

Supporting information for:

Access to valuable building blocks by selective ring-opening of itaconic anhydride using lipase catalysis.

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Table of Contents:

1 Materials and methods	S2
2 Synthesis of racemic alcohols [19, 22-25]	S2
3 <i>PCL</i> -catalyzed regioselective ring-opening of IAn using several primary alcohols	S4
4 Crystallographic data and structure refinement details for monoesters itaconates: 2e, 6e and 9e.	S7
5 <i>PCL</i> -catalyzed regioselective and enantioselective opening ring of (IAn) using a set of arylalkyl carbinols as chiral nucleophiles	S7
6 NMR spectra of racemic alcohols [19, 22-25]	S9
7 NMR spectra of all the monoesters itaconates	S14
8 Chromatograms of <i>PCL</i> -catalyzed regio and enantioselective ring-opening of chiral secondary alcohol	S31

1. Materials and methods:

1.1. Chemicals and Materials

All reagents purchased from: Aldrich, were used as received. Alcohols [1-18] and [20-21] are commercially available. All used lipases are commercially available and used as purchased without any pre-treatment Lipase from *pseudomonas cepacia* (*Amano Lipase Ps, from burkhoderia cepacia*) (*PCL*; Specific activity up to 30000 U/g), *Candida rugosa* lipase (*CRL*; LA= 1170 U/mg) and the *Candida antarctica* lipase fraction B immobilized on acrylic resin (*CAL-B*; LA > 10000 U/g). Reactions were monitored by thin-layer chromatography (TLC) carried out on Silica gel 60F₂₅₄ plates type *MERCK* 5179, 250 mesh, using UV light (254 nm) the visualizing agent sing ultraviolet light (254 nm) as the visualizing agent and KMnO₄ solution as developing agents. The separation of the resulting alcohols and the remaining acetates was performed by liquid-liquid extraction.

1.2. Instrumentations

NMR spectra were recorded on Bruker spectrometers (300 MHz for ¹H, 75 MHz for ¹³C) instrument and calibrated using residual deuterated solvent as an internal reference (peak at 7.26 ppm in ¹H NMR and 3 peaks at 77 ppm in ¹³C NMR in the case of CDCl₃ and DMSO. The following abbreviations were used to designate multiplicities: s=singlet, d=doublet, t=triplet, q=quartet, m=multiplet, br: broad signal, dd=double-doublet. Chemical shifts were expressed in ppm and coupling constant (*J*) in Hz. The enantiomeric excesses were measured by a chiral stationary phase HPLC: *Chiralpak OJ-H*, *Chiralpak IA* or *Chiralpak IB* columns (4.6 × 250 mm). Retention times are reported in minutes. Mass spectra were taken by a MicrOTOF-Q Bruker spectrometer using electrospray ionization (ESI) analysis.

2. Synthesis of racemic alcohols [19, 22-25]:

The racemic alcohols [19, 22-25] were prepared from the corresponding commercial ketones using the following procedure: to solution of THF / H₂O (4/1, v/v), 6 equivalent of sodium borohydride (18 mmol) are added, and stirred at 0°C. 3 mmoles of the corresponding alcohol diluted in 20 mL of THF are added by drops. After consumption the ketone, the mixture was neutralized by addition of a solution of (HCl, 1N). Then the organic layer was removed, and the aqueous layer was extracted with ethyl acetate (3 x 80 mL). The combined organic layers were dried over anhydrous MgSO₄ and concentrated to give the corresponding alcohols with good yields. The ¹H and ¹³C NMR spectra of these products were in good agreement with the literature.

1,2-dihydroacenaphthylen-1-ol (rac-19)

¹H NMR (300 MHz, CDCl₃, 25 °C) δ 2 (s, OH), 3.26 (d, *J* = 17.7Hz, 1H), 3.82 (dd, *J* = 17.7, 7.1Hz, 1H), 5.74 (dd, *J* = 7.1, 2.1Hz, 1H), 7.32- 7.79 (m, H_{arm}). **¹³C NMR (75 MHz, CDCl₃)** δ 41.9, 74.4, 119.8, 120.3, 122.7, 125, 128, 128.5, 131.2, 137.1, 141.5, 145.7.

Chiral HPLC: Chiralpak IA. **Eluent (v,v)** : Hexane/ iPrOH. **Flow rate** : 0.5 mL/min. **Retention time** : t_R = 25.81 min, t_S = 27.71 min.

6-methyl-2,3-dihydro-1H-inden-1-ol (rac-22)

¹H NMR (300 MHz, CDCl₃, 25 °C) δ 1.95 (m, 1H), 2.2 (s, OH), 2.41 (s, 3H), 2.49 (m, 1H), 2.79 (m, 1H), 3.03 (m, 1H); 5.21 (t, 1H), 7.09- 7.25 (m, H_{arm}). **¹³C NMR (75 MHz, CDCl₃)** δ 21.2, 29.3, 36.1, 77, 124.6, 124.7, 129.1, 136.3, 140.2, 145.1.

Chiral HPLC: Chiralpak IA. **Eluent (v,v)** : Hexane/ iPrOH. **Flow rate:** 0.5 mL/min. **Retention time** : t_S = 17.63 min, t_R = 18.96 min.

6-methoxy-2,3-dihydro-1H-inden-1-ol (rac-23)

¹H NMR (300 MHz, CDCl₃, 25 °C) δ 1.71 (s, OH), 1.96 (m, 1H), 2.5 (m, 1H), 2.77 (m, 1H), 3 (m, 1H), 3.83 (s, 3H), 5.23 (t, *J* = 6.2 Hz, 1H), 6.83-7.15(m, H_{arm}). **¹³C NMR (75 MHz, CDCl₃)** δ 28.9, 36.6, 55.5, 77.4, 108.8, 115, 125.5, 135, 146.3, 159.

Chiral HPLC: Chiralpak IA. **Eluent (v,v)** : Hexane/ iPrOH. **Flow rate:** 0.5 mL/min. **Retention time** : t_S = 30.60 min, t_R = 32.87 min.

6-chloro-2,3-dihydro-1H-inden-1-ol (rac-24)

¹H NMR (300 MHz, CDCl₃, 25 °C) δ 1.94 (m, 1H), 2.3 (s, OH), 2.5 (m, 1H), 2.77 (m, 1H), 3 (m, 1H), 5.19 (t, *J* = 6.3Hz, 1H), 7.15- 7.36 (m, H_{arm}). **¹³C NMR (75 MHz, CDCl₃)** δ 29.2 ; 36.2, 76, 124.4, 125.9, 128.3, 132.3, 141.6, 146.9.

Chiral HPLC: Chiralpak IA. **Eluent (v,v)** : Hexane/ iPrOH. **Flow rate:** 0.5 mL/min. **Retention time:** t_S = 18.55 min, t_R = 21.85 min.

6-(trifluoromethyl)-2,3-dihydro-1H-inden-1-ol (rac-25)

¹H NMR (300 MHz, CDCl₃, 25 °C) δ 1.98 (m, 1H), 2.2 (s, OH), 2.5 (m, 1H), 3.03 (m, 2H), 5.26 (t, *J* = 6.3Hz, 1H), 7.34- 7.67 (m, H_{arm}). **¹³C NMR (75 MHz, CDCl₃)** δ 29.7, 35.9, 75.8, 121.2, 121.3, 125.2, 125.32, 125.37, 145.6, 147.3.

Chiral HPLC: Chiralpak IA. **Eluent (v,v)** : Hexane/ iPrOH. **Flow rate:** 0.5 mL/min. **Retention time:** t_S = 14.35 min, t_R = 16.31 min.

3. PCL-catalyzed regioselective ring-opening of IAn using several primary alcohols:

To equimolecular mixture of itaconic anhydride and the appropriate alcohol [1-15] diluted in 5 mL of TBME, the adequate amount of lipase is added. After 24 hours of stirring at room temperature, the lipase was removed by filtration. The no reacted alcohol was removed by liquid-liquid extraction. The obtained monoesters recovered pure after re-crystallization. Complete experimental data have been provided (NMR spectra and HRMS).

4-ethoxy-2-methylene-4-oxobutanoic acid (2e). White crystalline. mp 45 °C. ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 09.88 (s, 1H, OH), 6.48 (s, 1H, C=CH₂), 5.84 (s, 1H, C=CH₂), 4.18 (q, 2H, J=7.1 Hz, CH₂CH₃), 3.35 (s, 1H, CH₂-C=), 1.27 (t, 3H, J=7.1 Hz, CH₂CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 171.5, 170.6, 133.3, 130.78, 61.08, 37.36, 14.1. HRMS (ESI) m/z calcd for C₇H₁₀O₄ [M + Na⁺]:181.04, Found: 181.048.

4-isobutoxy-2-methylene-4-oxobutanoic acid (3e). White crystalline. mp 47 °C. ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 11.2 (s, 1H, OH), 6.46 (s, 1H, C=CH₂), 5.83 (s, 1H, C=CH₂), 3.88 (d, 2H, J = 6.7 Hz, O-CH₂CH), 3.35 (s, 2H, OOC-CH₂-C=), 1.93 (m, 1H), 0.91 (d, 6H, J = 6.7 Hz, 2CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 171.7, 170.6, 133.4, 130.6, 71.1, 37.3, 27.6, 18.9. HRMS (ESI) m/z calcd for C₉H₁₄O₄ [M + Na⁺]: 209.19, Found: 209.0792.

4-(3- Methylbutanoxy)-2-methylene-4-oxobutanoic acid (1e). Gummy liquid · ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 10 (s, 1H, OH), 6.45 (s, 1H, C=CH₂), 5.82 (s, 1H, C=CH₂), 4.13 (d, J = 6.8 Hz, 2H, O-CH₂CH), 3.34 (d, 2H, J = 5.4 Hz, OOC-CH₂-C=), 1.67 (m, 1H, CH₂-CH(CH₃)₂), 1.51 (q, 2H, J = 13.5, 6.7 Hz, CH-CH₂-CH₂), 0.90 (d, 6H, J = 6.5 Hz, CH₃-CH-CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 170.7, 133.4, 130.5, 63.7, 37.3, 37.1, 34.04, 24.9, 22.3. 16.2, 11.1 HRMS (ESI) m/z calcd for C₁₀H₁₆O₄ [M + Na⁺]:223.08, Found: 223.095.

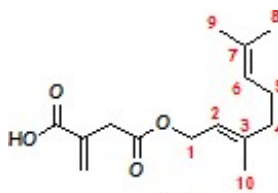
4-(hexyloxy)-2-methylene-4-oxobutanoic acid (4e). White crystalline. mp 42 °C. ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 09 (s, 1H, OH), 6.45 (s, 1H, C=CH₂), 5.82 (s, 1H, C=CH₂), 4.15 (t, 2H, J = 6.7 Hz, O-CH₂CH₂), 3.34 (s, 2H, OOC-CH₂-C=), 1.36 (m, 2H, O-CH₂-CH₂-(CH₂)₃-CH₃), 1.28 (m, 6H, CH₂-(CH₂)₃-CH₃), 0.90 (m, 3H, CH₂CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 170.7, 133.4, 130.5, 65.2, 37.3, 31.3, 28.4, 25.4, 22.4, 13.9. HRMS (ESI) m/z calcd for C₁₁H₁₈O₄ [M + Na⁺]:237.100, Found: 237.1102.

4-(heptyloxy)-2-methylene-4-oxobutanoic acid (5e). White crystalline. mp 53 °C. ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 8.3 (s, 1H, OH), 6.47 (s, 1H, C=CH₂), 5.84 (s, 1H, C=CH₂), 4.11 (t, 2H, J = 6.7 Hz, O-CH₂CH₂), 3.35 (s, 2H, CH₂-C=), 1.6 (m, 2H, CH₂-CH₂-O), 1.30 (m, 8H, O-CH₂-(CH₂)₄-CH₃), 0.89 (t, 3H, J = 9.2 Hz, CH₂CH₃). ¹³C NMR (75 MHz, CDCl₃) δ 171.5, 170.6, 133.3, 130.6, 65.2,

37.3, 31.7, 28.8, 28.5, 25.7, 22.5, 14.07. HRMS (ESI) m/z calcd for $C_{12}H_{20}O_4$ $[M + Na^+]$: 251.120, Found: 251.259.

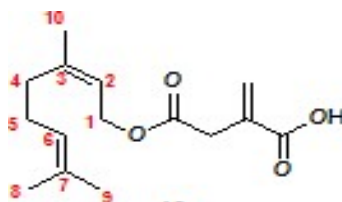
4-(decyloxy)-2-methylene-4-oxobutanoic acid (6e). White crystalline. mp 67 °C. 1H NMR (300 MHz, $CDCl_3$, 25 °C) δ 6.48 (s, 1H, $C=CH_2$), 5.84 (s, 1H, $C=CH_2$), 4.12 (t, 2H, $J = 6.7$ Hz, $O-CH_2CH_2$), 3.35 (s, 2H, $OOCCH_2-C=$), 1.6 (m, 2H, CH_2-CH_2-O), 1.30 (m, 14H, $OCH_2-CH_2-(CH_2)-CH_3$), 0.8 (t, 3H, $J = 6.7$ Hz, CH_2CH_3). ^{13}C NMR (75 MHz, $CDCl_3$) δ 171.3, 170.6, 133.3, 130.5, 65.2, 37.3, 31.8, 29.5, 29.4, 29.29, 29.21, 25.8, 22.6, 14.09. HRMS (ESI) m/z calcd for $C_{15}H_{26}O_4$ $[M + Na^+]$: 293.16, Found: 293.1730.

(E)-4-((3,7-dimethylocta-2,6-dien-1-yl)oxy)-2-methylene-4-oxobutanoic acid (Mono 4-geranyl Itaconate)(7e) :



Gummy liquid. 1H NMR (300 MHz, $CDCl_3$, 25 °C) δ 9.2 (s, 1H, OH), 6.48 (s, 1H, $C=CH_2$), 5.84 (s, 1H, $C=CH_2$), 5.35 (m, 1H, $CH=C^2H-CH_2$), 5.33 (m, 1H, $C=C^6H-CH_2$), 4.64 (d, 2H, $J = 7.1$ Hz, $O-C^1H_2CH$), 3.35 (s, 2H, $CH_2-C=$), 2(m, 4H, $C^4H_2-C^5H_2$), 1.7 (s, 6H, C^9H_3 , $C^{10}H_3$), 1.61 (s, 3H, $=CC^8H_3$). ^{13}C NMR (75 MHz, $CDCl_3$) δ 170.7, 170.6, 142.5, 133.4, 131.8, 130.4, 123.7, 117.9, 61.9, 39.5, 37.3, 26.2, 25.6, 17.6, 16.4. HRMS (ESI) m/z calcd for $C_{15}H_{22}O_4$ $[M + Na^+]$: 289.13, Found: 289.1408.

(Z)-4-((3,7-dimethylocta-2,6-dien-1-yl)oxy)-2-methylene-4-oxobutanoic acid (Mono 4-Néronyl Itaconate)(8e):



Gummy liquid. 1H NMR (300 MHz, $CDCl_3$, 25 °C) δ 6.48 (s, 1H, $C=CH_2$), 5.84 (s, 1H, $C=CH_2$), 5.35 (m, 1H, $CH=C^2H-CH_2$), 5.1 (m, 1H, $C=C^6H-CH_2$), 4.61 (d, 2H, $J = 7.2$ Hz, $O-C^1H_2CH$), 3.35 (s, 2H, $CH_2-C=$), 2.11 (m, 4H, $C^4H_2-C^5H_2$), 1.78 (s, 3H, $C^{10}H_3$), 1.69 (s, 3H, $=CC^9H_3$), 1.61 (s, H, $=CC^8H_3$). ^{13}C NMR (75 MHz, $CDCl_3$) δ 171.2, 170.5, 142.8, 133.3, 132.1, 130.6, 123.5, 118.9, 61.6, 37.2, 32.1, 26.6, 25.6, 23.4, 17.6. HRMS (ESI) m/z calcd for $C_{15}H_{22}O_4$ $[M + Na^+]$: 289.13, Found: 289.1408.

4-(benzyloxy)-2-methylene-4-oxobutanoic acid (9e). White crystalline. mp 80 °C. 1H NMR (300 MHz, $CDCl_3$, 25 °C) δ 10.3 (s, 1H, $COOH$), 7.37 (m, 5H, H_{arm}), 6.51 (s, 1H, $CH_2=C$), 5.86 (s, 1H, $CH_2=C$), 5.19 (s, 2H, $-COO-CH_2$), 3.43 (s, 2H, $-CH_2-COO$). ^{13}C NMR (75 MHz, $CDCl_3$) δ 171.5, 170.4, 135.6, 133.2, 130.9, 128.5, 128.2, 128.1, 66.8, 37.3. HRMS (ESI) m/z calcd for $C_{12}H_{12}O_4$ $[M + Na^+]$: 243.05, Found: 243.0641.

4-(2-phenylethoxy)-2-methylene-4-oxobutanoic acid (10e). White crystalline. mp 83 °C. ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 7.25 (m, 5H, H_{arm}), 6.48 (s, 1H, CH₂=C), 5.82 (s, 1H, CH₂=C), 4.34 (t, 2H, J = 7 Hz, COO-CH₂-CH₂-Ar), 3.35 (s, 2H, -CH₂-COO), 2.96 (t, 2H, J = 7.2 Hz, -CH₂-Ar). ¹³C NMR (75 MHz, CDCl₃) δ 171.3, 170.2, 137.5, 133.1, 130.9, 128.9, 128.5, 126.5, 65.5, 37.2, 34.9. HRMS (ESI) m/z calcd for C₁₃H₁₄O₄ [M + Na⁺]: 257.07, Found: 257.0787

4-(3-phenylethoxy)-2-methylene-4-oxobutanoic acid (11e). White crystalline. mp 86 °C. ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 8.6 (s, 1H, OH), 7.25 (m, 5H, H_{arm}), 6.49 (s, 1H, CH₂=C), 5.85 (s, 1H, CH₂=C), 4.15 (t, 2H, J = 6.5 Hz, COO-CH₂-(CH₂)₂-Ar), 3.37 (s, 2H, -CH₂-COO), 2.7 (m, 2H, CH₂-CH₂-Ar), 1.98 (m, 2H, -CH₂-CH₂-Ar). ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 170.7, 141.1, 133.3, 130.7, 128.4, 126.0, 64.3, 37.3, 32.0, 30.1. HRMS (ESI) m/z calcd for C₁₄H₁₆O₄ [M + Na⁺]: 271.08, Found: 271.0940.

4-(cinnamyloxy)-2-methylene-4-oxobutanoic acid (12e). White crystalline. mp 87 °C. ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 7.36 (m, 5H, H_{arm}), 6.6 (d, 1H, J = 15.9 Hz, CH=CH-Ph), 6.49 (s, 1H, CH₂=C), 6.31 (m, 1H, CH=CH-Ph), 5.87 (s, 1H, CH₂=C), 4.7 (dd, 2H, J = 6.4, 1 Hz, COO-CH₂-CH=CH-Ar), 3.4 (s, 2H, -CH₂-COO). ¹³C NMR (75 MHz, CDCl₃) δ 171.3, 170.4, 136.1, 134.3, 133.1, 130.9, 128.6, 128.0, 126.6, 122.8, 65.6, 37.3, 29.7. HRMS (ESI) m/z calcd for C₁₄H₁₄O₄ [M + Na⁺]: 269.07, Found: 269.0781

4-([1,1'-biphenyl]-4-ylmethoxy)-2-methylene-4-oxobutanoic acid (13e). White crystalline. mp 98 °C. ¹H NMR (300 MHz, CDCl₃) δ 3.4 (s, 2H, -CH₂-COO); 5.21 (s, 2H, COO-CH₂-Ph), 5.84 (s, 1H, CH₂=C); 6.48 (s, 1H, CH₂=C), 7.47-7.68 (m, 9H, H_{arm}). ¹³C NMR (75 MHz, CDCl₃) δ 170.7, 170.2, 140.8, 140.6, 139.7, 128.8, 128.7, 127.5, 127.3, 127.1, 66.5, 65.0, 37.5. HRMS (ESI) m/z calcd for C₁₈H₁₆O₄ [M + Na⁺]: 319.08, Found: 319.0928.

4-(furan-2-ylmethoxy)-2-methylene-4-oxobutanoic acid (14e). White crystalline. mp 75 °C. ¹H NMR (300 MHz, CDCl₃, 25 °C) δ 10 (s, 1H, OH), 7.43 (, 1H, CH=CH-O), 6.49 (s, 1H, CH₂=C), 6.42 (m, 1H, CH=CH), 6.37 (m, 1H, CH=CH) 5.85(s, 1H, CH₂=C), 5.12(s, 2H, COO-CH₂-CH=), 3.38 (s, 2H, -CH₂-COO). ¹³C NMR (75 MHz, CDCl₃) δ 171.4, 170.3, 149.1, 143.3, 133.03, 131.07, 110.8, 110.5, 58.7, 37.1. HRMS (ESI) m/z calcd for C₁₀H₁₀O₅ [M + Na⁺]:233.03, Found: 233.0421.

4. Crystallographic data and structure refinement details for monoesters itaconates: 2e, 6e and 9e .

X-ray diffraction data for compounds (6e: mo_v463), (2e: mo_v468) & (9e: mo_v471) were collected by using a VENTURE PHOTON100 CMOS Bruker diffractometer with Micro-focus IuS source Mo K α radiation ($\lambda = 0.71073 \text{ \AA}$). Crystals were mounted on a CryoLoop (Hampton Research) with Paratone-N (Hampton Research) as cryoprotectant and then flashfrozen in a nitrogen-gas stream at 100 K. For compounds, the temperature of the crystal was maintained at the selected value by means of a N-Helix Cryostream cooling device to within an accuracy of $\pm 1\text{K}$. The data were corrected for Lorentz polarization, and absorption effects. The structures were solved by direct methods using SHELXS-97¹ and refined against F^2 by full-matrix least-squares techniques using SHELXL-2018² with anisotropic displacement parameters for all non-hydrogen atoms. Hydrogen atoms were located on a difference Fourier map and introduced into the calculations as a riding model with isotropic thermal parameters. All calculations were performed by using the Crystal Structure crystallographic software package WINGX³.

The crystal data collection and refinement parameters are given in Table 3.

CCDC 2088738-2088740 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/structures.

5. *PCL-catalyzed regioselective and enantioselective opening ring of (1An) using a set of arylalkyl carbinols as chiral nucleophiles:*

To equimolecular mixture of itaconic anhydride and the appropriate alcohol (*rac*-16-25) diluted in 5mL of TBME, the adequate amount of lipase is added. After 24 hours of stirring at room temperature, the lipase was removed by filtration. The obtained monoester itaconate and the remained alcohols were easily separated by a simple alkaline washing. The two components were recovered by liquid-liquid extraction, the remaining (*S*)-alcohol was recovered in the organic layer and the monoester itaconate in the aqueous phase and then the chemical isolated yield was determined. The proportions of the isomeric mixture of the (α / β) mono esters itaconates recovered were checked by ¹H NMR analysis. Their quantifications were based on the chemical shifts of the double bond protons. The enantiomeric excesses of the (*R*)-monoesters itaconates are measured by chiral HPLC after saponification and obtaining the corresponding (*R*)-alcohols.

***(R)*-4-((1,2-dihydroacenaphthylen-1-yl)oxy)-2-methylene-4-oxobutanoic acid (19e)**

White powder. mp 140°C. ¹HNMR (300 MHz, CDCl₃, 25°C) δ 09.5 (s, 1H, OH), 7.56 (m, 6H, ar), ~~6.67 (dd, 1H, $J = 7.1 \text{ Hz}, 2.3 \text{ Hz}$, CH₂OOCCH),~~ 6.48 (s, 1H, C=CH₂), 5.84 (s, 1H, C=CH₂), , 3.84

¹ Sheldrick, G. M. SHELXS-97, Program for Crystal Structure Solution, University of Göttingen, Göttingen, Germany, 1997.

² G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Crystallogr., 2008, 64, 112-122

³ Farrugia, L. J. J. Appl. Cryst., 1999, 32, 837.

(dd, 1H, $J = 18$ Hz), 3.39 (s, 2H, CH_2OOC), 3.31 (m, 1H). ^{13}C NMR (75 MHz, CDCl_3): δ 171.58, 170.81, 141.67, 141.09, 137.89, 133.25, 131.15, 131.06, 128.16, 128.12, 125.5, 122.95, 121.93, 119.86, 76.4, 38.64, 37.56. HRMS (ESI) m/z calcd for $\text{C}_{17}\text{H}_{13}\text{O}_4$ [$\text{M} - 1$]:281.09, Found 281.081.

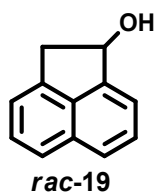
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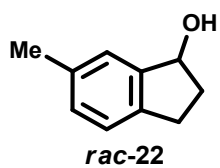
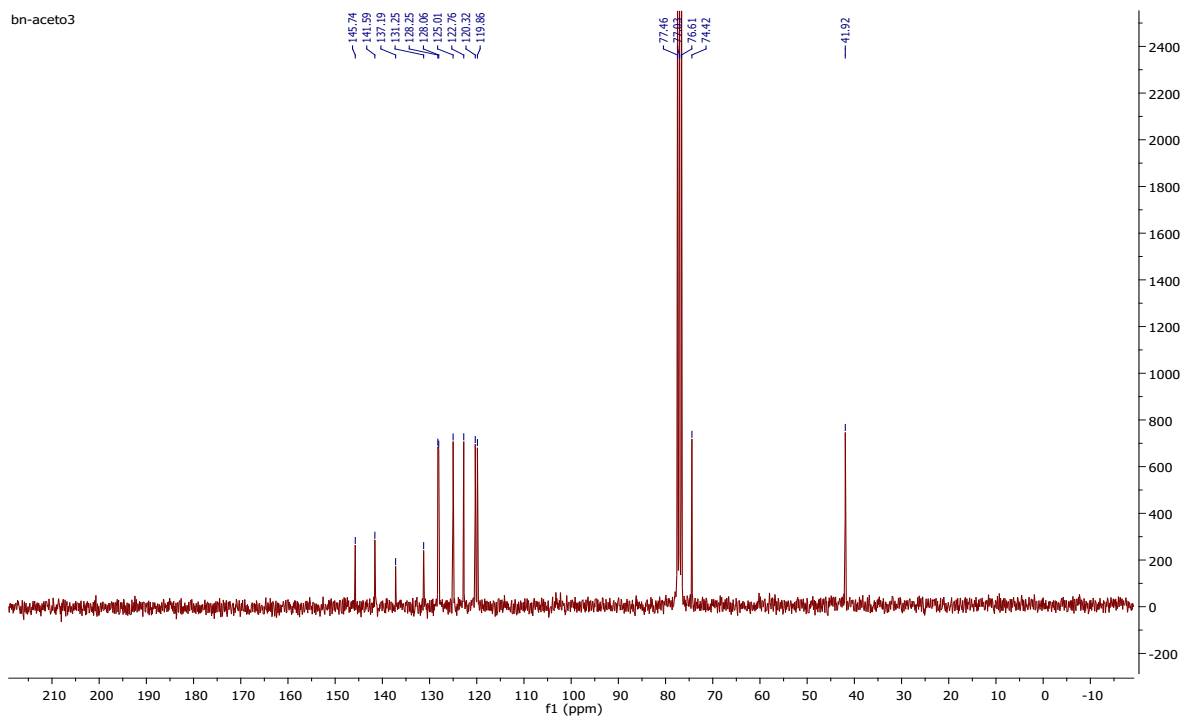
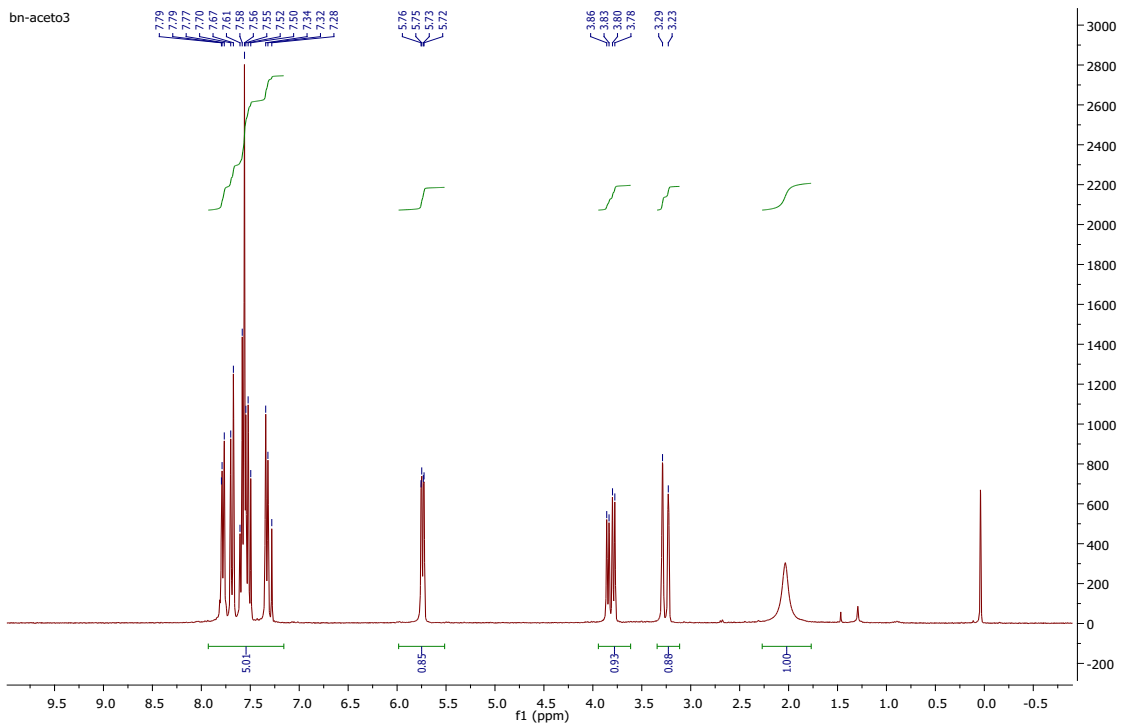
White powder. mp 106°C. ^1H NMR (300 MHz, CDCl_3 , 25°C) δ 7.42 (d, $J = 7.4$ Hz, 1H, H_{arm}), 7.29 (m, 2H, H_{arm}), 6.48 (s, 1H, $\text{C}=\underline{\text{CH}_2}$), 6.26 (dd, $J = 7.4$ Hz, 1H, CHOCO), 5.85 (s, 1H, $\text{C}=\underline{\text{CH}_2}$), 3.37 (s, 2H, CH_2O), 3.13 (m, 1H), 2.9 (m, 1H), 2.52 (m, 1H), 2.13 (m, 1H). ^{13}C NMR (75 MHz, CDCl_3) δ 171.15, 170.61, 144.41, 140.76, 133.32, 130.68, 129, 126.74, 125.59, 124.79, 79.07, 37.59, 32.17, 30.19. HRMS (ESI) m/z calcd for $\text{C}_{14}\text{H}_{14}\text{O}_4$ [$\text{M} - 1$]:245.09, Found 245.816.

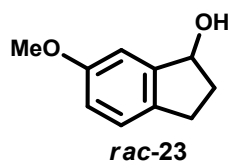
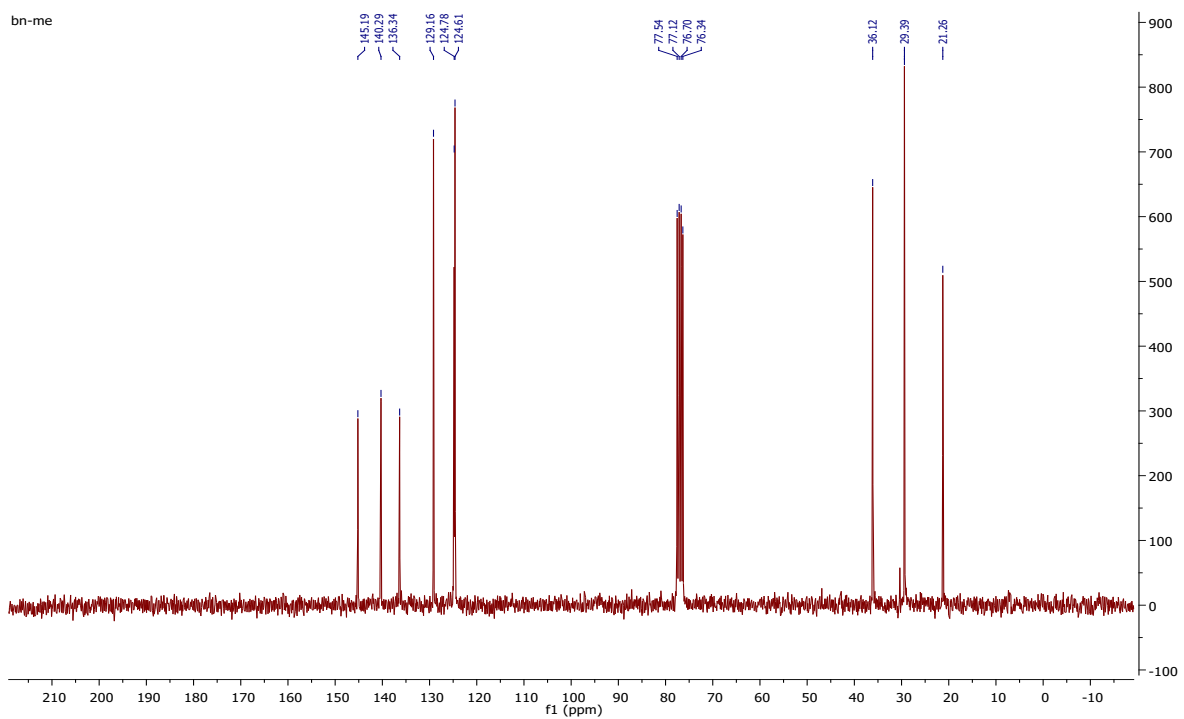
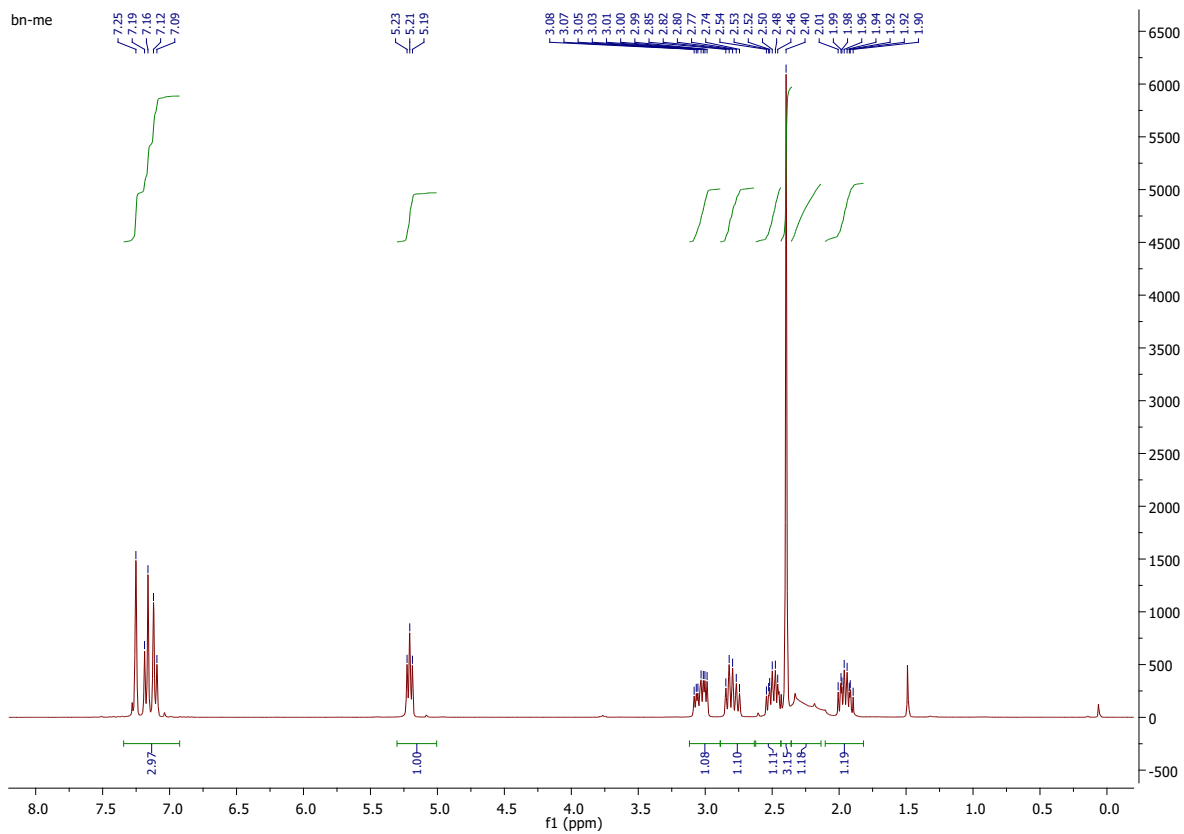
(R)-4-((6-methoxy-2,3-dihydro-1H-inden-1-yl)oxy)-2-methylene-4-oxobutanoic acid (23e)

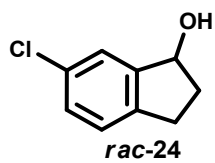
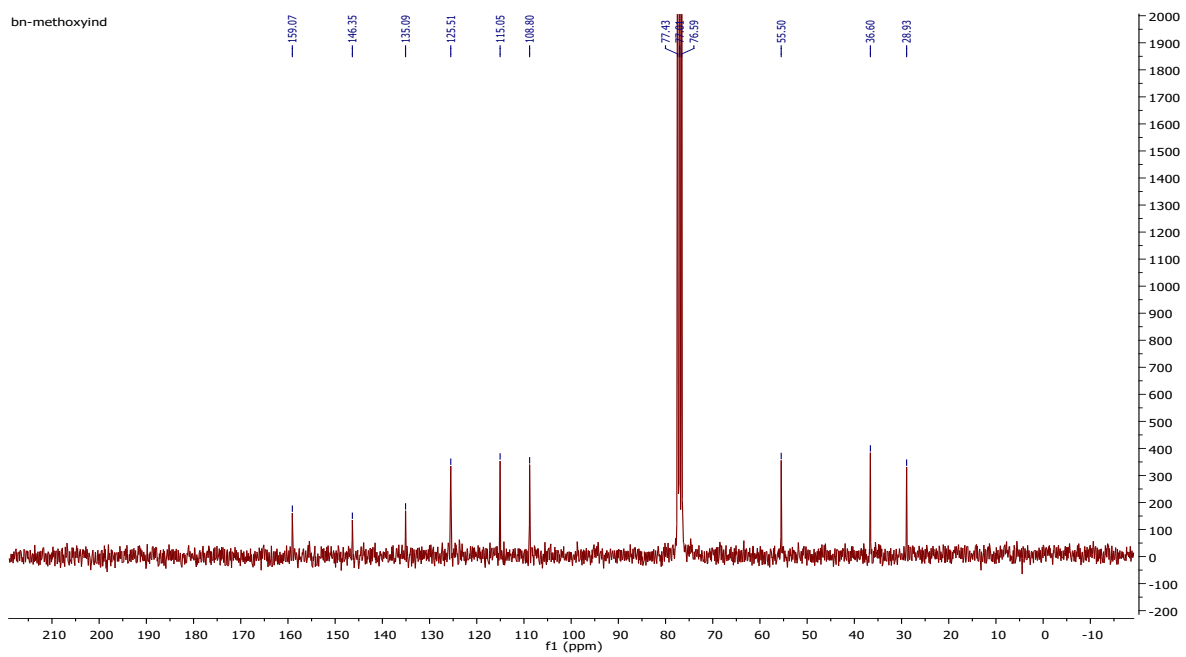
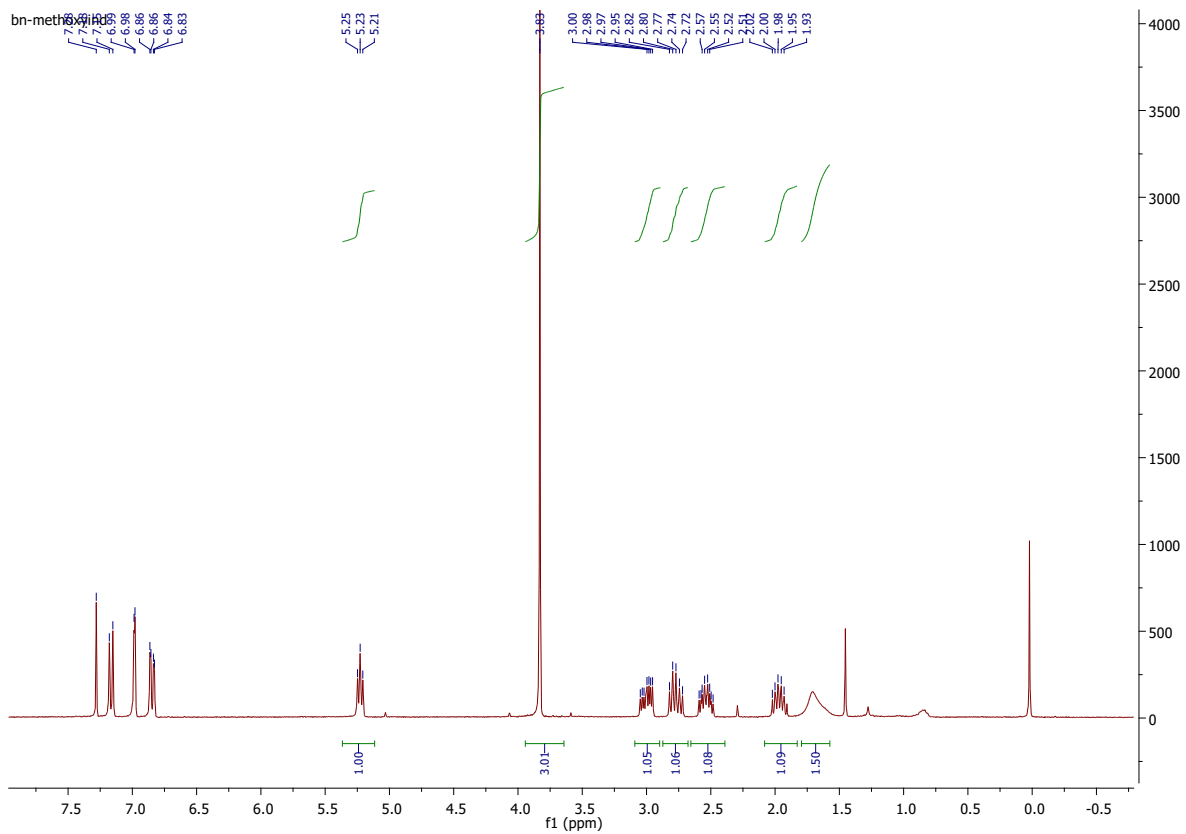
White powder. mp 110°C. ^1H NMR (300 MHz, CDCl_3 , 25°C) δ 7.17 (d, $J = 8.3$ Hz, 1H, H_{arm}), 6.9 (m, 2H, H_{arm}), 6.48 (s, 1H, $\text{C}=\underline{\text{CH}_2}$), 6.23 (dd, 1H, $J = 7.4$ Hz, CHOCO), 5.85 (s, 1H, $\text{C}=\underline{\text{CH}_2}$), 3.8 (s, 3H, CH_3O), 3.38 (s, 2H), 3.05 (m, 1H), 2.82 (m, 1H), 2.54 (m, 1H), 2.12 (m, 1H). ^{13}C NMR (75 MHz, CDCl_3) δ 171.28, 170.57, 158.93, 142, 136.23, 130.71, 125.37, 116, 109.76, 79.18, 55.48, 37.56, 32.69, 29.33. HRMS (ESI) m/z calcd for $\text{C}_{15}\text{H}_{16}\text{O}_5$ [$\text{M} - 1$]:275.10, Found 275.0924.

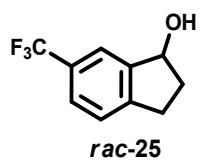
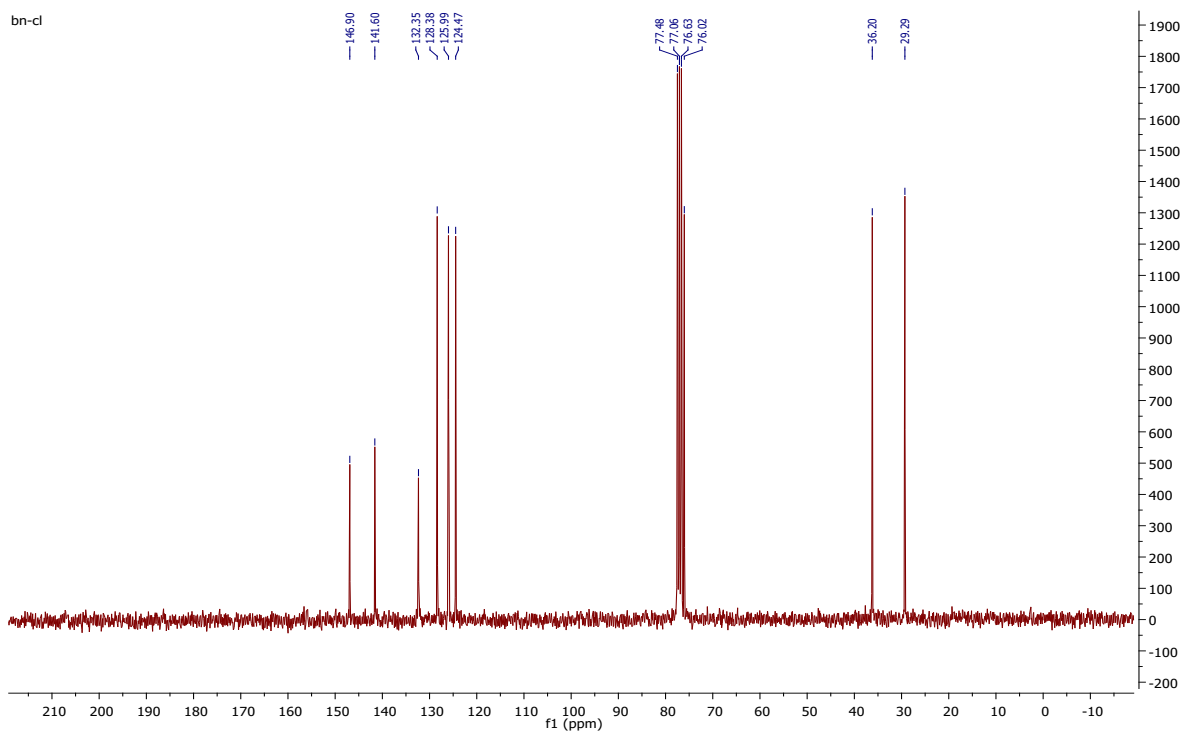
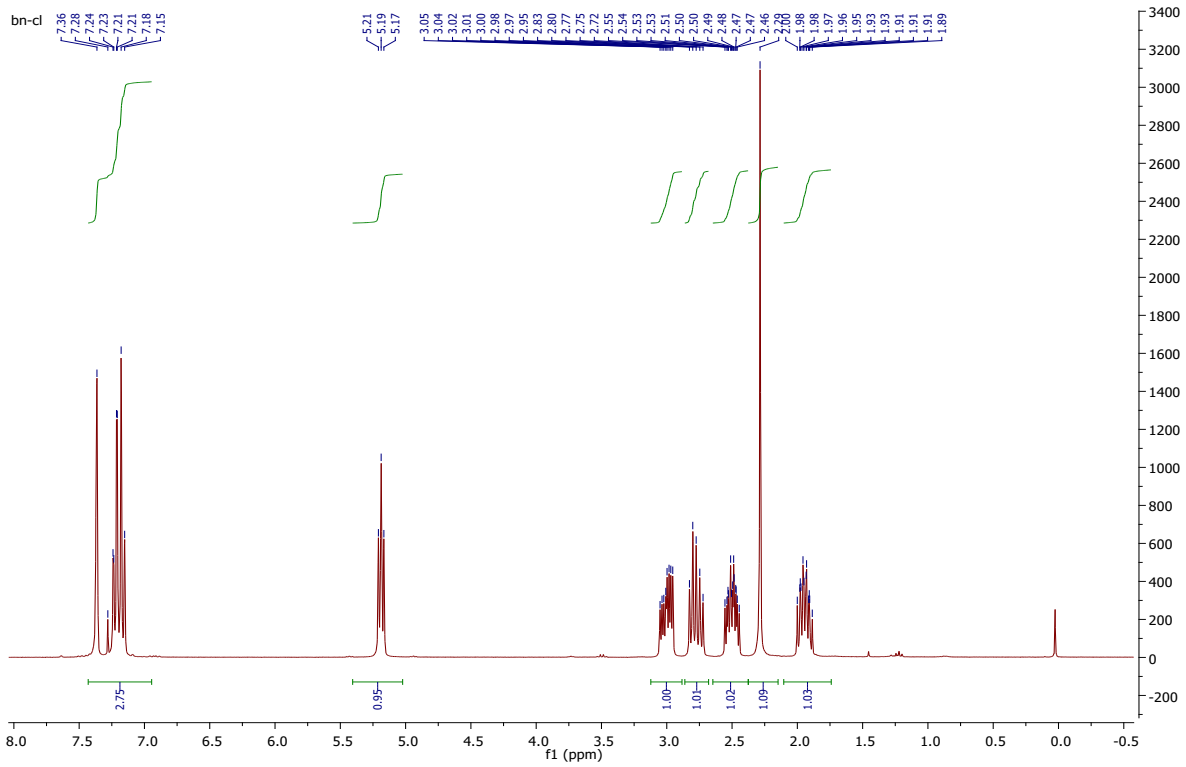
6. NMR spectra of racemic alcohols [19, 22-25]:

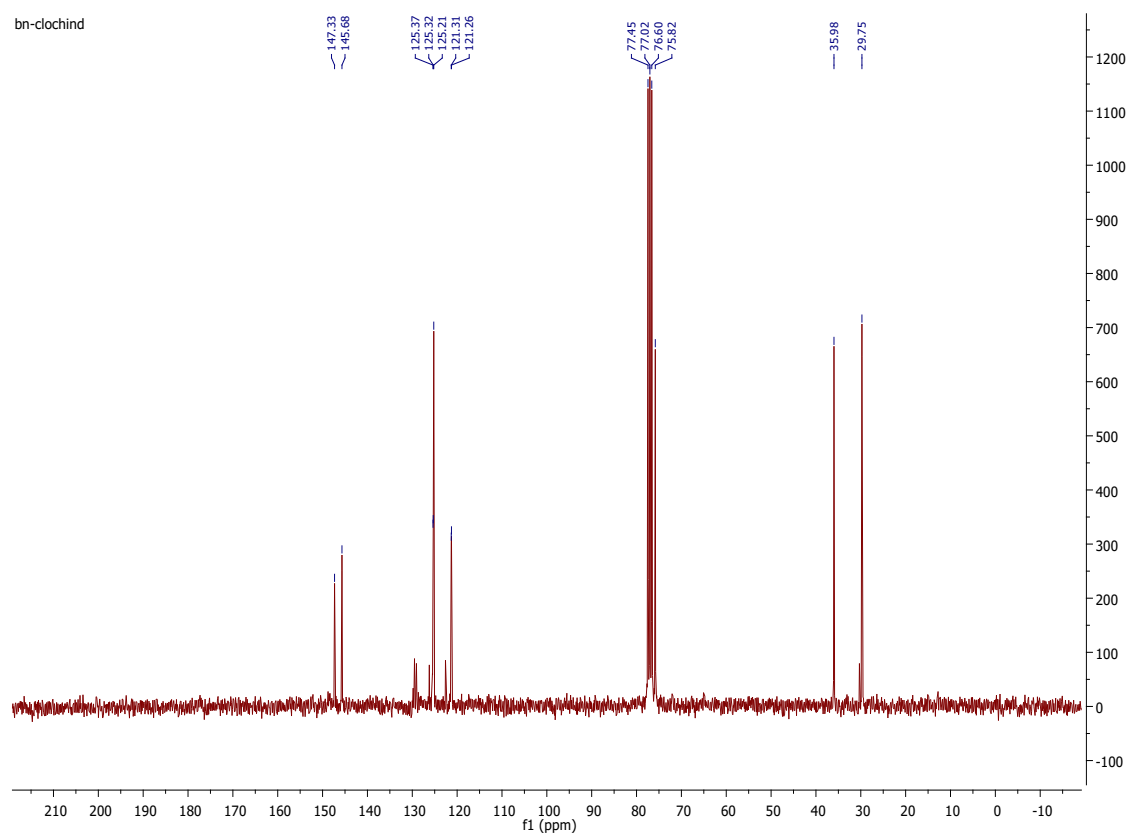
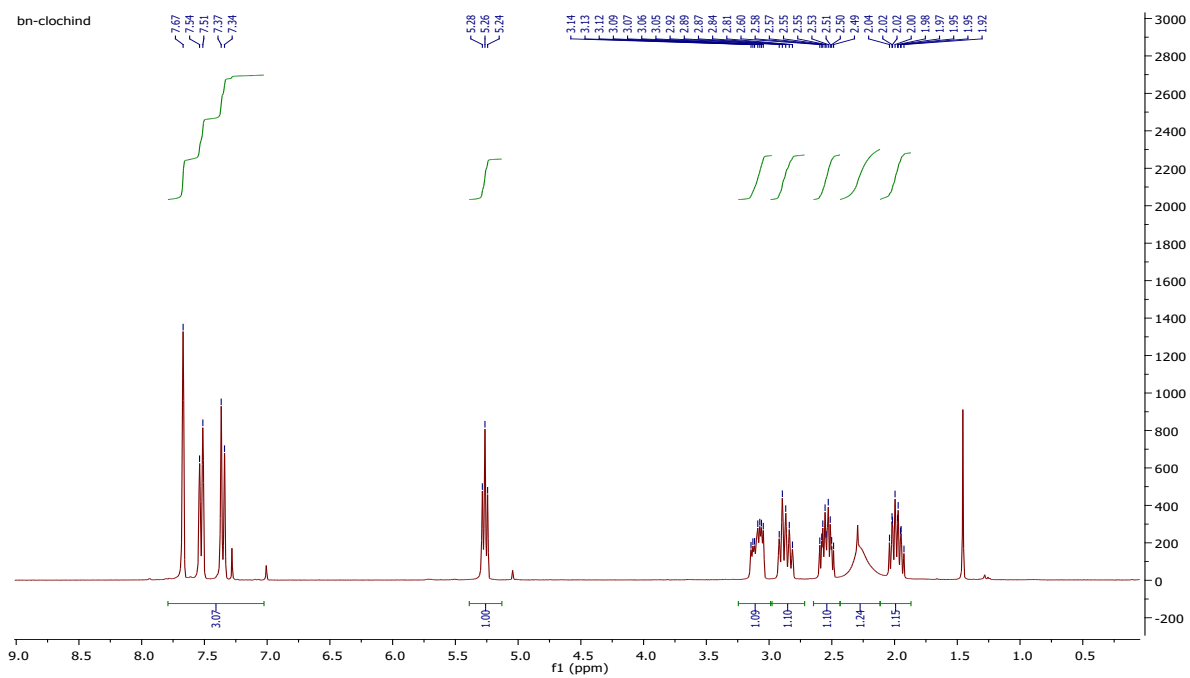




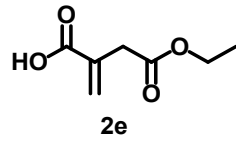




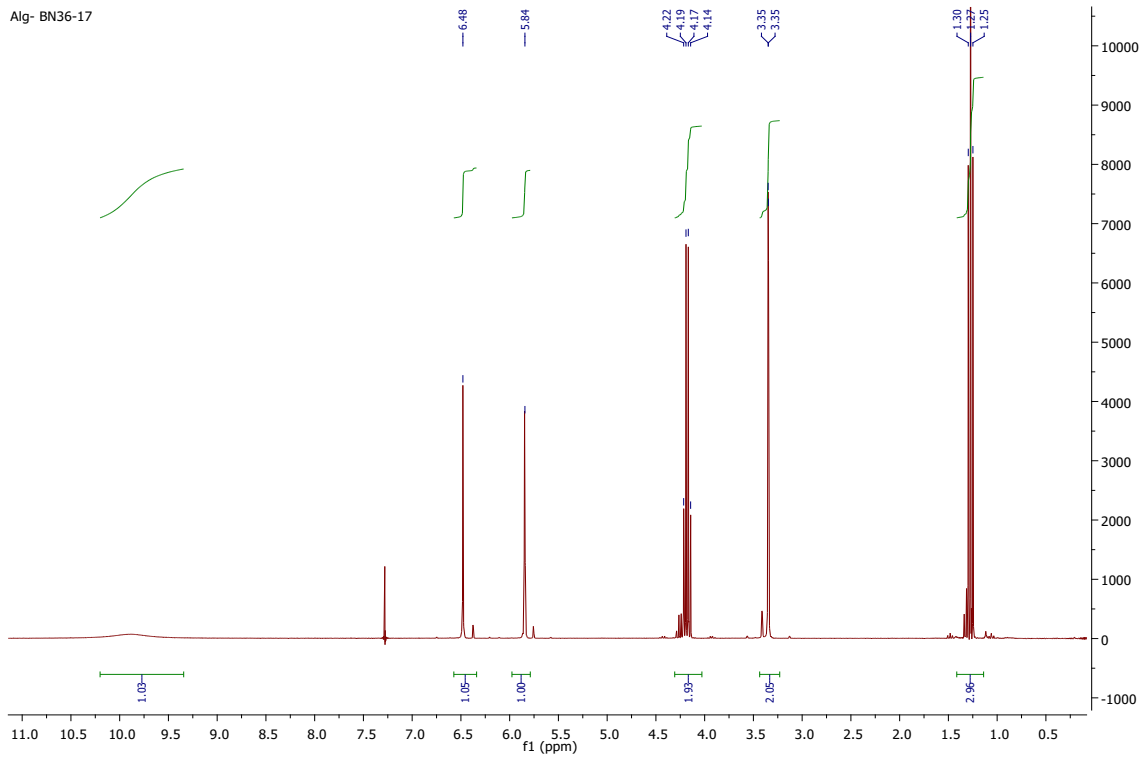




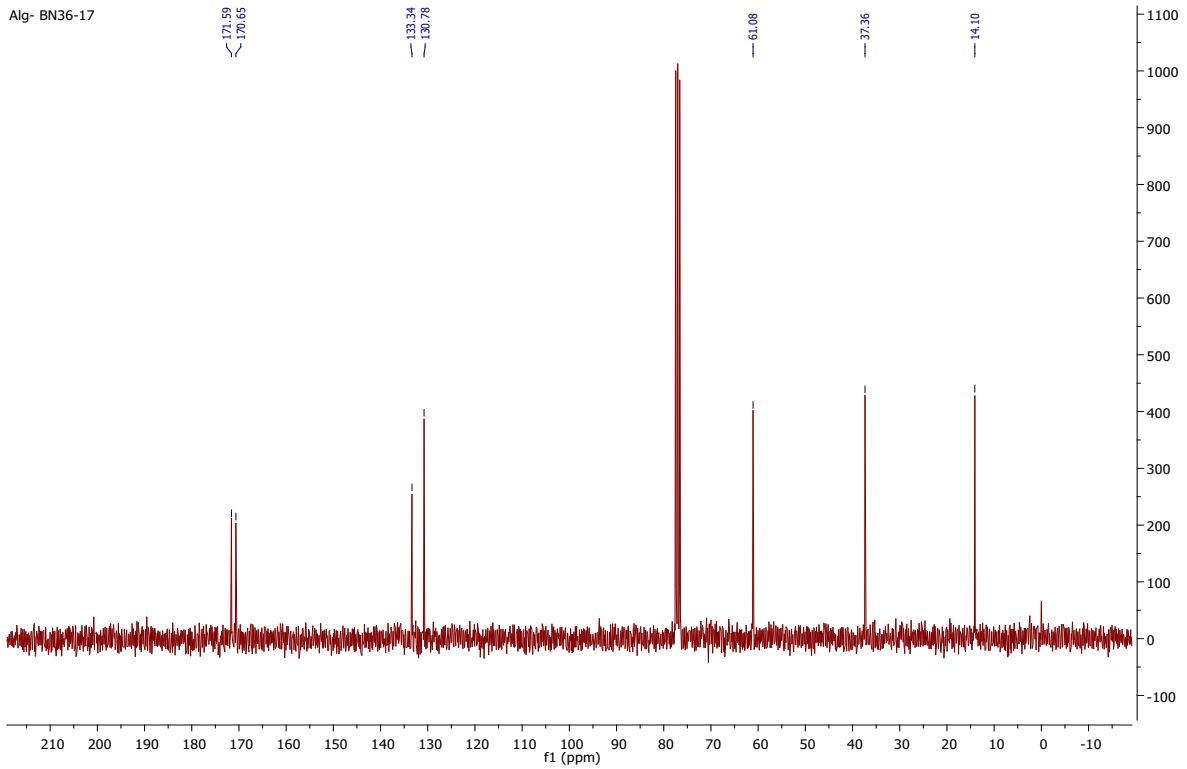
NMR spectra of all the monoesters itaconates:

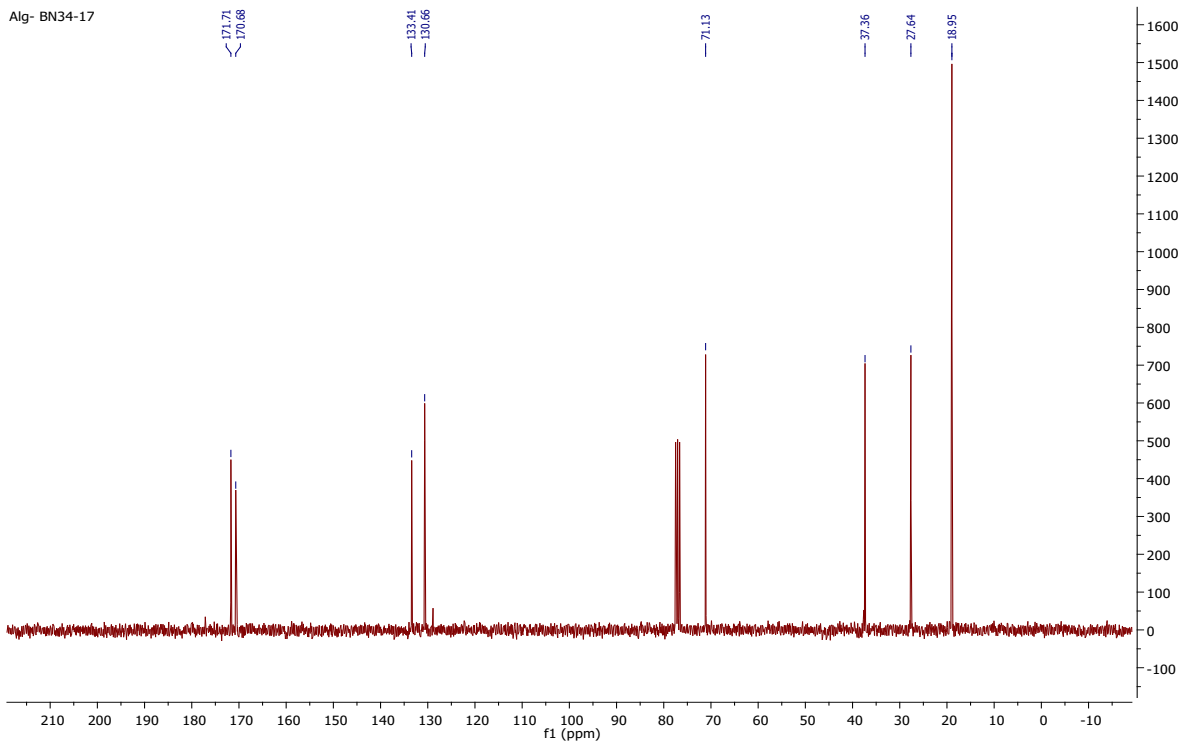
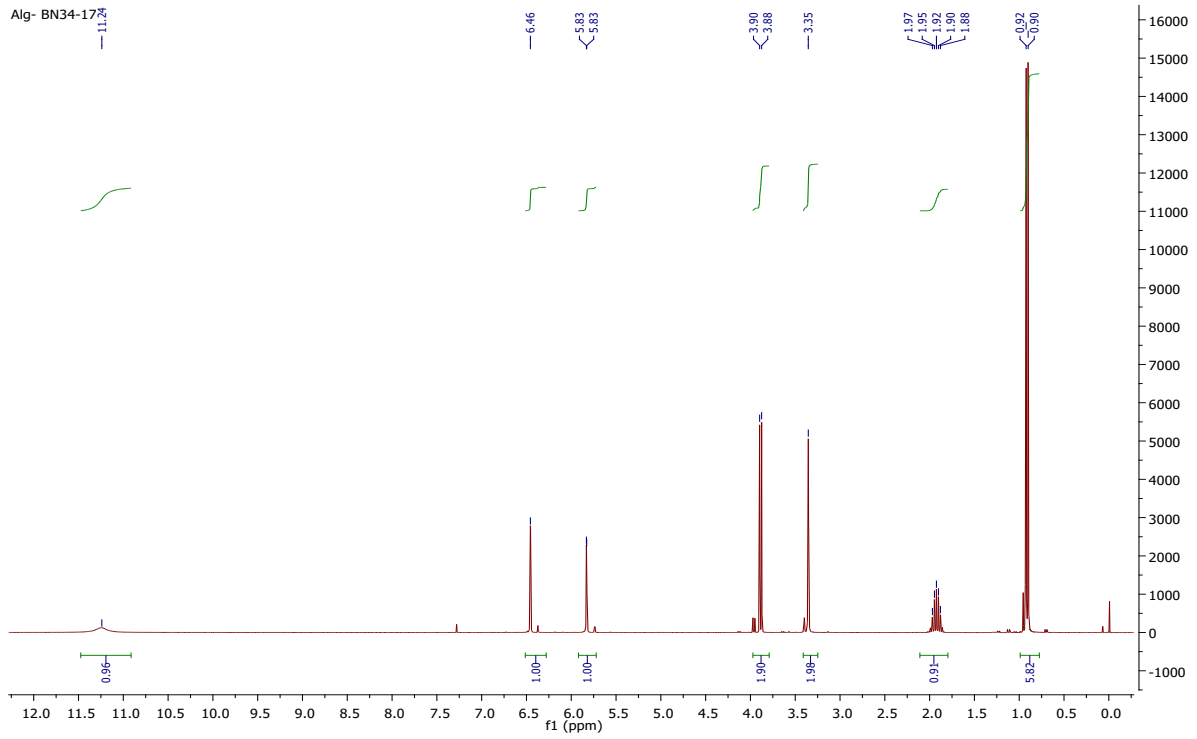
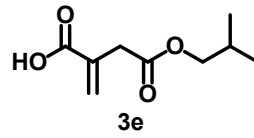


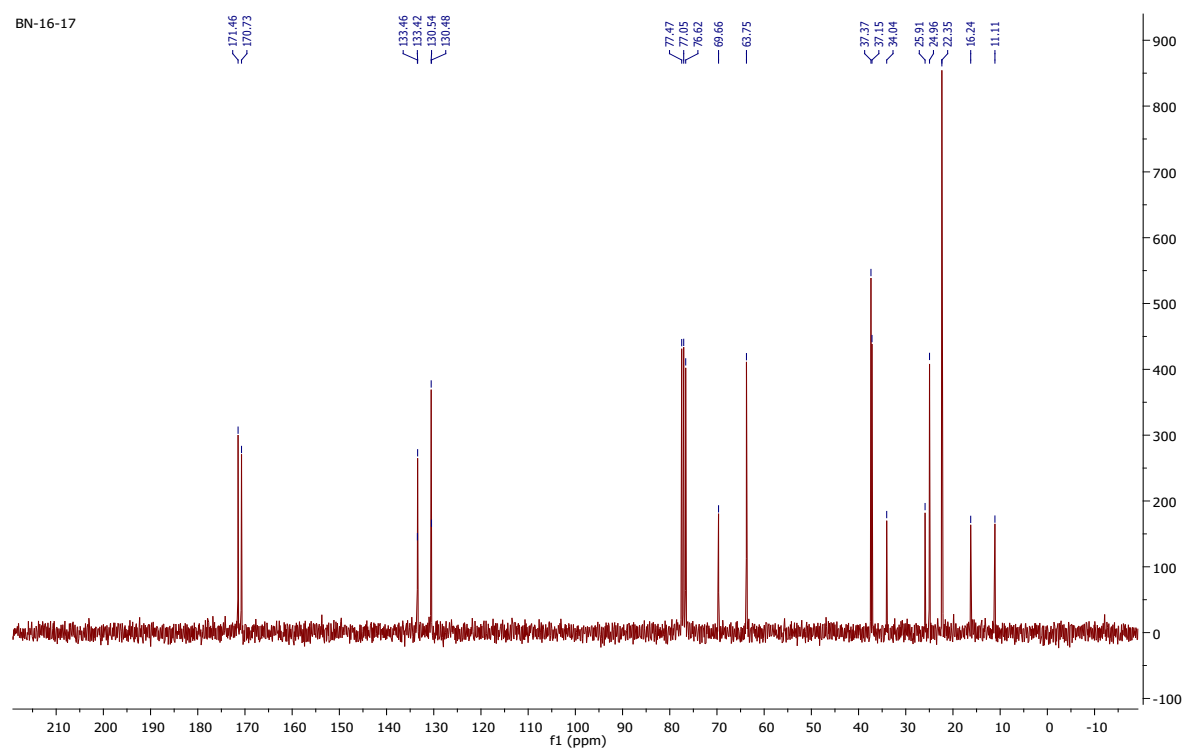
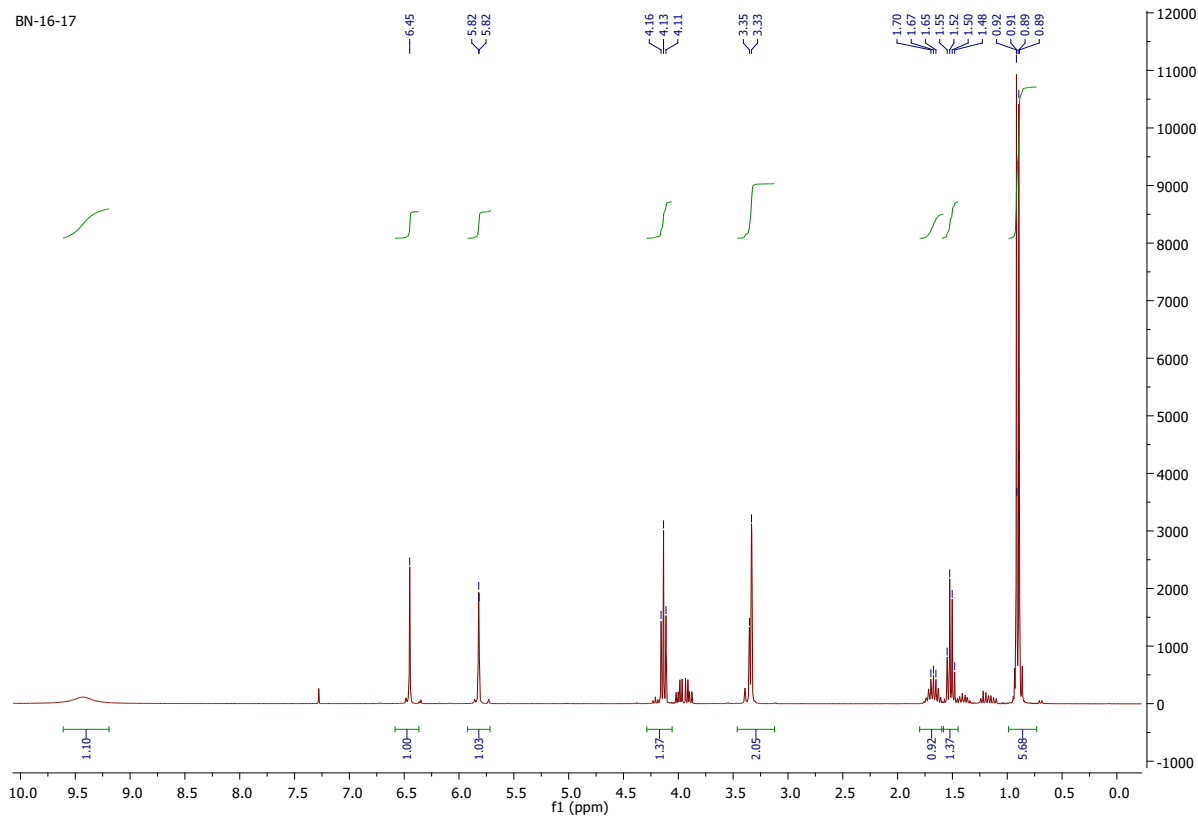
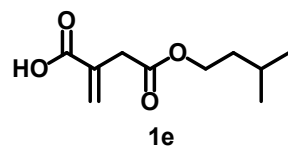
Alg- BN36-17

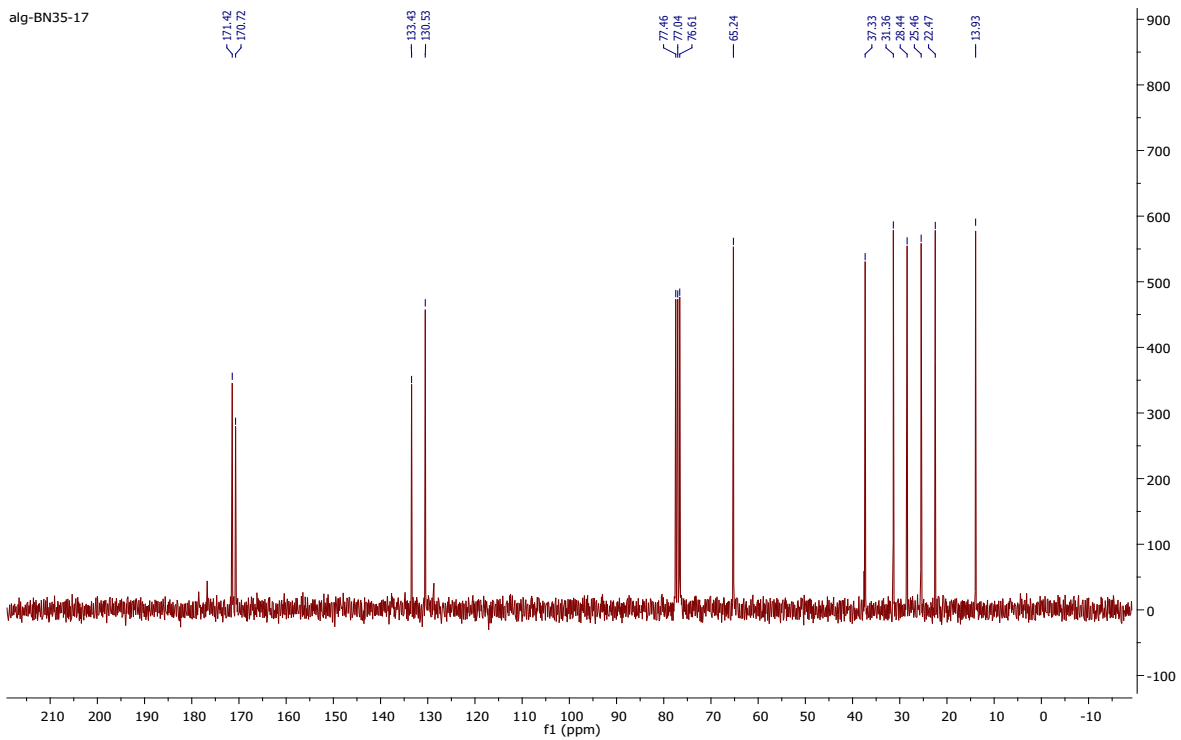
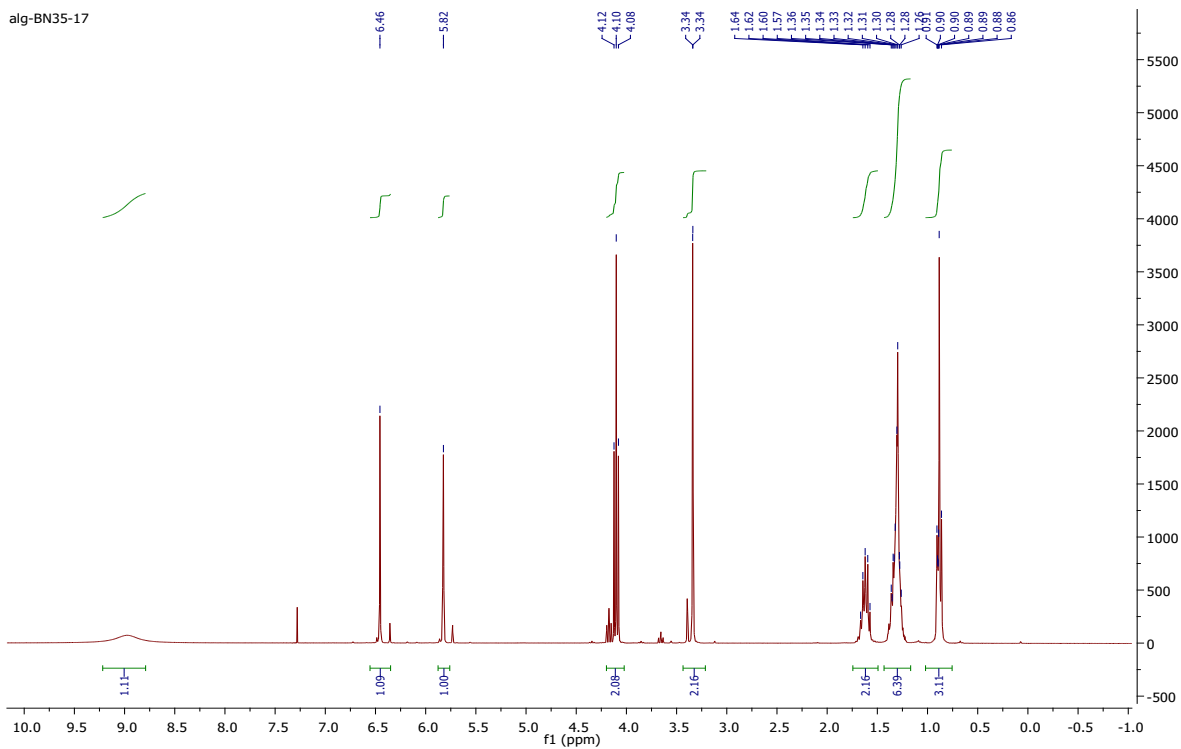
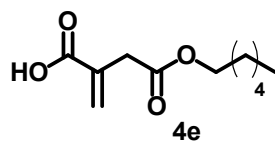


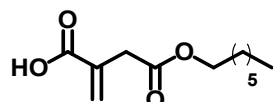
Alg- BN36-17





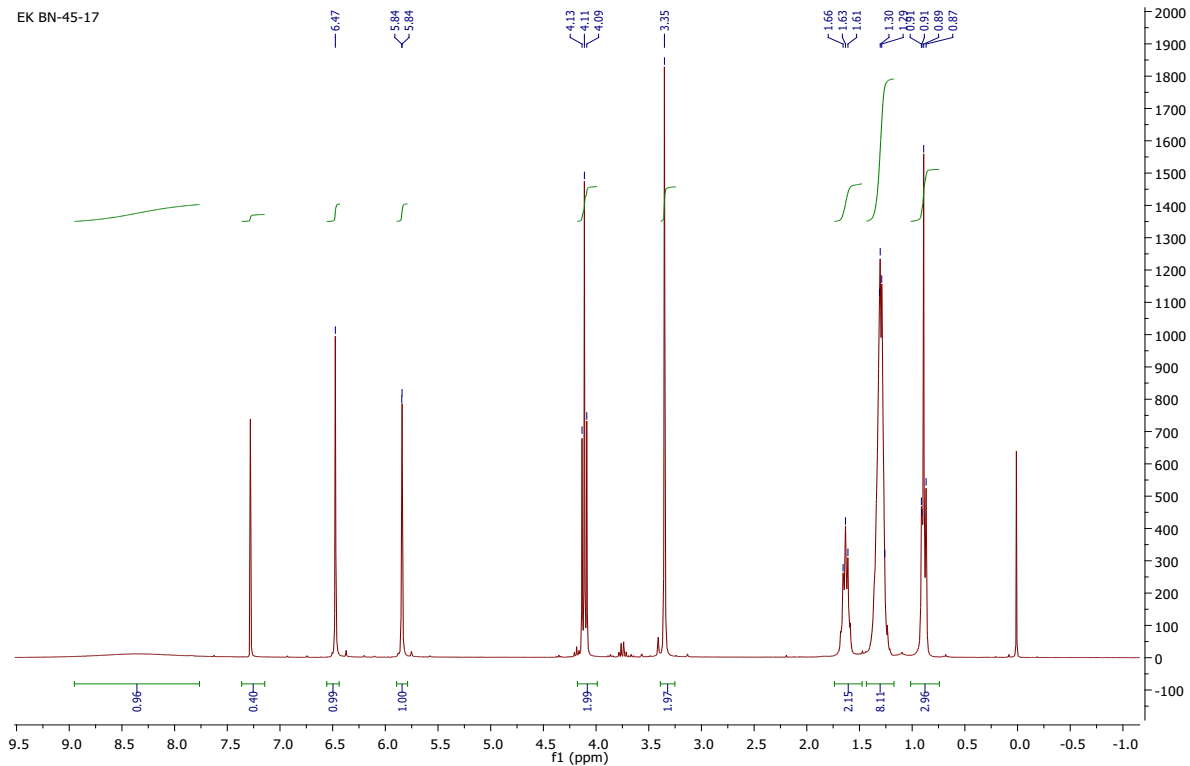




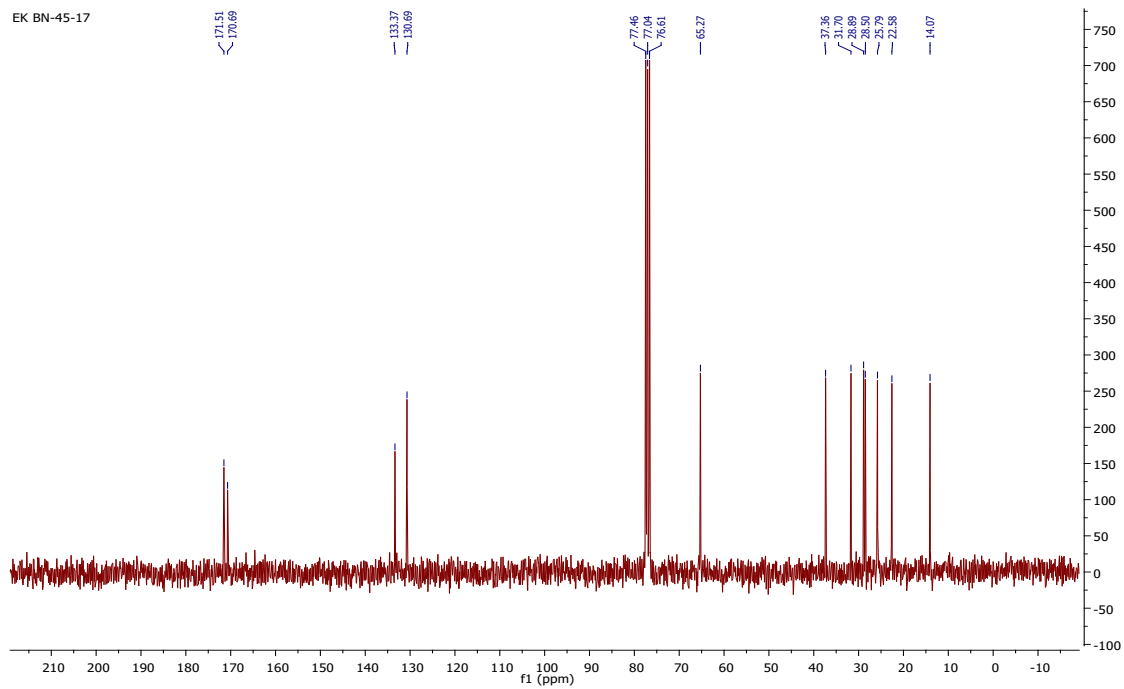


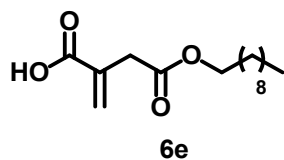
5e

EK BN-45-17

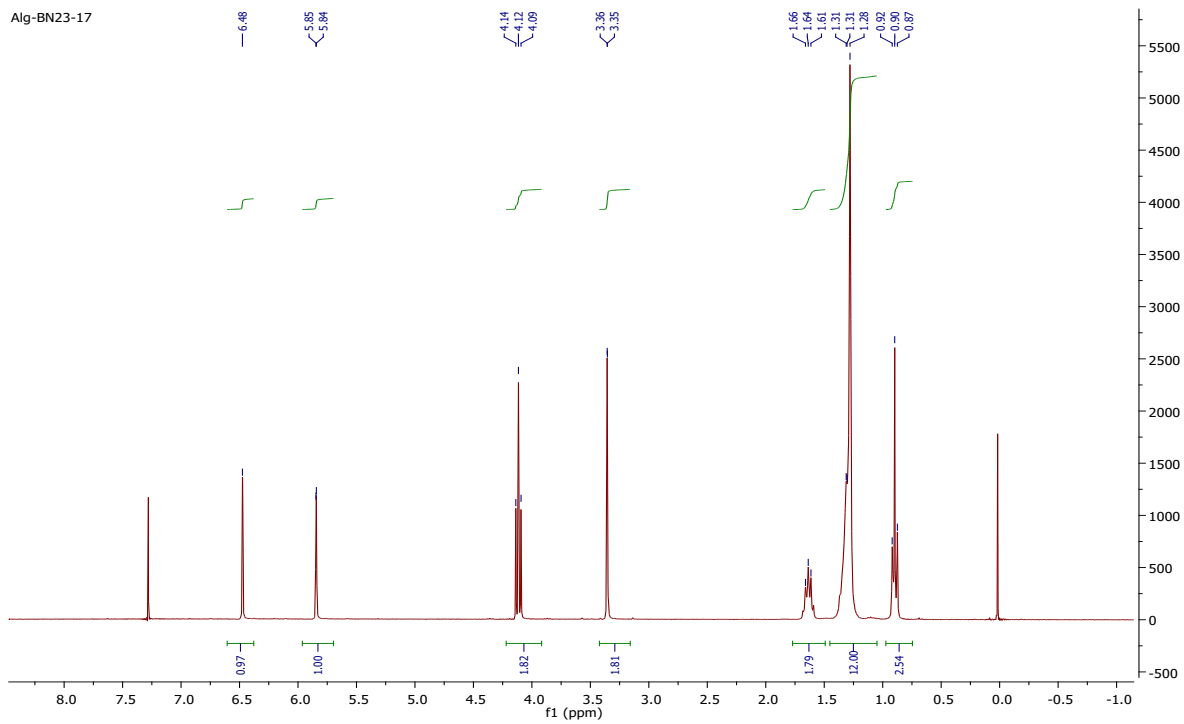


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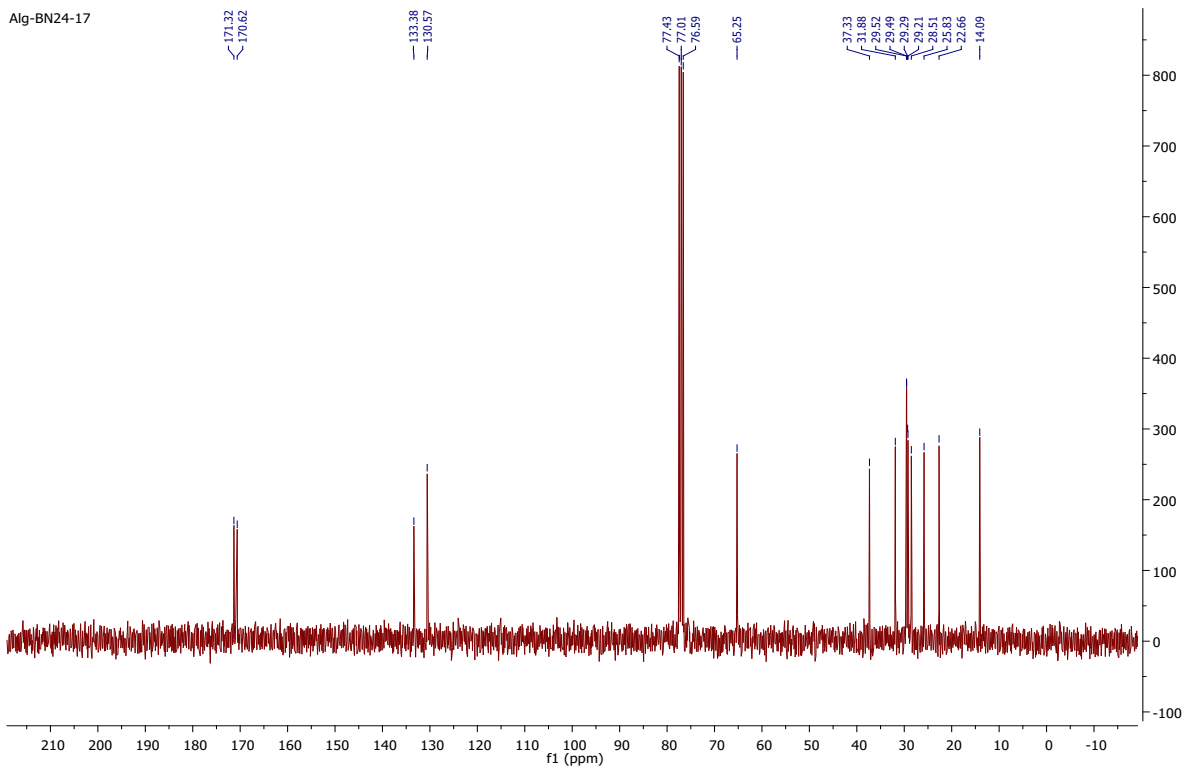


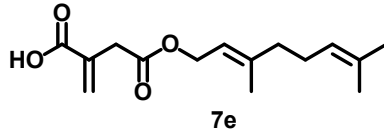


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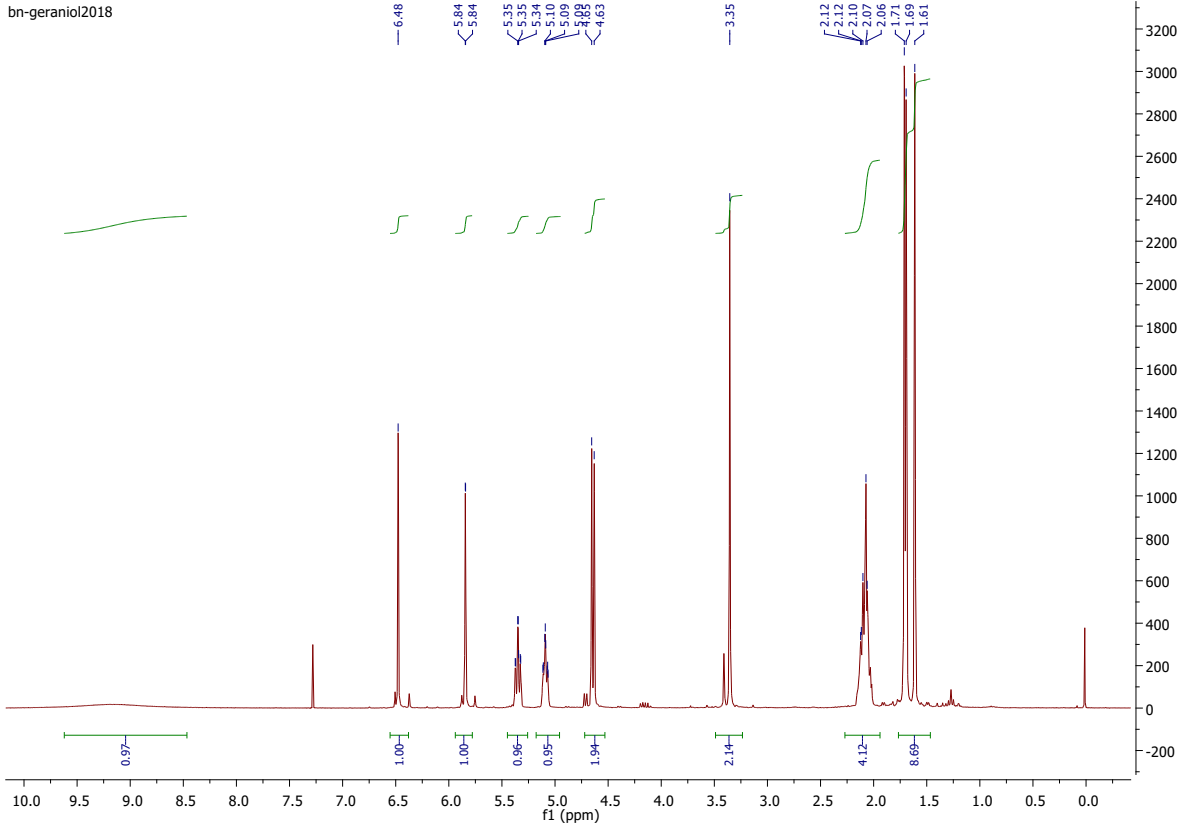


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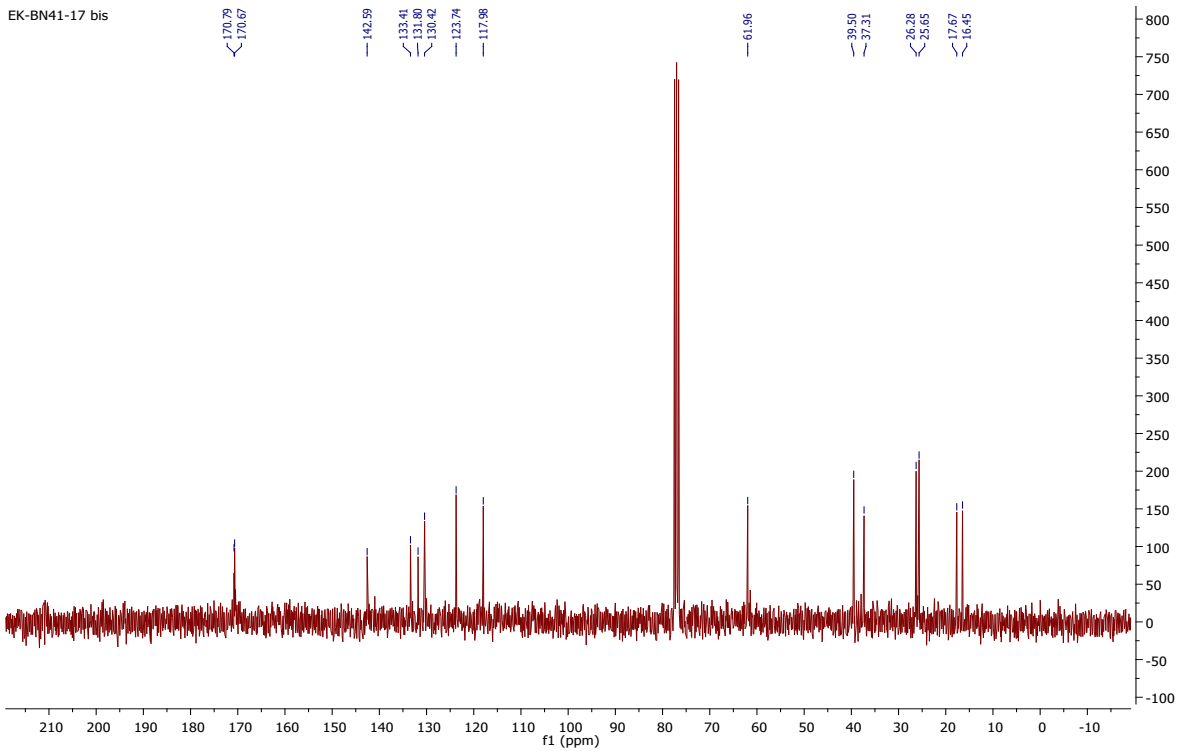


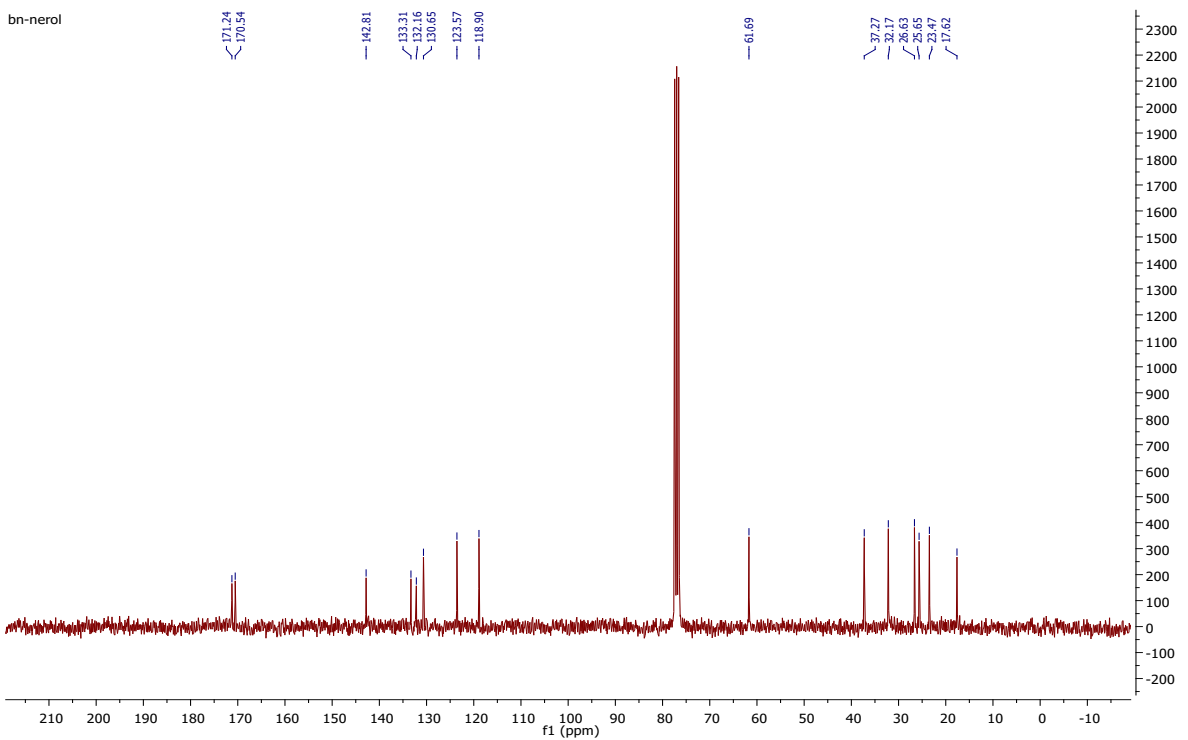
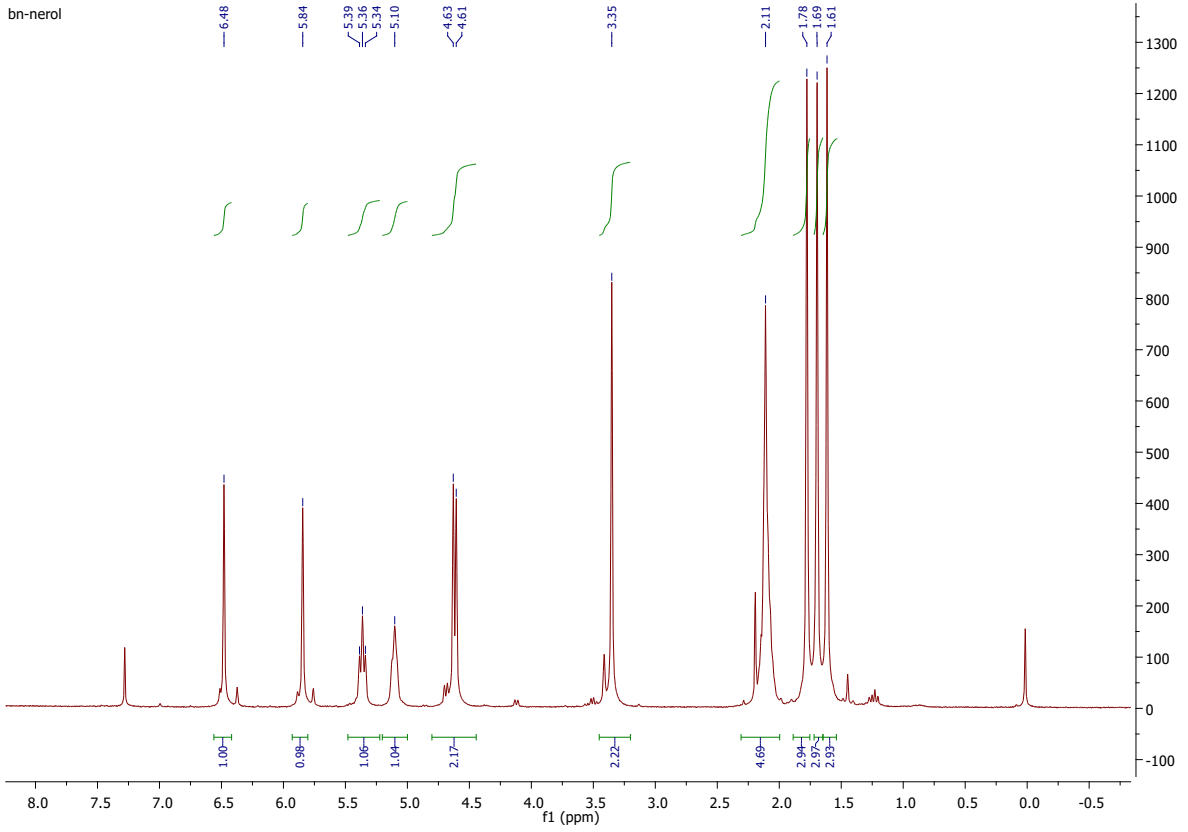
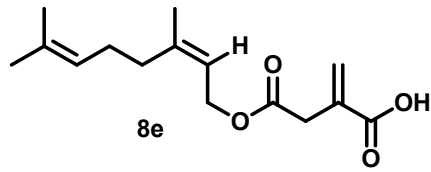


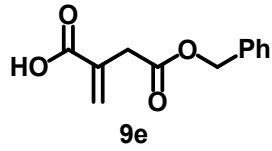
bn-geraniol2018



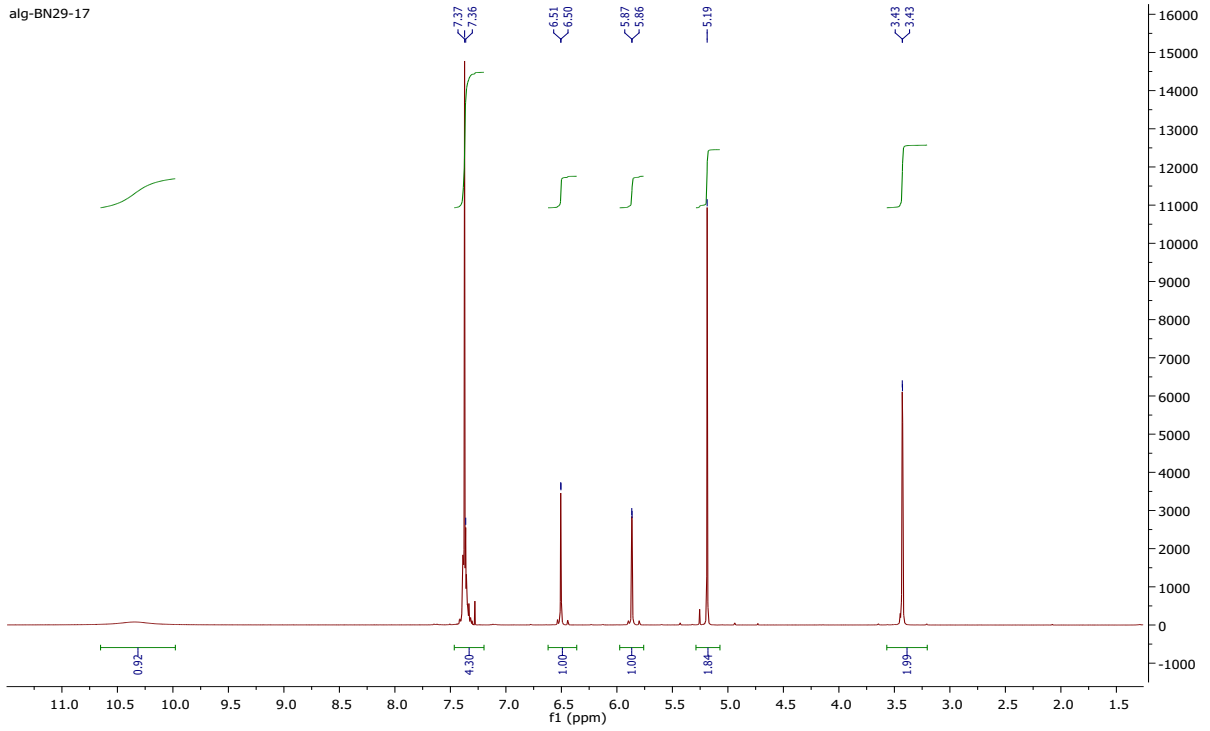
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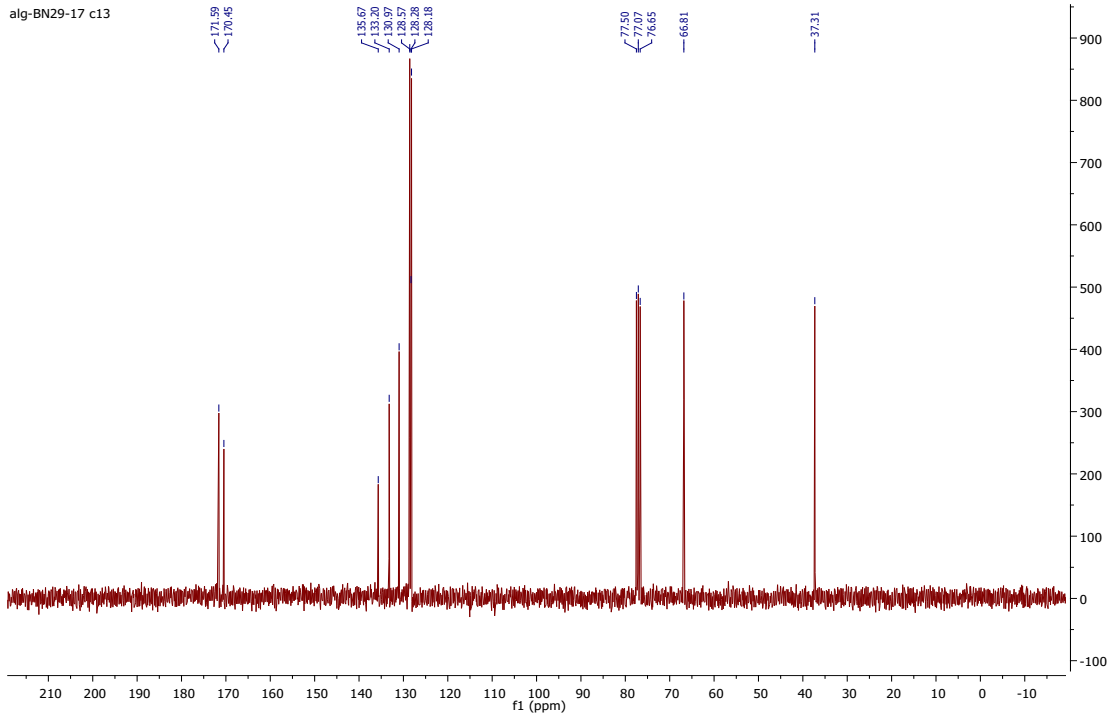


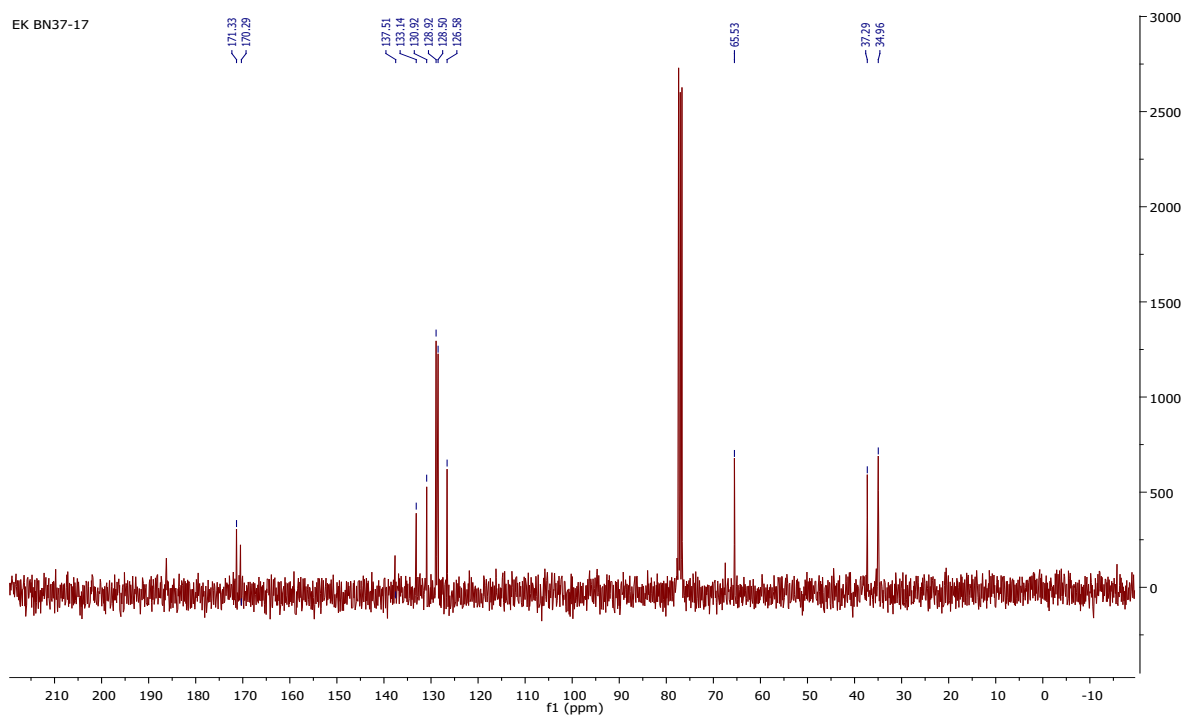
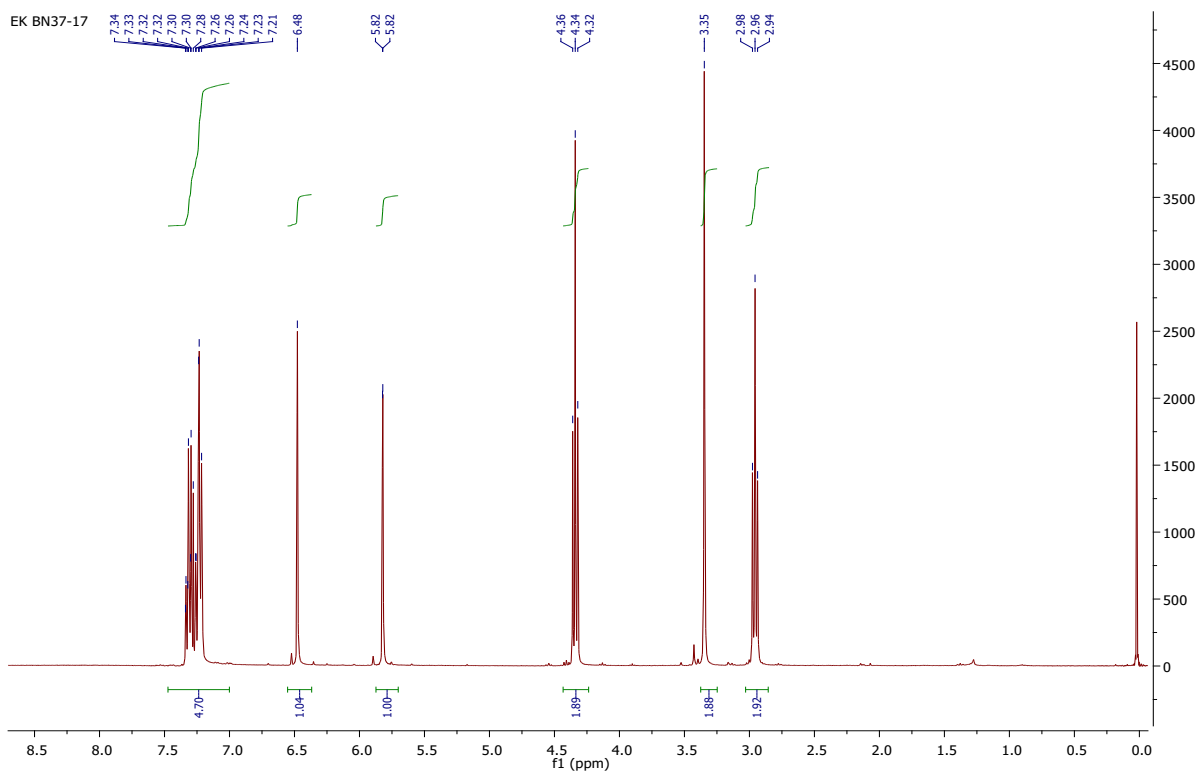
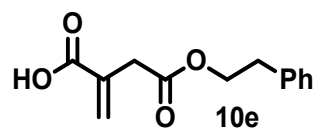


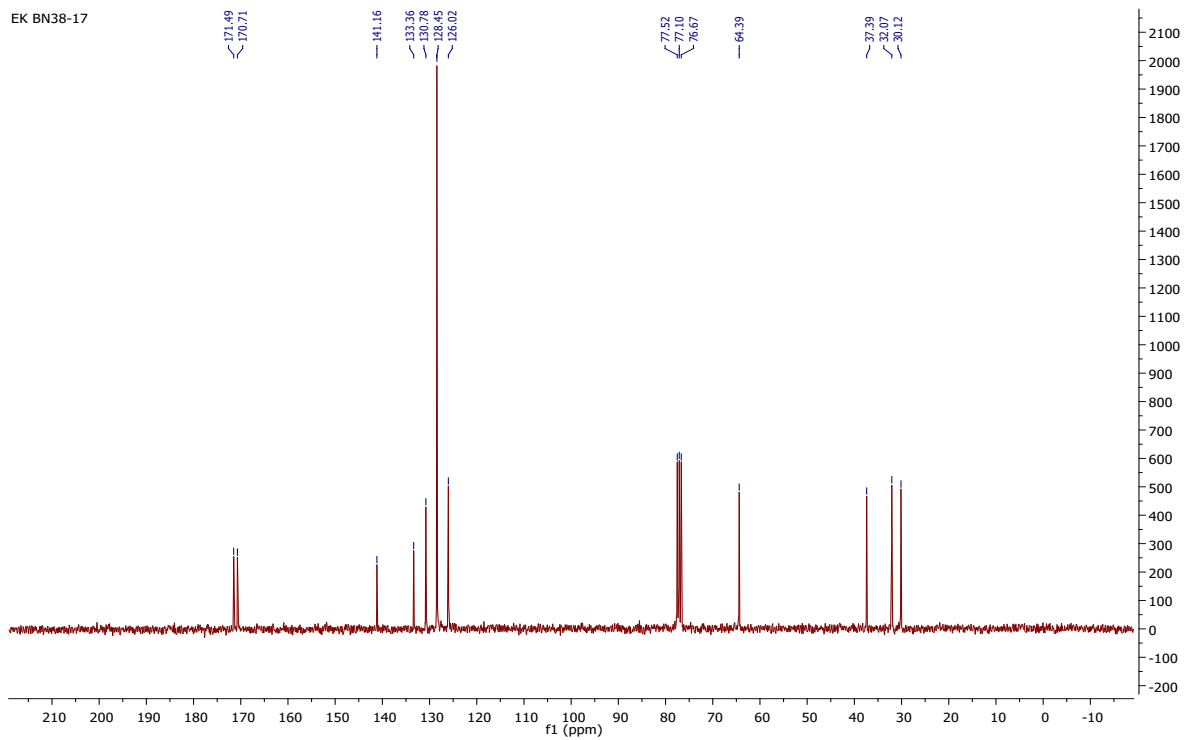
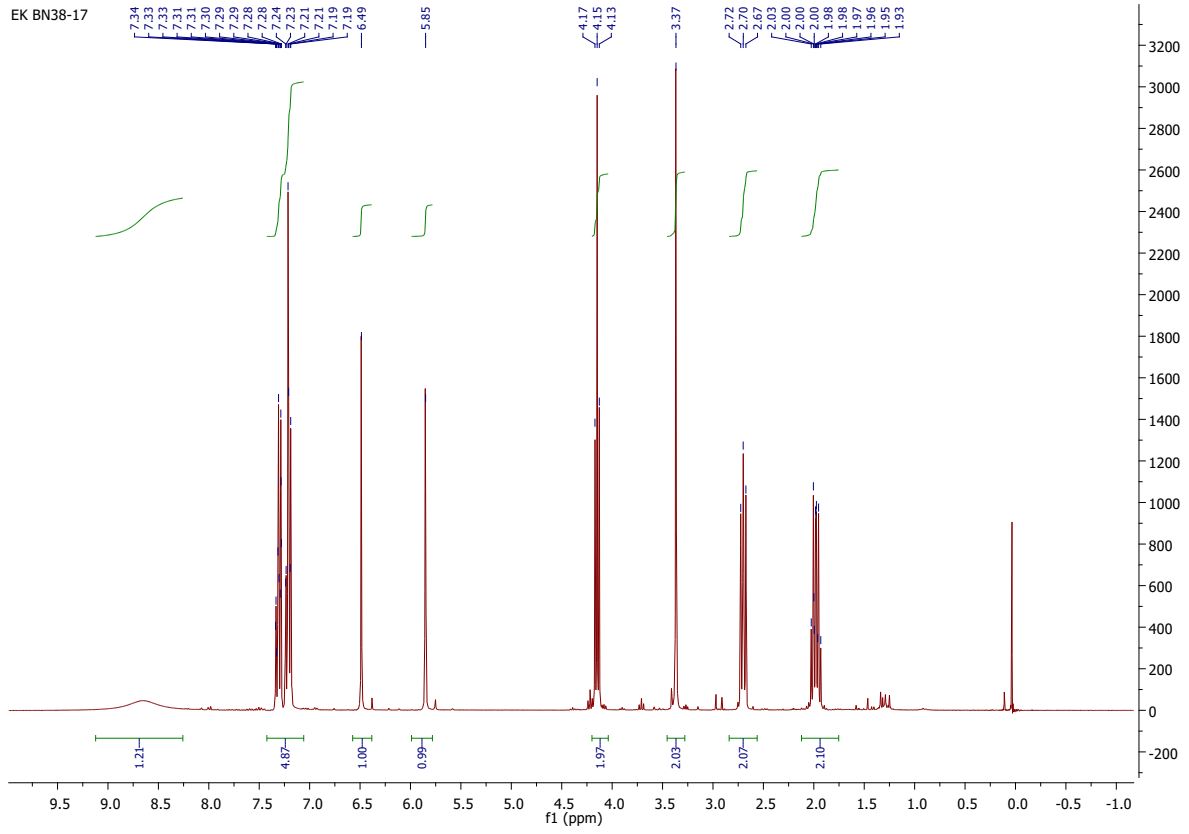
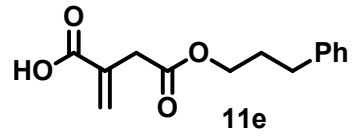
alg-BN29-17

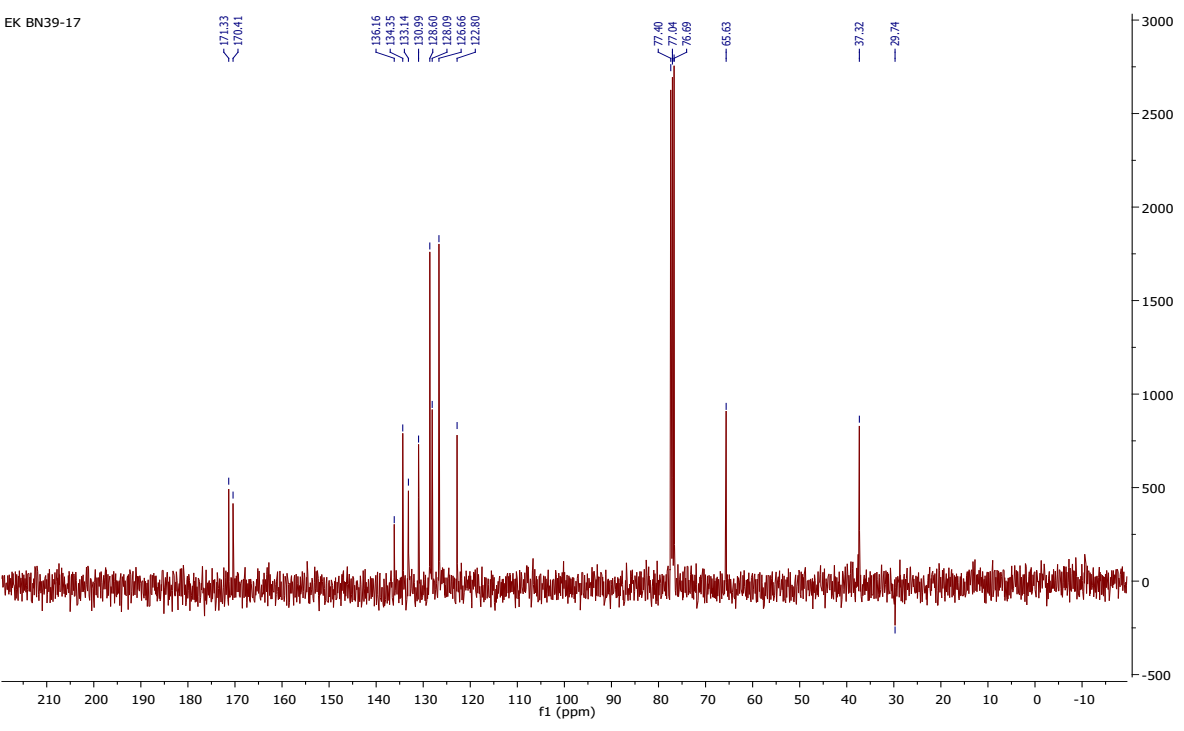
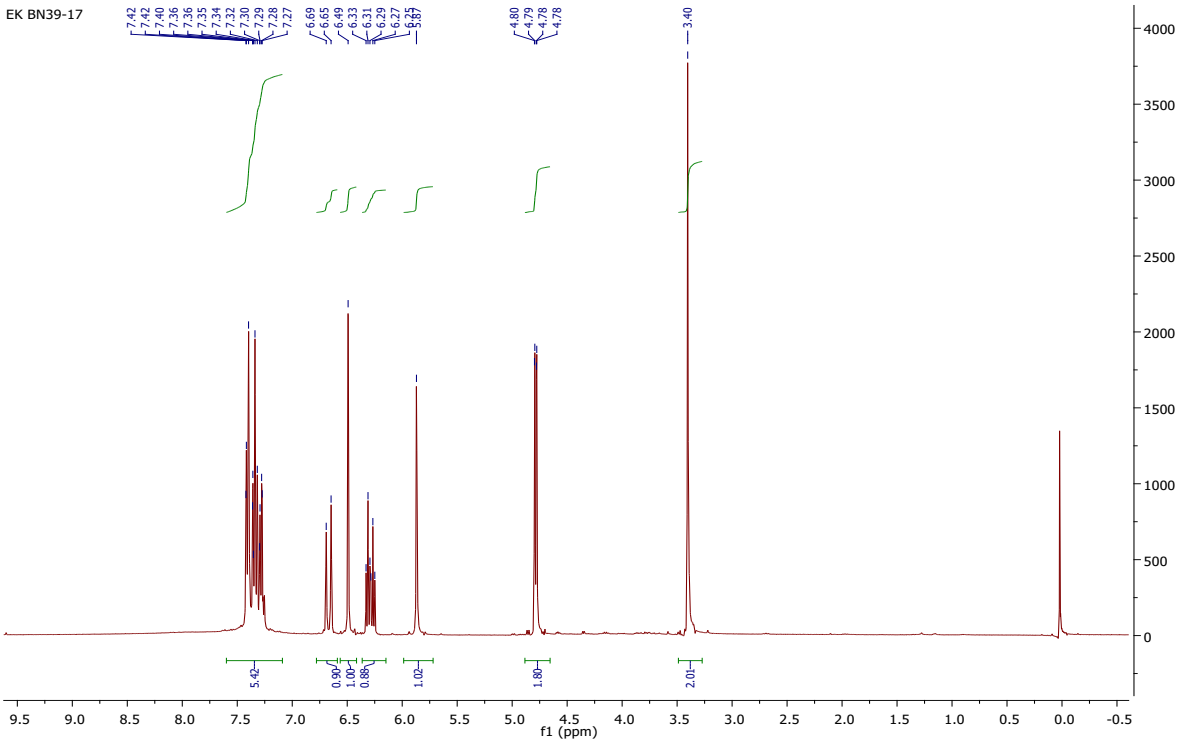
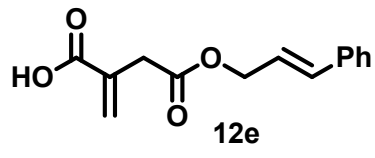


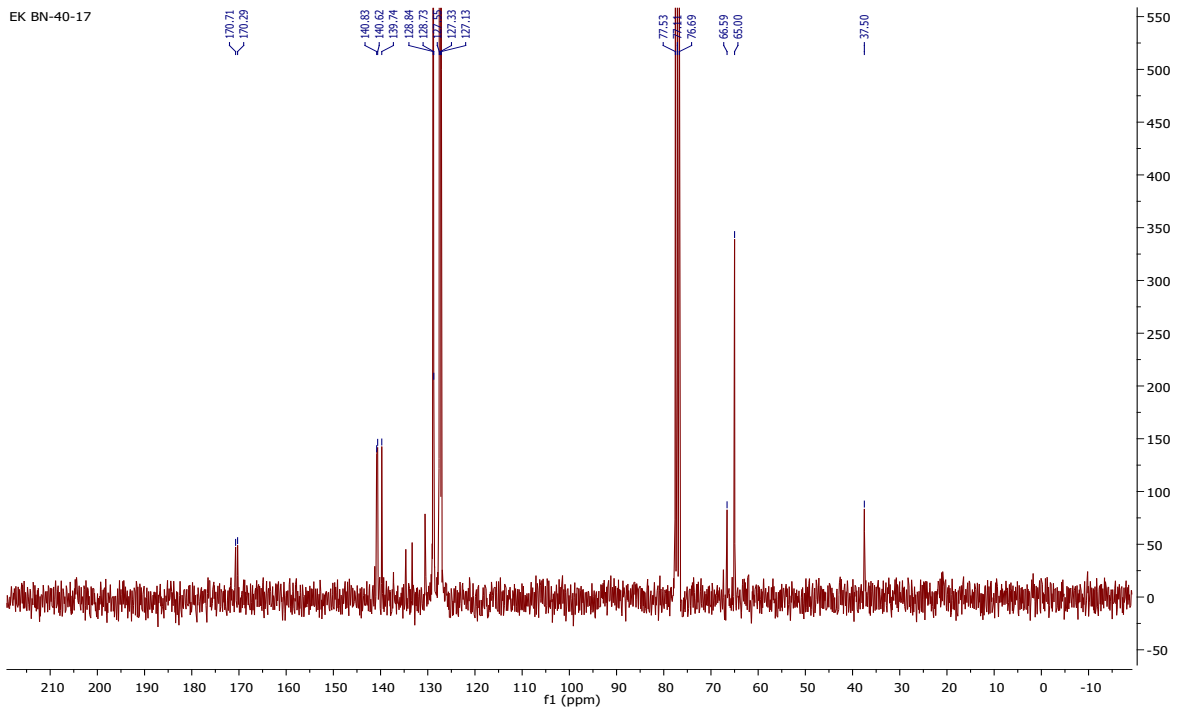
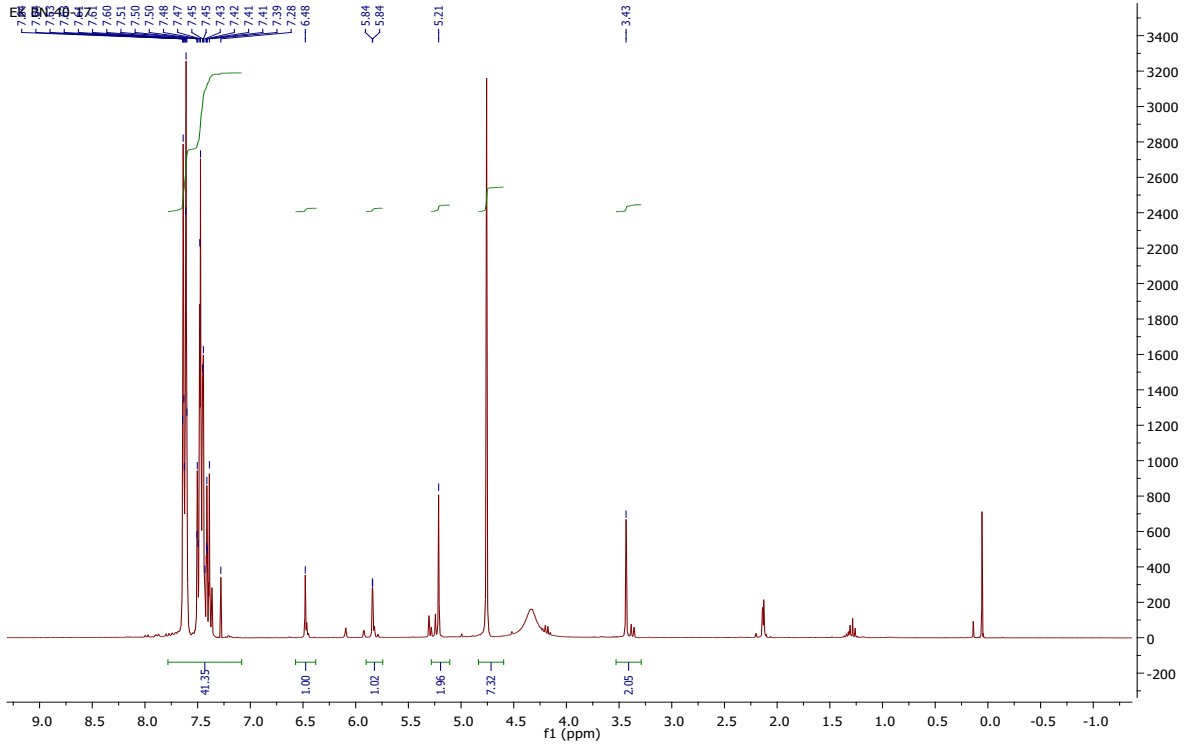
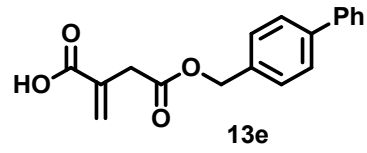
alg-BN29-17 c13

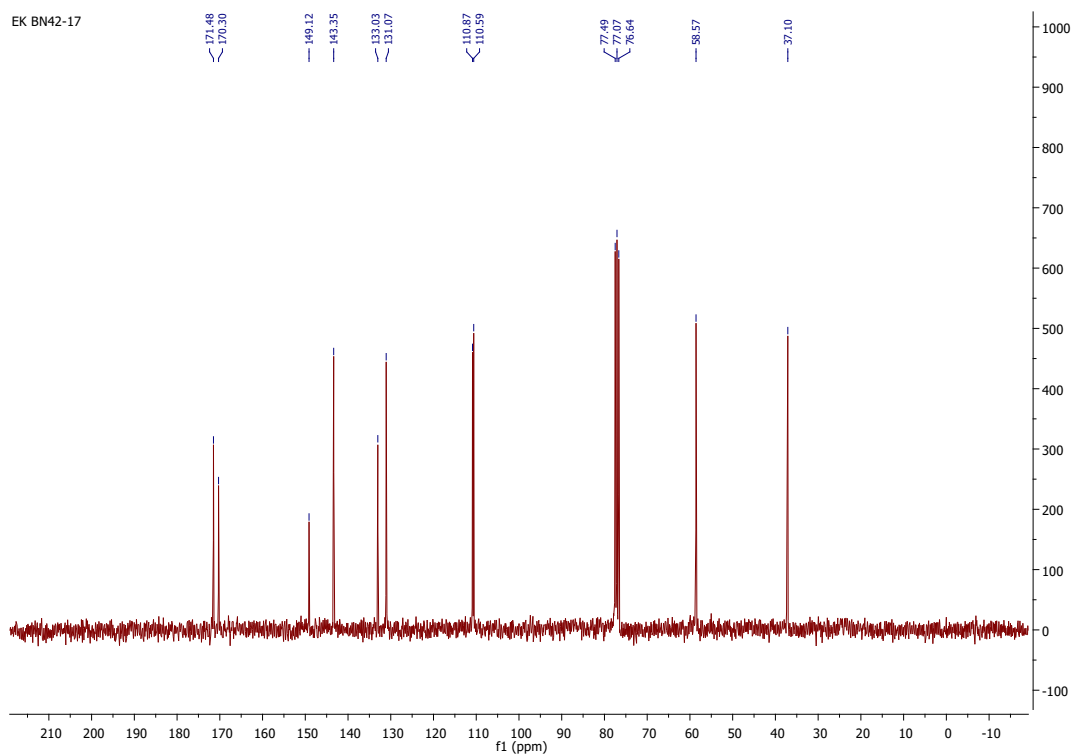
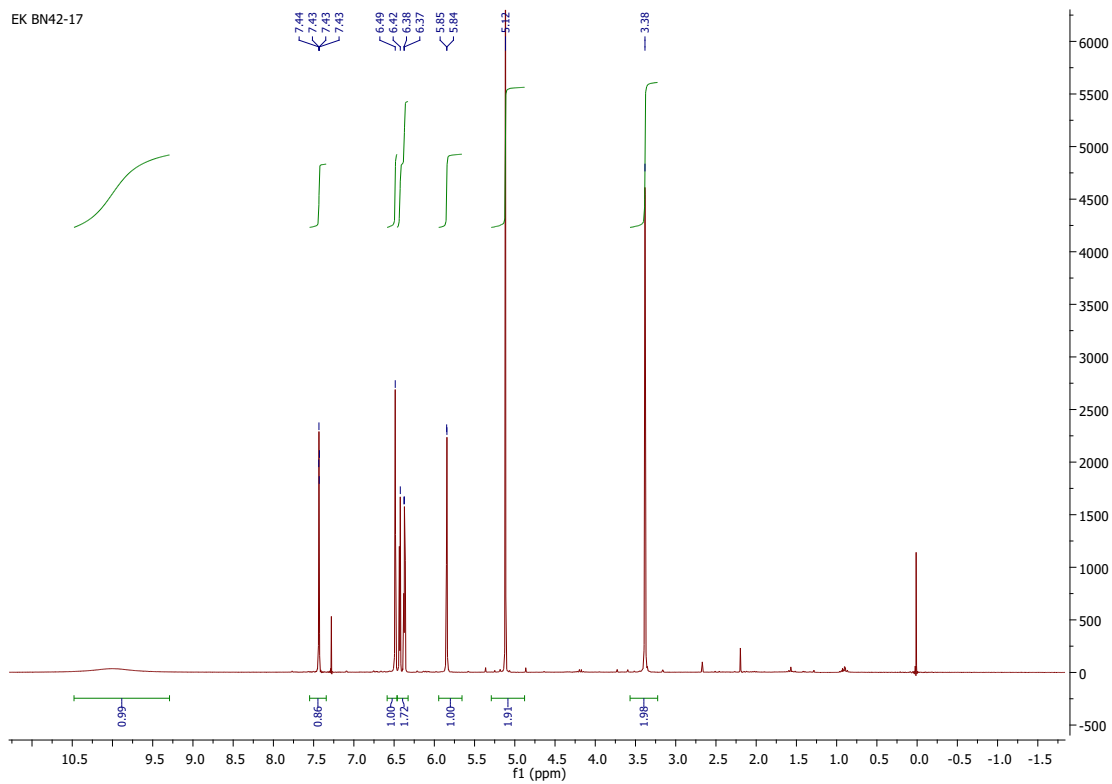
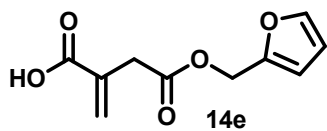


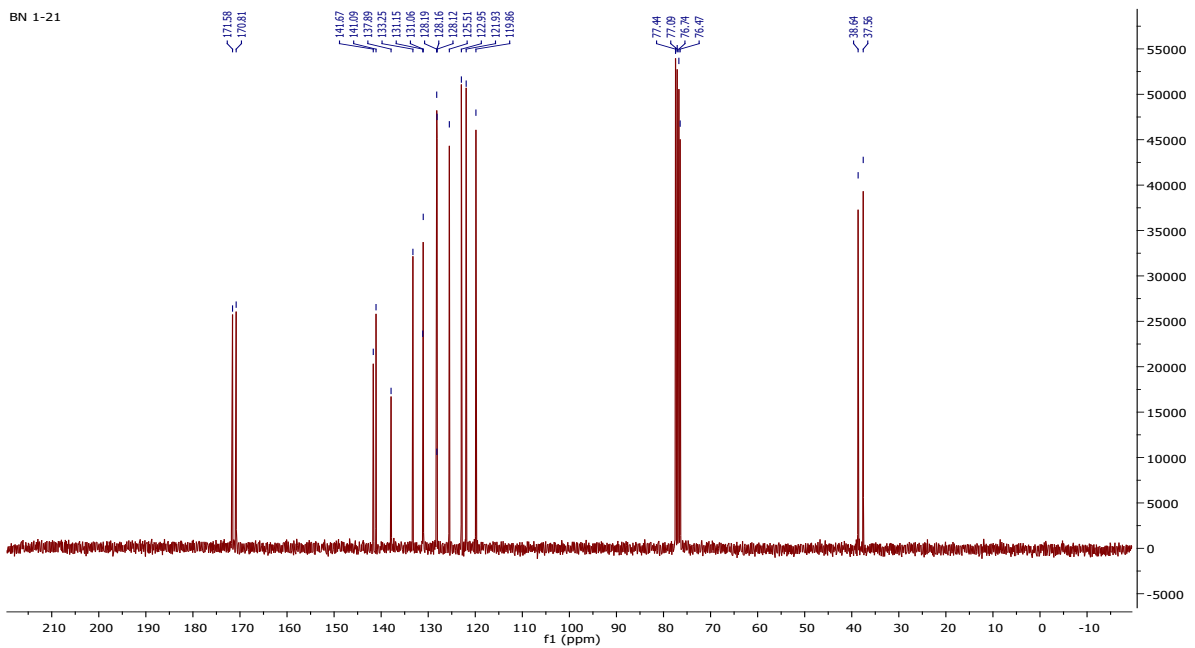
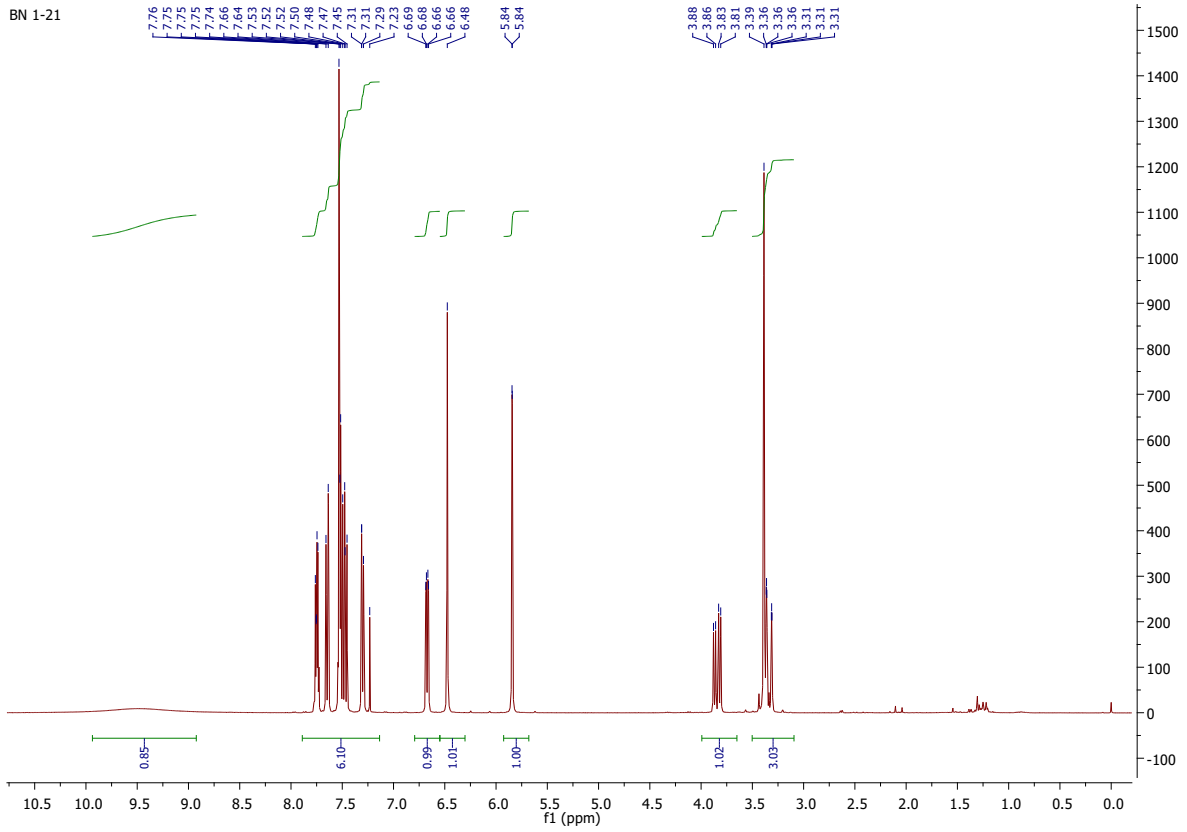
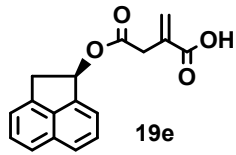


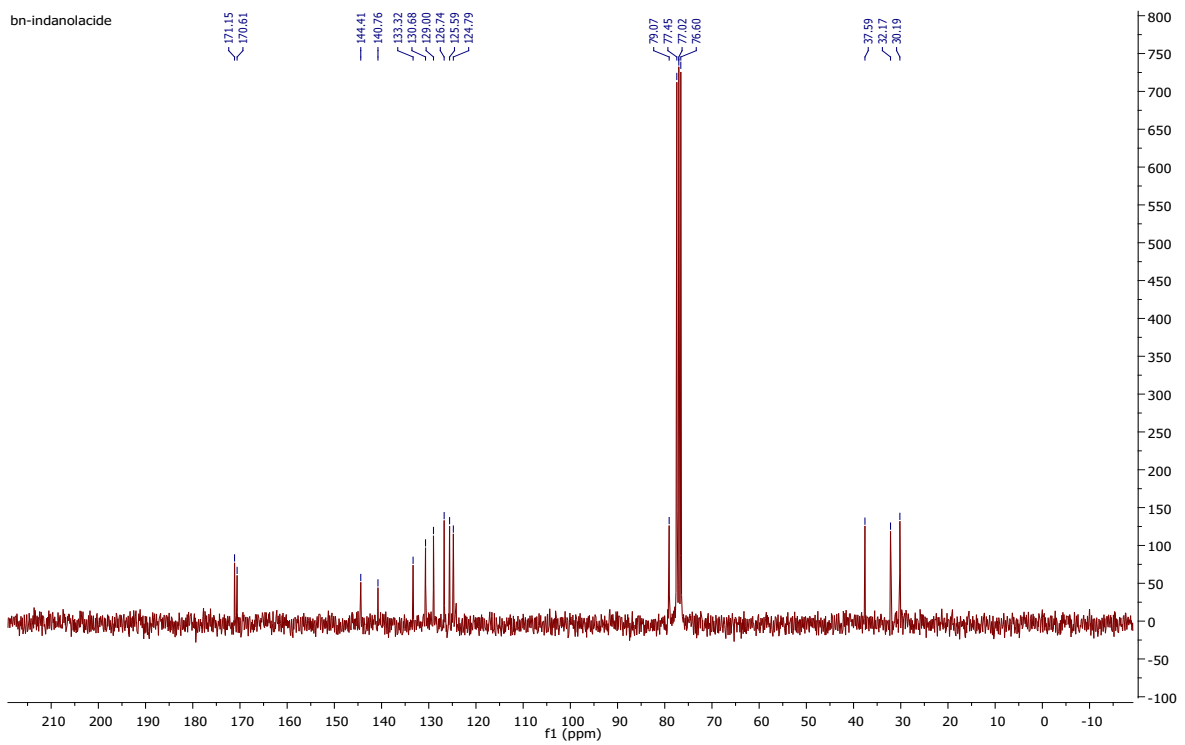
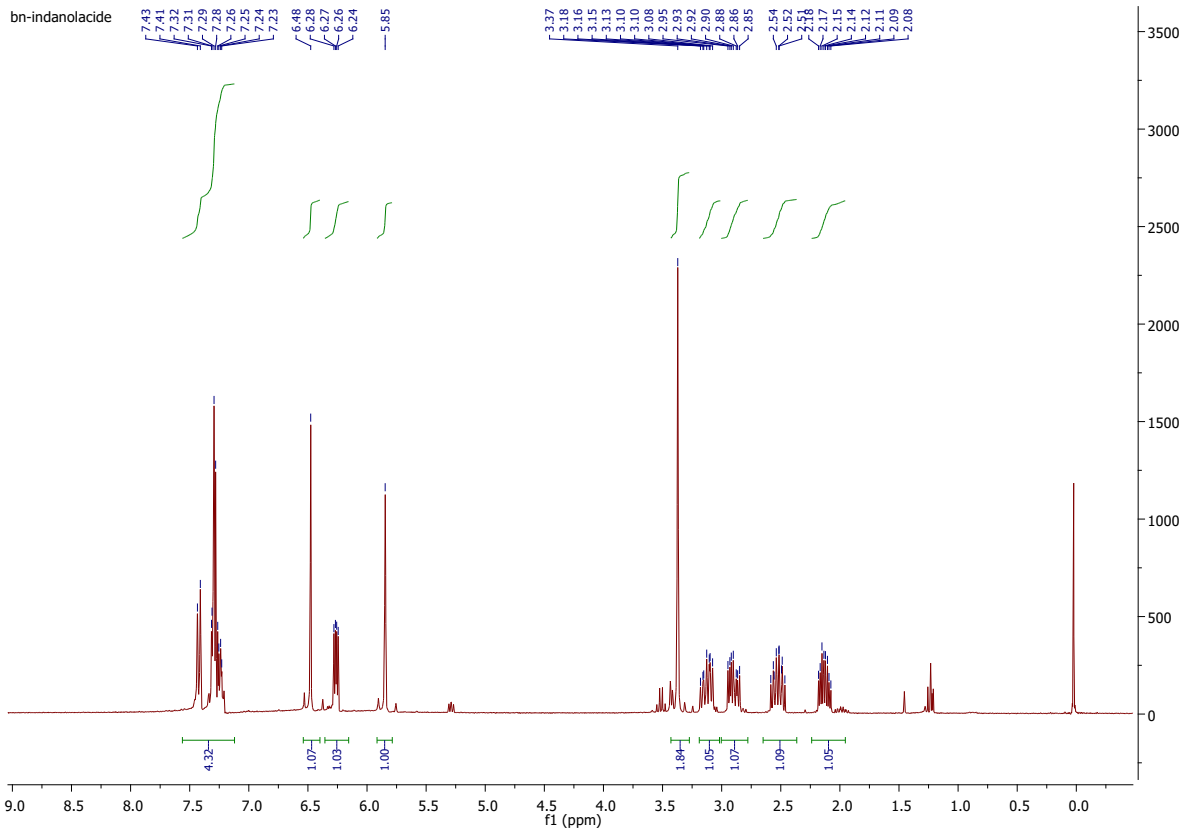
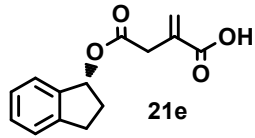


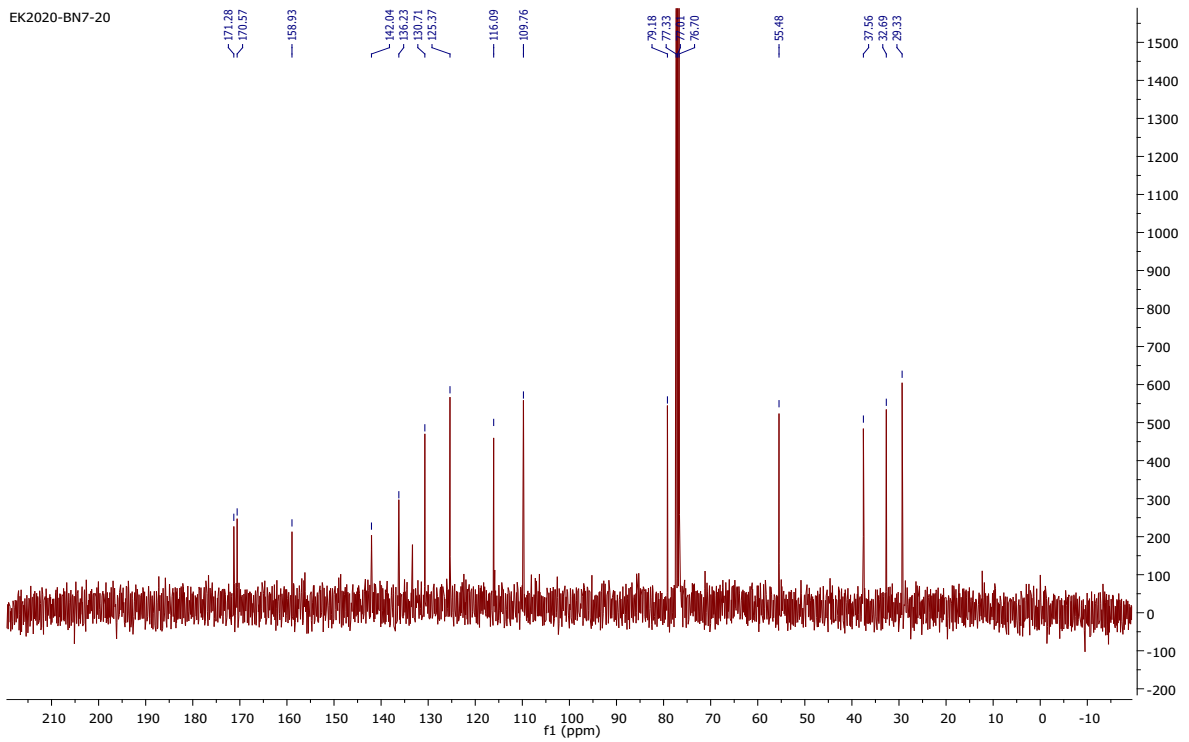
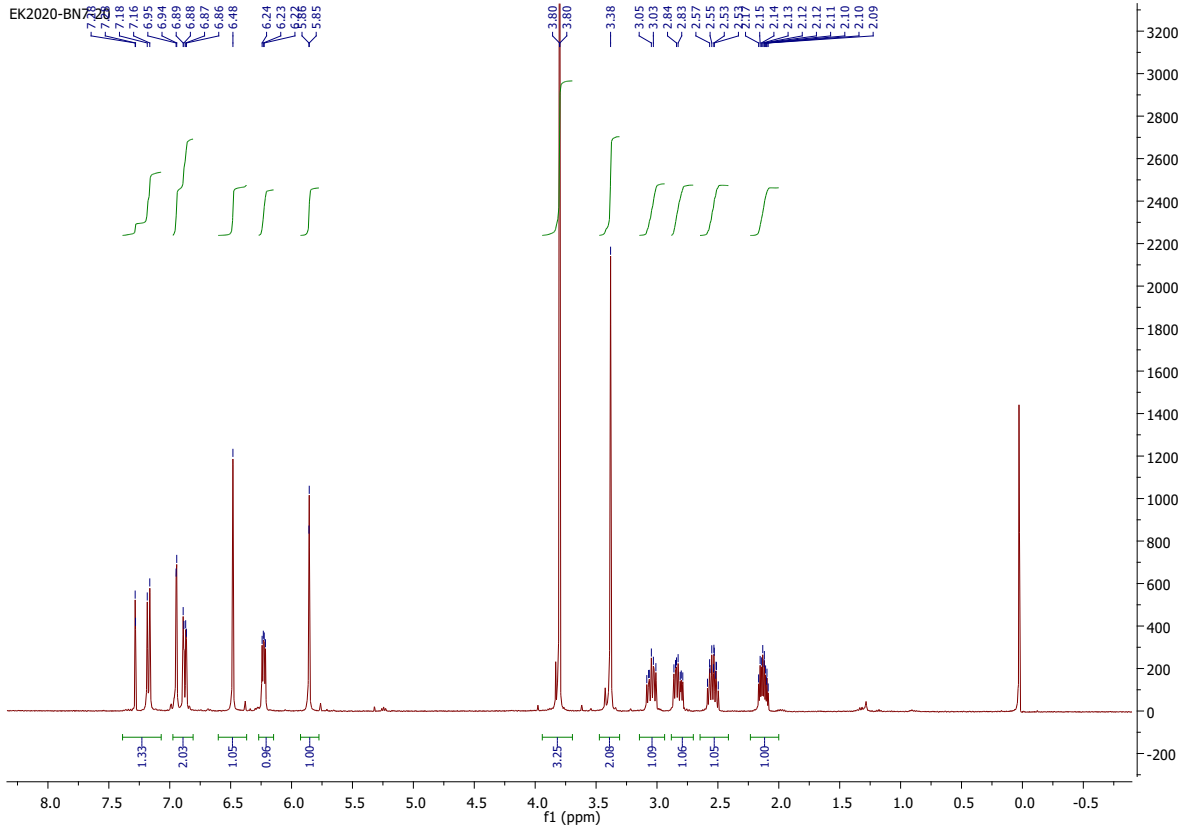
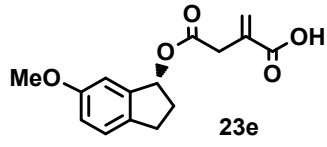






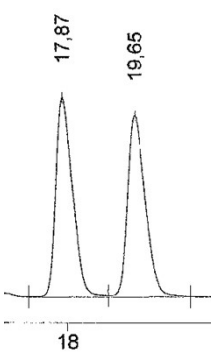

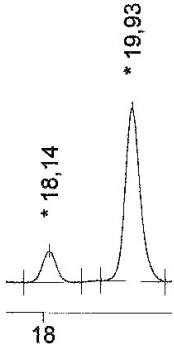
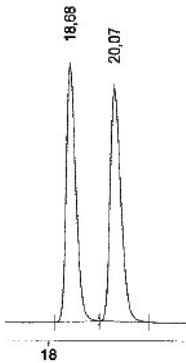
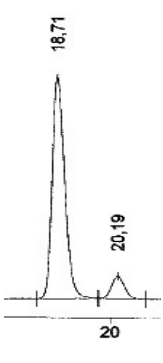
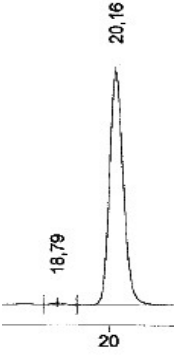
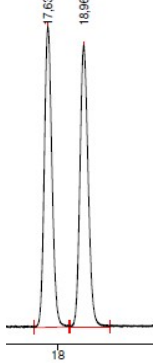
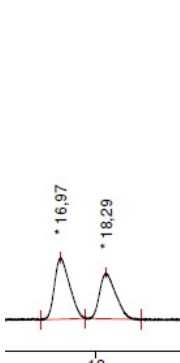
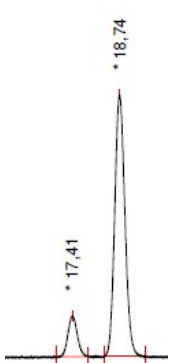

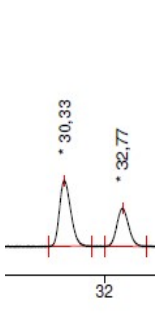
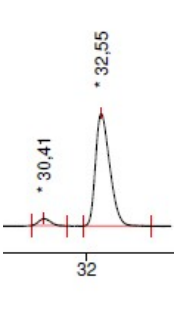


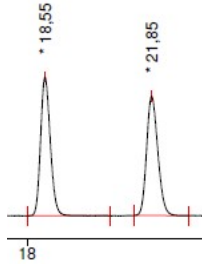
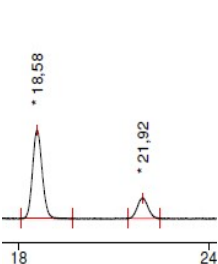
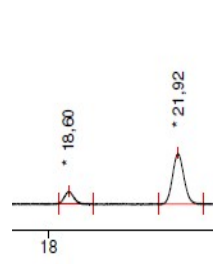




8 Chromatograms of PCL-catalyzed regio and enantioselective ring-opening of chiral secondary alcohol

Analysis's conditions	<i>racemic</i>	Unreacted alcohol $ee_{(S)}$	Reacted alcohol $ee_{(R)}$
<p><i>rac-16</i></p> <p>HPLC Chirale: colonne <i>Chiralpak IB</i>. $t_S=25.29$ min, $t_R=29.53$min. Eluent (v,v): hexane/iPrOH: 98/02; Flow rate: 0.5mL/min. $ee_S = 8\%$ (<i>S</i>)-16 $ee_p = 76\%$ (<i>R</i>)-16</p>			
<p><i>rac-17</i></p> <p>HPLC Chirale: colonne <i>Chiralpak OJ-H</i>. $t_S=19.97$ min, $t_R=20.32$ min. Eluent (v,v): hexane/iPrOH: 90/10; Flow rate: 0.8mL/min. $ee_S = 8.6\%$ (<i>S</i>)-17 $ee_p = 94.6\%$ (<i>R</i>)-17</p>			
<p><i>rac-18</i></p> <p>HPLC Chirale: colonne <i>Chiralpak IB</i>. $t_S=6.53$ min, $t_R=7.44$ min. Eluent (v,v): hexane/iPrOH: 75/25; Flow rate: 0.8mL/min. $ee_S = 11\%$ (<i>S</i>)-18 $ee_p = 79\%$ (<i>R</i>)-18</p>			
<p><i>rac 19</i></p> <p>HPLC Chirale: colonne <i>Chiralpak IA</i>. $t_R=25.81$ min, $t_S=27.71$min. Eluent (v,v): hexane/iPrOH: 95/05; Flow rate: 0.5mL/min. $ee_S = 99\%$ (<i>S</i>)-19 $ee_p = 95\%$ (<i>R</i>)-19</p>			

<p>rac-20</p> <p>HPLC Chirale: colonne <i>Chiralpak IA</i></p> <p>$t_S=17.87$ min, $t_R=19.65$ min.</p> <p>Eluant (v,v): hexane/iPrOH: 95/05;</p> <p>Flow rate: 0.5mL/min.</p> <p>$ee_S = 3.4\%$ (<i>S</i>)-20</p> <p>$ee_P = 72\%$ (<i>R</i>)-20</p>			
<p>rac-21</p> <p>HPLC Chirale: colonne <i>Chiralpak IA</i>.</p> <p>$t_S=18.68$ min, $t_R=20.07$ min.</p> <p>Eluant (v,v): hexane/iPrOH: 95/05;</p> <p>Flow rate: 0.5mL/min.</p> <p>$ee_S = 81\%$ (<i>S</i>)-21</p> <p>$ee_P = 98\%$ (<i>R</i>)-21</p>			
<p>rac-22</p> <p>HPLC Chirale: colonne <i>Chiralpak IA</i></p> <p>$t_S=17.63$ min, $t_R=18.96$ min.</p> <p>Eluant (v,v): hexane/iPrOH: 95/05;</p> <p>Flow rate: 0.5mL/min.</p> <p>$ee_S = 12\%$ (<i>S</i>)-22</p> <p>$ee_P = 75\%$ (<i>R</i>)-22</p>			
<p>rac-23</p> <p>HPLC Chirale: colonne <i>Chiralpak IA</i>.</p> <p>$t_S=30.6$ min, $t_R=32.87$ min. Eluant (v,v): hexane/iPrOH: 95/05; Flow rate: 0.5mL/min.</p> <p>$ee_S = 24\%$ (<i>S</i>)-23</p> <p>$ee_P = 90\%$ (<i>R</i>)-23</p>			

<p>rac-24</p> <p>HPLC Chirale: colonne <i>Chiralpak IA</i>. $t_S = 18.55$ min, $t_R = 21.85$ min. Eluant (v,v): hexane/<i>i</i>PrOH: 95/05; Flow rate: 0.5mL/min.</p> <p>ee_S = 59% (S)-24</p> <p>ee_P = 71% (R)-24</p>			
<p>rac-25</p> <p>HPLC Chirale: colonne <i>Chiralpak IA</i>. $t_S = 14.35$ min, $t_R = 16.31$ min. Eluant (v,v): hexane/<i>i</i>PrOH: 95/05; Flow rate: 0.5mL/min.</p> <p>ee_S = 45% (S)-25</p> <p>ee_P = 70% (R)-25</p>	