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SUPPLEMENTAL MATERIALS

for

Novel (E)-3-(1-substituted-1H-indazol-5-yl)-N-hydroxypropenamides as Histone Deacetylase Inhibitors: Design, Synthesis and Structure-Activity

Relationships

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S1
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S20

1. Chemistry

Thin layer chromatography which was performed using Whatman[®] 250 µm Silica Gel GF Uniplates and visualized under UV light at 254 and 365 nm, was used to check the progress of reactions and preliminary evaluation of compounds' homogeneity. Melting points were measured using a **Gallenkamp Melting Point Apparatus (LabMerchant,** London, United Kingdom) and are uncorrected. Purification of compounds was carried out using crystallization methods and/or open silica gel column flash chromatography employing Merck silica gel 60 (240 to 400 mesh) as stationary phase. Nuclear magnetic resonance spectra (¹H NMR) were recorded on a Bruker 500 MHz spectrometer with DMSO-*d*₆ as solvent unless otherwise indicated. Tetramethylsilane was used as an internal standard. Chemical shifts are reported in parts per million (ppm), downfield from tetramethylsilane. Mass spectra with different ionization modes including electron ionization (EI), Electrospray ionization (ESI), were recorded using PE Biosystems API2000 (Perkin Elmer, Palo Alto, CA, USA) and Mariner® (Azco Biotech, Inc. **Oceanside, CA, USA)** mass spectrometers, respectively. The elemental (C, H, N) analyses were performed on a Perkin Elmer model 2400 elemental analyzer. All reagents and solvents were purchased from Aldrich or Fluka Chemical Corp. (Milwaukee, WI, USA) or Merck unless noted otherwise. Solvents were used directly as purchased unless otherwise indicated.

2. Biology

2.1. Cytotoxicity assay

The cytotoxicity of the synthesized compounds was evaluated against three human cancer cell lines, including SW620 (colon cancer), PC3 (prostate cancer), and NCI-H23 (lung cancer). The cell lines were purchased from a Cancer Cell Bank at the Korea Research Institute of Bioscience and Biotechnology (KRIBB). The media, sera and other reagents that were used for cell culture in this assay were obtained from GIBCO Co. Ltd. (Grand Island, New York, USA). The cells were culture in DMEM (Dulbecco's Modified Eagle Medium) until confluence. The cells were then trypsinized and suspended at 3×10^4 cells/mL of cell culture medium. On day 0, each well of the 96-well plates was seeded with 180 µL of cell suspension. The plates were then incubated in a 5% CO₂ incubator at 37 °C for 24 h. Compounds were initially dissolved in dimethyl sulfoxide (DMSO) and diluted to appropriate concentrations by culture medium. Then 20 µL of each compounds' samples, which were prepared as described above, were added to each well of the 96-well plates, which had been seeded with cell suspension and incubated for 24-h, at various concentrations. The plates were further incubated for 48 h. Cytotoxicity of the compounds was measured by the colorimetric method, as described previously ^[34] with slight modifications.^[35-37] The IC₅₀ values were calculated using a Probits method ^[38] and were averages of three independent determinations (SD ≤ 10%).

2.2. HDAC enzymes assay

The HDAC enzymes (Hela cell nuclear extract) were purchased from Enzo Life Sciences Inc. (Farmingdale, New York, USA). The HDAC enzymatic assay was performed using a Fluorogenic HDAC Assay Kit (Enzo Life Sciences Inc.) according to the manufacturer's instructions. The testing protocol was similar to that previously reported elsewhere.^[39-40] Briefly, HDAC enzymes were incubated with vehicle or various concentrations of the assayed samples or SAHA for 30 min at 37°C in the presence of an HDAC fluorimetric substrate. The HDAC assay developer (which produces a fluorophore in reaction mixture) was added, and the fluorescence was measured using VICTOR (PerkinElmer, Waltham, MA, USA) with excitation at 360 nm and emission at 460 nm. The measured activities were subtracted by the vehicle-treated control enzyme activities and IC₅₀ values were calculated using GraphPad Prism (GraphPad Software, San Diego, CA, USA).

3. ALL ¹H & ¹³C NMR SPECTRA OF THE COMPOUNDS

¹H NMR of compound 5a



¹³C NMR of compound 5a



¹H NMR of compound 5b



¹³C NMR of compound 5b



¹H NMR of compound 5c



¹³C NMR of compound 5c



¹H NMR of compound 5d



¹³C NMR of compound 5d



¹H NMR of compound 5e



¹H NMR of compound 5f



¹³C NMR of compound 5f



¹H NMR of compound 5g



¹H NMR of compound 5h



¹³C NMR of compound 5h



¹H NMR of compound 5i



¹³C NMR of compound 5i



¹H NMR of compound 7a



¹H NMR of compound 7b



¹³C NMR of compound 7b



¹H NMR of compound 7c



¹³C NMR of compound 7c



¹H NMR of compound 7d



¹³C NMR of compound 7d



¹H NMR of compound 7e



¹H NMR of compound 7f



¹³C NMR of compound 7f



¹H NMR of compound 7g





¹H NMR of compound 7h



¹³C NMR of compound 7h



¹H NMR of compound 7i



¹³C NMR of compound 7i



4. 2D NMR for representative final compound. HMBC of 5a







