Electronic Supplementary Information

Fragment Optimization and Elaboration Strategies- The Discovery of Two Lead Series of PRMT5/MTA Inhibitors from Five Fragment Hits

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General experimental and chemical procedures

All chemicals were purchased from commercial suppliers and used as received unless otherwise indicated. Nuclear magnetic resonance (NMR) spectroscopy was performed on a Brüker Avance III 400 MHz spectrometer, where proton NMR (¹H NMR) spectra, carbon NMR (¹³C NMR) spectra and fluorine (¹⁹F NMR) were acquired at 400, 100 MHz and 376.5 MHz respectively. All spectra were recorded in deuterated dimethyl sulfoxide (DMSO-*d*₆), deuterated methanol (CD₃OD-*d*₄) or deuterated chloroform (CDCl₃) obtained from Cambridge Isotope Laboratories Inc. Chemical shifts (δ) were measured in parts per million (ppm) and referenced against the internal reference peaks. Coupling constants (*J*), when given, are reported in hertz. Multiplicities are reported using the following abbreviations: s = singlet, d = doublet, dd = doublet of doublets, t = triplet, q = quartet, m = multiplet (range of multiplet is given), br = broad signal, dt = doubletof triplets. Final Compounds were purified by reverse phase high-performance liquid chromatography (prep-HPLC) by either of the following conditions: HCl condition - Phenomenex Luna C18 75 x 30 mm x 3 µm; mobile phase: [water (0.05% HCl)-ACN]; B%: 10%-40%, 6.5 min. Basic condition - Waters Xbridge 150 x 25 mm x 5 µm; mobile phase: [water (0.05% ammonia hydroxide v/v)-acetonitrile]; B%: 3%-40%, 10 min. Ammonium bicarbonate condition - Waters Xbridge 150 x 25 mm x 5; mobile phase: water (10 mM ammonium bicarbonate) acetonitrile; B: 2%-40%, 10 min. The purity for test compounds was determined by highperformance liquid chromatography (HPLC) on a LC-20AB Shimadzu instrument. HPLC conditions were as follows: Kinetex C18 LC Column 4.6 x 50 mm, 5 µm, 10%-80% ACN (0.0375% TFA) in water (0.01875% TFA), 4 min run, flow rate 1.5 mL/min, UV detection ($\lambda =$ 220, 215, 254 nm) or XBridge C18, 2.1 x 50 mm, 5 µm, 10%-80% ACN in water buffered with 0.025% ammonia, 4 min run, flow rate 0.8 mL/min, UV detection ($\lambda = 220, 215, 254$ nm), or Kinetex EVO C18 100 x 4.6 mm, 2.6 µm, 0%-60% (or 10%-80%) ACN (0.0375% TFA) in water (0.01875% TFA) 10 min run, flow rate 1.0 mL/min, UV detection ($\lambda = 220, 254$ nm) or Eclipse plus C18 150 x 4.6 mm, 3.5 µm, 10%–80%) ACN (0.0375% TFA) in water (0.01875% TFA) 15 min run, flow rate 1.0 mL/min, UV detection ($\lambda = 220, 254$ nm). The mass spectra were obtained using liquid chromatography mass spectrometry (LCMS) on a LCMS-2020 Shimadzu instrument using electrospray ionization (ESI). LCMS conditions were as follows: Kinetex EVO C18 30 x 2.1 mm, 5 µm, 5%–95% ACN (0.0375% TFA) in water (0.01875% TFA), 1.5 min run, flow rate 1.5 mL/min, UV detection ($\lambda = 220, 254$ nm), or Kinetex EVO C18 2.1 x 30 mm, 5 μ m, 5%– 95% ACN in water buffered with 0.025% ammonia, 1.5 min run, flow rate 1.5 mL/min, UV detection ($\lambda = 220, 254$ nm). High resolution mass measurements were carried out on an Agilent 1290LC & 6530Q-TOF series with ESI. All compounds are > 95% pure by HPLC.

Example 6: 6-methyl-1*H*-pyrrolo[3,2-*b*]pyridin-5-amine.



To a solution of 6-bromo-1*H*-pyrrolo[3,2-*b*]pyridin-5-amine (150 mg, 707 µmol, 1.00 *eq*.) and methylboronic acid (212 mg, 3.54 mmol, 5.00 *eq*.) in 1,4-dioxane (2.5 mL) and water (0.5 mL) was added potassium carbonate (293 mg, 2.12 mmol, 3.00 *eq*.) and Pd(dppf)Cl₂ -DCM (58 mg, 70.7 µmol, 0.10 *eq*.). The reaction mixture was stirred at 80 °C for 16 hours under nitrogen atmosphere before being diluted with water (50 mL) and filtered. The filtrate was concentrated. The residue was purified by *prep*-HPLC (basic condition) and lyophilized to afford **6** (21.9 mg, 148 µmol, 21% yield) as a yellow solid. LCMS [M+1] ⁺: 148.3. ¹H NMR (400 MHz, DMSO-*d*₆) $\delta = 10.63$ (br s, 1H), 7.30 (s, 1H), 7.19 (t, *J* = 2.8 Hz, 1H), 6.11 (dt, *J* = 0.8, 2.4 Hz, 1H), 5.06 (s, 2H), 2.13 (s, 3H); HRMS (ESI, +ve ion) *m/z* calcd for C₈H₉N₃ 147.0796; found 147.0787; HPLC Rt 1.433 min, 99.7%.

Example 7: 6-cyclopropyl-1*H*-pyrrolo[3,2-*b*]pyridin-5-amine.



To a solution of 6-bromo-1*H*-pyrrolo[3,2-*b*]pyridin-5-amine (150 mg, 707 µmol, 1.00 *eq*.) and cyclopropylboronic acid (304 mg, 3.54 mmol, 5.00 *eq*.) in 1,4-dioxane (2.5 mL) and water (0.5 mL) was added potassium carbonate (293 mg, 2.12 mmol, 3.00 *eq*.) and Pd(dppf)Cl₂ -DCM (58 mg, 70.7 µmol, 0.10 *eq*.). The mixture was stirred at 80 °C for 16 hours under nitrogen atmosphere before being diluted with water (50 mL) and filtered. The filtrate was concentrated. The residue was purified by *prep*-HPLC (basic condition) and lyophilized to afford 7 (8.3 mg, 47.4 µmol, 7% yield) as a white solid. LCMS [M+1] ⁺: 174.1. ¹H NMR (400 MHz, DMSO-*d*₆) δ = 10.61 (br s, 1H), 7.21 (t, *J* = 2.8 Hz, 1H), 7.18 (s, 1H), 6.11 (ddd, *J* = 0.8, 2.0, 2.8 Hz, 1H), 5.20 (s, 2H), 1.78 - 1.69 (m, 1H), 0.92 - 0.85 (m, 2H), 0.54 - 0.47 (m, 2H); HRMS (ESI, +ve ion) *m/z* calcd for C₁₀H₁₁N₃ 173.0953; found 173.0941; HPLC Rt 1.319 min, 99.0%.

Example 8: 6-(trifluoromethyl) -1H-pyrrolo[3,2-b]pyridin-5-amine.



Step 1: To a mixture of 2-methoxy-6-methyl-5-nitro-3-(trifluoromethyl)pyridine (prepared following the method from WO2018215316) (500 mg, 2.12 mmol, 1.00 *eq*.) in acetonitrile (10 mL) was added chlorotrimethylsilane (1.15 g, 10.6 mmol, 1.34 mL, 5.00 *eq*.) and sodium iodide (1.59 g, 10.6 mmol, 5.00 *eq*.). The reaction mixture was stirred at 70 °C for 2 hours. The mixture was concentrated and the residue was purified by reversed-phase HPLC (formic acid condition) to afford 6-methyl-5-nitro-3-(trifluoromethyl)pyridin-2-ol (290 mg, 1.26 mmol, 60% yield) as a black solid. LCMS [M+1] +: 223.0. ¹H NMR (400 MHz, DMSO-*d*₆) δ = 13.23 (s, 1H), 8.46 (s, 1H), 2.70 (s, 3H).

Step 2: To a mixture of 6-methyl-5-nitro-3-(trifluoromethyl)pyridin-2-ol (250 mg, 1.13 mmol, 1.00 *eq.*) in acetonitrile (6 mL) was added DBU (343 mg, 2.25 mmol, 339 µL, 2.00 *eq.*), (4-methoxyphenyl)methanamine (463 mg, 3.38 mmol, 437 µL, 3.00 *eq.*) and BOP (647 mg, 1.46 mmol, 1.30 *eq.*). The mixture was stirred at 30 °C for 2 hours before being concentrated and purified by flash silica gel chromatography (Ethyl acetate / Petroleum ether 0-20%) to afford *N*-[(4-methoxyphenyl)methyl]-6-methyl-5-nitro-3-(trifluoromethyl)pyridin-2-amine (280 mg, 818 µmol, 73% yield) as a yellow oil. LCMS [M+1] +: 341.9. ¹H NMR (400 MHz, CDCl₃) δ = 8.48 (s, 1H), 7.26 (s, 2H), 6.96 - 6.84 (m, 2H), 5.73 (s, 1H), 4.75 (d, *J* = 5.2 Hz, 2H), 3.81 (s, 3H), 2.85 (s, 3H).

Step 3: To a solution of *N*-[(4-methoxyphenyl)methyl]-6-methyl-5-nitro-3-(trifluoromethyl)pyridin 2 aming (250 mg, 733 µmol, 1,00 gg) in DMF (4,00 mJ

(trifluoromethyl)pyridin-2-amine (250 mg, 733 μ mol, 1.00 eq.) in DMF (4.00 mL) was added *N*,*N*-dimethyl formamide dimethyl acetal (436 mg, 3.66 mmol, 486 μ L, 5.00 eq.). The mixture was stirred at 90 °C for 3 hours. The reaction mixture was poured into brine (50 mL), extracted with ethyl acetate (10 mL × 3). The combined organic layers were washed with water (30 mL), dried over anhydrous sodium sulfate, filtered and concentrated to afford 6-[(*E*)-2-(dimethylamino)vinyl]-*N*-[(4-methoxyphenyl)methyl]-5-nitro-3-(trifluoromethyl)pyridin-2-

amine (290 mg, 702 µmol, 96% yield) as a brown solid. LCMS [M+1] ⁺: 397.2. ¹H NMR (400 MHz, CDCl₃) δ = 8.45 (s, 1H), 8.00 (d, J = 2.4 Hz, 1H), 7.24 (d, J = 8.8 Hz, 2H), 6.93 - 6.82 (m, 2H), 6.47 (d, J = 12.0 Hz, 1H), 5.52 (s, 1H), 4.67 (d, J = 5.2 Hz, 2H), 3.79 (s, 3H), 3.03 (d, J = 6.4 Hz, 6H).

Step 4: A suspension of Fe (42.3 mg, 757 μ mol, 6.00 *eq.*) in acetic acid (1.0 mL) was stirred at 25 °C for 0.5 hour and then 6-[(*E*)-2-(dimethylamino)vinyl]-*N*-[(4-methoxyphenyl)methyl]-5-nitro-3-(trifluoromethyl)pyridin-2-amine (50.0 mg, 126 μ mol, 1.00 *eq.*) was added. The reaction mixture was stirred at 25 °C for 1 hour, filtered and concentrated to afford *N*-[(4-methoxyphenyl)methyl]-6-(trifluoromethyl)-1*H*-pyrrolo[3,2-b]pyridin-5-amine (30 mg, crude, 67% pure by LCMS) as a brown solid. LCMS [M+1] +: 322.1.

Step 5: To a mixture of *N*-[(4-methoxyphenyl)methyl]-6-(trifluoromethyl)-1*H*-pyrrolo[3,2b]pyridin-5-amine (30.0 mg, crude, 67% pure by LCMS) in dichloromethane (3.0 mL) was added trifluoroacetic acid (1.0 mL). The mixture was stirred at 25 °C for 16.5 hours. The mixture was basified to pH 7 with ammonium hydroxide and concentrated. The crude material was purified by *prep*-HPLC (basic condition) to afford **8** (2.5 mg, 12.0 µmol, 10% yield over 2 steps) as an off-white solid. LCMS [M+1] ⁺: 202.1. ¹H NMR (400 MHz, CD₃OD-*d*₄) δ = 7.89 (s, 1H), 7.54 (d, *J* = 3.2 Hz, 1H), 6.32 (dd, *J* = 0.8, 3.2 Hz, 1H); ¹⁹F NMR (376.5 MHz, CD₃OD-*d*₄) δ -64.4 (s, 1F); HPLC Rt 1.323 min, 95.1%.

Example 9: *N*-[(5-amino-6-bromo-1*H*-pyrrolo[3,2-*b*]pyridin-2-yl)methyl]pyrimidine-4-carboxamide.



Step 1: To a solution of 3-bromo-2-chloro-6-methyl-5-nitro-pyridine (19.0 g, 75.5 mmol, 1.00 *eq.*) and 1-(4-methoxyphenyl)-*N*-[(4-methoxyphenyl)methyl]methanamine (23.3 g, 90.6 mmol, 1.20 *eq.*) in THF (190 mL) was added sodium carbonate (9.61 g, 90.6 mmol, 1.20 *eq.*). The mixture was stirred at 75 °C for 16 h. The mixture was concentrated, diluted with ethyl acetate (500 mL), washed with water (200 mL), dried over sodium sulfate, filtered, and concentrated. The residue was purified by silica gel chromatography (Ethyl acetate / Petroleum ether 0-50 %) to afford 3-bromo-*N*,*N*-bis[(4-methoxyphenyl)methyl]-6-methyl-5-nitro-pyridin-2-amine (33.0 g, 68.4 mmol, 91% yield) as a yellow solid. LCMS [M+1] +: 472.0. ¹HNMR (400 MHz, CDCl₃) δ = 8.53 (s, 1H), 7.20 (d, *J* = 8.8 Hz, 4H), 6.94 - 6.80 (m, 4H), 4.75 (s, 4H), 3.82 (s, 6H), 2.76 (s, 3H).

Step 2: To a mixture of 3-bromo-*N*,*N*-bis[(4-methoxyphenyl)methyl]-6-methyl-5-nitro-pyridin-2-amine (10.0 g, 21.1 mmol, 1.00 *eq*.) and diethyl oxalate (9.20 g, 63.5 mmol, 8.60 mL, 3.00 *eq*.) was added DABCO (3.87 g, 25.4 mmol, 3.83 mL, 1.20 *eq*.). The mixture was stirred at 30 °C for 16 h. The mixture was diluted with ethyl acetate (500 mL). Acetic acid (4.0 mL) was added. The resulting solution was washed with water (300 mL), dried over sodium sulfate, filtered, and concentrated. The residue was purified by silica gel chromatography (Ethyl acetate / Petroleum ether 0-50 %) to afford ethyl (*Z*)-3-[6-[bis](4-methoxyphenyl)methyl]amino]-5-bromo-3-nitro-2-pyridyl]-2-hydroxy-prop-2-enoate (7.20 g, 10.6 mmol, 51% yield, 85% purity by LCMS) as a yellow solid. LCMS [M+1] ⁺: 572.2. ¹H NMR (400 MHz, CDCl₃) δ = 8.70 - 8.63 (m, 1H), 7.53 (s, 1H), 7.07 - 6.98 (m, 4H), 6.88 - 6.83 (m, 4H), 4.64 - 4.54 (m, 4H), 4.38 (q, *J* = 7.2 Hz, 2H), 3.81 (s, 6H), 1.43 - 1.30 (m, 3H).

Step 3: To a solution of ethyl (*Z*)-3-[6-[bis](4-methoxyphenyl)methyl]amino]-5-bromo-3-nitro-2-pyridyl]-2-hydroxy-prop-2-enoate (5.10 g, 8.91 mmol, 1.00 *eq*.) in THF (15 mL), ethanol (90 mL) and water (10 mL) was added ammonium chloride (571 mg, 10.6 mmol, 1.20 *eq*) followed by iron powder (1.99 g, 35.6 mmol, 4.00 *eq*.) at 25 °C. The mixture was then stirred at 60 °C for 6 hours. After such time the mixture was diluted with dichloromethane (600 mL) and water (600 mL), stirred for 10 min, filtered and the organic layer was separated, dried over sodium sulfate, filtered and concentrated. The residue was purified by silica gel chromatography (Ethyl acetate / Petroleum ether 0-80 %) to afford ethyl 5-[bis [(4-methoxyphenyl) methyl] amino]-6-bromo-1*H*pyrrolo [3, 2-*b*] pyridine-2-carboxylate (2.10 g, 3.88 mmol, 44% yield) as a yellow solid. LCMS [M+1] ⁺: 526.1. ¹H NMR (400 MHz, CDCl₃) *δ* = 8.92 (brs, 1H), 7.94 (s, 1H), 7.32 (d, *J* = 8.4 Hz, 4H), 7.20 (d, *J* = 1.2 Hz, 1H), 6.89 - 6.78 (m, 4H), 4.44 (q, *J* = 7.2 Hz, 2H), 4.36 (s, 4H), 3.78 (s, 6H), 1.43 (t, *J* = 7.2 Hz, 3H).

Step 4: To a solution of ethyl 5-[bis[(4-methoxyphenyl)methyl]amino]-6-bromo-1*H*-pyrrolo[3,2*b*]pyridine-2-carboxylate (900 mg, 1.72 mmol, 1.00 *eq*.) in THF (20 mL) was added lithium aluminum hydride (195 mg, 5.15 mmol, 3.00 *eq*.) at 0 °C. The mixture was stirred at 25 °C for 0.5 hour. The mixture was diluted with tetrahydrofuran (100 mL). Sodium sulfate decahydrate (5.0 g) was added. The resulting solution was stirred for 0.5 hour. The resulting mixture was filtered and the filtrate concentrated to afford [5-[bis](4-methoxyphenyl)methyl]amino]-6bromo-1*H*-pyrrolo[3,2-*b*]pyridin-2-yl]methanol (827 mg, crude, 76% purity by LCMS) as a brown solid. LCMS [M+1] ⁺: 484.2. ¹HNMR (400 MHz, CDCl₃) δ = 8.53 (br s, 1H), 7.66 - 7.58 (m, 1H), 7.22 (br d, *J* = 8.4 Hz, 4H), 6.91 (s, 1H), 6.71(d, *J* = 8.4 Hz, 4H), 4.62 (s, 2H), 4.20 (s, 4H), 3.66 (s, 6H).

Step 5: To a solution of [5-[bis[(4-methoxyphenyl)methyl]amino]-6-bromo-1*H*-pyrrolo[3,2*b*]pyridin-2-yl]methanol (827 mg, crude, 76% purity by LCMS) in dichloromethane (20 mL) was added thionyl chloride (1.02 g, 8.57 mmol, 621 μ L) and DMF (125 mg, 1.71 mmol, 131 μ L) at 0 °C. The mixture was stirred at 25 °C for 0.5 hour. The mixture was concentrated to afford 6bromo-2-(chloromethyl)-*N*-[(4-methoxyphenyl)methyl]-1*H*-pyrrolo [3,2-*b*] pyridin-5-amine (652 mg, crude) as a black solid. Which was used directly in the next step.

Step 6: Ammonia gas was passed through ethanol (20 mL) at 0 °C for 10 min. 6-bromo-2-(chloromethyl)-*N*-[(4-methoxyphenyl)methyl]-1*H*-pyrrolo[3,2-*b*]pyridin-5-amine (652 mg, crude) in methanol (15 mL) was added to the ammonia solution. The mixture was stirred at 25 °C for 16 hours. The mixture was concentrated, and the residue was purified by silica gel chromatography (methanol / dichloromethane 0-50%, 5% ammonium hydroxide) to afford 2- (aminomethyl)-6-bromo-*N*-[(4-methoxyphenyl) methyl]-1*H*-pyrrolo [3, 2-*b*] pyridin-5-amine, (75 mg, 159 µmol, 9% yield over 3 steps) as a brown solid. LCMS [M+1] +: 363.1.

Step 7: To a solution of 2-(aminomethyl)-6-bromo-*N*-[(4-methoxyphenyl) methyl]-1*H*-pyrrolo
[3, 2-*b*] pyridin-5-amine (70 mg, 193 μmol, 1.00 *eq*.) and pyrimidine-4-carboxylic acid (36 mg, 290 μmol, 1.50 *eq*.) in dichloromethane (4.0 mL) was added HATU (110 mg, 290 μmol, 1.50

eq.) and diisopropylethylamine (581 µmol, 101 µL, 3.00 *eq.*) at 25 °C. The mixture was stirred at 25 °C for 1h. The mixture was concentrated and the residue purified by silica gel chromatography (Petroleum ether / Ethyl acetate 0-100 %) to afford *N*-[[6-bromo-5-[(4-methoxyphenyl)methylamino]-1*H*-pyrrolo[3,2-*b*]pyridin-2-yl]methyl]pyrimidine-4-carboxamide (40 mg, 77.0 µmol, 40% yield, 90% purity by LCMS) as brown solid. LCMS [M+1] +: 467.1. To a solution of *N*-[[6-bromo-5-[(4-methoxyphenyl)methylamino]-1*H*-pyrrolo[3,2-*b*]pyridin-2-yl]methyl]pyrimidine-4-carboxamide (61.4 µmol, 1.00 *eq.*) in dichloromethane (2.5 mL) was added trifluoroacetic acid (0.5 mL) at 10 °C. The mixture was stirred at 30 °C for 6 hrs. The mixture was basified to pH 8 by addition of saturated sodium bicarbonate, extracted with dichloromethane (100 mL), dried over sodium sulfate, filtered, and concentrated. The residue was purified by silica gel chromatography (dichloromethane/ methanol 0-10%) to afford **9** (8.9 mg, 24 µmol, 12% yield over 3 steps) as an off-white solid. LCMS [M+1] +: 346.9; ¹H NMR (400 MHz, CD₃OD-*d*₄) δ = 9.30 (d, J = 1.2 Hz, 1H), 9.05 (d, J = 5.2 Hz, 1H), 8.32 (s, 1H), 8.12 (dd, J = 1.2, 5.2 Hz, 1H), 6.43 (s, 1H), 4.81 - 4.77 (m, 2H); HPLC Rt 1.376 min, 95.7%.

Example 13: 2-amino-1-methyl-1*H*-benzo[*d*]imidazole-7-carbonitrile hydrochloride.



Step 1: To a solution of 2-chloro-3-nitro-benzonitrile (250 mg, 1.37 mmol, 1.0 *eq.*) in ethanol (2.5 mL) was added methylamine in ethanol (42.5 mg, 1.37 mmol, 2.5 M, 10.0 *eq.*). The reaction mixture was stirred at 15 °C for 12 hours then concentrated. The residue was diluted with water (50 mL) and extracted with ethyl acetate (3 x 60 mL). The combined organic extracts were dried over sodium sulfate, filtered, and concentrated to produce 2-(methylamino)-3-nitro-benzonitrile (240 mg, 1.35 mmol, 99% yield) as a yellow solid. ¹H NMR (400 MHz, CDCl₃) δ = 8.51 (s, 1H), 8.39 (d, *J* = 8.8 Hz, 1H), 7.76 (d, *J* = 7.6 Hz, 1H), 6.72 (t, *J* = 8.0 Hz, 1H), 3.49 (d, *J* = 5.2 Hz, 3H).

Step 2: To a solution of 2-(methylamino)-3-nitro-benzonitrile (240 mg, 1.35 mmol, 1.00 eq.) in water (3.0 mL) was added iron powder (378 mg, 6.77 mmol, 5.00 eq.) and hydrochloric acid (6.00 M, 903 μ L, 4.00 eq.). The reaction mixture was stirred at 15 °C for 12 hours and after such

time the mixture was concentrated. The residue was diluted with water (50 mL) and extracted with ethyl acetate (2 x 100 mL). The combined organic extracts were dried over sodium sulfate, filtered and concentrated to give 3-amino-2-(methylamino)benzonitrile (140 mg, 0.95 mmol, 70% yield) as a black solid. ¹H NMR (400 MHz, CDCl₃) δ = 6.99 (dd, *J* = 1.6, 7.6 Hz, 1H), 6.91 - 6.85 (m, 1H), 6.84 - 6.77 (m, 1H), 4.06 - 3.42 (m, 2H), 3.32 - 3.08 (m, 1H), 3.01 (s, 3H).

Step 3: To a solution of 3-amino-2-(methylamino)benzonitrile (140 mg, 0.95 mmol, 1.00 eq.) in ethanol (3.0 mL) was added cyanogen bromide (202 mg, 1.90 mmol, 2.00 eq.). The reaction mixture was stirred at 15 °C for 2 hours. The mixture was then concentrated, and the residue diluted with water (50 mL) and extracted with ethyl acetate (100 x 2 mL). The combined organic extracts were dried over sodium sulfate, filtered, and concentrated. The residue was purified by *prep*-HPLC (HCl condition) to afford **13** (30 mg, 171 µmol, 18% yield) as a white solid. LCMS $[M+1]^+$: 173.0. ¹H NMR (400 MHz, DMSO-*d*₆) δ = 13.26 (s, 1H), 9.07 (s, 2H), 7.78 - 7.60 (m, 2H), 7.39 (t, *J* = 8.0 Hz, 1H), 3.85 (s, 3H); HPLC Rt 1.283 min, 99.4%.

Compound 19: 5-bromo-2-(2,5-dimethyl-1*H*-pyrrol-1-yl)-1-methyl-1*H*-benzo[*d*]imidazole-7-carbonitrile.



Step 1: To a solution of 5-bromo-2-(methylamino)benzonitrile (15.0 g, 71.1 mmol, 1.00 *eq.*) in acetonitrile (300 mL) was added nitronium tetrafluoroborate (8.78 mL, 85.3 mmol, 1.20 *eq.*) at 0 °C. The reaction was stirred at 20 °C for 14 hours. The reaction mixture was diluted with water (100 mL) and stirred for 5 min. The aqueous phase was extracted with ethyl acetate (150 mL × 3). The combined organic phase was dried with anhydrous sodium sulfate, filtered, and concentrated in vacuum to give a residue. The residue was purified by column chromatography (SiO₂, ethyl acetate in petroleum ether/ 1-33%) to produce 5-bromo-2-(methylamino)-3-nitrobenzonitrile (6.00 g, 23.4 mmol, 33% yield) as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ = 8.90 - 8.88 (m, 1H), 8.87 - 8.85 (m, 1H), 3.71 (s, 3H).

Step 2: To a solution of 5-bromo-2-(methylamino)-3-nitrobenzonitrile (5.00 g, 19.5 mmol, 1.00 *eq.*) in ethyl acetate (50 mL) and water (1.5 mL) was added acetic acid (15 mL) and iron powder

(10.9 g, 195 mmol, 10.0 eq.). The reaction was stirred at 60 °C for 1 hour. The reaction mixture was diluted with water (100 mL) and stirred for 5 min. The aqueous phase was extracted with ethyl acetate (200 mL \times 3). The combined organic phase was dried over anhydrous sodium sulfate, filtered and concentrated in vacuum to afford 3-amino-5-bromo-2-

(methylamino)benzonitrile (4.10 g, 18.1 mmol, 93% yield) as a black brown gum. LCMS [M+1] +: 227.9. ¹H NMR (400 MHz, DMSO- d_6) δ = 6.84 (m, 2H), 5.31 (br s, 2H), 4.48 (m, 1H), 3.06 (br d, J = 4.8 Hz, 3H).

Step 3: To a solution of 3-amino-5-bromo-2-(methylamino)benzonitrile (4.10 g, 18.1 mmol, 1.00 *eq.*) in ethyl alcohol (45 mL) was added cyanogen bromide (3.84 g, 36.3 mmol, 2.67 mL, 2.00 *eq.*). The reaction was stirred at 20 °C for 2 hours. The reaction mixture was then diluted with water (50 mL) and stirred for 5 min. The aqueous phase was extracted with ethyl acetate (40 mL \times 3). The combined organic phase was dried over anhydrous sodium sulfate, filtered, and concentrated in vacuum to give a residue. The residue was purified by column chromatography (SiO₂, ethyl acetate in petroleum ether 30-50%, then methanol in ethyl acetate 10%) to give 2-amino-6-bromo-3-methyl-benzimidazole-4-carbonitrile (1.80 g, 6.77 mmol, 37% yield) as a dark brown solid. LCMS [M+1]⁺: 252.8. ¹H NMR (400 MHz, DMSO-*d*₆) δ = 7.54 (d, *J* = 1.6 Hz, 1H), 7.44 (d, *J* = 2.0 Hz, 1H), 7.10 (s, 2H), 3.74 (s, 3H).

Step 4: To a solution of 2-amino-6-bromo-3-methyl-benzimidazole-4-carbonitrile (300 mg, 1.19 mmol, 1.00 *eq.*) in toluene (18 mL) and *N*,*N*-dimethylformamide (6 mL) was added 4methylbenzenesulfonic acid hydrate (13.6 mg, 71.7 µmol, 0.06 *eq.*) and hexane-2,5-dione (682 mg, 5.97 mmol, 701 µL, 5.00 *eq.*) at 10 °C. The mixture was stirred at 140 °C for 16 hours fitted with a Dean-Stark trap. The mixture was poured into water (30 mL) and extracted with ethyl acetate (15 mL × 3). The combined organic layers were washed with water (10 mL) and brine (10 mL), dried over anhydrous sodium sulfate, filtered, and concentrated in vacuum. The residue was purified by silica gel chromatography (ethyl acetate in petroleum ether 20%) to afford **19** (150 mg, 454 µmol, 38% yield) as a white solid. LCMS [M+1]⁺: 329.0/331.0. ¹H NMR (400 MHz, CDCl₃) δ =8.35 (d, *J* = 2.0 Hz, 1H), 8.13 (d, *J* = 2.0 Hz, 1H), 5.96 (s, 2H), 3.66 (s, 3H), 1.98 (s, 6H).

Compound 21: 2-(2,5-dimethylpyrrol-1-yl)-3-methyl-6-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)benzimidazole-4-carbonitrile.



To a solution of **19** (600 mg, 1.82 mmol, 1.00 *eq.*) and 4,4,5,5-tetramethyl-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1,3,2-dioxaborolane (1.39 g, 5.47 mmol, 3.00 *eq.*) in 1,4-dioxane (9 mL) was added potassium acetate (537 mg, 5.47 mmol, 3.00 *eq.*), followed by [1,1'-bis(diphenylphosphino)ferrocene]dichloropalladium(II) (200 mg, 273 µmol, 0.15 *eq.*) under a nitrogen atmosphere. The mixture was stirred at 80 °C for 16 hours. The mixture was poured into water (20 ml) and extracted with ethyl acetate (20 mL × 3). The combined organic layers were dried over anhydrous sodium sulfate, filtered, and concentrated in vacuum. The residue was purified by silica gel chromatography (ethyl acetate in petroleum ether 25%) to afford **21** (710 mg, crude) as a white solid that was used in the next steps without further purification. LCMS [M+1]⁺: 377.2/295.1. ¹H NMR (400 MHz, CDCl₃) δ =8.40 (s, 1H), 8.06 (s, 1H), 5.90 (s, 2H), 3.67 (s, 3H), 1.97 (s, 6H), 1.31 (s, 12H).

Compounds 28-31



Step 1: To a solution of 2,5-dibromopyridin-3-amine (25.0 g, 99.2 mmol, 1.00 *eq*.) in pyridine (250 mL) was added 2,2-dimethylpropanoyl chloride (18.0 g, 149 mmol, 18.3 mL, 1.50 *eq*.) at 0

°C. The mixture was stirred at 20 °C for 0.5 hr. The reaction mixture was diluted with water (200 mL) and extracted with ethyl acetate (200 mL × 3). The combined organic phases were washed with brine (100 mL), dried over sodium sulfate, filtered, and concentrated to give a residue. The residue was purified by column chromatography (SiO₂, Ethyl acetate / Petroleum ether 2-5%) to give *N*-(2,5-dibromo-3-pyridyl)-2,2-dimethyl-propanamide (33.0 g, 98.2 mmol, 99% yield) as a yellow solid. ¹H NMR (400 MHz, CDCl₃) δ = 8.95 (d, *J* = 2.4 Hz, 1H), 8.14 (d, *J* = 2.4 Hz, 1H), 8.01 (br s, 1H), 1.35 (s, 9H).

Step 2: A mixture of N-(2,5-dibromo-3-pyridyl)-2,2-dimethyl-propanamide (33.0 g, 98.2 mmol, 1.00 eq.), tri-butyl(1-ethoxyvinyl)stannane (28.4 g, 78.6 mmol, 26.5 mL, 0.80 eq.) and Pd(PPh₃)₄ (11.4 g, 9.82 mmol, 0.10 eq.) in toluene (1.32 L) was degassed and stirred at 80 °C for 12 hours under nitrogen atmosphere. The reaction mixture was concentrated under reduced pressure to give a residue. The residue was purified by column chromatography (SiO₂, Ethyl acetate / Petroleum ether 2-5%) to give N-[5-bromo-2-(1-ethoxyvinyl)-3-pyridyl]-2,2-dimethylpropanamide (16.0 g, 47.6 mmol, 49% yield) as a white solid. ¹H NMR (400 MHz, CDCl₃) $\delta =$ 9.19 (br s, 1H), 9.03 - 8.99 (m, 1H), 8.34 - 8.27 (m, 1H), 5.06 (dd, J = 2.8, 5.2 Hz, 1H), 4.63 -4.55 (m, 1H), 4.13 - 4.00 (m, 2H), 1.52 - 1.44 (m, 3H), 1.32 - 1.27 (m, 9H). A mixture of N-[5bromo-2-(1-ethoxyvinyl)-3-pyridyl]-2,2-dimethyl-propanamide (9.50 g, 29.0 mmol, 1.00 eq.) in hydrochloric acid/dioxane (4 M, 38.0 mL, 5.24 eq.) was stirred at 20 °C for 10 minutes. The reaction mixture was concentrated under reduced pressure to give a residue. The residue was diluted with water (50 mL) and extracted with ethyl acetate (50 mL \times 3). The combined organic phases were washed with saturated sodium bicarbonate aqueous solution (50 mL \times 2), brine (50 mL), dried over sodium sulfate, filtered, and concentrated under reduced pressure to give a residue. The residue was purified by column chromatography (SiO₂, Ethyl acetate / Petroleum ether 2-5%) to give N-(2-acetyl-5-bromo-3-pyridyl)-2,2-dimethyl-propanamide (6.90 g, 17.4 mmol, 60% yield) as a white solid. ¹H NMR (400 MHz, CDCl₃) $\delta = 11.82$ (br s, 1H), 9.44 (d, J =2.0 Hz, 1H), 8.39 (d, J = 2.0 Hz, 1H), 2.77 (s, 3H), 1.36 (s, 9H).

Step 3: To a solution of acetonitrile (1.84 g, 44.9 mmol, 2.36 mL, 2.10 *eq.*) in THF (45 mL) was added lithium diisopropylamide (2 M, 22.5 mL, 2.10 *eq.*) in a dropwise fashion at -78 °C. After stirring for 0.5 hour, a solution of *N*-(2-acetyl-5-bromo-3-pyridyl)-2,2-dimethyl-propanamide (6.40 g, 21.4 mmol, 1.00 *eq.*) in THF (20 mL) was added to the reaction mixture. The reaction

mixture was stirred at -78 °C for 30 minutes. The reaction mixture was quenched with water (50 mL) and extracted with dichloromethane (50 mL × 3). The combined organic extracts were washed with brine (50 mL), dried over sodium sulfate, filtered, and concentrated to give *N*-[5-bromo-2-(2-cyano-1-hydroxy-1-methyl-ethyl)-3-pyridyl]-2,2-dimethyl-propanamide (8.10 g, crude) as a brown oil, which was used in the next step directly without further purification. ¹H NMR (400 MHz, CDCl₃) δ = 10.11 (br s, 1H), 8.98 (d, *J* = 2.0 Hz, 1H), 8.23 (d, *J* = 2.0 Hz, 1H), 4.46 (br s, 1H), 3.31 - 3.21 (d, *J* = 16.4 Hz, 1H), 3.17 - 3.06 (d, *J* = 16.4 Hz, 1H), 1.68 (s, 3H), 1.30 (s, 9H).

Step 4: A solution of *N*-[5-bromo-2-(2-cyano-1-hydroxy-1-methyl-ethyl)-3-pyridyl]-2,2dimethyl-propanamide (0.95 g, crude) in hydrochloric acid (3 M, 3.80 mL) was heated at 160 °C for 5 minutes in a microwave. The resulting mixture was basified to pH 9 with saturated sodium bicarbonate (10 mL). A precipitate formed and the precipitate was filtered and washed with water to give 7-bromo-4-methyl-1*H*-1,5-naphthyridin-2-one (5.00 g, 20.9 mmol) as a brown solid. ¹H NMR (400 MHz, DMSO- d_6) δ = 11.75 (br s, 1H), 8.55 (d, *J* = 2.0 Hz, 1H), 7.82 (d, *J* = 2.0 Hz, 1H), 6.68 (d, *J* = 0.8 Hz, 1H), 2.43 (d, *J* = 0.8 Hz, 3H).

Step 5: A mixture of 7-bromo-4-methyl-1*H*-1,5-naphthyridin-2-one (2.50 g, 10.5 mmol, 1.00 *eq.*) and phosphorus oxychloride (41.3 g, 269 mmol, 25 mL) was stirred at 120 °C for 3 hours. The reaction mixture was then concentrated under reduced pressure to give a residue. The residue was diluted with ethyl acetate (50 mL) and ice water (50 mL). The aqueous phase was separated and extracted with ethyl acetate (50 mL × 3). The combined organic extracts were washed with saturated sodium bicarbonate aqueous solution (50 mL), brine (50 mL), dried over sodium sulfate, filtered, and concentrated to give 7-bromo-2-chloro-4-methyl-1,5-naphthyridine, (2.5 g, 9.71 mmol, 92% yield) as a brown solid which was used in the next step directly without further purification. ¹H NMR (400 MHz, DMSO-*d6*) δ = 9.08 (d, *J* = 2.4 Hz, 1H), 8.66 (d, *J* = 2.4 Hz, 1H), 7.79 (d, *J* = 1.2 Hz, 1H), 2.72 (d, *J* = 1.2 Hz, 3H).

Compound 28: 7-bromo-N-[(4-methoxyphenyl)methyl]-4-methyl-1,5-naphthyridin-2-amine.

Step 6: To a solution of 7-bromo-2-chloro-4-methyl-1,5-naphthyridine (0.80 g, 3.11 mmol, 1.00 *eq.*) in dimethylsulfoxide (8.0 mL) was added potassium fluoride (541 mg, 9.32 mmol, 218 μ L, 3.00 *eq.*) and (4-methoxyphenyl)methanamine (852 mg, 6.21 mmol, 804 μ L, 2 *eq.*). The mixture

was stirred at 130 °C for 2 hours. The reaction mixture was diluted with water (10 mL) and extracted with ethyl acetate (10 mL \times 3). The combined organic extracts were washed with brine (10 \times 2 mL), dried over sodium sulfate, filtered, and concentrated to give **28** (1.30 g, crude, 86% purity) as a yellow oil which was used in the next step directly without further purification. LCMS [M+1]⁺: 358.1.

Compound 29: 7-bromo-*N*,*N*-bis[(2,4-dimethoxyphenyl)methyl]-4-methyl-1,5-naphthyridin-2-amine.

Step 7: A mixture of 7-bromo-2-chloro-4-methyl-1,5-naphthyridine (1.00 g, 3.88 mmol, 1.00 *eq.*), 1-(2,4-dimethoxyphenyl)-*N*-[(2,4-dimethoxyphenyl)methyl]methanamine (2.46 g, 7.77 mmol, 2.00 *eq.*), potassium fluoride (677 mg, 11.7 mmol, 273 µL, 3.00 *eq.*) in dimethyl sulfoxide (10 mL) was degassed and stirred at 130 °C for 12 hours under nitrogen atmosphere. The mixture was diluted with brine (10 mL), extracted with ethyl acetate (20 mL × 2) and the combined organic phases were dried over sodium sulfate, filtered, concentrated in vacuo. The residue was purified by column chromatography (SiO₂, ethyl acetate / petroleum ether 5-30%) to give **29** (1.70 g, 3.08 mmol, 79% yield) as a white solid. LCMS [M+1]⁺: 540.0. ¹H NMR (400 MHz, CDCl₃) δ = 8.54 (d, *J* = 2.0 Hz, 1H), 8.12 (d, *J* = 2.4 Hz, 1H), 7.10 (br d, *J* = 7.2 Hz, 2H), 6.85 (d, *J* = 0.8 Hz, 1H), 6.48 (d, *J* = 2.4 Hz, 2H), 6.41 (d, *J* = 2.4 Hz, 1H), 6.39 (d, *J* = 2.0 Hz, 1H), 4.84 (br s, 4H), 3.79 (d, *J* = 2.4 Hz, 12H), 2.58 (s, 3H), 1.60 (s, 1H).

Compound 30: [6-[bis[(2,4-dimethoxyphenyl)methyl]amino]-8-methyl-1,5-naphthyridin-3yl]boronic acid.

Step 8: A mixture of compound **29** (300 mg, 557 μ mol, 1.00 *eq.*), 4,4,5,5-tetramethyl-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-1,3,2-dioxaborolane (212 mg, 836 μ mol, 1.50 *eq.*), potassium acetate (164 mg, 1.67 mmol, 3.00 *eq.*) and Pd(dppf)₂-DCM(45.5 mg, 55.7 μ mol, 0.10 *eq*) in 1,4-dioxane (5.0 mL) was degassed stirred at 110 °C for 3 hours under a nitrogen atmosphere. The cooled reaction mixture was filtered and concentrated under vacuum to give a residue. The residue was washed with methyl alcohol (20 mL) and filtered to give **30** (450 mg, crude) as a black solid. The material was used directly in the next step without further purification. LCMS [M+1]⁺: 503.9

Compound 31, 6-[bis[(2,4-dimethoxyphenyl)methyl]amino]-8-methyl-1,5-naphthyridin-3-ol.

Step 9: A mixture of compound **29** (200 mg, 371 µmol, 1.00 *eq*), Pd₂(dba)₃ (34.0 mg, 37.1 µmol, 0.100 *eq*), *t*-BuXphos (31.6 mg, 74.3 µmol, 0.200 *eq*) and potassium hydroxide (208 mg, 3.71 mmol, 10.0 *eq*) in dioxane (1.5 mL) and water (1.5 mL) was degassed and stirred at 100 °C for 12 hours under nitrogen atmosphere. The pH of the reaction mixture was adjusted to pH 7 with HCl (1 N). Then the mixture was poured into water (30 mL) and extracted with ethyl acetate (15 mL × 3). The combined organic layers were dried over sodium sulfate, filtered and concentrated under reduced pressure to give a residue. The residue was purified by column chromatography (SiO₂, ethyl acetate / petroleum ether 10-50%) to give **31** (170 mg, 354 µmol, 95% yield) as a yellow solid. ¹H NMR (400 MHz, CDCl₃) δ = 7.85 - 7.77 (m, 1H), 7.10 – 7.01 (m, 3H), 6.65 (s, 1H), 6.41 - 6.33 (m, 4H), 4.75 (s, 4H), 3.68 (d, *J* = 9.2 Hz, 12H), 2.58 (s, 3H).

Example 32: 7-(isothiazol-4-yl)-4-methyl-1,5-naphthyridin-2-amine.



A mixture of **28** (158 mg, 0.44 mmol, 1.00 eq.), isothiazol-4-ylboronic acid (114 mg, 0.89 mmol, 2.00 eq.), Pd(dppf)Cl₂ (32 mg, 44 µmol, 0.10 eq.) and cesium carbonate (143 mg, 0.89 mmol, 2.00 eq.) in dioxane (2.0 mL) and water (0.40 mL) was degassed and purged with nitrogen atmosphere for 3 times, and then the mixture was stirred at 100 °C for 1 hour under nitrogen atmosphere. The reaction mixture was diluted with water (2.0 mL) and extracted with ethyl acetate (2.0 mL × 3). Combined organic phase was washed with brine (2.0 mL), dried, filtered and concentrated to give a residue which was used into next step directly without further purification. The obtained residue was mixture with TFA (2.0 mL) and stirred at 70 °C for 6 hours. The reaction mixture was concentrated under reduced pressure to give a residue. The residue was basified with ammonium hydroxide (0.10 mL) to pH=9. The solution was purified by prep-HPLC (neutral condition) to give **32** (4.3 mg, 17.7 µmol, 4% yield over 2 steps) was obtained as a white solid. LCMS [M+1]⁺: 243.2; ¹H NMR (400 MHz, CD₃OD-*d*₄) δ = 9.37 (s, 1H), 9.04 (s, 1H), 8.90 (d, J = 2.0 Hz, 1H), 8.13 (d, J = 2.0 Hz, 1H), 6.94 (d, J = 1.2 Hz, 1H), 2.67 - 2.64 (d, J = 1.2 Hz, 3H).

Example 33: 4-methyl-7-(3-pyridyloxy)-1,5-naphthyridin-2-amine.



Step 1: A mixture of 6-[bis[(2,4-dimethoxyphenyl)methyl]amino]-8-methyl-1,5-naphthyridin-3ol (60 mg, 126 µmol, 1.00 eq.), 3-bromopyridine (60 mg, 379 µmol, 37 µL, 3.0 eq.), CuI (4.8 mg, 25 µmol, 0.20 eq.), 2-(dimethylamino) acetic acid (5.2 mg, 51 µmol, 0.40 eq.) and cesium carbonate (127 mg, 391 µmol, 3.10 eq.) in dioxane (1.0 mL) was degassed and purged with nitrogen 3 times, and then the mixture was stirred at 100 °C for 12 hrs. The reaction mixture was filtered and concentrated under reduced pressure to give a residue. The residue was purified by prep-TLC (SiO₂, dichloromethane: methyl alcohol = 20:1) to give compound *N*,*N*-bis[(2,4dimethoxyphenyl)methyl]-4-methyl-7-(3-pyridyloxy)-1,5-naphthyridin-2-amine (46 mg, 83 µmol, 66% yield) as a yellow solid. LCMS [M+1] ⁺: 553.2.

Step 2: A mixture of *N*,*N*-bis[(2,4-dimethoxyphenyl)methyl]-4-methyl-7-(3-pyridyloxy)-1,5naphthyridin-2-amine (46 mg, 83 µmol, 1.00 eq.) and trifluoroacetic acid (1.00 mL) was stirred at 80 °C for 0.5 hrs. The mixture was concentrated under vacuum. Then ammonium hydroxide (2.0 mL) was added, and the mixture was stirred for 0.5 hrs. The mixture was concentrated, and the formed residue was purified by prep-HPLC (ammonia hydroxide conditions) to give **33** (13 mg, 53 µmol, 64% yield) as a white solid. LCMS [M+1] +: 253.2; ¹H NMR (400 MHz, CDCl₃) δ = 8.56 (d, J = 2.8 Hz, 1H), 8.53 (d, J = 2.8 Hz, 1H), 8.49 (dd, J = 4.4, 1.2 Hz, 1H), 7.46 - 7.43 (m, 1H), 7.36 - 7.32 (m, 1H), 7.31 (d, J = 2.4 Hz, 1H), 6.73 (d, J = 1.2 Hz, 1H), 4.78 (br, s, 2H), 2.70 (d, J = 0.8 Hz, 3H).

Example 34: 4-methyl-7-(3-pyridylmethyl)-1,5-naphthyridin-2-amine.



Step 1: A mixture of **30** (400 mg, 795 μmol, 1. 0 *eq.*), 3-(chloromethyl)pyridine (203 mg, 1.59 mmol, 2.00 *eq.*), cyclopentyl(diphenyl)phosphane;dichloromethane;dichloropalladium;iron (64.9 mg, 80 μmol, 0.1 *eq.*) and potassium carbonate (220 mg, 1.59 mmol, 2.00 *eq.*) in 1,4-dioxane (4.0

mL) and water (0.8 mL) was degassed and purged with nitrogen 3 times, and then stirred at 120 °C for 2 hours. The reaction mixture was filtered and concentrated under reduced pressure and the formed residue was purified by column chromatography (SiO₂, petroleum ether/ethyl acetate 10/1 to 0/1) to give *N*,*N*-bis[(2,4-dimethoxyphenyl)methyl]-4-methyl-7-(3-pyridylmethyl)-1,5-naphthyridin-2-amine (160 mg, 287 µmol, 36% yield) as a yellow solid. LCMS [M+1] +: 551.5 **Step 2:** A solution of *N*,*N*-bis[(2,4-dimethoxyphenyl)methyl]-4-methyl-7-(3-pyridylmethyl)-1,5-naphthyridin-2-amine (150 mg, 272 µmol, 1.00 *eq*.) in trifluoroacetic acid (1.00 mL) was stirred at 70 °C for 0.5 hour. The reaction mixture was then diluted with dichloromethane, filtered and the filtrate concentrated under reduced pressure. The residue was purified by *prep*-HPLC (ammonium bicarbonate condition) to give **34** (36 mg, 142 µmol, 52% yield) as a white solid. LCMS [M+1] +: 251.3; ¹H NMR (400 MHz, CD₃OD-*d*₄) δ = 8.52 (d, *J* = 2.0 Hz, 1H), 8.45 (d, *J* = 2.4 Hz, 1H), 8.42 (dd, *J* = 1.6, 4.8 Hz, 1H), 7.76 (d, *J* = 8.0 Hz, 1H), 7.64 (d, *J* = 2.0 Hz, 1H), 7.40 (dd, *J* = 4.8, 8.0 Hz, 1H), 6.87 (d, *J* = 0.8 Hz, 1H), 4.20 (s, 2H), 2.60 (s, 3H); HPLC Rt 1.538 min, 99.5%.

Example 35: 4-methyl-7-[(1-methylpyrazol-4-yl)methyl]-1,5-naphthyridin-2-amine.



Step 1: A mixture of **30** (60 mg, 119 μmol, 1.00 *eq.*), 4-(chloromethyl)-1-methyl-pyrazole (40 mg, 238 μmol, 2.00 *eq.*),

cyclopentyl(diphenyl)phosphane;dichloromethane;dichloropalladium;iron (10 mg, 12 μ mol, 0.10 eq.) and potassium carbonate (49 mg, 358 μ mol, 3.00 eq.) in DMF (1.0 mL) was degassed and purged with nitrogen for 3 times. The mixture was then stirred at 100 °C for 2 hours. The reaction mixture was diluted with water (10 mL) and then extracted with ethyl acetate (20 mL × 3). The combined organic layers were washed with brine (50 mL × 2), dried with sodium sulfate, filtered, and concentrated under reduced pressure. The formed residue was purified by *prep*-TLC (SiO₂, petroleum ether/ethyl acetate 1/1) to give *N*,*N*-bis[(2,4-dimethoxyphenyl)methyl]-4-methyl-7-[(1-methylpyrazol-4-yl)methyl]-1,5-naphthyridin-2-amine (10 mg, 18 μ mol, 15% yield) as a white solid. LCMS [M+1] +: 554.6.

Step 2: A solution of *N*,*N*-bis[(2,4-dimethoxyphenyl)methyl]-4-methyl-7-[(1-methylpyrazol-4-yl)methyl]-1,5-naphthyridin-2-amine (10 mg, 18 µmol, 1.00 *eq*.) in trifluoroacetic acid (0.5 mL) was stirred at 70 °C for 0.5 hour. The reaction mixture was then diluted with dichloromethane, filtered, and the filtrate concentrated under reduced pressure. The residue was purified by prep-HPLC (ammonium bicarbonate condition) to give **35** (1.6 mg, 6.1 µmol, 34% yield) as a white solid. LCMS [M+1] +: 254.3; ¹H NMR (400 MHz, CD₃OD-*d*₄) δ = 8.43 (d, *J* = 2.0 Hz, 1H), 7.68 - 7.63 (m, 1H), 7.47 (s, 1H), 7.36 (s, 1H), 6.87 (d, *J* = 0.8 Hz, 1H), 3.99 (s, 2H), 3.85 (s, 3H), 2.60 (d, *J* = 0.8 Hz, 3H); HPLC Rt 1.947 min, 99.6%.

Example 36: N-[(6-amino-8-methyl-1,5-naphthyridin-3-yl)methyl]benzamide.



Step 1: A mixture of **29** (1.70 g, 3.16 mmol, 1.00 *eq.*), potassium (*N*-Bocaminomethyl)trifluoroborate (1.50 g, 6.31 mmol, 2.00 *eq.*), [2-(2aminophenyl)phenyl]palladium(1+);bis(1-adamantyl)-butyl-phosphane;methanesulfonate (230 mg, 316 μmol, 0.10 *eq.*), sodium carbonate (1.00 g, 9.47 mmol, 3.0 *eq.*) in water (15 mL) and dioxane (75 mL) was degassed and purged with nitrogen 3 times. The mixture was then stirred at 100 °C for 12 hours under nitrogen atmosphere. The mixture was concentrated and the residue was triturated with petroleum ether/ethyl acetate 10/1 (50 mL) for 0.5 hour to give *tert*-butyl *N*-[[6-[bis[(2,4-dimethoxyphenyl)methyl]amino]-8-methyl-1,5-naphthyridin-3yl]methyl]carbamate, **29a** (1.60 g, 2.72 mmol, 86% yield) as a white solid. LCMS [M+1] ⁺: 589.3.

Step 2: A mixture of 29a (1.50 g, 2.55 mmol, 1.00 eq.) in trifluoroacetic acid (5.0 mL) was stirred at 70 °C for 1 hour. The mixture was concentrated under reduced pressure and the residue was diluted with water (10 mL) and washed ethyl acetate (20 mL \times 2). The pH of the aqueous phase was then adjusted to pH 9 with sodium bicarbonate and exacted with ethyl acetate (100 mL \times 10). The combined organic phases were washed with brine (30 mL \times 2), dried over

anhydrous sodium sulfate, filtered, and concentrated under reduced pressure to give a 7-(aminomethyl)-4-methyl-1,5-naphthyridin-2-amine, **29b** (280 mg, 1.49 mmol, 58% yield) as a yellow solid. The residue was used for the next step without further purification. LCMS [M+1] ⁺: 189.1; ¹H NMR (400 MHz, CD₃OD- d_4) $\delta = 8.55$ (d, J = 2.0 Hz, 1H), 7.85 (d, J = 2.0 Hz, 1H), 6.92 (d, J = 1.0 Hz, 1H), 4.07 (s, 2H), 2.62 (d, J = 0.8 Hz, 3H). **Step 3**: To a solution of benzoic acid (16 mg, 128 µmol, 1.00 *eq.*) in DMF (1.0 mL) was added HATU (73 mg, 191 µmol, 1.50 *eq.*) and triethylamine (53 µL, 383 µmol, 3.00 *eq.*). The mixture was stirred at 20 °C for 0.5 hour and then **29b** (31 mg, 166 µmol, 1.30 *eq.*) was added to the mixture and stirred at 20°C for 0.5 hour. The mixture was concentrated under reduced pressure to give a residue. The residue was purified by *prep*-HPLC (HCl condition) to give **36** (7.5 mg, 25 µmol, 19% yield, HCl salt) as a white solid. LCMS [M+1] ⁺: 293.2; ¹H NMR (400 MHz,

CD₃OD-*d*₄) δ = 8.82 (d, *J* = 2.0 Hz, 1H), 7.98 (br d, *J* = 2.0 Hz, 1H), 7.94 - 7.83 (m, 2H), 7.61 - 7.55 (m, 1H), 7.53 - 7.46 (m, 2H), 7.12 (s, 1H), 4.79 (s, 2H), 2.75 (s, 3H); HPLC Rt 1.667 min, 95.9%.

Example 37: N-((6-amino-8-methyl-1,5-naphthyridin-3-yl)methyl)pyrimidine-4-carboxamide.



Step 1: To a solution of **29a** (780 mg, 1.32 mmol, 1.00 *eq.*) in dichloromethane (10 mL) was added zinc bromide (895 mg, 3.97 mmol, 199 μ L, 3.00 *eq.*). The mixture was stirred at 25 °C for 12 hours. The reaction mixture was concentrated, and the formed residue was diluted with water (20 mL) and extracted with ethyl acetate (20 mL × 3). The combined organic layers were washed with brine (50 mL), dried over anhydrous sodium sulfate, filtered, and concentrated under reduced pressure. The residue was triturated with ethyl acetate (10 mL) at for 30 minutes to give

7-(aminomethyl)-*N*,*N*-bis[(2,4-dimethoxyphenyl)methyl]-4-methyl-1,5-naphthyridin-2-amine **29c** (480 mg, 982 μmol, 74% yield) as a yellow solid. LCMS [M+1] ⁺: 489.1.

Step 2: To a solution of **29c** (217 mg, 443 μ mol, 1.10 *eq*.) and pyrimidine-4-carboxylic acid (50.0 mg, 403 μ mol, 1.00 *eq*.) in dichloromethane (2.0 mL) was added T3P (50% in ethyl acetate, 2.01 mmol, 1.20 mL, 5.00 *eq*.) and triethylamine (1.21 mmol, 168 μ L, 3.00 *eq*.). The mixture was stirred at 20 °C for 2 hours. The mixture was concentrated under reduced pressure to give a residue. The residue was purified by *prep*-TLC (SiO₂, dichloromethane: methyl alcohol = 10:1) to give *N*-[[6-[bis[(2,4-dimethoxyphenyl)methyl]amino]-8-methyl-1,5-naphthyridin-3-yl]methyl]pyrimidine-4-carboxamide (80 mg, 135 μ mol, 33% yield) as a white solid. LCMS [M+1] +: 595.2.

Step 3: To a solution of *N*-[[6-[bis](2,4-dimethoxyphenyl)methyl]amino]-8-methyl-1,5naphthyridin-3-yl]methyl]pyrimidine-4-carboxamide (80 mg, 135 µmol, 1.00 *eq*) in trifluoroacetic acid (1.50 mL) was stirred at 70 °C for 0.5 hour. The mixture was concentrated under reduced pressure to give a residue. The residue was purified by *prep*-HPLC (HCl condition)) to give **37** (17 mg, 57 µmol, 43% yield) as a white solid. LCMS [M+1] + =295.1; ¹H NMR (400 MHz, DMSO-*d*₆) δ = 14.68 - 14.08 (m, 1H), 9.92 (br t, *J* = 6.2 Hz, 1H), 9.48 - 9.24 (m, 2H), 9.12 (d, *J* = 5.0 Hz, 1H), 8.80 (d, *J* = 2.0 Hz, 1H), 8.48 - 8.24 (m, 1H), 8.08 - 8.00 (m, 2H), 7.16 (d, *J* = 1.0 Hz, 1H), 4.68 (br d, *J* = 6.2 Hz, 2H), 2.64 (d, *J* = 0.8 Hz, 3H); HPLC Rt 1.440 min, 100%.

PRMT5/MTA X-ray Co-crystallography

PRMT5/MTA protein was expressed, purified, and crystallized as previously described.¹ Crystals were harvested and incubated in a soak solution consisting of reservoir and 5-10 mM fragment for 2-5 hours, dipped in cryoprotectant (soak solution plus 20% ethylene glycol) and flash frozen in pucks for data collection. X-ray diffraction data was collected at the following beamlines: Advanced Photon Source (APS) NE-CAT ID-E, Swiss Light Source (SLS) PXII/X10SA and PXIII/X06DA, and Diamond Light Source (DLS) I03 (Table ESI-1). PRMT5 data sets indexed in the I222 space group and the images were integrated and scaled using HKL2000.² The PRMT5 crystals were highly anisotropic and STARANISO server was used to calculate ellipsoidal completeness for several structures.³ Structures were determined by molecular replacement using Phaser⁴ and refined through iterative rounds of automated refinement using phenix.refine⁵ and manual refitting in Coot.⁶ Coordinates and structure factors are deposited in the PDB with accession codes: 7UY1, 7UYF, 7ZUP, 7ZUQ, 7ZUU, 7ZUY, 7ZV2, 7ZVL, 7ZVU, 8CSG, 8CTB.

Table ESI-1. X-ray data collection and refinement statistics for PRMT5 crystal structures						
Compound	Fragment 5	Hisegumpehet 73	Firsaynmyeliet 94			
PDB ID	870.59G	7820/IB	772LDKOF			
Data Collection						
Beamline	PX111/X069A(SLS)	APRSSIDGE	PXIII/20060043(SLS)			
Wavelength (Å)	0.999	0.979	0.999			
Detector	PILATUS 6M	EIGHGEREOIGIM	ERGIEAZTIXIS 260M			
Resolution (Å) ^a	46 - 2.66 2.82 (2.73) 46 - 2.66 (2.91-2.66) 2.48)	44.7 -261 (2.64-2.39) (2.61 (2.64-2.61) (2.75-2.39)	109,2 - 2.48 108.6 (2.82 (3.0) - 2.82) (2.84 - 2.48) - 2.82)			
Space Group	1222	1222	1222			
Unit Cell a,b,c (Å)	100.Z, 138.2, 178.0	1998005, 11337824, 11778835	102.9;, 138.2,, 178.3			
# Unique Refls	43203524 (18337024))	2218803971 ((1402972))	29233 (6669)			
Redundancy	2.8 (2.8)	2.7 (1.9)	5.5 (5.4)			
Completeness (%)	95.7 (98.2)	76.1 (23.7)	98.5 (99.5)			
< 1/s _i >	17.4 (3.2)	9.4 (1.5)	16.9 (3.3)			
R _{merge} ^b	0.06 (0.44)	0.13 (0.60)	0.09 (0.45)			
CC _{1/2} ^c	0.95	0.99	0.93			
Refinement						
R _{Free} (%) ^e	23.9	28.2	27.2			
R (%) ^d	21.0	23.2	22.1			
B Average (Å ²)	28.7	75.5	33.2			
RMS Deviations						
Bond Lengths(Å)	0.011	0.001	0.007			
Bond Angles (°)	1.6	0.38	1.2			
Ramachandran (%)						
Favored	99.6	93.1	99.6			
Outliers	0.2	0.4	0.1			

Table ESI-1 (continued). X-ray data collection and refinement statistics for PRMT5 crystal						
structures Redundancy	5.0 (5.1)	11.5 (11.5)	23 (22)			
Completendess (%)	Eggagn(øle.4)8	Exaggipte 22	Exagypie 25			
PDB ID	2 5 20(p ^{3.1)}	¹² 72 1 29)	¹ 7 2 (10 ⁸⁾			
Data Collection						
Beamline	DLS 103	DLS 103	DLS 103			
Refinement (Å)	0.919	0.919	0.919			
Betector	Eiger2% 16M	Eiger28 2 16M	EigerᢓᠯᢆX £ 16M			
$R(\%)^d$	109.5 -21.98 (2.26-	109.21-91.94	109.28-91.94			
B Average (Å ²)	1.4978)	(2.165.1394)	(2.1375.1694)			
RMS Deviations	1222	1222	1222			
Unit Cell a,b,c (Å) Bond Lengths(Å)	104.2, <u>138,6</u> , 178.8 0.003	104.5, <u>138,7</u> , 178.9 0.008	105.2, <u>139,2,</u> 178.6 0.005			
# Unique Refls ´ Bond Angles (°)	39692 (1984) 1.0	49072 (2455) 1.6	60214 (3012) 1.3			
Redundancy Ramachandran	<u>11.9 (14.7)</u>	11.8 (13.1)	12.1 (14.1)			
ဖြံ့စွဲmpleteness (%)	93.2∆	93.6∆	94.9 [∆]			
Fall®ored	10 <i>q</i> 9(ჭ .9)	13 9 9(2 .7)	13.@p(௹.7)			
Bruttiers	0.1 6 . <u>⁄2</u> 1.5)	0.10.61.5)	0.10.41.4)			
CC _{1/2} °	0.99	0.99	0.99			
Refinement						
R _{Free} (%) ^e	27.0	27.5	26.5			
R (%) ^d	22.1	23.4	23.8			
B Average (Å ²)	30.7	33.1	30.6			
RMS Deviations						
Bond Lengths(Å)	0.007	0.007	0.014			

Bond Angles (°)	1.5	1.4		1.8		
Ramachandran						
Favored 99.3 Table ESI-1 (continued). X-ray data collection and refinement statistics for PRMT5 crystal structures. 0.7						
Compound	Example 26		e 26 Example 34			
PDB ID	7ZUY		7200			
Data Collection						
Beamline	DLS I03	3	DLS I03			
Wavelength (Å)	0.919	0.919		0.919		
Detector	Eiger2 XE	16M		Eiger2 XE 16M		
Resolution (Å)ª	109.7 - 1 (2.23-1.9	109.7 - 1.97 (2.23-1.97)		109.7 - 1.95 (2.17-1.95)		
Space Group	Ì1222	1222		I222		
Unit Cell a,b,c (Å)	105.2, 139.1	105.2, 139.1, 178.5		105.2, 139.2, 178.5		
# Unique Refls	48889 (2444)		60214 (3012)			
Redundancy	11.8 (13	11.8 (13.8)		12.1 (14.1)		
Completeness (%)	93.7△		94.6 ^Δ			
< 1/s;>	12.8 (1.7)		13.8 (1.7)			
R _{merge} ^b	0.10 (1.5)		0.1 (1.4)			
CC _{1/2} ^c	0.99		0.99			
Refinement						
R _{free} (%) ^e	26.9	26.9		28.0		
R (%) ^d	22.4		23.0			
B Average (Ų)	31.2		29.5			
RMS Deviations						
Bond Lengths(Å)	0.07			0.07		
Bond Angles (°)	1.5	1.5		1.4		
Ramachandran (%)						
Favored	99.7	99.7		99.6		
Outliers	0.3		0.4			
^a Numbers in parentheses refer to the highest resolution shell. ${}^{b}R_{merge} = \Sigma_{hkl}\Sigma_i I_{hkl,i} - \langle I_{hkl} \rangle /\Sigma_{hkl}\Sigma_i I_{hkl,l}$						
where $I_{hkl,i}$ is the scaled intensity of the <i>i</i> th measurement of reflection hkl, $\langle I_{hkl} \rangle$ is the average intensity						
for that reflection. °CC _{1/2} = Pearson Correlation Coefficient between two random half datasets.						
${}^{d}R = \Sigma_{hkl} F_{o} - F_{c} / \Sigma_{hkl} F_{o} x 100, e_{R_{free}}$ was calculated as for <i>R</i> , but on a test set comprising 5% of						

the data excluded from refinement.

^ΔEllipsoidal completeness was calculated using Staraniso.

References

Smith, C. R.; Aranda, R.; Bobinski, T. P.; Briere, D. M.; Burns, A. C.; Christensen, J. G.; Clarine, J.; Engstrom, L. D.; Gunn, R. J.; Ivetac, A.; Jean-Baptiste, R.; Ketcham, J. M.; Kobayashi, M.; Kuehler, J.; Kulyk, S.; Lawson, J. D.; Moya, K.; Olson, P.; Rahbaek, L.; Thomas, N. C.; Wang, X.; Waters, L. M.; Marx, M. A., Fragment-Based Discovery of MRTX1719, a Synthetic Lethal Inhibitor of the PRMT5*MTA Complex for the Treatment of MTAP-Deleted Cancers. *J Med Chem* 2022, *65* (3), 1749-1766.

2. Otwinowski, Z.; Minor, W., Processing of X-ray diffraction data collected in oscillation mode. In *Methods Enzymol*, 1997/01/01 ed.; 1997; Vol. 276, pp 307-26.

Tickle, I. J.; Flensburg, C.; Keller, P.; Paciorek, W.; Sharff, A.; Smart, O.; Vornhein,
 C.; Bricogne, G. STARANISO, 3.344; Global Phasing Limited: 2022.

4. McCoy, A. J.; Grosse-Kunstleve, R. W.; Adams, P. D.; Winn, M. D.; Storoni, L. C.; Read, R. J., Phaser crystallographic software. *J Appl Crystallogr* **2007**, *40* (Pt 4), 658-674.

5. Liebschner, D.; Afonine, P. V.; Baker, M. L.; Bunkoczi, G.; Chen, V. B.; Croll, T. I.;

Hintze, B.; Hung, L. W.; Jain, S.; McCoy, A. J.; Moriarty, N. W.; Oeffner, R. D.; Poon, B.

K.; Prisant, M. G.; Read, R. J.; Richardson, J. S.; Richardson, D. C.; Sammito, M. D.;

Sobolev, O. V.; Stockwell, D. H.; Terwilliger, T. C.; Urzhumtsev, A. G.; Videau, L. L.;

Williams, C. J.; Adams, P. D., Macromolecular structure determination using X-rays, neutrons and electrons: recent developments in Phenix. *Acta Crystallogr D Struct Biol* **2019**, *75* (Pt 10), 861-877.

6. Emsley, P.; Cowtan, K., Coot: model-building tools for molecular graphics. *Acta Crystallogr D Biol Crystallogr* **2004**, *60* (Pt 12 Pt 1), 2126-32.