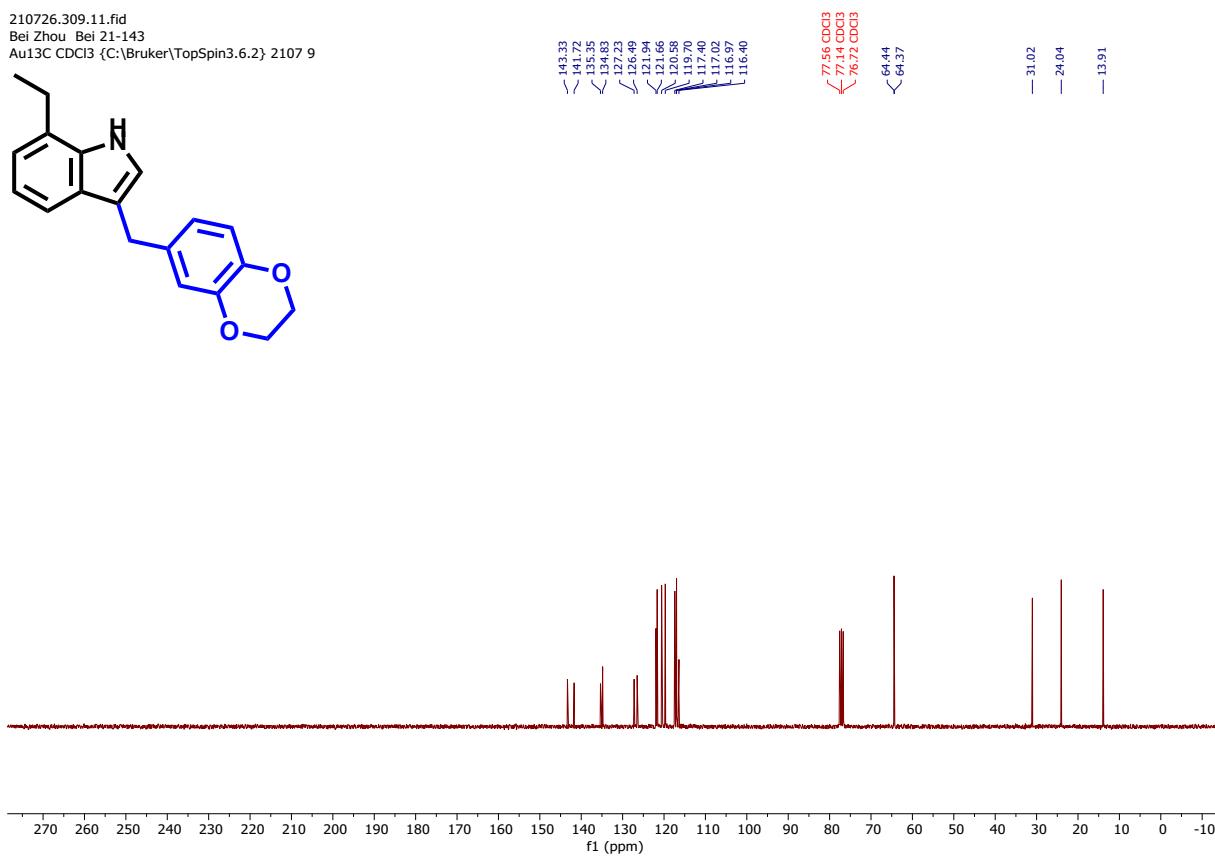
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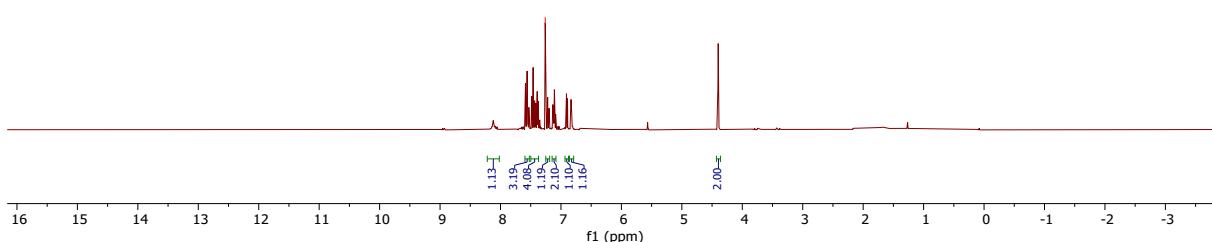
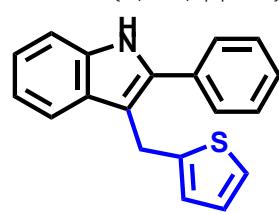
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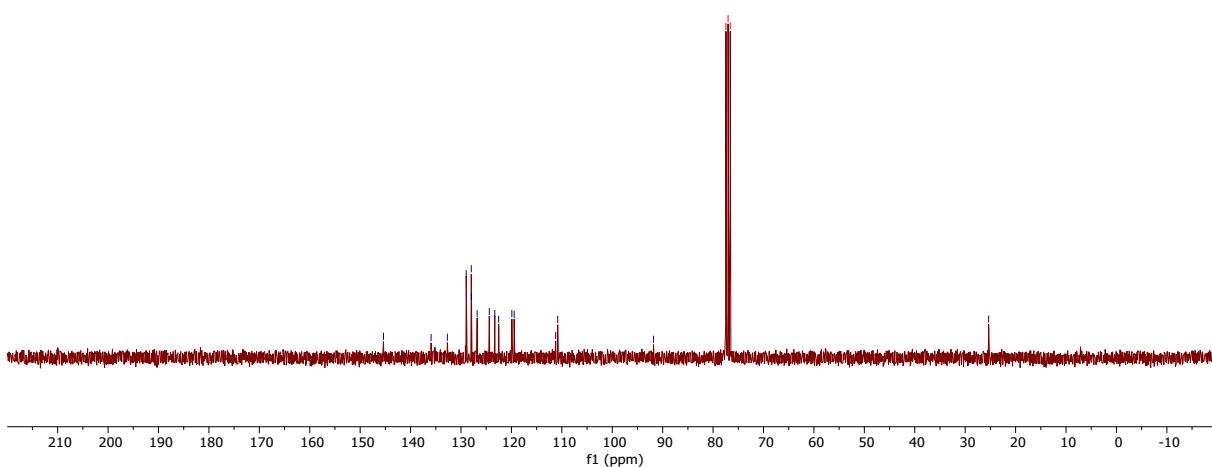
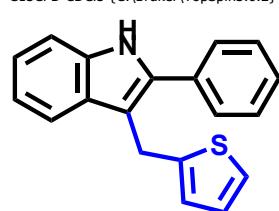
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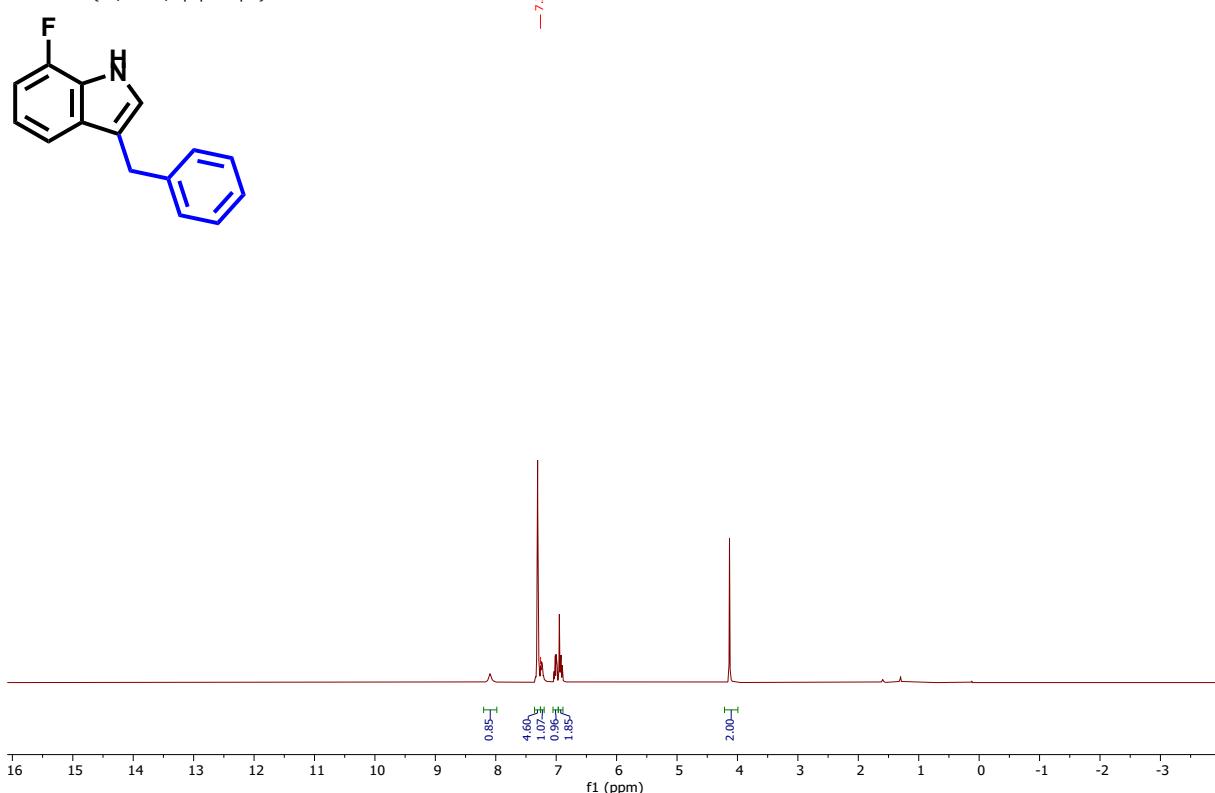
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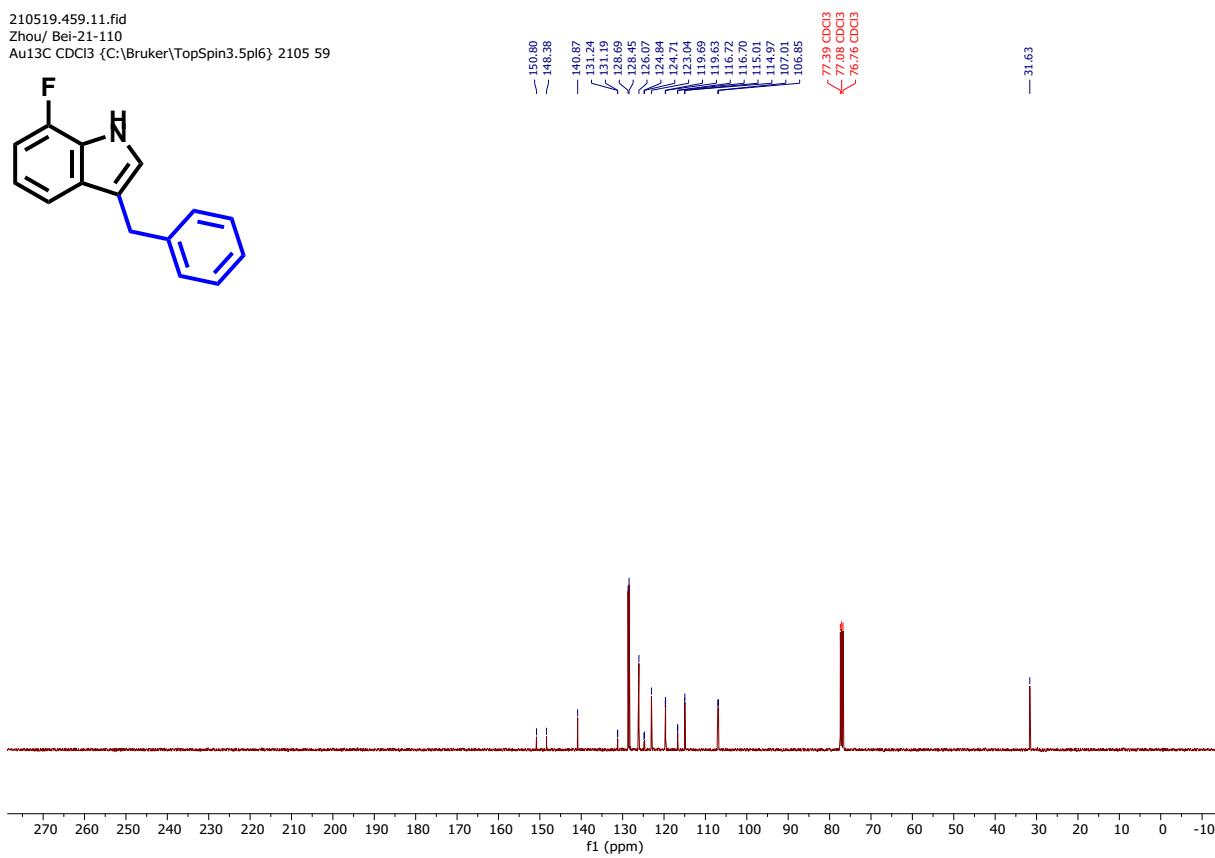
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C13CPD CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2107 44



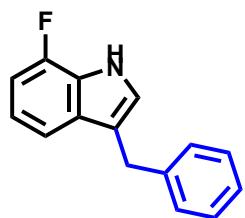
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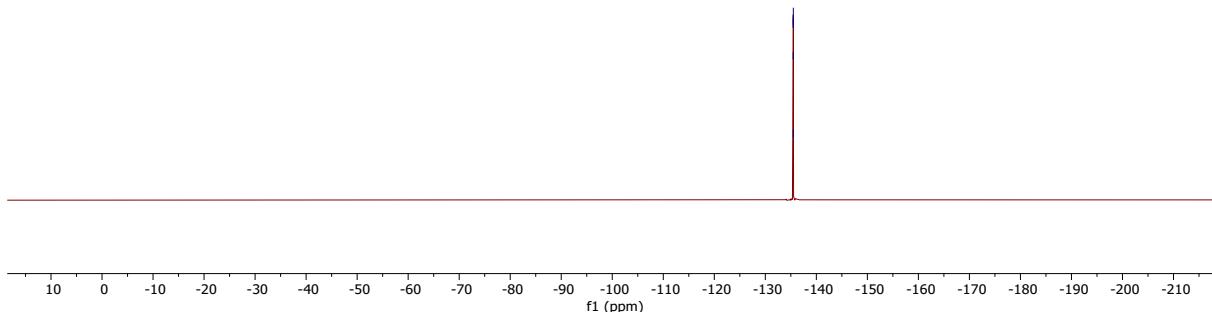
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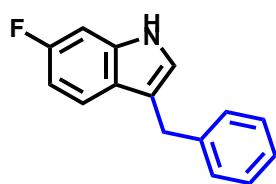
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Zhou/ Bei 21-110  
F19 CDCl<sub>3</sub> {C:\Bruker\TopSpin3.5pl6} 2105 26



-135.45  
-135.46  
-135.48  
-135.49  
-135.50

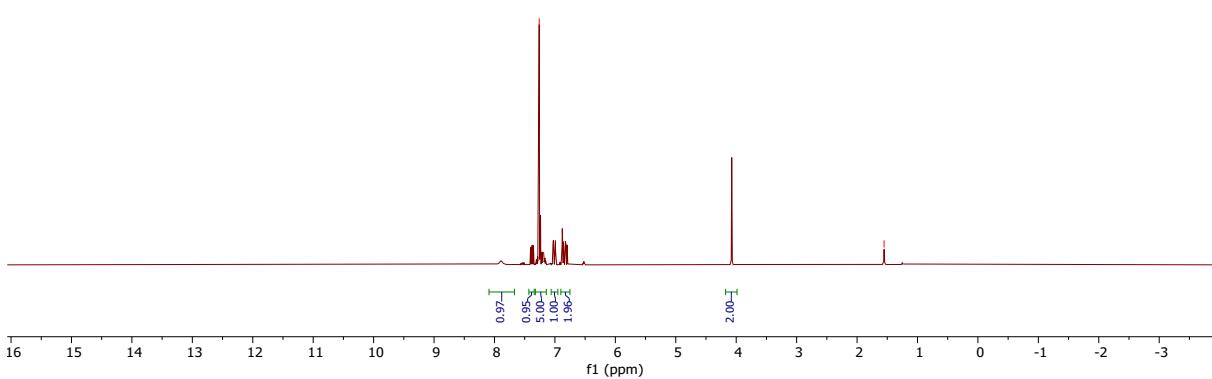


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Au1H CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 18

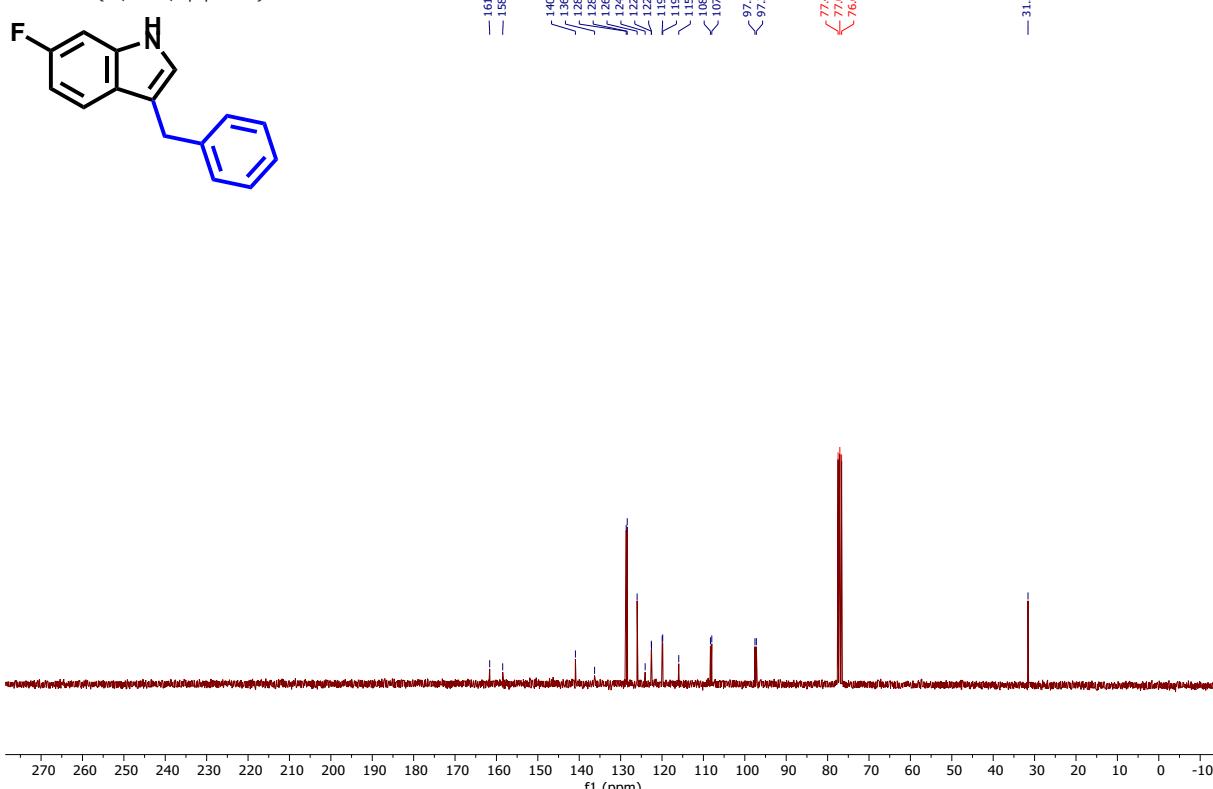


— 7.36 CDCl<sub>3</sub>

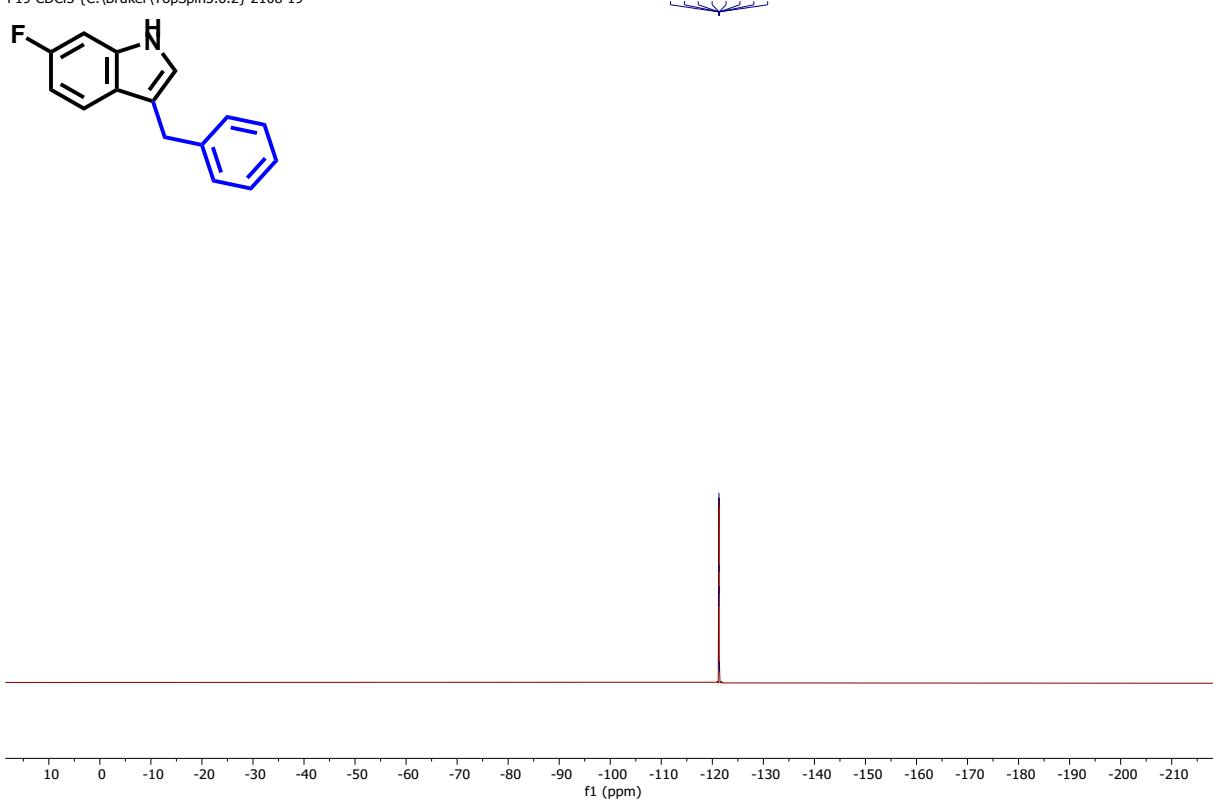
— 1.55 H<sub>2</sub>O



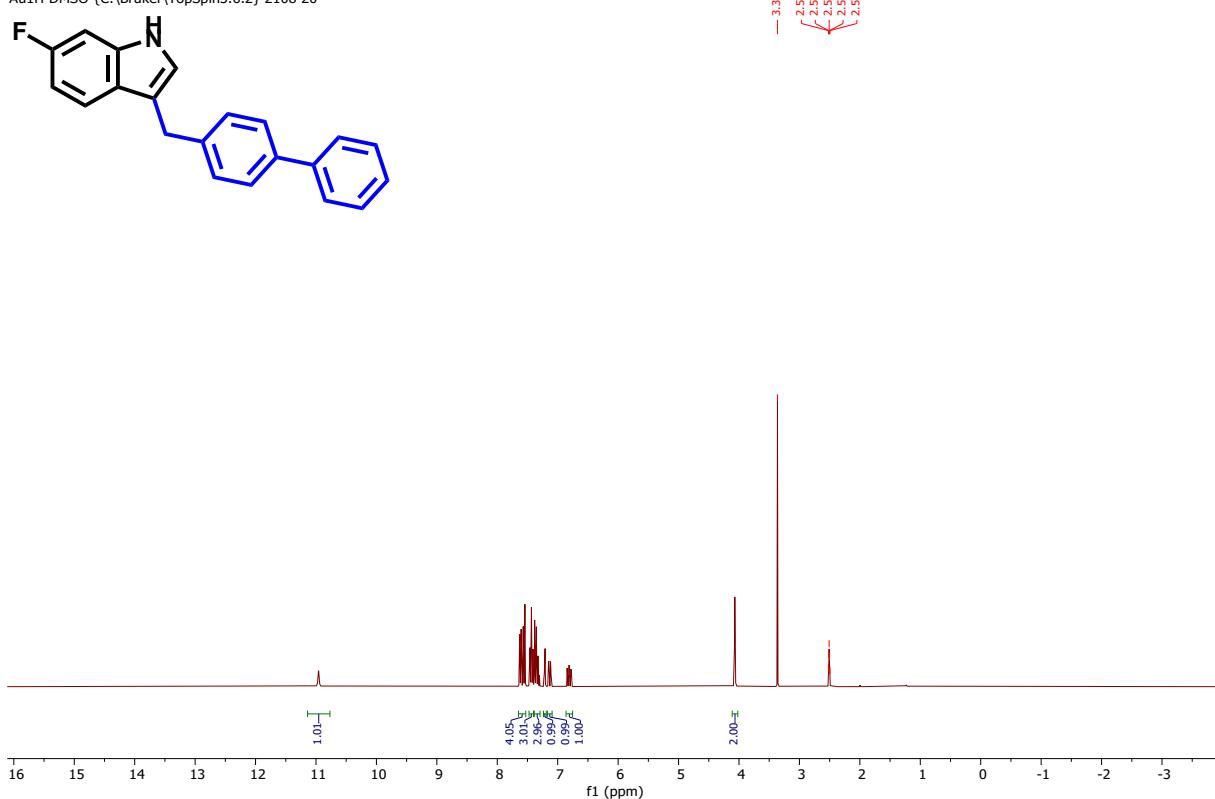
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Au13C CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 18



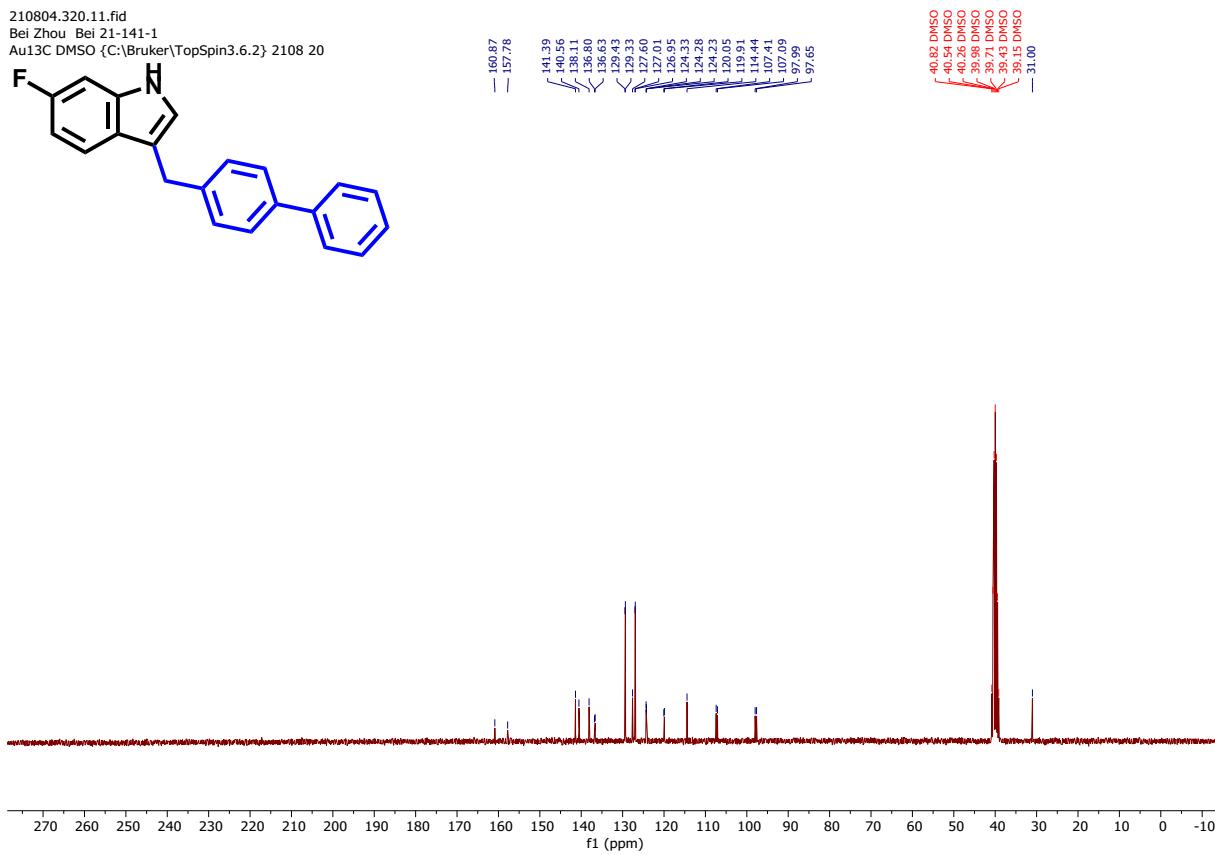
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F19 CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 19



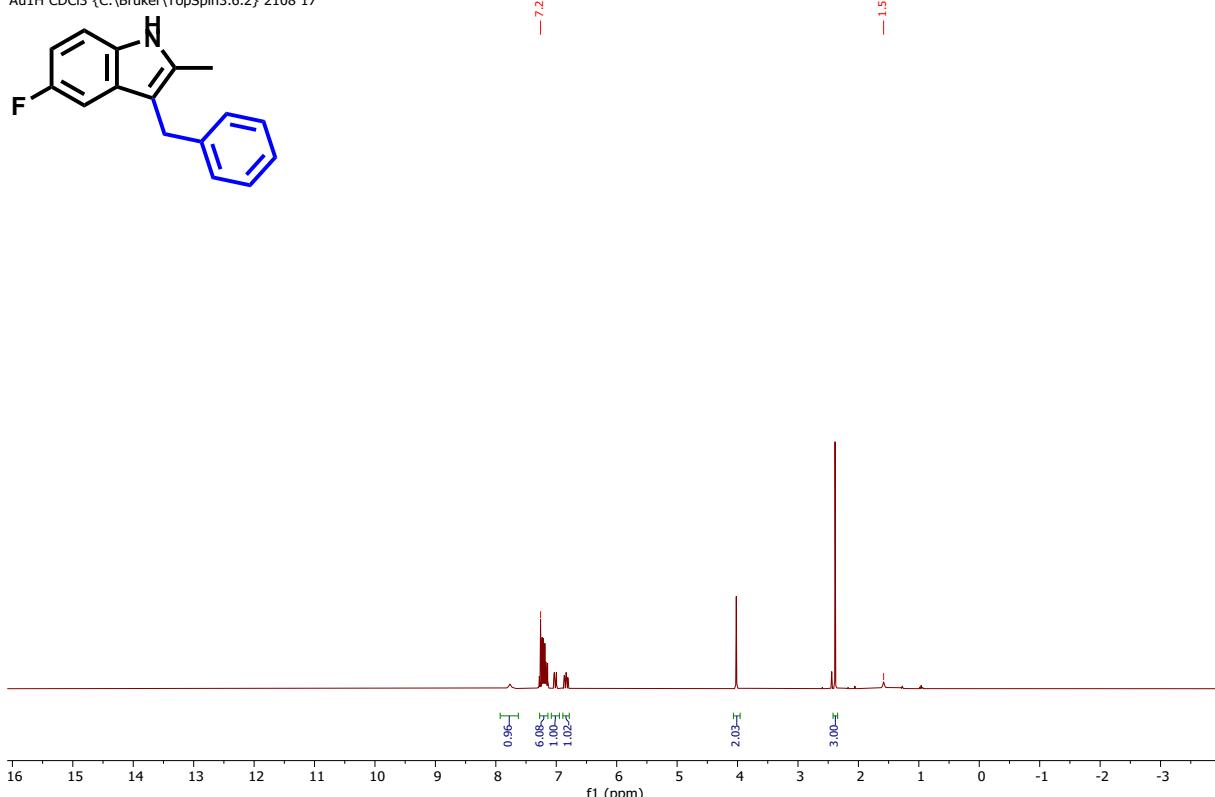
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Au1H DMSO {C:\Bruker\TopSpin3.6.2} 2108 20



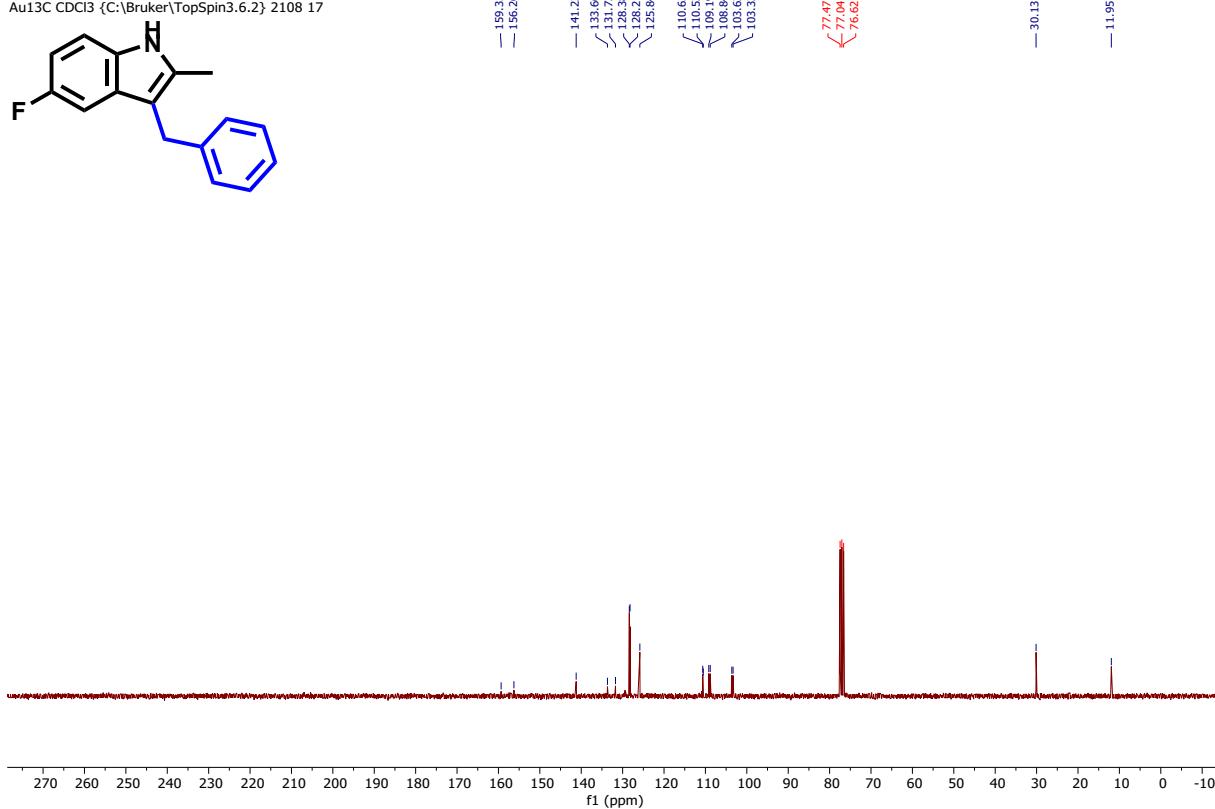
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Au13C DMSO {C:\Bruker\TopSpin3.6.2} 2108 20



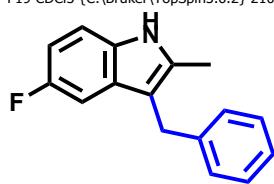
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Au1H CDCl3 {C:\Bruker\TopSpin3.6.2} 2108 17



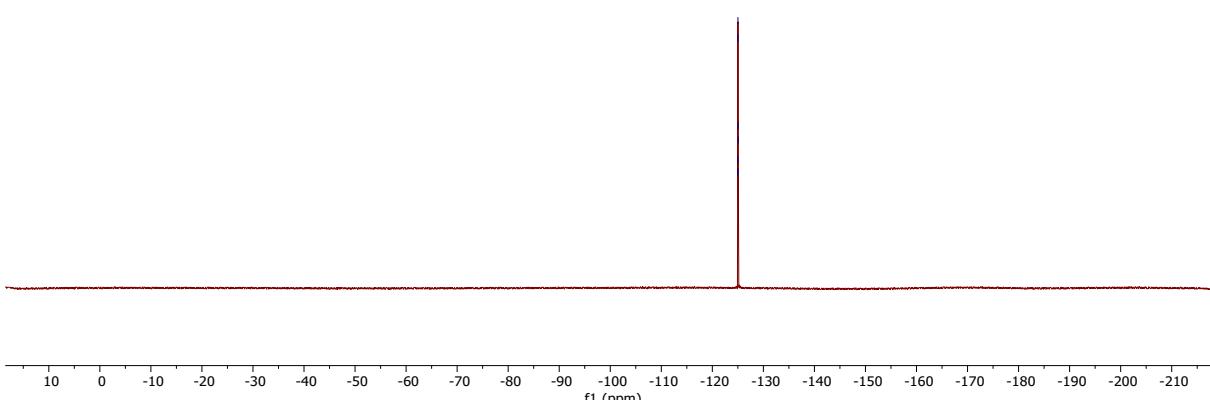
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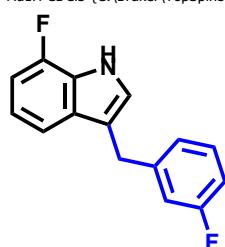
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F19 CDCl3 {C:\Bruker\TopSpin3.6.2} 2108 18



-124.98  
-125.00  
-125.02  
-125.03  
-125.05  
-125.07

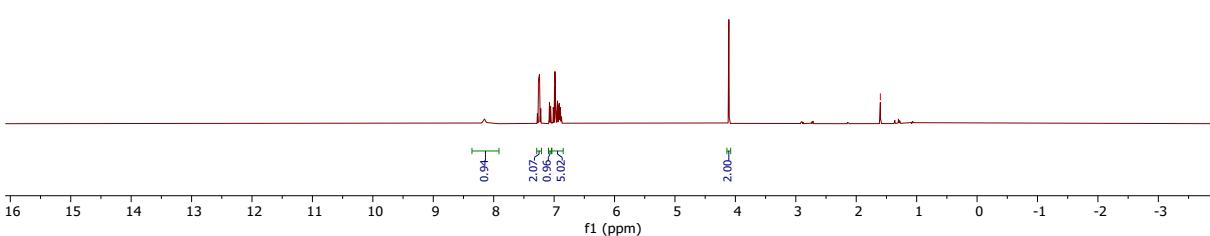


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Au1H CDCl3 {C:\Bruker\TopSpin3.5pl6} 2106 18

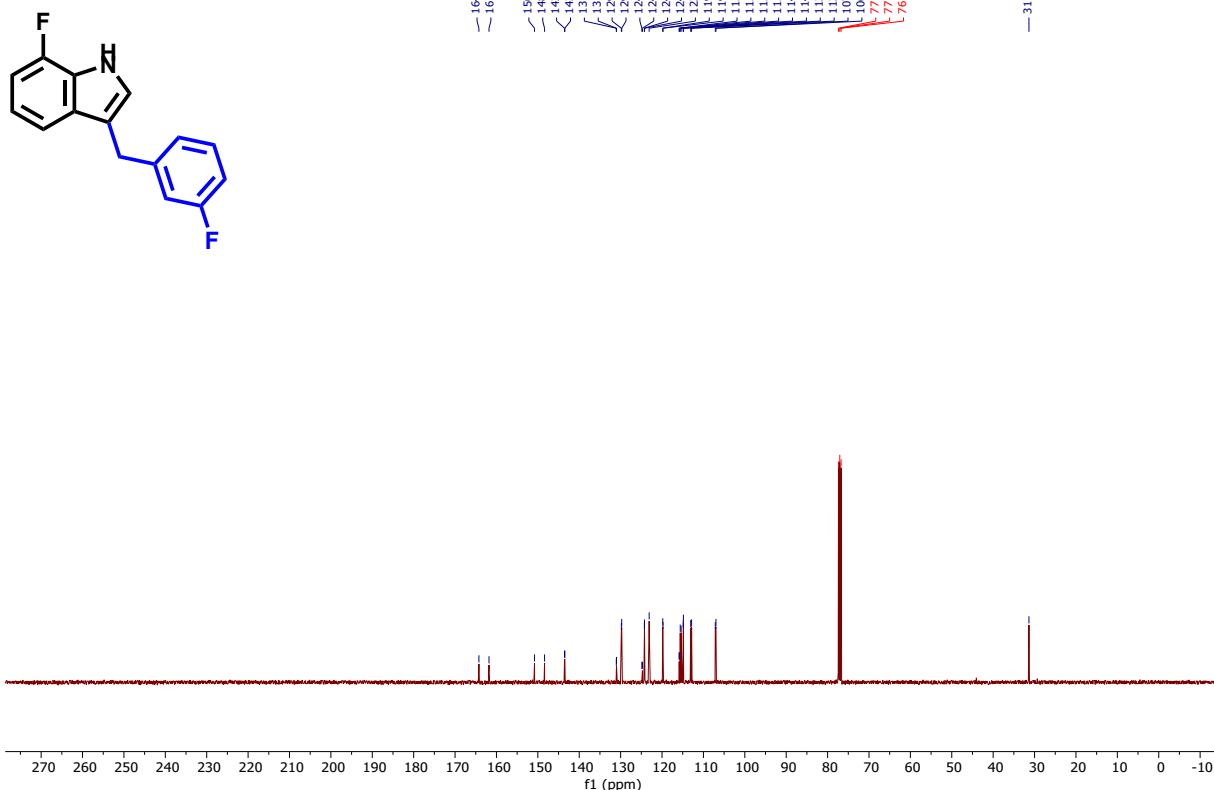


-7.36 CDCl3

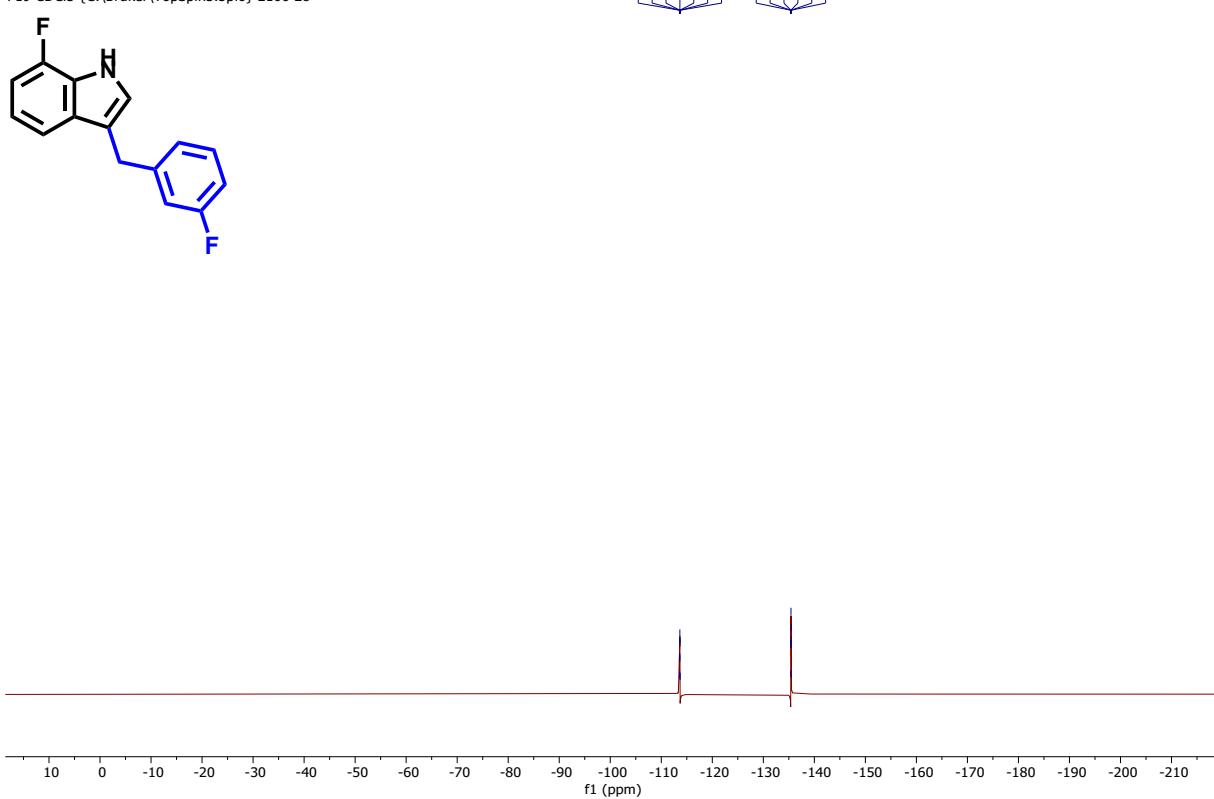
-1.60 H2O



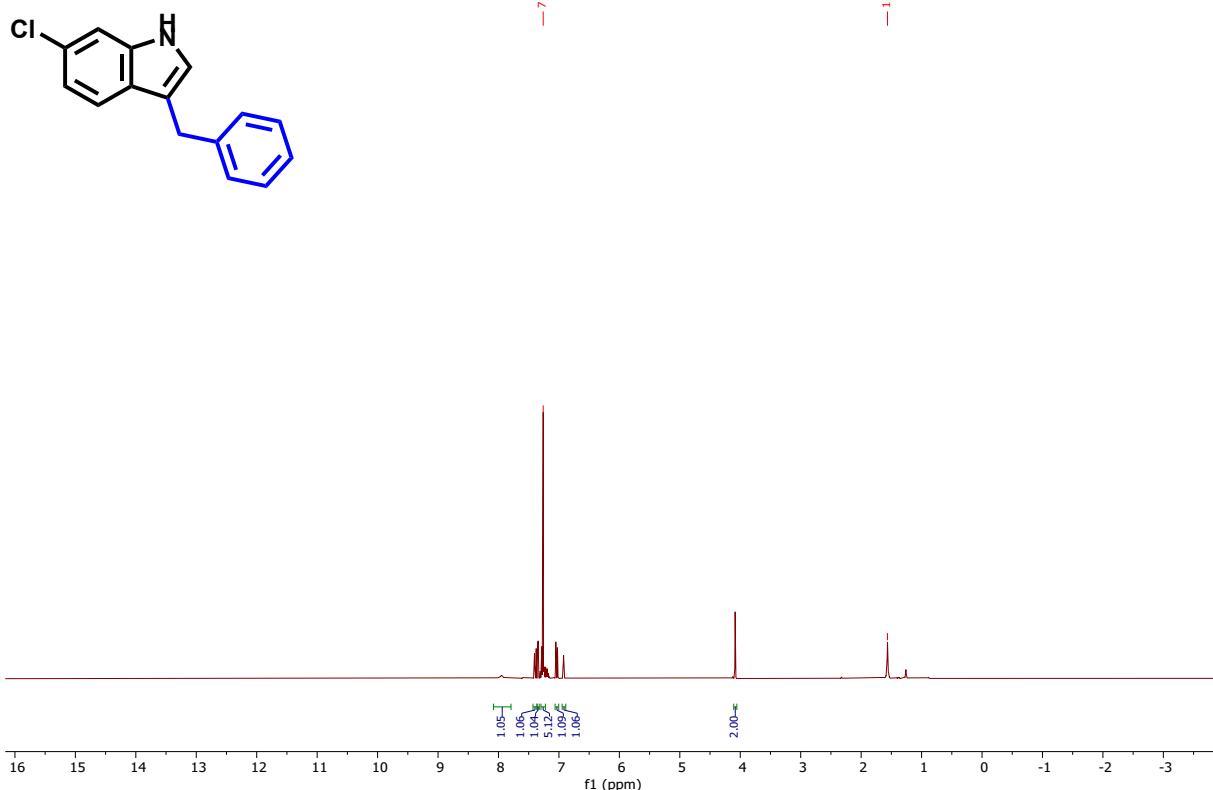
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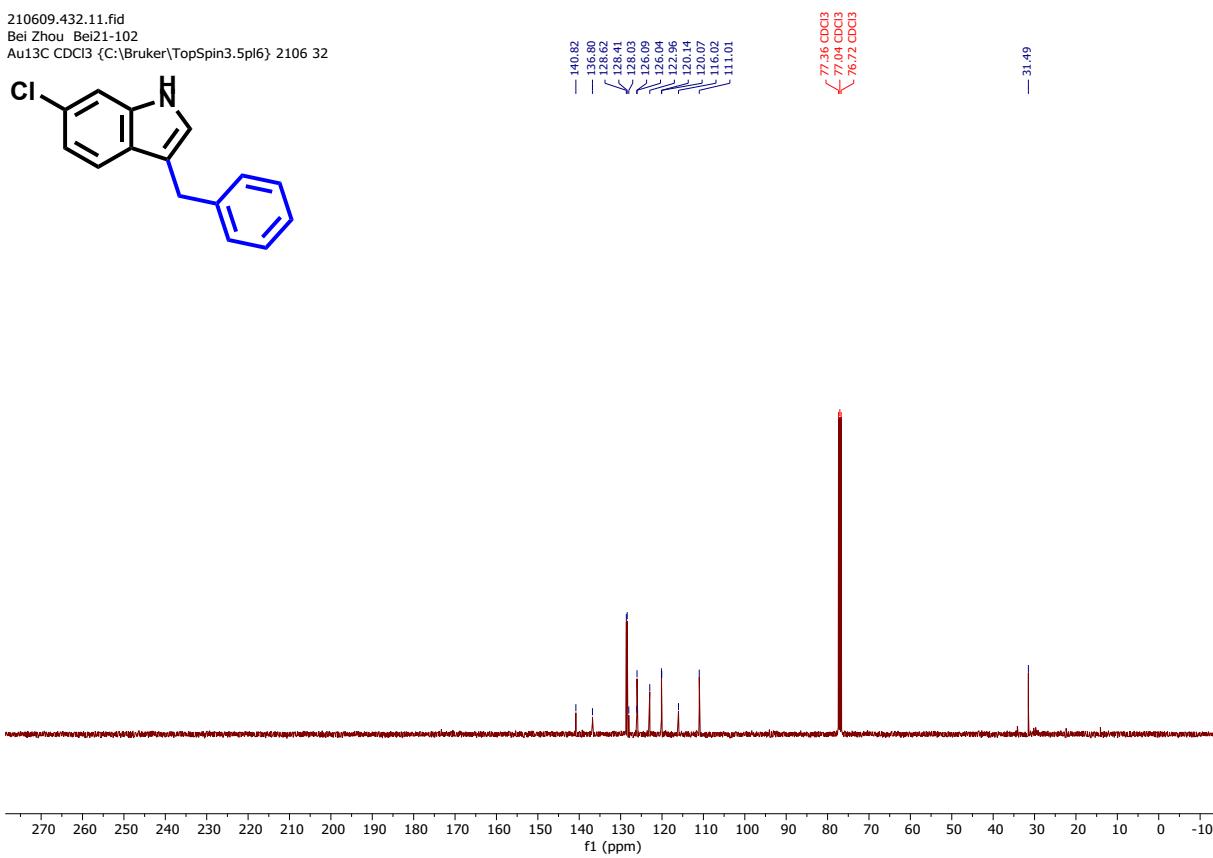
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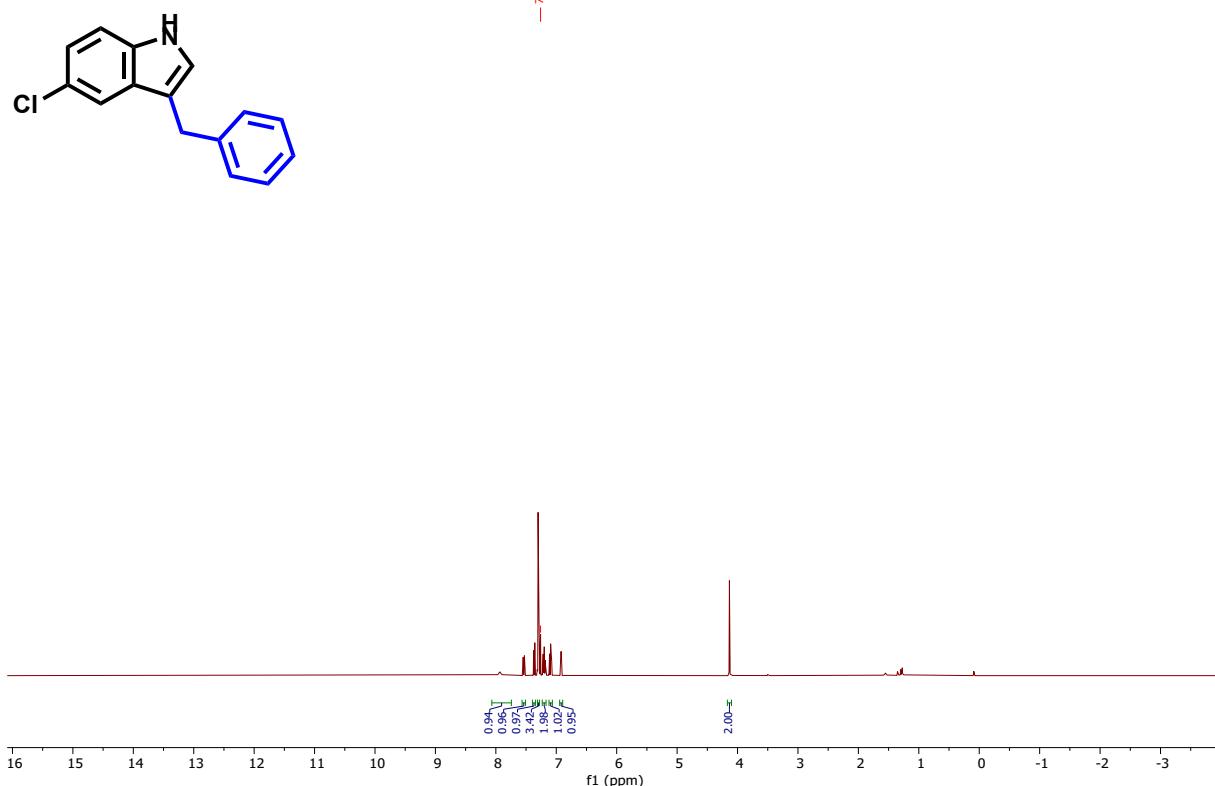
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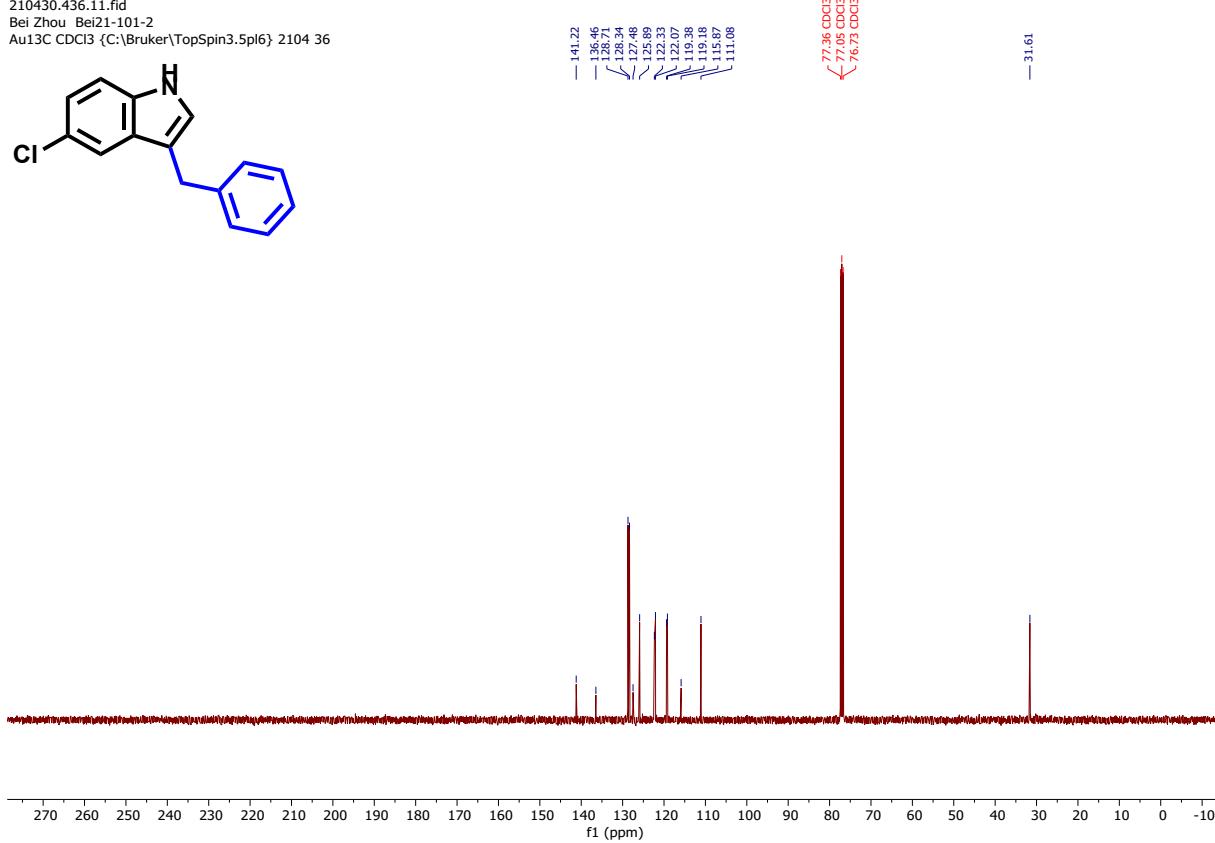
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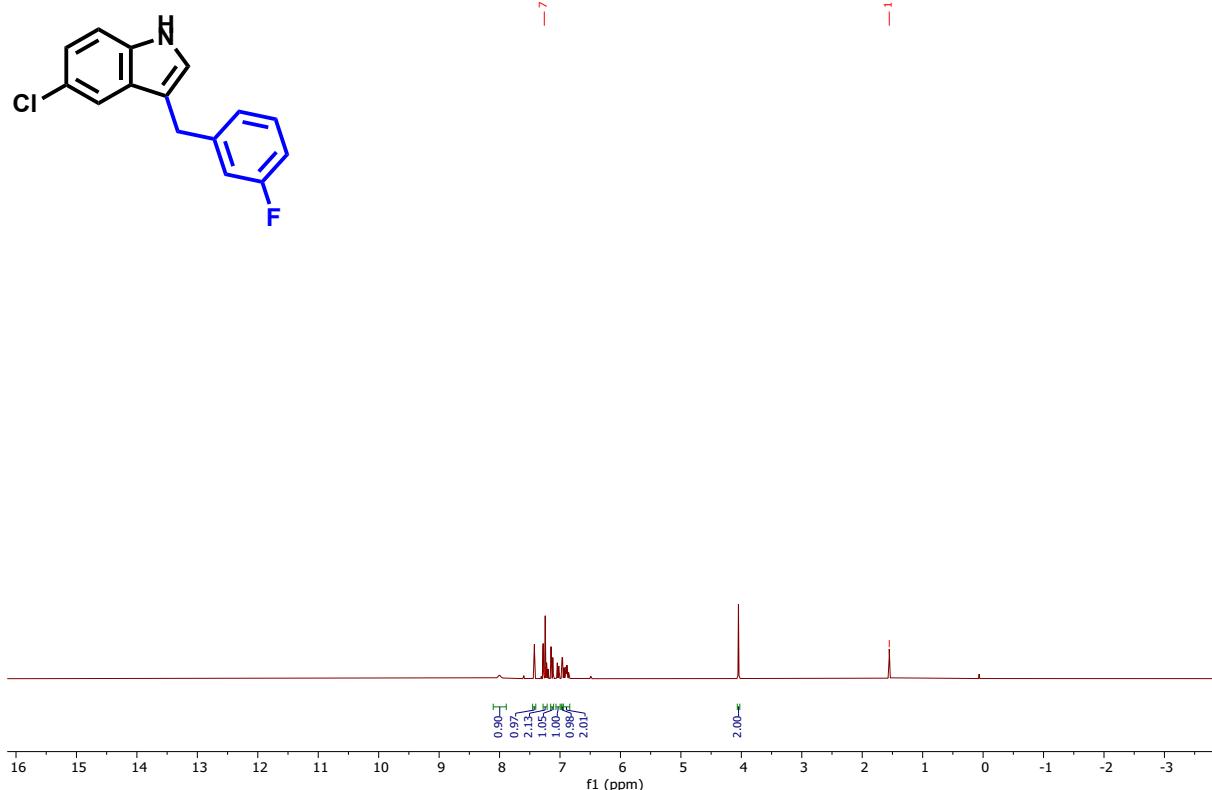
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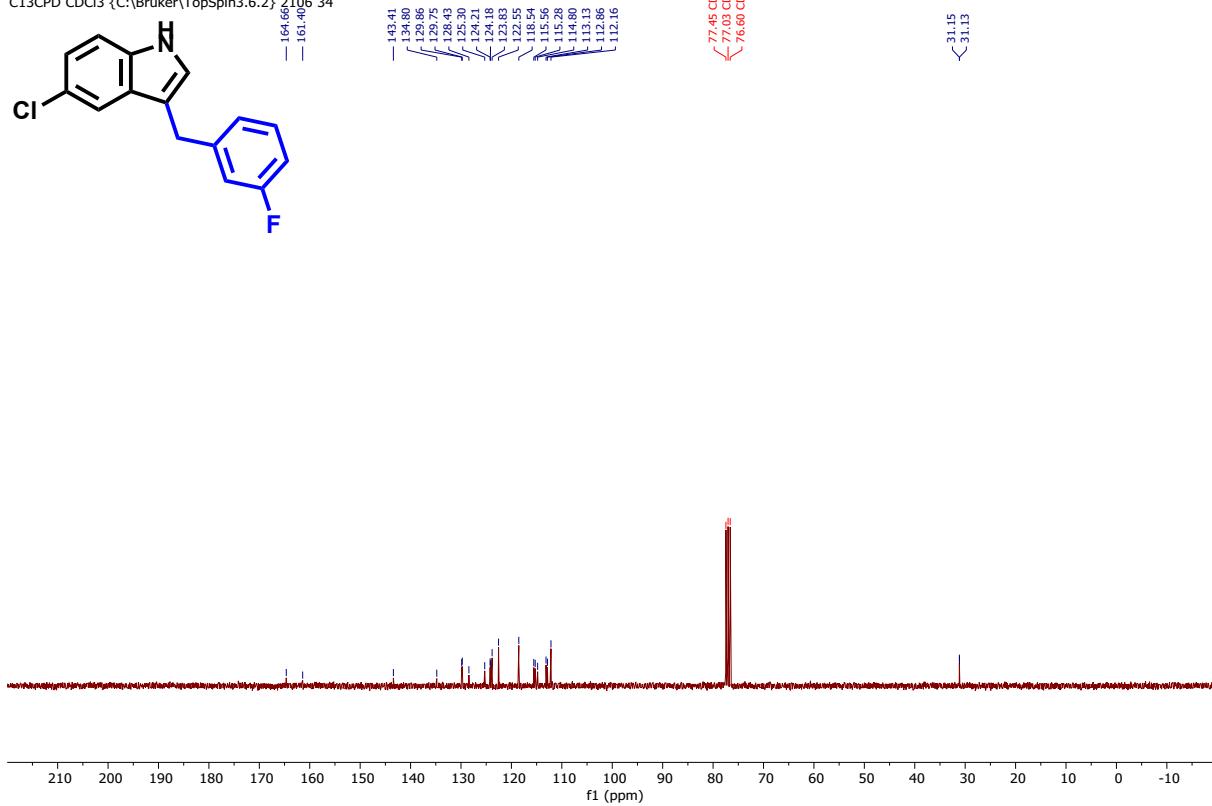
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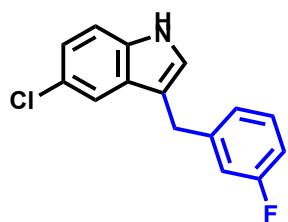
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PROTON CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2106 34



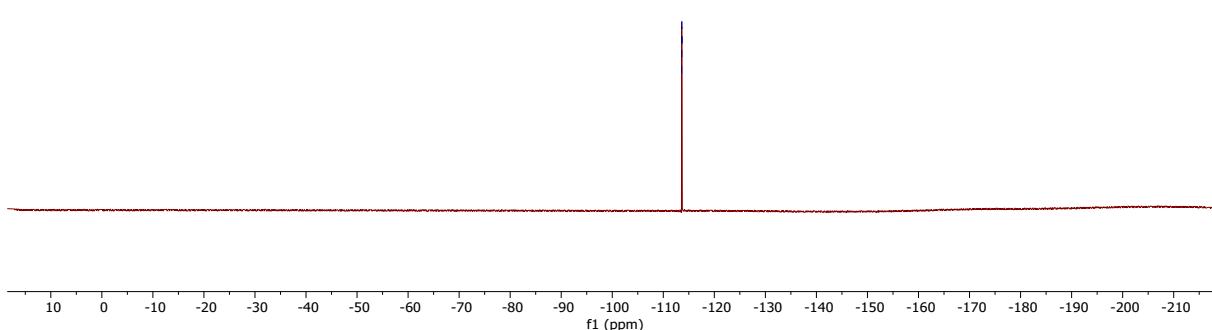
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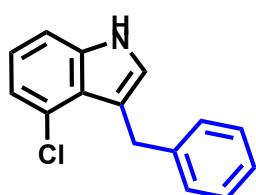
210630.f352.10.fid  
Bei Zhou Bel 21-132  
F19 CDCl3 {C:\Bruker\TopSpin3.6.2} 2106 52



-13.60  
-13.62  
-13.63  
-13.64  
-13.65  
-13.66  
-13.67



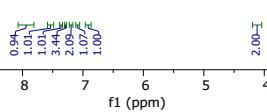
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Bei Zhou Bel 21-111-1  
PROTON CDCl3 {C:\Bruker\TopSpin3.6.2} 2108 38



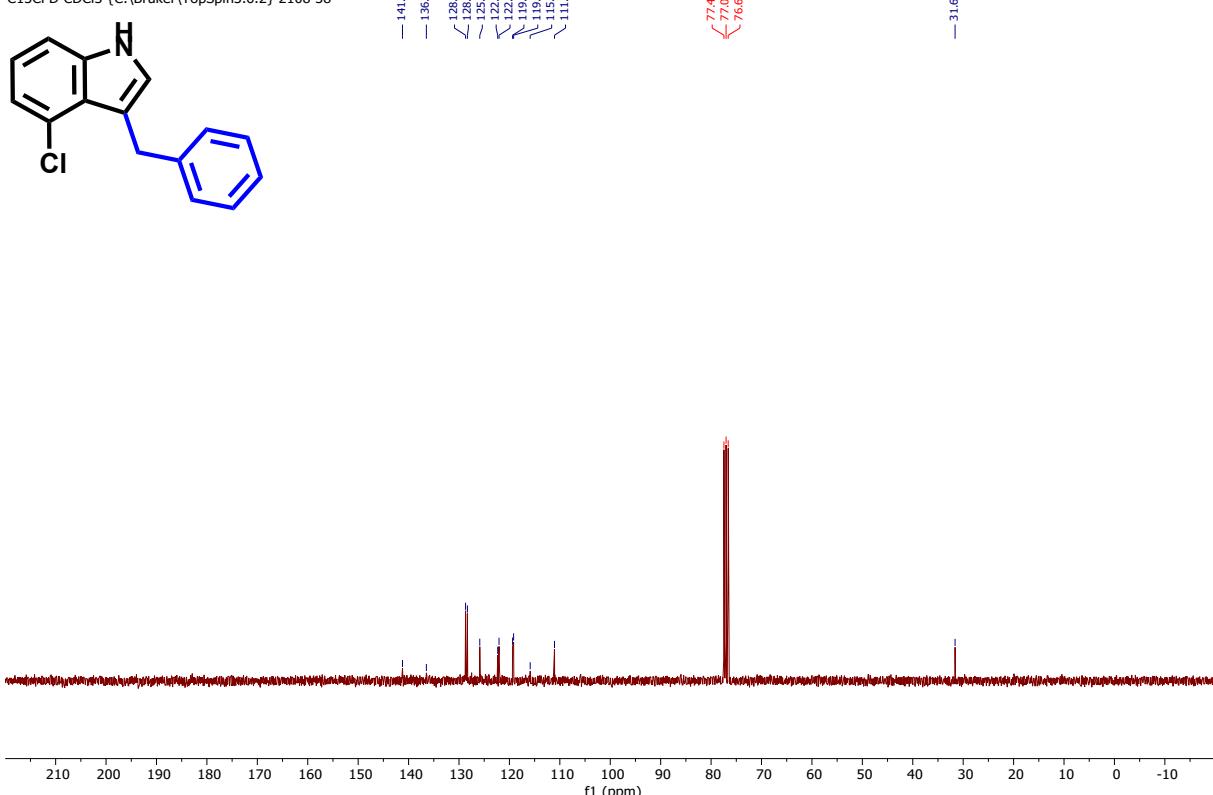
— 7.26 CDCl3

— 1.56 H<sub>2</sub>O

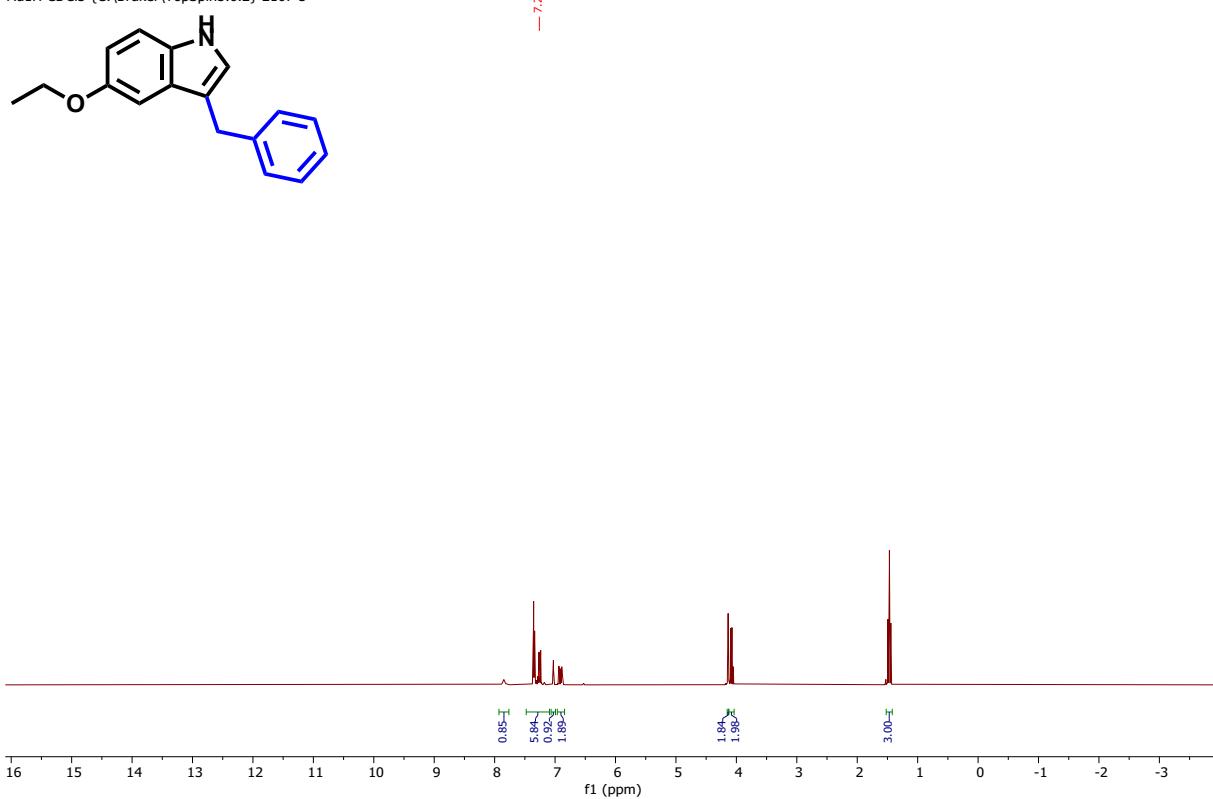
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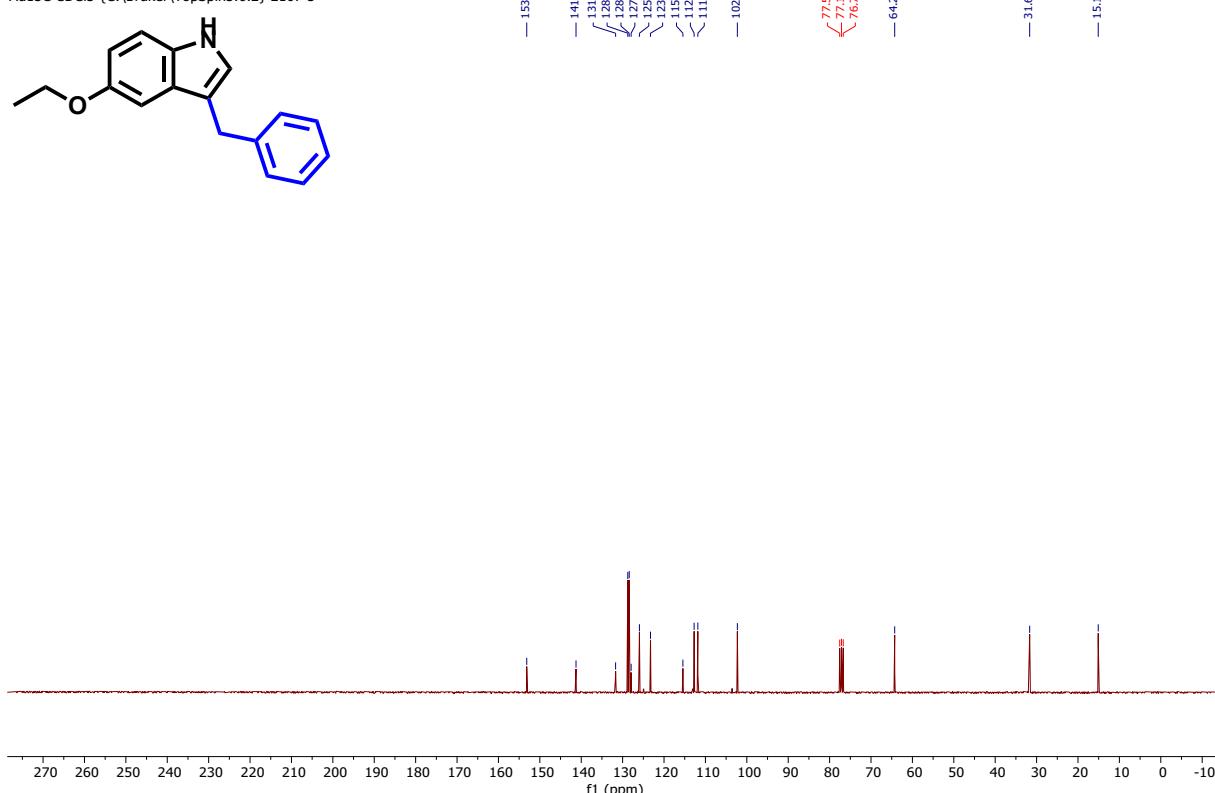
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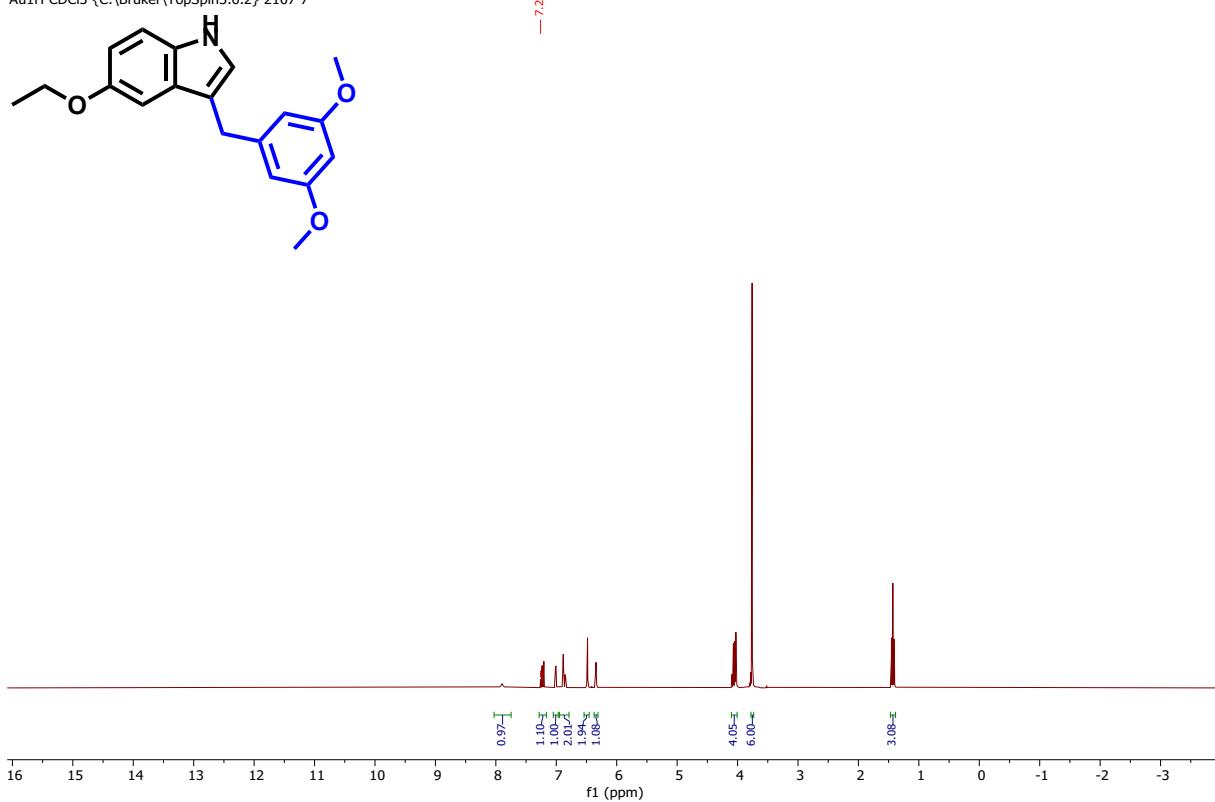
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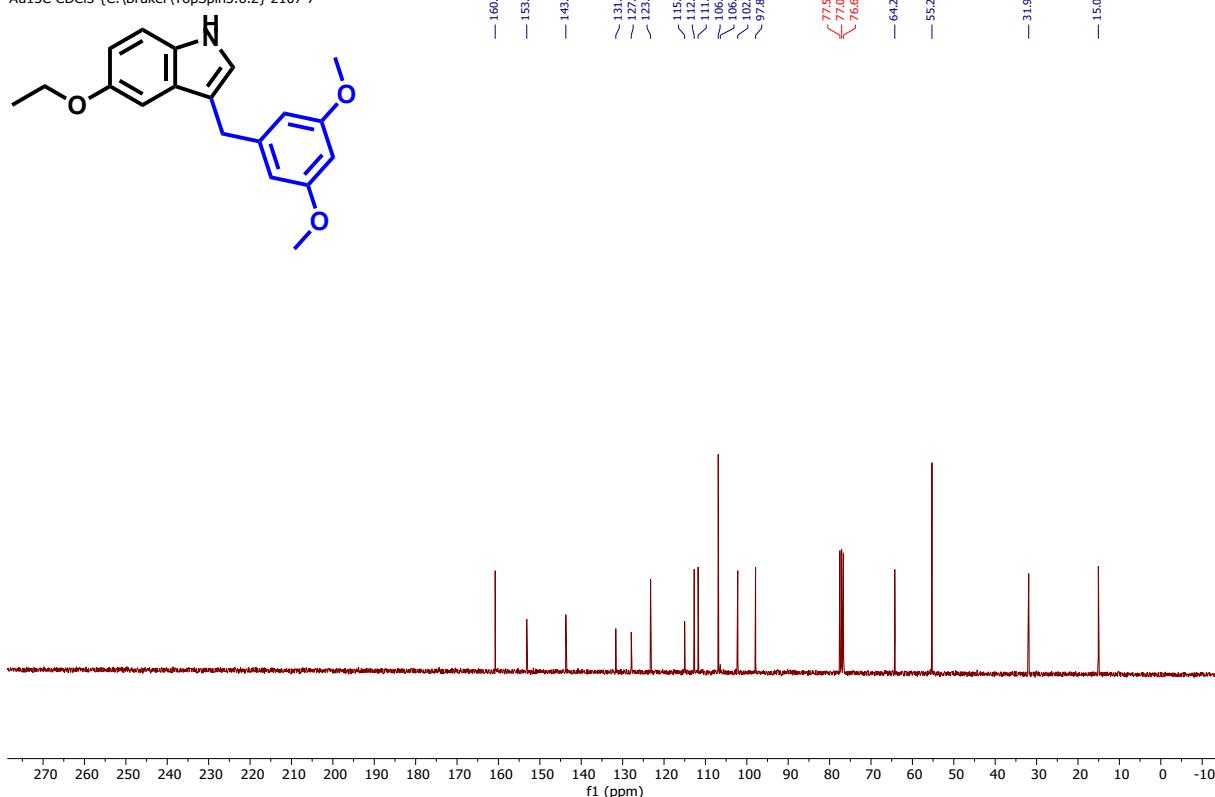
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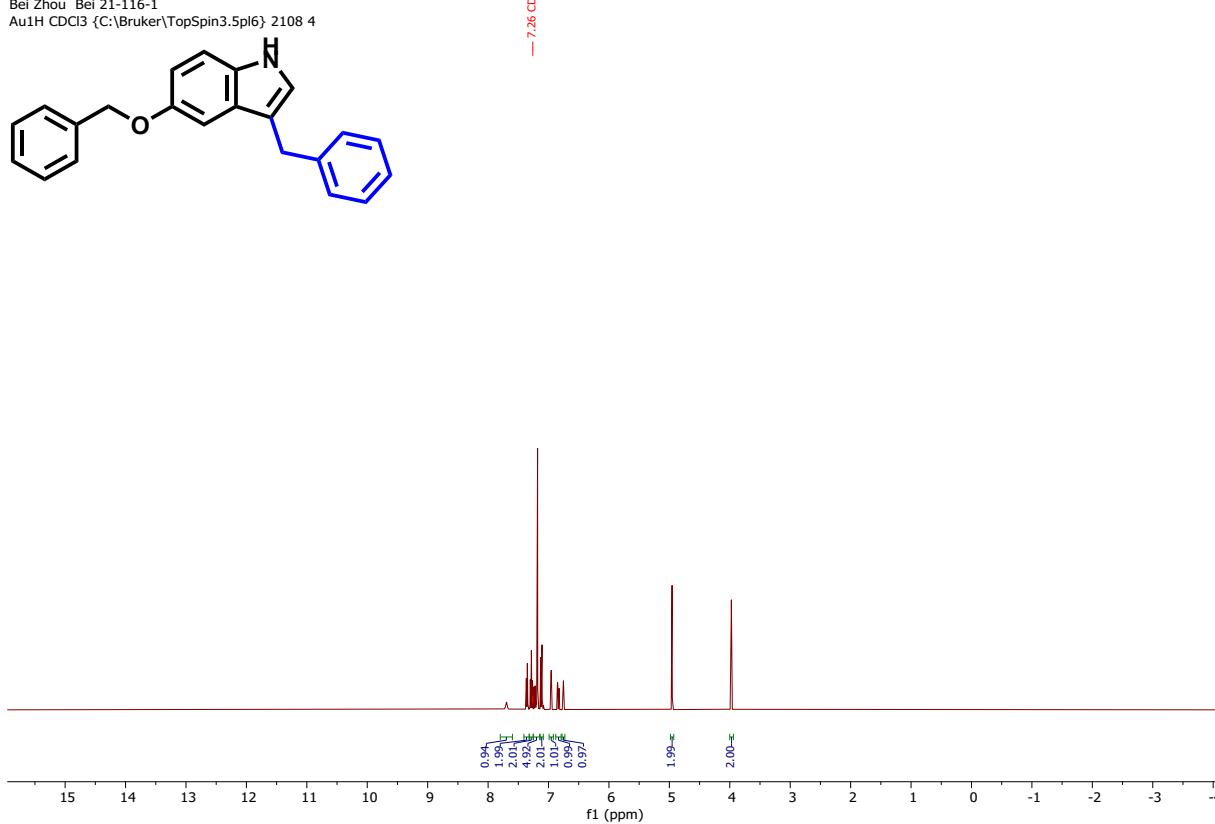
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Bei Zhou Bei 21-144  
Au1H CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2107 7



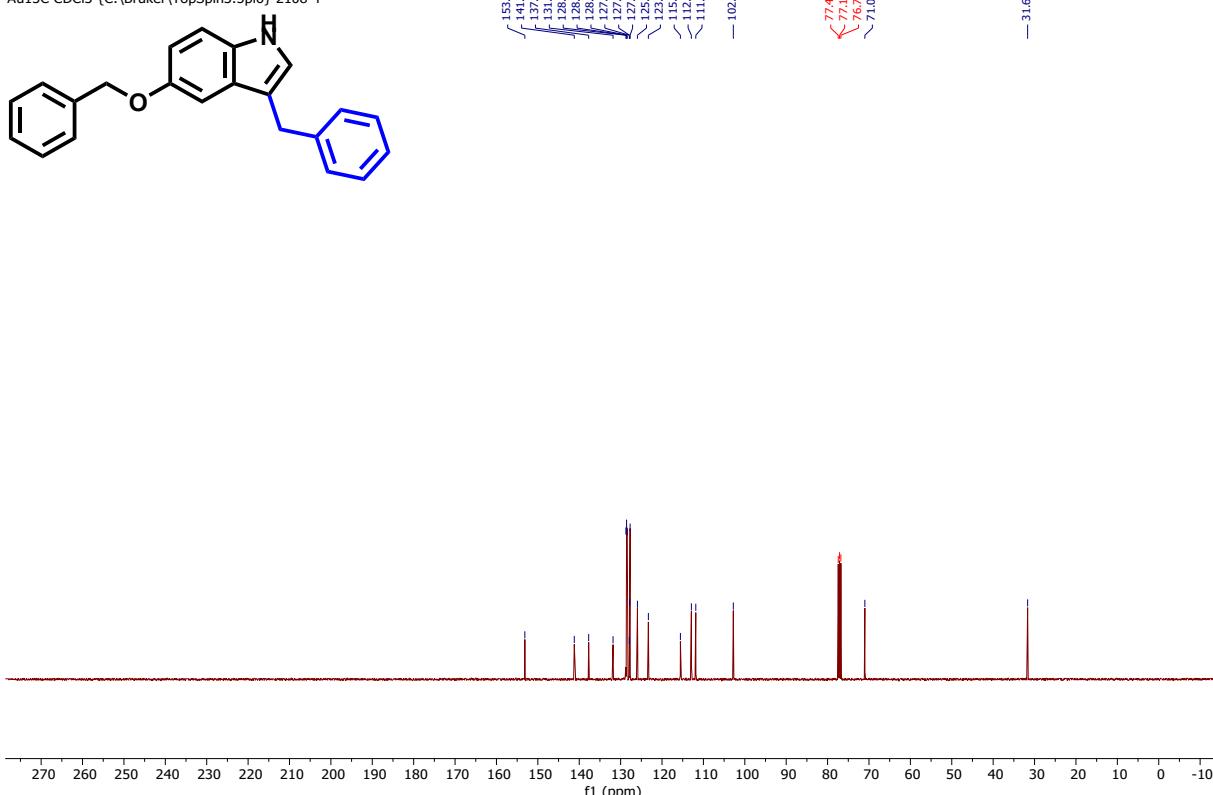
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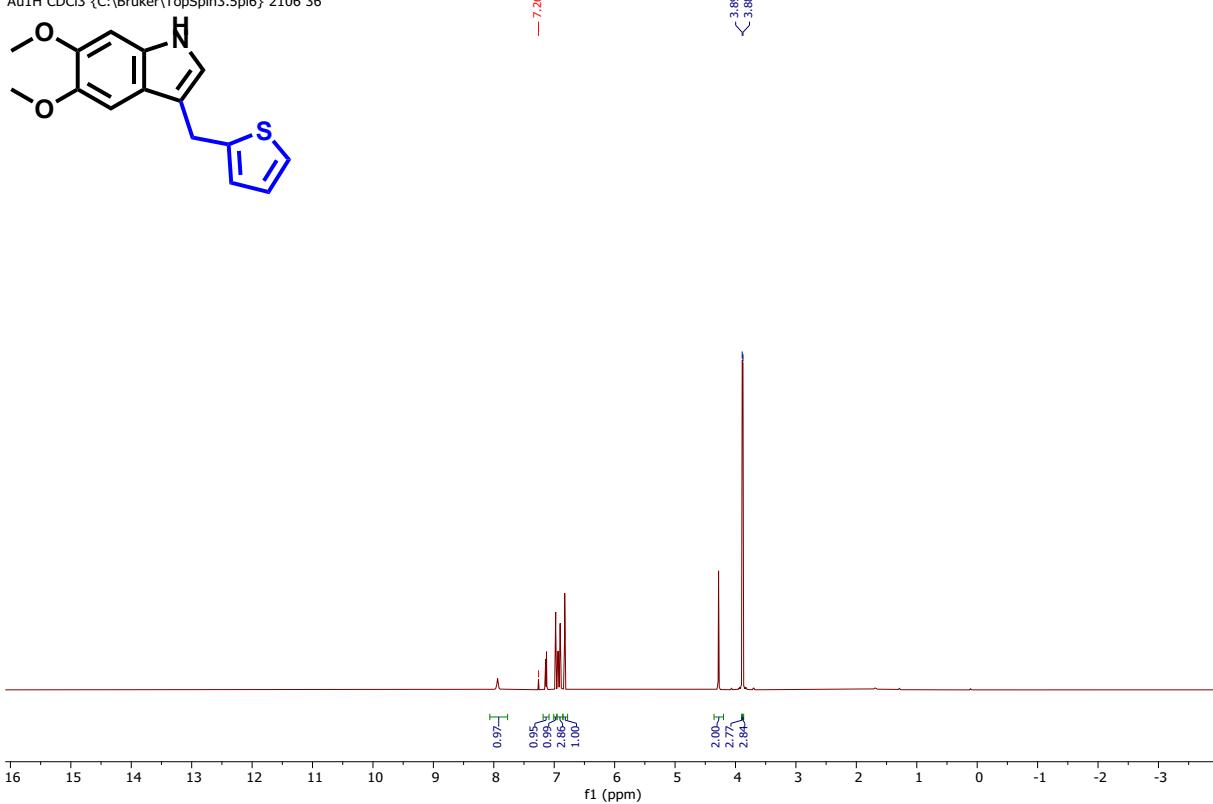
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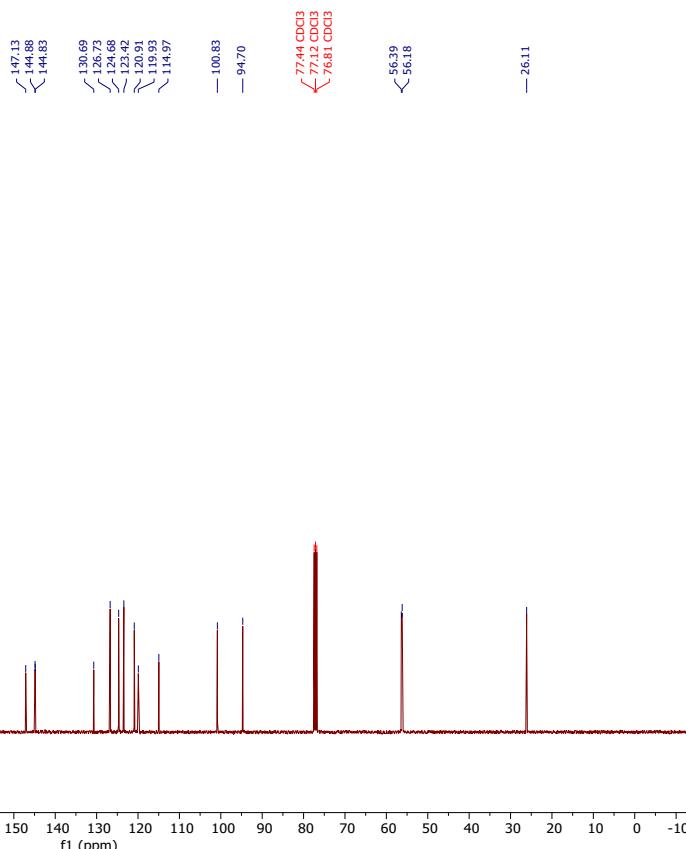
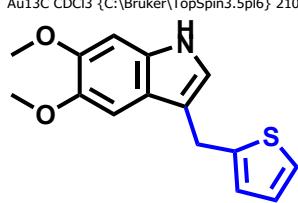
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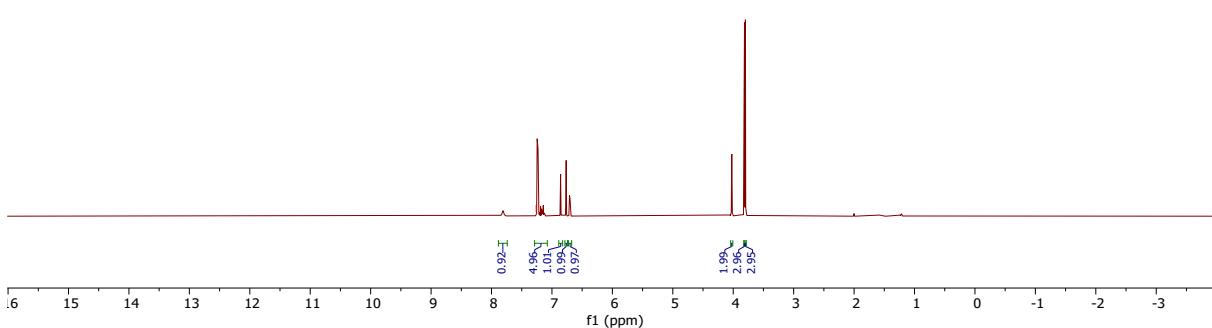
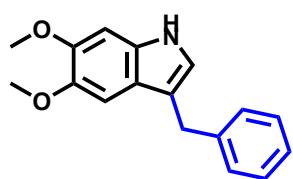
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Bei Zhou Bei 21-137-2  
Au1H CDCl<sub>3</sub> {C:\Bruker\TopSpin3.5pl6} 2106 36



210621.436.11.fid  
Bei Zhou Bei 21-137-2  
Au13C CDCl<sub>3</sub> {C:\Bruker\TopSpin3.5pl6} 2106 36



210804.330.10.fid  
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Au1H CDCl<sub>3</sub> {C:\Bruker\TopSpin3.6.2} 2108 30



210804.330.11.fid  
Bei Zhou Bei 21-118  
Au13C CDCl3 {C:\Bruker\TopSpin3.6.2} 2108 30

