

Catalytic hydroesterification of lignin: A versatile and efficient entry into fully biobased tunable materials

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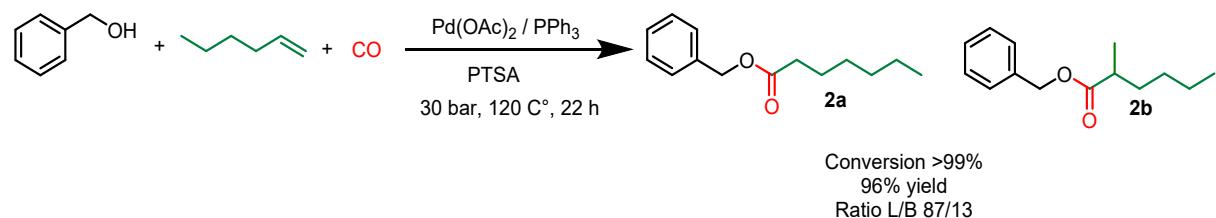
Supporting information

Contents

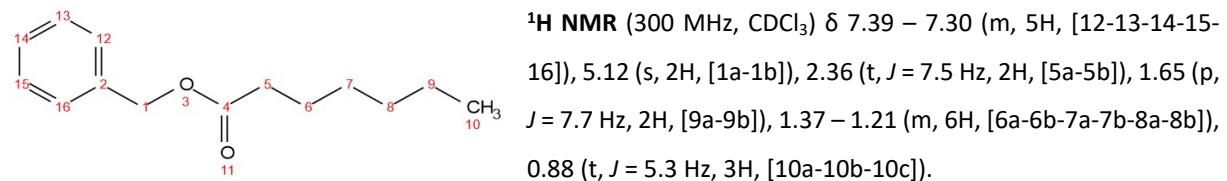
• Model molecules	2
Hydroesterification of 1-hexene with benzyl alcohol.....	2
Hydroesterification of 1-hexene with Guaiacol.....	4
Hydroesterification of butadiene with guaiacol.....	5
Model molecule synthesis (benzyl alcohol hydroesterification with butadiene).....	9
• Table SI 1: Supplementary experiments of Table 1 – Optimization of the palladium catalyzed hydroesterification of 1-hexene with lignin	11
• ^{31}P NMR spectrum of technical lignins (Kraft, Organosolv, Soda) and modified lignins.....	11
• Correction factor calculation in ^{31}P NMR	12
• Table SI 2 ^{31}P Analysis of Technical lignins : Kraft, Organosolv, and Soda Lignins – titration of the hydroxyl groups.....	12
• ^{31}P NMR spectra of Technical lignins : Kraft, Organosolv, and Soda Lignins	13
• NMR spectra and DSC of Table 2: Scope of the hydroesterification of 1-hexene on lignins from different extraction processes.....	14
• NMR spectra of Table 3: Scope of olefin length in hydroesterification on Kraft lignin	18
• Figure SI 29: DSC Comparison of modified Kraft lignin with olefins.....	21
• NMR spectra of Table 4: hydroesterification of fatty acids, fatty ester and estragole on kraft lignin	21
• Table SI 3: Supplementary catalytic experiments of butadiene hydroesterification with Kraft lignin	26
• Table SI 4 - SEC analyses: the number-average molecular weight (M_n), the weight-average molecular weight (MW), and molar mass dispersity (D), for native and modified lignins	27
• Figure SI 40 - Size exclusion chromatography (SEC) of lignin modified by hydroesterification of butadiene	28
• 2D HSQC NMR of Technical and modified lignin by 1-hexene hydroesterification	28

Model molecules

Hydroesterification of 1-hexene with benzyl alcohol



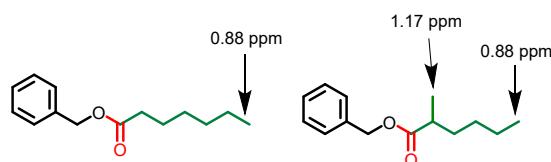
Benzyl alcohol (0.375 mL, 3.6 mmol), palladium acetate (6 mg, 0.027 mmol), triphenylphosphine (175 mg, 0.67 mmol), and paratoluenesulfonic acid (115 mg, 0.67 mmol) were introduced into an autoclave under a nitrogen stream. Distilled and degassed THF (10 mL) and degassed hexene (3 mL, 24 mmol) were added to the autoclave. The autoclave was pressurized with carbon monoxide (30 bar) and kept under stirring at 120 °C for 22 hours. After cooling the reactor to room temperature and release of carbon monoxide, the solvent was evaporated under vacuum, and the crude product was purified by column chromatography on silica using petroleum ether/ethyl acetate mixtures as eluent to afford the desired product in 96% yield. CAUTION: Carbon monoxide is a highly toxic gas and should be handled with extreme care and proper equipment.



¹³C NMR (75 MHz, CDCl₃) δ 173.8 [4], 136.3 [2], 128.7 [12-16], 128.29 [13-15], 128.27 [14], 66.2 [1], 34.5 [5], 31.6 [8], 28.9 [7], 25.1 [6], 22.6 [9], 14.1 [10].

HMRS m/z: Calcd for [M+H]⁺ C₁₄H₂₁O₂ 221.15415; Found 221.0836 ($\Delta m = 0.07055$ m/z).

General method for determination of L/B ratio on the hydroesterification of 1-hexene products: Comparison between the integration of the terminal CH₃ of both isomers (at 0.88 ppm) and the CH₃ of the branched compound (at 1.17 ppm) on ¹H NMR spectrum. See example in figure SI 1. **Ratio L/B: 87/13**



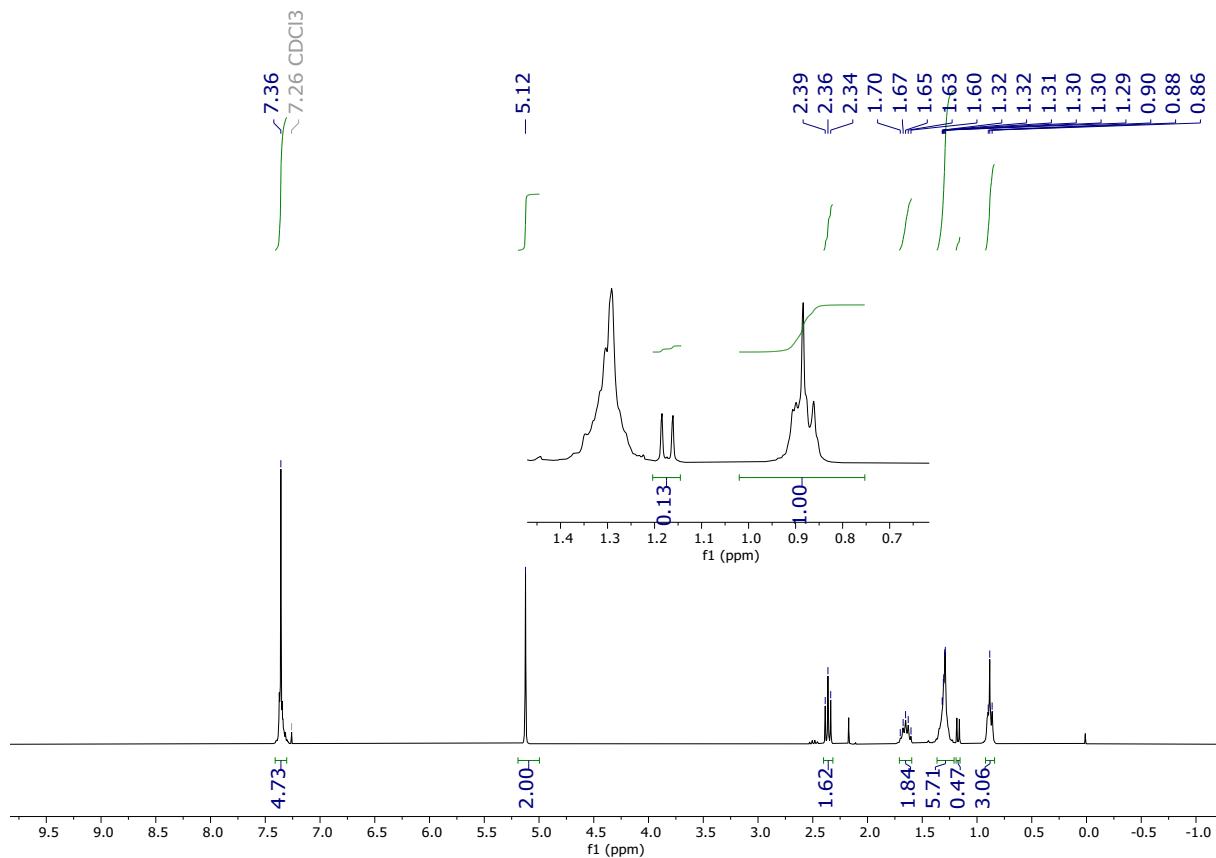


Figure SI 1: ^1H NMR spectrum of the compound **2a + 2b** in CDCl_3 .

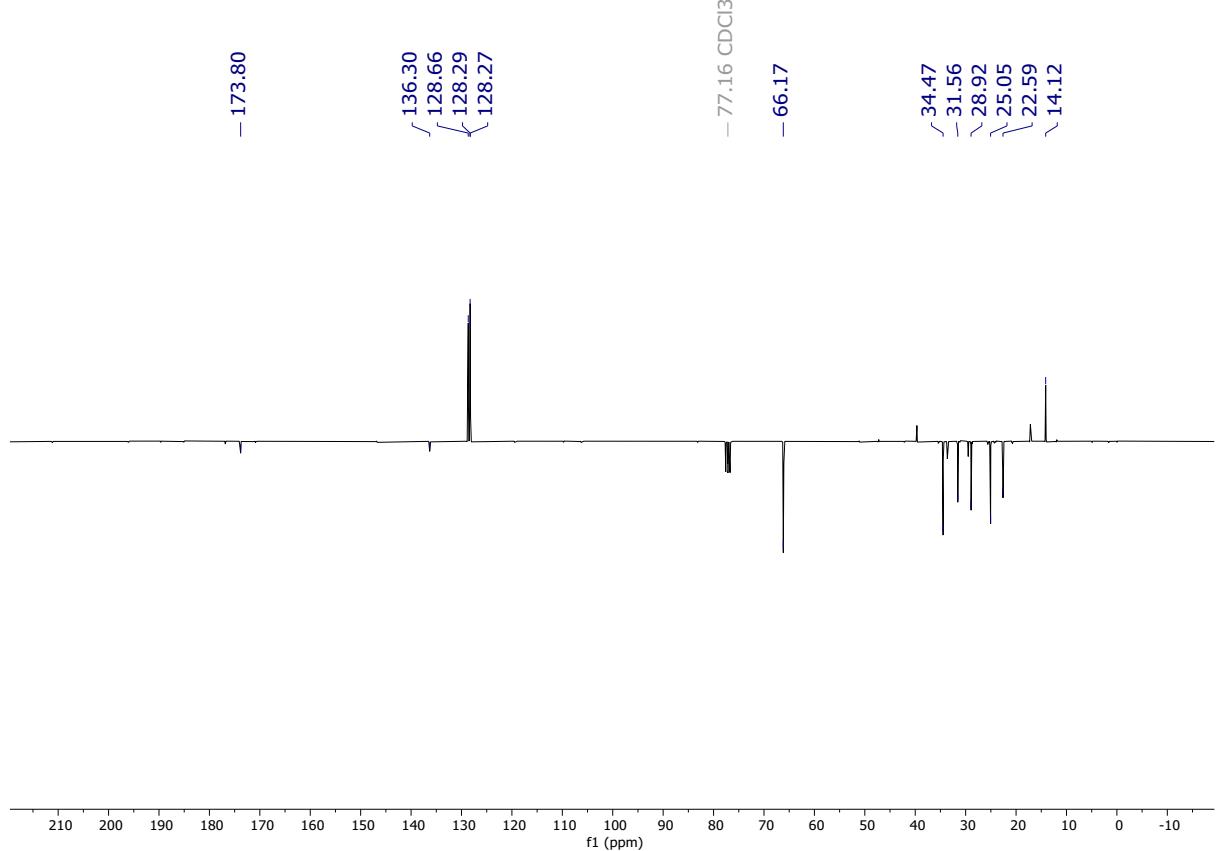
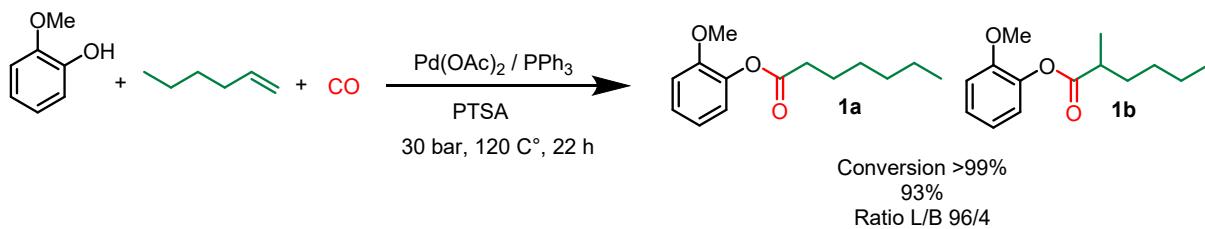
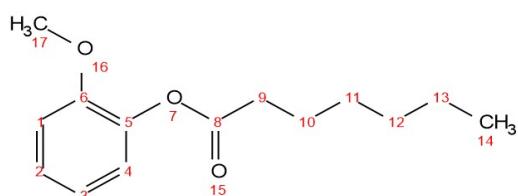


Figure SI 2: ^{13}C NMR spectrum of the compound **2a + 2b** in CDCl_3 .

Hydroesterification of 1-hexene with Guaiacol



This product was synthesized using the same procedure as the one used for the synthesis of compound **1** with benzyl alcohol and was obtained in (93% yield).



¹H NMR (300 MHz, CDCl₃) δ 7.24 – 7.15 (m, 1H, [1]), 7.06 – 6.90 (m, 3H, [2-3-4]), 3.82 (s, 3H, [17a-17b-17c]), 2.59 (t, J = 7.5 Hz, 2H, [9a-9b]), 1.78 (p, J = 7.2 Hz, 2H, [13a-13b]), 1.49 – 1.25 (m, 6H, [10a-10b-11a-11b-12a-12b]), 0.92 (t, J = 6.8 Hz, 3H, [14a-14b-14c]).

¹³C NMR (75 MHz, CDCl₃) δ 172.0 [8], 151.3 [6], 140.0 [5], 126.9 [2], 123.0 [3], 120.9 [4], 112.5 [1], 55.9 [17], 34.2 [9], 31.6 [12], 28.9 [11], 25.1 [10], 22.7 [13], 14.16 [14].

HMRS m/z: Calcd for [M+H]⁺ C₁₄H₂₁O₃ 237.14907; Found 237.1481 (Δm = 0.001 m/z).

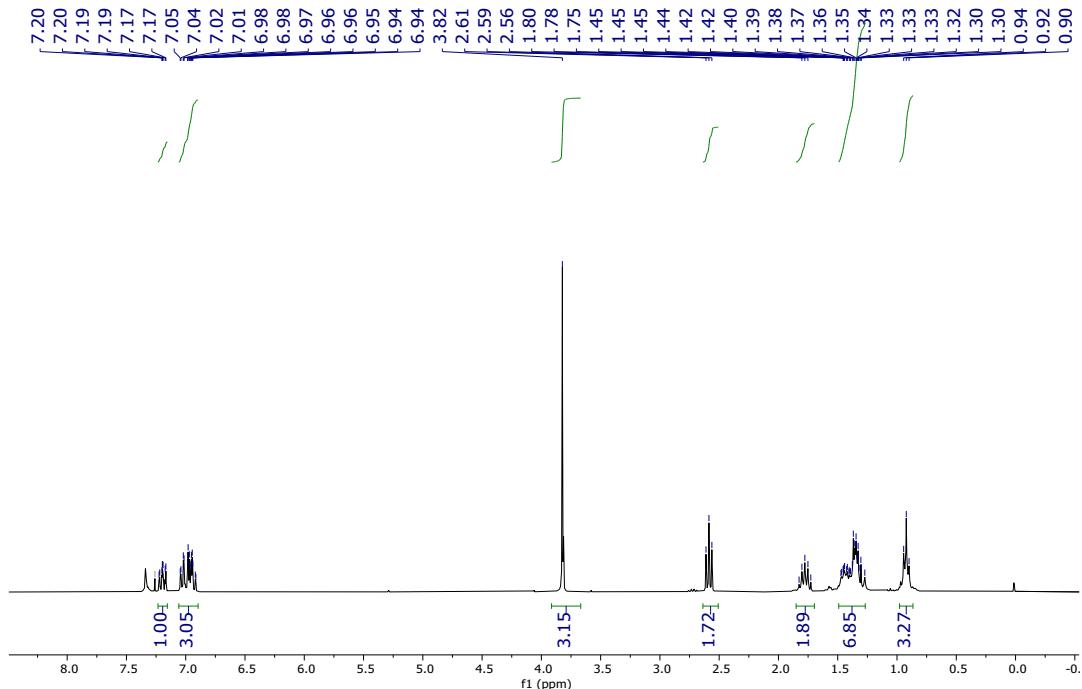


Figure SI 3: ¹H NMR spectrum of the compound **1a + 1b** in CDCl₃.

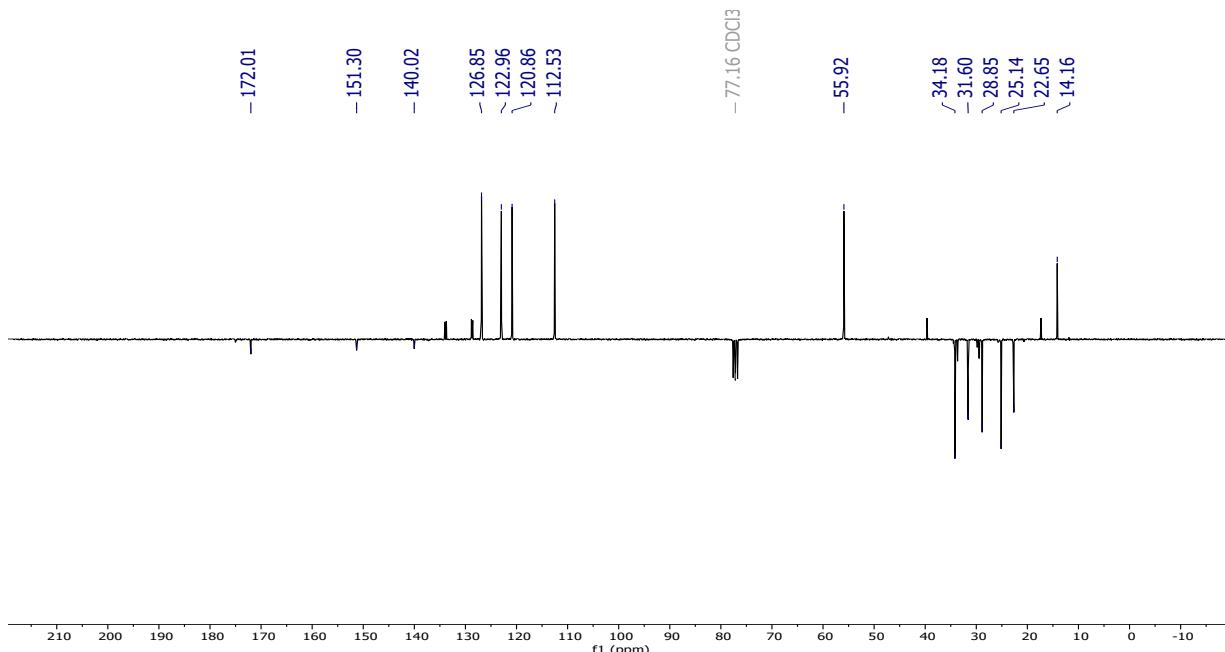
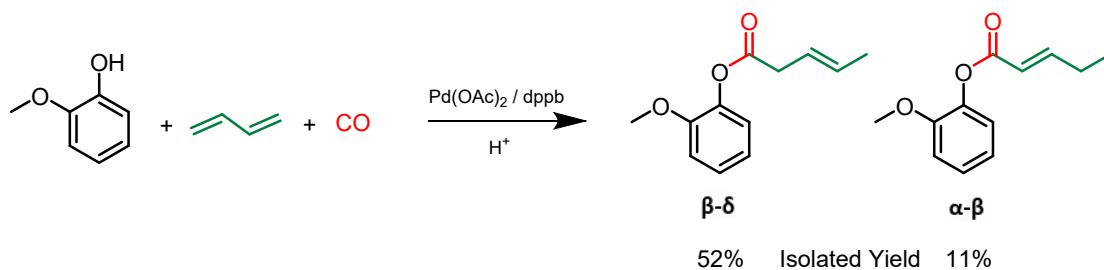
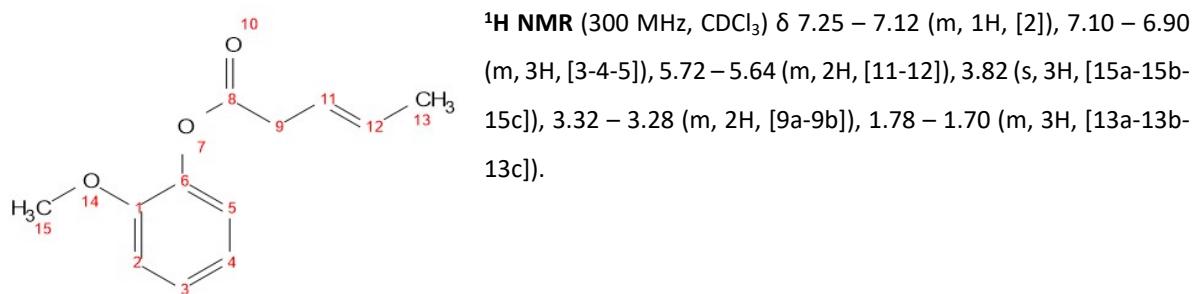


Figure SI 4: ^{13}C NMR spectrum of the compound **1a + 1b** in CDCl_3

Hydroesterification of butadiene with guaiacol



An autoclave was charged with palladium acetate (20 mg, 0.09 mmol), 1,4-bis(diphenylphosphino)butane (230 mg, 0.54 mmol) and benzoic acid (330 mg, 2.7 mmol). After performing three vacuum/ N_2 cycles, dry THF (3 mL) and degassed guaiacol (1 mL, 9 mmol) were added to the autoclave. The reactor was cooled to -40°C , and then (6 mL, 3.6g, 66 mmol) of butadiene was condensed in a Schlenk tube at -40 and added to the reaction mixture *via* a cannula. The autoclave was pressurized with 25 bar of carbon monoxide and heated at 120°C for 24 h. After cooling, the excess gas was evacuated, the solvent was evaporated and the crude was purified on silica gel chromatographic column (95/5, petroleum ether/ethyl acetate). Two isomers have been isolated, 2-methoxyphenyl-pent-3-enoate ($\beta\text{-}\delta$) in 52% yield and 2-methoxyphenyl-pent-2-enoate ($\alpha\text{-}\beta$) in 11% yield. CAUTION: Carbon monoxide is a highly toxic gas and should be handled with extreme care and proper equipment.



¹³C NMR (75 MHz, CDCl₃) δ 170.0 [8], 151.0 [6], 139.7 [1], 129.6 [11], 126.7 [3], 122.7 [12], 122.3 [4], 120.6 [5], 112.3 [2], 55.6 [15], 37.5 [9], 17.9 [3].

HMRS m/z: Calcd for [M+H]⁺ C₁₂H₁₅O₃ 207,1021; Found 207,1020, ($\Delta m = 0,0001$ m/z).

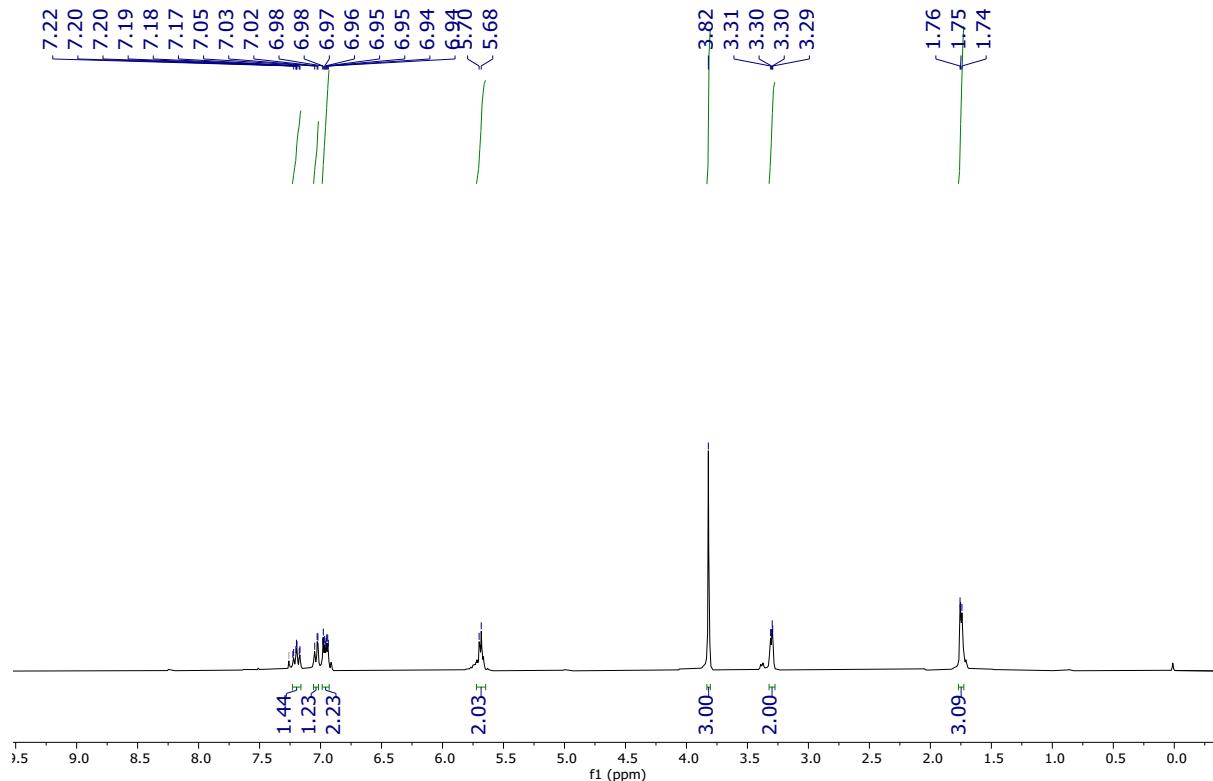


Figure SI 5: ¹H NMR spectrum of the compound β - δ in CDCl₃.

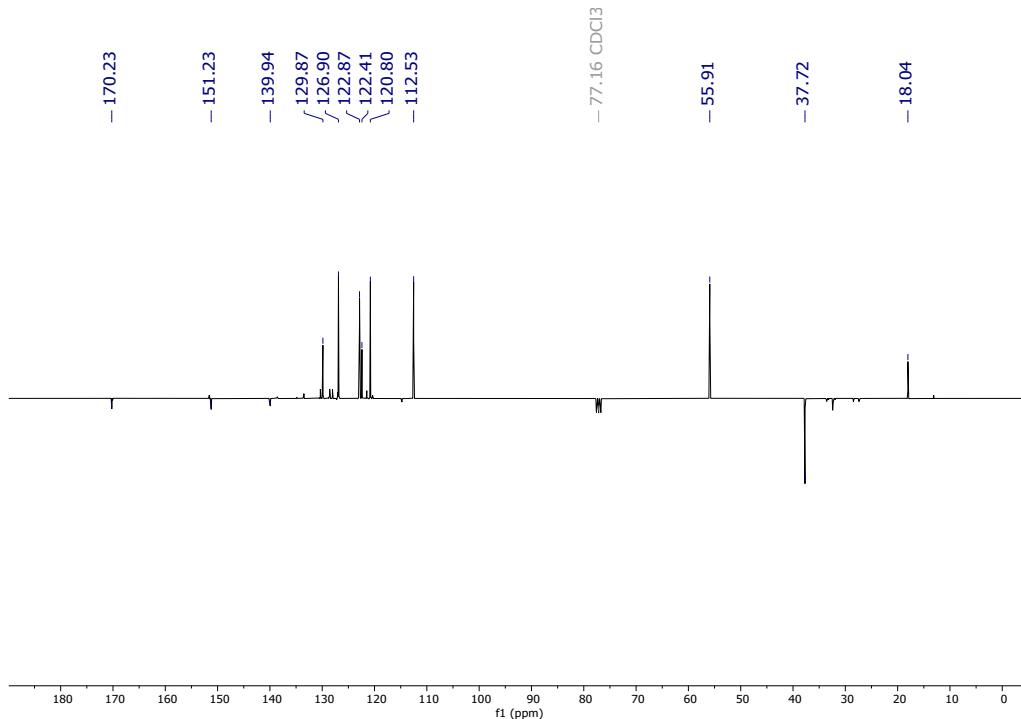


Figure SI 6: ^{13}C NMR spectrum of the compound β - δ in CDCl_3 .

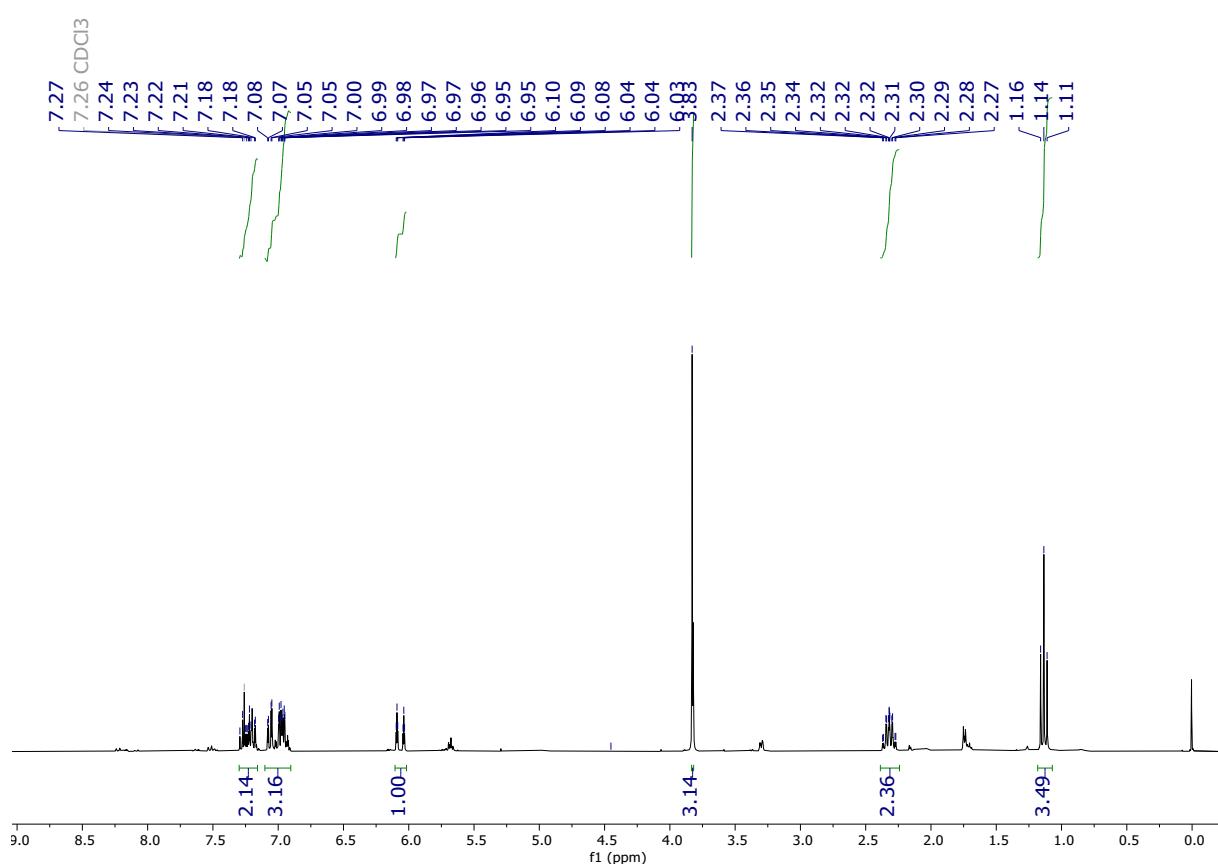
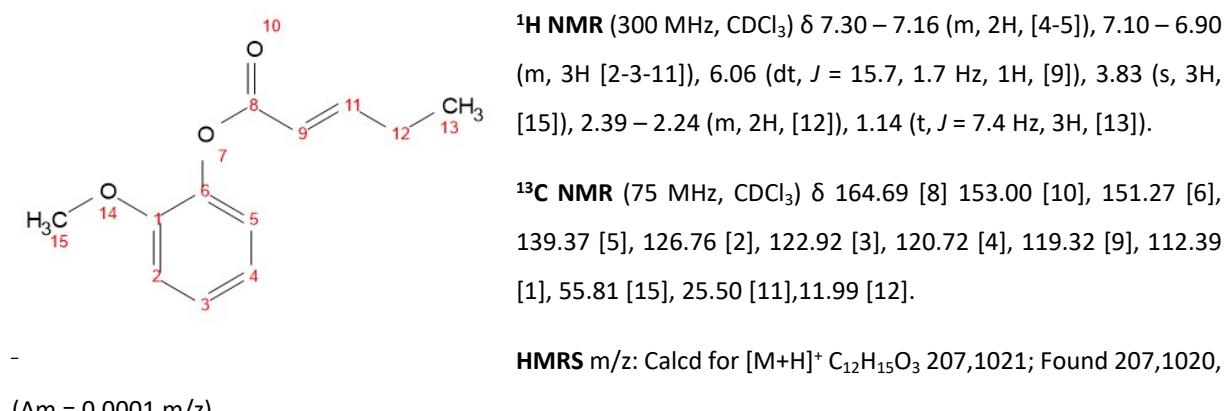


Figure SI 7: ^1H NMR spectrum of the compound α - β in CDCl_3 .

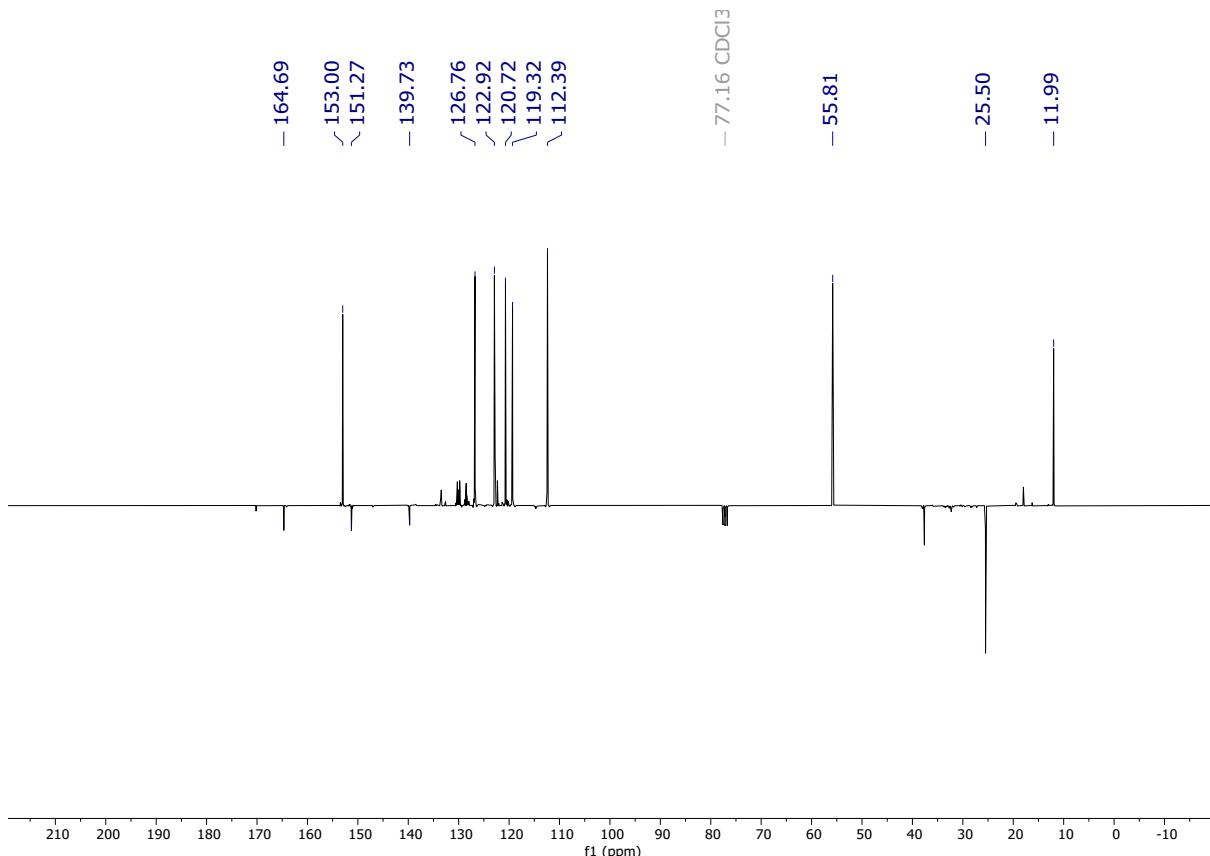
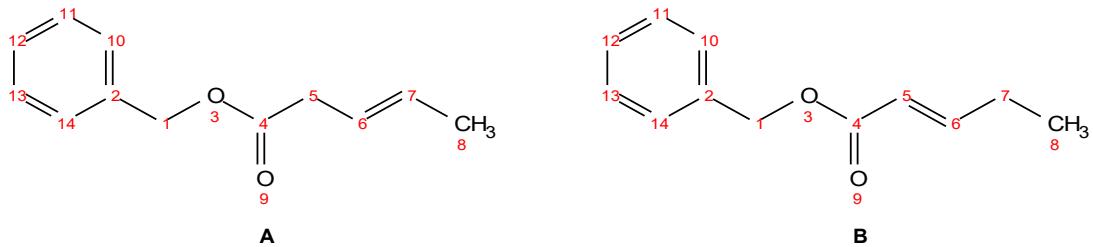


Figure SI 8: ^{13}C NMR spectrum of the compound α - β in CDCl_3 .

Model molecule synthesis (benzyl alcohol hydroesterification with butadiene)

An autoclave was charged with palladium acetate (20 mg, 0.09 mmol), 1,4-bis(diphenylphosphino)butane (230 mg, 0.54 mmol) and benzoic acid (330 mg, 2.7 mmol). After performing three vacuum/ N_2 cycles, dry THF (3 mL) and degassed benzyl alcohol (1 mL, 9 mmol) were added to the autoclave. The reactor was cooled to -40 °C, and then (6 mL, 3.6g, 66 mmol) of butadiene was condensed in a Schlenk tube at -40 and added to the reaction mixture *via* a cannula. The autoclave was pressurized with 25 bar of carbon monoxide and heated at 120°C for 24 h. After cooling, the excess gas was evacuated, the solvent was evaporated and the crude was purified on silica gel chromatographic column (95/5, petroleum ether/ethyl acetate). A mixture of two isomers A and B was obtained in **94% yield with ratio of 60% A and 40% B.** CAUTION: Carbon monoxide is a highly toxic gas and should be handled with extreme care and proper equipment.



^1H NMR (300 MHz, CDCl_3) δ 7.42 – 7.29 (m, 5H, [10-11-12-13-14, A and B]), 7.08 (dt, $J = 15.7, 6.3$ Hz, 1H, [6, A]), 5.93- 5.83 (m, 1H, [5, B]), 5.62 – 5.52 (m, 2H, [6-7, A]), 5.18 (s, 2H, [1, B]), 5.13 (s, 2H, [1, A]), 3.20 – 3.00 (m, 2H, [5, A]), 2.30 – 2.18 (m, 2H, [7, B]), 1.72 – 1.68 (m, 3H, [8, A]), 1.07 (t, $J = 7.4$ Hz, 3H, [8, B]).

¹³C NMR (75 MHz, CDCl₃) δ 172.02 [4, A], 166.62 [4, B] 151.37 [6, B], 136.22 [2, A], 136.0 [2, B], 129.55 [6, A], 128.55 [10-14, A], 128.21 [11-13, B], 128.15 [12, A+B], 122.62 [7, A], 120.11 [5, B], 66.35 [1, A], 66.01 [1, B], 38.04 [5, A], 25.34 [7, B] 17.92 [8, A], 12.11 [8, B].

HMRS m/z: Calcd for $[M+H]^+$ C₁₂H₁₅O₂ 191,1072; Found 191,1068, ($\Delta m = 0,0004$ m/z).

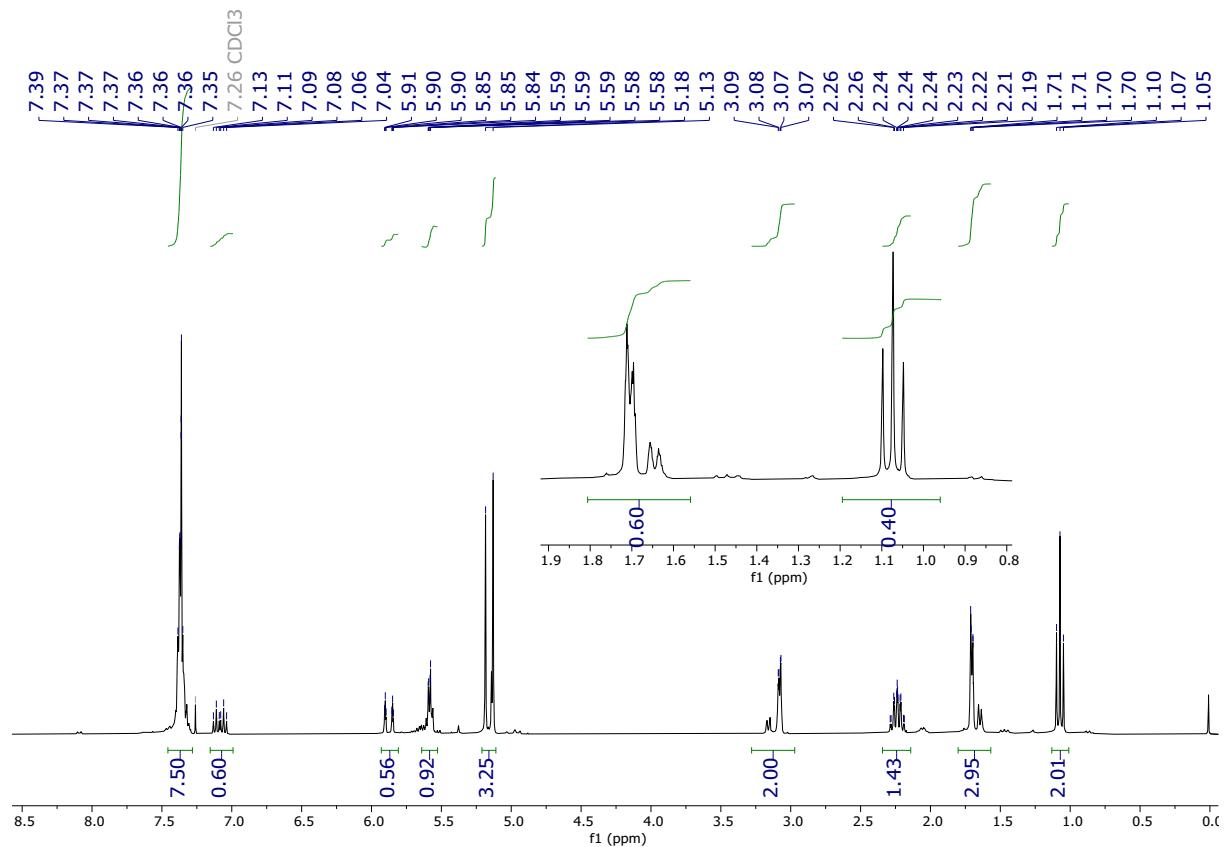


Figure SI 9: ^1H NMR spectrum of the compound A + B 60/40 ratio in CDCl_3 .

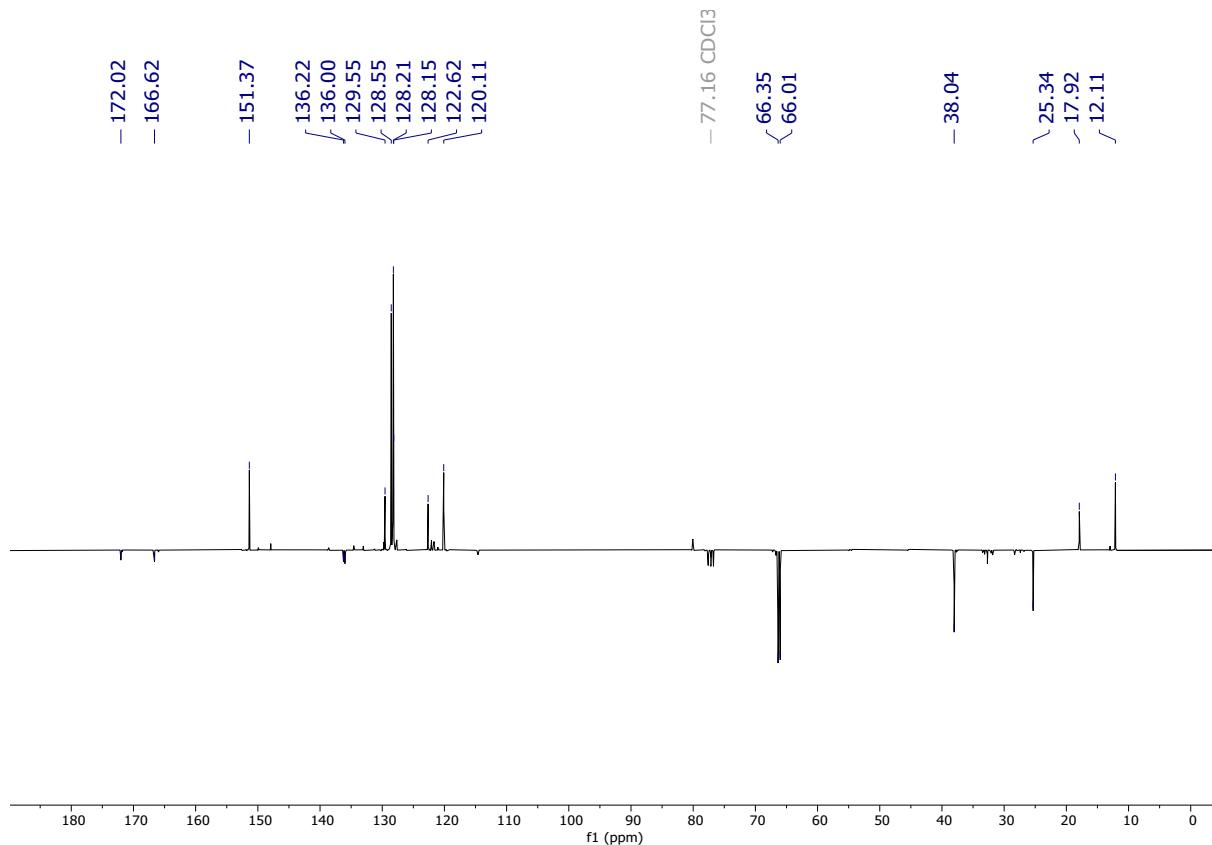


Figure SI 10: ^{13}C NMR spectrum of the compound **A + B** in CDCl_3 .

Table SI 1: Supplementary experiments of Table 1 – Optimization of the palladium catalyzed hydroesterification of 1-hexene with lignin

Entry	Pd(OAc) ₂ (Eq/OH)	PPPh ₃ (eq/Pd)	PTSA (eq/Pd)	T (°C)	THF (mL)	t (h)	Conv. ^[a] OH _{total}	%W ^[b]
1	1 %	11	25	120	10	22 h	9 %	52 %
2	1 %	15	25	120	10	22 h	9 %	52 %
3	1 %	25	25	120	5	22 h	42 %	58 %
4	1 %	25	25	120	20	22 h	56 %	35 %
5 ^[c]	2 %	13	13	120	10	22 h	57 %	96 %
6 ^[c]	2 %	13	13	120	10	48 h	59 %	81 %
7 ^{[c][d]}	2 %	13	13	120	10	22 h	60 %	74 %

Conditions: 0.5g of Kraft lignin, 8 eq/OH of 1-hexene, 30 bars of CO a) by ^1H NMR with pentafluorobenzaldehyde as internal standard: Conversion of total OH groups - comparison of the amount of chains grafted onto the lignin with the quantity of available hydroxyl groups. b) Weight of lignin recovered in proportion to the amount of starting lignin. c) reaction with 3 eq/OH of 1-hexene d) reaction with 10 bars of CO

Reducing the amount of PPPh₃ considerably decreases the substitution degree of the hydroxyl groups. 61% of lignin's hydroxyl groups were converted with 25 equivalents of PPPh₃ and this value was reduced to 9% with 11 or 15 equivalents of phosphine (entries **1** and **2**). The large amount of ligand required for the reaction can be explained by the oxidation and sulphidation of phosphine caused by impurities present in the kraft lignin as this

has been previously demonstrated in the case of the telomerization reaction with the same material.¹ It is elsewhere known that the presence of large quantities of strong acid is responsible of phosphine protonation, thus limiting the availability of the ligand to coordinate the metal.

³¹P NMR spectrum of technical lignins (Kraft, Organosolv, Soda) and modified lignins.

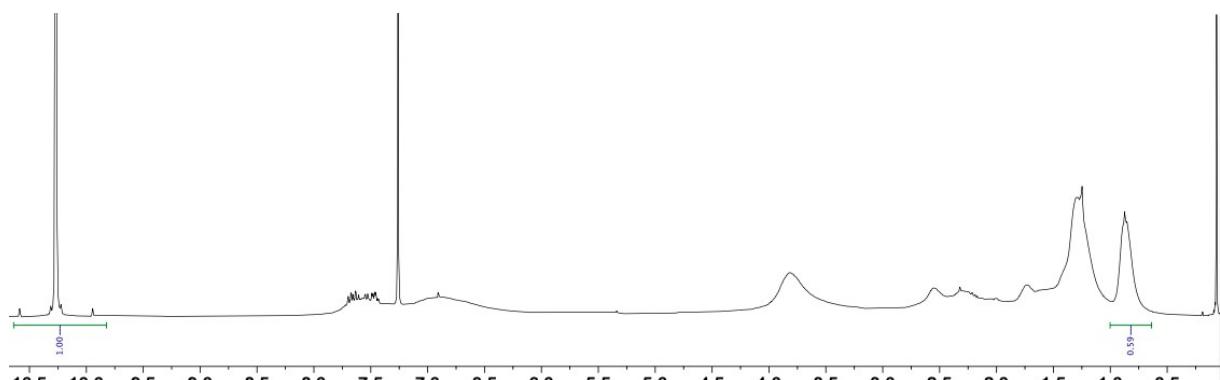
³¹P NMR experiments method: Lignin (30 mg) was dissolved in a CDCl₃ / Pyridine mixture (1:1.6 v/v, 0.5 mL). The phosphorylation reagent, 2-chloro-4,4,5,5-tetramethyl-1,3,2-dioxaphospholane (150 µL), and the internal standard N-hydroxy-6-norbornene-2,3-dicarboximide (100 µL of 0.1 M solution in 1:1.6 v/v CDCl₃ / Pyridine mixture) were added successively. Chromium(III) acetylacetone (100 µL of 0.014M solution) in the same CDCl₃ / Pyridine mixture was added to the solution in order to homogenize and accelerate phosphorus relaxation. Spectra were recorded with a 15 sec relaxation time and an average number of 512 scans. Chemical shifts are relative to the signal of the phospholane hydrolysis product at 132.2 ppm. The integrated value of the internal standard was used for calculation of the absolute amount of each functional group. This method includes a correction factor for integrations of modified lignins considering the reduced lignin content.

Correction factor calculation in ³¹P NMR

The calculation of this coefficient is performed by calculating the mass of initial lignin in the final product by the calculation of the esters weight added.

$$\%_{ligninweight} = \frac{n_{esters/g}}{1 - (\frac{M_{esters}}{M})}$$

The mole content of esters per gram of lignin is determined by ¹H NMR using pentafluorobenzaldehyde as internal standard.



$$n_{esters/g} = \frac{n_{PFB} \times I_{CH_3}}{3}$$

¹ C. Dumont, R. M. Gauvin, F. Belva and M. Sauthier, *ChemSusChem*, 2018, **11**, 1649–1655.

$\%_{\text{ligninweight}}$: mass percentage of lignin in the product after hydroesterification

$n_{\text{esters/g}}$: number of moles of esters per gram of product after hydroesterification

M_{esters} : molar mass of the atoms added through the hydroesterification (depends on the olefin used)

n_{PFB} : number of moles of internal standard pentafluorobenzaldehyde added to the sample

I_{CH_3} : Integration of the ^1H NMR signal between 0.6 - 1 ppm corresponding to the terminal CH_3 of the ester chains.

Table SI 2 ^{31}P Analysis of Technical lignins : Kraft, Organosolv, and Soda Lignins – titration of the hydroxyl groups

Entry	Technical Lignin	mmol/g of hydroxyl groups			
		Aliphatic OH	Phenolic OH	Total OH	Carboxylic Acid
1	Kraft	1.9	4.2	6.1	0.5
2	Soda	1.5	3.9	5.4	1.0
3	Organosolv	2.3	3.7	6.0	2.3

^{31}P NMR spectra of Technical lignins : Kraft, Organosolv, and Soda Lignins

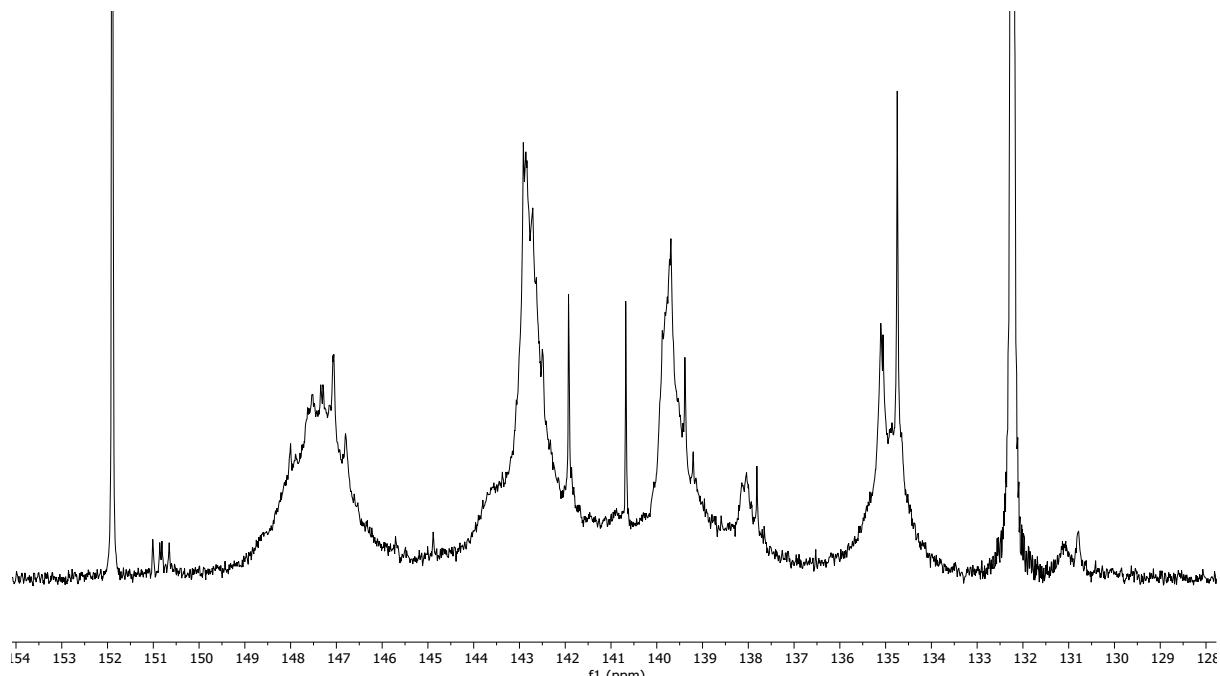


Figure SI 11: ^{31}P NMR spectrum of native Kraft lignin.

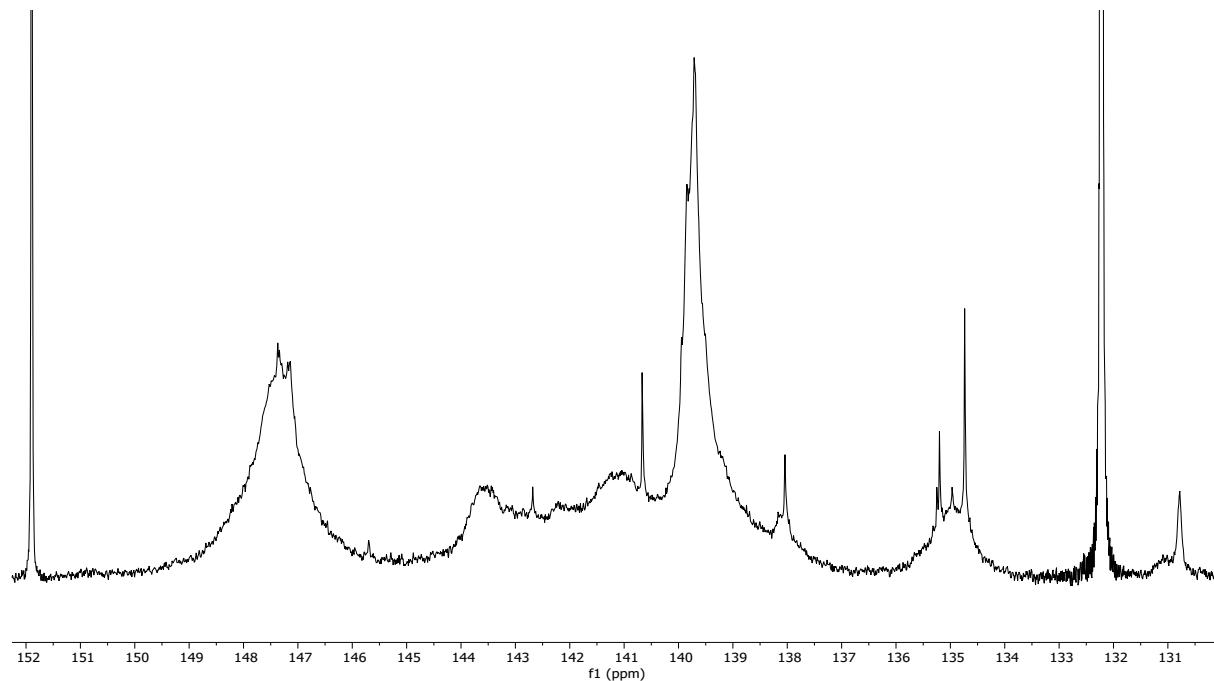


Figure SI 12: ^{31}P NMR spectrum of native organosolv lignin.

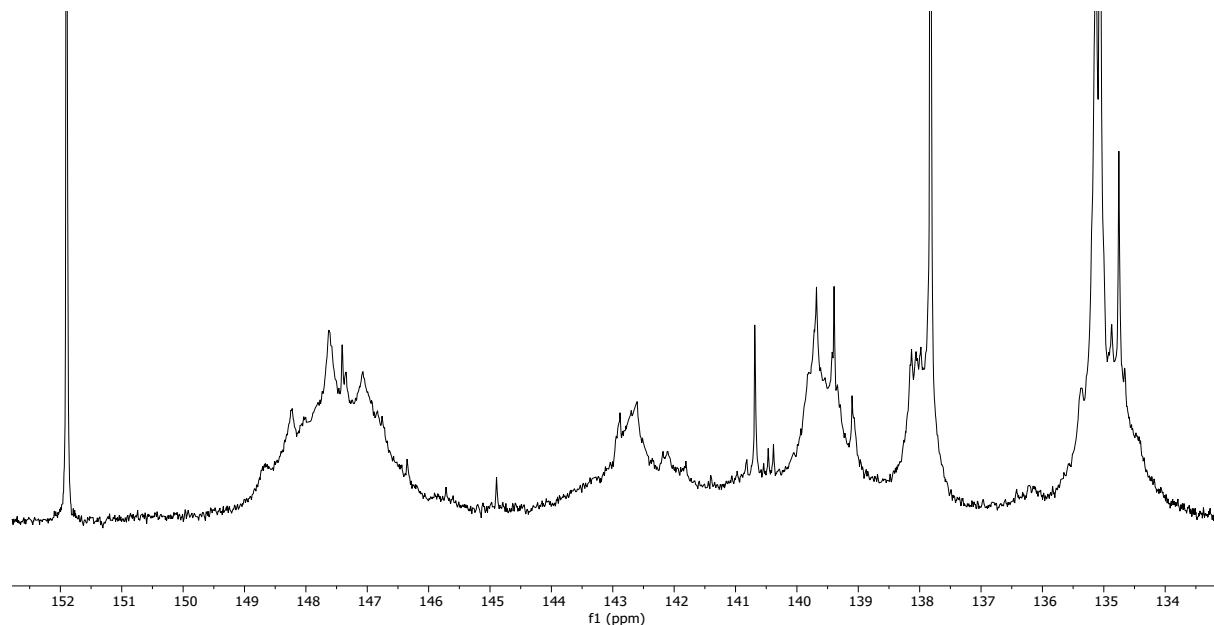


Figure SI 13: ^{31}P NMR spectrum of native soda lignin.

NMR spectra and DSC of Table 2: Scope of the hydroesterification of 1-hexene on lignins from different extraction processes

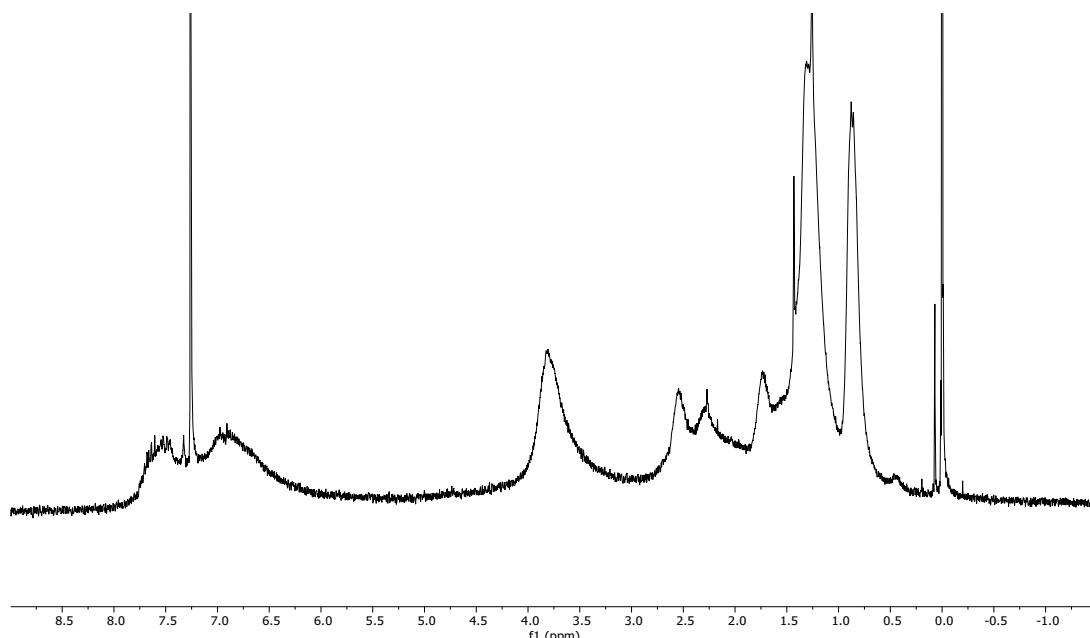


Figure SI 14: ^1H NMR spectrum of the reaction with **Kraft lignin** in CDCl_3 .

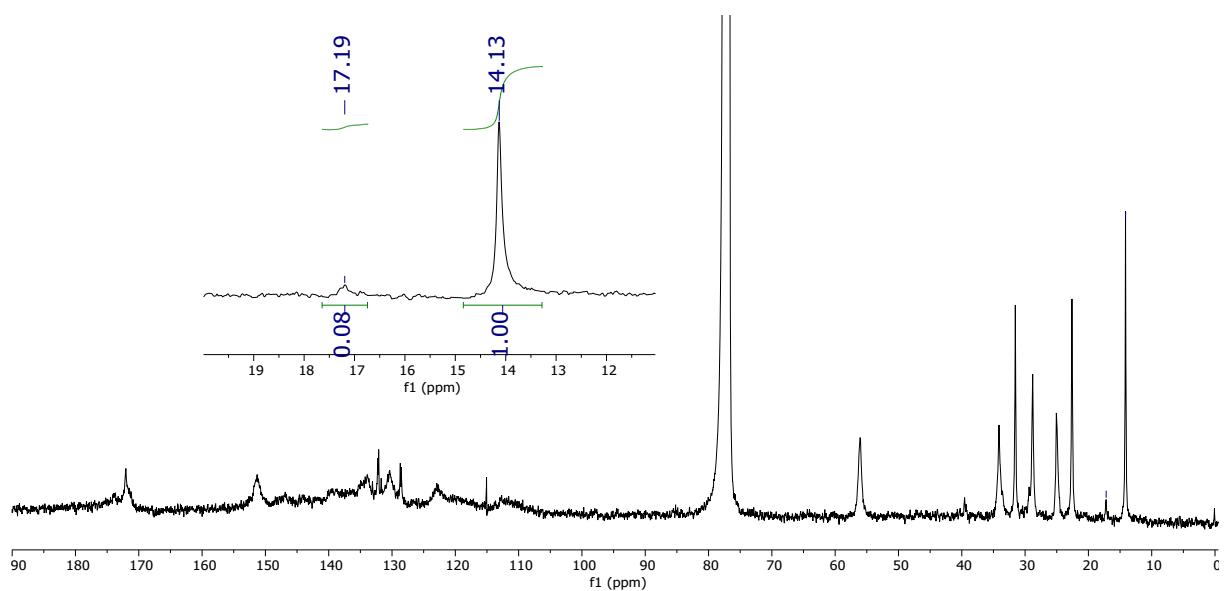
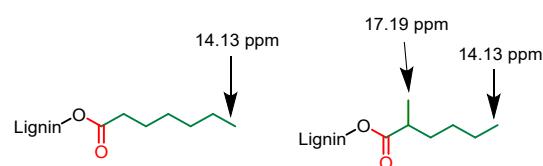


Figure SI 15: ^{13}C Quantitative NMR spectrum of the reaction with **Kraft lignin** in CDCl_3 .

General method for determination of L/B ratio on lignins: Comparison between the integration of the terminal CH_3 of both isomers (at 14.13 ppm) and the CH_3 of the branched compound (at 17.19 ppm) on ^{13}C quantitative NMR spectrum. See example in figure SI 16. **Ratio L/B : 92/8**



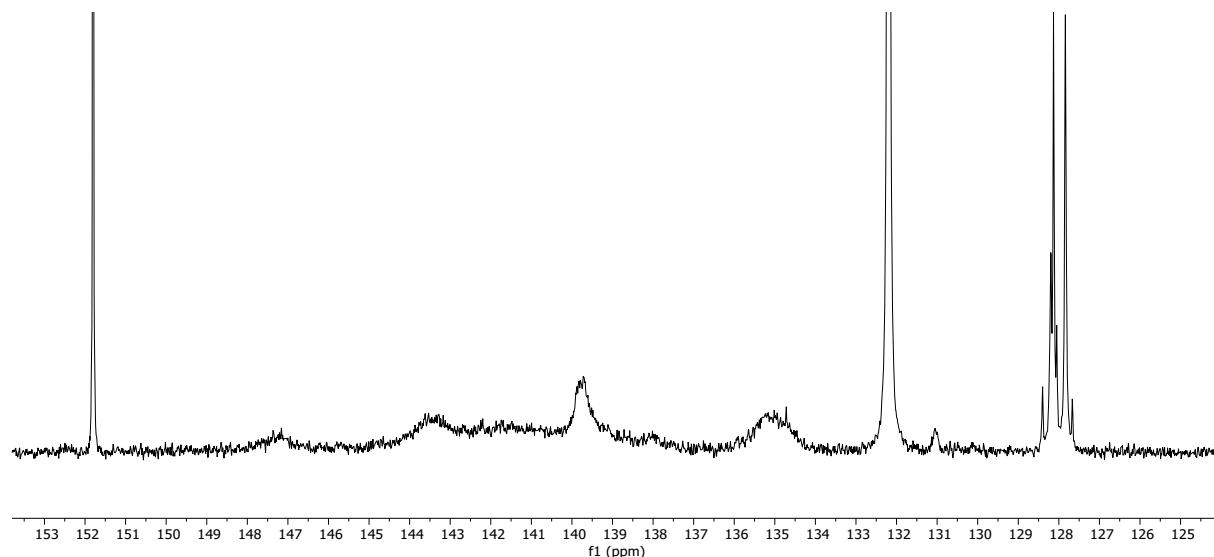


Figure SI 16: ^{31}P NMR spectrum of the reaction with **Kraft lignin**.

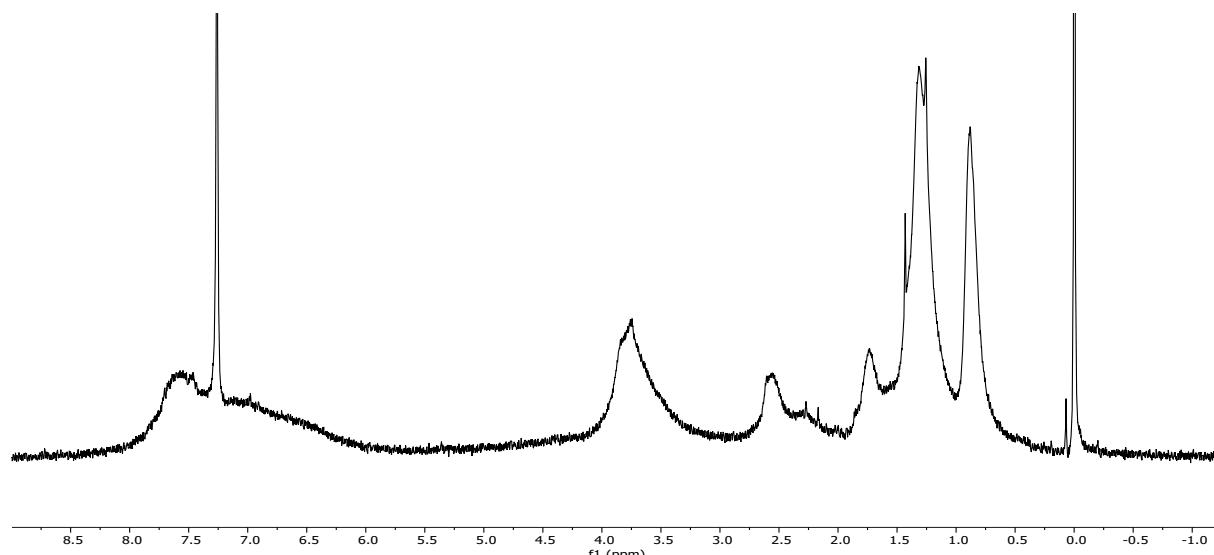


Figure SI 17: ^1H NMR spectrum of the reaction with **Organosolv lignin** in CDCl_3 .

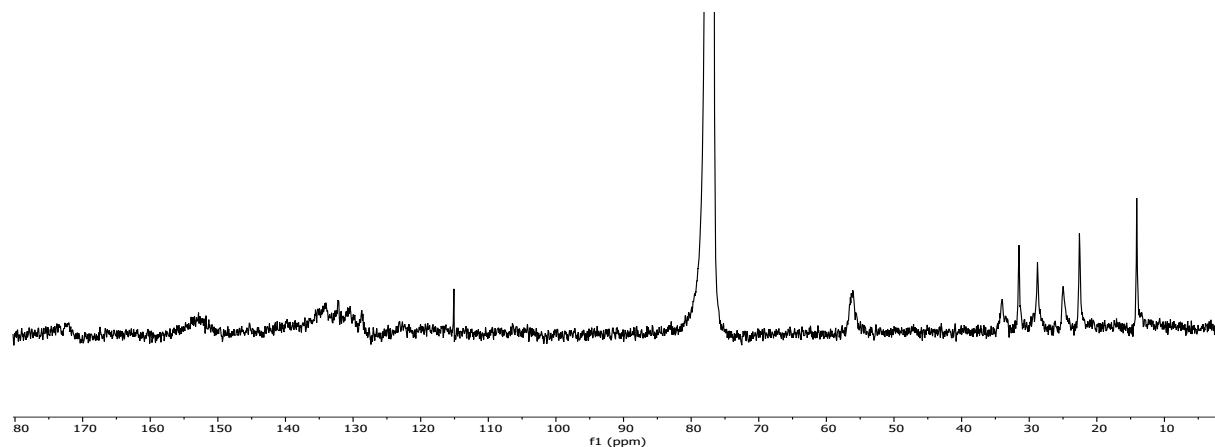


Figure SI 18: ^{13}C Quantitative NMR spectrum of the reaction with **Organosolv lignin** in CDCl_3 .

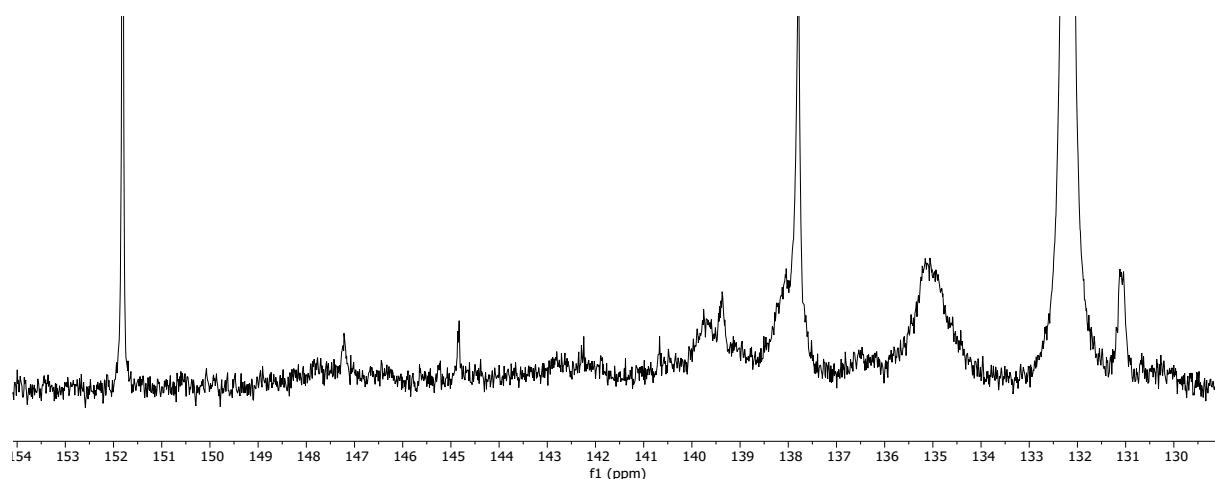


Figure SI 19: ^{31}P NMR spectrum of the reaction with **Organosolv lignin**.

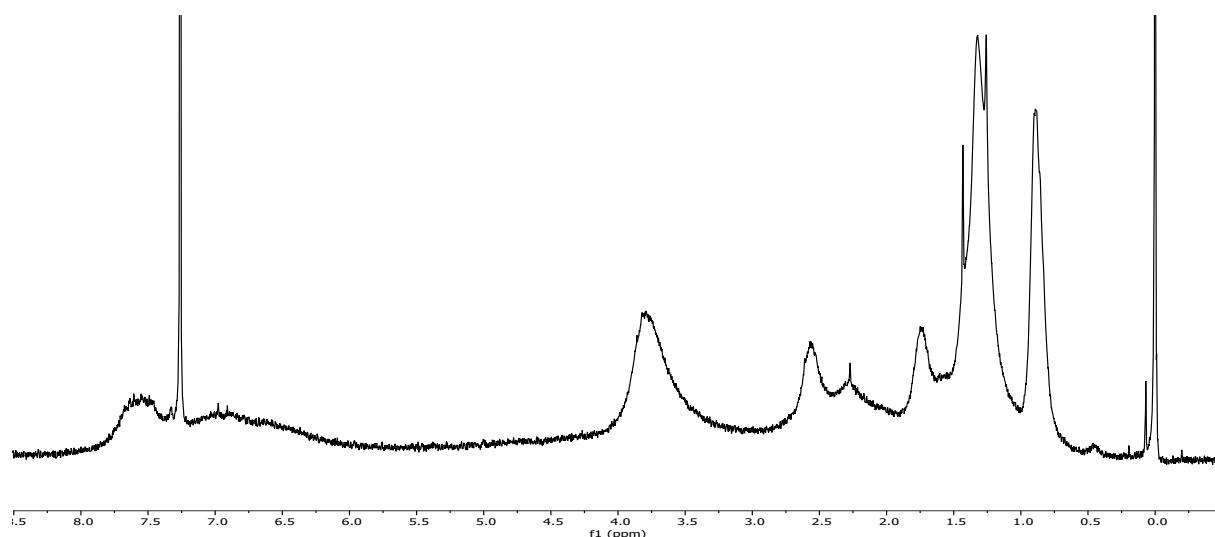


Figure SI 20: ^1H NMR spectrum of the reaction with **Soda lignin** in CDCl_3 .

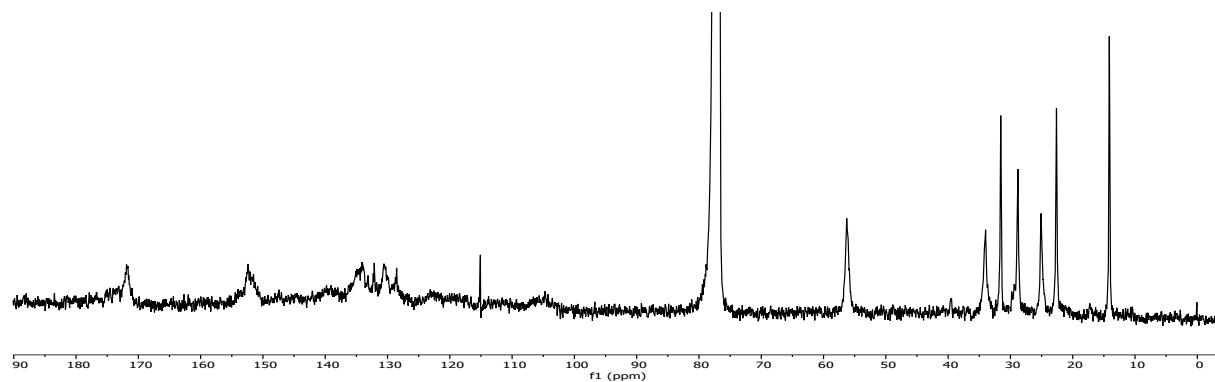


Figure SI 21: ¹³C Quantitative NMR spectrum of the reaction with **Soda lignin** in CDCl_3 .

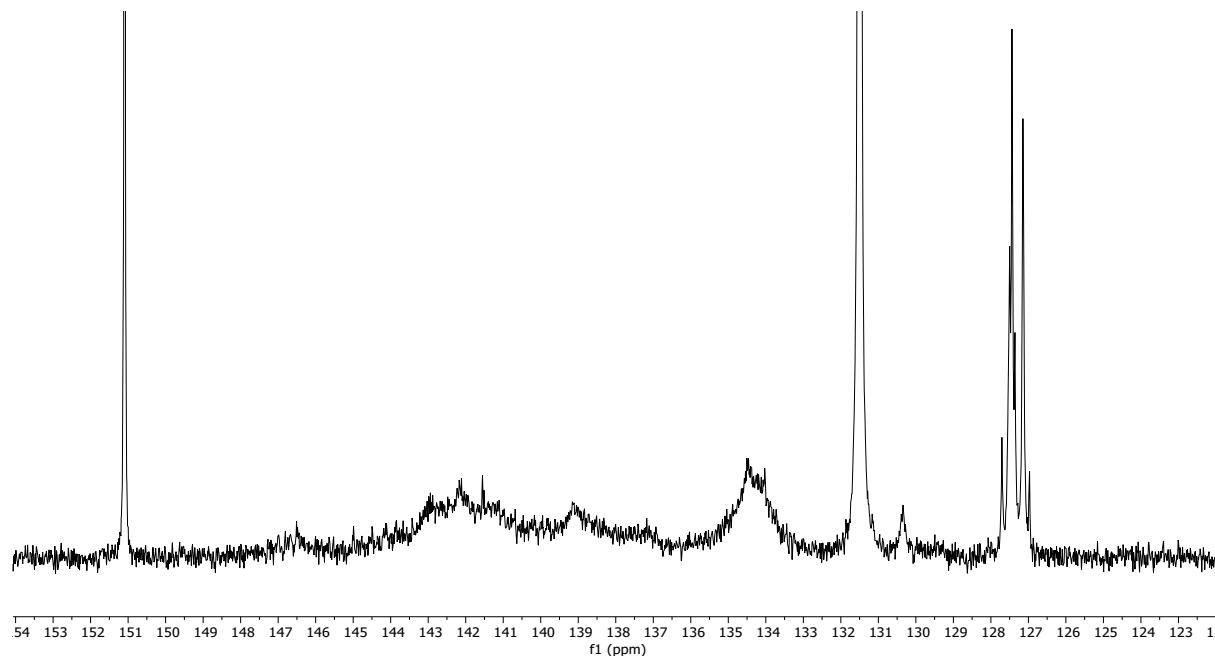


Figure SI 22: ³¹P NMR spectrum of the reaction with **Soda lignin**.

NMR spectra of Table 3: Scope of olefin length in hydroesterification on Kraft lignin

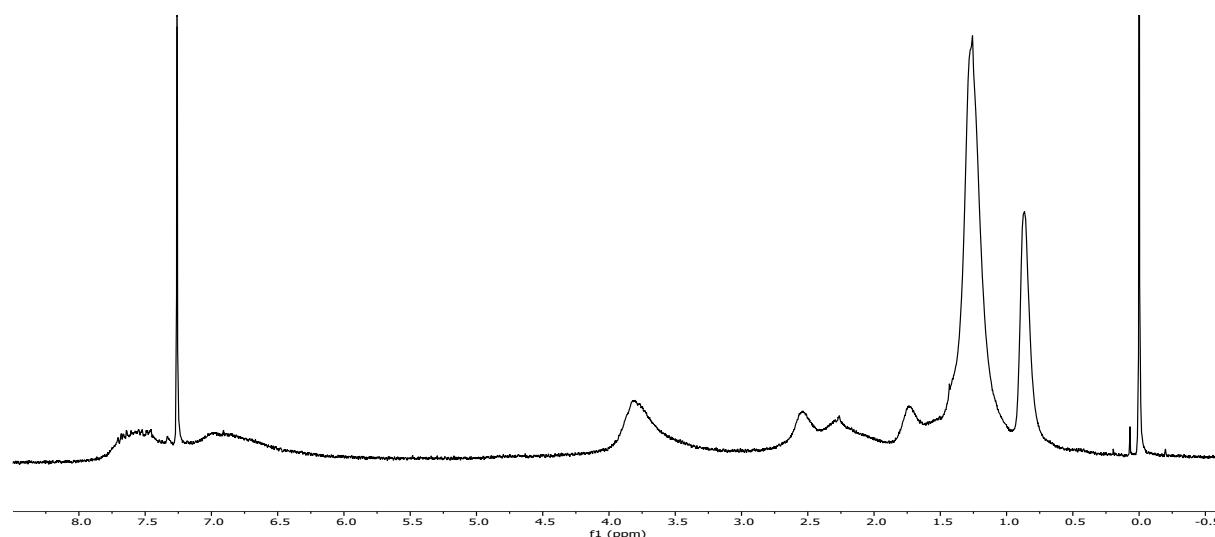


Figure SI 23: ^1H NMR spectrum of the **1-octene** hydroesterification on lignin in CDCl_3 .

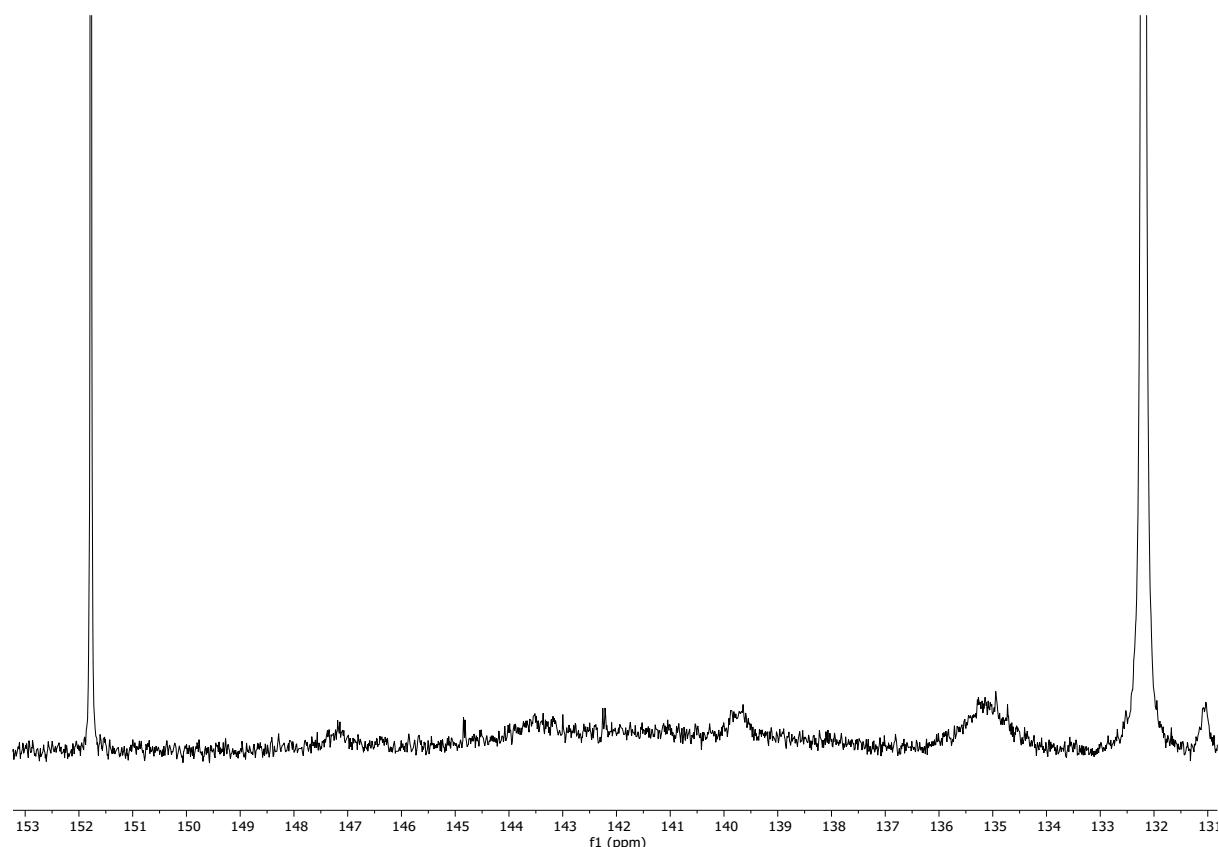


Figure SI 24: ^{31}P NMR spectrum of the **1-octene** hydroesterification on lignin.

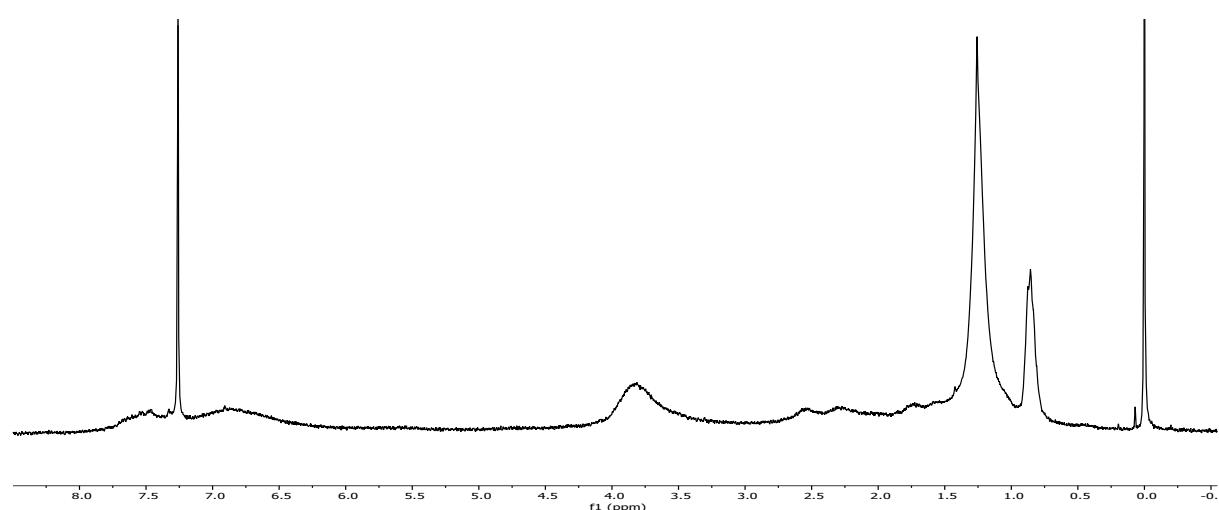


Figure SI 25: ^1H NMR spectrum of the **1-decene** hydroesterification on lignin in CDCl_3 .

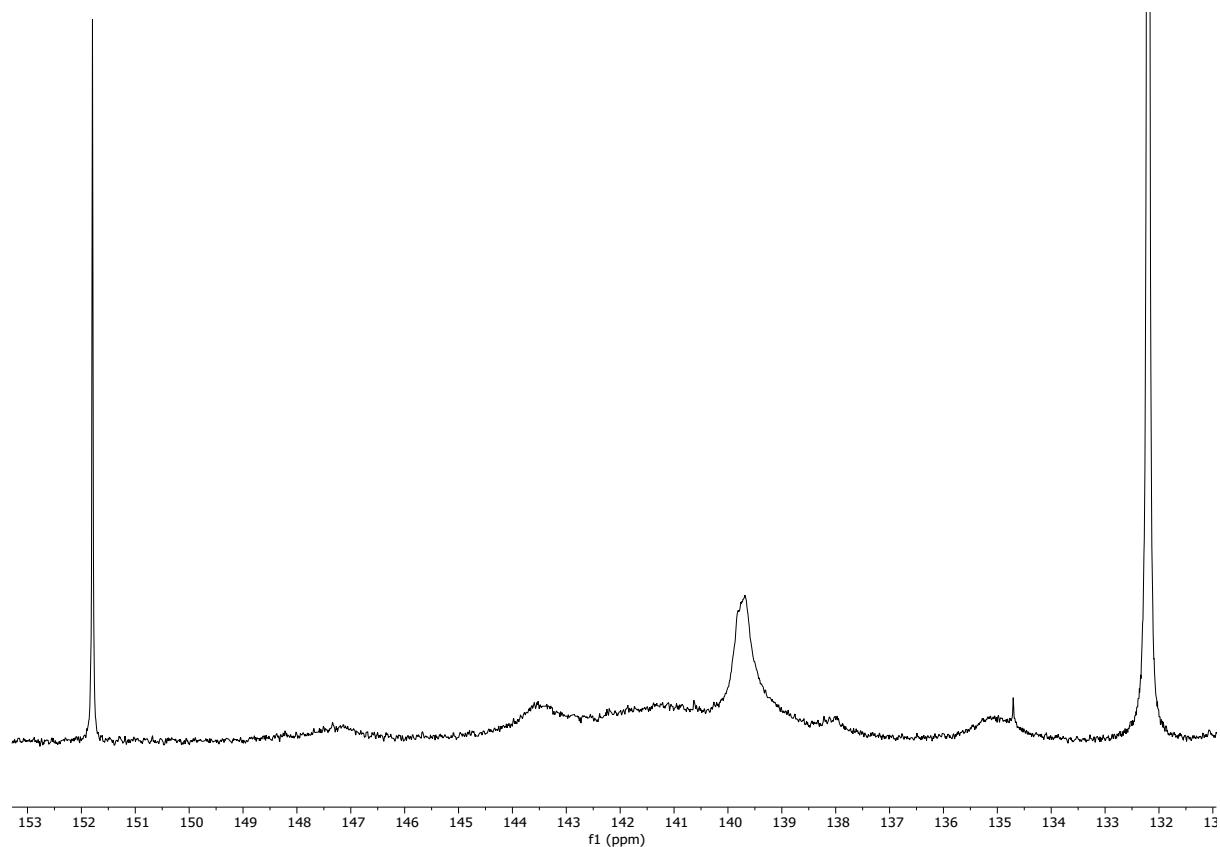


Figure SI 26: ^{31}P NMR spectrum of the **1-decene** hydroesterification on lignin.

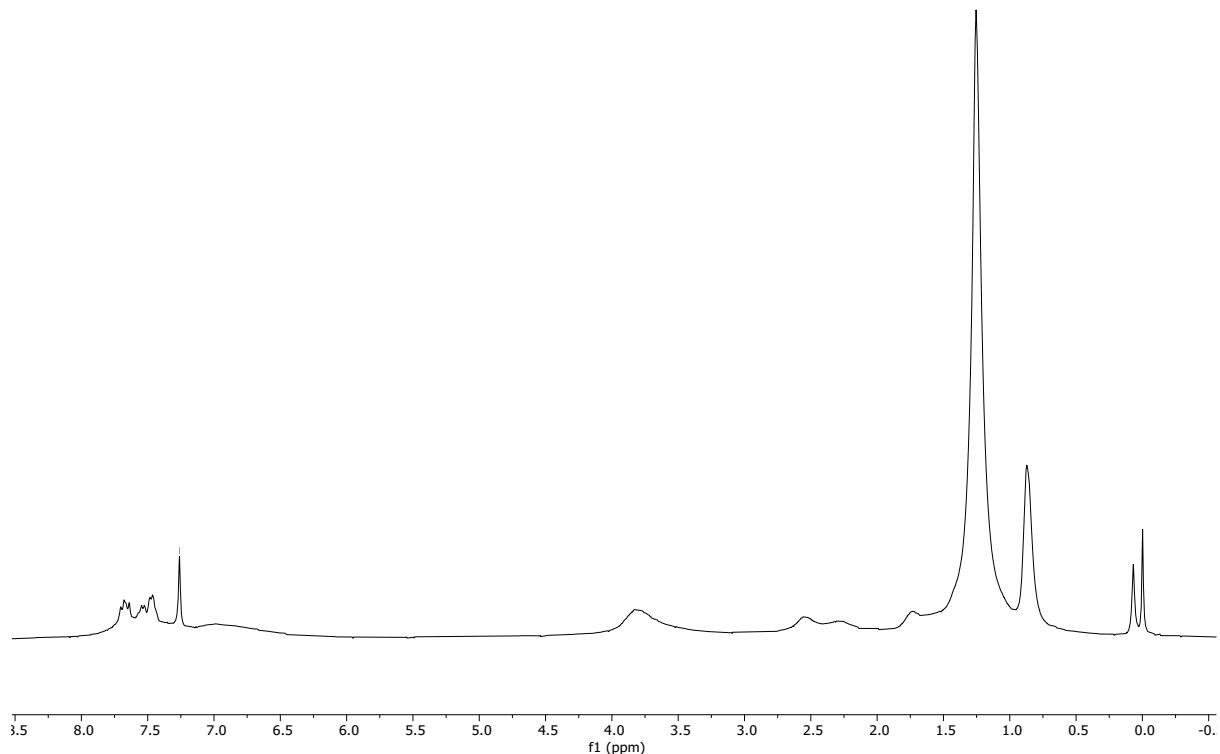


Figure SI 27: ^1H NMR spectrum of the **1-dodecene** hydroesterification on lignin in CDCl_3 .

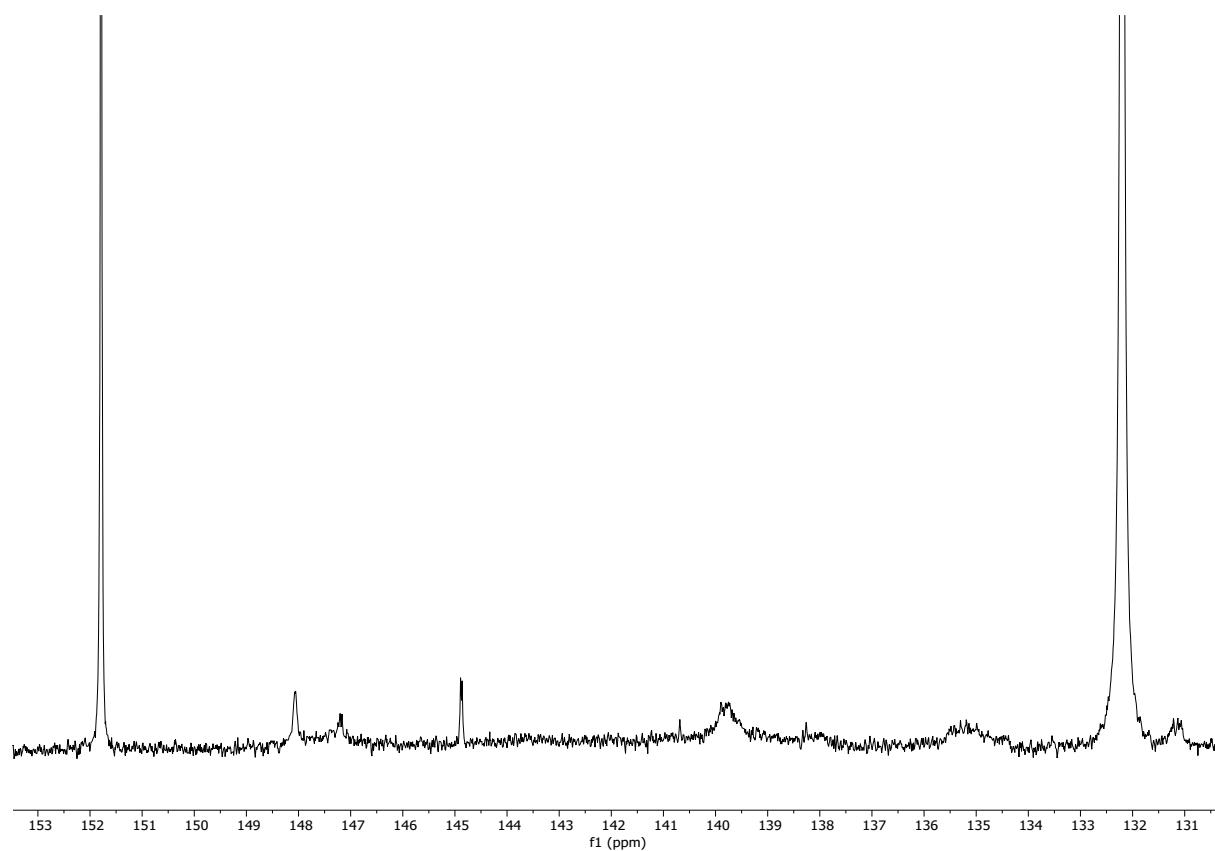


Figure SI 28: ^{31}P NMR spectrum of the **1-dodecene** hydroesterification on lignin.

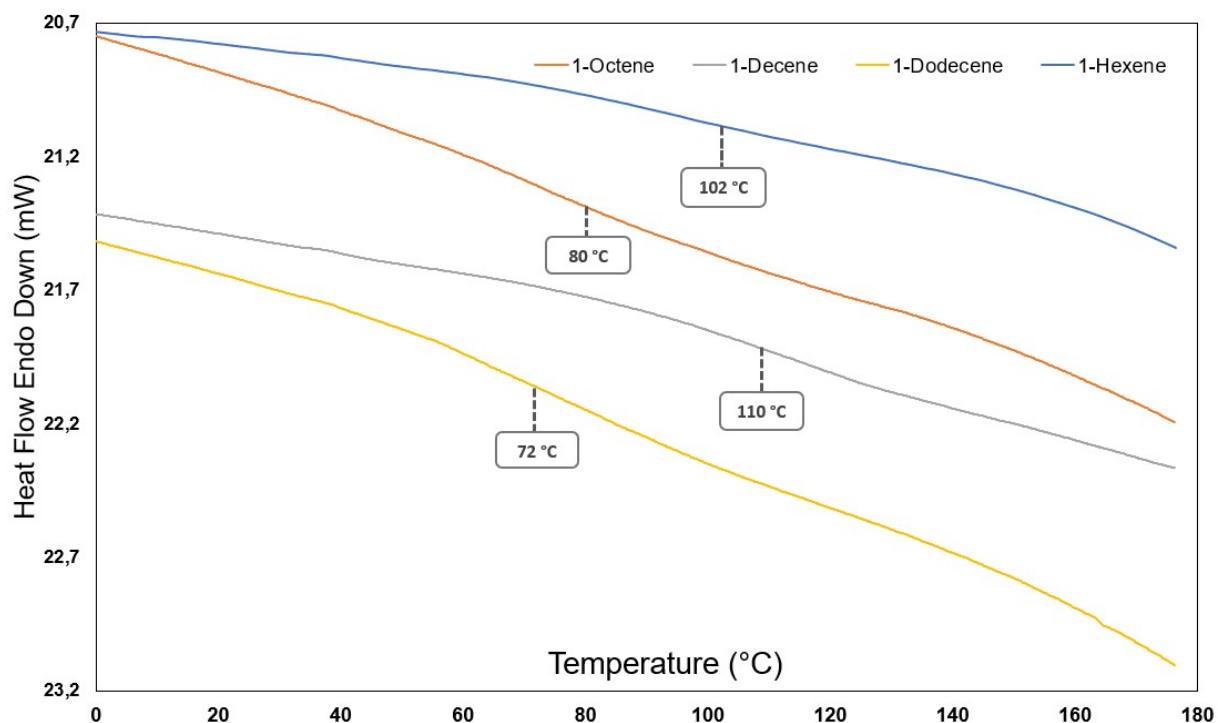


Figure SI 29: DSC Comparison of modified Kraft lignin with olefins.

NMR spectra of Table 4: hydroesterification of fatty acids, fatty ester and estragole on kraft lignin

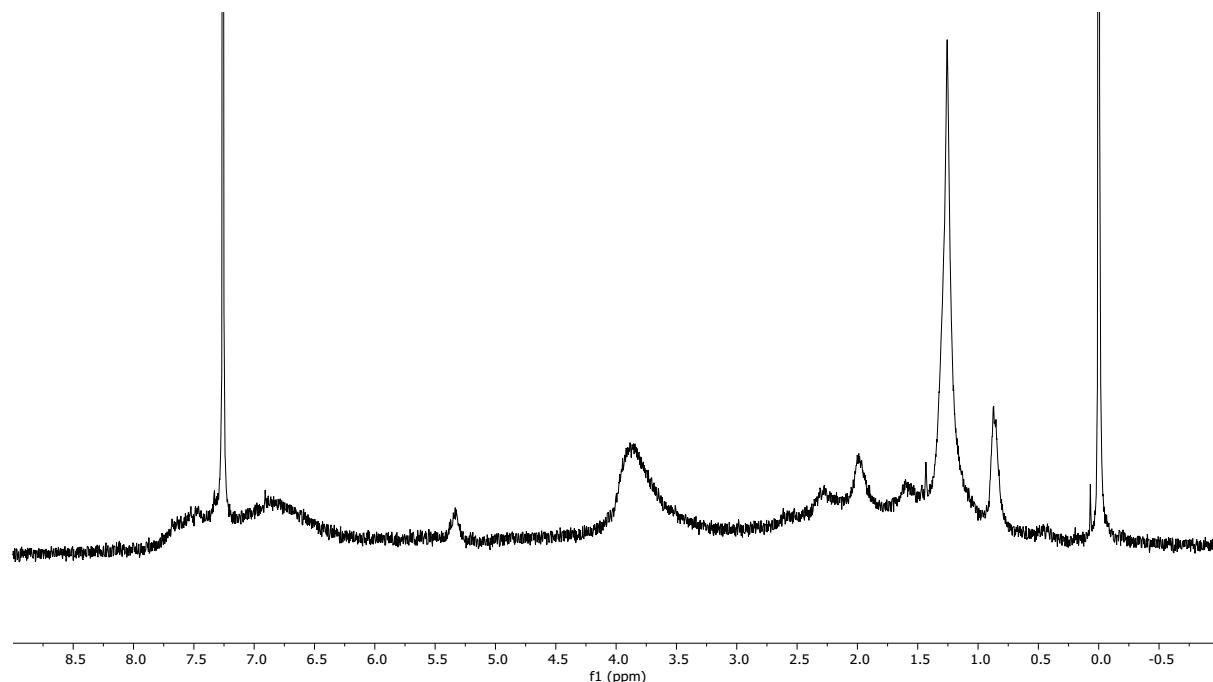


Figure SI 30: ¹H NMR spectrum of the **oleic acid** hydroesterification on lignin in CDCl_3 .

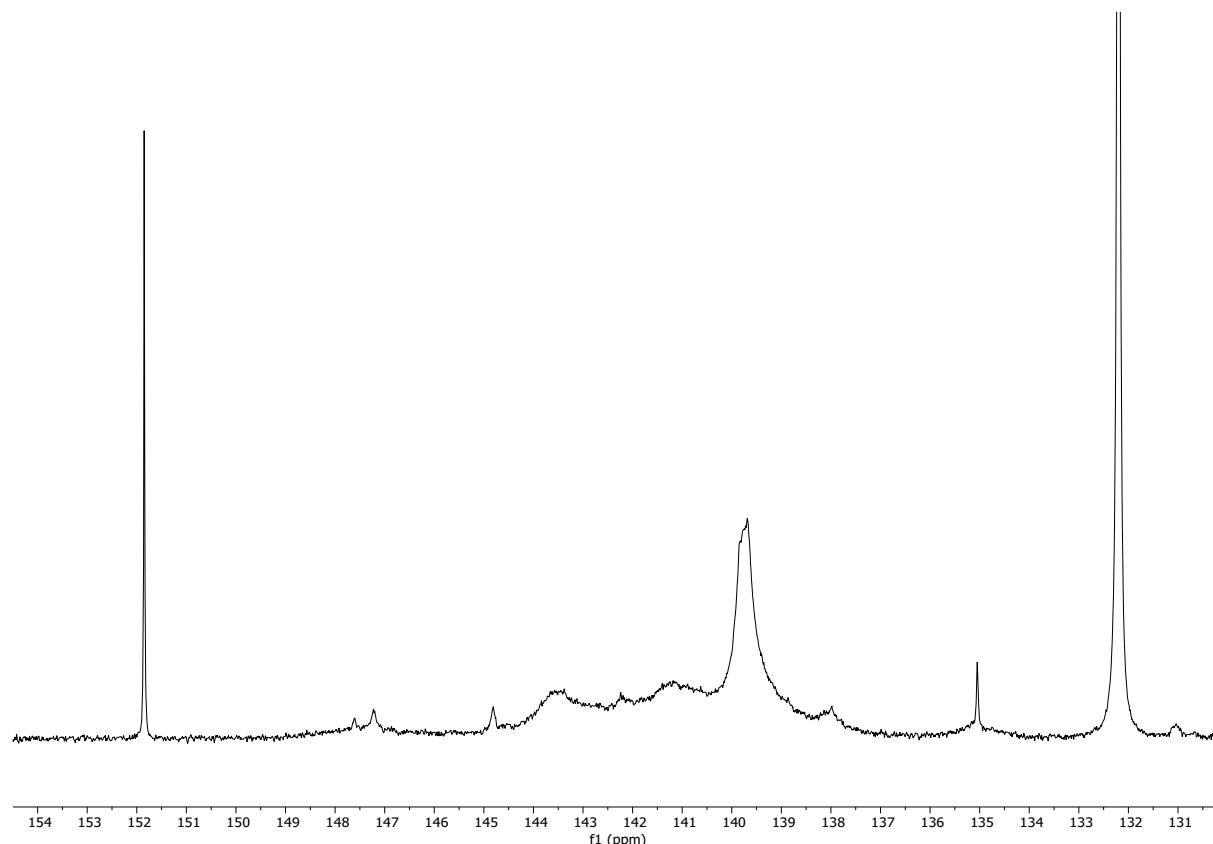


Figure SI 31: ³¹P NMR spectrum of the **oleic acid** hydroesterification on lignin.

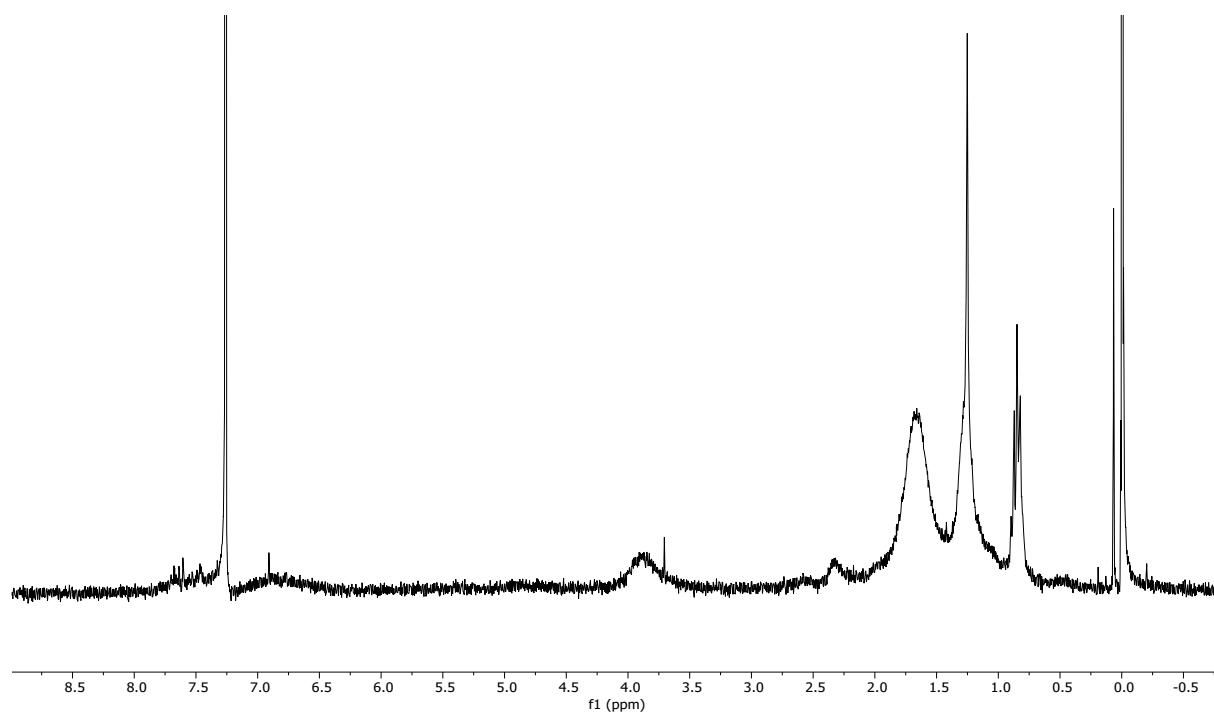


Figure SI 32: ¹H NMR spectrum of the **undecylenic acid** hydroesterification on lignin in CDCl₃.

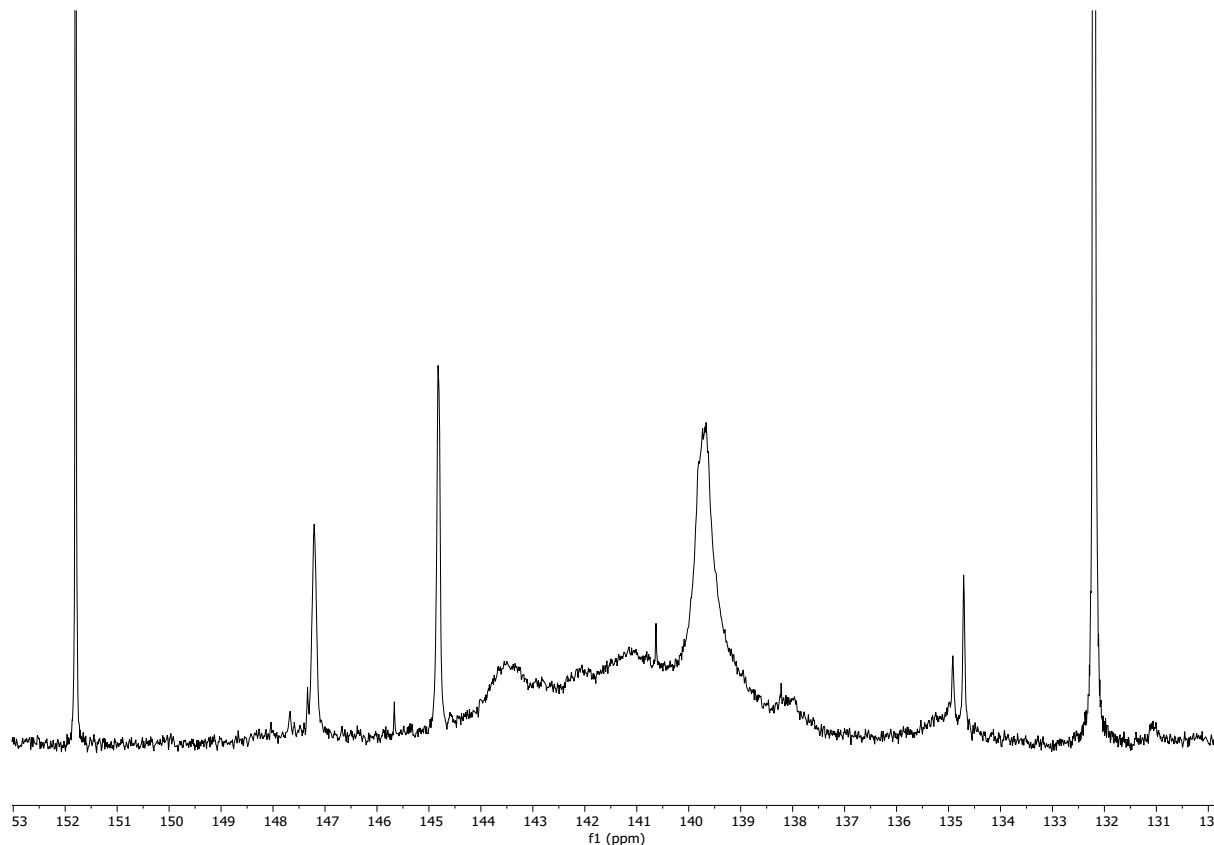


Figure SI 33: ³¹P NMR spectrum of the **undecylenic acid** hydroesterification on lignin.

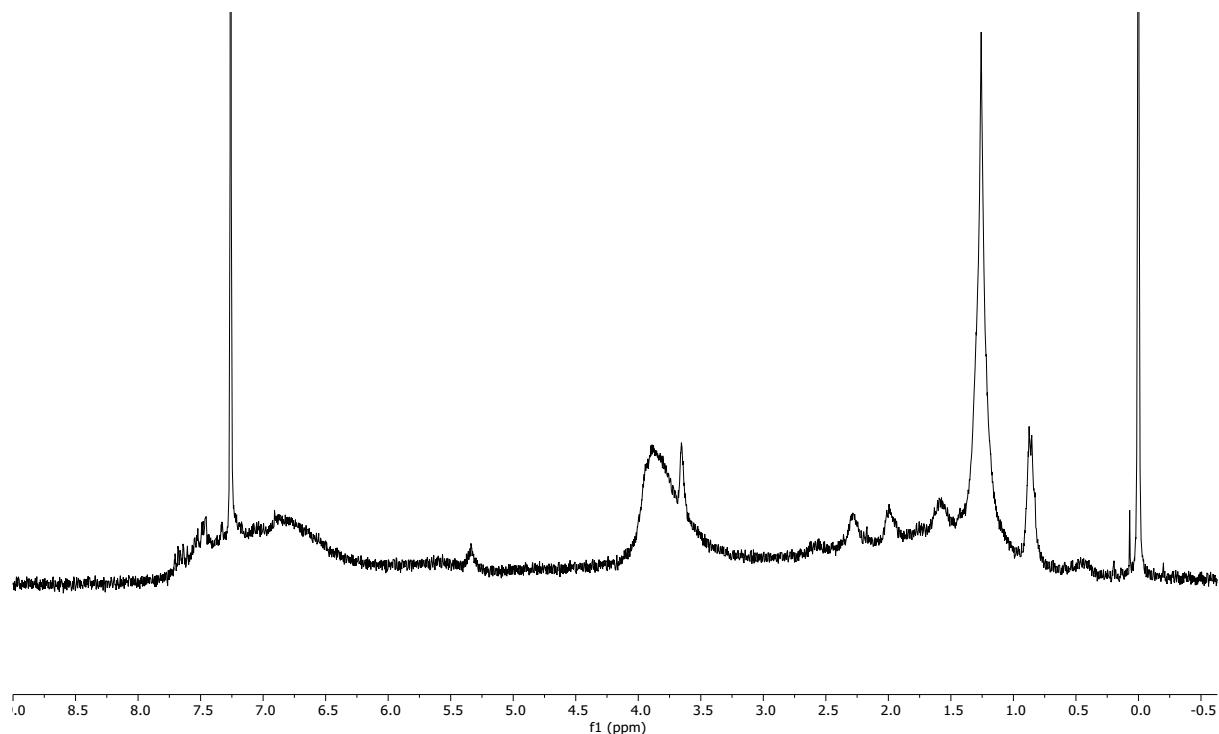


Figure SI 34: ¹H NMR spectrum of the **methyl oleate** hydroesterification on lignin in CDCl₃.

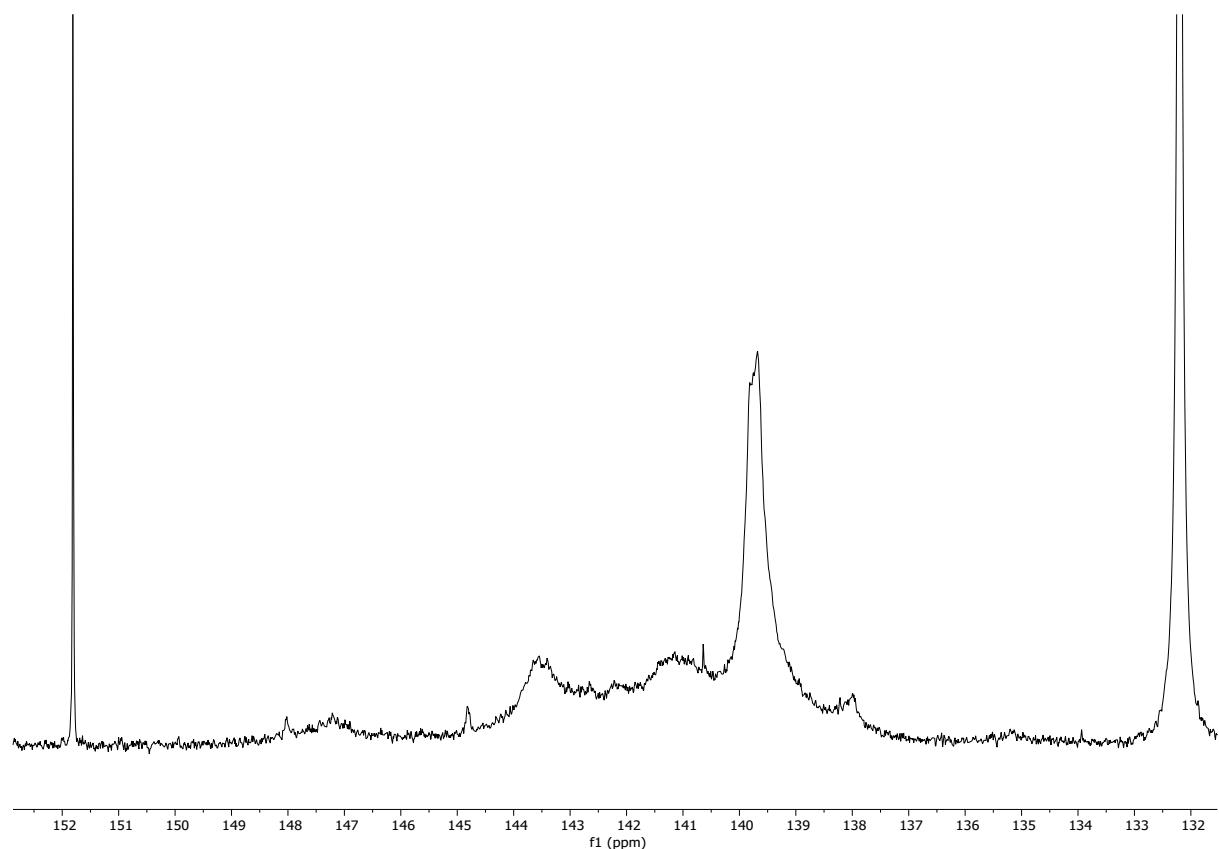


Figure SI 35: ³¹P NMR spectrum of the **methyl oleate** hydroesterification on lignin.

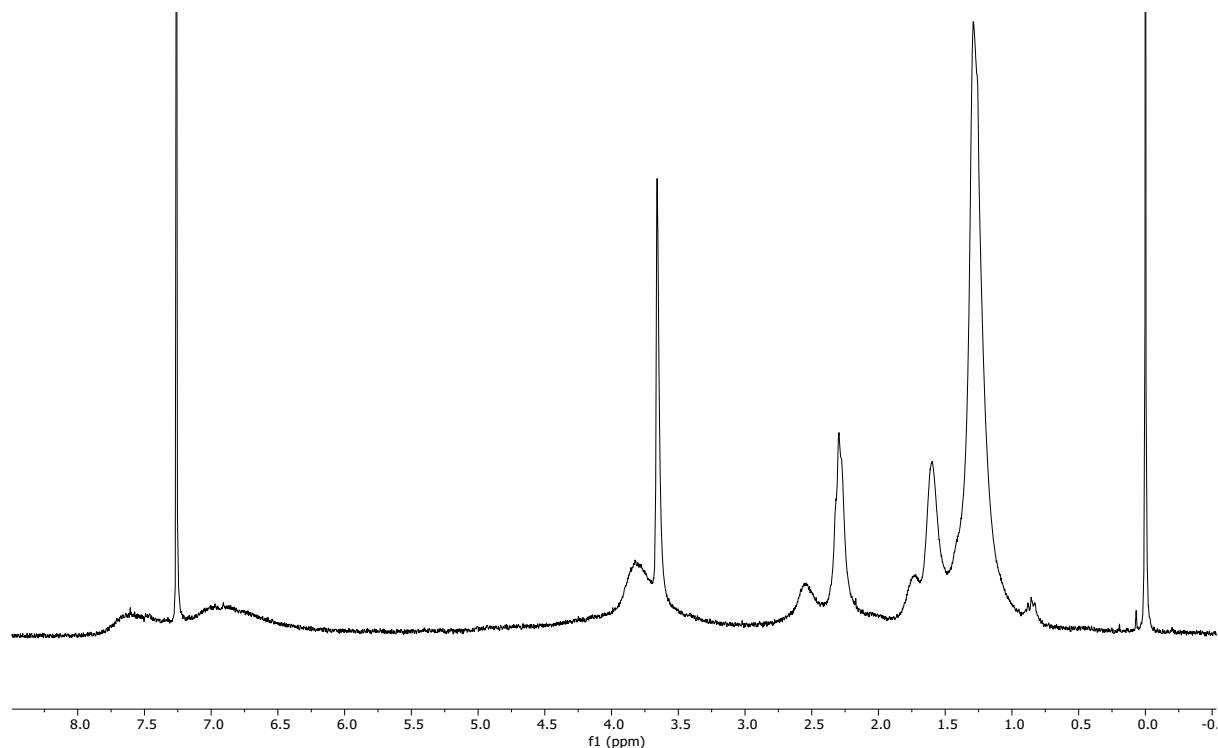


Figure SI 36: ¹H NMR spectrum of the **Methyl 10-undecenoate** hydroesterification on lignin in CDCl₃.

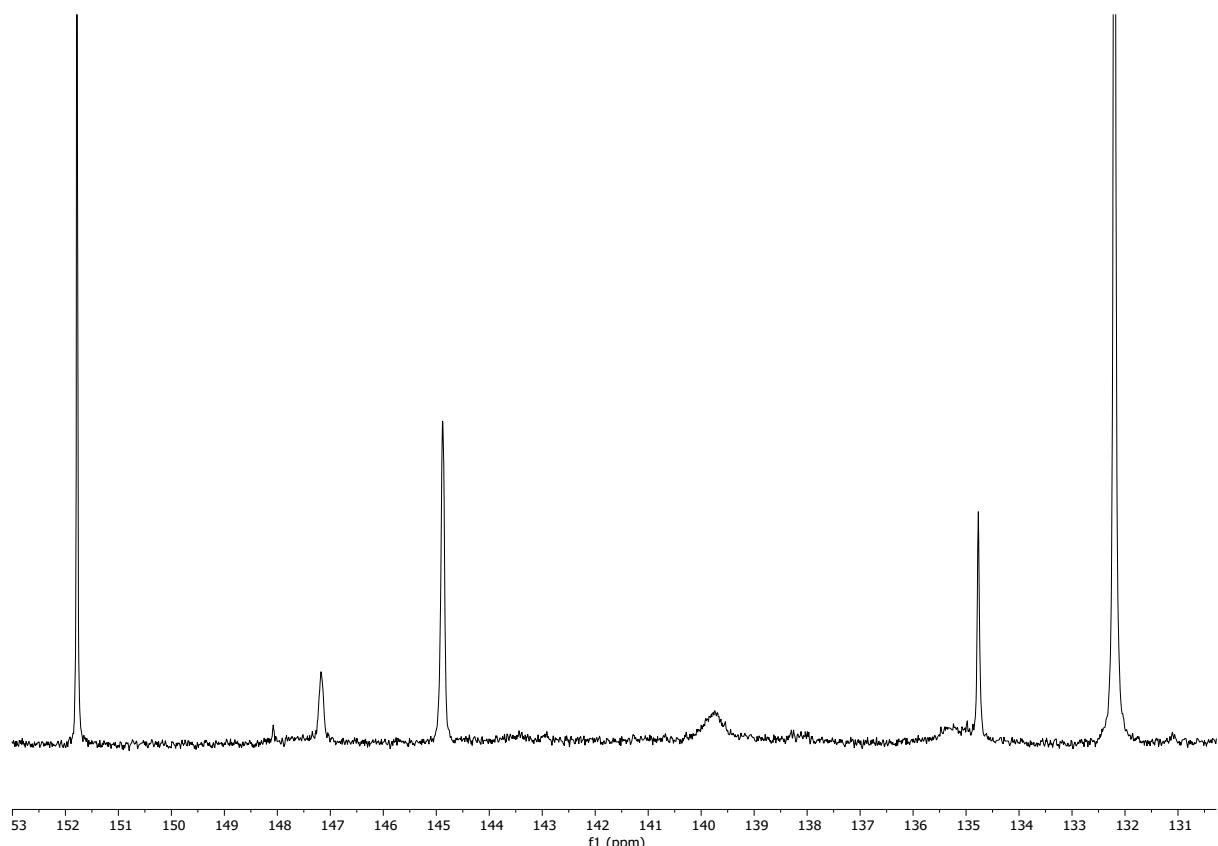


Figure SI 37: ³¹P NMR spectrum of the **Methyl 10-undecenoate** hydroesterification on lignin.

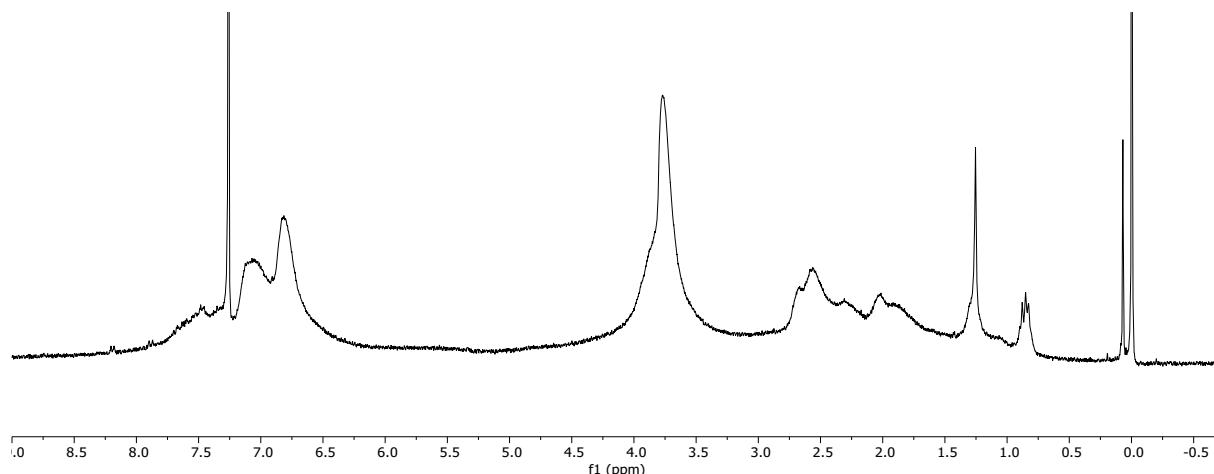


Figure SI 38: ^1H NMR spectrum of the **estragole** hydroesterification on lignin in CDCl_3 .

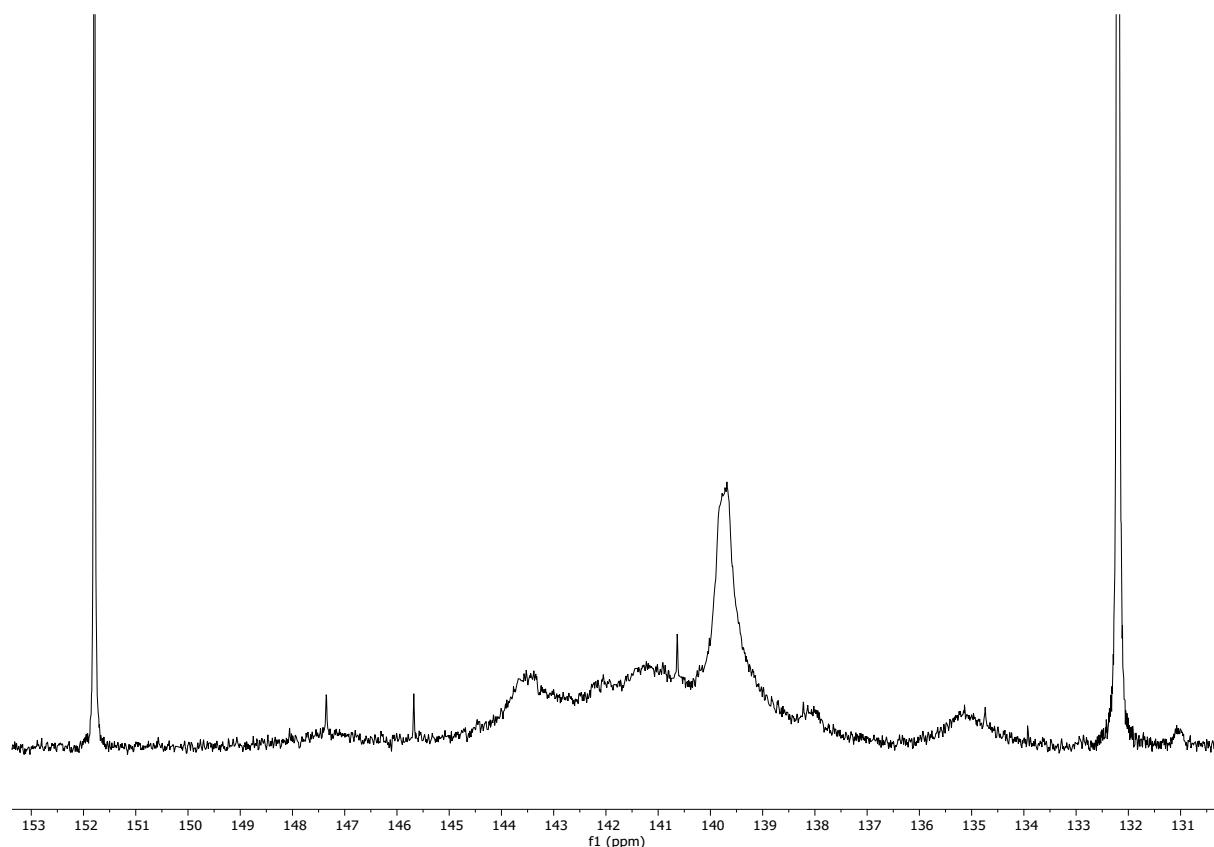


Figure SI 39: ^{31}P NMR spectrum of the **estragole** hydroesterification on lignin.

Table SI 3: Supplementary catalytic experiments of butadiene hydroesterification with Kraft lignin

Entry	$\text{Pd}(\text{OAc})_2$ (eq/OH)	Benzoic Acid (eq/Pd)	T (°C)	Time	Substitution rate ^[a]
1	1%	30	80	15 h	0 %
2	1%	30	90	15 h	5 %
3	1%	30	100	15 h	5 %

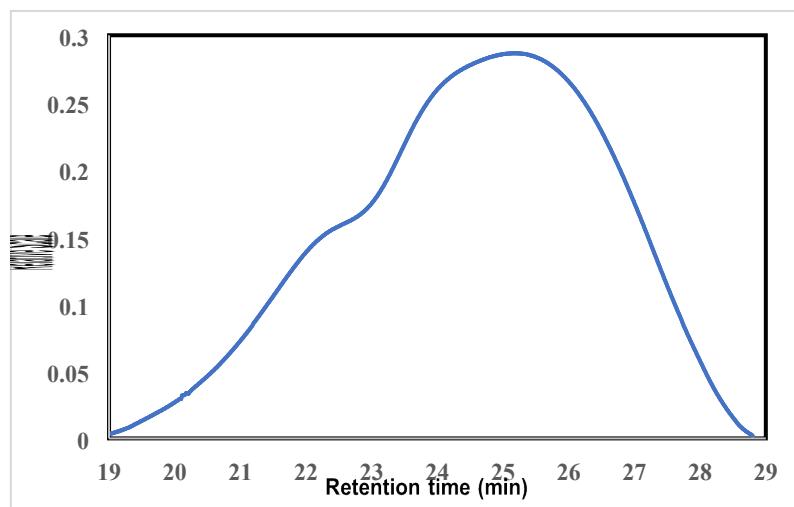
4	1%	30	120	15 h	43 %
5	1%	0	120	15 h	6 %
7	0,5%	30	120	15 h	13 %
8	1%	30	120	15 h	43 %
9	1,6%	30	120	15 h	67 %
10	0,5%	30	110	48 h	47 %
11	1%	30	110	48 h	46 %

Conditions: 500 mg Kraft lignin, 6eq/Pd dppb, 7 eq/OH butadiene, 10 mL distilled and degassed THF, 50 bar CO. (a) determined by ^1H NMR with pentafluorobenzaldehyde as internal standard using the αCH_2 of the ester between 2.8 ppm and 3.45 ppm. Substitution rate as a function of total alcohols present in lignin.
 [b] 400 mg of Kraft lignin.

Table SI 4 - SEC analyses: the number-average molecular weight (M_n), the weight-average molecular weight (M_w), and molar mass dispersity (D), for native and modified lignins

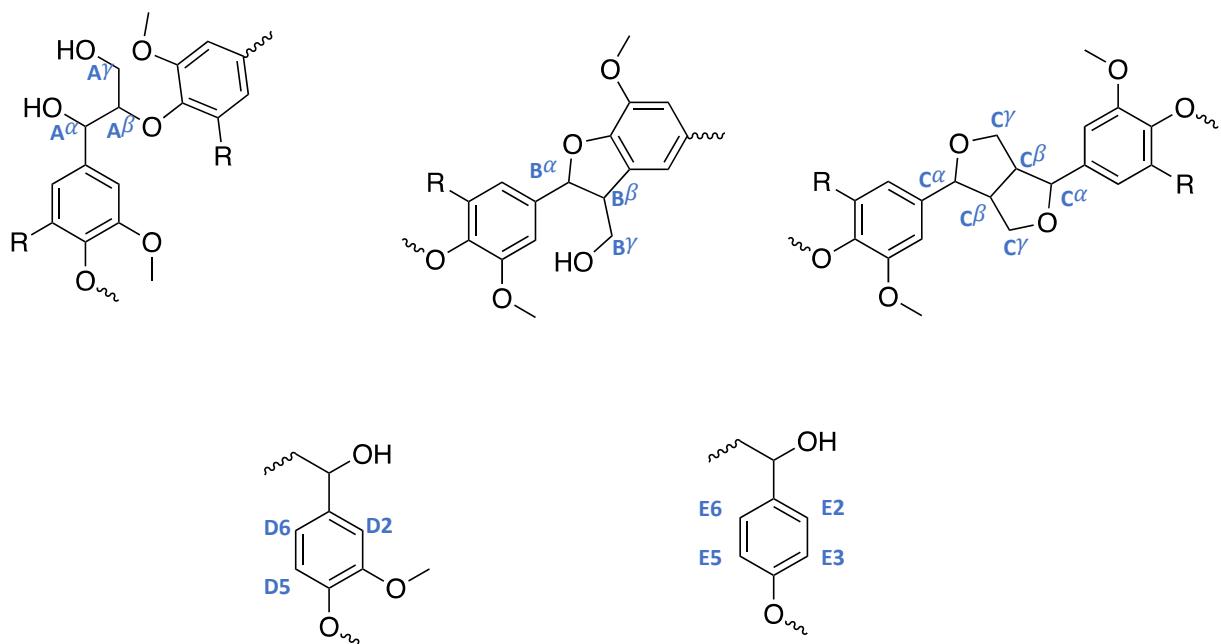
Lignin	olefin	M_n	M_w	Dispersity $D = M_w/M_n$
Acetylated Kraft	-	1800	5600	3.1
Acetylated Soda	-	1100	2700	2.5
Acetylated Organosolv	-	1400	3700	2.6
Kraft	Hexene	2600	4400	1.7
Soda	Hexene	2800	5100	1.8
Organosolv	Hexene	2100	3500	1.6
Kraft	Octene	3000	5100	1.7
Kraft	Decene	2500	4000	1.6
Kraft	Dodecene	3100	6700	2.2
Kraft	Methyl oleate	1800	2900	1.6
Kraft	Methyl 10-undecenoate	4000	7700	1.9
Kraft	Oleic acid	2000	3300	1.7
Kraft	4-Allylanisole	2300	4400	1.9
Kraft	Undecylenic acid	2600	4700	1.8
Kraft	Butadiene	5100	12000	2.3

Figure SI 40 - Size exclusion chromatography (SEC) of lignin modified by hydroesterification of butadiene



2D HSQC NMR of Technical and modified lignin by 1-hexene hydroesterification

NMR interpretation based on the study: S. Constant, H. L. J. Wienk, A. E. Frissen, P. de Peinder, R. Boelens, D. S. van Es, R. J. H. Grisel, B. M. Weckhuysen, W. J. J. Huijgen, R. J. A. Gosselink and P. C. A. Bruijnincx, Green Chem., 2016, 18, 2651–2665.



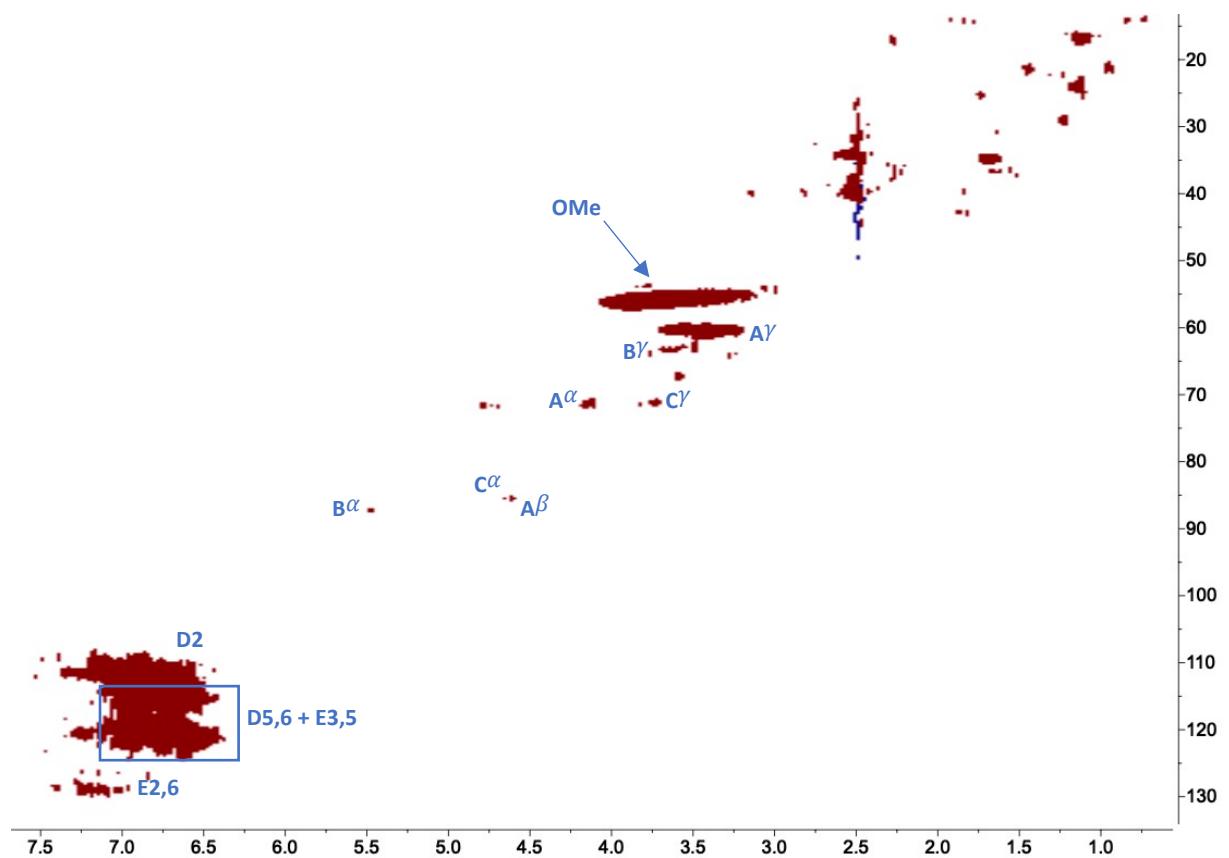


Figure SI 41: 2D HSQC NMR spectrum of technical Kraft lignin.

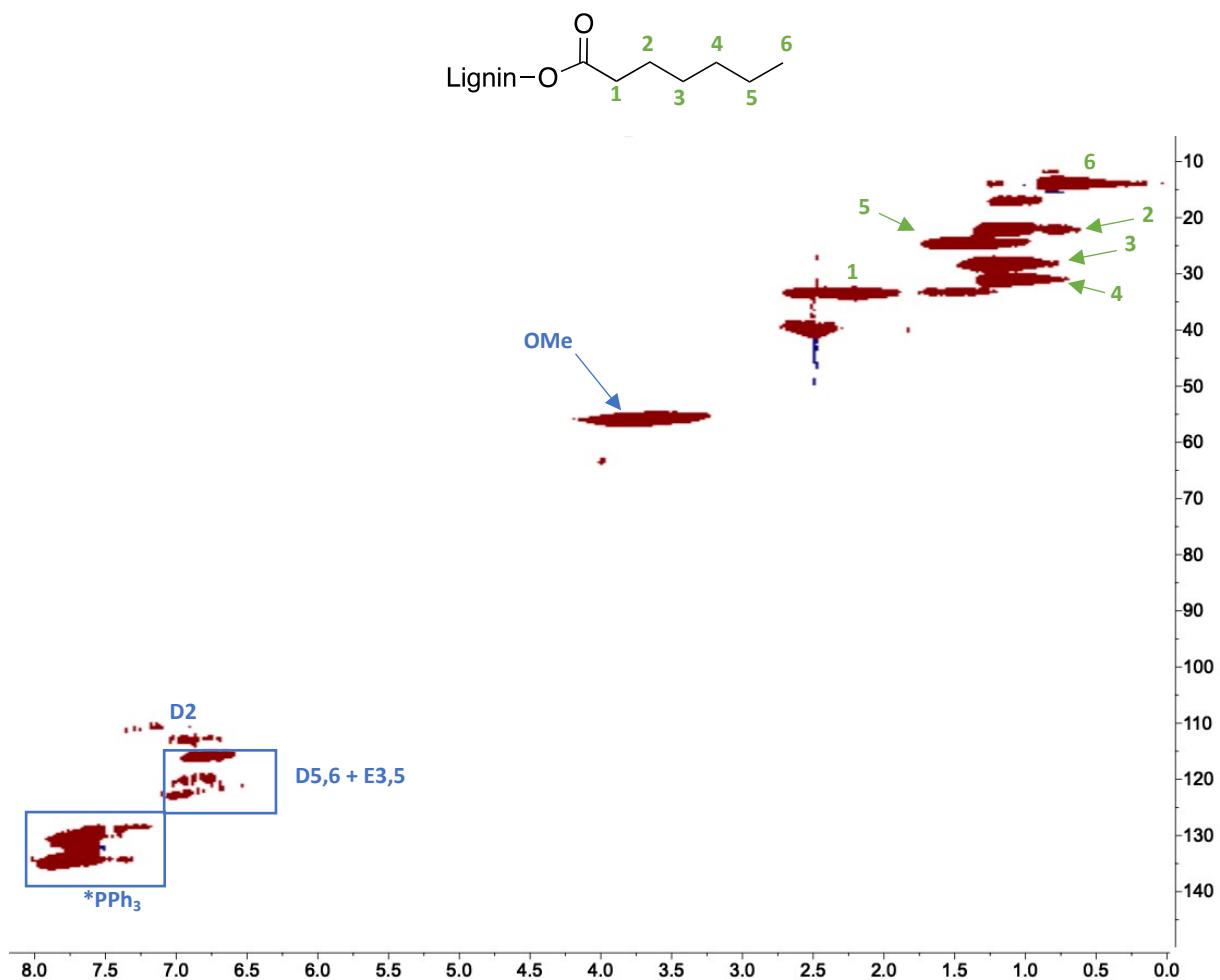


Figure SI 42: 2D HSQC NMR spectrum of the 1-hexene hydroesterification on Kraft lignin