Supporting Information

Utilizing Biocatalysis and a Sulfolane-mediated Reductive Acetal Opening to Access Nemtabrutinib from Cyrene

Nadine Kuhl,*^a Ben W. H. Turnbull,*^a Yining Ji,*^b Reed T. Larson,^a Michael Shevlin,^a Christopher K. Prier,^a Cheol K. Chung, ^a Richard Desmond,^a Erik Guetschow,^b Cyndi Qixin He,^c Tetsuji Itoh,^a Jeffrey T. Kuethe,^a Justin A. Newman,^b Mikhail Reibarkh,^b Nelo R. Rivera,^b Gao Shang,^a Zhixun Wang,^a Daniel Zewge,^b David A. Thaisrivongs^a

^a Process Research and Development, Merck & Co., Inc., Rahway, New Jersey 07065, United States

^b Analytical Research and Development, Merck & Co., Inc., Rahway, New Jersey 07065, United States

^c Computational and Structural Chemistry, Merck & Co., Inc., Rahway, New Jersey 07065, United States

E-mail: nadine.kuhl@merck.com, ben.turnbull@merck.com, yining.jichen@merck.com

Table of Contents

Contents

1. General Experimental Details	3
2. Biocatalytic Transamination	5
2.1. Reductive Amination of Cyrene TM	5
2.2. Screening of Transaminase Enzymes	5
2.3. Transaminase Reactions on Gram-Scale	7
2.4. Characterization of Compound 4	8
2.5. Synthesis and Characterization of 3a ·TsOH and 3b ·TsOH	9
2.6. Synthesis of Cyrene Amine Salts1	1
2.7. Synthesis of Bn-3a and Bn-3b 1	2
3. Reductive Acetal Opening	4
3.1. Reaction Optimization using Bn-3a 14	4
3.2. Solvent Screening using Bn-3a 1	5
3.3. Synthesis and Characterization of Bn-1a and Bn-1b1	7
3.4. Screening of Cyrene Amine Salts	0
3.5. Synthesis and Characterization of 1a TsOH	0
4. Mechanistic Experiments	2
4.1. Reaction Profile for the Synthesis of 1a · TsOH by ¹ H NMR Spectroscopy2	2
4.2. Identification and Characterization of Diborane by NMR Spectroscopy	3

	4.3. Delayed Cyrene Amine Addition Experiment	25
	4.3. Dependence of the Induction Period on Water Content	25
	4.4 Importance of Sulfolane for Diborane Formation	27
	4.5. Reaction Kinetics – Reaction Rate Dependence on Sulfolane	28
	4.6. Reductive Acetal Opening with other Sulfones	28
	4.7. Reaction with BF ₃ ·THF	29
5.	Computational Studies	30
	5.1. Computational Methods	30
	5.2. Calculated Activation Free Energy Difference for the Reduction of Cyrene TM with BH ₄ ⁻	30
	5.3. Calculated Thermodynamics for the Reaction of Cyrene TM with Isopropylamine	31
	5.4. Calculated Solvent-Sulfolane Displacement Equilibria	31
6.	X-Ray Crystallography data for 3a·TsOH and 1a·TsOH	34
	6.1. Crystal Data and Structure Refinement for Compound 3a · TsOH (CCDC 2215969)	34
	6.2. Crystal Data and Structure Refinement for Compound 1a·TsOH (CCDC 2215968)	36
7.	NMR Spectra	38

1. General Experimental Details

Unless otherwise noted, all reactions were performed under N₂-atmosphere. Heating of reaction mixtures was performed using a temperature-controlled hotplate equipped with stirring and an active thermocouple. Stirring of reaction mixtures was performed using magnetic stirring, unless noted otherwise. Evaporation and concentration *in vacuo* steps were performed using variable vacuum *via* a vacuum-controlled (ca. 400–40 mmHg) rotary evaporator. Column chromatography was performed using a Teledyne ISCO CombiFlash® Rf+ chromatography system using prepacked single-use silica packed cartridges (RediSep Rf Gold Normal-Phase Silica, 20–40 micron average particle size, 60 Å average pore size).

Materials.

Reagents were purchased in reagent grade from commercial suppliers and used without further purification, unless otherwise described. CyreneTM was purchased from Sigma-Aldrich or Circa Group directly. Sulfolane was purchased from Sigma-Aldrich or Alfa Aesar and used as received. Anhydrous solvents (acetonitrile, anisole, cyclopentyl methyl ether, dichloromethane, 1,4-dioxane, DMSO, and 1,2-dichloroethane, *N*,*N*-dimethylacetamide, *N*-methyl-2-pyrrolidone, 1,2-dimethoxyethane) were obtained from Sigma-Aldrich as part of their Sure/SealTM bottles product line. NMR kinetic experiments utilized anisole-*d*₈ which was purchased from Sigma-Aldrich. CD₂Cl₂, CD₃OD, and DMSO-*d*₆ were purchased from Cambridge Isotope Laboratories in sealed ampules and used as received.

Instrumentation.

Nuclear Magnetic Resonance Spectroscopy (NMR). Proton nuclear magnetic resonance (¹H NMR) spectra, carbon nuclear magnetic resonance (¹³C NMR) spectra, and proton-decoupled fluorine nuclear magnetic resonance (¹⁹F NMR) spectra were recorded at 25 °C on a Bruker 500 MHz AVANCE III HD spectrometer equipped with a SmartProbe. Chemical shifts for proton and carbon are reported in parts per million downfield from tetramethylsilane and are referenced to residual proton resonances of the NMR solvent according to values reported in the literature.^{1 19}F NMR spectra are not calibrated by an internal reference. Data are represented as follows: chemical shift, multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants (*J*) quoted in hertz (Hz) and integration. Analysis of crude reaction mixtures during the reaction optimization for amine **3a** and **1a** and wt% purity analysis of isolated products was performed by quantitative ¹H NMR using the following parameters: D1 = 60 s, NS = 8, and either maleic acid, CH₂Br₂ or 1,3,5-trimethoxybenzene as the internal standard. Analysis of reaction profiles, and quantification of starting material, product, and byproducts, was done using quantitative ¹H NMR (D1 = 20 seconds, number of scans = 8, 10-degree flip angle), and ¹⁹F NMR (D1 = 10 seconds, number of scans = 4, 10-degree flip angle) analysis. Octafluoronaphthalene (0.49 equiv) was used as an internal standard for ¹⁹F NMR.

Gas Chromatography (GC). GC-FID analysis was performed using a DB-624 (6% cyanopropylphenyl/94% dimethyl polysiloxane) GC column (20 m x 0.18 mm, 1 μ m) and operating at an injection port temperature of 220 °C and an FID detector temperature of 250 °C. The oven temperature was increased at a rate of 20 °C/minute from 80 °C at t = 0 to 250 °C, and then held for 10 minutes using a constant hydrogen flow of 1 mL/minute. Typical Retention Times: Compound **3a** (4.8 minutes), Compound **3b** (4.7 minutes).

UPLC. Analysis of reactions to optimize the reaction conditions to access amine **Bn-1a** was performed by UPLC analysis using an Agilent 1290 system equipped with a UV detector ($\lambda = 210$ nm) and an Eclipse

¹ Fulmer, G. R.; Miller, A. J. M. Sherden, N. H.; Gottlieb, H. E.; Nudelman, A.; Stoltz, B. M. Bercaw, J. E.; Goldberg, K. I. *Organometallics* **2010**, *29*, 2176–2179.

Plus C18 column (50 mm x 2.1 mm, 1.8 μ m) operating at 35 °C with a flow of 1 mL/min. The binary eluent mixture of eluent A = 0.5 mM Na₂B₄O₇ in H₂O and eluent B = acetonitrile was used with a 1.5-2 min gradient starting from 95%A and 5%B to 5%A and 95% B.

UPLC Method for HRMS. HRMS sample analysis was performed using a Waters Acquity UPLC system interfaced with Thermo Scientific Orbitrap Mass Analyzer and an ESI source temperature set to 263 °C. Samples were dissolved in acetonitrile at a 0.01 mM concentration, and 2 μ L injection volumes were used for analysis. The Waters UPLC was equipped with an ACQUITY UPLCTM CSH Column (130 Å pore size, 1.7 μ m particle size, 2.1 mm internal diameter, 150 mm length; part number 186005408) operating at a 0.4 mL/min flow rate of a binary eluent mixture (eluent A and B, prepared as described below). A column temperature of 45 °C was used for all analytes. A column temperature of 60 °C was used for compound 4. The 5 min method used the following eluent gradient: a gradient from 90% A/10% B at t = 0 min held until 0.5 min, to 10% A/90% B at t = 1.0 min held until 4.0 min, and a subsequent gradient to achieve 90% A and 10% B at t = 4.1 min and final hold until t = 5.0 min. Eluent Preparation: A = 0.1% v/v formic acid in Acetonitrile.

Abbreviations. ATA = Amine Transaminase, Bn = Benzyl, Boc = *tert*-butyloxycarbonyl, calc. = calculation, CyreneTM = (1S,5R)-6,8-dioxabicyclo[3.2.1]octan-4-one, DMSO = dimethyl sulfoxide, ESI = electrospray ionization, equiv = equivalents, GC = gas chromatography, UPLC = ultra-high pressure liquid chromatography, HRMS = high resolution mass spectrometry, PLP = (4-Formyl-5-hydroxy-6-methylpyridin-3-yl)methyl dihydrogen phosphate or pyridoxal-5'-phosphate, QToF = quadrupole time of flight, Tf = Trifluoromethanesulfonate, 2-MeTHF = 2-methyltetrahydrofuran, THF = tetrahydrofuran, Ts = *para*-toluenesulfonate.

Safety. Please note that reaction conditions to form **Bn-1a** and **1a**·**TsOH** require handling of BH_3 ·THF or involve the *in-situ* generation of H_2 and diborane gas. H_2 gas is flammable, BH_3 ·THF and diborane are highly reactive, flammable and acutely toxic. Appropriate PPE and engineering controls should be used when performing those reactions. Adequate process safety analysis and equipment selection should be performed prior to scaling-up those reaction conditions. For more information see reference 22 in the manuscript.

2. Biocatalytic Transamination

2.1. Reductive Amination of CyreneTM

A 40 mL vial equipped with a stir bar was charged with CyreneTM (500 mg, 0.4 mL, 3.9 mmol, 1.0 equiv). A solution of NH₄OAc (3.0 g, 39.0 mmol, 10.0 equiv) in methanol (8 mL) was added followed by heatactivated 3 Å molecular sieves (MS) (550 mg). Sodium cyanoborohydride (245 mg, 3.9 mmol, 1.0 equiv) was added and the reaction was stirred at room temperature for 5.5 hours until full conversion was observed by ¹H NMR. An aliquot was diluted 4 times with methanol, filtered, and analyzed by GC to determine the product distribution (Scheme S1, Figure S1).



Scheme S1. Reductive amination of Cyrene[™] with NaBH₃CN.

Figure S1. Gas chromatogram (GC) of the reductive amination of Cyrene[™] with NaBH₃CN.

2.2. Screening of Transaminase Enzymes

General Procedure 1. CyreneTM (500 mg, 0.4 mL, 3.9 mmol, 1.0 equiv) and (4-formyl-5-hydroxy-6methylpyridin-3-yl)methyl dihydrogen phosphate (PLP, 10 mg, 40 µmol, 1 mol%) were dissolved in 9.6 mL buffer solution (0.1 M Na₂B₄O₇ 10·H₂O, 1 M isopropylamine, 2.5 equiv relative to CyreneTM, pH 9.5) to reach a total volume of about 10 mL and a CyreneTM concentration of 50 g/L. 500 µL (25 mg, 0.39 mmol, 1.0 equiv of CyreneTM) of this solution were added to 2.5 mg of transaminase variant (lyophilized cell-free powder, 10 wt% relative to CyreneTM) in vials and incubated at 35 °C with shaking. After 20 hours, 100 µL of each reaction was diluted with 600 µL of 0.5% maleic acid (internal standard) in D₂O for analysis by quantitative ¹H NMR (see Figure S2 for a representative ¹H NMR and Table S1 and S2 for results.) Scheme S2. Biocatalytic transaminase reaction.



Table (21	Corooning	of	Cod	avia®	trancominaça	01077710000
I able a	51.	Screening	01	Cou	exis	transammase	enzymes.

Entry	Transaminase Enzyme	Assay Yield [%]	3a:3b d.r.
1	ATA-007	11	1:1
2	ATA-009	10	1.1:1
3	ATA-013	65	2.5:1
4	ATA-014	74	2.3:1
5	ATA-015	68	1.8:1
6	ATA-016	100	1.7:1
7	ATA-024	100	1.3:1
8	ATA-036	67	1:1.6
9	ATA-234	86	1:6.9
10	ATA-273	100	1:4.8
11	ATA-303 ²	95	1.9:1
12	ATA-426 ³	57	17:1
13	CDX-010	91	1.5:1
14	CDX-017 ⁴	72	1.1:1

Table S2. ATA-426 enzyme loading study.

Entry ^a	ATA-426 Loading [wt%]	Assay Yield [%]	3a:3b d.r.
1	20	71%	17:1
2	15	62%	18:1
3	10	57%	17:1
4	5	42%	>20:1

^{*a*} Reaction conditions according to general protocol 1, above.

² Limanto, J. et al. Org. Lett. 2014, 16, 2716–2719.
³ Yasuda, N. et al. Org. Process Res. Dev. 2017, 21, 1851–1858.
⁴ Savile, C. K. et al. Science 2010, 329, 305–309.



Figure S2. ¹H NMR at 500 MHz in D_2O of the crude reaction mixture for entry 14 of Table S1. Integration of the singlet at 5.58 ppm was used to calculate the combined assay yield of **3a** and **3b**. The diastereomeric ratio was determined via integration of the doublets at 4.15 ppm (**3a**) and 4.08 ppm (**3b**).

2.3. Transaminase Reactions on Gram-Scale

Representative procedure for Entry 7 of Table 1. In a 100 mL Mettler Toledo EasyMax vessel equipped with an overhead stirrer, (4-formyl-5-hydroxy-6-methylpyridin-3-yl)methyl dihydrogen phosphate (PLP) (41 mg, 160 µmol, 0.7 mol%) was dissolved in 0.1 M Na₂B₄O₇·10 H₂O buffer (60 mL, pH 9.5) containing isopropylamine (4.4 mL, 51.5 mmol, 2.2 equiv). CyreneTM (3.0 g, 23.4 mmol, 1.0 equiv) and transaminase enzyme ATA-426 (600 mg, 20 wt%) were then added at room temperature. The mixture was heated to 45 °C and agitated for 21 h. During the reaction, vacuum (400-450 mbar) and airflow were applied to remove the acetone generated. The pH was monitored throughout the reaction using a pH probe. The pH was adjusted manually over the course of the reaction to keep the pH above 7.5 (with 5 N aqueous NaOH) yielding a solution of **3a** (2.76 g, 21.4 mmol, 91% assay yield, 24:1 d.r.). The assay yield was determined by diluting an aliquot of the reaction mixture by a factor of 6 using a 0.5% solution of maleic acid (internal standard) in D₂O.

Higher concentrations of the borate buffer, 0.25 M and 0.40 M concentrations were also explored following the reaction conditions above using a 100 g/L Cyrene concentration. The assay yields were 95% and 94%, respectively, and a pH adjustment was still required. Using a 0.1 M borate buffer at 100 g/L Cyrene yielded **3a** in 93% assay yield, see Section 2.5). Therefore, higher buffer concentrations did not provide a significant benefit.

2.4. Characterization of Compound 4



1-((1*S*,5*R*)-4-hydroxy-6,8-dioxabicyclo[3.2.1]octan-4-yl)propan-2-one (4). A 2.5 M solution of *n*-butyllithium in hexanes (62.3 mL, 156 mmol, 0.95 equiv) was added over 2 h and 15 minutes to a solution of diisopropylamine (21.8 mL, 156 mmol, 0.95 equiv) in cyclopentyl methyl ether (210 mL) in a 500 mL round bottom flask at 0 °C. The resulting solution was stirred for 1 h and then cooled to -78 °C. Acetone (10.8 mL,

148 mmol, 0.9 equiv) was added slowly to maintain the reaction temperature below -68 °C and the resulting mixture was stirred for an additional 2 h at -78 °C. CyreneTM (16.8 mL, 164 mmol, 1.00 equiv) in cyclopentyl methyl ether (10 mL) was added and the reaction was allowed to stir overnight at -78 °C. The reaction was then quenched with a saturated aqueous solution of NH₄Cl (200 mL) at room temperature. The phases were separated, and the organic phase was dried over anhydrous Na₂SO₄, filtered, and concentrated to obtain the crude product as an oil. The crude product was purified by column chromatography using hexanes/ethyl acetate to yield compound **4**, a mixture of two diastereomers, as a pale-yellow oil (8.8 g, 94.5 wt% purity, 44.7 mmol, 27% yield, 1.2:1 d.r.). The isolated material contained 4.2 GC area% CyreneTM that could not be separated.

¹**H** NMR (500 MHz, CD₂Cl₂) δ 5.10 (s, 1H), 5.08 (s, 1H), 4.46 (br s, 2H), 3.84 – 3.79 (m, 2H), 3.76 – 3.70 (m, 2H), 3.17 (br s, 2H), 2.85 (d, *J* = 16.3 Hz, 1H), 2.63 (d, *J* = 16.3 Hz, 1H), 2.56 (s, 2H), 2.18 (s, 3H), 2.16 (s, 3H), 2.09 – 2.01 (m, 1H), 1.88 – 1.57 (m, 6H), 1.55 – 1.49 (m, 1H), 1.45 – 1.40 (m, 1H) ppm. ¹³C NMR (126 MHz, CD₂Cl₂) δ 210.18 (CO), 209.47 (CO), 104.37 (CH), 103.90 (CH), 73.73 (CH), 73.33 (CH), 72.17 (C_q), 71.19 (C_q), 68.32 (CH₂), 67.38 (CH₂), 49.93 (CH₂), 47.43 (CH₂), 32.32 (CH₃), 32.30 (CH₃), 29.85 (CH₂), 28.40 (CH₂), 27.34 (CH₂), 25.64 (CH₂) ppm. HRMS m/z calc. for [C₉H₁₅O₄] ([M+H]⁺): 187.0965, found: 187.0961.

2.5. Synthesis and Characterization of 3a TsOH and 3b TsOH

NH₂

3a•HOTs

.0

(1S,4R,5R)-6,8-Dioxabicyclo[3.2.1]octan-4-aminium 4-methylbenzene-1-sulfonate (3a·TsOH). In a 1 L jacketed round bottom flask, (4-formyl-5-hydroxy-6-• HOTs methylpyridin-3-yl)methyl dihydrogen phosphate (PLP) (1.0 g, 4.0 mmol, 1.0 mol%) was dissolved in 0.1 M Na₂B₄O₇ ·10 H₂O buffer (500 mL, pH 9.5) containing 1.56 M isopropylamine (67 mL, 778 mmol, 2 equiv). Transaminase enzyme ATA-426 (10 g,

20 wt%) was then added and dissolved at room temperature. Cyrene[™] (49.9 g, 389 mmol, 1.0 equiv) was then added, and the mixture was heated to 33-37 °C and agitated with overhead stirring for 27 h. During the reaction, vacuum (400-450 mbar) and airflow were applied to remove the acetone generated. The pH was monitored at various points during the reaction using a pH probe and was adjusted if needed to keep the pH between 7.9 and 8.6 (four additions of a total of 17.5 mL of 50 wt% aqueous NaOH) yielding a solution of **3a** (46.8 g, 362 mmol, 93% assay yield, 17:1 d.r.). The assay yield was determined by diluting an aliquot of the reaction mixture by a factor of 6 using a 0.5% solution of maleic acid (internal standard) in D₂O.



Figure S3. ¹H NMR at 500 MHz in D₂O of the crude reaction mixture. The * indicates compound 4.

Next, the solution pH was adjusted to 13.4 by adding 50 wt% aqueous NaOH solution (28 mL). The mixture was cooled to 10-15 °C and potassium carbonate (83.3 g) was added in 3 portions to the stirring solution maintaining the temperature below 25 °C. 2-MeTHF (300 mL) was added, and the resulting mixture was stirred at room temperature overnight to allow for enzyme denaturing. The denatured protein solids were then filtered off. The filter cake was washed with 2-MeTHF (3 x 50 mL). The two phases of the filtrate were separated, and the aqueous phase was extracted with 2-MeTHF (75 mL). The combined organic layers were then concentrated under reduced pressure to remove isopropylamine to <0.5 mol% with respect to the Cyrene[™] amine. The resulting crude solution was then diluted with 2-MeTHF to a total volume of 200 mL, and the water content (KF) was adjusted to 3-3.6 wt% by adding water. A solution of para-toluenesulfonic acid monohydrate (73.6 g, 387 mmol, 0.99 equiv) in 2-MeTHF (150 mL) was added dropwise over 4 h at 40 °C, the resulting slurry was cooled to room temperature and aged for 2 h. The solids were filtered, washed with wet 2-MeTHF (50 mL, KF = 2 wt%) followed by dry 2-MeTHF (50 mL), and then dried overnight at 50 °C under vacuum with nitrogen sweep to yield **3a**·**TsOH** as a white solid (92.1 g, 99.4 wt% purity, 306 mmol, 79% yield, >20:1 d.r.).

¹**H NMR (500 MHz, DMSO-***d*₆) δ 8.02 (br s, 3H), 7.57 – 7.39 (m, 2H), 7.14 – 7.13 (m, 2H), 5.40 (s, 1H), 4.59 – 4.58 (m, 1H), 3.97 (d, *J* = 7.4 Hz, 1H), 3.74 – 3.56 (m, 1H), 3.17 – 2.99 (m, 1H), 2.29 (s, 3H), 2.14 – 1.94 (m, 2H), 1.62 – 1.50 (m, 1H), 1.49 – 1.35 (m, 1H) ppm. ¹³C NMR (126 MHz, DMSO-*d*₆) δ 145.24 (C_q), 137.94 (C_q), 128.17 (CH), 125.47 (CH), 98.61 (CH), 73.07 (CH), 66.99 (CH₂), 47.44 (CH), 23.21 (CH₂), 20.78 (CH₃), 18.59(CH₂) ppm. HRMS m/z calc. for [C₆H₁₂NO₂] ([M+H]⁺): 130.0863, found: 130.0861.

The relative and absolute stereochemistry of $3a \cdot TsOH$ was confirmed by X-ray crystallography, see Section 6.1.



Boc-3b

tert-Butyl ((1*S*,4*S*,5*R*)-6,8-Dioxabicyclo[3.2.1]octan-4-yl)carbamate (Boc-3b). In a 1 L 3-necked vessel equipped with an overhead stirrer, (4-formyl-5-hydroxy-6-methylpyridin-3-yl)methyl dihydrogen phosphate (PLP) (600 mg, 2.4 mmol, 1.0 mol%) was dissolved in 0.1 M Na₂B₄O₇·10 H₂O buffer (600 mL, pH 9.5) containing 1.50 M isopropylamine (50 mL, 585 mmol, 2.5 equiv). ATA-234 (4.5 g, 15 wt%) was then added

and dissolved at room temperature. CyreneTM (30 g, 234 mmol, 1.0 equiv) was added, and the mixture was heated to 45 °C and agitated for 22 h. During the reaction, vacuum (400 mbar) and nitrogen flow were applied to remove the acetone generated. The stream was then cooled to room temperature and the pH was adjusted to pH 12 using 50 wt% aqueous NaOH. To remove isopropylamine, the reaction mixture was distilled at 80 mbar and 48 °C jacket temperature using an air sweep. During the distillation additional 50 wt% aqueous NaOH was charged to maintain the pH at 12. THF (200 mL) was charged, and the distillation was continued until isopropylamine was < 0.5 wt% by ¹H NMR. The remaining aqueous solution was filtered over celite to remove the denatured enzyme yielding an aqueous solution of **3b** (195 mmol, 83% assay yield, 5:1 d.r.).

To the aqueous solution at pH 12 was then added to a solution of di-*tert*-butyl dicarbonate (Boc₂O, 51.1 g, 234 mmol, 1.0 equiv.) in THF (70 mL) *via* syringe pump over 40 min upon which the pH dropped to pH 7.55. The pH was adjusted to pH 12 using 50 wt% aqueous NaOH and the reaction was stirred at room temperature for 22 h. Methyl *tert*-butyl ether (500 mL) was then charged, and the mixture was stirred for 30 min. The mixture was filtered using methyl *tert*-butyl ether, the layers of the filtrate were separated, and the aqueous layer was extracted with methyl *tert*-butyl ether (300 mL). The combined organic layers were washed with 20 wt% aqueous NaCl solution (300 mL) and then concentrated. The isolated solid was recrystallized twice from methylcyclohexane to yield **Boc-3b** as a white solid (32.3 g, 141 mmol, 60% yield, 44:1 d.r).

¹**H NMR (500 MHz, DMSO**-*d*₆) δ 6.71 (d, J = 7.5 Hz, NH, 1H), 5.14 (s, 1H), 4.47 (s, 1H), 3.76 (d, J = 7.1 Hz, 1H), 3.61- 3.59 (m, 1H), 3.38 – 3.34 (m, 1H), 1.77 – 1.67 (m, 1H), 1.67 – 1.47 (m, 3H), 1.37 (s, 9H) ppm; ¹³**C NMR (126 MHz, DMSO**-*d*₆) δ 154.95 (CO), 101.06, 77.77 (C(CH₃)₃), 72.12, 67.51 (CH₂), 50.59, 28.18 (CH₃), 27.46 (CH₂), 21.42 (CH₂) ppm. **HRMS** m/z calc. for [C₁₁H₂₀NO₄] ([M+H]⁺): 230.1387, found: 230.1383.



(1S,4S,5R)-6,8-Dioxabicyclo[3.2.1]octan-4-aminium 4-methylbenzene-1-sulfonate (3b·TsOH). Boc-3b (31.7 g, 138 mmol, 1.0 equiv) was dissolved in a solution of paratoluenesulfonic acid monohydrate (79 g, 415 mmol, 3 equiv) in 2-MeTHF (159 mL) and the reaction was heated to 35 °C overnight. The resulting slurry was cooled to room temperature and the solids were filtered, washed with 2-MeTHF (100 mL), and then

dried under vacuum yielding 3b·TsOH as a white solid (35.1 g, 99.2 wt% purity, 116 mmol, 84% vield, >20:1 d.r.).

¹**H** NMR (500 MHz, DMSO- d_6) δ 7.91 (s, 3H), 7.48 (d, J = 8.1 Hz, 2H), 7.12 (d, J = 7.9 Hz, 2H), 5.39 (s, 1H), 4.60 (s, 1H), 3.90 (d, J = 7.3 Hz, 1H), 3.70 (t, J = 6.2 Hz, 1H), 3.15 (br s, 1H), 2.29 (s, 3H), 1.91 - $1.74 (m, 2H), 1.66 - 1.57 (m, 2H) ppm; {}^{13}C NMR (126 MHz, DMSO-d_{6}) \delta 145.65 (C_{a}), 137.62 (C_{a}), 128.04$ (CH), 125.46 (CH), 98.88 (CH), 72.39 (CH), 67.84 (CH₂), 49.68 (CH), 26.56 (CH₂), 20.76 (CH₃), 20.66 (CH₂) ppm. **HRMS** m/z calc. for $[C_6H_{12}NO_2]$ ($[M+H]^+$): 130.0863, found: 130.0861.

2.6. Synthesis of Cyrene Amine Salts



3a•HOTf

(1S,4R,5R)-6,8-Dioxabicyclo[3.2.1]octan-4-aminium trifluoromethanesulfonate (3a·HOTf). A 9.86 wt% solution of 3a (78.4 g, 59.9 mmol, 1.0 equiv.) in 2-MeTHF was charged into a 250 mL round bottom flask and then concentrated under vacuum (100-150 mbar) at a heating bath temperature of 40 °C. 2-MeTHF (50 mL) was added, and the process was repeated to remove water and isopropylamine. The residue was

dissolved in dichloromethane (50 mL) and the resulting solution was cooled in an ice bath. A solution of TfOH (5.3 mL, 59.9 mmol, 1.0 equiv) in dichloromethane (70 mL) was added slowly via an addition funnel and the mixture was allowed to warm to room temperature overnight. The resulting slurry was filtered under nitrogen atmosphere and the solid was washed with dichloromethane and dried under vacuum to yield **3a·HOTf** as a light yellow solid (16.5 g, 98.3 wt% purity, 58.2 mmol, 97% yield, >20:1 d.r.).

¹**H NMR (500 MHz, DMSO-** d_6) δ 7.93 (s, NH₃, 3H), 5.38 (s, 1H), 4.72 - 4.48 (m, 1H), 3.98 (d, J = 7.3 Hz, 1H), 3.77 - 3.60 (m, 1H), 3.12 (s, 1H), 2.20 - 1.87 (m, 2H), 1.60 - 1.47 (m, 1H), 1.50 - 1.38 (m, 1H) ppm; ¹⁹F{¹H} NMR (471 MHz, DMSO- d_6) δ -77.74 ppm; ¹³C NMR (126 MHz, DMSO- d_6) δ 120.67 (q, J = 322.4 Hz, CF₃), 98.60 (C6), 73.07 (C2), 66.99 (C1), 47.39 (C5), 23.21 (C3), 18.60 (C4) ppm.



(1S,4R,5R)-6,8-Dioxabicyclo[3.2.1]octan-4-aminium hydrochloride (3a·HCl). A 250 mL 3-neck round bottom equipped with an overhead stirrer was charged with a 1.15 M solution of 3a (20 mL, 23 mmol, 1 equiv) in 2-MeTHF. 2-MeTHF (24 mL) was added, and the solution was heated to 50 °C. HCl in cyclopentyl methyl ether (7.7 ml, 23.0 mmol,

1.0 equiv) was added at 48 °C over 1 hour. The resulting slurry was cooled to room temperature and aged overnight. The slurry was filtered and the solid was washed with EtOH (2 x 10 mL). After drying under vacuum, 3a·HCl was obtained as a white crystalline solid (2.89 g, 99.8 wt% purity, 17.4 mmol, 76% yield, >20:1 d.r.).

¹**H NMR (500 MHz, DMSO-d₆)** δ 8.30 (s, 3H, NH₃, 3H), 5.45 (s, 1H), 4.59 (d, J = 3.6 Hz, 1H), 4.11 - 3.87 (m, 1H), 3.77 - 3.55 (m, 1H), 3.15 - 2.92 (m, 1H), 2.19 - 1.89 (m, 2H), 1.69 - 1.53 (m, 1H), 1.45-1.42 (m, 1H) ppm; ¹³C NMR (126 MHz, DMSO-d₆) δ 98.61, 72.99, 66.95, 47.31, 23.25, 18.55 ppm.



(1*S*,4*R*,5*R*)-6,8-Dioxabicyclo[3.2.1]octan-4-aminium hydrobromide (3a·HBr). A 9.86 wt% solution of 3a (87.9 g, 67.1 mmol, 1.0 equiv.) in 2-MeTHF was charged into a 250 mL round bottom flask and then concentrated under vacuum (100-150 mbar) at 40 °C. 2-MeTHF (50 mL) was added, and the process was repeated to remove water and isopropylamine. 2-MeTHF (70 mL) was added followed by a slow addition of aqueous

HBr (9.1 mL, 81.0 mmol, 1.2 equiv.) over 5-10 minutes at 40 °C. The biphasic mixture was then concentrated at 40 °C, 2-MeTHF (50 mL) was then added, and the process was repeated to remove excess water. The residue was dissolved in 2-MeTHF (50 mL) and heated to 40 °C. Methanol (5 mL) was added and the solution was allowed to cool to room temperature overnight. The resulting slurry was filtered and washed 2-MeTHF (2 x 20 mL). The isolated solid was recrystallized from 2-MeTHF (60 mL) and methanol (6 mL) at 40 °C, followed by a recrystallization in methanol (25 mL) at 60 °C. 2-MeTHF (10 mL) was added at room temperature to improve the recovery. The resulting solid was filtered, washed with 2-MeTHF (2 x 20 mL), and dried under vacuum to yield **3a·HBr** as a white solid (7.86 g, 100 wt% purity, 37.4 mmol, 56% yield, >20:1 d.r.).

¹**H NMR (500 MHz, DMSO**-*d*₆) δ 8.09 (s, 3H), 5.42 (s, 1H), 4.60 (d, J = 4.6 Hz, 1H), 3.98 (dd, J = 7.3 Hz, J = 0.6 Hz, 1H), 3.74 - 3.60 (m, 1H), 3.16 - 3.03 (m, 1H), 2.19 - 1.83 (m, 2H), 1.67 - 1.50 (m, 1H), 1.49 - 1.36 (m, 1H) ppm; ¹³**C NMR (126 MHz, DMSO**-*d*₆) δ 98.52, 73.02, 67.00, 47.36, 23.24, 18.54 ppm.

2.7. Synthesis of Bn-3a and Bn-3b

(1S,4R,5R)-N-benzyl-6,8-Dioxabicyclo[3.2.1]octan-4-amine (Bn-3a). A 250 mL round "O NHBn bottom flask was charged with 3a·TsOH (6.0 g, 19.9 mmol, 1.0 equiv.) and methanol (7 mL). Sodium acetate (3.3 g, 39.8 mmol, 2.0 equiv) and MgSO₄ (1.2 g, 10.0 mmol, Bn-3a 0.5 equiv) were added under nitrogen atmosphere. Benzaldehyde (2.0 mL, 19.9 mmol, 1.0 equiv) was added and the slurry was stirred at room temperature for 1 hour. Sodium cyanoborohydride (2.5 g, 39.8 mmol, 2.0 equiv) was added in one portion and the reaction was heated to 50 °C overnight. The cooled reaction was filtered using ethyl acetate and the filtrated was concentrated under vacuum. The resulting residue was dissolved in ethyl acetate (50 mL), washed with water (50 mL), and then with a 2 M aqueous K₂CO₃ solution (2 x 25 mL). The organic phase was dried over MgSO₄, filtered, and concentrated The crude oil was then purified by column chromatography under vacuum. using dichloromethane/methanol to yield Bn-3a as a colorless oil (3.79 g, 96.7 wt% purity, 16.73 mmol, 84% yield, >20:1 d.r.).

¹**H** NMR (500 MHz, CD₂Cl₂) δ 7.37 – 7.33 (m, 2H), 7.33 – 7.28 (m, 2H), 7.26 – 7.20 (m, 1H), 5.34 – 5.30 (s, 1H), 4.42 (s, 1H), 3.89 (d, *J* = 7.0 Hz, 1H), 3.87 (d, *J* = 13.3 Hz, 1H), 3.76 (d, *J* = 13.3 Hz, 1H), 3.71 (ddd, *J* = 7.0 Hz, *J* = 5.2 Hz, *J* = 1.3 Hz, 1H), 2.63 – 2.52 (m, 1H), 2.05 – 1.93 (m, 1H), 1.84 (tt, *J* = 13.4 Hz, *J* = 5.4 Hz, 1H), 1.75 – 1.63 (m, 2H), 1.45 – 1.33 (m, 1H) ppm; ¹³C NMR (126 MHz, CD₂Cl₂) δ 141.55, 128.80, 128.55, 127.31, 103.39, 73.82, 67.39, 55.44, 51.62, 25.73, 20.02 ppm. LCMS m/z calc. for [C₁₃H₁₈NO₂] ([M+H]⁺): 220.1, found: 220.2.



(1*S*,4*R*,5*R*)-*N*-benzyl-6,8-Dioxabicyclo[3.2.1]octan-4-aminium 4-methylbenzene-1sulfonate (Bn-3a·TsOH). A 500 mL round bottom flask was charged with 3a·TsOH (10.0 g, 33.2 mmol, 1.0 equiv.) and methanol (120 mL). Sodium acetate (5.4 g, 66.4 mmol, 2.0 equiv) and MgSO₄ (2.0 g, 16.6 mmol, 0.5 equiv) were added under re Renzeldebyde (3.4 mL 33.2 mmol, 1.0 equiv) was added and the slurry was stirred

nitrogen atmosphere. Benzaldehyde (3.4 mL, 33.2 mmol, 1.0 equiv) was added and the slurry was stirred

at room temperature for 1.5 hours. Sodium cyanoborohydride (4.2 g, 66.4 mmol, 2.0 equiv) was added in one portion and the reaction was heated to 50 °C overnight. The cooled reaction was filtered using ethyl acetate and the filtrated was concentrated under vacuum. The resulting residue was dissolved in ethyl acetate (100 mL), washed with water (50 mL), and then with a 2 M aqueous K₂CO₃ solution (2 x 50 mL). The organic phase was dried over MgSO₄, filtered, and concentrated under vacuum. The crude oil was dissolved in 2-MeTHF (15 mL) and a solution of *para*-toluenesulfonic acid monohydrate (6.3 g, 33.2 mmol, 1.0 equiv) in 2-MeTHF (15 mL) was added at room temperature over 30 minutes using an addition funnel. Additional 2-MeTHF (40 mL) was added via the addition funnel and the resulting slurry was stirred overnight at room temperature. The solid was filtered, washed with 2-MeTHF (20 mL, then 30 mL), and dried under vacuum to yield **Bn-3a·TsOH** as a white solid (10.2 g, 93.1 wt% purity, 24.3 mmol, 73% yield). ¹H NMR (500 MHz, DMSO-d₆) δ 8.95 (br s, NH₂, 2H), 7.56 - 7.50 (m, 2H), 7.50 - 7.40 (m, 5H), 7.18 -7.05 (m, 2H), 5.63 (s, 1H), 4.64 (br s, 1H), 4.28 (d, J = 13.1 Hz, 1H), 4.22 (d, J = 13.1 Hz, 1H), 3.99 (d, J = 13.1 Hz, 1H), = 7.0 Hz, 1H), 3.68 (ddd, J = 7.0 Hz, J = 5.1 Hz, J = 1.1 Hz, 1H), 3.09 (br s, 1H), 2.29 (s, 3H), 2.19 - 2.06 (m, 1H), 2.05 - 1.95 (m, 1H), 1.85 - 1.76 (m, 1H), 1.45 (dd, J = 13.7 Hz, J = 5.8 Hz, 1H) ppm; ¹³C NMR (**126 MHz, DMSO-***d*₆) δ 145.77 (C_a), 137.54 (C_a), 131.61 (C_a), 130.24 (CH), 129.08 (CH), 128.67 (CH), 128.01 (CH), 125.47 (CH), 97.55 (CH), 73.16 (CH), 67.20 (CH₂), 53.84 (CH), 48.88 (CH₂), 23.32 (CH₂), 20.75 (CH₃), 16.78 (CH₂) ppm. LCMS m/z calc. for [C₁₃H₁₈NO₂] ([M+H]⁺): 220.1, found: 220.2. HRMS m/z calc. for $[C_{13}H_{18}NO_2]$ ($[M+H]^+$): 220.1332, found: 220.1330.



(1*S*,4*S*,5*R*)-*N*-benzyl-6,8-Dioxabicyclo[3.2.1]octan-4-aminium 4-methylbenzene-1sulfonate (Bn-3b·TsOH). A 500 mL round bottom flask was charged with 3b·TsOH (10.0 g, 33.2 mmol, 1.0 equiv.) and methanol (120 mL). Sodium acetate (5.4 g, 66.4 mmol, 2.0 equiv) and MgSO₄ (2.0 g, 16.6 mmol, 0.5 equiv) were added under

nitrogen atmosphere. Benzaldehyde (3.4 mL, 33.2 mmol, 1.0 equiv) was added and the slurry was stirred at room temperature for 1 hour. Sodium cyanoborohydride (4.2 g, 66.4 mmol, 2.0 equiv) was added in one portion and the reaction was heated to 50 °C overnight. The cooled reaction was filtered using ethyl acetate and the filtrated was concentrated under vacuum. The resulting residue was dissolved in ethyl acetate (100 mL), washed with water (100 mL), and then with a 2 M aqueous K_2CO_3 solution (2 x 50 mL). The organic phase was dried over MgSO₄, filtered, and concentrated under vacuum. The crude oil was dissolved in 2-MeTHF (15 mL) and a solution of *para*-toluenesulfonic acid monohydrate (6.3 g, 33.2 mmol, 1.0 equiv) in 2-MeTHF (15 mL) was added at room temperature over 30 minutes using an addition funnel. Additional 2-MeTHF (40 mL) was added *via* the addition funnel and the resulting slurry was stirred overnight at room temperature. The solid was filtered, washed with 2-MeTHF (2 x 20 mL), and dried under vacuum to yield **Bn-3b-TsOH** as a white solid (11.4 g, 93.4 wt% purity, 27.2 mmol, 82% yield).

¹**H** NMR (500 MHz, DMSO-*d*₆) δ 8.92 (d, J = 28.2 Hz, NH₂, 2H), 7.55 – 7.38 (m, 7H), 7.11 (d, J = 7.8 Hz, 2H), 5.63 (s, 1H), 4.62 (d, J = 4.6 Hz, 1H), 4.20 (d, J = 13.8 Hz, 1H), 4.15 (d, J = 13.8 Hz, 1H), 3.94 (d, J = 7.4 Hz, 1H), 3.78 – 3.67 (m, 1H), 3.21 (br s, 1H), 2.29 (s, 3H), 2.09 – 1.96 (m, 1H), 1.83 – 1.64 (m, 3H) ppm; ¹³C NMR (126 MHz, DMSO-*d*₆) δ 145.76 (C_q), 137.55 (C_q), 131.73 (C_q), 130.09 (CH), 129.04 (CH), 128.68 (CH), 128.01 (CH), 125.47 (CH), 97.85 (CH), 72.60 (CH), 67.97 (CH₂), 56.13 (CH), 47.79 (CH₂), 26.62 (CH₂), 20.75 (CH₃), 19.02 (CH₂) ppm. LCMS m/z calc. for [C₁₃H₁₈NO₂] ([M+H]⁺): 220.1, found: 220.2. HRMS m/z calc. for [C₁₃H₁₈NO₂] ([M+H]⁺): 220.1332, found: 220.1330.

3. Reductive Acetal Opening

3.1. Reaction Optimization using Bn-3a

General Procedure 2. A tared 4 mL vial equipped with a stir bar was charged with **Bn-3a** (50 mg, 225 μ mol, 1 equiv.) followed by 0.5 mL of the respective solvent under N₂ atmosphere. The reductant, either triethylsilane (108 μ l, 674 μ mol, 3 equiv.) or BH₃·THF (1.0 M, 674 μ l, 674 μ mol, 3 equiv), was added followed by the addition of the Lewis or Bronsted Acid such as BF₃·OEt₂ (57 μ l, 449 μ mol, 2 equiv). The vial was then placed into a heating block and heated to 50 °C for 18 h before it was carefully quenched with methanol (200 μ L) and heated again to 50 °C for 2 hours. The reactions were analyzed by HPLC after diluting a weighed aliquot (80 μ L) into a 50 mL volumetric flask using acetonitrile.

Scheme S3. Acetal opening of Bn-3a. For results, see Table 2.



3.2. Solvent Screening using Bn-3a

TMDSO

Me₂SiHCl

51

6

Microscale high-throughput experimentation was conducted as previously described⁵ in aluminum 96-well microtiter plates containing 8×30 mm glass vial inserts in a nitrogen-purged glovebox with $O_2 < 10$ ppm.

Survey of Silanes and Solvents for the Ring Opening of Bn-3a.

To each well in a 96-well plate, 100 μ L of a 0.1 M stock solution of **Bn-3a** free base (2.19 mg, 10 μ mol) in 1,2-dichloroethane (DCE) was added followed by removal of the volatiles under a flow of nitrogen. 100 μ L of the desired solvent was added to each well, followed by 60 μ mol (6 equiv) of the desired silane (9.58 μ L Et₃SiH, 12.29 μ L ^{*i*}Pr₃SiH, 11.07 μ L (EtO)₃SiH, 9.61 μ L (EtO)₂MeSiH, 8.26 μ L (EtO)Me₂SiH, 7.40 μ L PhSiH₃, 10.60 μ L TMDSO, or 6.66 μ L Me₂SiHCl) and 7.40 μ L (60 μ mol, 6 equiv.) of BF₃·OEt₂. The plate was sealed, stirred using magnetic tumble stirring, and heated to 40 °C overnight (Figure S4).

AY	MeCN	DMSO	DMA	NMP	sulfolane	PC	DME	DCE	2-MeTHF	CPME	PhCF₃	PhMe
Et₃SiH	25.7	0.0	0.0	0.0	93.2	28.0	0.6	3.0	0.3	0.7	1.7	1.2
′Pr₃SiH	12.4	0.5	0.6	0.6	5.1	6.0	0.6	1.0	0.6	0.4	0.5	0.5
(EtO)₃SiH	62.7	0.6	1.3	1.0	50.0	44.7	13.8	0.9	1.1	0.6	0.9	0.6
(EtO) ₂ MeSiH	1.1	0.0	0.7	0.0	1.2	1.0	0.0	0.0	0.0	0.0	0.0	0.0
(EtO)Me ₂ SiH	37.1	0.0	0.0	0.0	30.2	16.2	0.0	0.5	0.0	0.0	0.3	0.3
PhSiH₃	23.5	0.0	0.0	0.0	83.4	13.7	0.4	1.2	0.0	0.0	0.0	0.5
TMDSO	22.3	0.6	1.0	0.3	92.2	8.6	1.5	4.9	0.4	0.4	1.9	3.0
Me ₂ SiHCl	47.1	0.0	1.8	1.8	32.3	42.3	19.2	2.8	10.6	1.7	3.0	1.4
Product d.r.	MeCN	DMSO	DMA	NMP	sulfolane	PC	DME	DCE	2-MeTHF	CPME	$PhCF_3$	PhMe
Et₃SiH	5				599	22						
′Pr₃SiH	19				2	2						
(EtO)₃SiH	121				14	19	91					
(EtO) ₂ MeSiH		_										
(EtO)Me ₂ SiH	92				6	5						
PhSiH₃	30				542	88						

Figure S4. *Top:* Assay yields (AY) as determined by UPLC analysis against an internal standard for the silane and solvent combination tested. *Bottom:* **Bn-1a:Bn-1b** ratio as determined by UPLC analysis. MeCN = acetonitrile, DMA = N,N-dimethylacetamide, NMP = N-methyl-2-pyrrolidone, PC = propylene carbonate, DME = 1,2-dimethoxyethane, CPME = cyclopentyl methyl ether, TMDSO = 1,1,3,3-tetramethyldisiloxane.

56

6

22

1

0

0

601

7

Survey of Cosolvents for the Ring Opening of Bn-3a[.]TsOH.

To each well in a 96-well plate, 100 μ L of a well-stirred 0.1 M stock slurry of **3a**·**TsOH** (3.91 mg, 10 μ mol) in 1,2-dichloroethane (DCE) was added followed by removal of the volatiles under a flow of nitrogen. 10, 25, 50, or 75 μ L of gently warmed sulfolane was added to the desired wells followed by 90, 75, 50, or 25 μ L of the desired cosolvents. 9.58 μ L (60 μ mol, 6 equiv.) of Et₃SiH or 10.60 μ L (60 μ mol, 6 equiv.) of

⁵ Shevlin, M. Practical High-Throughput Experimentation for Chemists. ACS Med. Chem. Lett. 2017, 8, 601–607.

TMDSO was added to the desired wells, followed by 7.40 μ L (60 μ mol, 6 equiv.) of BF₃·OEt₂ to each well. The plate was sealed, stirred using magnetic tumble stirring, and heated to 40 °C overnight (Figure S5).

AY	DME	2-MeTHF	CPME	MTBE	PhOMe	PhCF₃	PhCl	CH_2CI_2	PhMe	DMSO	DMA	NMP	_
10%	0.0	0.0	0.0	0.0	93.7	94.1	96.0	93.2	87.9	0.0	0.0	0.0	
25%	0.2	0.0	0.0	0.0	>99	>99	>99	>99	>99	0.8	0.6	0.5	Et SiLl
50%	0.6	0.4	0.4	0.3	95.2	>99	>99	>99	>99	0.8	0.5	0.5	
75%	>99	0.3	0.0	0.0	26.4	3.8	>99	>99	>99	0.8	0.9	0.8	
10%	0.5	0.2	0.8	0.4	0.6	0.2	0.7	22.1	3.8	0.0	0.5	0.2	
25%	0.2	0.4	0.3	0.0	88.1	0.5	84.3	98.5	78.5	0.5	0.3	0.0	TMDGO
50%	0.4	0.4	0.6	0.4	>99	>99	>99	>99	0.7	0.6	0.6	0.5	TNDSO
75%	0.5	0.5	0.5	0.4	0.4	>99	>99	>99	>99	0.6	0.4	0.3	
Product d.r.	DME	2-MeTHF	CPME	MTBE	PhOMe	PhCF₃	PhCl	CH_2CI_2	PhMe	DMSO	DMA	NMP	_
10%					>500	>500	>500	>500	>500				
25%					>500	>500	>500	>500	>500				Et SiLl
50%					135.6	>500	>500	>500	>500				
75%	70.5				>500		>500	>500	>500				
10%								17.1					
25%					125.2		63.5	117.4	33.7				TMDGO
50%					>500	>500	>500	>500					
75%						>500	>500	>500	>500				

Figure S5. *Top:* Assay yields (AY) as determined by UPLC analysis against an internal standard for the sulfolane-solvent mixtures tested. *Bottom:* **Bn-1a:Bn-1b** ratio as determined by UPLC analysis. DMA = N,N-dimethylacetamide, NMP = N-methyl-2-pyrrolidone, DME = 1,2-dimethoxyethane, CPME = cyclopentyl methyl ether, MTBE = methyl *tert*-butyl ether, TMDSO = 1,1,3,3-tetramethyldisiloxane.

3.3. Synthesis and Characterization of Bn-1a and Bn-1b

((2S,5R)-5-(benzylamino)tetrahydro-2H-pyran-2-yl)methanol (Bn-1a). Bn-3a (1.0 g, 4.6 mmol, 1.0 equiv) was weight into a 100 mL 2-neck round bottom flask followed by acetonitrile (10 mL) and 2,2,2-trifluoroacetic acid (1.0 mL, ′NHBn 13.8 mmol, 3.0 equiv). BH₃·THF (1.0 M, 13.8 mL, 13.8 mmol, 3.0 equiv) was Bn-1a added and the mixture was stirred at room temperature until no gas evolution was

observed. The reaction was heated to 50 °C for 6 h. The reaction was quenched via the dropwise addition of methanol (1.7 mL, 41.5 mmol, 9.0 equiv) and then stirred for 45 min before allowing the reaction to cool to room temperature (95% assay yield determined by UPLC). The reaction was concentrated to dryness and the residue was dissolved in ethyl acetate (10 mL). The organic layer was washed with 20 wt% aqueous Na_2CO_3 solution (15 mL) and H_2O (10 mL). The aqueous phases were back extracted with ethyl acetate (10 mL), the combined organic layers were dried over MgSO₄ and concentrated under vacuum. The crude oil was purified by column chromatography (dichloromethane/methanol 100/1-9/1) yielding Bn-1a as a colorless oil (824 mg, 3.7 mmol, 81% yield, >20:1 d.r.).

¹**H NMR (500 MHz, CD₂Cl₂)** δ 7.32 – 7.29 (m, 4H), 7.26 - 7.22 (m, 1H), 4.06 (ddd, J = 10.7 Hz, J = 4.5 Hz, *J* = 2.3 Hz, 1H), 3.82 (d, *J* = 13.1 Hz, 1H), 3.77 (d, *J* = 13.1 Hz, 1H), 3.53 (dd, *J* = 11.4 Hz, *J* = 3.2 Hz, 1H), 3.43 (dd, J = 11.4 Hz, J = 7.1 Hz, 1H), 3.37 – 3.32 (m, 1H), 3.06 (t, J = 10.7 Hz, 1H), 2.64 (tt, J = 10.5 Hz, J = 4.2 Hz, 1H, 2.10 - 2.06 (m, 1H), 1.62 - 1.54 (m, 1H), 1.44 - 1.21 (m, 2H) ppm. ¹³C NMR (126 MHz, CD₂Cl₂) δ 141.50 (C_g), 128.87 (CH), 128.53 (CH), 127.41 (CH), 78.56 (CH), 72.97 (CH₂), 66.32 (CH₂), 53.85 (CH), 51.66 (CH₂), 31.20 (CH₂), 27.24 (CH₂) ppm. LCMS m/z calc. for [C₁₃H₂₀NO₂] ([M+H]⁺): 222.15, found: 222.2.



((2S,5R)-5-(benzylamino)tetrahydro-2H-pyran-2-yl)methanol methylbenzenesulfonate (Bn-1a TsOH). A 40 mL vial equipped with a stir bar

was charged with Bn-3a·TsOH (1.89 g, 4.49 mmol, 1.00 equiv), sulfolane (4.0 mL), anisole (6.0 mL) and triethylsilane (3.6 mL, 22.5 mmol, 5.0 equiv). BF₃·OEt₂ (1.1 mL, 9.0 mmol, 2.0 equiv) was added and the reaction was heated to

50 °C for 18 h. Methanol (4 mL) was added carefully, and the mixture was heated to 50 °C for 2 h. The mixture was then allowed to cool to room temperature overnight. The resulting slurry was filtered and the solid was washed with 2-MeTHF (4 mL, 6 mL, and 2 mL) and dried under vacuum to yield Bn-1a·TsOH as a white solid (943 mg, 2.40 mmol, 53% yield, >20:1 d.r.).

¹H NMR (500 MHz, Methanol- d_4) δ 7.70 (d, J = 8.1 Hz, 2H), 7.51 - 7.46 (m, 5H), 7.23 (d, J = 8.1 Hz, 2H), 4.28 (d, J = 12.9 Hz, 1H), 4.28-4.25 (ddd, J = 10.6 Hz, J = 4.5 Hz, J = 2.4 Hz, 1H), 4.21 (d, J = 12.9 Hz, 1H), 3.55 - 3.49 (m, 2H), 3.43 - 3.39 (m, 2H), 3.31 - 3.26 (m, 1H), 2.40-2.35 (m, 1H), 2.37 (s, 3H), 1.83 (dt, J = 13.6 Hz, J = 4.7 Hz, 1H), 1.66 (qd, J = 12.4 Hz, J = 4.2 Hz, 1H), 1.51 - 1.43 (m, 1H) ppm. ¹³C NMR (126 MHz, Methanol-d₄) δ 143.60 (C_q), 141.67 (C_q), 132.60 (C_q), 130.91 (CH), 130.80 (CH), 130.39 (CH), 129.81 (CH), 126.97 (CH), 79.51 (CH), 68.18 (CH₂), 65.54 (CH₂), 54.70 (CH), 50.01(CH₂), 27.22 (CH₂), 27.13 (CH₂), 21.29 (CH₃) ppm. LCMS m/z calc. for [C₁₃H₂₀NO₂] ([M+H]⁺): 222.2, found: 222.2. HRMS m/z calc. for [C₁₃H₂₀NO₂] ([M+H]⁺): 222.1489, found: 222.1486.

4-



Figure S6. The relative stereochemistry of **Bn-1a** was assigned by rotating-frame nuclear Overhauser effect spectroscopy (ROESY) NMR in methanol- d_4 at 500 MHz. The key through-space correlations are highlighted.

HO O NHBn 100 Bn-1b for

((2S,5S)-5-(benzylamino)tetrahydro-2H-pyran-2-yl)methanol (Bn-1b). A 100 mL round bottom flask was charged with Bn-3b·TsOH (1.2 g, 2.9 mmol, 1.0 equiv) and methanol (10 mL). A 50 wt% solution of NaOH (0.15 mL, 2.9 mmol, 1.0 equiv) was added and the mixture was stirred at room temperature for 1 h. The solution was concentrated to dryness, the residue was suspended in 2-

MeTHF (10 mL), MgSO₄ was added, and the obtained slurry was filtered. The filtrate was concentrated to dryness to obtain **Bn-3b** as a crude oil (647 mg, 95.1 wt% purity, 2.8 mmol, 98% yield). **Bn-3b** (621 mg, 2.8 mmol, 1.0 equiv) was dissolved in acetonitrile (6 mL) in a 40 mL vial equipped with a stir bar. 2,2,2-trifluoroacetic acid (0.7 mL, 8.5 mmol, 3.0 equiv) was added followed by BH₃·THF (1.0 M, 8.5 mL, 3.0 equiv) and the mixture was stirred at room temperature until gas evolution subsided. The reaction was heated to 50 °C for 23 h. The reaction was quenched *via* the dropwise addition of methanol (1.0 mL, 24.7 mmol, 8.7 equiv) and then stirred for 35 minutes before allowing the reaction to cool to room temperature. The reaction was concentrated to dryness and the residue was dissolved in ethyl acetate (10 mL). The organic layer was washed with 20 wt% aqueous Na₂CO₃ solution (10 mL) and H₂O (6 mL). The aqueous phases were back extracted with ethyl acetate (3 x 8 mL), the combined organic layers were dried over MgSO₄ and concentrated under vacuum. The crude oil was purified twice by column chromatography (dichloromethane/methanol 100/0 – 9/1, then 4 vol% isopropylamine in dichloromethane) yielding **Bn-1b** as a colorless oil (447 mg, 91.1 wt% purity, 1.9 mmol, 66% yield based on **Bn-3b**, >20:1 d.r.).

¹**H NMR (500 MHz, CD₂Cl₂)** δ 7.37 – 7.35 (m, 2H), 7.33 – 7.31 (m, 2H), 7.25 – 7.22 (m, 1H), 3.96 (dt, *J* = 11.6 Hz, *J* = 2.1 Hz, 1H), 3.79 (s, 2H), 3.56 – 3.38 (m, 4H), 2.69 – 2.57 (m, 1H), 2.14 (br s, 2H), 1.99 –

1.94 (m, 1H), 1.72 – 1.53 (m, 2H), 1.37 – 1.33 (m, 1H) ppm. ¹³C NMR (126 MHz, CD₂Cl₂) δ 141.33, 128.65, 128.47, 127.14, 78.73, 70.59, 66.24, 51.22, 51.13, 27.06, 22.61 ppm. HRMS m/z calc. for [C₁₃H₂₀NO₂] ([M+H]⁺): 222.1489, found: 222.1469.



Figure S7. The relative stereochemistry of **Bn-1b** was assigned by nuclear Overhauser effect spectroscopy (NOESY) NMR in methanol- d_4 at 500 MHz. The key through-space correlations are highlighted.

3.4. Screening of Cyrene Amine Salts

General Procedure 3. A 4 mL vial equipped with a stir bar was charged with the requisite salt of 3a (664 μ mol, 1.0 equiv) and suspended in sulfolane (0.4 mL) and anisole (0.6 mL) under N₂ atmosphere. Triethylsilane (318 μ l, 1.99 mmol, 3.0 equiv) and BF₃·OEt₂ (168 μ l, 1.33 mmol, 2.0 equiv) were then charged and the vial was placed into a heating block and heated to 50 °C for 18 h before it was carefully quenched with methanol (400 μ L) and heated again to 50 °C for 2 hours. Maleic acid (664 μ mol, 1.0 equiv) was then added as an internal standard and the mixture was analyzed by quantitative ¹H NMR in methanol-*d*₄.

Scheme S4. Salt screen of 3a.



•TsOH

1a•TsOH

HO

		Solvent KF	r = 300 ppm	Solvent KF = 1500 ppm		
Entry	3a Salt	1a [%]	1a:1b d.r.	1a [%]	1a:1b d.r.	
1	None	85	>20:1	78	>20:1	
2	ТѕОН	90	>20:1	91	>20:1	
3	TfOH	64	>20:1	54	18:1	
4	HCl	81	>20:1	41	6:1	
5	HBr	68	10:1	44	4:1	

3.5. Synthesis and Characterization of 1a TsOH

((2S,5R)-5-(aminotetrahydro-2H-pyran-2-yl)methanol

4-



equiv) were then charged sequentially and the mixture was heated to 40 °C for 18 hours. The resulting homogenous solution was quenched with methanol (12 mL) at such a rate to maintain the internal temperature below 45 °C before the subsequent quenched solution was heated to 60 °C for *ca.* 2 hours. To this mixture was charged crystalline **1a**·**TsOH** (60 mg, 1 wt%) and the resulting seed bed was aged for *ca.* 1 hour at 60 °C before being cooled to 20 °C over 6 hours and then aged a further 18 hours at this temperature. The slurry was filtered, and the wet cake was washed with a solution of 9:1 v/v 2-MeTHF:methanol (2 x 12 mL). The cake was dried under vacuum with an N₂ sweep for *ca.* 18 hours to provide **1a**·**TsOH** as a white solid (4.55 g, 15.0 mmol, 75% yield, >20:1 d.r.).

¹**H NMR (500 MHz, DMSO-***d*₆) δ 7.85 (s, 3H), 7.50 – 7.49 (m, 2H), 7.14 – 7.12 (m, 2H), 4.65 (br s, 1H), 3.96 (ddd, J = 10.7 Hz, J = 4.4 Hz, J = 2.2 Hz, 1H), 3.36 (dd, J = 11.3 Hz, J = 5.9 Hz, 1H), 3.30 (dd, J = 1.2 Hz, 1H), 3.40 (dd, J = 1.2 Hz, 1H

11.3 Hz, J = 4.7 Hz, 1H), 3.24 – 3.19 (m, 1H), 3.21 (t, J = 10.7 Hz, 1H), 3.05 (s, 1H), 2.29 (s, 3H), 2.06 – 2.01 (m, 1H), 1.75 – 1.64 (m, 1H), 1.48 (qd, J = 12.6 Hz, J = 4.2 Hz, 1H), 1.34 – 1.19 (m, 1H) ppm. ¹³C **NMR (126 MHz, DMSO-***d*₆) δ 145.37 (C_q), 137.84 (C_q), 128.13 (CH), 125.46 (CH), 77.80 (CH), 67.61 (CH₂), 63.84 (CH₂), 46.18 (CH), 27.17 (CH₂), 26.06 (CH₂), 20.77 (CH₃) ppm. **HRMS** m/z calc. for [C₆H₁₄NO₂] ([M+H]⁺): 132.1019, found: 132.1017.

The relative and absolute stereochemistry was confirmed by X-ray crystallography, see Section 6.2.

4. Mechanistic Experiments

4.1. Reaction Profile for the Synthesis of 1a · TsOH by ¹H NMR Spectroscopy

General Procedure 4. A 1 mL volumetric flask was charged with **3a**·**TsOH** (65 mg, 0.216 M, 1.0 equiv) and octafluoronaphthalene (30 mg, 0.4 equiv), 0.4 mL of a 2:3 (v:v) mixture of sulfolane:anisole- d_8 was added followed by addition of Et₃SiH (106 µL, 3 equiv). Another 0.2 mL of the solvent mixture was added, followed by BF₃·OEt₂ (55 µL, 2 equiv). The mixture was diluted up to the 1 mL mark of the volumetric flask using the 2:3 (v:v) sulfolane:anisole- d_8 mixture. The solution became homogenous after agitation. A 0.6 mL aliquot from this solution was transferred into a 5 mm NMR tube. The NMR tube was capped and sealed using Parafilm M, where the cap and tube meet. The sample was brought out of the N₂-filled glovebox for NMR analysis.



Scheme S5. Acetal Opening of 3a · TsOH.

Figure S8. Following general procedure 4, temporal concentration profiles monitored by ¹H NMR spectroscopy for the reaction in Scheme S5 were recorded. [**3a**·**TsOH**]₀ = 0.216 M, BF₃·OEt₂ (2 equiv, 0.431 M), Et₃SiH (3 equiv, 0.656 M), 2:3 (v/v) sulfolane:anisole- d_8 , and 50 °C.



Figure S9. ¹⁹F{¹H} NMR spectrum of the end of the reaction mixture in Scheme S5. $[3a \cdot TsOH]_0 = 0.216 \text{ M}, BF_3 \cdot OEt_2 (2 \text{ equiv}, 0.431 \text{ M}), Et_3SiH (3 \text{ equiv}, 0.656 \text{ M}), 2:3 (v/v) sulfolane:anisole-$ *d* $₈ and 50 °C. Octafluoronaphthalene (0.49 equiv) was used as an internal standard (IS) for ¹⁹F{¹H} NMR. The singlet at ca. –176 ppm is identified as Et_3SiF by ¹⁹F-¹H NMR spectroscopy, with$ *J*_{Si-F} = 288 Hz (d) and*J* $_{Si-H} = 6.4 Hz (hept). The chemical shifts match with commercially available Et_3SiF (CAS: 358-43-0).$



4.2. Identification and Characterization of Diborane by NMR Spectroscopy

Figure S10. Following general procedure 4, temporal concentration profiles monitored by ¹¹B{¹H} NMR spectroscopy for the reaction in Scheme S5 were recorded. [**3a**·**TsOH**]₀ = 0.216 M, BF₃·OEt₂ (2 equiv, 0.431 M), Et₃SiH (3 equiv, 0.656 M), 2:3 (v/v) sulfolane:anisole- d_8 , and 50 °C. A known solution of NaBH₄ in methanol- d_4 in a capillary was used as an internal standard (IS) for ¹¹B NMR. *Note:* Quantification of boron species is semiquantitative due to a non-flat baseline caused by the probe and tube glass background signal.

All diborane NMR structure elucidation experiments have been performed on a Bruker 600 MHz instrument equipped with a 5mm Prodigy broadband cryoprobe (LN₂-cooled) and an Avance III HD console. All NMR spectra have been acquired at 3 °C of a sample generated as per general procedure 4 but without substrate **3a**·**TsOH**. Band-selective ¹H decoupling in ¹¹B spectra has been achieved using a low-power (0.001W) single-frequency continuous-wave (cw) irradiation.



Figure S11. A. ¹H spectrum without ¹¹B decoupling; resonances at 3.64 ppm (1:1:1:1 quartet with J = 132.9 Hz) and -0.73 ppm (very broad peak) are detected. **B.** ¹H{¹¹B} spectrum; both resonances at 3.64 ppm and -0.73 ppm become singlets.



Figure S12. ¹¹B NMR spectra recorded with and without ¹H decoupling. **A.** ¹¹B{¹H} spectrum; the boron resonance of interest is a singlet. **B.** ¹¹B spectrum without ¹H decoupling; the boron resonance is a tt with $J_{\text{HB}} = 132.5$ Hz and 45.4 Hz. **C.** ¹¹B spectrum with band-selective ¹H decoupling at -0.73 ppm; the boron resonance is a triplet with $J_{\text{HB}} = 132.5$ Hz. **D.** ¹¹B spectrum with band-selective ¹H decoupling at 3.64 ppm; the boron resonance is a triplet with $J_{\text{HB}} = 45.4$ Hz.

Based on the ¹H and ¹¹B peak multiplicity analysis in Figures S11 and S12, as well as the corresponding ¹H and ¹¹B chemical shifts, it was concluded that J_{HB} coupling constants and chemical shifts match those reported in reference 27 of the manuscript for the B₂H₆ diborane molecule.

4.3. Delayed Cyrene Amine Addition Experiment

420 μ L of 2:3 (v/v) sulfolane/anisole- d_8 , BF₃·OEt₂ (27.3 μ L), and Et₃SiH (52 μ L) were added to an NMR tube. The solution was heated at 50 °C and the reaction progress was monitored by NMR spectroscopy over time. Once the formation of B₂H₆ was detected by ¹¹B{¹H} NMR (with a characteristic signal around 17 ppm), **3a**·**TsOH** (46.4 mg) was immediately added. The subsequent NMR monitoring revealed immediate product formation.



Figure S13. Reaction profiles monitored by ¹H NMR spectroscopy for the reductive acetal opening of **3a·TsOH**, with a delayed addition of **3a·TsOH**. [**3a·TsOH**] = 0.216 M, [BF₃·OEt₂] = 0.431 M, [Et₃SiH] = 0.656 M, 2:3 (v/v) sulfolane:anisole- d_8 , 50 °C.

4.3. Dependence of the Induction Period on Water Content

We found that the duration of the induction period increased with increased water content in the reaction mixture (Figure S14A). With the observation of H_2 generation during the induction period, we thus concluded that residual water is quenched during the induction period *via* the release of H_2 before an increase in diborane concentration can be observed. A linear correlation between the water content and the Et₃SiF concentration at the end of the induction period further suggested that an intermediate species generated from Et₃SiH and BF₃·OEt₂ is the active species reacting with residual water.

General procedure 4 was followed by adding the corresponding additional amount of water: no additional water (460 ppm), 0.1 equiv (974 ppm water), and 0.2 equiv (1529 ppm).



Figure S14. A. Et₃SiF and B₂H₆ concentration profiles monitored by ¹¹B{¹H} and ¹⁹F{¹H} NMR spectroscopy for the reaction of BF₃·OEt₂ (0.431 M) with Et₃SiH (0.656 M) to probe the water effect (0, 0.1 and 0.2 equiv). **B.** Linear correlation between water equivalents and the concentration of Et₃SiH at the end of the induction period. Octafluoronaphthalene (0.49 equiv) was used as an internal standard for ¹⁹F{¹H} NMR.

4.4 Importance of Sulfolane for Diborane Formation

Sulfolane appears essential for the productive reaction between $BF_3 \cdot OEt_2$ and Et_3SiH to form diborane, which is required for the reductive acetal opening. Strong coordinating solvents such as THF, dioxane and DMSO were shown to suppress the benefit of sulfolane in the acetal opening reaction (Table S4).

able 54. Solvent encets of the dectal opening of call 13011 (see Scheme 55).							
Entry	Solvent	B ₂ H ₆ Formation	Et₃SiF Formation	1a Formation			
1	Anisole	No	No	No			
2	Anisole:Sulfolane	Yes	Yes	Yes			
3	DCM:Sulfolane	Yes	Yes	Yes			
4	DCM	No	No	No			
5	THF	No	No	No			
6	THF:Sulfolane	No	No	No			
7	Dioxane:Sulfolane	No	No	No			
8	DMSO	No	No	No			
9	DMSO:Sulfolane	No	No	No			

Table S4. Solvent effects of the acetal opening of 3a · TsOH (see Scheme S5).

^aExperiments were performed following general procedure 4 using the solvent systems listed.

In non-coordinating solvents such as anisole and dichloromethane, the addition of sulfolane is crucial for reactivity. The formation of B_2H_6 from $BF_3 \cdot OEt_2$ and Et_3SiH was not feasible in pure anisole and only occurred upon addition of sulfolane.

Delayed Sulfolane Addition Procedure: To a 1 mL volumetric flask was added Et₃SiH (106 μ L, 1.5 equiv), 0.4 mL of anisole- d_8 was added, followed by BF₃·OEt₂ (55 μ L, 1 equiv). The mixture was diluted up to 1 mL mark of the volumetric flask using anisole- d_8 . A 0.5 mL aliquot from this solution was transferred into a 5 mm NMR tube. The NMR tube was capped and sealed using Parafilm M, where the cap and tube meet. The sample was brought out of the N₂-filled glovebox for NMR analysis at 50 °C. No diborane formation was observed until upon the later sulfolane addition (0.1 mL).



Figure S15. Temporal concentration profiles monitored by ¹H and ¹¹⁹F{¹H} NMR spectroscopy for the reaction of BF₃.OEt₂ (0.431 M) with Et₃SiH (0.656 M) in anisole- d_8 with a late addition of sulfolane.

4.5. Reaction Kinetics – Reaction Rate Dependence on Sulfolane

The ratio between anisole and sulfolane dictates the homogeneity of the final reaction mixture. A high ratio of sulfolane seems to lead to biphasic mixtures due to the low solubility of Et₃SiH in the mixture.

Entry	Anisole (mL)	Sulfolane (mL)	Notes
1	0.2	0.8	Biphasic (low solubility of Et ₃ SiH)
2	0.4	0.6	Biphasic (low solubility of Et ₃ SiH)
3	0.6	0.4	homogeneous
4	0.8	0.2	homogeneous
5	0.9	0.1	homogeneous
6	1	0	low solubility of 3a·TsOH

Table S5. Effect of the solvent ratio on the homogeneity of the reaction mixture.

For homogeneous mixtures (anisole:sulfolane 0.6:0.4, 0.8:0.2, and 0.9:0.1; v:v), the acetal opening reaction was performed following general procedure 4 (Scheme S5). The solvent ratio was shown to significantly impact the length of the induction period and the overall conversion rate of **3a**·**TsOH**. The induction period increased significantly for lower sulfolane concentrations while the reaction rate for Et₃SiF formation as well as the conversion of **3a**·**TsOH** decreased. These results further corroborate the important role of sulfolane to initiate the formation of diborane and Et₃SiF.



Figure S16. Temporal concentration profiles monitored by ¹H and ¹⁹F{¹H} NMR spectroscopy for the reaction in Scheme S5 using different ratios of anisole- d_8 :sulfolane (0.6:0.4, 0.8:0.2, and 0.9:0.1; v:v). [**3a·TsOH**]₀ = 0.216 M, BF₃·OEt₂ (2 equiv), Et₃SiH (3 equiv), and 50 °C.

4.6. Reductive Acetal Opening with other Sulfones

General Procedure 6. A 4 mL vial equipped with a stir bar was charged with **3a** \cdot **TsOH** (200 mg, 664 µmol, 1 equiv.) and suspended in the reaction solvent (0.6 mL) under N₂ atmosphere and the respective sulfone (4.2 mmol, 6.3 equiv) was added followed by triethylsilane (0.32 mL, 1.99 mmol, 3.00 equiv) and BF₃ \cdot OEt₂ (0.17 mL, 1.33 mmol, 2.00 equiv). The reaction vials were then placed into a heating block and heated to 50 °C for 18 h before it was carefully quenched with methanol (0.4 mL) and heated again to 50 °C for 2 hours. CH₂Br₂ (46 µL, 664 µmol, 1 equiv) was then added as an internal standard and the mixture was analyzed by quantitative ¹H NMR in methanol-*d*₄.

Scheme S6. Acetal opening of 3a · TsOH with different sulfones.



Table S6. Acetal opening of **3a** · **TsOH** with different sulfones.

Entry	Sulfone	Solvent	1a [%]
1	sulfolane	anisole	93
2	^{<i>i</i>} PrSO ₂ Me	anisole	92
3	^{<i>i</i>} Pr ₂ SO ₂ ⁶	anisole	87
4	Ph ₂ SO ₂ ^a	1,2-dichloroethane	20
5	$Tol_2SO_2^a$	1,2-dichloroethane	47

^a Reactions were performed on a 100 mg (332 µmol) scale.

4.7. Reaction with BF₃·THF

When BF_3 ·THF was used instead of BF_3 ·OEt₂, the induction period was found to be about 1.7 days long. 90% conversion of **3a**·TsOH was eventually observed after heating at 50 °C for 8 days.



Figure S17. Following general procedure 4, temporal concentration profiles were monitored by ¹H NMR spectroscopy for the reaction in Scheme S5 using BF₃·THF. [**3a·TsOH**]₀ = 0.216 M, BF₃·THF (2 equiv, 0.431 M), Et₃SiH (3 equiv, 0.656 M), 2:3 (v/v) sulfolane:anisole- d_8 , and 50 °C.

⁶ Pr₂SO₂ was synthesized following a literature procedure: Balicki, R. Synth. Commun. **1999**, 29, 2235–2239.

5. Computational Studies

5.1. Computational Methods

All density functional theory (DFT) calculations were performed using Gaussian 16⁷. Geometry optimizations and frequency calculations were performed at the M06-2X/6-31+G(d,p) level of theory^{8,9}, with the SMD model¹⁰ to account for solvation effects. Normal vibrational mode analysis confirmed the optimized structures are minima or transition structures. Transition structures are verified by intrinsic reaction coordinate (IRC) calculations. Truhlar's quasiharmonic correction was used to compute molecular entropies to reduce the error caused by the breakdown of the harmonic oscillator approximation, by setting all positive frequencies that are less than 100 cm⁻¹ to 100 cm⁻¹.¹¹ M06-2X/6-311+G(d,p) single-point energies were computed on the M06-2X-optimized structures. MacroModel¹² was used to perform conformational searches with the OPLS3e force field.¹³ 3D renderings of stationary points were generated using CYLview 1.0¹⁴ and PyMOL 2.3¹⁵.

5.2. Calculated Activation Free Energy Difference for the Reduction of Cyrene[™] with BH4⁻



Figure S18. Reaction of CyreneTM with BH₄-

⁷ Gaussian 16, Revision C.01, Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Petersson, G. A.; Nakatsuji, H.; Li, X.; Caricato, M.; Marenich, A. V.; Bloino, J.; Janesko, B. G.; Gomperts, R.; Mennucci, B.; Hratchian, H. P.; Ortiz, J. V.; Izmaylov, A. F.; Sonnenberg, J. L.; Williams-Young, D.; Ding, F.; Lipparini, F.; Egidi, F.; Goings, J.; Peng, B.; Petrone, A.; Henderson, T.; Ranasinghe, D.; Zakrzewski, V. G.; Gao, J.; Rega, N.; Zheng, G.; Liang, W.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Throssell, K.; Montgomery, J. A., Jr.; Peralta, J. E.; Ogliaro, F.; Bearpark, M. J.; Heyd, J. J.; Brothers, E. N.; Kudin, K. N.; Staroverov, V. N.; Keith, T. A.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A. P.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Millam, J. M.; Klene, M.; Adamo, C.; Cammi, R.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Farkas, O.; Foresman, J. B.; Fox, D. J. Gaussian, Inc., Wallingford CT, 2016.

⁸ Zhao, Y.; Truhlar, D. G. Theor. Chem. Acc. 2008, 120, 215.

⁹ Zhao, Y.; Truhlar, D. G. Acc. Chem. Res. 2008, 41, 157.

¹⁰ Marenich, A. V.; Cramer, C. J.; Truhlar, D. G. J. Phys. Chem. B 2009, 113, 6378.

¹¹ Zhao, Y.; Truhlar, D. G. Phys. Chem. Chem. Phys. 2008, 10, 2813.

¹² Schrödinger Release 2019-3: MacroModel, Schrödinger, LLC, New York, NY, 2019.

¹³ Roos, K.; Wu, C.; Damm, W.; Reboul, M.; Stevenson, J. M.; Lu, C.; Dahlgren, M. K.; Mondal, S.; Chen, W.; Wang, L.; Abel, R.; Friesner, R. A.; Harder, E. D. *J. Chem. Theory Comput.* **2019**, 1863–1874.

¹⁴ Legault, C. Y. CYLview, version 1.0b; Université de Sherbrooke: Quebec, Canada, 2009; <u>http://www.cylview.org</u>.

¹⁵ The PyMOL Molecular Graphics System, Version 2.0 Schrödinger, LLC.

5.3. Calculated Thermodynamics for the Reaction of Cyrene[™] with Isopropylamine

Scheme S7. Calculated thermodynamics for the transamination reaction of CyreneTM.

$$\begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & & \\ & & & & \\ & & & & \\ & & & & & \\ & & & & \\ & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & & & & & \\ & &$$

5.4. Calculated Solvent-Sulfolane Displacement Equilibria



Figure S19. A. Computed enthalpies and free energies of reaction with BF₃. M06-2X/6-311+G(d,p)/SMD(anisole) // M06-2X/6-31+G(d,p)/SMD(anisole). Energies are in kcal/mol.

To understand the solvation effects of this reaction, we computed the reaction equilibria between sulfolane, and each coordinating solvent screened experimentally to BF₃ (Figure S19A). The results show that anisole is the easiest to displace by sulfolane, with a strong energy preference of 4.4 kcal/mol favoring the sulfolane-BF₃ complex. This contrasts with the DMSO equilibrium, which has a ΔG of 13.7 kcal/mol preferring coordination to DMSO. This trend is consistent with the experimental solvent screen (Table S4), where anisole and dichloromethane are the only solvents in which B₂H₆ forms in the presence of sulfolane.

Sulfo	olane			Н	-0.10003	2.22046	-0.09293
S	0.76706	0.00000	0.00000	Н	-0.44829	1.40742	1.46593
С	-0.44348	1.29841	0.37868	Н	-1.83968	0.91965	-1.23028
С	-1.77018	0.75399	-0.14979	Н	-2.61017	1.26439	0.32614
С	-1.77017	-0.75400	0.14979	Н	-2.61016	-1.26442	-0.32611
С	-0.44347	-1.29840	-0.37872	Н	-1.83962	-0.91967	1.23029
0	1.51797	0.35081	-1.21516	Н	-0.10000	-2.22046	0.09284
0	1.51791	-0.35082	1.21520	Н	-0.44830	-1.40735	-1.46598

Cartesian Coordinates

BF3-sulfolane

В	-1.87323	-0.22466	0.01353
F	-2.09313	-0.32970	1.37240
F	-1.22686	-1.35636	-0.47836
F	-2.99269	0.10897	-0.69713
0	-0.88506	0.97275	-0.19673
S	0.63689	0.85005	-0.05710
0	1.27673	2.15743	0.00049
С	1.31749	-0.18598	-1.37431
С	1.87541	-1.41917	-0.65215
С	2.26660	-0.98919	0.77299
С	1.08060	-0.20148	1.33323
Н	2.09643	0.43328	-1.82713
Н	0.52477	-0.39039	-2.09504
Н	2.73469	-1.80756	-1.20138
Н	1.10810	-2.19502	-0.60940
Н	3.16178	-0.35896	0.75528
Н	2.46938	-1.85130	1.41064
Н	0.22196	-0.83936	1.55745
Н	1.29885	0.45903	2.17471
BF ₃ -	anisole		
В	1.83807	0.04672	-0.51972
F	1.98426	1.40425	-0.35460
F	1.27690	-0.31215	-1.71220
F	2.96324	-0.66062	-0.18396
0	0.75390	-0.40948	0.58402
С	1.12130	-0.14912	1.97031
Η	0.45815	-0.74261	2.59860
Н	1.01900	0.91685	2.17797
С	-3.27517	0.18702	-0.29027

С	-2.74773	-1.10356	-0.25007	
С	-1.39893	-1.29958	0.04378	
С	-0.61616	-0.18372	0.28864	
С	-1.10693	1.11283	0.25543	
С	-2.45819	1.28980	-0.03992	
Н	-4.32513	0.33471	-0.52174	
Н	-3.38204	-1.96052	-0.45169	
Н	-0.95484	-2.28916	0.07314	
Н	-0.45150	1.95761	0.44082	
Н	-2.86824	2.29364	-0.07917	
Н	2.15435	-0.47355	2.07792	
BF ₃ -	dioxane			
В	1.66239	-0.00000	-0.03870	
F	1.88971	0.00008	1.32074	
F	2.08032	1.15114	-0.65731	
F	2.08033	-1.15121	-0.65718	
С	-2.02651	1.16276	-0.24322	
С	-0.59879	1.20681	0.25355	
С	-0.59881	-1.20683	0.25355	
С	-2.02652	-1.16275	-0.24322	
Н	-2.04188	1.19481	-1.34229	
Н	-2.56777	2.02780	0.14579	
Н	-0.54543	1.21297	1.34686	
Н	-0.04817	2.04840	-0.16487	
Н	-0.04821	-2.04843	-0.16488	
Н	-0.54545	-1.21299	1.34686	
Н	-2.04189	-1.19479	-1.34229	
Н	-2.56780	-2.02778	0.14578	
0	0.08113	-0.00001	-0.21614	
0	-2.68798	0.00001	0.22143	

BF ₃ -DMSO			
В	-1.44999	-0.03794	0.03579
F	-1.70607	-1.02137	-0.91394
F	-1.42029	1.22269	-0.57179
F	-2.32458	-0.09018	1.09882
0	-0.06316	-0.28586	0.62847
S	1.13596	-0.05767	-0.38895
С	2.32578	-1.23523	0.23915
С	1.84910	1.47931	0.19090
Н	3.26808	-1.07335	-0.29002
Н	1.93983	-2.23246	0.02185
Н	2.02119	1.40923	1.26674
Н	1.13401	2.26999	-0.04167
Н	2.44354	-1.08561	1.31406
Н	2.78239	1.64673	-0.35261
BF ₃ -	-Et ₂ O_1		
В	0.94755	-0.69851	-0.02473
F	1.82909	-0.20855	-0.95932
F	0.59725	-2.01499	-0.22712
F	1.35017	-0.46130	1.27536
0	-0.38103	0.11043	-0.25097
С	-1.50469	-0.22706	0.62189
С	-0.30031	1.51273	-0.65795
Н	-1.69027	0.63900	1.26220
Н	-1.16191	-1.05521	1.24218
Н	0.20636	1.50232	-1.62230
С	-2.70162	-0.60808	-0.21616

Н	-3.53404	-0.85920	0.44728
Н	-3.01945	0.21385	-0.86385
Н	-2.47214	-1.47988	-0.83356
С	0.42534	2.35912	0.36350
Н	-0.04325	2.29338	1.34943
Н	0.38543	3.40192	0.03573
Н	1.47311	2.06440	0.44911
Н	-1.33424	1.83189	-0.80269
BF ₃ -T	ΉF		
В	1.35424	0.00651	0.04125
F	1.29236	-0.25056	1.39986
F	1.86619	1.25208	-0.23911
F	1.94791	-1.01614	-0.66141
0	-0.12472	0.05159	-0.46938
С	-0.91196	-1.19006	-0.34465
С	-2.17851	-0.75266	0.36871
С	-0.92901	1.21749	-0.06734
Н	-0.30517	-1.91123	0.20194
Н	-1.08931	-1.53740	-1.36371
Н	-3.02242	-1.39444	0.11025
Н	-2.02995	-0.78041	1.45232
Н	-0.61721	1.49787	0.94207
Н	-0.70229	2.01323	-0.77489
С	-2.35042	0.69189	-0.10911
Н	-2.73803	0.71844	-1.13198
Н	-3.00775	1.28039	0.53311

6. X-Ray Crystallography data for 3a TsOH and 1a TsOH 6.1. Crystal Data and Structure Refinement for Compound 3a TsOH (CCDC 2215969)

A single crystal, grown from a water and 2-MeTHF solution, was selected for single crystal X-ray data analysis. The crystal was a colorless rod with dimensions of 0.15 mm x 0.09 mm x 0.09mm. Data collection was performed on a Bruker Apex II system at 293K. The unit cell was determined to be orthorhombic in space group $P2_12_12_1$. The structure contained one molecule of *para*-toluenesulfonic acid and one molecule of **3a** in the crystallographic asymmetric unit.

The absolute configuration was established by anomalous-dispersion effects in diffraction measurements on the crystal and confirmed that the stereochemistry was as shown.

Crystallographic data is summarized in Table S8. Figure S20 shows a thermal ellipsoid representation of Compound **3a**·**TsOH** with thermal ellipsoids set at the 30% probability level. Significant disorder was observed for the oxygens of *para*-toluenesulfonic acid. Coordinates, refinement details, and structure factors have been deposited with the Cambridge Crystallographic Data Centre (CCDC 2215969).



Figure S20. Thermal ellipsoid representation of Compound **3a·TsOH** with thermal ellipsoids set at the 30% probability level. Disorder in the *para*-toluenesulfonic acid was removed for clarity.

Identification code	mdp019	
Empirical formula	C13H19NO5S	
Formula weight	301.35	
Temperature	293(2) K	
Radiation	$CuK\alpha$ ($\lambda = 1.54178$)	
Crystal system	orthorhombic	
Space group	P2 ₁ 2 ₁ 2 ₁	
Unit Cell Dimensions	$a = 6.1009(3) \text{ Å} \alpha = 90^{\circ}$	
	$b = 12.5179(6) \text{ Å} \beta = 90^{\circ}$	
	$c = 19.3009(11) \text{ Å} \gamma = 90^{\circ}$	
Volume	1474.02(13) (Å ³)	
Ζ	4	
Density	1.358 (g/cm ³)	
Absorption coefficient	2.129 mm ⁻¹	
F(000)	640.0	
Crystal size	$0.15\times0.09\times0.09~mm^3$	
20 range for data collection	8.418° to 133.232°	
Index ranges	$-5 \le h \le 7, -14 \le k \le 14, -22 \le l \le 22$	
Reflections collected	9588	
Independent reflections	2602 [$R_{int} = 0.0306, R_{sigma} = 0.0291$]	
Completeness to Θ =66.616°	0.999	
Absorption correction	multi-scan	
Max. and min. transmission	0.7528 and 0.6754	
Refinement Method	Full-matrix least-squares on F ²	
Data/restraints/parameters	2602/0/211	
Goodness-of-fit on F ²	1.077	
Final R indexes [I>=2σ (I)]	$R_1 = 0.0431, wR_2 = 0.1072$	
Final R indexes [all data]	$R_1 = 0.0479, wR_2 = 0.1108$	
Flack parameter	0.064(11)	
Largest diff. peak/hole	0.24/-0.28 eÅ ⁻³	

Table S7. Crystal data and structure refinement for Compound 3a TsOH [CCDC 2215969].

6.2. Crystal Data and Structure Refinement for Compound 1a TsOH (CCDC 2215968)

A single crystal, grown from a saturated solution of acetonitrile and water, was selected for single crystal X-ray data analysis. The crystal was a colorless irregular shape with dimensions of 0.1 mm x 0.09 mm x 0.09mm. Data collection was performed on a Bruker Apex II system at 100K. The unit cell was determined to be orthorhombic in space group $P2_12_12_1$. The structure contains one molecule **1a**, one molecule of *para*-toluenesulfonic acid, and two water molecules in the crystallographic asymmetric unit.

The absolute configuration was established by anomalous-dispersion effects in diffraction measurements on the crystal and confirmed that the stereochemistry was as shown.

Crystallographic data is summarized in Table S9. Figure S21 shows a thermal ellipsoid representation of Compound **1a**·**TsOH** with thermal ellipsoids set at the 30% probability level. Coordinates, refinement details, and structure factors have been deposited with the Cambridge Crystallographic Data Centre (CCDC 2215968).



Figure S21. Thermal ellipsoid representation of Compound 1a TsOH with thermal ellipsoids set at the 30% probability level.

Identification code	mdq012
Empirical formula	C ₁₃ H ₂₅ NO ₇ S
Formula weight	339.40
Temperature	296.15 K
Radiation	$CuK\alpha$ ($\lambda = 1.54184$)
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
Unit Cell Dimensions	$a = 7.9459(9) \text{ Å} \alpha = 90^{\circ}$
	$b = 13.5525(12) \text{ Å} \beta = 90^{\circ}$
	$c = 15.5352(9) \text{ Å} \gamma = 90^{\circ}$
Volume	1672.9(3) (Å ³)
Ζ	4
Density	1.348 (g/cm ³)
Absorption coefficient	2.022 mm ⁻¹
F(000)	728.0
Crystal size	$0.1\times0.09\times0.09\ mm^3$
20 range for data collection	8.658° to 133.028°
Index ranges	$-9 \le h \le 9, -16 \le k \le 13, -18 \le l \le 9$
Reflections collected	9594
Independent reflections	2943 [$R_{int} = 0.0435$, $R_{sigma} = 0.0440$]
Completeness to Θ =66.514°	1.000
Absorption correction	multi-scan
Max. and min. transmission	0.7528 and 0.6738
Refinement Method	Full-matrix least-squares on F ²
Data/restraints/parameters	2943/0/293
Goodness-of-fit on F ²	1.068
Final R indexes [I>=2σ (I)]	$R_1 = 0.0311, wR_2 = 0.0810$
Final R indexes [all data]	$R_1 = 0.0334, wR_2 = 0.0831$
Flack parameter	0.031(12)
Largest diff. peak/hole	0.23/-0.17 eÅ ⁻³

 Table S8. Crystal data and structure refinement for 1a TsOH [CCDC 2215968].

7. NMR Spectra

¹H NMR at 500 MHz of 3a·HOTs in DMSO-d₆:



¹H NMR at 500 MHz of Boc-3b in DMSO-*d*₆:



¹H NMR at 500 MHz of 3b·HOTs in DMSO-d₆:



¹H NMR at 500 MHz of Bn-3a in CD₂Cl₂:



¹H NMR at 500 MHz of Bn-3a·HOTs in DMSO-d₆:



42

¹H NMR at 500 MHz of Bn-3b·HOTs in DMSO-d₆:



¹H NMR at 500 MHz of 3a·HOTf in DMSO-d₆:



44

¹⁹F NMR at 471 MHz of 3a·HOTf in DMSO-*d*₆:



¹H NMR at 500 MHz of 3a·HCl in DMSO-d₆:





46

¹H NMR at 500 MHz of 3a·HBr in DMSO-*d*₆:





¹H NMR at 500 MHz of 4 in CD₂Cl₂: (contains CyreneTM)

¹³C NMR at 126 MHz of 4 in CD₂Cl₂: (contains CyreneTM)



¹H NMR at 500 MHz of Bn-1a·in CD₂Cl₂:



¹³C NMR at 126 MHz of Bn-1a in CD₂Cl₂:







¹³C NMR at 126 MHz of Bn-1a·TsOH in methanol-d4:



¹H NMR at 500 MHz of Bn-1b·in CD₂Cl₂:





¹H NMR at 500 MHz of 1a · TsOH in DMSO-d₆:

