

**Fluorescently-Tagged Magnetic Protein Nanoparticles for High-Resolution Optical and Ultra-High
Field Magnetic Resonance Dual-Modal Cerebral Angiography**

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Table ST1. Magnetic Properties of NPs measured by SQUID at 1.5T applied magnetic field.

	Magnetic moment (emu/g)	Coercivity (Oe)	Retentivity (emu/g)
c-FeMNPs	0.135	34.62	9.4×10^{-4}
f-MPNPs	0.085	26.51	11.2×10^{-4}

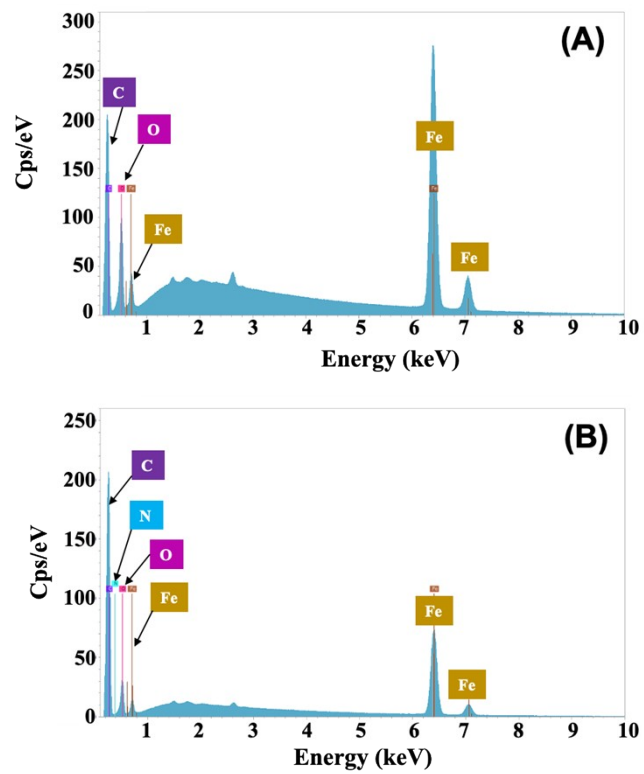


Figure S1. EDX survey spectra of magnetic NPs showing elemental distributions. (A) c-FeMNPs and (B) f-MPNPs.

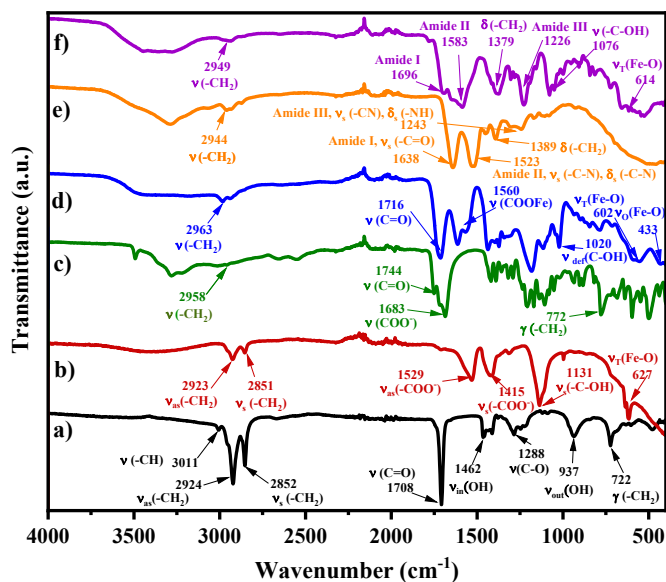


Figure S2. FTIR spectra of MNPs as well as some intermediates (with glossary of terms: ν = Stretching vibration; δ = Bending vibration; γ = Wagging, twisting, rocking; **def** = Deformation; **as** = Asymmetric; **s** = Symmetric).

a) Pristine oleic acid; $-\text{CH}_2$ asymmetric stretching vibration at 2924 cm^{-1} and symmetric vibration at 2852 cm^{-1} , and stretching vibration of $\text{C}=\text{O}$ (of prominent $-\text{COOH}$ group), and in-plane stretching vibration of $-\text{OH}$ at 1462 cm^{-1} and out-plane stretching vibration of $-\text{OH}$ at 1462 cm^{-1} are characteristic peaks of oleic acid. **b)** o-FeMNPs; after several washes and centrifugation to ensure excess oleic acid and oleyl alcohol are removed, the FTIR spectrum of oleic acid coated FeMNPs (o-FeMNPs) reveals additional peaks of $-\text{COO}^-$ asymmetric stretching vibration at 2924 cm^{-1} and symmetric vibration at 2852 cm^{-1} , and high intensity symmetric vibration of $-\text{C-OH}$, and symmetric vibration for tetragonal Fe-O confirming oleic acid coating on the surface of FeMNPs. **c)** Pristine sodium citrate; stretching vibration ($-\text{C}=\text{O}$) at 1744 cm^{-1} , and stretching vibration ($-\text{COO}^-$) at 1683 cm^{-1} , and rocking vibration ($-\text{CH}_2$) are characteristic peaks of sodium citrate. **d)** c-FeMNPs; citric acid coated FeMNPs reveals shift in stretching vibration ($-\text{C}=\text{O}$) to 1716 cm^{-1} , and shift in stretching vibration of ($-\text{COO}^-$) to 1560 cm^{-1} after interaction with Fe(III) ions on the nanoparticle surface. Appearance of additional peaks deformation vibration of ($-\text{C-OH}$), stretching vibration of tetrahedral Fe-O at 602 cm^{-1} and octahedral Fe-O at 433 cm^{-1} confirms citric acid surface modification on FeMNPs devoid of free citrate. **e)** Pristine bovine serum albumin (the model protein); characteristic peaks of the protein are Amide I at 1638 cm^{-1} , Amide II at 1523 cm^{-1} and Amide III at 1243 cm^{-1} . **f)** MNPs; protein conjugation with c-FeMNPs shows shift in Amide I to 1696 cm^{-1} , Amide II to 1583 cm^{-1} and Amide III to 1226 cm^{-1} , and additional peaks of stretching vibration (C-OH) at 1076 cm^{-1} and of stretching vibration of tetrahedral Fe-O peak at 614 cm^{-1} confirm the covalent conjugation of c-FeMNPs and protein molecules.

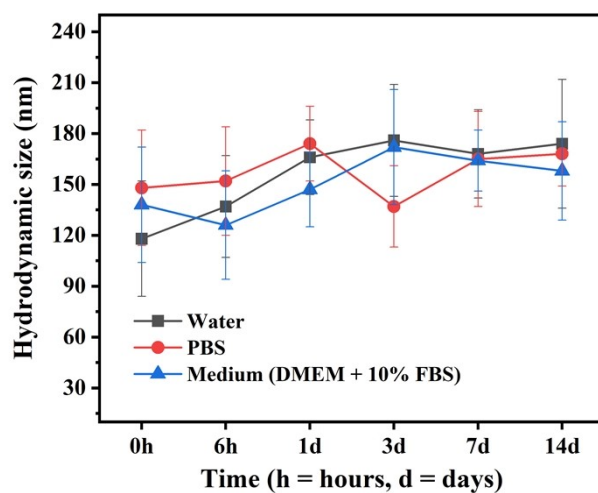


Figure S3. Colloidal stability of f-MPNPs. Colloidal stability was analyzed in water, PBS, and cell culture media (DMEM + 10% FBS). A known concentration of NPs was prepared in water, PBS, and DMEM media and stored at 25 °C. (n = 5 for each media) The hydrodynamic size was measured at predetermined time intervals for a 14-day study period.

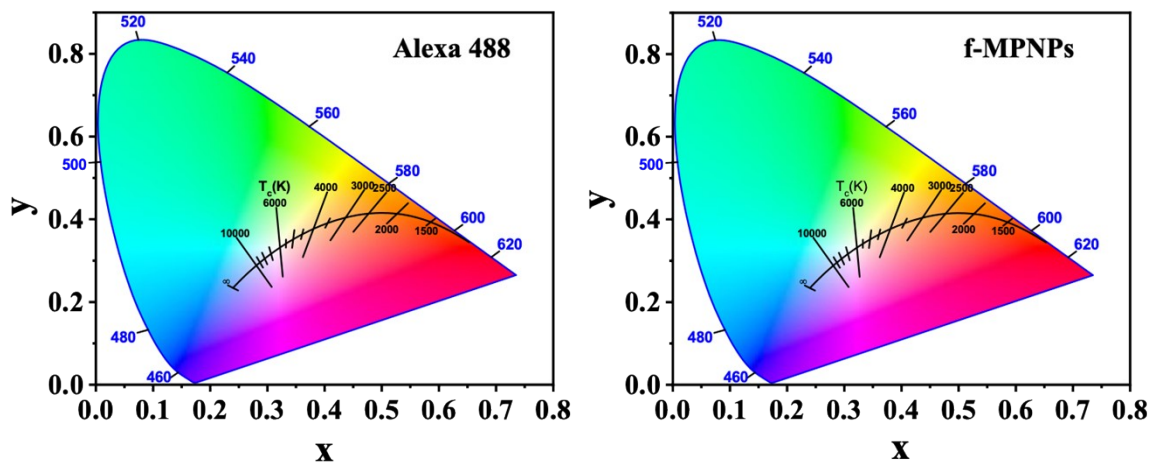


Figure S4. CIE fluorescent plot of fluorescent dye with and without conjugation; **(A)** Alexa Fluor 488 dye and **(B)** Alexa Fluor 488 conjugated MPNPs (f-MPNPs).

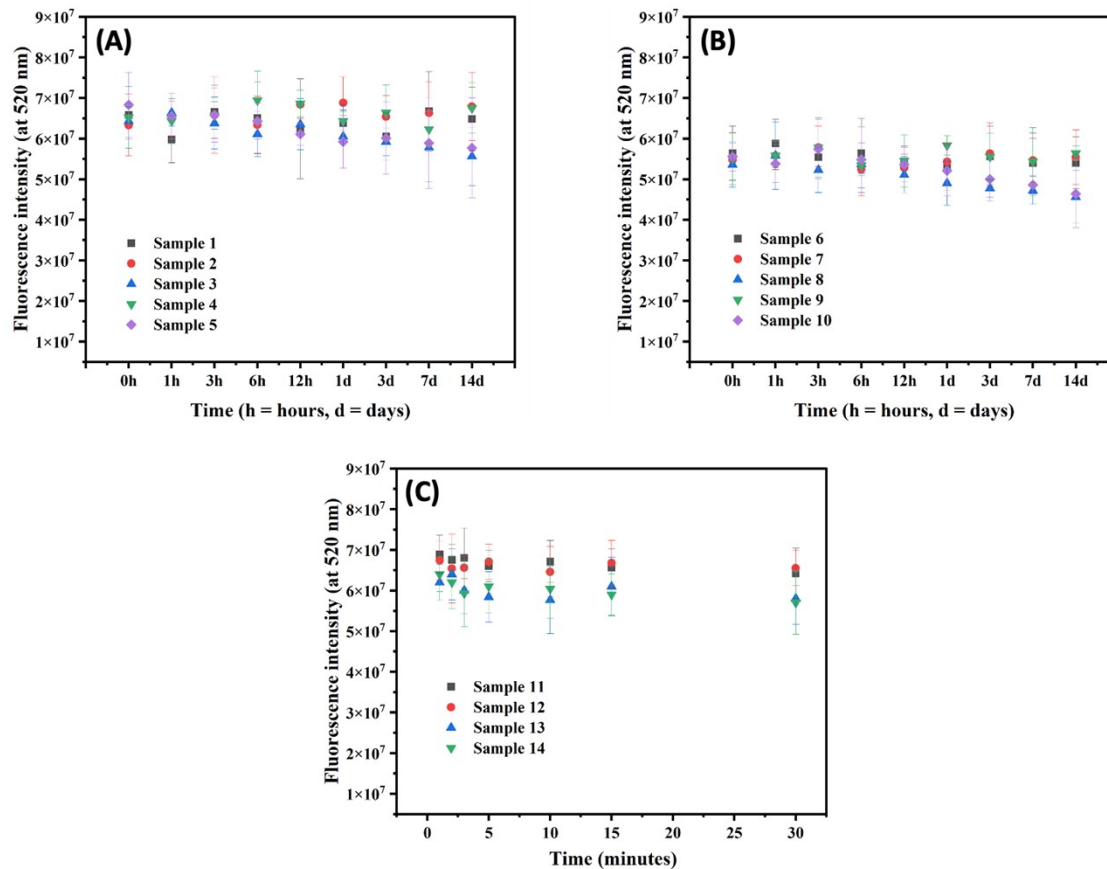


Figure S5. Photostability of f-MPNPs. The photostability of f-MPNPs was evaluated at different conditions. The samples were preconditioned at various conditions (**Solvent/Temperature/Light**) and then photostability tests were performed by recording the fluorescence emission at 520 nm by excitation at 488 nm wavelength. Two solvents were selected: PBS and rat blood plasma (Plasma). Three light conditions were applied: dark, bright light (daylight lamp) and illumination under LED light source of wavelength 488 nm. We selected three temperature conditions: 4°C (storage temperature), 25°C (room temperature) and 37°C (body temperature). **(A)** f-MPNPs were dissolved in PBS and different conditions were applied; **Sample 1** - PBS/4°C/Dark; **Sample 2** - PBS/25°C/Dark, **Sample 3** - PBS/37°C/Dark, **Sample 4** - PBS/25°C/Bright, and **Sample 5** - PBS/37°C/Bright. **(B)** f-MPNPs were dissolved in plasma and different conditions were applied; **Sample 6** - Plasma/4°C/Dark, **Sample 7** - Plasma/25°C/Dark, **Sample 8** - Plasma/37°C/Dark, **Sample 9** - Plasma/25°C/Bright, and **Sample 10** - Plasma/37°C/Bright. **(C)** f-MPNPs solutions were irradiated with 488 nm LED light for definite time: **Sample 11** - PBS/25°C/LED, **Sample 12** - PBS/37°C/LED, **Sample 13** - Plasma/25°C/LED, and **Sample 14** - Plasma/37°C/LED. (n = 5 for each sample)

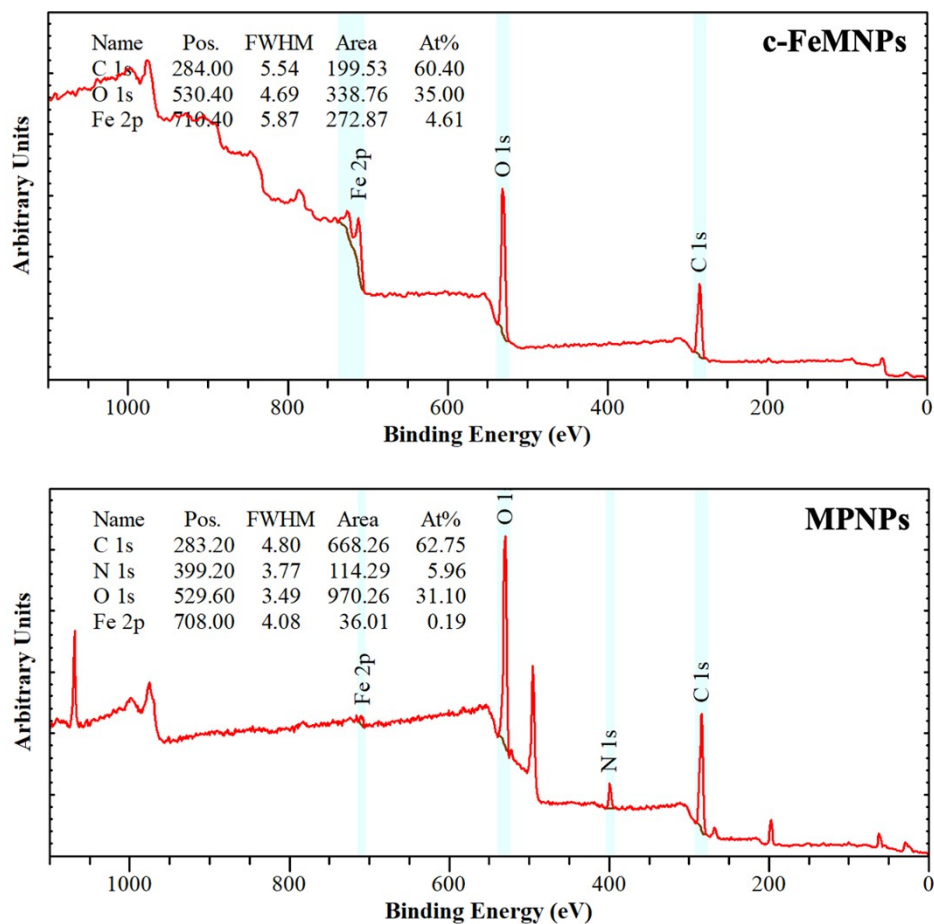


Figure S6. XPS survey spectra of c-FeMNPs and MPNPs; (A) c-FeMNPs, reveals elemental composition as C 1s, O 1s and Fe 2p, and (B) MPNPs, reveals elemental composition as C 1s, O 1s, Fe 2p and N 1s peaks (some additional peaks denote Na and Cl elements from HCl, NaOH and saline solution).

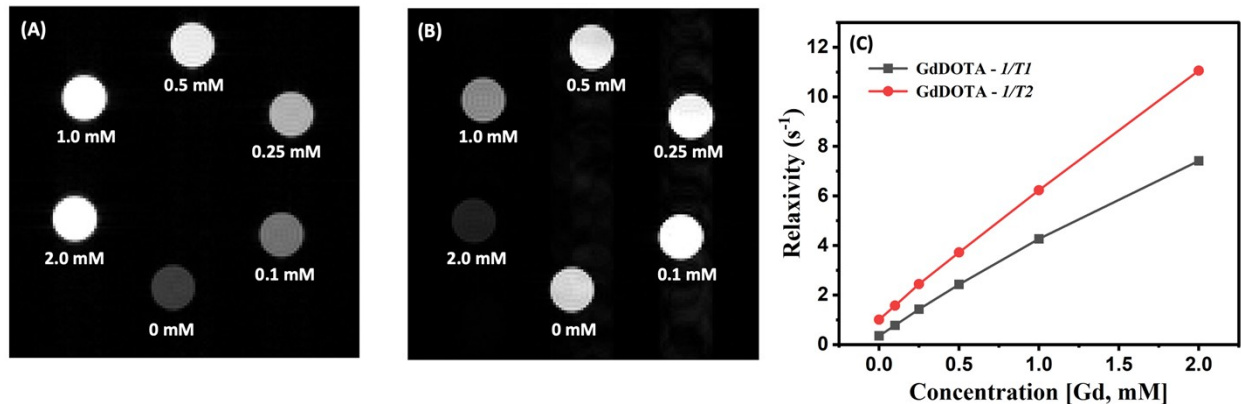


Figure S7. Evaluation of MRI properties of GdDOTA. (A) T1 weighted ($T1w$) (B) T2 weighted ($T2w$) images of GdDOTA for different Gd concentrations. (C) Relaxivity evaluation of Gd concentration; plots of GdDOTA; $1/T1$ (black ■), $1/T2$ (red ●). The slope indicates the specific relaxivity (r). The series of concentrations of GdDOTA reveals longitudinal relaxivity (r_1) = 4.26, transverse relaxivity (r_2) = 6.23, and the $r_2/r_1 = 1.46$, at 11.7T.

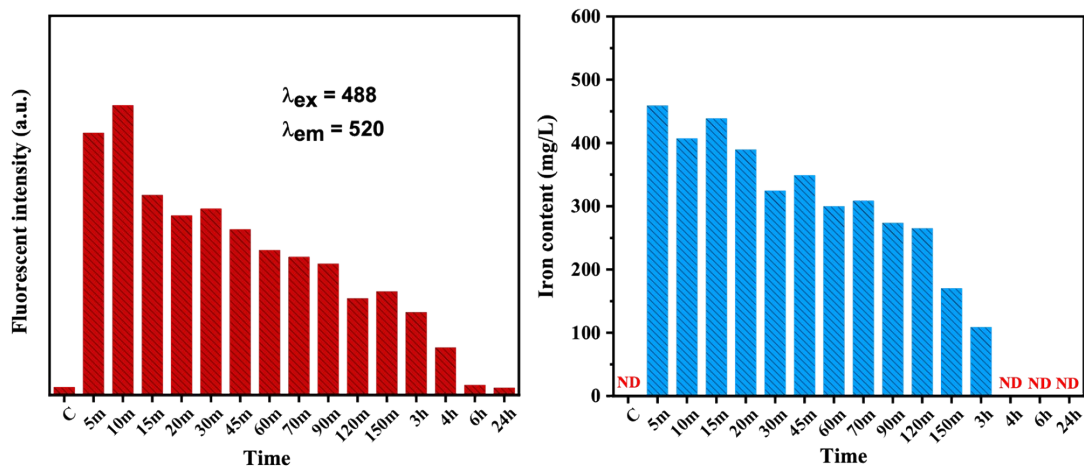


Figure S8. Blood half-life and iron content of MNPs. **(A)** Blood half-life of f-MPNPs estimated by measuring fluorescence intensity ($\lambda_{ex} = 488$, $\lambda_{em} = 520$) by plate reader and **(B)** iron content in blood plasma by ICP-MS. Both methods reveal a circulation half-life of 90-100 minutes from rat blood samples.

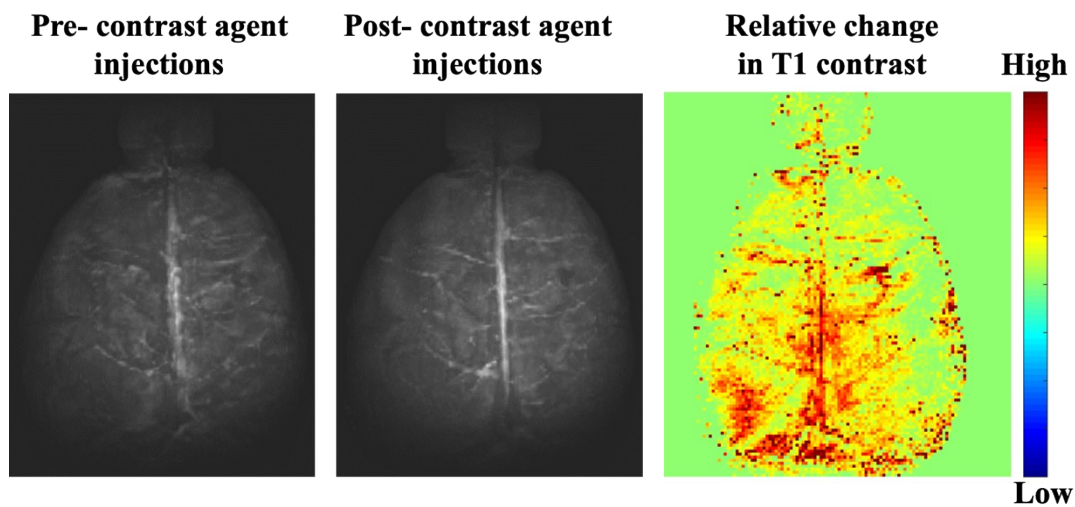


Figure S9. Cerebral angiography by real-time in vivo MRA of rats. Data from rat brain treated with f-MPNPs demonstrates enhanced 30-40% T1 contrast enhancement using 3D-FLASH sequence. (Scale = low to high represents 0 to 100 % arbitrary unit)

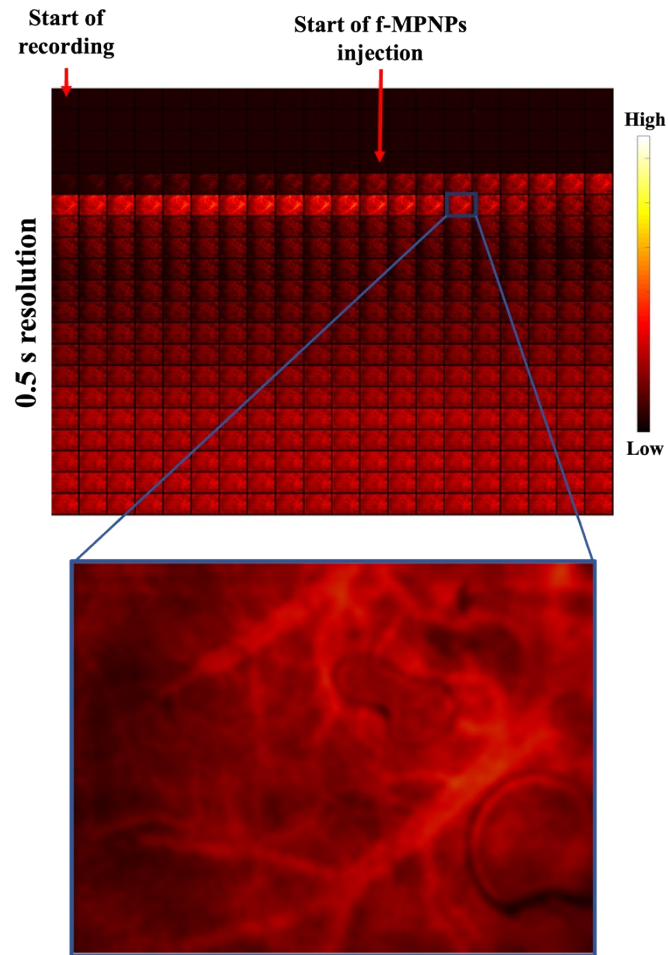


Figure S10. Cerebral angiography by real-time in vivo fluorescence imaging of rat brain after intravenously injection with f-MPNPs (image resolution = 0.5s). Scale = 1mm. (Scale = low to high represents 0 to 100 % arbitrary unit)

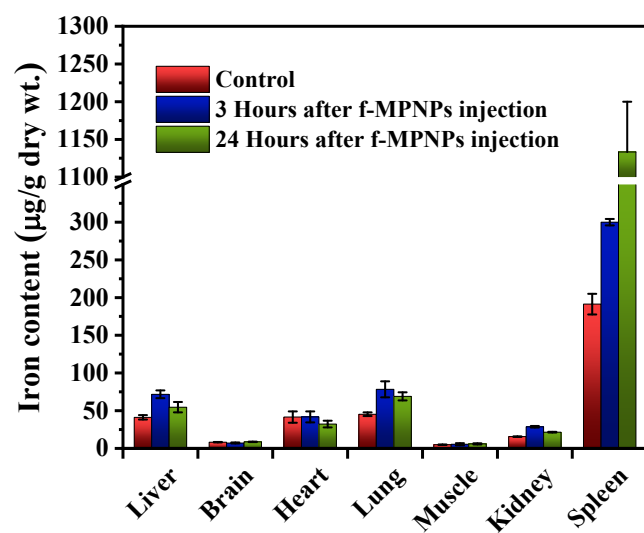


Figure S11. Biodistribution pattern for 3h and 24 h after intravenous injection of the f-MPNPs in rats.