

Engineered Protein-Iron Oxide Hybrid Biomaterial for MRI-traceable Drug Encapsulation

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SUPPORTING INFORMATION

Determination of Q protein's binding capacity for doxorubicin

The optimal Q protein-to-doxorubicin (Dox) binding ratio was assessed *via* spectrophotometric assay using Q_{WT} protein at 10 μ M and 0-100 μ M DMSO-dissolved Dox, in 50 mM PB pH 7.4 containing 1% v/v DMSO. The fluorescence intensity of all conditions was assessed spectrophotometrically at 600 nm following overnight (16 hr) binding at 300 rpm and room temperature (**Figure S4**). While an initial plateau in fluorescence intensity occurred at 1:2, the fluorescence was fully saturated at 1:7.5 Q:Dox. Therefore, a 1:5 Q:Dox ratio was considered the near-saturation ratio and used for all Dox binding studies.

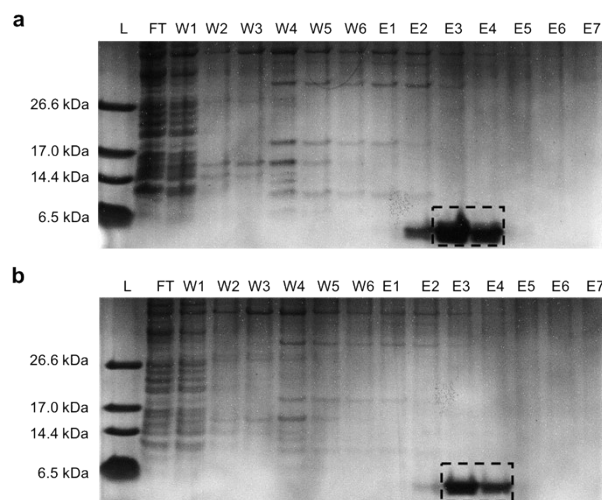


Figure S1. 12% SDS-PAGE of (a) Q_{WT} and (b) Q_{AHA} purified under denaturing conditions *via* immobilized metal affinity chromatography with the following samples: L: ladder, FT: flow through, W1-W3: washes 1-3 using purification buffer with 0 mM imidazole, W4-W6: washes 4-6 using 5 mM imidazole, and E1-E7: elutions 1-7 containing 10, 20, 50, 100, 200, 500, and 500 mM imidazole, respectively. Dashed boxes illustrate elution fractions collected.

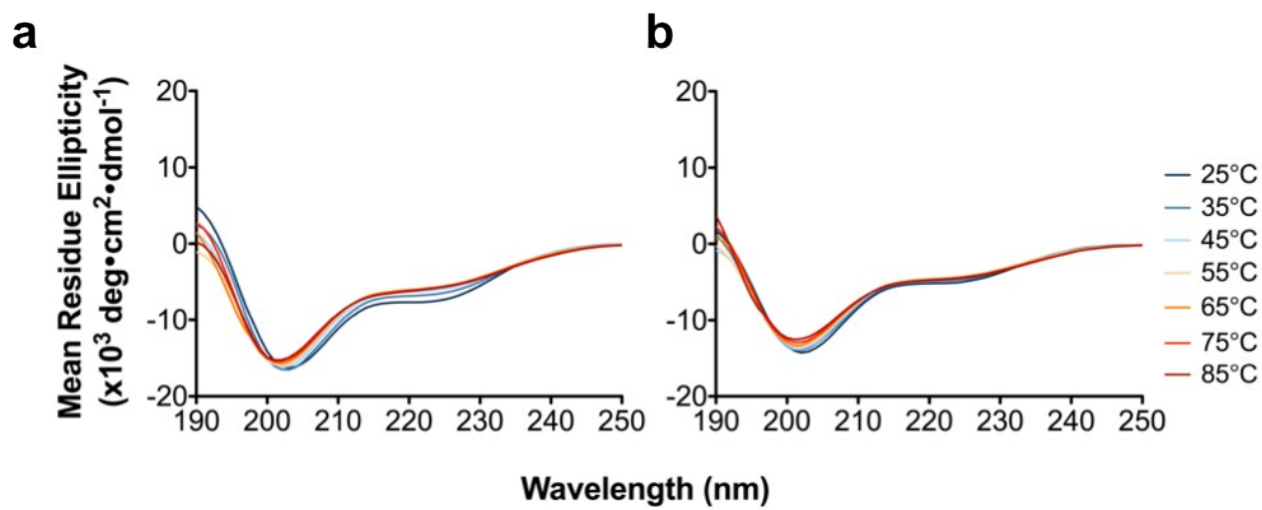


Figure S2. Circular dichroism (CD) of (a) Q_{WT} and (b) Q_{AHA} at pH 4.0 acquired every 10°C from 25°C to 85°C. Data depict the average of three independent trials.

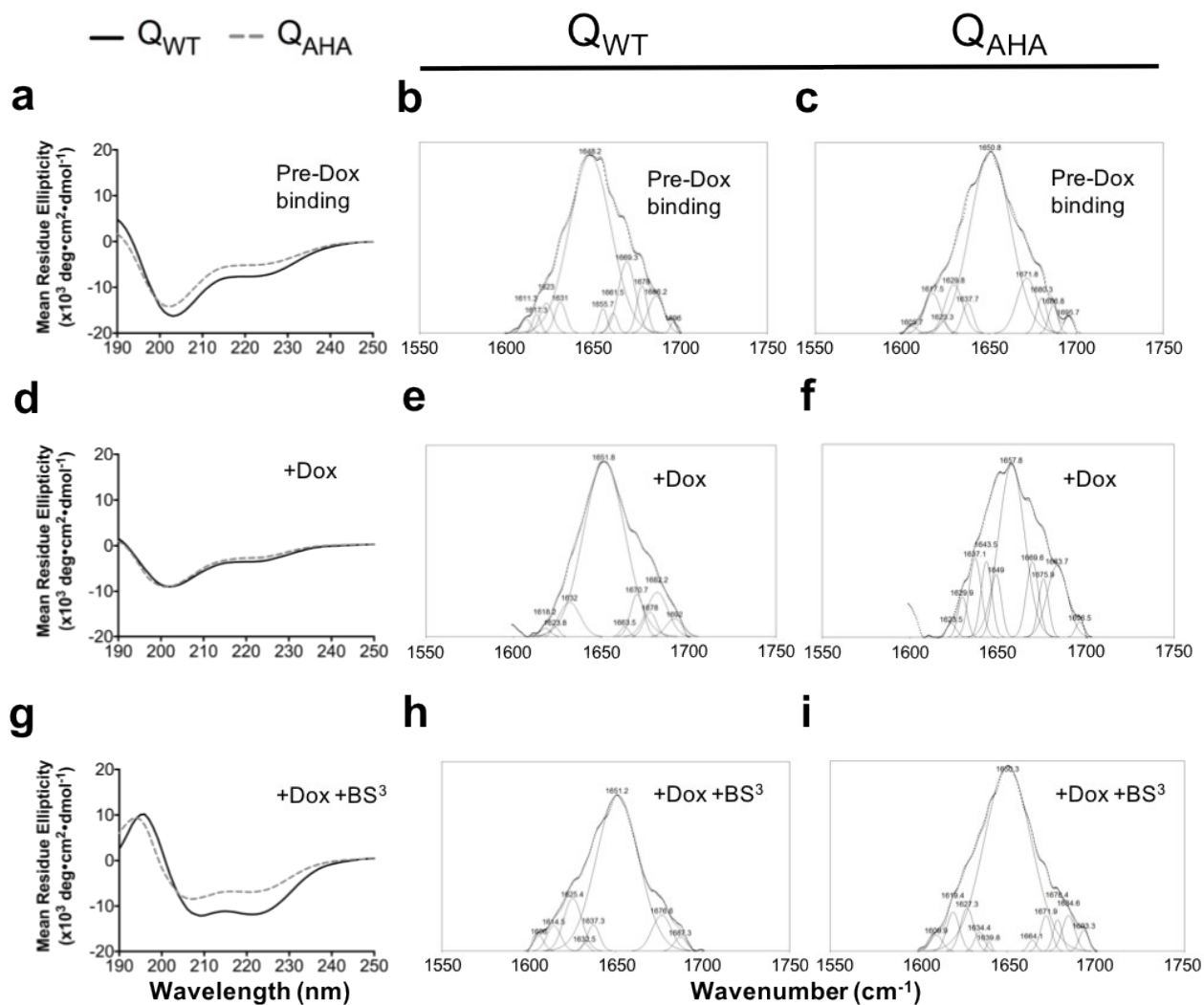


Figure S3. Secondary structure of Q proteins pre- and post-Dox binding and crosslinking. CD wavelength scans and ATR-FTIR spectra acquired for Q_{WT} and Q_{AHA} (a-c) at pH 4.0 prior to Dox binding, (d-f) Dox-bound, and (g-i) Dox-bound and crosslinked. All CD data depict the average of three independent trials and all ATR-FTIR spectra represent two independent trials.

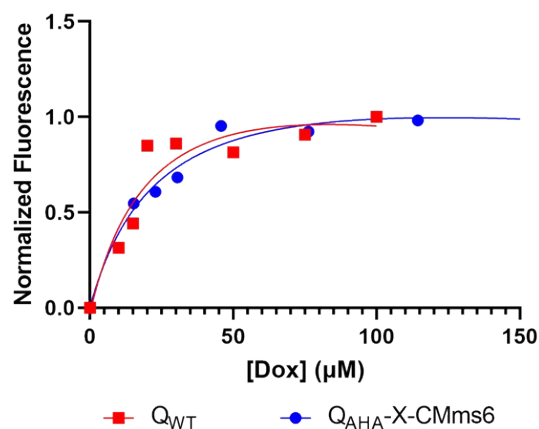


Figure S4. Assessment of Q_{WT}:Dox and Q_{AHA-X-CMms6}:Dox binding ratio. Normalized fluorescence intensity following overnight (16 hr) incubation of 10 μM Q_{WT} protein bound to increasing concentrations of Dox to assess the optimal molar ratio for Dox binding by Q. Binding affinities (K_d) are 24.3 μM Dox and 26.0 μM Dox for Q_{WT} and Q_{AHA-X-CMms6} respectively indicating saturation at 50 μM or a 1:5 ratio of Dox-to-protein.

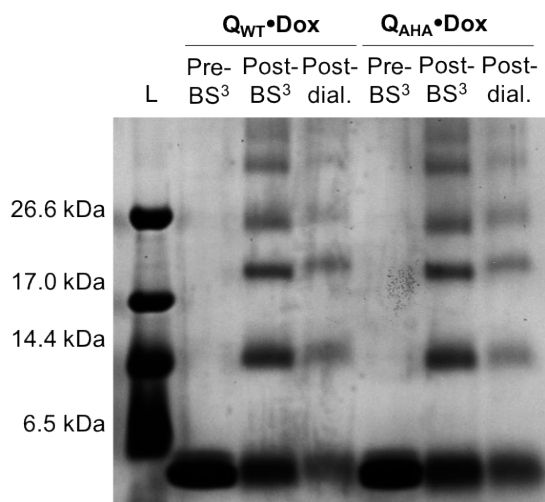


Figure S5. 12% SDS-PAGE including ladder (L) and Dox-bound Q_{WT} and Q_{AHA} mesofibers (Q_{WT}•Dox and Q_{AHA}•Dox) pre- and post-BS³ chemical crosslinking and following subsequent dialysis.

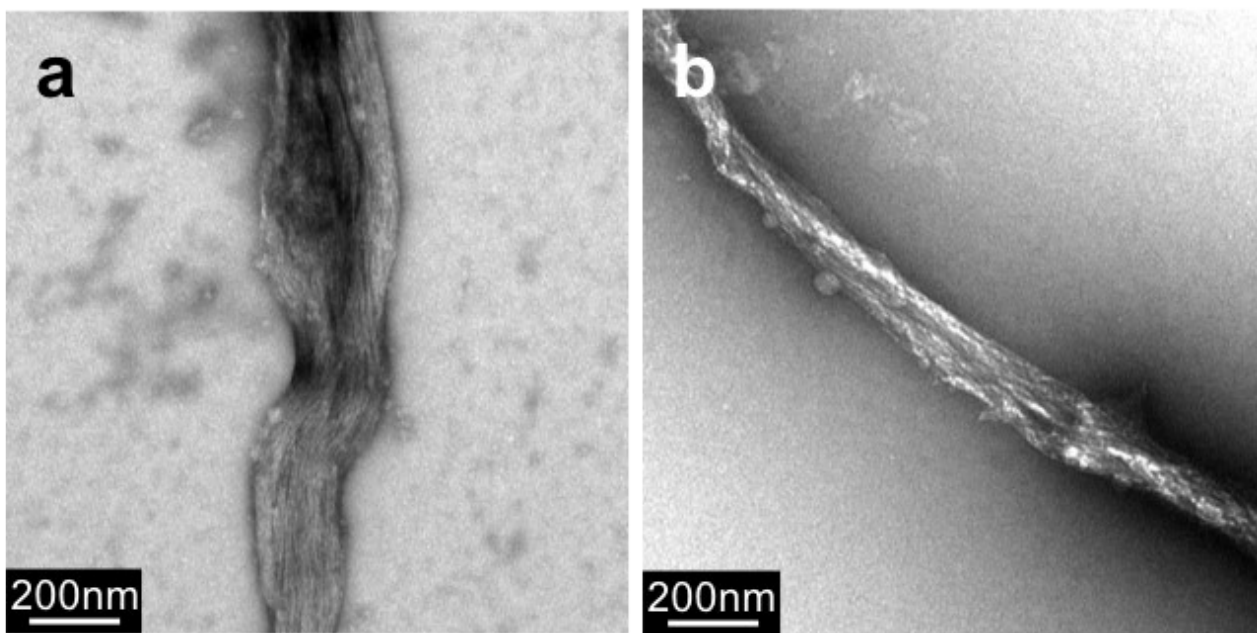


Figure S6. Fiber maintenance under Dox binding conditions in the absence of Dox. Uranyl-acetate-stained TEM following overnight incubation in 50 mM PB pH 7.4 + 1% DMSO of (a) Q_{WT} and (b) Q_{AHA} to confirm nanofiber assembly is maintained under binding conditions in the absence of Dox.

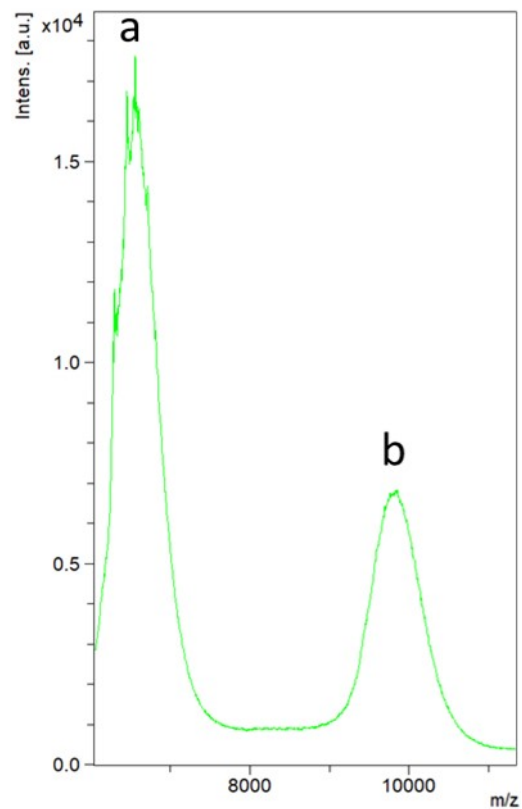


Figure S7. MALDI-TOF spectra after crosslinking of Q_{AHA} and clicking to CMms6 and before dialysis showing a m/z difference corresponding to 2.9 kDa. **a.** Unreacted Q_{AHA} 6.4 kDa. **b)** Q_{AHA} reacted with one CMms6 peptide

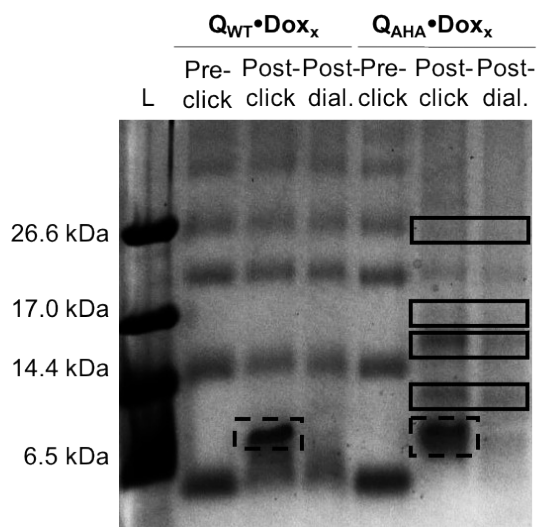


Figure S8. SDS-PAGE assessment of azide-alkyne cycloaddition reaction. 12% SDS-PAGE including ladder (L) and crosslinked Q_{WT} and Q_{AHA} mesofibers (Q_{WT}•Dox_x and Q_{AHA}•Dox_x) pre and post-copper-catalyzed azide-alkyne cycloaddition reaction with alkyne-bearing CMms6 peptide and following subsequent dialysis. Unreacted prg-CMms6 is shown with dashed boxes while solid boxes highlight the clicked product, dubbed Q_{AHA}-X-CMms6.

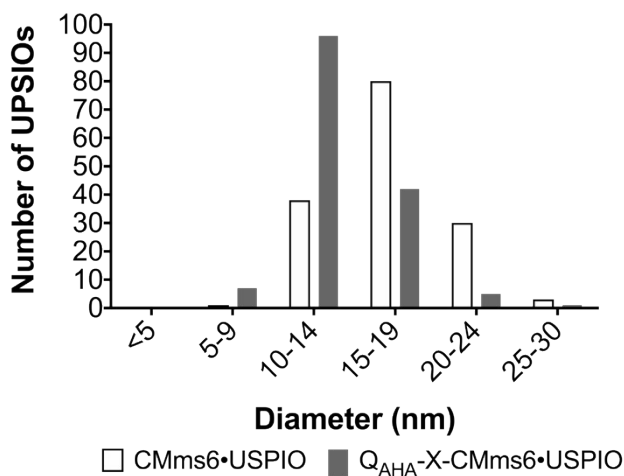
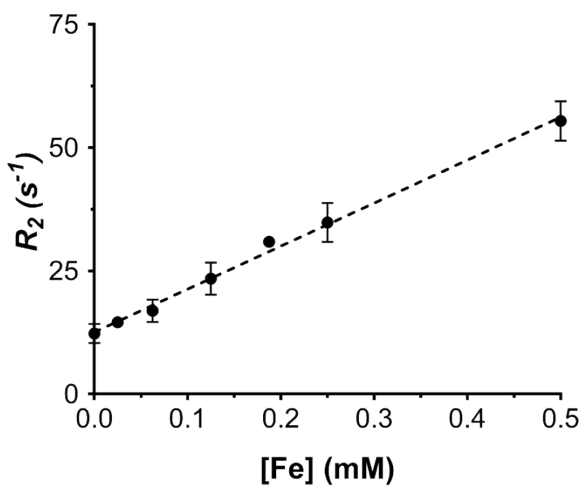


Figure S9. Size distribution of USPIOs visualized on brightfield TEM following co-precipitation reaction in the presence of CMms6 alone (CMms6•USPIO) or CMms6-conjugated Q_{AHA}-X-CMms6 (Q_{AHA}-X-CMms6•USPIO).

Feraheme



Q_{AHA}-X-CMms6•USPIO

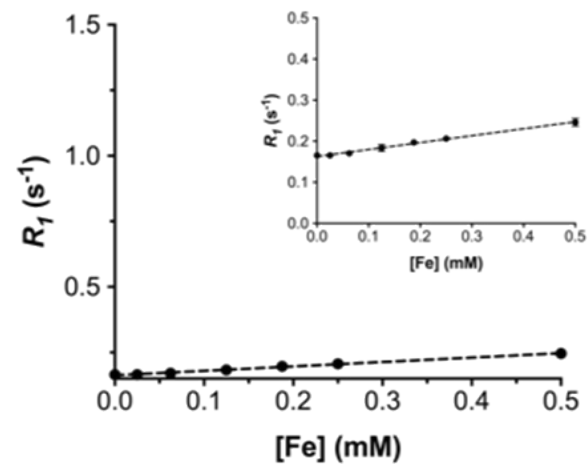
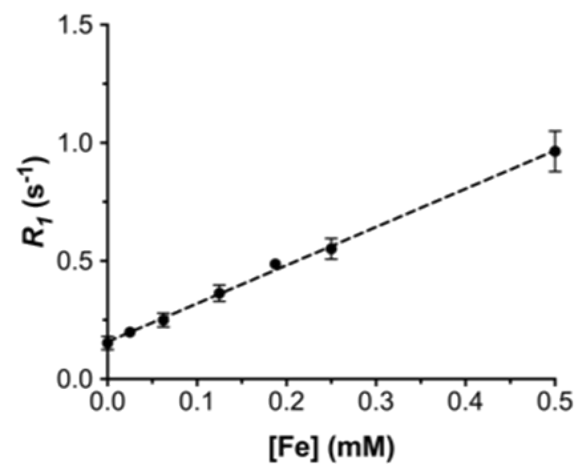
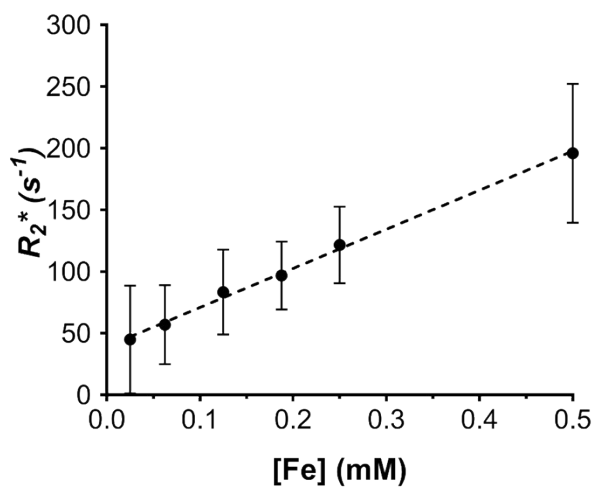
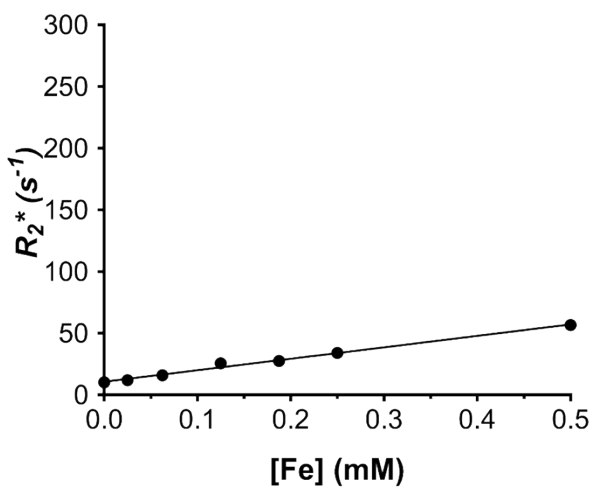
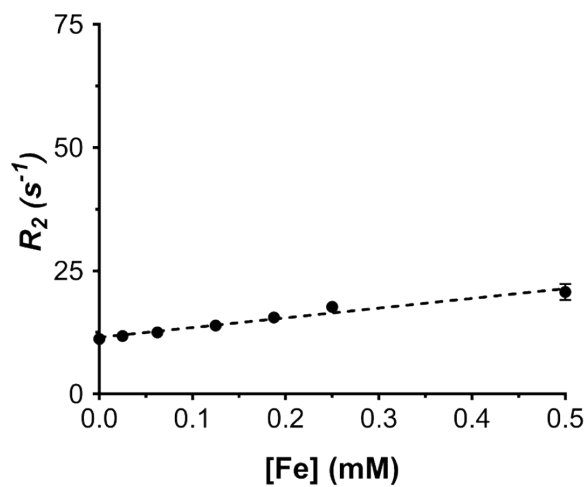


Figure S10. Relaxation curves from magnetic resonance relaxometry. Relaxation rates, R_2 , R_2^* and R_1 , as a function of iron concentration in Feraheme and Q_{AHA}-X-CMms6•USPIO. All data depict the average and standard deviation of three independent trials. Note the low R_1 relaxivity of Q_{AHA}-X-CMms6•USPIO reflected by the weak slope in comparison to Feraheme. This was confirmed *in vivo* in tissue muscle

Table S1. Protein expression yield. Average Q_{WT} and Q_{AHA} yields from expression in M15MA *E.coli*. Data is reported as the mean value \pm standard deviation.

Protein	Yield (mg L ⁻¹)
Q _{WT}	17.82 \pm 4.44 (N=14)
Q _{AHA}	8.70 \pm 1.58 (N=12)

Table S2. AHA Incorporation into the Q protein. Percent AHA incorporated into Q_{AHA} determined by MALDI-TOF MS and amino acid analysis. MALDI-TOF MS data is reported as the mean value \pm standard deviation of six independent trails.

Method	AHA Incorporation (%)
MALDI-TOF MS	88.53 \pm 5.03 (N=6)
Amino Acid Analysis (UC Davis Genome Center)	90.00

Table S3. Secondary structure of Q proteins in response to temperature. Mean residue ellipticity values from CD of Q_{WT} and Q_{AHA} at pH 4.0 from 25°C to 85°C and corresponding predicted secondary structure composition from CONTIN/LL assessment of the spectra. Data is reported as the average of three independent trials.

		$\Theta \times 10^3$ (deg·cm ² ·dmol ⁻¹)			% composition		
		Θ_{222}	Θ_{\min}	$\Theta_{222} / \Theta_{\min}$	α -helix	β -content	unordered
25°C	Q _{WT}	-7.63	-16.28	0.47	56	21	23
	Q _{AHA}	-5.15	-14.25	0.36	55	24	21
35°C	Q _{WT}	-6.76	-16.54	0.41	67	33	-
	Q _{AHA}	-4.80	-13.95	0.34	55	26	20
45°C	Q _{WT}	-6.19	-16.13	0.38	55	17	27
	Q _{AHA}	-4.59	-13.77	0.33	51	49	-
55°C	Q _{WT}	-5.92	-15.93	0.37	69	31	-
	Q _{AHA}	-4.50	-13.43	0.33	49	39	12
65°C	Q _{WT}	-5.89	-15.68	0.38	51	28	21
	Q _{AHA}	-4.54	-13.29	0.34	52	34	14
75°C	Q _{WT}	-5.94	-15.43	0.38	46	18	36
	Q _{AHA}	-4.62	-12.86	0.36	50	25	25
85°C	Q _{WT}	-6.02	-15.24	0.39	54	34	13
	Q _{AHA}	-4.64	-12.51	0.37	48	19	32

Table S4. Mean residue ellipticity values from CD of Q proteins. Mean residue ellipticity values of Q_{WT} and Q_{AHA} pre and post-Dox binding and chemical crosslinking *via* BS³. Data is reported as the average of three independent trials.

	$\Theta \times 10^3 \text{ (deg}\cdot\text{cm}^2\cdot\text{dmol}^{-1}\text{)}$		
	Θ_{222}	Θ_{\min}	$\Theta_{222} / \Theta_{\min}$
Q_{WT}	-7.63	-16.28	0.47
Q_{AHA}	-5.15	-14.25	0.36
Q_{WT}•Dox	-3.50	-8.97	0.39
Q_{AHA}•Dox	-2.64	-9.06	0.30
Q_{WT}•Dox_x	-11.80	-12.16	0.97
Q_{AHA}•Dox_x	-6.87	-8.44	0.81

Table S5. IC₅₀ values of Dox and Q_{AHA}-X-CMms6•Dox treatment of MCF-7 cells *in vitro*. IC₅₀ values calculated from CCK8 cell viability assays of MCF-7 cells treated *in vitro* with Dox and Q_{AHA}-X-CMms6•Dox for 24 hr and 48 hr. Data is reported as the mean value ± standard deviation of three independent trials.

Time	Treatment	IC ₅₀ (μM)
24 hr	Dox	2.92 ± 0.23 (N = 3)
	Q _{AHA} -X-CMms6•Dox	2.74 ± 0.31 (N = 3)
48 hr	Dox	1.28 ± 0.22 (N = 3)
	Q _{AHA} -X-CMms6•Dox	0.48 ± 0.11 (N = 3)

Table S6. Lattice d -spacing of USPIOs. Calculated lattice d -spacing from detectable small angle electron diffraction rings for USPIOs organized by Q_{AHA}-X-CMms6 (Q_{AHA}-X-CMms6•USPIO) compared to that reported for magnetite. Data is reported as the mean value \pm standard deviation of three independent trials.

Ring	Q _{AHA} -CMms6•USPIO d -spacing (Å)	Magnetite (<i>Bragg et al</i>)[1]
1	1.150 \pm 0.00	1.18
2	1.33 \pm 0.00	1.32
3	1.37 \pm 0.00	1.37
4	1.52 \pm 0.00	1.51
5	1.65 \pm 0.00	1.64
6	1.74 \pm 0.00	1.74
7	2.12 \pm 0.01	2.11
8	2.55 \pm 0.00	2.54
9	2.95 \pm 0.02	2.97
10	4.80 \pm 0.04	4.82

1. Bragg, W.H., *The Structure of Magnetite and the Spinels*. Nature, 1915. **95**(2386): p. 561-561.