

## **Electronic Supplementary Information**

### **A novel signal-on fluorometric sensor based on metal ions-mediated carbon dots for formaldehyde determination and lysosome-targeted bioimaging**

Xi Zhou <sup>abc</sup>, Yun Hu <sup>bc</sup>, Yufeng Cao <sup>a</sup>, Yuan Liu <sup>\*a</sup>, Tao Qian <sup>\*a</sup>

a School of Chemistry and Chemical Engineering, Nantong University, Nantong 226019, China

b Institute of Chemical Industry of Forestry Products, Chinese Academy of Forestry, Nanjing  
210042, China

c Co-Innovation Center of Efficient Processing and Utilization of Forest Resources, Nanjing  
Forestry University, Nanjing 210037, China

\* Corresponding author. E-mail address: liuyuan1105@ntu.edu.cn (Yuan Liu);  
qiantao@ntu.edu.cn (Tao Qian)

### **The fluorescence quantum yield measurement**

The fluorescence QY of CDs was estimated by the equation[1]:

$$QY_L = QY_R [(I_L \times A_R)/(I_R \times A_L)] [(N_L)/(N_R)]^2$$

where the subscripts *L* refers to CDs and *R* is the reference, *QY* is the quantum yield (Quinine sulfate in 0.1M H<sub>2</sub>SO<sub>4</sub> solution was chosen as reference, the standard *QY* of which is 54% at 360 nm excitation), *I* refers to the integrated intensity of the emission, *N* represents the refractive index of the medium, *A* is the absorbance at corresponding excitation wavelength.

### **MTT viability assay**

Human uterine cancer cell (HeLa) was seeded into 96-well plates with Dulbecco's modified Eagle's medium (DMEM) for 24 h at a density of 10<sup>4</sup> cells/150 µL in an incubator (37 °C, 5% CO<sub>2</sub>). And the culture medium was replaced by DMEM containing NSCDs and Ag-M-NSCDs, respectively, with different concentrations (0, 0.01, 0.05, 0.1, 0.25, 0.5, 0.8, 1.0 mg/mL) for another 24 h. Then, the cells were washed by 20 mL PBS buffer (pH 7.4) and incubated with MTT solution (5 mg/mL) for 4 h. After that, the culture medium was removed, followed by the addition of 150 µL DMSO and shaken for 10 min at room temperature. The optical density (*OD*) was measured by a microplate reader (ELx800, Biotek, USA) at 490 nm. The cell viability was estimated according to the following equation:

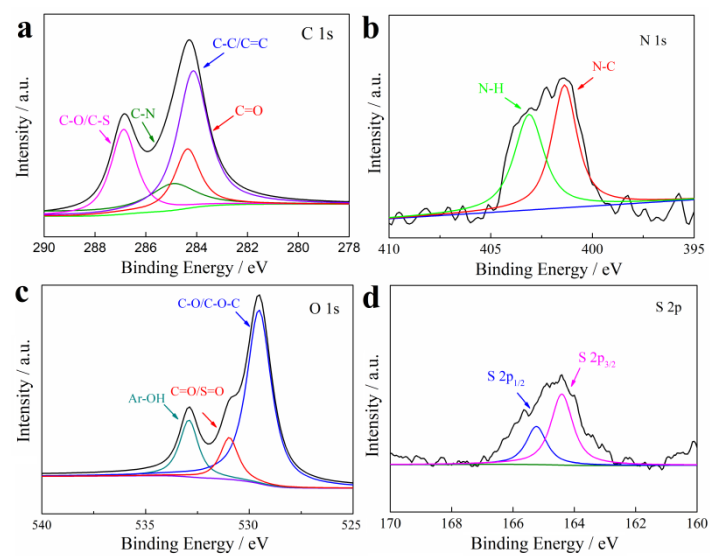
$$\text{Cell viability (\%)} = (OD_{\text{treated}} / OD_{\text{control}}) \times 100\%$$

where *OD*<sub>treated</sub> was obtained in the presence of NSCDs/Ag-M-NSCDs, and *OD*<sub>control</sub> was obtained in the absence of NSCDs/Ag-M-NSCDs.

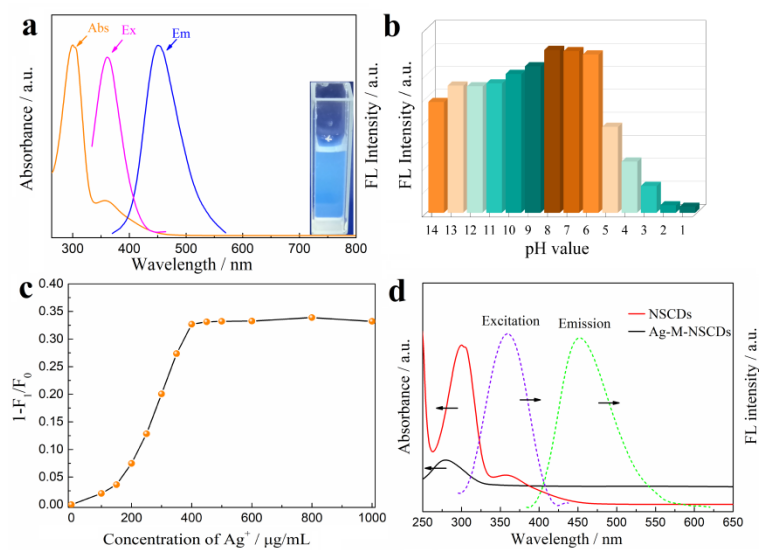
### **Cellular imaging analysis**

The HeLa was first cultured in DMEM containing 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin at 37 °C in 5% CO<sub>2</sub> condition. Before the imaging experiment, all the cells were allowed to adhere to a confocal dish for 12 h. For the fluorescence imaging experiment of Ag-M-NSCDs and formaldehyde (FA), the cells were incubated with 0.1 mg/mL Ag-M-NSCDs for 6 h and washed with a fresh DMEM. Then, 500 µM FA was added for further incubation of 1 h. For the localization experiment, the HeLa cells were incubated with 0.1 mg/mL Ag-M-NSCDs for 6 h and washed three times with PBS buffer. Then 75 nM Lyso-Traker Cys Deep Red was added in serum-free medium and co-cultured with the cells for 1 h. Finally, the residual probe was

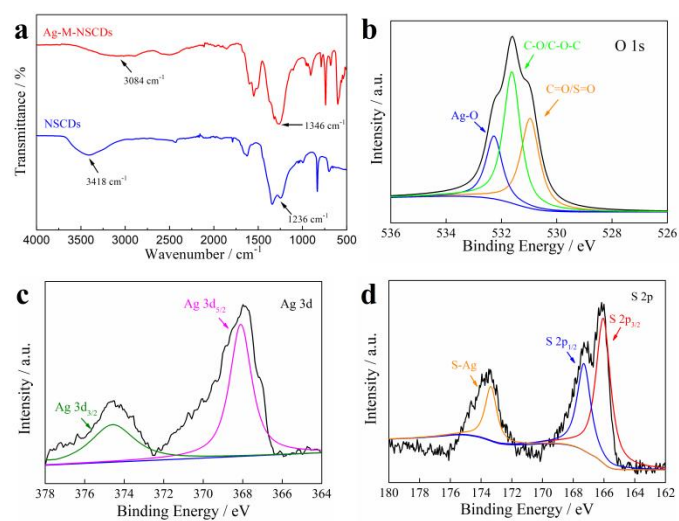
washed three times before imaging under a TCS SP8 (Leica, Germany) confocal microscopy system ( $Ex = 405$  nm; blue channel,  $Em = 420-460$  nm; green channel,  $Em = 520-560$  nm; red channel,  $Ex = 561$  nm,  $Em = 575-630$  nm for Lyso-Traker Cys Deep Red).



**Fig. S1** The high-resolution XPS spectra of (a)C 1s, (b)N 1s, (c) O 1s and (d)S 2p in NSCDs.



**Fig. S2** (a) The UV-vis absorption spectrum, fluorescence excitation and emission spectra of NSCDs. The inset is photograph of NSCDs under UV lamp at 365 nm. (b) Fluorescence intensities of NSCDs versus different pHs. (c) The influence of  $Ag^+$  concentration on the quenching efficiency of Ag-M-NSCDs. (d) The UV-vis absorption spectra of NSCDs, Ag-M-NSCDs and  $AgNO_3$  solution, fluorescence excitation and emission spectra of Ag-M-NSCDs.



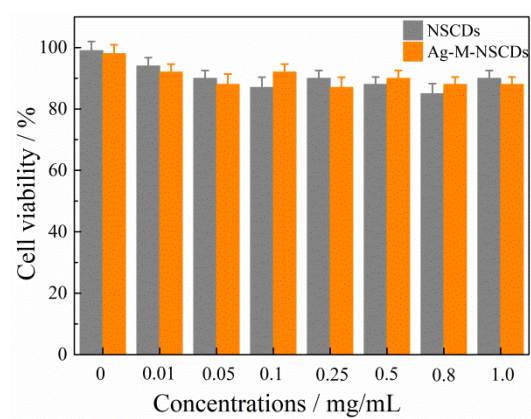
**Fig. S3** (a) The FT-IR spectra of NSCDs and Ag-M-NSCDs, the high-resolution XPS spectra of (b) O 1s, (c) Ag 3d and (d) S 2p in Ag-M-NSCDs.

**Table S1** Determinations of FA in tap water

Sample	Add/ $\mu\text{M}$	Measured/ $\mu\text{M}$		Recovery/%	RSD/%
		HPLC-MS	This work		
1	0.00	6.23	6.75	/	3.22
2	50.00	47.89	48.21	96.42	2.58
3	250.00	257.56	259.25	103.70	5.83
4	500.00	532.22	529.10	105.82	5.11

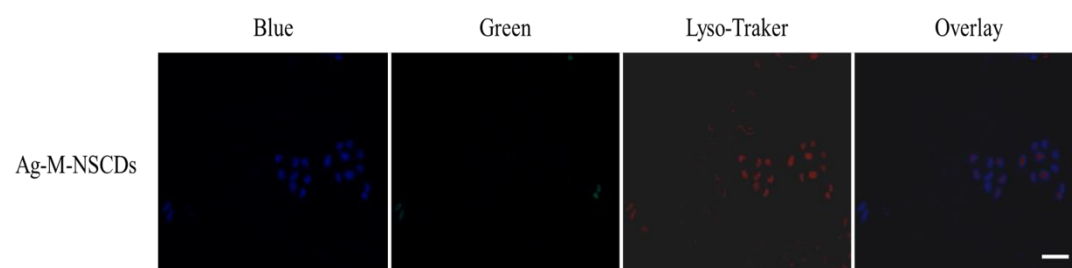
**Table S2** Determinations of FA in PBS

Sample	Add/ $\mu\text{M}$	Measured/ $\mu\text{M}$		Recovery/%	RSD/%
		HPLC-MS	This work		
1	0.00	/	/	/	3.31
2	50.00	44.75	45.89	91.78	4.65
3	250.00	248.21	255.48	102.19	5.47
4	500.00	483.34	524.35	104.87	4.77

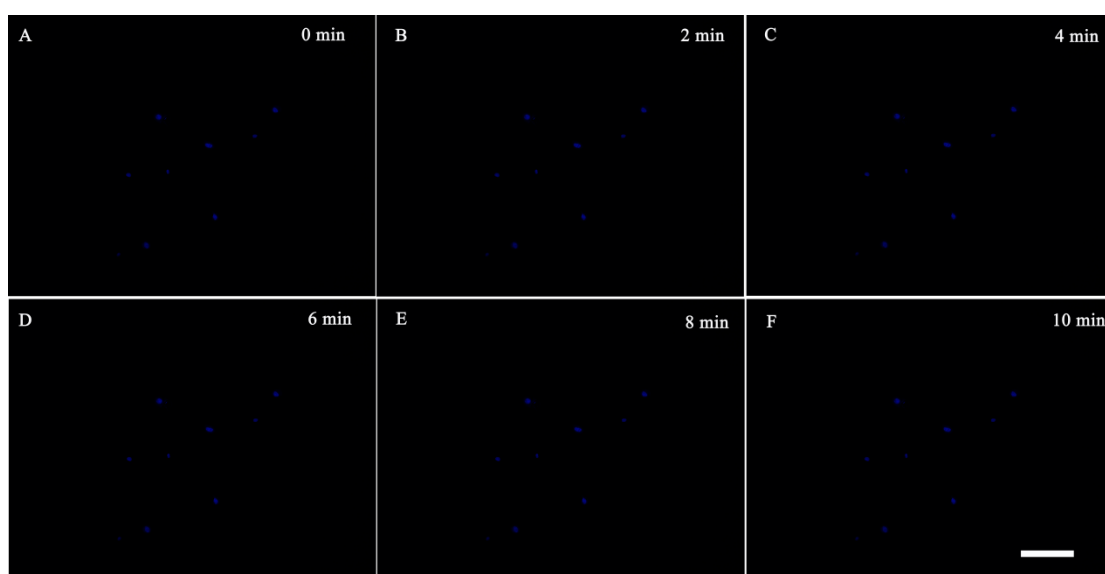


**Fig. S4** Cell viability of HeLa cells after incubated with a series concentrations of NSCDs and Ag-M-NSCDs, respectively.





**Fig. S5** Fluorescence images of HeLa cells incubated with Ag-M-NSCDs and the Lyso-Traker Cys Deep Red.



**Fig. S6** Fluorescence images of HeLa cells after incubation in the medium containing Ag-M-NSCDs and then with a continuous scan of 10 min.

## References

- [1] X.Y. Chai, H. He, H.H. Fan, X.H. Kang and X.P. Song, A hydrothermal-carbonization process for simultaneously production of sugars, graphene quantum dots, and porous carbon from sugarcane bagasse, *Bioresour. Technol.*, 2019, 282, 142-147.