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Supporting Information

Self-assembled Mg(OH)₂ gels driving to MgO nanoribbons

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SI.1 – Materials, gel preparation and further analysis

Materials

Magnesium nitrate hexahydrate ($Mg(NO_3)_2.6H_2O$, 96 %), magnesium sulphate ($MgSO_4.7H_2O$), isopropanol (99 %), sodium hydroxide (NaOH, 98 %), and glycerol (99.5 %) were purchased from Synth (Brazilian company) and used without further purification. Water was doubly distilled and deionized through a Milli-Q system.

Preparation of Mg hydroxide/oxide nanoparticles in Glycerol Isopropanol solvent – a Survey

Standard gel preparation: firstly, glycerol:isopropanol 3:1 mole ratio solvent (GI) was mixed before salt dissolution. After that 0.1 mol of $Mg(NO_3)_2.6H_2O$ was dissolved into GI (30 mL) at room temperature, and then 0.2mol of NaOH aqueous solution (5mL) was added at once under stirring at 600 rpm. Mixing these reagents a whitish $Mg(OH)_2$ gel was formed almost immediately, Fig. 1a. In order to remove glycerol, isopropanol and residual salts, $Mg(OH)_2$ gel was washed with large amount of deionized water four times under stirring (200 ml in each time), Fig.S1 b-



Figure S1. Mg(OH)₂ gel: (a) as-prepared and (b) centrifuged, and (c) washed and centrifuged.

In addition, at varying concentration of the magnesium salt, while keeping 1:2 salt:base ratio, Mg(OH)₂ gels having different apparent viscosities from stiff to viscous solutions were obtained as amount of reagent decreased as well as distinct translucency, Fig.2 a-c. Notice, MgSO₄.7H₂O also provides a good gel being an alternative salt. However, a better investigation must be made regarding rheological properties and salt interference on gel features and on the MgO nanoparticles characteristics as well.



Figure S2. $Mg(OH)_2$ gel (Mg²⁺ salt mole amount in 30 mL glycerol/isopropanol): (a) sulfate precursor, (b) 2/5 Mg²⁺ nitrate, and (c) 1/5 Mg²⁺nitrate.

Xerogel preparation

Washed Mg(OH)₂ hydrogel was dried in an oven at 70°C for 4 h. A white powder was formed, and PXRD analysis indicates Brucite Mg(OH)₂ structure (space group P-3m1, number 164), see SI-2, Fig.S3.

Lyophilization (freeze-dried)

 $Mg(OH)_2$ hydrogel was frozen for 3 h at -60 °C to then promoting the lyophilization for 15 h at 40 below zero under pressure of 0.09 mbar. A white powder was formed, and PXRD analysis indicates Brucite $Mg(OH)_2$ structure (space group P-3m1, number 164), see SI-2, Fig.S3.

Thermal treatment and infrared

Mg(OH)₂ xerogel and hydrogel were calcinated in an oven at 400°C for 3h under atmospheric pressure in order to obtain MgO nanoparticles (Space group Fm-3m, number 225), see SI-2, Fig.S4.

SI.2 - PXRD analysis

Structural characterization for $Mg(OH)_2$ and MgO are presented on the Table S1 and S2. X'Pert Highscore from PANalytical/Malvern was utilized for the whole profile fitting of the XRD profiles. ICSD database was used for the structural model, the profiles' intensities were simulated with a pseudo-Voigt function without asymmetry and the background was fitted with a 10th

polynomial function. Isotropic temperature factors were considered, and the samples' displacement and absorption were also corrected. Average crystallite size was calculated from Scherrer formula and the lattice microstrain was determined by Stokes-Wilson equation [1-7]. Mg(OH)₂ and MgO diffractograms are presented Fig.S3 and S4.



Figure S3. Mg(OH)₂ structures. Symbols I_{exp} experimental data; I_{calc} calculated profile from Rietveld analysis, I_{diff} difference between calculated and experimental profiles.



Figure S4. MgO structures. Symbols I_{exp} experimental data; I_{calc} calculated profile from Rietveld analysis, I_{diff} difference between calculated and experimental profiles.

Brucite Mg(OH)₂ (space group P-3m1, number 164) is a layered material made of a lattice with a central plane of Mg atoms surrounded by planes of OH groups in the (00l) direction [9], Fig. S5. For Mg(OH)₂ some different strategies were employed in order to fit the whole pattern due to anisotropic broadening of the (00l) peaks. Lyophilized and xerogel Mg(OH)₂ structures stretch the (hk0) planes and give rise to anisotropic crystallites with an similar preferred orientation, though some structural differences may be noted. Different thermal treatments, drying and lyophilization, led to few differences in the lattices: lyophilized Mg(OH)₂ particles have higher lattice volume while those from xerogel have smaller crystallite sizes owing to larger lattice parameters and micro-strains (Table S1).



Figure S5. Mg(OH)₂ and MgO crystal structures (VESTA software) [8].

MgO periclase cubic structure (Space group Fm-3m, number 225) is usually formed by the Mg(OH)₂ decomposition around 300 °C [10-13]. Although MgO particles showed different morphologies their XRD profiles did not show issues regarding anisotropic broadening, but still retaining a small preferred orientation of (hk0) peaks for both of leave- or ribbon-shaped morphologies [9,10]. Lattice strain and volume had very small differences, indicating a robust MgO formation after calcination of the xerogel or hydrogel.

Table S1. X-Ray Diffraction complete results: samples' crystallographic phases, phase fraction,phases' lattices parameters (a,b, c) and cell volume (V). Scherrer average crystallite size $(D_{(hkl)})$ and, lattice microstrain (ϵ).

Sample	Phase	Lattice parameters		۱/ /Å ³ ۱	D _(hkl)	c (%)
	weight (%)	a=b (Å)	c (Å)	V _{cell} (A ⁻)	(nm)	ε(%)
xerogel	100% brucite	3.144(9)	4.672(8)	40.00(4)	8(3)	2.0(7)
hydrogel lyophilized	100% brucite	3.150(11)	4.811(4)	41.33(5)	7.0(3)	1.6(9)
Dried xerogel	100% periclase	a=b=c= 4.2194(6)		75.12(9)	11.0(4)	0.45(4)
Hydrogel calcined	100% periclase	a=b=c= 4.2229(5)		75.30(8)	8.5(7)	0.39(7)

Sample	Rexp	Rp	Rwp	GOF	X ²
xerogel	6.2	12.16	15.16	5.98	2.45
hydrogel lyophilized	5.96	10.05	13.02	4.77	2.18
Dried xerogel	6.96	8.58	11.71	2.83	1.68
Hydrogel calcined	6.98	5.75	7.93	1.29	1.14

 Table S2. Rietveld convergence indices for each sample.

SI-3 - SEM

Mg(OH)₂ particles obtained after lyophilization of the hydrogel prepared directly in water. Individual particles are seen with a very trapezoidal shapes, Fig.S6.



Figure S6. SEM images of the lyophilized Mg(OH)₂ gel.



Figure S7. SEM images of MgO ribbons from calcined hydrogel that was prepared with Mg₂SO₄.

SI-4 - EDS/TEM images

In order to characterize the formation of MgO spheres EDS spectra was collected, Fig. S8.







Figure S8. Top: EDS/TEM measurements: confirmation of MgO spherical particles after sonication. Bottom: grid background EDS image.

SI-5 – Dynamic Light Scattering (DLS) measurements

Sample	sonication time	Average particle size		
	(min)	(nm)		
The same	0	not measured		
sample	20	185 ± 58		
described in fig 5 (main	60 (run 1)	110 ± 30		
article)	60 (run 2)	185 ± 36		

Table S3. DLS results for the MgO nanoribbons after sonication.



Figure S9. Above: MgO before sonication, after 20 min of sonication. Below: DLS after 60 min of sonication, showing the average particle size.

SI-6 – Nitrogen Adsorption Analysis (NAI) measurements

The NAI (Fig. S10) showed an open hysteresis with small pores format [13,14]. The specific surface area after 60 min of sonication was 45 m²/g with adsorbed volume at $P/P_0 = 0.95$ of 0.085 cm³/g. The pore size distribution showed an average pore size of ~20 nm.



Figure S10: Left: Nitrogen adsorption isotherm. Right: Pore size distribution.

Instruments and Methods

The X-ray diffraction (XRD) experiments were carried out with the dried nanoparticles in powder form with a PANalytical Empyrean diffractometer with a PIXEL3D detector, copper tube (λ =1.5418 Å) and a Ni filter. The equipment operated at 40 kV and 30 mA, with 20 from 20° to 70° with a 0.02° step, counting time of 5 s/step. ize the difference between the observed and simulated powder diffraction patterns. The Rietveld R values are used as indicators of the model convergence.

Electron microscopy images were obtained to evaluate nanoparticles size and morphology. Scanning electron microscopy (SEM) images were obtained with a field emission electron microscopy JEOL JSM-7401F with an acceleration tension of 5.0 kV, SEI secondary electron detector, working distance ranging from 3.0 mm and resolution of 1.5 nm and High-resolution transmission electronic microscopy (HRTEM) images were obtained with a JEM-2100-JEOL microscope with 0.23 nm of resolution point with 80 to 200 kV accelerating voltage. For the experiments, 1 mg of hydroxide/oxide magnesium nanoparticles were dispersed in 10 mL of isopropanol by sonication in an ultrasonic bath for 1 h.

Freeze drying Edwards Super Modulyo Edwards super modulyo freeze dryer. DLS experiments were DLS Delsa Nano C Particle Analyzer da Beckman Coulter. DLS sample preparation consisted of isopropanol suspensions.

 N_2 adsorption-desorption measurements were recorded on a Quantachrome Nova Win adsorption analyzer, sample after sonication for 60 min was previously sonicated in isopropanol and dried in a petri dish on a fume hood for 2h. Sample was degassed at 200 °C and isotherms were measured at 77 K. Specific surface area was calculated via Brunauer Emmett Teller (BET) method and the pore size distribution was determined using Density Functional Theory (DFT) method, assuming a slit-shaped pore geometry. The total pore volume (V_{0.98}) was determined from the aggregation of N₂ vapor adsorbed at a relative pressure of 0.99.

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