

Supplementary Information

Atropselective Synthesis of *N*-Aryl Pyridones via Dynamic Kinetic Resolution Enabled by Non-Covalent Interactions

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1 General Experimental

1.1 Naming and Numbering of Compounds

Systematic compound names are those generated by ChemBioDraw™ Ultra version 15.1.0.144 (Perkin Elmer) following IUPAC nomenclature.

1.2 Solvents and Reagents

Reactions were carried out under a nitrogen atmosphere in oven-dried glassware unless otherwise stated. Standard inert atmosphere techniques were used in handling all air- and moisture-sensitive reagents. Where necessary toluene and DMF (from commercial sources) were degassed prior to use by sparging with argon or nitrogen (15 min). Anhydrous and oxygen-free THF was obtained by distillation from Na/benzophenone. Other solvents and reagents were used directly as received from commercial suppliers.

1.3 Chromatography

Flash column chromatography was carried out using Fluorochem 60 40-63 micron silica gel. Thin-layer chromatography was carried out using Merck Kieselgel 60 F254 (230-400 mesh) fluorescent treated silica, visualized under UV light (254 nm) or by staining with aqueous potassium permanganate solution, ninhydrin or ceric ammonium molybdate solutions.

1.4 Analytical Techniques

¹H and ¹³C NMR spectra were recorded using a Bruker Ascend 600 or Avance 400 MHz spectrometer running TopSpin™ 3.6.3 software and are quoted in ppm for measurement against tetramethylsilane. Where no tetramethylsilane was present, spectra are referenced relative to the residual non-deuterated solvent peaks. Unless otherwise stated spectra were acquired at 298 K. Topspin™ was used for processing and viewing NMR data. Chemical shifts (δ) are given in parts per million (ppm), and coupling constants (J) are given in Hertz (Hz). The ¹H NMR spectra are reported as follows: δ / ppm (number of protons, multiplicity, coupling constant J / Hz (where appropriate), assignment). Multiplicity is abbreviated as follows: s = singlet, br = broad, d = doublet, t = triplet, q = quartet, m = multiplet. The numbering scheme used for NMR assignment is arbitrary and does not follow any particular convention. The ¹³C NMR spectra are reported in δ / ppm. Where necessary or appropriate, two-dimensional (COSY, HSQC, HMBC, NOESY or ROESY) NMR experiments were used to assist the assignment of signals in the ¹H and ¹³C NMR spectra. In some cases, complete assignment of spectra was not possible (in particular, aromatic CHs corresponding to multiple phenyl groups overlapped significantly); in these cases only a partial assignment is reported.

High-performance liquid chromatography (HPLC) was conducted using an Agilent 1220 Infinity II instrument using an isocratic acetonitrile/water eluent mixture.

Liquid chromatography-mass spectrometry (LCMS) analysis was conducted using an instrument comprising an Agilent 1260 HPLC (equipped with Infinity II quaternary pump, vial sampler, integrated column compartment and variable wavelength detector) and 6125C MSD single quadrupole mass spectrometer. Samples were analysed using an Agilent Infinitylab poroshell 120 column (2.7 μ m, 2.1 x 150 mm) under an acetonitrile/water gradient with 0.1% HCOOH additive.

Infra-red (IR) spectra were recorded on an Agilent Cary 630 spectrometer equipped with an attenuated total reflectance (ATR) accessory. Samples were deposited on the ATR as a thin film. Only selected maximum absorbances (v_{\max}) of the most intense peaks are reported (cm^{-1}).

High resolution mass spectra (HRMS) were recorded by Analytical Services and Environmental Projects (ASEP) at Queen's University Belfast on a Waters LCT Premier ToF mass spectrometer using the electrospray ionisation (ESI) technique.

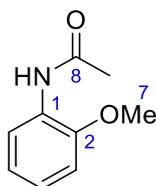
Optical rotations were recorded at the sodium D-line (589 nm) using a Perkin Elmer 341 polarimeter at a temperature of 20 °C and are reported in degrees using concentrations (*c*) in g·100 mL⁻¹. Reported values are the average of eight readings.

Melting points were determined for compounds where a preparative recrystallization was carried out. These were acquired on a Stuart SMP10 digital melting point apparatus. Values are given in °C and are uncorrected.

2 Experimental Procedures and Characterization Data

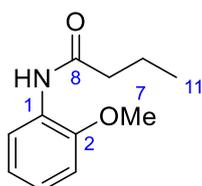
2.1 Acetamides (1)

N-(2-Methoxyphenyl)acetamide (1a)



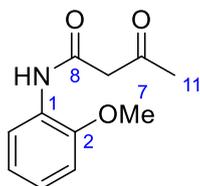
To a solution of 2-aminoanisole (4.58 g, 40.6 mmol, 1.0 equiv.) in dichloromethane (150 mL) was added Ac₂O (4.6 mL, 48.7 mmol, 1.2 equiv.), flushed with Ar and was stirred for 20 mins at RT. The reaction mixture was quenched with saturated NaHCO₃ aqueous solution (ca. 3 x 10 mL/mmol), extracted with dichloromethane (ca. 3 x 10 mL/mmol) and dried over anhydrous magnesium sulfate. The solvent was removed *in vacuo* affording **1a** as an off-white fluffy solid (6.51 g, 97%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 8.35 (1H, dd, *J* = 8.1, 1.3 Hz, H6), 7.76 (1H, s, NH), 7.03 (1H, td, *J* = 7.8, 1.5 Hz, H5), 6.95 (1H, td, *J* = 7.9, 1.3 Hz, H4), 6.87 (1H, dd, *J* = 7.9, 1.0 Hz, H3), 3.87 (3H, s, H7), 2.20 (3H, s, H9). **¹³C NMR** (101 MHz CDCl₃): δ_C = 168.1 (C8), 147.6 (C1), 127.7 (C2), 123.6 (C6), 121.1 (C5), 118.8 (C4), 109.9 (C3), 55.7 (C7), 25.0 (C9). **IR**: ν_{max} (thin film): 3246, 2967, 1654, 1595, 1461, 1248, 1113, 1021, 745, 708 cm⁻¹. **HRMS** (ES⁺): found 166.0868; C₉H₁₂NO₂, [M+H]⁺ requires 166.0868. The data obtained are consistent with those reported in the literature.¹

N-(2-Methoxyphenyl)butyramide (1b)



To a solution of *o*-anisidine (3.55 g, 28.9 mmol, 1.0 equiv.) in pyridine (35 mL) was added dropwise a solution of butyryl chloride (3.59 mL, 34.6 mmol, 1.2 equiv.) in THF (175 mL) at 0 °C. The mixture was stirred at 0 °C for 3 hours. The reaction mixture was reduced *in vacuo* and crude residue extracted with EtOAc (100 mL), washed with H₂O (75 mL), followed by 1M HCl aqueous solution (75 mL), saturated NaHCO₃ aqueous solution (75 mL) and saturated NaCl aqueous solution (75 mL). The organic extracts were dried over anhydrous magnesium sulfate and reduced *in vacuo* to an orange residue. The crude product was purified by flash column chromatography (silica gel 50 g, EtOAc/PE 10 → 20%) affording **1b** as a yellow oil (5.11 g, 91%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 8.40 (1H, dd, *J* = 7.9, 1.3 Hz, H6), 7.76 (1H, s, NH), 7.03 (1H, td, *J* = 7.6, 1.6 Hz, H5), 6.96 (1H, td, *J* = 7.8, 1.5 Hz, H4), 6.87 (1H, dd, *J* = 8.0, 1.5 Hz, H3), 3.88 (3H, s, H7), 2.37 (2H, t, *J* = 7.3 Hz, H9), 1.77 (2H, sex, *J* = 7.4 Hz, H10), 1.01 (3H, t, *J* = 7.4 Hz, H11). **¹³C NMR** (101 MHz CDCl₃): δ_C = 171.1 (C8), 147.6 (C1), 127.8 (C2), 123.4 (C6), 121.1 (C5), 119.7 (C4), 109.8 (C3), 55.7 (C7), 40.0 (C9), 19.11 (C10), 13.8 (C11). **IR**: ν_{max} (thin film): 3317, 2959, 1669, 1599, 1520, 1457, 1248, 1025, 745 cm⁻¹. **HRMS** (ES⁺): found 194.1185; C₁₁H₁₆NO₂, [M+H]⁺ requires 194.1181. The data obtained are consistent with those reported in the literature.²

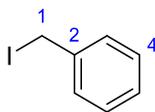
N-(2-Methoxyphenyl)-3-oxobutanamide (**1c**)



A neat mixture of 2-aminoanisole (2.72 g, 22 mmol) and ethyl acetoacetate (5.6 mL, 44 mmol) and potassium *tert*-butoxide (240 mg, 2.2 mmol) was heated to 130 °C. After 2 h the reaction mixture was cooled to room temperature and the crude residue was purified by flash column chromatography (silica gel, 0% → 20% EtOAc/dichloromethane) to afford the product **1c** as an off-white solid (1.94 g, 42%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 9.22 (1H, s, NH), 8.32 (1H, dd, *J* = 7.9, 1.5 Hz, H6), 7.06 (1H, td, *J* = 8.0, 1.5 Hz, H5), 6.95 (1H, td, *J* = 8.0, 1.2 Hz, H4), 6.89 (1H, dd, *J* = 8.1, 1.0 Hz, H3), 3.92 (3H, s, H7), 3.60 (2H, s, H9), 2.33 (3H, s, H11). **¹³C NMR** (101 MHz CDCl₃): δ_C = 204.4 (C10), 163.2 (C8), 148.3 (C1), 127.4 (C2), 124.1 (C6), 121.0 (C5), 120.1 (C4), 110.1 (C3), 55.8 (C7), 50.9 (C9), 25.0 (C11). **IR**: ν_{max} (thin film): 3295, 2940, 1714, 1669, 1599, 1524, 1461, 1356, 1252, 1118, 1025, 745 cm⁻¹. **HRMS** (ES⁺): found 208.0965; C₁₁H₁₄NO₃, [M+H]⁺ requires 208.0974. The data obtained are consistent with those reported in the literature.³

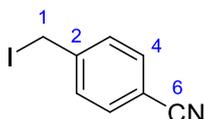
2.2 Benzyl Iodides

(Iodomethyl)benzene (**S1**)



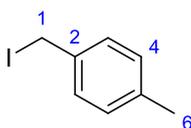
NaI (4.50 g, 30.0 mmol, 2.0 equiv.) was dissolved in acetone (20 mL) at 0 °C. To the solution, was added BnBr (1.80 mL, 15.0 mmol, 1.0 equiv.) and the resulting mixture was stirred at 0 °C for 6 h. The reaction mixture was washed with brine (50 mL) and extracted with Et₂O (3 x 50 mL), dried over anhydrous magnesium sulfate and the solvent was removed *in vacuo* giving a pink oil. PE (*ca.* 1 mL) was added and crude product placed on ice as pink crystals formed. Mother liquor was removed via pipette at 0 °C affording **S1** as a pink solid (3.06 g, 94%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.39-7.36 (2H, m, H3), 7.31-7.21 (3H, m, H4, H5), 4.46 (2H, s, H1). **¹³C NMR** (101 MHz CDCl₃): δ_C = 139.3 (C2), 128.8 (C3), 128.7 (C4), 127.9 (C5), 5.7 (C1). **IR**: ν_{max} (thin film): 3026, 1699, 1453, 1211, 1155, 749, 689 cm⁻¹. **HRMS** (ES⁺): found 218.9705; C₇H₈I, [M+H]⁺ requires 218.9796. The data obtained are consistent with those reported in the literature.⁴

4-(Iodomethyl)benzonitrile (**S2**)



NaI (3.0 g, 20.0 mmol, 2.0 equiv.) was dissolved in acetone (12 mL) at 0 °C. To the solution, was added 4-cyanobenzyl bromide (1.96 g, 10.0 mmol, 1.0 equiv.) and the resulting mixture was stirred at 0 °C for 4 h. The reaction mixture was washed with brine (30 mL) and extracted with Et₂O (3 x 30 mL), dried over anhydrous magnesium sulfate and the solvent was removed *in vacuo*. The residue was purified by trituration (petroleum ether) to afford **S2** as a pale yellow solid (2.26 g, 93%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.60-7.57 (2H, m, H4), 7.48-7.45 (2H, m, H3), 4.43 (2H, s, H1). **¹³C NMR** (101 MHz CDCl₃): δ_C = 144.7 (C2), 132.6 (2C, C4), 129.7 (2C, C3), 118.5 (C6), 111.6 (C5), 2.8 (C1). **IR**: ν_{max} (thin film): 2221, 1707, 1602, 1505, 1159, 842, 734 cm⁻¹. **HRMS** (ES⁺): found 243.9889; C₈H₇IN, [M+H]⁺ requires 243.9798. The data obtained are consistent with those reported in the literature.⁵

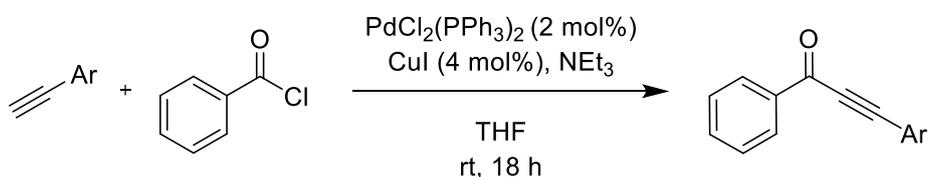
1-(Iodomethyl)-4-methylbenzene (S3)



NaI (3.0 g, 20.0 mmol, 2.0 equiv.) was dissolved in acetone (12 mL) at 0 °C. To the solution, was added 4-methylbenzyl bromide (1.85 g, 10.0 mmol, 1.0 equiv.) and the resulting mixture was stirred at 0 °C for 4 h. The reaction mixture was washed with brine (30 mL) and extracted with Et₂O (3 x 30 mL), dried over anhydrous magnesium sulfate and the solvent was removed *in vacuo*. The crude residue was purified by recrystallization from hot petroleum ether to afford **S3** (1.41 g, 68%) as a white crystalline solid. **M.p.** = 46-48 °C (petroleum ether), **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.28 (2H, d, *J* = 8.1 Hz, H3), 7.11 (2H, d, *J* = 7.8 Hz, H4), 4.46 (2H, s, H1), 2.32 (3H, s, H6). **¹³C NMR** (101 MHz CDCl₃): δ_C = 137.8 (C2), 136.3 (C5), 129.6 (2C, C4), 128.7 (2C, C3), 21.3 (C6), 6.2 (C1). **IR**: ν_{max} (thin film): 2911, 1610, 1513, 1427, 1207, 1148, 812 cm⁻¹. **HRMS** (ES⁺): found 232.9787; C₈H₁₀I, [M+H]⁺ requires 232.9782. The data obtained are consistent with those reported in the literature.⁵

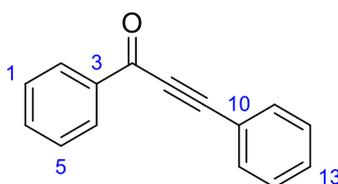
2.3 Alkynones (2)

General Procedure A (Preparation of Prop-2-yn-1-ones)



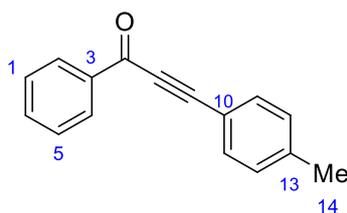
A flask was charged with benzoyl chloride, PdCl₂(PPh₃)₂, and Et₃N, THF was added, and the resulting solution stirred for 10 min after which CuI was added changing the colour (substrate dependent) upon stirring for an additional 10 min. Arylacetylene was added and the flask was sealed and left to stir at room temperature for 18 h. The reaction mixture was quenched with 0.1 N HCl (*ca.* 2 x 10 mL/mmol), washed with saturated NH₄Cl aqueous solution (*ca.* 10 mL/mmol), extracted with dichloromethane (*ca.* 3 x 10 mL/mmol), and dried over anhydrous magnesium sulfate. The solvent was removed *in vacuo* resulting in a crude solid which was purified by column chromatography (silica gel, 2.5% → 5% Et₂O/PE) affording the desired alkynone.

1,3-Diphenylprop-2-yn-1-one (2a)



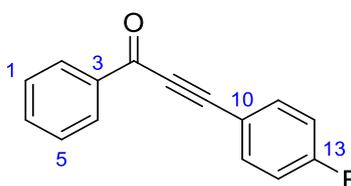
Prepared according to **General Procedure A** using benzoyl chloride (3.18 g, 27.3 mmol, 1.2 equiv.), PdCl₂(PPh₃)₂ (319 mg, 460 μmol, 2.0 mol%) and Et₃N (3.63 mL, 27.3 mmol, 1.2 equiv.), THF (40 mL), CuI (174 mg, 0.91 mmol, 4.0 mol%) and phenylacetylene (2.50 g, 22.8 mmol, 1.0 equiv.) **2a** was obtained as a yellow oil (4.21 g, 90%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 8.23 (2H, dd, *J* = 8.6, 1.4 Hz, PhH), 7.69 (2H, m, PhH), 7.64 (1H, t, *J* = 7.6 Hz, PhH), 7.54-7.47 (3H, m, PhH) 7.45-7.41 (2H, m, PhH). **¹³C NMR** (101 MHz CDCl₃): δ_C = 178.0, 136.9, 134.1, 133.1 (2C), 130.8, 129.6 (2C), 128.7 (2C), 128.6 (2C), 120.1, 93.1, 86.9. **IR**: ν_{max} (thin film): 3060, 2195, 1636, 1490, 1446, 1312, 1282, 1207, 756, 689 cm⁻¹. **HRMS** (ES⁺): found 207.0815; C₁₅H₁₁O, [M+H]⁺ requires 207.0810. The data obtained are consistent with those reported in the literature.⁶

1-Phenyl-3-(*p*-tolyl)prop-2-yn-1-one (**2b**)



Prepared according to **General Procedure A** using benzoyl chloride (1.32 mL, 11.4 mmol, 1.5 equiv.), PdCl₂(PPh₃)₂ (105 mg, 150 μmol, 2.0 mol%) and Et₃N (1.30 mL, 9.4 mmol, 1.25 equiv.), THF (16 mL), CuI (29 mg, 0.15 mmol, 2.0 mol%) and 4-methylphenylacetylene (0.96 mL, 7.58 mmol, 1.0 equiv.) **2b** was obtained as a yellow solid (1.39 g, 84%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 8.24-8.22 (2H, m, PhH), 7.65-7.58 (3H, m, PhH, ArH), 7.52 (2H, t, *J* = 7.8 Hz, ArH), 7.23 (2H, d, *J* = 7.9 Hz, ArH), 2.41 (3H, s, CH₃). **¹³C NMR** (101 MHz CDCl₃): δ_C = 178.0, 141.5, 137.0, 134.0, 133.1 (2C), 129.6 (2C), 129.5 (2C), 128.6 (2C), 117.1, 93.8, 86.8, 21.8. **IR**: ν_{max} (thin film): 2191, 1632, 1599, 1312, 1170, 1006, 812, 697 cm⁻¹. **HRMS** (ES⁺): found 221.0966; C₁₆H₁₃O, [M+H]⁺ requires 221.0966. The data obtained are consistent with those reported in the literature.⁷

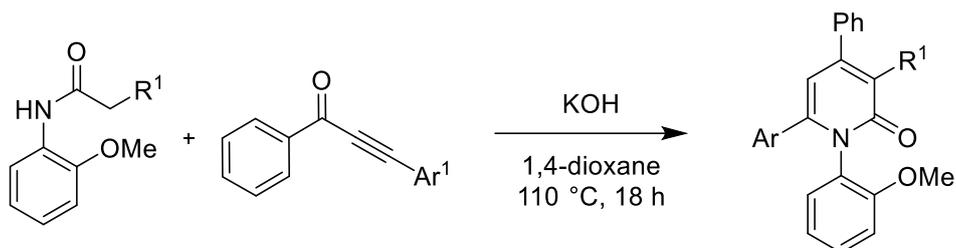
3-(4-Fluorophenyl)-1-phenylprop-2-yn-1-one (**2c**)



Prepared according to **General Procedure A** using benzoyl chloride (0.58 mL, 5.0 mmol, 1.5 equiv.), PdCl₂(PPh₃)₂ (46 mg, 66 μmol, 2.0 mol%), Et₃N (0.69 mL, 4.9 mmol, 1.5 equiv.), THF (8 mL), CuI (13 mg, 66 μmol, 2.0 mol%) and 4-fluorophenyl acetylene (398 mg, 3.31 mmol, 1.0 equiv.). **2c** was obtained as a lime green foam (598 mg, 81%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 8.21 (2H, dd, *J* = 8.5, 1.5 Hz, PhH), 7.72-7.62 (3H, m, PhH, ArH), 7.52 (2H, t, *J* = 7.8 Hz, ArH), 7.13 (2H, t, *J* = 8.5 Hz, ArH). **¹³C NMR** (101 MHz CDCl₃): δ_C = 177.9, 164.1 (d, ¹J_{CF} = 250.1 Hz), 136.8, 135.4 (2C, d, ³J_{CF} = 8.9 Hz), 134.2, 129.6 (2C), 128.7 (2C), 116.3 (d, ⁴J_{CF} = 3.4 Hz), 116.3 (2C, d, ²J_{CF} = 22.4 Hz), 92.0, 86.8. **¹⁹F NMR** (376 MHz, CDCl₃): δ_F = -106.0. **IR**: ν_{max} (thin film): 2199, 1640, 1595, 1233, 1010, 834, 693 cm⁻¹. **HRMS** (ES⁺): found 225.0712; C₁₅H₁₀FO, [M+K]⁺ requires 225.0716.⁷

2.4 2'-Methoxyphenyl-2-pyridones

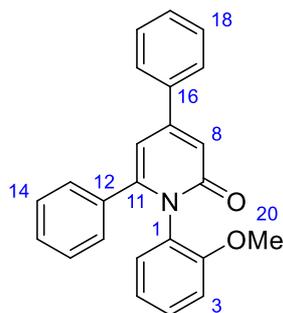
General Procedure B (*Preparation of 2'-methoxyphenyl-2-pyridones*)



This reaction was conducted by analogy to the method reported by Tang and Pen.⁸ An oven-dried microwave tube was charged with prop-2-yn-1-one derivatives (1.0 equiv.), 1,4-dioxane (ca. 0.3 M), amide (1.0 equiv.) and KOH (1.0 equiv., freshly ground up *via* pestle and mortar). The tube was sealed and stirred at 80 °C for 24-36 h, then allowed to cool to RT. The reaction mixture was diluted with H₂O (ca. 5 mL/mmol) and extracted with EtOAc (ca. 3 x 5 mL/mmol) and dried

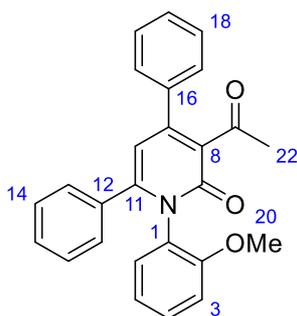
over anhydrous magnesium sulfate. The solvent was removed *in vacuo* resulting in a crude oil which was purified by column chromatography (silica gel, 2.5% → 5% MeOH/dichloromethane) affording the desired product.

1-(2-Methoxyphenyl)-4,6-diphenylpyridin-2(1H)-one (S4)



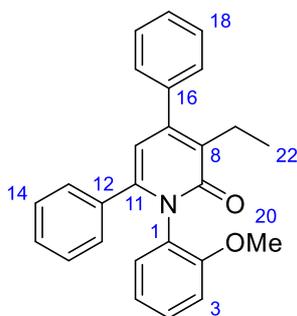
Prepared according to **General Procedure B** using alkynone **2a** (1.77 g, 8.60 mmol, 1.2 equiv.), amide **1a** (1.18 g, 7.17 mmol, 1.0 equiv.), KOH (402 mg, 7.17 mmol, 1.0 equiv.) and 1,4-dioxane (28 mL). Crude mixture purified by flash column chromatography (silica gel, 2.5% → 5% MeOH/dichloromethane) affording **S4** as a cream foam (1.24 g, 49%). Impure fractions were recrystallized from CHCl₃/PE giving off-white crystals (112 mg, 4%), in total 1.35 g, 53%. M.p. = 147-149 °C, **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.68 (2H, dd, *J* = 7.9 Hz, H13), 7.50-7.42 (3H, m, H14, 15), 7.23-7.14 (6H, m, H4, 17, 18, 19), 7.09 (1H, dd, *J* = 7.7, 1.5 Hz, H6), 6.93 (1H, d, *J* = 2.0 Hz, H10), 6.87 (1H, td, *J* = 7.6, 1.0 Hz, H5), 6.80 (1H, d, *J* = 8.2 Hz, H3), 6.52 (1H, d, *J* = 1.6 Hz, H8), 3.71 (3H, s, H20). **¹³C NMR** (101 MHz CDCl₃): δ_C = 163.3 (C7), 154.5 (C9), 151.3 (C2), 150.2 (C11), 137.8 (C16), 135.8 (C12), 130.2 (C19), 129.9 (C15), 129.4 (C4), 129.0 (2C, C18), 128.6 (2C, C17), 128.5 (C5), 127.6 (C1), 127.5 (2C, C14), 126.9 (2C, C13), 120.6 (C3), 116.1 (C6), 111.8 (C8), 107.0 (C10), 55.5 (C20). **IR**: ν_{max} (thin film): 1654, 1587, 1498, 1364, 1271, 1118, 1025, 861, 760, 700 cm⁻¹. **HRMS** (ES⁺): found 354.1431; C₂₄H₂₀NO₂, [M+H]⁺ requires 354.1494. The data obtained are consistent with those reported in the literature.⁸

3-Acetyl-1-(2-methoxyphenyl)-4,6-diphenylpyridin-2(1H)-one (S5)



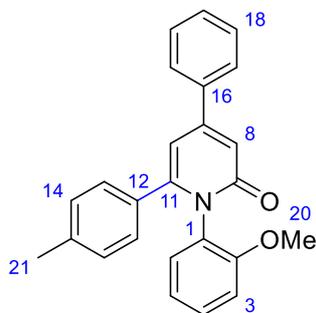
Prepared according to **General Procedure B** using alkynone **2a** (450 mg, 2.18 mmol, 1.2 equiv.), amide **1c** (377 mg, 1.82 mmol, 1.0 equiv.), KOH (102 mg, 1.82 mmol, 1.0 equiv.) and 1,4-dioxane (7 mL). Crude mixture purified by flash column chromatography (silica gel, 2.5% → 5% MeOH/dichloromethane) affording **S5** as a yellow foam (306 mg, 43%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.44-7.39 (5H, m, H17, 18, 19), 7.24-7.16 (6H, m, H13, 14, 15), 7.12 (1H, dd, *J* = 7.9, 1.8 Hz, H6), 6.89 (1H, td, *J* = 7.6, 1.2 Hz, H5), 6.79 (1H, dd, *J* = 8.5, 1.2 Hz, H3), 6.30 (1H, s, H10), 3.71 (3H, s, H20), 2.49 (3H, s, H22). **¹³C NMR** (101 MHz CDCl₃): δ_C = 202.4 (C21), 160.6 (C7), 154.4 (C2), 150.8 (C11), 150.8 (C16), 150.8 (C9), 138.0 (C8), 135.0 (C9), 130.1 (C19), 130.0 (C15), 129.4 (C12), 129.0 (C4), 128.9 (C5), 128.6 (2C, C17), 128.4 (2C, C18), 128.0 (2C, C14), 127.6 (2C, C13), 126.9 (C1), 120.6 (C3), 111.8 (C6), 109.7 (C10), 55.5 (C20), 31.8 (C22). **IR**: ν_{max} (thin film): 3060, 2243, 1699, 1640, 1572, 1528, 1271, 1025, 909, 756, 730 cm⁻¹. **HRMS** (ES⁺): found 396.1593; C₂₆H₂₂NO₃, [M+H]⁺ requires 396.1600.

3-Ethyl-1-(2-methoxyphenyl)-4,6-diphenylpyridin-2(1H)-one (S6)



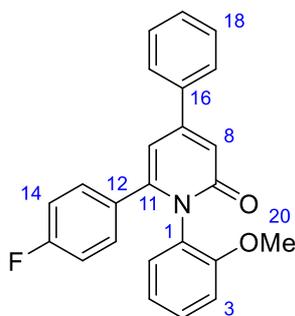
Prepared according to **General Procedure B** using alkyne **2a** (450 mg, 2.18 mmol, 1.2 equiv.), amide **1b** (352 mg, 1.82 mmol, 1.0 equiv.), KOH (102 mg, 1.82 mmol, 1.0 equiv.) and 1,4-dioxane (7 mL). Crude mixture purified by flash column chromatography (silica gel, 2.5% → 5% MeOH/dichloromethane) affording **S6** as an orange foam (359 mg, 52%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.47-7.37 (5H, m, H17, 18, 19), 7.22-7.09 (7H, m, H6, 13, 14, 15), 6.87 (1H, td, *J* = 7.6, 1.2 Hz, H5), 6.80 (1H, dd, *J* = 8.3, 1.1 Hz, H3), 6.17 (1H, s, H10), 3.71 (3H, s, H20), 2.66-2.52 (2H, m, H21), 1.18 (3H, t, *J* = 7.4 Hz, H22). **¹³C NMR** (101 MHz CDCl₃): δ_C = 163.0 (C7), 154.6 (C2), 149.2 (C11), 146.2 (C9), 139.9 (C12), 135.8 (C16), 130.8 (C8), 130.2 (C19), 129.6 (C4), 128.6 (2C, C18), 128.3 (2C, C17), 128.2 (C1), 128.2 (C15), 128.1 (2C, C14), 127.8 (C5), 127.4 (2C, C13), 120.5 (C3), 111.7 (C6), 109.7 (C10), 55.5 (C20), 21.7 (C21), 13.6 (C22). **IR**: ν_{max} (thin film): 2929, 2236, 1643, 1498, 1271, 1237, 1025, 909, 726, 700 cm⁻¹. **HRMS** (ES⁺): found 382.1809; C₂₆H₂₄NO₂, [M+H]⁺ requires 382.1807.

1-(2-Methoxyphenyl)-4-phenyl-6-(*p*-tolyl)pyridin-2(1H)-one (S7)



Prepared according to **General Procedure B** using alkyne **2b** (600 mg, 2.68 mmol, 1.2 equiv.), amide **1a** (368 mg, 2.23 mmol, 1.0 equiv.), KOH (125 mg, 2.23 mmol, 1.0 equiv.) and 1,4-dioxane (9 mL). Crude mixture purified by flash column chromatography (silica gel, 2.5% → 5% MeOH/dichloromethane) affording **S7** as an off-white foam (711 mg, 86 %). **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.68-7.65 (2H, m, H17), 7.49-7.41 (3H, m, H18, 19), 7.22 (1H, td, *J* = 8.2, 1.6 Hz, H6), 7.09-7.06 (3H, m, H4, 13), 6.96 (2H, d, *J* = 8.1 Hz, H14), 6.90-6.86 (2H, m, H5, 10), 6.81 (1H, dd, *J* = 8.2, 1.1 Hz, H3), 6.49 (1H, d, *J* = 2.1 Hz, H8), 3.72 (3H, s, H20), 2.26 (3H, s, H21). **¹³C NMR** (101 MHz CDCl₃): δ_C = 163.4 (C7), 154.7 (C9), 151.3 (C2), 150.4 (C11), 138.4 (C16), 137.9 (C15), 133.0 (C12), 130.2 (C19), 129.8 (C4), 129.4 (C5), 128.9 (2C, C13), 128.5 (2C, C14), 128.3 (2C, C17), 127.8 (C1), 126.9 (2C, C18), 120.6 (C3), 115.9 (C6), 111.9 (C8), 109.0 (C10), 55.6 (C20), 21.2 (C21). **IR**: ν_{max} (thin film): 2922, 2236, 1654, 1572, 1498, 1364, 1267, 1025, 905, 723 cm⁻¹. **HRMS** (ES⁺): found 368.1649; C₂₅H₂₃NO₂, [M+H]⁺ requires 368.1651.

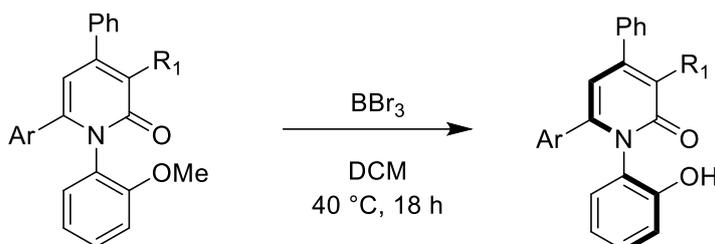
6-(4-Fluorophenyl)-1-(2-methoxyphenyl)-4-phenylpyridin-2(1H)-one (S8)



Prepared according to **General Procedure B** using alkyne **2c** (450 mg, 2.01 mmol, 1.2 equiv.), amide **1a** (276 mg, 1.67 mmol, 1.0 equiv.), KOH (94 mg, 1.67 mmol, 1.0 equiv.) and 1,4-dioxane (7 mL). Crude mixture purified by flash column chromatography (silica gel, 2.5% → 5% MeOH/dichloromethane) affording **S8** as a cream foam (530 mg, 85%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.68-7.65 (2H, m, H17), 7.50-7.42 (3H, m, H18, 19), 7.26-7.15 (3H, m, H4, 13), 7.09 (1H, dd, *J* = 7.8, 1.7 Hz, H6), 6.92-6.80 (5H, m, H3, 5, 10, 14), 6.48 (1H, d, *J* = 1.9 Hz, H8), 3.72 (3H, s, H20). **¹³C NMR** (101 MHz CDCl₃): δ_C = 163.2 (C7), 162.5 (d, ¹*J*_{CF} = 249.0 Hz, C15), 154.4 (C2), 151.3 (C9), 149.1 (C11), 137.6 (C16), 131.8 (d, ⁴*J*_{CF} = 3.6 Hz, C12), 130.5 (d, ³*J*_{CF} = 8.5 Hz, C13), 130.1 (C19), 130.1 (C4), 129.5 (C5), 129.0 (2C, C17), 127.4 (C1), 126.4 (2C, C18), 120.7 (C3), 116.4 (C6), 114.7 (d, ²*J*_{CF} = 21.7, C14), 111.9 (C8), 107.1 (C10), 55.5 (C20). **IR**: ν_{max} (thin film): 2922, 2236, 1654, 1606, 1498, 1364, 1222, 1159, 1025, 723, 697 cm⁻¹. **HRMS** (ES⁺): found 743.2714; C₂₄H₁₉FNO₂, [2M+H]⁺ requires 743.2710.

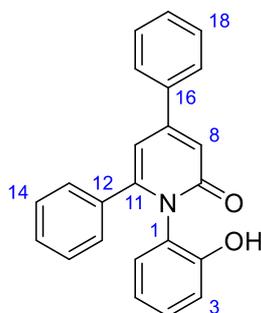
2.5 2'-Hydroxyphenyl-2-pyridones (3)

General Procedure C (Preparation of 2'-hydroxyphenyl-2-pyridones)



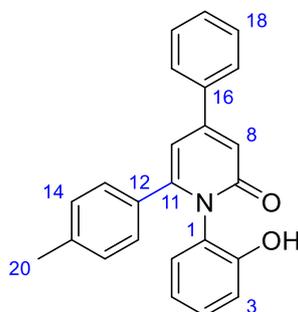
An oven-dried microwave tube was charged with pyridone derivatives (1.0 equiv.) and flushed with Ar. To this was added dichloromethane (*ca.* M) followed by BBr₃ (5.3 equiv.) and the tube was sealed and stirred at 40 °C for 18 h, then allowed to cool to RT. In some cases, as the tube cooled down the desired 2'-hydroxyphenyl-2-pyridone crashed out of solution. The solid was collected by filtration and washed with cold dichloromethane affording the desired 2'-hydroxyphenyl-2-pyridone derivative. Each mother liquor was reduced *in vacuo* and a further crop of desired enamine was retained either by recrystallization or flash column chromatography. If no crystallisation occurred purification was carried out by flash column chromatography.

1-(2-Hydroxyphenyl)-4,6-diphenylpyridin-2(1H)-one (3a)



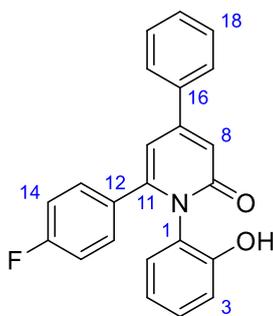
Prepared according to **General Procedure C** using pyridone **S4** (1.00 g, 2.83 mmol, 1.0 equiv.), BBr₃ (15.0 mL, 15.0 mmol, 5.3 equiv., 1M in dichloromethane), and dichloromethane (32 mL). **3a** obtained via trituration with dichloromethane as a beige solid (876 mg, 92%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 8.16 (1H, s, OH), 7.72-7.66 (2H, m, H17), 7.54-7.45 (3H, m, H18, 19), 7.26-7.16 (5H, m, H13, 14, 15), 7.05-6.99 (2H, m, H4, 10), 6.94 (1H, dd, *J* = 8.2, 1.1 Hz, H6), 6.73 (1H, d, *J* = 2.2 Hz, H8), 6.64 (1H, dd, *J* = 7.9, 1.8 Hz, H3), 6.57 (1H, td, *J* = 7.3, 1.5 Hz, H5). **¹³C NMR** (101 MHz CDCl₃): δ_C = 164.8 (C7), 153.4 (C9), 152.6 (C2), 150.6 (C11), 137.0 (C16), 135.9 (C3), 130.0 (C4), 129.9 (C19), 129.2 (3C, C1, 17), 128.8 (C15), 128.3 (2C, C14), 128.2 (2C, C18), 126.9 (2C, C13), 120.5 (C5), 119.7 (C6), 115.3 (C8), 110.1 (C10). **IR**: ν_{max} (thin film): 3056, 1654, 1531, 1587, 1498, 1271, 1025, 760, 700 cm⁻¹. **HRMS** (ES⁺): found 701.2413; C₂₃H₁₇NaNO₂, [2M+Na]⁺ requires 701.2416.

1-(2-Hydroxyphenyl)-4-phenyl-6-(*p*-tolyl)pyridin-2(1H)-one (3b)



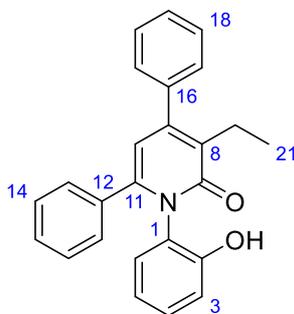
Prepared according to **General Procedure C** using pyridone **S7** (500 mg, 1.36 mmol, 1.0 equiv.), BBr₃ (7.21 mL, 7.21 mmol, 5.3 equiv., 1M in dichloromethane), and dichloromethane (16 mL). Crude mixture purified by flash column chromatography (silica gel, 1% → 5% MeOH/dichloromethane) affording **3b** as a white foam (374 mg, 78%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 8.03 (1H, s, OH), 7.69-7.66 (2H, m, H17), 7.52-7.45 (3H, m, H18, 19), 7.11-7.05 (3H, m, H4, 13), 7.0-6.97 (4H, m, H6, 10, 14), 6.71 (1H, d, *J* = 2.0 Hz, H8), 6.64 (1H, dd, *J* = 7.9, 1.8 Hz, H3), 6.60 (1H, td, *J* = 7.2, 1.6 Hz, H5), 2.26 (3H, s, H20). **¹³C NMR** (101 MHz CDCl₃): δ_C = 164.8 (C7), 153.5 (C2), 152.6 (C9), 150.8 (C11), 138.9 (C15), 137.2 (C16), 133.0 (C12), 129.9 (C3), 129.7 (C4), 129.1 (2C, C17), 129.0 (C19), 128.9 (2C, C18), 128.8 (C1), 128.2 (2C, C13), 126.9 (2C, C14), 120.3 (C5), 119.3 (C6), 115.0 (C10), 109.8 (C8), 21.3 (C20). **IR**: ν_{max} (thin film): 3058, 2713, 1640, 1602, 1505, 1457, 1362, 1278, 915, 723, 697 cm⁻¹. **HRMS** (ES⁺): found 354.1457; C₂₄H₂₀NO₂, [M+H]⁺ requires 354.1494.

6-(4-Fluorophenyl)-1-(2-hydroxyphenyl)-4-phenylpyridin-2(1H)-one (3c)



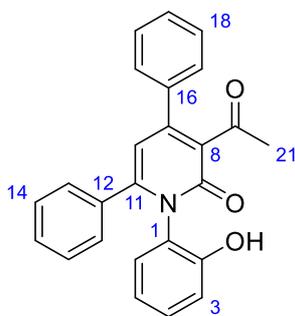
Prepared according to **General Procedure C** using pyridone **S8** (500 mg, 1.35 mmol, 1.0 equiv.), BBr₃ (7.13 mL, 7.13 mmol, 5.3 equiv., 1M in dichloromethane), and dichloromethane (16 mL). Crude mixture purified by flash column chromatography (silica gel, 1% → 5% MeOH/dichloromethane) affording **3c** as an off-white solid (473 mg, 98%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 8.28 (1H, s, OH), 7.69-7.65 (2H, m, H17), 7.53-7.46 (3H, m, H18, 19), 7.25-7.21 (2H, m, H13), 7.04-6.99 (2H, m, H4, 10), 6.90-6.85 (3H, m, H6, 14), 6.69 (1H, dd, *J* = 2.0 Hz, H8), 6.65-6.59 (2H, m, H3, 5). **¹³C NMR** (101 MHz CDCl₃): δ_C = 164.6 (C7), 162.7 (d, ¹*J*_{CF} = 249.9 Hz, C15), 153.5 (C2), 152.6 (C9), 149.7 (C11), 137.1 (C16), 131.8 (d, ⁴*J*_{CF} = 3.5 Hz, C12), 130.3 (d, ³*J*_{CF} = 8.6 Hz, C13), 129.9 (C3), 129.9 (C4), 129.1 (2C, C17), 128.8 (C19), 127.8 (C1), 126.9 (2C, C18), 120.2 (C5), 118.9 (C6), 115.6 (C8), 115.3 (d, ²*J*_{CF} = 21.6 Hz, C14), 109.6 (C10). **IR**: ν_{max} (thin film): 3060, 1643, 1602, 1558, 1502, 1367, 1230, 909, 831 cm⁻¹. **HRMS** (ES⁺): found 715.2405; C₂₃H₁₇FNO₂, [2M+H]⁺ requires 715.2408.

3-Ethyl-1-(2-hydroxyphenyl)-4,6-diphenylpyridin-2(1H)-one (3d)



Prepared according to **General Procedure C** using pyridone **S6** (330 mg, 865 μmol, 1.0 equiv.), BBr₃ (4.58 mL, 4.58 mmol, 5.3 equiv., 1M in dichloromethane), and dichloromethane (10 mL). Crude mixture purified by flash column chromatography (silica gel, 1% → 5% MeOH/dichloromethane) affording **3d** as an orange foam (301 mg, 95%). **¹H NMR** (400 MHz, CDCl₃): δ_H = 7.85 (1H, s, OH), 7.50-7.38 (5H, m, H13, 14, 15), 7.21-7.06 (7H, m, H4, 6, 17, 18, 19), 6.63 (1H, dd, *J* = 8.0, 1.6 Hz, H3), 6.59-6.55 (1H, m, H5), 6.42 (1H, s, H8), 2.75-7.54 (2H, m, H20), 1.20 (3H, t, *J* = 7.5 Hz, H21). **¹³C NMR** (101 MHz CDCl₃): δ_C = 164.7 (C7), 153.6 (C2), 150.7 (C9), 146.5 (C11), 139.1 (C12), 135.9 (C16), 130.6 (C1), 129.9 (C8), 129.6 (C3), 129.4 (C4), 128.5 (2C, C17), 128.4 (C19), 128.3 (C15), 128.2 (4C, C14, 18), 128.0 (2C, C13), 128.0 (2C, C13), 120.5 (C5), 119.8 (C6), 113.1 (C10), 21.6 (C20), 13.6 (C21). **IR**: ν_{max} (thin film): 2963, 1632, 1599, 1565, 1457, 1259, 1025, 905, 726, 697 cm⁻¹. **HRMS** (ES⁺): found 368.1645; C₂₅H₂₂NO₂, [M+H]⁺ requires 368.1651.

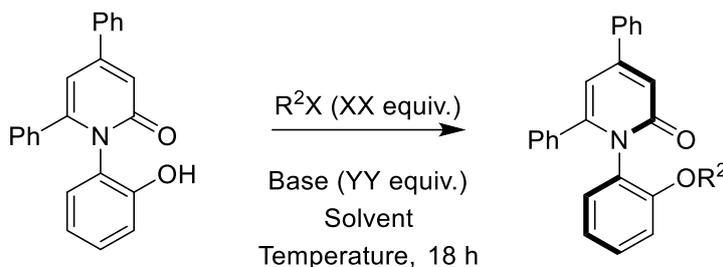
3-Acetyl-1-(2-hydroxyphenyl)-4,6-diphenylpyridin-2(1H)-one (3e)



Prepared according to **General Procedure C** using pyridone **S5** (300 mg, 759 μmol , 1.0 equiv.), BBr_3 (4.02 mL, 4.02 mmol, 5.3 equiv., 1M in dichloromethane), and dichloromethane (10 mL). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 5% MeOH/dichloromethane) affording **3e** as a yellow foam (236 mg, 82%). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.52-7.43 (5H, m, H13, 14, 15), 7.40 (1H, s, OH), 7.25-7.17 (5H, m, H17, 18, 19), (5H, m, H13, 14, 15), 7.117.07 (1H, m, H4), 7.00 (1H, dd, J = 8.2, 1.3 Hz, H6), 6.65 (1H, dd, J = 8.0, 1.8 Hz, H8), 6.61 (1H, td, J = 7.1, 1.4 Hz, H3), 6.53 (1H, s, H10), 2.44 (3H, s, H21). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 201.9 (C20), 161.8 (C7), 153.4 (C2), 151.1 (C9), 150.8 (C11), 137.1 (C16), 135.1 (C12), 130.1 (C3), 129.5 (C19), 129.2 (C15), 129.0 (C4), 129.0 (C8), 128.9 (2C, 17), 128.3 (2C, C18), 128.3 (C1), 128.2 (2C, C14), 128.0 (2C, C13), 120.7 (C5), 119.5 (C6), 112.2 (C10), 31.8 (C21). **IR**: ν_{max} (thin film): 3183, 1699, 1628, 1565, 1528, 1367, 1103, 909, 756 cm^{-1} . **HRMS** (ES+): found 763.2812; $\text{C}_{25}\text{H}_{20}\text{NO}_3$, $[\text{2M}+\text{H}]^+$ requires 763.2808.

2.6 O-Benzylated N-(2-phenoxy)pyridones (4)

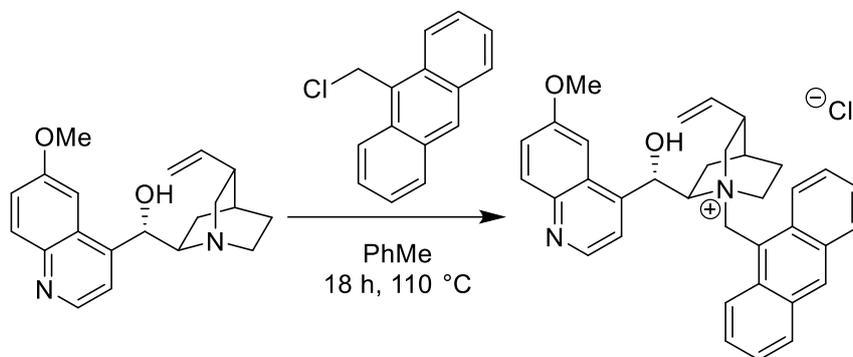
2.6.1 Optimisation method using chiral catalyst



Sample vials were charged with *N*-(2-phenoxy)pyridone (14 mg, 41.0 μmol , 1.0 equiv.) and catalyst (10-15 mol%). Solvent (0.5-1.5 mL) was added followed by 50% aqueous base (5.0 equiv.) and R^2X ($\text{X} = \text{Br}, \text{I}$ or OTs, 0.6-5.0 equiv.) as the vial was sealed and stirred at the desired temperature for the time stated. At various intervals, the resulting solution was subjected to preparative TLC and enantiomeric excess determined by HPLC (Chiralpak, 60% MeCN/ H_2O).

2.6.2 Catalyst synthesis

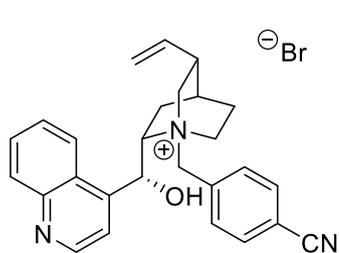
Various catalysts were synthesised and used during optimisation. *O*-Alkylation was performed as described by Corey *et al.*⁹ and hydrogenation was performed as described by Tang *et al.*¹⁰



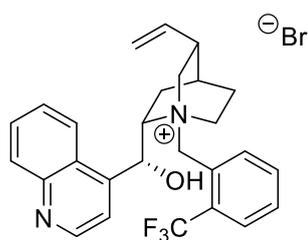
A flask was charged with quinidine (200 mg, 616 μmol , 1.0 equiv.), 9-chloromethyl anthracene (139 mg, 616 μmol , 1.0 equiv.) and toluene (10 mL). The reaction mixture was stirred at 110 $^{\circ}\text{C}$ for 18 h as a cream precipitate formed from a yellow solution. The crude product was removed from the heat, allowed to cool and then Et_2O (3 x 25 mL) was added and resulting mixture stirred for 10 mins. Product purified by filtration, washing with cold Et_2O (3 x 25 mL), affording QD-1 as a yellow solid (178 mg, 52%). The data obtained are consistent with those reported in the literature.¹¹

$^1\text{H NMR}$ (400 MHz, CDCl_3): δ_{H} = 9.07 (1H, d, J = 9.0 Hz), 8.50 (1H, d, J = 8.9 Hz), 8.30-8.24 (2H, m), 8.12-8.09 (1H, m), 8.00 (1H, s, OH), 7.95-7.91 (1H, m), 7.65 (1H, d, J = 8.8 Hz), 7.57 (1H, d, J = 8.3 Hz), 7.38-7.26 (3H, m), 7.20-7.12 (2H, m), 7.07-7.05 (1H, m), 6.95 (1H, d, J = 13.2 Hz), 6.56 (1H, d, J = 13.3 Hz), 5.74-5.65 (1H, m), 5.09 (1H, d, J = 10.5 Hz), 4.98-4.94 (1H, m), 4.75-4.70 (1H, m), 4.57-4.51 (1H, m), 4.14 (1H, t, J = 11.4 Hz), 3.84 (3H, s, OMe), 2.81-2.70 (1H, m), 2.35-2.21 (2H, m), 1.87-1.72 (3H, m), 1.63 (1H, s), 1.44-1.38 (1H, m), 1.00-0.88 (1H, m). **$^{13}\text{C NMR}$** (101 MHz, CDCl_3): δ_{C} = 157.7, 147.2, 144.5, 143.2, 135.8, 133.1, 132.9, 131.5, 131.0, 130.5, 130.3, 128.7, 128.5, 127.8, 127.6, 127.3, 126.1, 125.0, 124.9, 124.8, 121.4, 120.5, 118.3, 117.4, 104.1, 68.3, 65.5, 56.6, 56.2, 54.5, 54.3, 38.1, 26.3, 24.2, 22.6. **IR**: ν_{max} (thin film): 3652, 3052, 2199, 1621, 1505, 1449, 1241, 1028, 905, 723 cm^{-1} . **HRMS** (ES+): found 515.2687; $\text{C}_{35}\text{H}_{35}\text{N}_2\text{O}_2$, $[\text{M}-\text{Cl}]^+$ requires 515.2699. $[\alpha]_{\text{D}}^{293}$ +380.2 (c = 0.3, CHCl_3).

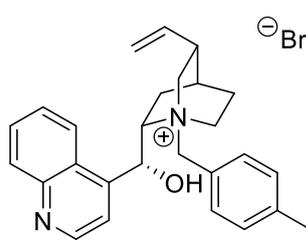
2.6.3 Chiral catalyst library



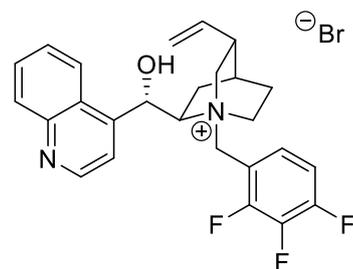
CD-1



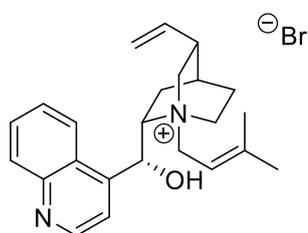
CD-2



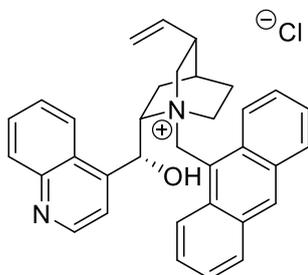
CD-3



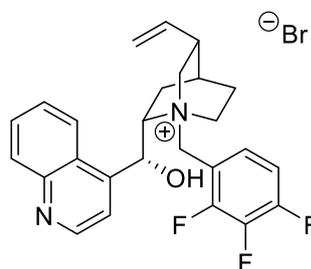
CN-1



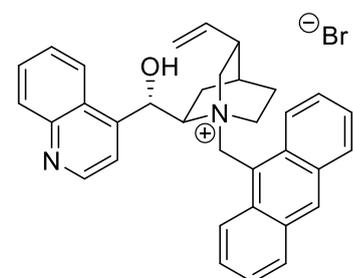
CD-4



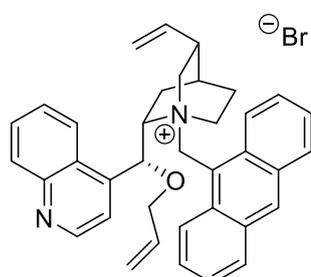
**CD-5
(C1)**



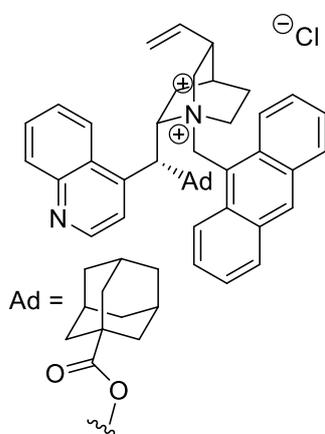
CD-6



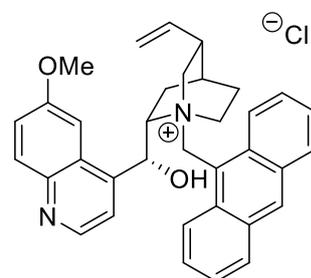
CN-2



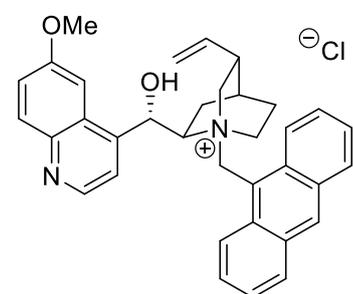
**CD-7
(C2)**



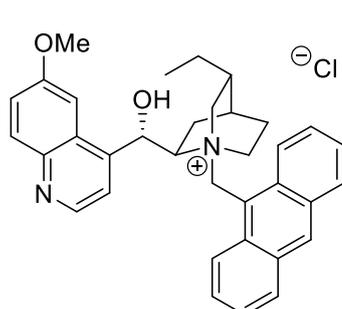
CD-8



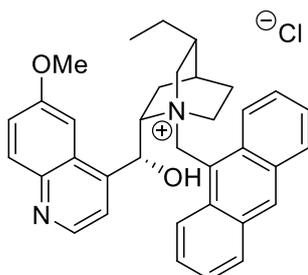
**QN-1
(C3)**



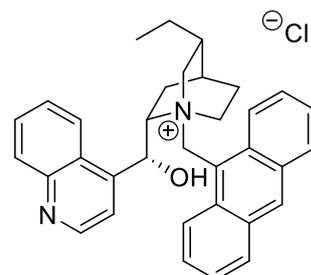
**QD-1
(C4)**



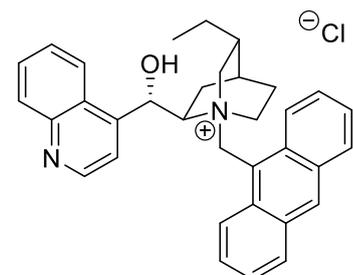
QD-2



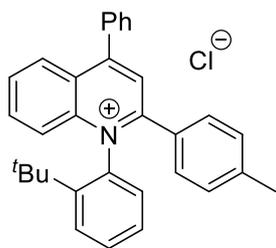
QN-2



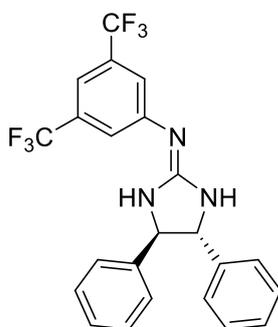
CD-9



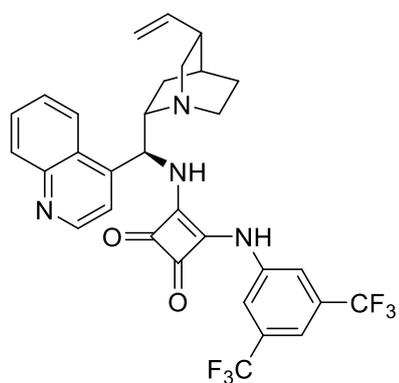
CN-3



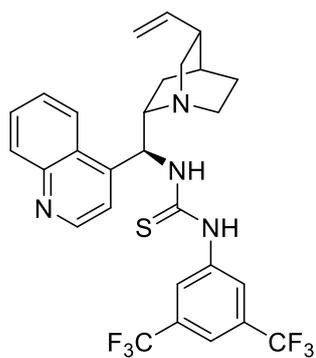
Quinolinium-1



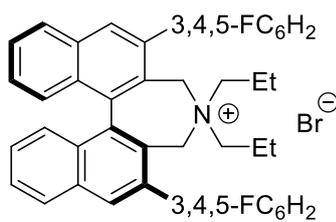
Cat-1



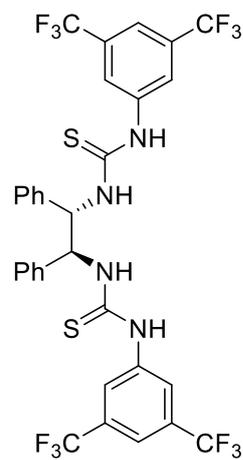
Cat-2



Cat-3



Maruoka-1



Cat-4

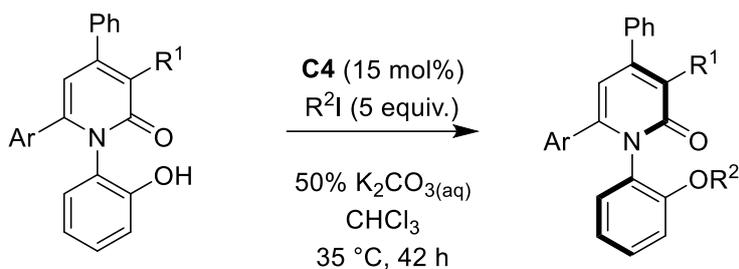
2.6.4 Optimisation results

Table S1. Optimisation of dynamic kinetic resolution of pyridones. For reaction conditions see section 2.6.1 above. In each case, base was always 50% aqueous solution. Enantiomeric excesses were determined after 18 h by HPLC on a chiral stationary phase.

Entry	Solvent	Base	Bn-X (eq.)	TLC Conversion (%)	Catalyst	Temp/ °C	ee
1	PhH	K ₃ PO ₄	BnBr (1.2 eq.)	<25	CD-1	rt	16%
2	PhH	K ₃ PO ₄	BnBr (1.2 eq.)	<25	CD-2	rt	11%
3	PhH	K ₃ PO ₄	BnBr (1.2 eq.)	<25	CD-3	rt	16%
4	PhH	K ₃ PO ₄	BnBr (1.2 eq.)	<25	CD-4	rt	1%
5	PhH	K ₃ PO ₄	BnBr (1.2 eq.)	<25	CD-5	rt	-12%
6	PhH	K ₃ PO ₄	BnBr (1.2 eq.)	<25	CD-6	rt	9%
7	PhH	K ₃ PO ₄	BnBr (0.6 eq.)	<25	CN-1	rt	-3%
8	PhH	K ₃ PO ₄	BnBr (0.6 eq.)	<25	Maruoka-1	rt	24%
9	PhH	K ₃ PO ₄	BnBr (5.0 eq.)	<25	Maruoka-1	rt	23%
10	CHCl ₃	K ₃ PO ₄	BnBr (0.6 eq.)	<25	Cat-1	rt	0%
11	CHCl ₃	K ₃ PO ₄	BnBr (0.6 eq.)	<25	Cat-2	rt	6%
12	CHCl ₃	K ₃ PO ₄	BnBr (0.6 eq.)	<25	Cat-3	rt	18%
13	CHCl ₃	K ₃ PO ₄	BnBr (0.6 eq.)	<25	Cat-4	rt	28%
14	CHCl ₃	K ₃ PO ₄	BnBr (0.6 eq.)	<25	Quinolinium-1	rt	-7%
15	CHCl ₃	K ₃ PO ₄	BnBr (0.6 eq.)	<25	Maruoka-1	rt	29%
17	CHCl ₃	K ₃ PO ₄	BnBr (0.6 eq.)	25-50	CD-8	rt	-2%
18	CHCl ₃	K ₃ PO ₄	BnBr (0.6 eq.)	<25	CD-5	rt	-58%
19	CHCl ₃	KOH	BnBr (0.6 eq.)	<25	CD-5	rt	-8%
20	CHCl ₃	Cs ₂ CO ₃	BnBr (0.6 eq.)	<25	CD-5	rt	-66%
21	CHCl ₃	K ₂ CO ₃	BnBr (0.6 eq.)	<25	CD-5	rt	-66%
22	DCE	K ₂ CO ₃	BnBr (0.6 eq.)	<25	CD-5	rt	-40%
23	Et ₂ O	K ₂ CO ₃	BnBr (0.6 eq.)	<25	CD-5	rt	-2%
24	<i>m</i> -xylene	K ₂ CO ₃	BnBr (0.6 eq.)	<25	CD-5	rt	-30%
25	THF	K ₂ CO ₃	BnBr (0.6 eq.)	<25	CD-5	rt	-8%
26	PhMe	K ₂ CO ₃	BnBr (0.6 eq.)	<25	CD-5	rt	-22%
27	CHCl ₃	K ₂ CO ₃	BnBr (0.6 eq.)	<25	CN-2	rt	68%
28	CHCl ₃	K ₂ CO ₃	BnOTs (0.6 eq.)	25-50	CD-5	rt	-51%
29	CHCl ₃	K ₂ CO ₃	BnI (0.6 eq.)	<25	CD-5	rt	-73%
30	CHCl ₃	K ₂ CO ₃	BnI (0.6 eq.)	<25	CD-7	rt	-33%
31	CHCl ₃	K ₂ CO ₃	BnI (0.6 eq.)	50-75	CD-5	50	-45%
32	CHCl ₃	K ₂ CO ₃	BnI (0.6 eq.)	<25	CD-5	0	-54%
33	CHCl ₃	K ₂ CO ₃	BnI (5.0 eq.)	<25	QN-1	rt	-76%
34	CHCl ₃	K ₂ CO ₃	BnI (5.0 eq.)	<25	QD-1	rt	85%
35	CHCl ₃	K ₂ CO ₃	BnI (5.0 eq.)	<25	QD-2	rt	57%
36	CHCl ₃	K ₂ CO ₃	BnI (5.0 eq.)	50-75	QD-1	35	80%

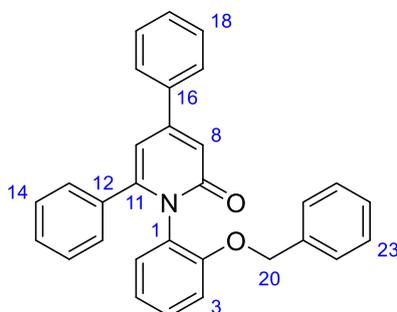
2.6.5 Enantioselective Synthesis of 4a-4o

General Procedure D (Preparation of 2-arylpiperidones)



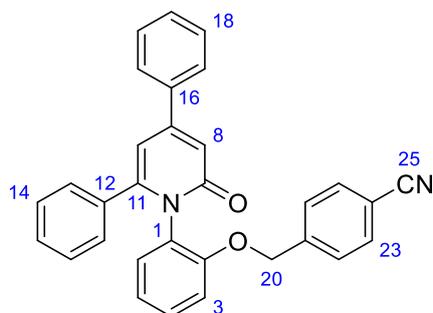
A round-bottomed flask was charged with 2'-hydroxyphenyl-2-pyridone (1.0 equiv.) and QD-1 (10 mol%). CHCl₃ (ca. 0.3 M) was added followed by 50% aqueous K₂CO₃ (5.0 equiv.) and R²I (5.0 equiv.) as the flask was sealed and stirred at 35 °C for 42 h. The reaction mixture was subjected to preparative TLC and enantiomeric excess analysis determined by HPLC (CHIRALPAK, 60% MeCN/H₂O) after 18 h, at which time an additional portion of QD-1 (5 mol%) was added. Piperidine (4.0 equiv.) was added and reaction mixture was stirred for 30 mins before addition of 3M HCl (ca. 1 mL). Crude product was extracted with EtOAc (3 x10 mL/mmol), dried over anhydrous magnesium sulfate and the solvent removed *in vacuo* resulting in a crude solid. Purification by column chromatography (silica gel, 1% → 2.5% MeOH/dichloromethane) afforded the desired product.

1-(2-(Benzyloxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4a)



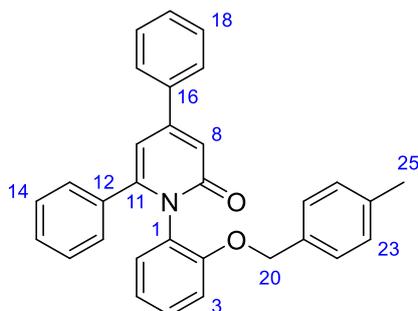
General Procedure D: Pyridone **3a** (100 mg, 295 μ mol, 1.0 equiv.), QD-1 (24.4 mg, 44.3 μ mol, 15 mol%). CHCl₃ (11 mL) was added followed by 50% aqueous K₂CO₃ (205 μ L, 1.48 mmol, 5.0 equiv.) and R²I-**S1** (184 μ L, 1.48 mmol, 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% → 2.5% MeOH/dichloromethane) affording **4a** as a yellow foam (98 mg, 77%, 75% ee). **¹H NMR** (400 MHz, CDCl₃): δ_{H} = 7.69-7.66 (2H, m, H17), 7.50-7.44 (3H, m, H18, 19), 7.33-7.24 (5H, m, H13, 14, 15), 7.21-7.11 (7H, m, H4, 6, 22, 23, 24), 6.94 (1H, d, J = 2.0 Hz, H10), 6.9. (1H, td, J = 7.6, 1.2 Hz, H5), 6.84 (1H, dd, J = 8.4, 1.1 Hz, H3), 6.49 (1H, d, J = 2.1 Hz, H8), 5.09-4.93 (2H, m, H20). **¹³C NMR** (101 MHz CDCl₃): δ_{C} = 163.4 (C7), 153.8 (C9), 151.4 (C2), 150.3 (C11), 137.8 (C16), 136.8 (C21), 135.7 (C12), 130.4 (C19), 129.9 (C4), 129.4 (C15), 129.0 (2C, C17), 128.8 (2C, C23), 128.5 (C24), 128.5 (2C, C18), 128.1 (C1), 127.7 (C6), 127.6 (2C, C14), 126.8 (2C, C13), 126.7 (2C, C22), 120.9 (C5), 116.1 (C10), 113.4 (C3), 107.0 (C8), 70.2 (C20). **HPLC:** Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_{R} (major) = 36.4 min, t_{R} (minor) = 15.5 min. **IR:** ν_{max} (thin film): 3060, 2236, 1654, 1572, 1531, 1490, 1364, 1267, 1230, 909, 730, 697 cm⁻¹. **HRMS** (ES⁺): found 430.1810; C₃₀H₂₄NO₂, [M+H]⁺ requires 430.1807. $[\alpha]_{\text{D}}^{293}$ +33.3 (c = 0.6, CHCl₃).

4-((2-(2-Oxo-4,6-diphenylpyridin-1(2H)-yl)phenoxy)methyl)benzonitrile (4b)



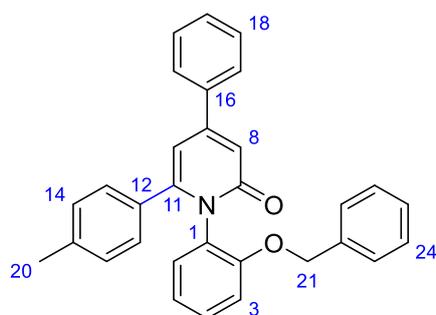
General Procedure D: Pyridone **3a** (50.0 mg, 141 μmol , 1.0 equiv.), QD-1 (11.7 mg, 21.2 μmol , 15 mol%). CHCl_3 (6 mL) was added followed by 50% aqueous K_2CO_3 (98.0 μL , 707 μmol , 5.0 equiv.) and $\text{R}^2\text{I-S2}$ (89 mg, 707 μmol , 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4b** as a yellow solid (40 mg, 80%, 67% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.69-7.66 (2H, m, H17), 7.60 (2H, d, J = 8.4 Hz, H23), 7.52-7.46 (3H, m, H18, 19), 7.42 (2H, d, J = 8.6 Hz, H22), 7.24-7.13 (7H, m, H4, 6, 13, 14, 15), 6.94-6.91 (2H, m, H5, 10), 6.78 (1H, dd, J = 8.3, 1.0 Hz, H3), 6.53 (1H, d, J = 2.0 Hz, H8), 5.16-5.05 (2H, m, H20). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 163.3 (C7), 153.3 (C9), 151.6 (C2), 150.2 (C11), 142.2 (C21), 137.6 (C16), 135.6 (C12), 132.4 (2C, C23), 130.6 (C4), 130.0 (C19), 129.7 (C15), 129.1 (2C, C17), 128.7 (C5), 128.6 (2C, C14), 128.3 (C1), 127.8 (2C, C17), 127.1 (2C, C13), 126.8 (2C, C22), 121.6 (C6), 118.7 (C25), 115.9 (C10), 113.3 (C3), 111.6 (C24), 107.2 (C8), 69.3 (C20). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 33.4 min, t_{R} (minor) = 13.7 min. **IR:** ν_{max} (thin film): 3056, 2228, 1651, 1490, 1449, 1271, 909, 820, 726, 697 cm^{-1} . **HRMS** (ES⁺): found 455.1785; $\text{C}_{31}\text{H}_{23}\text{N}_2\text{O}_2$, $[\text{M}+\text{H}]^+$ requires 455.1760. $[\alpha]_{\text{D}}^{293}$ +43.6 (c = 0.5, CHCl_3).

1-(2-((4-Methylbenzyl)oxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4c)



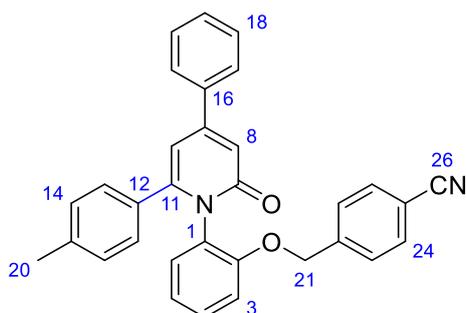
General Procedure D: 2-hydroxypyridone **3a** (60 mg, 177 μmol , 1.0 equiv.), QD-1 (14.6 mg, 26.6 μmol , 15 mol%). CHCl_3 (6 mL) was added followed by 50% aqueous K_2CO_3 (122 μL , 884 μmol , 5.0 equiv.) and $\text{R}^2\text{I-S3}$ (205 mg, 884 μmol , 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4c** as an off-white solid (60 mg, 76%, 70% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.69-7.65 (2H, m, H17), 7.50-7.45 (3H, m, H18, 19), 7.22-7.11 (11H, m, H4, 6, 13, 14, 15, 22, 23), 6.94 (1H, d, J = 2.0 Hz, H10), 6.89 (1H, td, J = 7.5, 1.1 Hz, H5), 6.84 (1H, dd, J = 8.4, 1.0 Hz, H3), 6.48 (1H, d, J = 2.0 Hz, H8), 5.05-4.88 (2H, m, H20), 2.33 (3H, s, H25). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 163.4 (C7), 153.9 (C9), 151.3 (C2), 150.3 (C11), 137.8 (C16), 137.4 (C24), 135.7 (C12), 133.8 (C21), 130.4 (C4), 129.8 (C19), 129.4 (C15), 129.1 (2C, C23), 129.0 (2C, C17), 128.8 (2C, C18), 128.5 (C5), 128.1 (C1), 127.6 (2C, C14), 126.9 (2C, C13), 126.9 (2C, C22), 120.8 (C6), 116.1 (C8), 113.5 (C3), 107.0 (C10), 70.2 (C20), 21.2 (C25). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 53.5 min, t_{R} (minor) = 19.9 min. **IR:** ν_{max} (thin film): 3056, 2922, 2236, 1654, 1572, 1531, 1490, 1364, 1267, 998, 801, 697, 723 cm^{-1} . **HRMS** (ES⁺): found 444.1976; $\text{C}_{31}\text{H}_{26}\text{NO}_2$, $[\text{M}+\text{H}]^+$ requires 444.1964. $[\alpha]_{\text{D}}^{293}$ +27.4 (c = 0.4, CHCl_3).

1-(2-(Benzyloxy)phenyl)-4-phenyl-6-(*p*-tolyl)pyridin-2(1*H*)-one (4d)



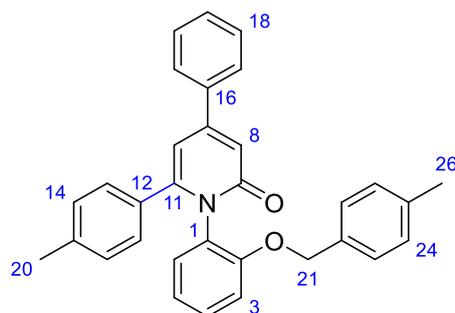
General Procedure D: Pyridone **3d** (100 mg, 262 μmol , 1.0 equiv.), QD-1 (21.6 mg, 39.3 μmol , 15 mol%). CHCl_3 (11 mL) was added followed by 50% aqueous K_2CO_3 (182 μL , 1.31 mmol, 5.0 equiv.) and $\text{R}^2\text{I-S1}$ (163 μL , 1.31 mmol, 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4d** as a yellow foam (82 mg, 70%, 77% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): $\delta_{\text{H}} = 7.68\text{--}7.65$ (2H, m, H17), 7.50–7.44 (3H, m, H18, 19), 7.33–7.24 (5H, m, H23, 24, 25), 7.22–7.14 (2H, m, H4, 6), 7.06–7.04 (2H, m, H13), 6.95–6.89 (4H, m, H5, 10, 14), 6.85 (1H, dd, $J = 8.4, 1.2$ Hz, H3), 6.48 (1H, d, $J = 2.0$ Hz, H8), 5.10–4.94 (2H, m, H21), 2.26 (3H, s, H20). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): $\delta_{\text{C}} = 163.5$ (C7), 153.8 (C9), 151.4 (C2), 150.4 (C11), 138.4 (C16), 137.9 (C12), 136.9 (C21), 132.9 (C15), 130.4 (C19), 129.8 (C4), 129.4 (C25), 129.0 (2C, C13), 128.7 (2C, C17), 128.4 (2C, C24), 128.3 (2C, C14), 128.3 (C1), 127.7, 126.9 (2C, C18), 126.7 (2C, C23), 120.9 (C5), 115.9 (C10), 113.5 (C3), 107.1 (C8), 70.2 (C20), 21.2 (C20). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, $\lambda = 250$ nm, t_{R} (major) = 40.2 min, t_{R} (minor) = 18.2 min. **IR:** ν_{max} (thin film): 3030, 2236, 1654, 1572, 1498, 1364, 1267, 905, 723 cm^{-1} . **HRMS** (ES $^+$): found 444.1956; $\text{C}_{31}\text{H}_{26}\text{NO}_2$, $[\text{M}+\text{H}]^+$ requires 444.1964. $[\alpha]_{\text{D}}^{293} +43.8$ ($c = 0.6$, CHCl_3).

4-((2-(2-Oxo-4-phenyl-6-(*p*-tolyl)pyridin-1(2*H*)-yl)phenoxy)methyl)benzonitrile (4e)



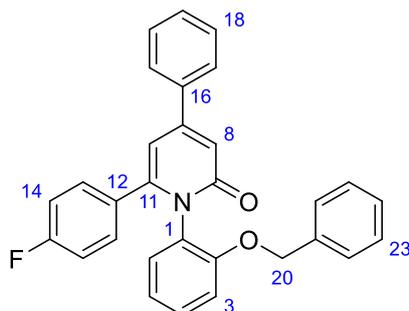
General Procedure D: Pyridone **3d** (100 mg, 283 μmol , 1.0 equiv.), QD-1 (23.4 mg, 42.5 μmol , 15 mol%). CHCl_3 (11 mL) was added followed by 50% aqueous K_2CO_3 (196 μL , 1.42 mmol, 5.0 equiv.) and $\text{R}^2\text{I-S2}$ (344 mg, 1.42 mmol, 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4e** as a white foam (114 mg, 86%, 63% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): $\delta_{\text{H}} = 7.68\text{--}7.65$ (2H, m, H17), 7.60 (2H, d, $J = 8.1$ Hz, H24), 7.52–7.46 (3H, m, H18, 19), 7.42 (2H, d, $J = 8.2$ Hz, H23), 7.21 (1H, td, $J = 8.3, 1.7$ Hz, H4), 7.14 (1H, dd, $J = 7.8, 1.7$ Hz, H6), 7.04 (2H, d, $J = 8.2$ Hz, H13), 6.98–6.92 (4H, m, H5, 10, 14), 6.79 (1H, dd, $J = 8.3, 1.0$ Hz, H3), 6.51 (1H, d, $J = 2.0$ Hz, H8), 5.15–5.02 (2H, m, H21), 2.26 (3H, s, H20). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): $\delta_{\text{C}} = 163.4$ (C7), 153.3 (C9), 151.6 (C2), 150.3 (C11), 142.2 (C22), 138.6 (C16), 137.7 (C15), 132.7 (C12), 132.3 (2C, C24), 130.6 (C3), 129.8 (C4), 129.6 (C19), 129.1 (2C, C13), 128.5 (2C, C17), 128.4 (3C, C1, 14), 127.1 (2C, C18), 126.8 (2C, C23), 121.6 (C5), 118.7 (C26), 115.7 (C8), 113.4 (C3), 111.6 (C25), 107.1 (C10), 69.4 (C21), 21.2 (C20). **$^{19}\text{F NMR}$** (376 MHz, CDCl_3): $\delta_{\text{F}} = -111.7$. **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, $\lambda = 250$ nm, t_{R} (major) = 39.6 min, t_{R} (minor) = 16.6 min. **IR:** ν_{max} (thin film): 3056, 2922, 2228, 1654, 1572, 1498, 1453, 1364, 1267, 998, 909, 816 723 cm^{-1} . **HRMS** (ES $^+$): found 469.1940; $\text{C}_{32}\text{H}_{25}\text{N}_2\text{O}_2$, $[\text{M}+\text{H}]^+$ requires 469.1916. $[\alpha]_{\text{D}}^{293} +53.1$ ($c = 0.4$, CHCl_3).

1-(2-((4-Methylbenzyl)oxy)phenyl)-4-phenyl-6-(*p*-tolyl)pyridin-2(1*H*)-one (4f)



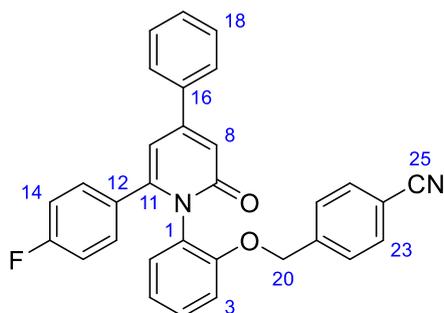
General Procedure D: Pyridone **3d** (100 mg, 283 μmol , 1.0 equiv.), QD-1 (23.4 mg, 42.5 μmol , 15 mol%). CHCl_3 (11 mL) was added followed by 50% aqueous K_2CO_3 (196 μL , 1.42 mmol, 5.0 equiv.) and $\text{R}^{21}\text{-S3}$ (328 mg, 1.42 mmol, 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4f** as a yellow foam (86 mg, 66%, 58% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.68-7.65 (2H, m, H17), 7.50-7.44 (3H, m, H18, 19), 7.21-7.10 (6H, m, H4, 6, 13, 14), 7.07-7.04 (2H, m, H23), 6.95-6.85 (5H, m, H3, 5, 11, 24), 6.47 (1H, d, J = 2.0 Hz, H8), 5.05-4.89 (2H, m, H21), 2.33 (3H, s, H20), 2.26 (3H, s, H26). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 162.4 (C7), 152.9 (C9), 150.3 (C2), 149.4 (C11), 137.4 (C16), 136.9 (C15), 136.3 (C25), 132.8 (C22), 131.8 (C12), 129.3 (C4), 128.7 (C19), 128.3 (C5), 128.1 (2C, C24), 127.9 (2C, C13), 127.6 (2C, C17), 127.3 (2C, C14), 127.2 (C1), 125.8 (2C, C18), 125.8 (2C, C23), 119.8 (C6), 114.8 (C8), 112.5 (C3), 105.6 (C10), 69.2 (C21), 20.2 (C20), 20.1 (C26). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 62.8 min, t_{R} (minor) = 23.8 min. **IR:** ν_{max} (thin film): 3026, 2922, 2236, 1654, 1498, 1453, 1364, 1263, 998, 801, 723, 697 cm^{-1} . **HRMS** (ES $^+$): found 458.2122; $\text{C}_{32}\text{H}_{28}\text{NO}_2$, $[\text{M}+\text{H}]^+$ requires 458.2120. $[\alpha]_{\text{D}}^{293}$ +39.2 (c = 0.5, CHCl_3).

1-(2-(Benzyloxy)phenyl)-6-(4-fluorophenyl)-4-phenylpyridin-2(1*H*)-one (4g)



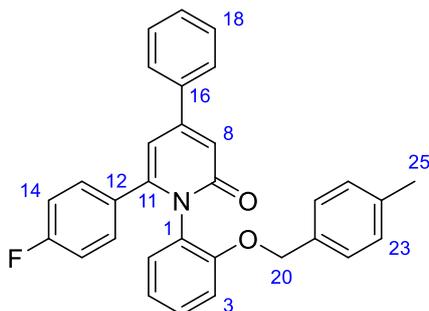
General Procedure D: Pyridone **3e** (50.0 mg, 141 μmol , 1.0 equiv.), QD-1 (11.7 mg, 21.2 μmol , 15 mol%). CHCl_3 (6 mL) was added followed by 50% aqueous K_2CO_3 (98.0 μL , 707 μmol , 5.0 equiv.) and $\text{R}^{21}\text{-S1}$ (89 μL , 707 μmol , 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4g** as a yellow solid (40 mg, 64%, 65% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.68-7.65 (2H, m, H17), 7.51-7.45 (3H, m, H18, 19), 7.35-7.27 (5H, m, H23, 24, 25), 7.24-7.19 (1H, m, H4), 7.16-7.11 (3H, m, H6, 13), 6.95-6.90 (2H, m, H5, 10), 6.88-6.80 (3H, m, H3, H14), 6.46 (1H, d, J = 1.9 Hz, H8), 5.09-4.92 (2H, m, H21). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 163.5 (C7), 162.5 (d, $^1J_{\text{CF}}$ = 249.0 Hz, C15), 153.6 (C9), 151.4 (C2), 149.2 (C11), 137.7 (C16), 136.7 (C21), 131.8 (d, $^4J_{\text{CF}}$ = 3.6 Hz, C12), 130.7 (d, $^3J_{\text{CF}}$ = 8.4 Hz, C13), 130.4 (C19), 130.0 (C4), 129.5 (C24), 129.0 (2C, C17), 128.5 (2C, C23), 128.0 (C1), 127.8 (C6), 126.8 (2C, C18), 126.8 (2C, C23), 121.1 (C5), 116.3 (C10), 114.7 (d, $^2J_{\text{CF}}$ = 21.7 Hz, C14), 113.6 (C3), 107.1 (C8), 70.2 (C20). **$^{19}\text{F NMR}$** (376 MHz, CDCl_3): δ_{F} = -112.1. **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 41.8 min, t_{R} (minor) = 16.8 min. **IR:** ν_{max} (thin film): 3060, 2922, 2236, 1654, 1605, 1498, 1271, 1222, 1159, 998, 730, 697 cm^{-1} . **HRMS** (ES $^+$): found 448.1711; $\text{C}_{30}\text{H}_{23}\text{FNO}_2$, $[\text{M}+\text{H}]^+$ requires 448.1713. $[\alpha]_{\text{D}}^{293}$ +29.6 (c = 0.5, CHCl_3).

4-((2-(6-(4-Fluorophenyl)-2-oxo-4-phenylpyridin-1(2H)-yl)phenoxy)methyl)benzonitrile (4h)



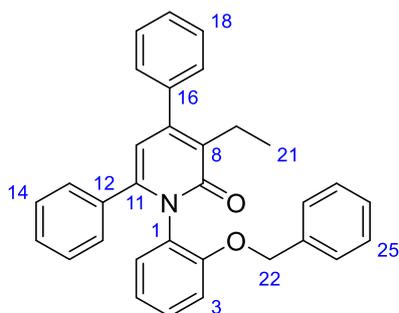
General Procedure D: Pyridone **3e** (100 mg, 280 μmol , 1.0 equiv.), QD-1 (23.3 mg, 42.0 μmol , 15 mol%). CHCl_3 (11 mL) was added followed by 50% aqueous K_2CO_3 (193 μL , 1.40 mmol, 5.0 equiv.) and $\text{R}^2\text{I-S2}$ (340 mg, 1.40 mmol, 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4h** as a yellow solid (114 mg, 86%, 57% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.68-7.65 (2H, m, H17), 7.62-7.59 (2H, m, H23), 7.52-7.46 (3H, m, H18, 19), 7.43 (2H, d, J = 8.6 Hz, H22), 7.22 (1H, td, J = 8.0, 1.8 Hz, H4), 7.16-7.11 (3H, m, H6, 13), 6.97-6.93 (2H, m, H5, 10), 6.87-6.80 (3H, m, H3,14), 6.50 (1H, d, J = 2.0 Hz, H8), 5.14-5.02 (2H, m, H20). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 163.9 (C7), 162.6 (d, $^1J_{\text{CF}}$ = 249.0 Hz, C15), 153.2 (C9), 151.6 (C2), 149.1 (C11), 142.0 (C21), 137.4 (C16), 132.3 (2C, C23), 131.6 (d, $^4J_{\text{CF}}$ = 3.5 Hz, C12), 130.6 (C6), 130.6 (d, $^3J_{\text{CF}}$ = 8.1 Hz, C13), 130.1 (C4), 129.7 (C19), 129.1 (2C, C17), 128.2 (C1), 127.1 (2C, C18), 126.8 (2C, C22), 121.8 (C5), 118.7 (C25), 116.2 (C8), 114.9 (d, $^2J_{\text{CF}}$ = 21.7 Hz, C14), 113.5 (C3), 111.7 (C25), 107.2 (C10), 69.4 (C20). **$^{19}\text{F NMR}$** (376 MHz, CDCl_3): δ_{F} = -111.7. **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 38.7 min, t_{R} (minor) = 14.5 min. **IR:** ν_{max} (thin film): 3060, 2922, 2228, 1654, 1606, 1498, 1364, 1222, 909, 820, 726, 697 cm^{-1} . **HRMS** (ES+): found 473.1670; $\text{C}_{31}\text{H}_{22}\text{FN}_2\text{O}_2$, $[\text{M}+\text{H}]^+$ requires 473.1665. $[\alpha]_{\text{D}}^{293}$ +42.9 (c = 0.6, CHCl_3).

6-(4-Fluorophenyl)-1-(2-((4-methylbenzyl)oxy)phenyl)-4-phenylpyridin-2(1H)-one (4i)



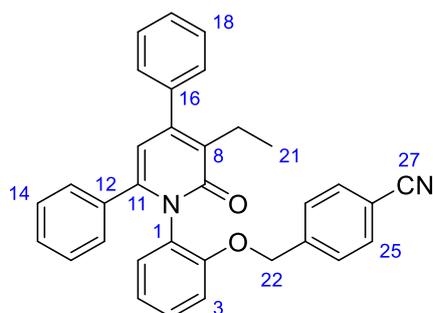
General Procedure D: Pyridone **3e** (100 mg, 280 μmol , 1.0 equiv.), QD-1 (23.3 mg, 42.0 μmol , 15 mol%). CHCl_3 (11 mL) was added followed by 50% aqueous K_2CO_3 (193 μL , 1.40 mmol, 5.0 equiv.) and $\text{R}^2\text{I-S3}$ (324 mg, 1.40 mmol, 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4i** as an off-white foam (81 mg, 63%, 51% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.68-7.65 (2H, m, H17), 7.51-7.44 (3H, m, H18, 19), 7.23-7.19 (3H, m, H4, 22), 7.15-7.11 (5H, m, H6, 13, 23), 6.94-6.80 (5H, m, H3, 5, 10, 14), 6.45 (1H, d, J = 2.0 Hz, H8), 5.05-4.88 (2H, m, H20), 2.33 (3H, s, H25). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 163.8 (C7), 162.5 (d, $^1J_{\text{CF}}$ = 248.7 Hz, C15), 153.8 (C9), 151.3 (C2), 149.2 (C11), 137.7 (C16), 137.6 (C24), 133.7 (C22), 131.8 (d, $^4J_{\text{CF}}$ = 3.6 Hz, C12), 130.7 (d, $^3J_{\text{CF}}$ = 8.5 Hz, C13), 130.3 (C4), 130.0 (C19), 129.5 (C5), 129.2 (2C, C23), 129.0 (2C, C17), 128.0 (C1), 126.9 (2C, C18), 126.8 (2C, C22), 121.0 (C6), 116.3 (C8), 114.7 (d, $^2J_{\text{CF}}$ = 21.6 Hz, C14), 112.5 (C3), 107.1 (C10), 70.3 (C20), 21.2 (C25). **$^{19}\text{F NMR}$** (376 MHz, CDCl_3): δ_{F} = -112.2. **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 61.4 min, t_{R} (minor) = 21.6 min. **IR:** ν_{max} (thin film): 3056, 2922, 2236, 1654, 1608, 1498, 1364, 1222, 909, 726, 697 cm^{-1} . **HRMS** (ES+): found 462.1872; $\text{C}_{31}\text{H}_{25}\text{FNO}_2$, $[\text{M}+\text{H}]^+$ requires 462.1869. $[\alpha]_{\text{D}}^{293}$ +24.7 (c = 0.6, CHCl_3).

1-(2-(Benzyloxy)phenyl)-3-ethyl-4,6-diphenylpyridin-2(1H)-one (4j)



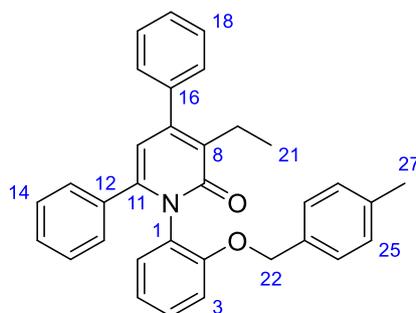
General Procedure D: Pyridone **3c** (100 mg, 272 μmol , 1.0 equiv.), QD-1 (22.5 mg, 40.8 μmol , 15 mol%). CHCl_3 (11 mL) was added followed by 50% aqueous K_2CO_3 (188 μL , 1.36 mmol, 5.0 equiv.) and $\text{R}^{21}\text{-S1}$ (169 μL , 1.36 mmol, 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4j** as a yellow foam (66 mg, 53%, 40% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.48-7.37 (5H, m, H17, 18, 19), 7.36-7.27 (5H, m, H13, 14, 15), 7.21-7.09 (7H, m, H4, 6, 24, 25, 26), 6.92-6.85 (2H, m, H3, 5), 6.14 (1H, s, H10), 5.11-4.92 (2H, m, H22), 2.64-2.55 (2H, m, H20), 2.45 (3H, t, J = 7.3 Hz, H21). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 163.1 (C7), 153.9 (C2), 149.2 (C16), 146.3 (C11), 140.0 (C16), 137.0 (C23), 135.7 (C12), 130.8 (C8), 130.5 (C4), 129.5 (C26), 129.0 (C1), 128.8 (2C, C17), 128.4 (2C, C18), 128.3 (2C, C25), 128.2 (C19), 128.0 (2C, C14), 127.8 (C15), 127.6 (C6), 127.5 (2C, C13), 126.7 (2C, C24), 120.9 (C5), 113.4 (C3), 109.8 (C10), 70.2 (C22), 21.7 (C20), 13.6 (C21). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 69.9 min, t_{R} (minor) = 30.2 min. **IR:** ν_{max} (thin film): 2929, 2240, 1643, 1602, 1494, 1449, 1375, 1271, 1226, 1025, 723, 697 cm^{-1} . **HRMS** (ES+): found 458.2115; $\text{C}_{32}\text{H}_{28}\text{NO}_2$, $[\text{M}+\text{H}]^+$ requires 458.2120. $[\alpha]_{\text{D}}^{293} +6.0$ (c = 0.5, CHCl_3).

4-((2-(3-Ethyl-2-oxo-4,6-diphenylpyridin-1(2H)-yl)phenoxy)methyl)benzonitrile (4k)



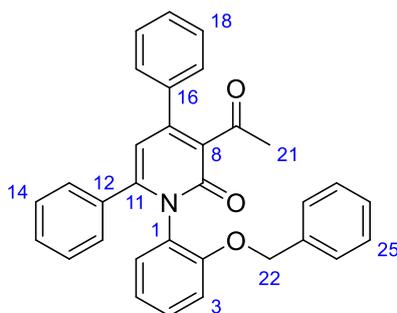
General Procedure D: Pyridone **3c** (50.0 mg, 136 μmol , 1.0 equiv.), QD-1 (11.3 mg, 20.4 μmol , 15 mol%). CHCl_3 (6 mL) was added followed by 50% aqueous K_2CO_3 (94.0 μL , 680 μmol , 5.0 equiv.) and $\text{R}^{21}\text{-S2}$ (165 mg, 680 μmol , 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4k** as a yellow foam (46 mg, 70%, 28% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.64-7.61 (2H, m, H17), 7.50-7.38 (7H, m, H18, 19, 24, 25), 7.22-7.09 (7H, m, H4, 6, 13, 14, 15), 6.93 (1H, td, J = 7.6, 1.2 Hz, H5), 6.81 (1H, dd, J = 8.4, 1.1 Hz, H3), 6.19 (1H, s, H10), 5.17-5.02 (2H, m, H22), 2.66-2.53 (2H, m, H20), 1.16 (1H, t, J = 7.5 Hz, H21). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 163.0 (C7), 153.4 (C2), 149.4 (C9), 146.2 (C11), 142.4 (C23), 139.7 (C12), 135.6 (C16), 132.3 (2C, C25), 130.8 (C8), 130.6 (C19), 129.6 (C4), 129.0 (C1), 128.7 (2C, C18), 128.5 (2C, C14), 128.3 (C15), 128.0 (C5), 127.9 (2C, C24), 127.6 (2C, C13), 127.0 (2C, C17), 121.5 (C6), 118.7 (C27), 113.2 (C3), 111.5 (C26), 110.0 (C10), 69.2 (C22), 21.7 (C20), 13.6 (C21). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 22.3 min, t_{R} (minor) = 17.4 min. **IR:** ν_{max} (thin film): 3056, 2929, 2228, 1640, 1605, 1539, 1494, 1446, 1274, 1021, 909, 726 cm^{-1} . **HRMS** (ES+): found 483.2078; $\text{C}_{33}\text{H}_{27}\text{N}_2\text{O}_2$, $[\text{M}+\text{H}]^+$ requires 483.2073. $[\alpha]_{\text{D}}^{293} +12.0$ (c = 0.4, CHCl_3).

3-Ethyl-1-(2-((4-methylbenzyl)oxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4l)



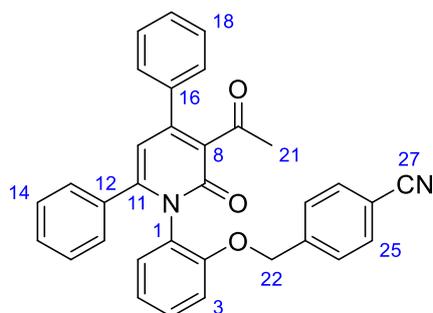
General Procedure D: Pyridone **3c** (50 mg, 136 μmol , 1.0 equiv.), QD-1 (11.3 mg, 20.4 μmol , 15 mol%). CHCl_3 (6 mL) was added followed by 50% aqueous K_2CO_3 (95.0 μL , 680 μmol , 5.0 equiv.) and $\text{R}^{21}\text{-S3}$ (158 mg, 680 μmol , 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4l** as a yellow foam (21 mg, 33%, 41% ee). $^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta_{\text{H}} = 7.48\text{--}7.37$ (5H, m, H17, 18, 19), 7.23–7.07 (11H, m, H4, 6, 13, 14, 15, 22, 23), 6.90–6.86 (2H, m, H3, 5), 6.14 (1H, s, H10), 5.07–4.88 (2H, m, H22), 2.65–2.56 (2H, m, H20), 2.36 (3H, s, H27), 1.89 (3H, t, $J = 7.3$ Hz, H21). $^{13}\text{C NMR}$ (101 MHz CDCl_3): $\delta_{\text{C}} = 163.1$ (C7), 154.0 (C2), 149.2 (C9), 146.3 (C11), 140.0 (C26), 137.3 (C12), 135.8 (C23), 134.0 (C16), 130.8 (C8), 130.5 (C4), 129.5 (C19), 129.0 (2C, C25), 129.0 (C1), 128.8 (2C, C17), 128.3 (2C, C14), 128.1 (C5), 128.1 (2C, C18), 127.8 (C15), 127.5 (2C, C13), 126.9 (2C, C24), 120.8 (C6), 113.4 (C3), 109.8 (C10), 70.2 (C22), 21.7 (C20), 21.2 (C27), 13.6 (C21). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, $\lambda = 250$ nm, t_{R} (major) = 26.9 min, t_{R} (minor) = 25.0 min. **IR:** ν_{max} (thin film): 3026, 2922, 2240, 1643, 1606, 1539, 1474, 1271, 1226, 909, 726, 697 cm^{-1} . **HRMS** (ES+): found 472.2289; $\text{C}_{33}\text{H}_{30}\text{NO}_2$, $[\text{M}+\text{H}]^+$ requires 472.2277. $[\alpha]_{\text{D}}^{293} +8.54$ ($c = 0.4$, CHCl_3).

3-Acetyl-1-(2-(benzyloxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4m)



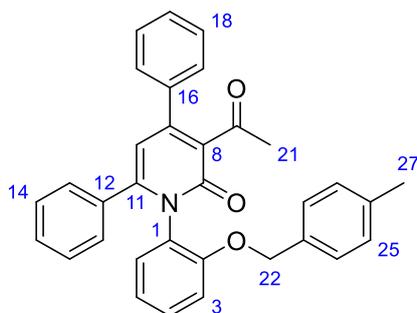
General Procedure D: Pyridone **3b** (100 mg, 262 μmol , 1.0 equiv.), QD-1 (21.6 mg, 39.3 μmol , 15 mol%). CHCl_3 (11 mL) was added followed by 50% aqueous K_2CO_3 (182 μL , 1.31 mmol, 5.0 equiv.) and $\text{R}^{21}\text{-S1}$ (163 μL , 1.31 mmol, 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4m** as a yellow foam (82 mg, 66%, 6% ee). $^1\text{H NMR}$ (400 MHz, CDCl_3): $\delta_{\text{H}} = 7.44\text{--}7.40$ (5H, m, H17, 18, 19), 7.37–7.31 (5H, m, H13, 14, 15), 7.23–7.18 (2H, m, H4, 26), 7.16–7.13 (5H, m, H6, 24, 25), 6.90–6.86 (2H, m, H3, 5), 6.27 (1H, s, H10), 5.11–4.95 (2H, m, H22), 2.45 (3H, s, H21). $^{13}\text{C NMR}$ (101 MHz CDCl_3): $\delta_{\text{C}} = 202.2$ (C2), 160.6 (C7), 153.8 (C9), 150.8 (C2), 150.7 (C11), 137.9 (C16), 136.7 (C21), 135.0 (C12), 130.3 (C19), 130.1 (C4), 129.3 (C8), 129.0 (C15), 128.9 (C26), 128.6 (2C, C17), 128.6 (2C, C18), 128.5 (2C, C24), 128.0 (2C, C14), 127.8 (C6), 127.7 (2C, C13), 127.5 (C1), 126.9 (2C, C24), 121.0 (C5), 113.4 (C3), 109.7 (C8), 70.4 (C22), 31.7 (C21). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, $\lambda = 250$ nm, t_{R} (major) = 11.3 min, t_{R} (minor) = 9.4 min. **IR:** ν_{max} (thin film): 3060, 2243, 1699, 1640, 1528, 1490, 1367, 1271, 909, 752, 730, 697 cm^{-1} . **HRMS** (ES+): found 472.1904; $\text{C}_{32}\text{H}_{26}\text{NO}_3$, $[\text{M}+\text{H}]^+$ requires 472.1913. $[\alpha]_{\text{D}}^{293} +1.3$ ($c = 0.5$, CHCl_3).

4-((2-(3-Acetyl-2-oxo-4,6-diphenylpyridin-1(2H)-yl)phenoxy)methyl)benzonitrile (4n)



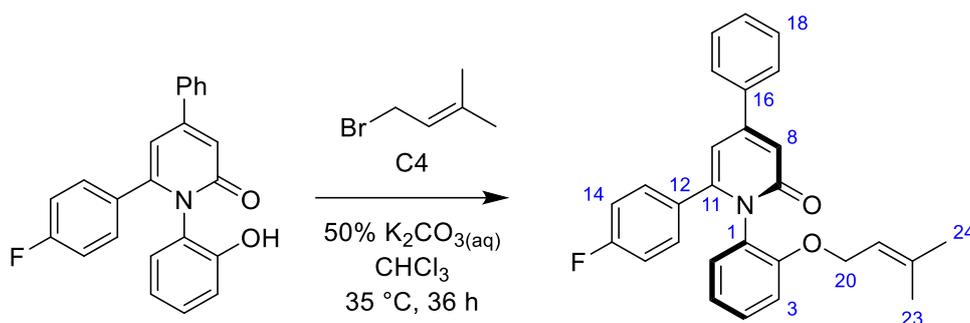
General Procedure D: Pyridone **3b** (50.0 mg, 131 μmol , 1.0 equiv.), QD-1 (10.8 mg, 19.7 μmol , 15 mol%). CHCl_3 (6 mL) was added followed by 50% aqueous K_2CO_3 (91.0 μL , 655 μmol , 5.0 equiv.) and $\text{R}^2\text{I-S2}$ (159 mg, 655 μmol , 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4n** as a yellow foam (45 mg, 69%, 3% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.67-7.64 (2H, m, H17), 7.46-7.40 (7H, m, H18, 19, 24, 25), 7.24-7.12 (7H, m, H4, 6, 13, 14, 15), 6.93 (1H, td, J = 7.6, 1.2 Hz, H5), 6.81 (1H, dd, J = 8.4, 1.2 Hz, H3), 6.31 (1H, s, H10), 5.16-5.05 (2H, m, H22). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 202.13 (C20), 160.5 (C7), 153.4 (C2), 150.6 (C11), 150.5 (C9), 142.0 (C23), 137.6 (C16), 134.9 (C12), 132.5 (2C, C25), 130.4 (C4), 130.2 (C19), 129.5 (C8), 129.2 (C15), 129.0 (C5), 128.8 (2C, C18), 128.5 (2C, C14), 127.9 (2C, C24), 127.9 (2C, C13), 127.6 (C1), 127.3 (2C, C17), 121.6 (C6), 118.7 (C27), 113.3 (C3), 111.8 (C26), 109.8 (C10), 69.5 (C22), 31.7 (C21). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 13.7 min, t_{R} (minor) = 10.9 min. **IR:** ν_{max} (thin film): 3060, 2922, 2228, 1699, 1640, 1602, 1528, 1271, 909, 756, 726 cm^{-1} . **HRMS** (ES $^{+}$): found 497.1862; $\text{C}_{33}\text{H}_{25}\text{N}_2\text{O}_3$, $[\text{M}+\text{H}]^{+}$ requires 497.1865. $[\alpha]_{\text{D}}^{293}$ +0.8 (c = 0.5, CHCl_3).

3-Acetyl-1-(2-((4-methylbenzyl)oxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4o)



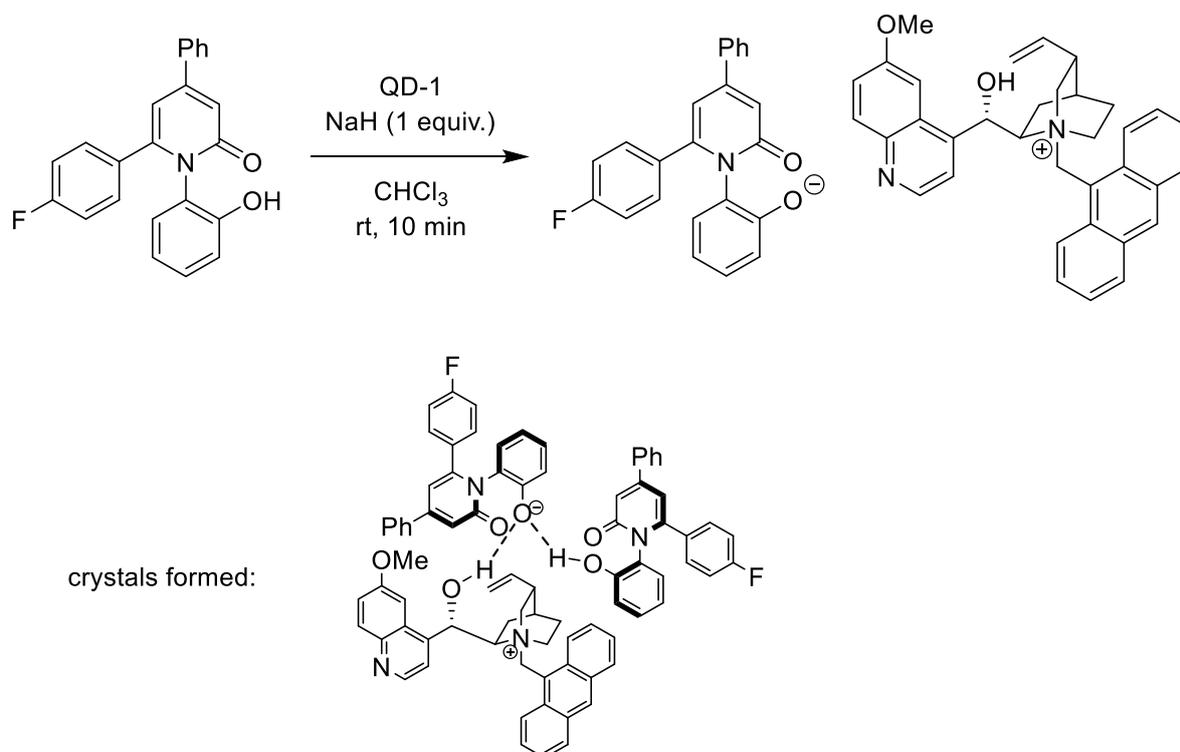
General Procedure D: Pyridone **3b** (50.0 mg, 131 μmol , 1.0 equiv.), QD-1 (10.8 mg, 19.7 μmol , 15 mol%). CHCl_3 (6 mL) was added followed by 50% aqueous K_2CO_3 (91.0 μL , 655 μmol , 5.0 equiv.) and $\text{R}^2\text{I-S3}$ (152 mg, 655 μmol , 5.0 equiv.). Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4o** as an off-white foam (32 mg, 54%, 5% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.42-7.39 (5H, m, H17, 18, 19), 7.24-7.11 (11H, m, H4, 6, 13, 14, 15, 22, 23), 6.90-6.87 (2H, m, H3, 5), 6.26 (1H, s, H10), 5.06-4.91 (2H, m, H22), 2.47 (3H, s, H21), 2.36 (3H, s, H27). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 202.2 (C20), 160.6 (C7), 153.9 (C2), 150.8 (C11), 150.7 (C16), 138.0 (C9), 137.6 (C26), 135.0 (C12), 133.7 (C23), 130.2 (C4), 130.0 (C5), 129.3 (C8), 129.2 (2C, C17), 129.0 (C19), 128.9 (C15), 128.6 (2C, C25), 128.6 (2C, C18), 128.0 (2C, C14), 127.7 (2C, C13), 127.5 (C1), 127.0 (2C, C24), 120.9 (C6), 113.4 (C3), 109.7 (C10), 70.4 (C22), 31.7 (C21), 21.2 (C27). **HPLC:** Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 17.5 min, t_{R} (minor) = 15.6 min. **IR:** ν_{max} (thin film): 3056, 2922, 1699, 1643, 1602, 1531, 1494, 1267, 1028, 801, 760 cm^{-1} . **HRMS** (ES $^{+}$): found 486.2066; $\text{C}_{33}\text{H}_{28}\text{NO}_3$, $[\text{M}+\text{H}]^{+}$ requires 486.2069. $[\alpha]_{\text{D}}^{293}$ +0.9 (c = 0.6, CHCl_3).

2.7 6-(4-Fluorophenyl)-1-(2-((3-methylbut-2-en-1-yl)oxy)phenyl)-4-phenylpyridin-2(1H)-one (4p)



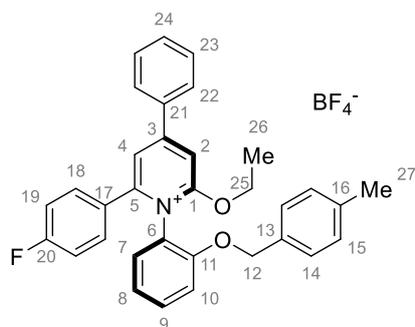
A round-bottomed flask was charged with 2'-hydroxyphenyl-2-pyridone **3e** (50 mg, 140 μmol , 1.0 equiv.), **C4** (7.7 mg, 14.0 μmol , 15 mol%). CHCl_3 (6 mL) was added followed by 50% aqueous K_2CO_3 (97.0 μL , 700 μmol , 5.0 equiv.) and 3,3-dimethylallyl bromide (81 μL , 700 μmol , 5.0 equiv.) as the flask was sealed and stirred at 35 $^\circ\text{C}$ for 18 h. Piperidine (4.0 equiv.) was added and reaction mixture was stirred for 30 mins before addition of 3M HCl (ca. 1 mL). Crude product was extracted with EtOAc (3 x10 mL/mmol), dried over anhydrous magnesium sulfate and the solvent removed *in vacuo* resulting in a crude solid. Crude mixture purified by flash column chromatography (silica gel, 1% \rightarrow 2.5% MeOH/dichloromethane) affording **4p** as a yellow foam (47 mg, 59%, 47% ee). **$^1\text{H NMR}$** (400 MHz, CDCl_3): δ_{H} = 7.69-7.66 (2H, m, H17), 7.50-7.42 (3H, m, H18, 19), 7.22-7.16 (3H, m, H4, 22), 7.10 (1H, dd, J = 7.7, 1.7 Hz, H6), 6.91-6.78 (5H, m, H3, 5, 10, 14), 6.47 (1H, d, J = 2.0 Hz, H8), 5.31-5.27 (1H, m, H21), 4.51-4.35 (2H, m, H20), 1.74 (3H, s, H24), 1.68 (3H, s, H23). **$^{13}\text{C NMR}$** (101 MHz CDCl_3): δ_{C} = 163.8 (C7), 162.5 (d, $^1J_{\text{CF}}$ = 248.7 Hz, C15), 153.7 (C9), 151.1 (C2), 149.2 (C11), 137.7 (C16), 137.0 (C22), 131.9 (d, $^4J_{\text{CF}}$ = 3.6 Hz, C12), 130.7 (d, $^3J_{\text{CF}}$ = 8.4 Hz, C13), 130.3 (C4), 129.9 (C19), 129.5 (C5), 129.0 (2C, C17), 127.7 (C1), 126.8 (2C, C18), 120.5 (C5), 119.8 (C21), 116.3 (C8), 114.6 (d, $^2J_{\text{CF}}$ = 21.8 Hz, C14), 113.2 (C3), 106.9 (C10), 65.5 (C20), 25.7 (C24), 18.3 (C23). **$^{19}\text{F NMR}$** (376 MHz, CDCl_3): δ_{F} = -112.3. **HPLC**: Chiralpak OD-RH, 60% MeCN/ H_2O , 0.75 mL/min, λ = 250 nm, t_{R} (major) = 29.5 min, t_{R} (minor) = 13.7 min. **IR**: ν_{max} (thin film): 3060, 2922, 1658, 1605, 1498, 1364, 1267, 1222, 998, 842, 749, 697 cm^{-1} . **HRMS** (ES $^+$): found 426.1867; $\text{C}_{28}\text{H}_{25}\text{NO}_2$, $[\text{M}+\text{H}]^+$ requires 426.1869. $[\alpha]_{\text{D}}^{293}$ +19.5 (c = 0.4, CHCl_3).

2.8 Catalyst-substrate co-crystal (6)



A flask was charged with pyridone **3c** (25 mg, 70.0 μmol , 1.0 equiv.) and evacuated/backfilled with Ar under vacuum. THF (3mL) was added followed by NaH (2.0 mg, 70.0 μmol , 1.0 equiv.) as the flask was sealed and stirred at room temperature for 10 minutes. Under Ar, QD-1 was added and reaction mixture left to stir for a further 10 minutes. The reaction mixture was washed through a celite pad with EtOAc (25 mL) and reduced in vacuo affording a yellow residue. Recrystallization from hot EtOAc was attempted but with no success, the solution was reduced to dryness and CHCl₃ was added. A small portion (ca. 0.2 mL) was removed and subjected to vapour diffusion, using n-hexane (ca. 1 mL) as the anti-solvent, affording yellow crystals.

2.9 Synthesis of Pyridinium Salt 7



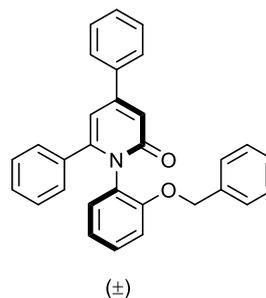
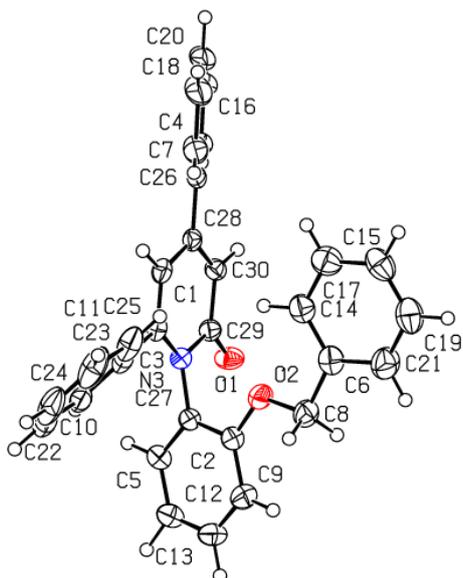
Triethyloxonium tetrafluoroborate (13 mg, 0.068 mmol) was added in one portion to a stirred solution of pyridone **4i** (15 mg, 0.032 mmol) in anhydrous dichloromethane (1 mL). After 1 h the reaction mixture was diluted with dichloromethane (5 mL) and washed with sodium bicarbonate (sat. aq., 5 mL). The aqueous phase was extracted with dichloromethane (3 x 5 mL). The organic extracts were combined, dried over anhydrous magnesium sulfate, filtered and concentrated *in vacuo*. The crude residue was purified by flash column chromatography (SiO₂, 2% MeOH in dichloromethane) to afford the product **7** salt as a beige solid (17 mg, 91%). δ_H (CDCl₃, 600 MHz): 7.93-7.89 (2H, m, ArC), 7.65 (1H, s, H2), 7.60-7.54 (4H, m, ArH), 7.50 (1H, s, H4), 7.33 (1H, t, *J* 7.9, ArH), 7.29-7.27 (2H, m, ArH), 7.14 (2H, d, *J* 7.8, H15), 7.07 (2H, d, *J* 7.8, H14), 6.99 (1H, t, *J* 7.7, ArH), 6.94-6.89 (3H, m, ArH), 5.02 (1H, dd, *J* 12.0, H12), 4.96 (1H, dd, *J* 12.0, H12'), 4.75-4.69 (1H, m, H25), 4.57-4.50 (1H, m, H25'), 2.34 (3H, s, H27), 1.30 (3H, t, *J* 7.0, H26); δ_C (CDCl₃, 151 Hz) 163.6 (d, *J* 252.6), 160.9, 160.4, 153.2, 152.2 (C11), 138.5, 135.0, 132.5, 132.03, 132.01, 131.4 (d, *J* 8.8, C18), 129.73, 129.68, 129.5 (C15), 128.4, 128.3, 127.0 (C14), 125.0, 121.7, 118.4 (C4), 115.5 (d, *J* 22.1, C19), 113.0, 107.3 (C2), 70.5 (C12), 69.9 (C25), 21.9 (C27), 13.9 (C26); δ_F (CDCl₃, 564 MHz): -108.7 (1F, s, Ar-F), 153.4 (4F, s, BF₄⁻); **HRMS (ES⁺)** found 490.2165 (C₃₃H₂₉NO₂F⁺, [M-BF₄]⁺ requires 490.2182), **HRMS (ES⁻)** found 86.9950 ([BF₄]⁻ requires 87.0029); 664.2233 (C₃₃H₂₉B₂NO₂F₉⁻, [M+BF₄]⁻ requires 664.2241). **HPLC**: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, *t_R* (major) = 54.9 min, *t_R* (minor) = 20.9 min, 49% ee; [α]_D²⁹³ +49.6 (c = 0.4, CHCl₃).

3 X-Ray Crystallography

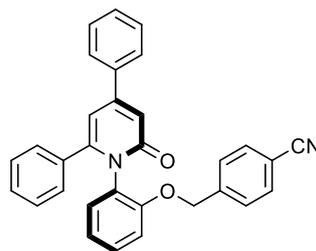
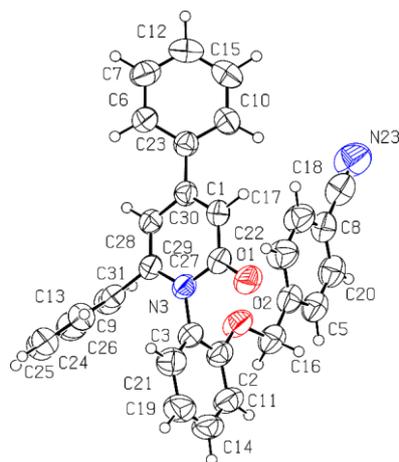
Low temperature¹² and room temperature single crystal X-ray diffraction studies for **4a**, **4b**, **4j**, **4m** were carried out using CuK α radiation on an Agilent Supernova diffractometer equipped with an area detector and graphite monochromator. Raw frame data were reduced using CrysAlisPRO¹³ and solved using SHELXT 2015.¹⁴ Full-matrix least-squares refinement of the structures were carried out using CRYSTALS.¹⁵ The absolute stereochemistry of **4b** was found using the method of Flack.¹⁶

X-ray diffraction studies for **3a** were conducted using CuK α on a Rigaku 007HF equipped with Varimax confocal mirrors and an AFC11 goniometer and HyPix 6000 detector, at the National Crystallography Service in the University of Southampton. Raw frame data were reduced using CrysAlisPRO^[2] and solved using SHELXT.¹⁴ Full-matrix least-squared refinement of the structure was carried out using SHELXL¹⁷ and visualised using Olex 1.3.¹⁸ A void containing disordered solvent molecules necessitated the use of a solvent mask. A mask was calculated, and 1559 electrons were found in a volume of 5917 Å³ in 1 void per unit cell. This is consistent with the presence of 3[CHCl₃] per asymmetric unit. One of the pyridone phenol moieties is disordered, so was refined over two positions (occupancies 0.63 and 0.37).

Full refinement details are given in the supplementary material (CIF). CCDC 2124211 (**4a**), 2124210 (**4b**), 2124208 (**4j**), 2124209 (**4m**) and 2124143 (**6**) contain the supplementary crystallographic data for this paper. These data are provided free of charge by The Cambridge Crystallographic Data Centre and copies can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif.

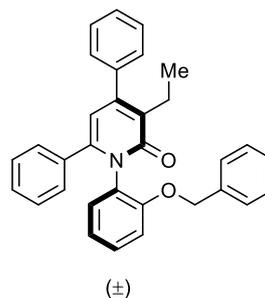
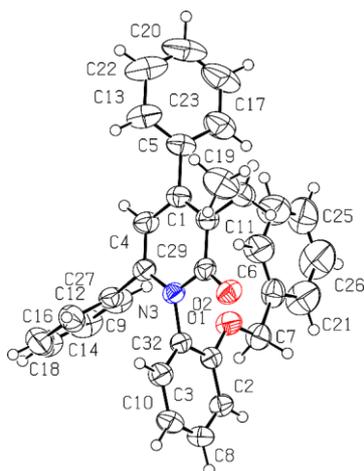
(±)-4a (CCDC 2124211)

Crystal data	
Chemical formula	$C_{30}H_{23}NO_2$
M_r	429.52
Crystal system, space group	Monoclinic, $I2/a$
Temperature (K)	100
a, b, c (Å)	18.54, 10.37, 24.28
β (°)	103.312 (3)
V (Å ³)	4542.64 (6)
Z	8
Radiation type	Cu $K\alpha$
μ (mm ⁻¹)	0.62
Crystal size (mm)	0.20 × 0.16 × 0.16
Data collection	
Diffractometer	Oxford Diffraction SuperNova
Absorption correction	Multi-scan <i>CrysAlis PRO</i> (Rigaku Oxford Diffraction, 2017)
T_{min}, T_{max}	0.78, 0.91
No. of measured, independent and observed [$I > 2.0\sigma(I)$] reflections	22074, 4384, 3995
R_{int}	0.019
$(\sin \theta/\lambda)_{max}$ (Å ⁻¹)	0.615
Refinement	
$R[F^2 > 2\sigma(F^2)], wR(F^2), S$	0.048, 0.125, 1.00
No. of reflections	4384
No. of parameters	299
H-atom treatment	H-atom parameters constrained
$\Delta\rho_{max}, \Delta\rho_{min}$ (e Å ⁻³)	1.05, -0.17

(+)-4b (CCDC 2124210)

Crystal data	
Chemical formula	$C_{62}H_{44}N_4O_4$
M_r	909.06
Crystal system, space group	Monoclinic, $P2_1$
Temperature (K)	299
a, b, c (Å)	9.3537 (3), 12.1471 (4), 11.0218 (3)
β (°)	104.661 (3)
V (Å ³)	1211.52 (6)
Z	1
Radiation type	Cu $K\alpha$
μ (mm ⁻¹)	0.62
Crystal size (mm)	0.16 × 0.16 × 0.12
Data collection	
Diffractometer	Oxford Diffraction SuperNova
Absorption correction	Multi-scan <i>CrysAlis PRO</i> (Rigaku Oxford Diffraction, 2017)
T_{min}, T_{max}	0.76, 0.93
No. of measured, independent and observed [$I > 2.0\sigma(I)$] reflections	6917, 4528, 4016
R_{int}	0.000
$(\sin \theta/\lambda)_{max}$ (Å ⁻¹)	0.615
Refinement	
$R[F^2 > 2\sigma(F^2)], wR(F^2), S$	0.037, 0.094, 0.95
No. of reflections	4528
No. of parameters	317
No. of restraints	1
H-atom treatment	H-atom parameters not refined
$\Delta\rho_{max}, \Delta\rho_{min}$ (e Å ⁻³)	0.18, -0.27
Absolute structure	Parsons, Flack & Wagner (2013), 1957 Friedel Pairs
Absolute structure parameter	0.02 (15)

(±)-4j (CCDC 2124208)



Crystal data

Chemical formula

C₃₂H₂₇NO₂

*M*_r

457.55

Crystal system, space group

Monoclinic, *I*2/a

Temperature (K)

293

a, *b*, *c* (Å)

21.811899 (14), 9.84050 (1), 23.848099 (14)

β (°)

101.3470 (18)

V (Å³)

5018.70 (3)

Z

8

Radiation type

Cu Kα

μ (mm⁻¹)

0.59

Crystal size (mm)

0.30 × 0.28 × 0.18

Data collection

Diffractometer

Oxford Diffraction SuperNova

Absorption correction

Multi-scan

CrysAlis PRO (Rigaku Oxford Diffraction, 2017)

*T*_{min}, *T*_{max}

0.67, 0.90

No. of measured, independent and observed [*I* > 2.0σ(*I*)] reflections

45155, 4846, 4399

*R*_{int}

0.021

(sin θ/λ)_{max} (Å⁻¹)

0.615

Refinement

R [*F*² > 2σ(*F*²)], *wR* (*F*²), *S*

0.041, 0.110, 0.88

No. of reflections

4846

No. of parameters

316

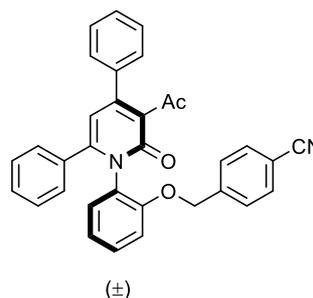
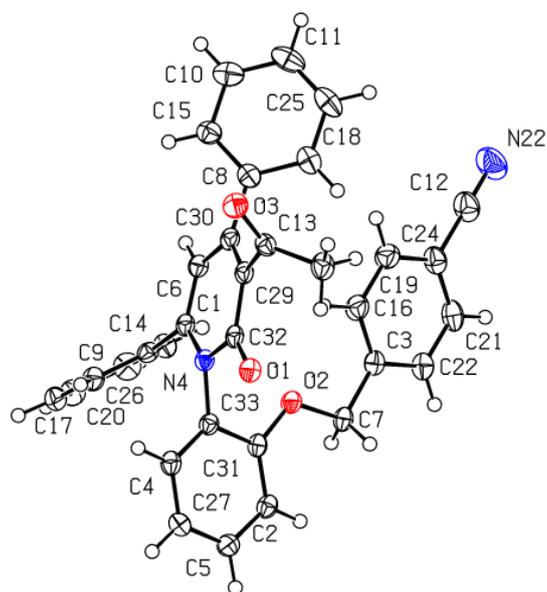
H-atom treatment

H-atom parameters constrained

Δρ_{max}, Δρ_{min} (e Å⁻³)

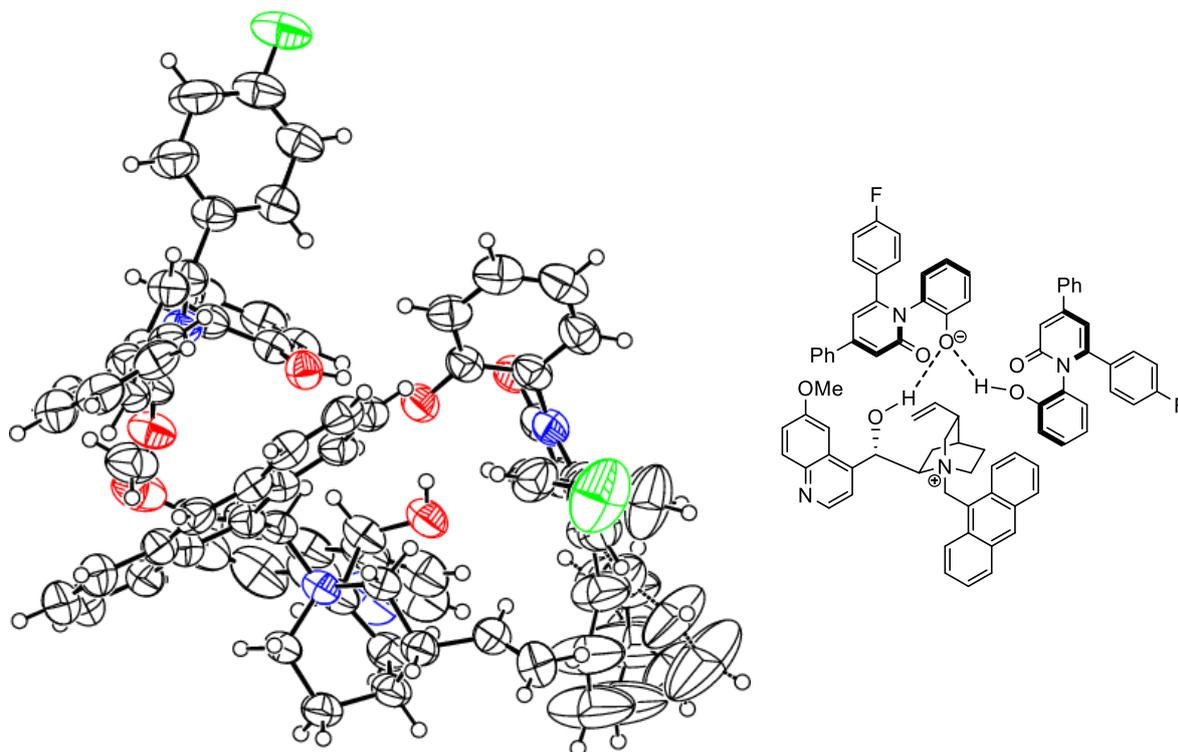
0.13, -0.18

(±)-4m (CCDC 2124209)



Crystal data	
Chemical formula	C ₁₃₂ H ₉₆ N ₈ O ₁₂
<i>M_r</i>	1986.26
Crystal system, space group	Monoclinic, <i>P2₁/n</i>
Temperature (K)	100
<i>a</i> , <i>b</i> , <i>c</i> (Å)	14.759600 (14), 9.835999 (10), 17.417799 (14)
β (°)	94.4720 (18)
<i>V</i> (Å ³)	2520.94 (1)
<i>Z</i>	1
Radiation type	Cu Kα
μ (mm ⁻¹)	0.67
Crystal size (mm)	0.30 × 0.10 × 0.10
Data collection	
Diffractometer	Oxford Diffraction SuperNova
Absorption correction	Multi-scan <i>CrysAlis PRO</i> (Rigaku Oxford Diffraction, 2017)
<i>T_{min}</i> , <i>T_{max}</i>	0.93, 0.93
No. of measured, independent and observed [<i>I</i> > 2.0σ(<i>I</i>)] reflections	23616, 4859, 4283
<i>R_{int}</i>	0.122
(sin θ/λ) _{max} (Å ⁻¹)	0.615
Refinement	
<i>R</i> [<i>F</i> ² > 2σ(<i>F</i> ²)], <i>wR</i> (<i>F</i> ²), <i>S</i>	0.052, 0.138, 0.96
No. of reflections	4859
No. of parameters	343
H-atom treatment	H-atom parameters constrained
Δρ _{max} , Δρ _{min} (e Å ⁻³)	0.33, -0.30

6 (CCDC 2124143)



Crystal data

Chemical formula $C_{23}H_{16}FNO_2 \cdot C_{23}H_{15}FNO_2 \cdot C_{35}H_{35}N_2O_2$ M_r 1229.37Crystal system, space group Trigonal, $R\bar{3}$

Temperature (K) 100

 a, c (Å) 28.7071 (4), 25.9528 (4) V (Å³) 18522.2 (6) Z 9Radiation type Cu $K\alpha$ μ (mm⁻¹) 0.53

Crystal size (mm) 0.22 × 0.04 × 0.04

Data collection

Diffractometer Rigaku 007HF
diffractometer equipped with Varimax confocal mirrors, an AFC11 goniometer and HyPix 6000 detector

Absorption correction Multi-scan
CrysAlis PRO 1.171.41.113a (Rigaku Oxford Diffraction, 2021) Empirical absorption correction using spherical harmonics, implemented in SCALE3 ABSPACK scaling algorithm.

 T_{\min}, T_{\max} 0.393, 1.000

No. of measured, independent and observed [$I > 2\sigma(I)$] reflections 72024, 14182, 10313

R_{int}	0.051
$(\sin \theta/\lambda)_{\text{max}}$ (\AA^{-1})	0.595
Refinement	
$R[F^2 > 2\sigma(F^2)],$ $wR(F^2), S$	0.040, 0.110, 1.02
No. of reflections	14182
No. of parameters	896
No. of restraints	841
H-atom treatment	H-atom parameters constrained
$\Delta\rho_{\text{max}}, \Delta\rho_{\text{min}}$ (e \AA^{-3})	0.15, -0.11
Absolute structure	Flack x determined using 3880 quotients $[(I+)-(I-)]/[(I+)+(I-)]$ (Parsons, Flack and Wagner, Acta Cryst. B69 (2013) 249-259).
Absolute structure parameter	0.01 (8)

4 Measurement of Rotational Barriers

The phenol starting materials **3a-e** underwent rapid racemisation at room temperature, necessitating the separation of racemic mixtures by HPLC on a chiral stationary phase immediately prior to the thermal racemisation analysis.

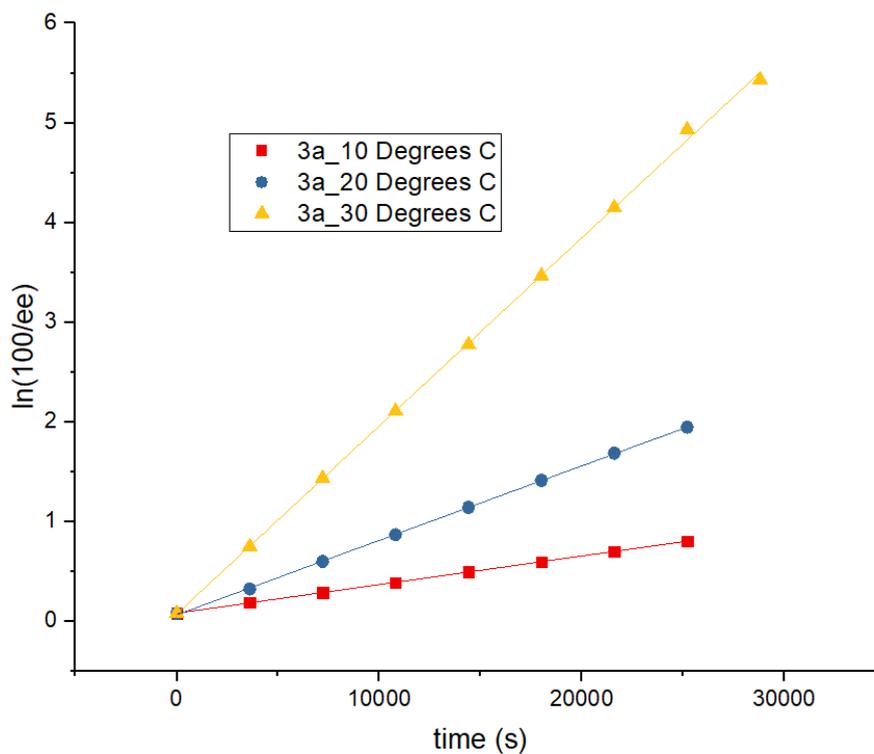
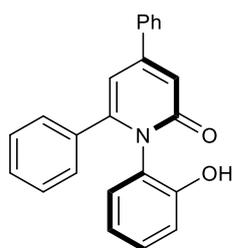
General Procedure E – Measurement of Barriers in Phenols 3a-3e

A sample of racemic 2'-hydroxyphenyl-2-pyridone was separated on HPLC (3a, d and e, Chiralpak OD-RH, 60:40 MeCN/H₂O, 0.75 mL/min) or (3b and c, SB-Cellulose, 20:80 IPA/hexane, 1 mL/min) and each enantiomer semi-preparatively collected to ensure two enantioenriched samples of each enantiomer of the respective 2'-hydroxyphenyl-2-pyridone. The samples were stored at 0 °C until sampled as soon as possible for HPLC analysis (under the same conditions required for initial enantiomer separation) over 7-12 hours with the column and the sample tray held at the desired temperature (10, 20 or 30 °C) until analysis was complete. Compounds were sampled in either MeCN (60 µL, 3a, d and e) or IPA (60 µL, 3b and c).

General Procedure F – Measurement of Barriers in Products 4a-4p

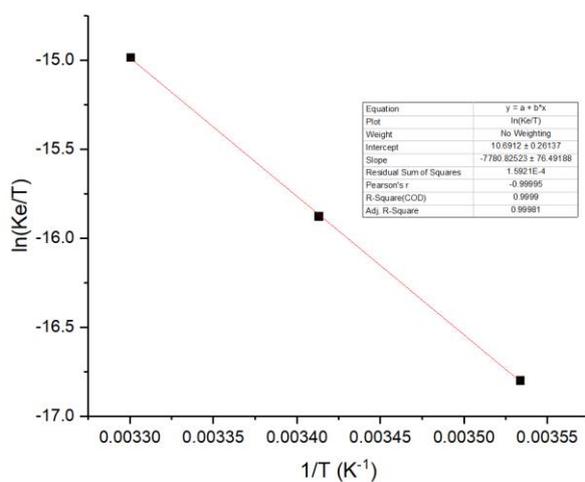
A sample of the benzylated product (1-2 mg) was dissolved in DMSO (80 µL), placed in a glass vial and immersed in a temperature-controlled oil bath ($t = 0$) set to the desired temperature (60, 70 and 80 °C). Samples were removed using the tip of a pasteur pipette at a series of time-points (e.g. $t = 1$ h, 2 h, 3 h, 4 h, 6 h, 16 h, 30 h, 54 h, 76 h). Each sample was added to an HPLC vial containing acetonitrile (1 mL) and its *ee* measured by HPLC on a chiral stationary phase (Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min).

3a



Equation	y = a + b*x		
	ln(100/ee)	ln(100/ee)	ln(100/ee)
Plot			
Weight	No Weighting		
Intercept	0.08202 ± 0.00118	0.06561 ± 0.00469	0.07507 ± 0.02999
Slope	2.86714E-5 ± 7.8339E-8	7.47416E-5 ± 3.11499E-7	1.88598E-4 ± 1.74965E-6
Residual Sum of Squares	2.00429E-5	3.16897E-4	0.01666
Pearson's r	0.99998	0.99995	0.9997
R-Square(COD)	0.99996	0.9999	0.9994
Adj. R-Square	0.99995	0.99988	0.99931

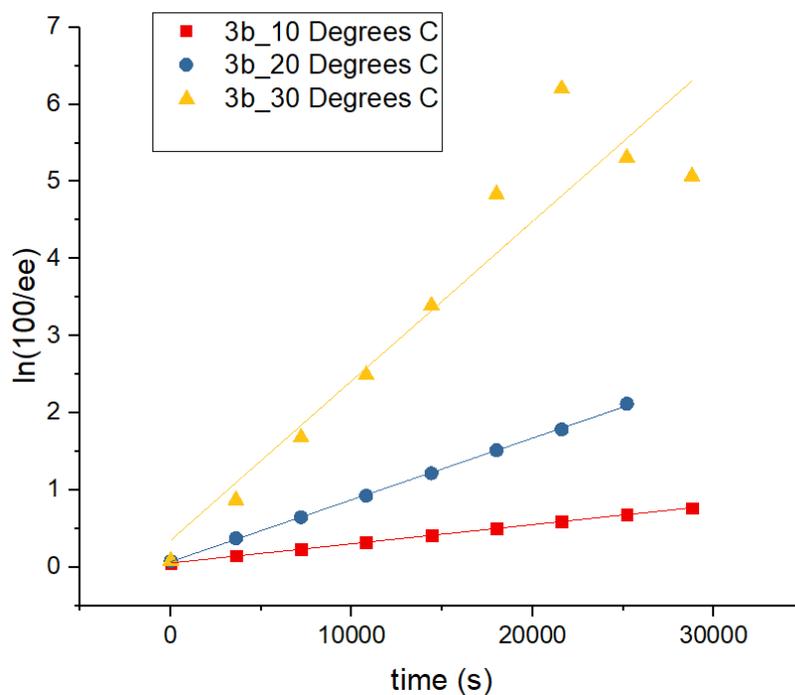
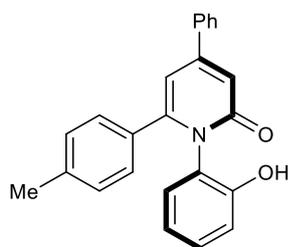
	T (K)		
	283	293	303
k_{rac} (s ⁻¹)	2.867E-05	7.474E-05	1.886E-04
k_{ent} (s ⁻¹)	1.434E-05	3.737E-05	9.430E-05
ΔG_T^\ddagger (kJ mol ⁻¹)	95.43	96.56	97.60
$t_{1/2}(rac)_T$ (min)	402.93	154.57	61.25



$$\Delta H_{ent}^\ddagger = 64.7 \text{ kJ mol}^{-1}$$

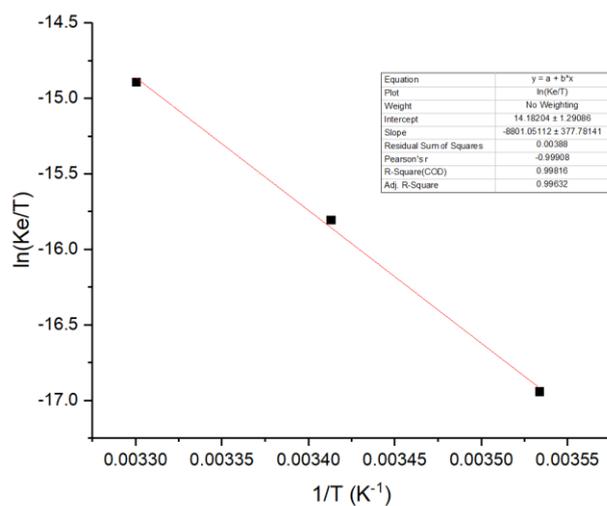
$$\Delta S_{ent}^\ddagger = -108.7 \text{ J K}^{-1} \text{ mol}^{-1}$$

3b



Equation	y = a + b*x		
	ln(100/ee)	ln(100/ee)	ln(100/ee)
Plot			
Weight	No Weighting		
Intercept	0.05367 ± 0.00242	0.07136 ± 0.01036	0.34948 ± 0.47954
Slope	2.4848E-5 ± 1.4133E-7	8.01546E-5 ± 6.8824E-7	2.06711E-4 ± 2.79789E-5
Residual Sum of Squares	1.08723E-4	0.00155	4.26104
Pearson's r	0.99989	0.99978	0.94145
R-Square(COD)	0.99977	0.99956	0.88633
Adj. R-Square	0.99974	0.99948	0.8701

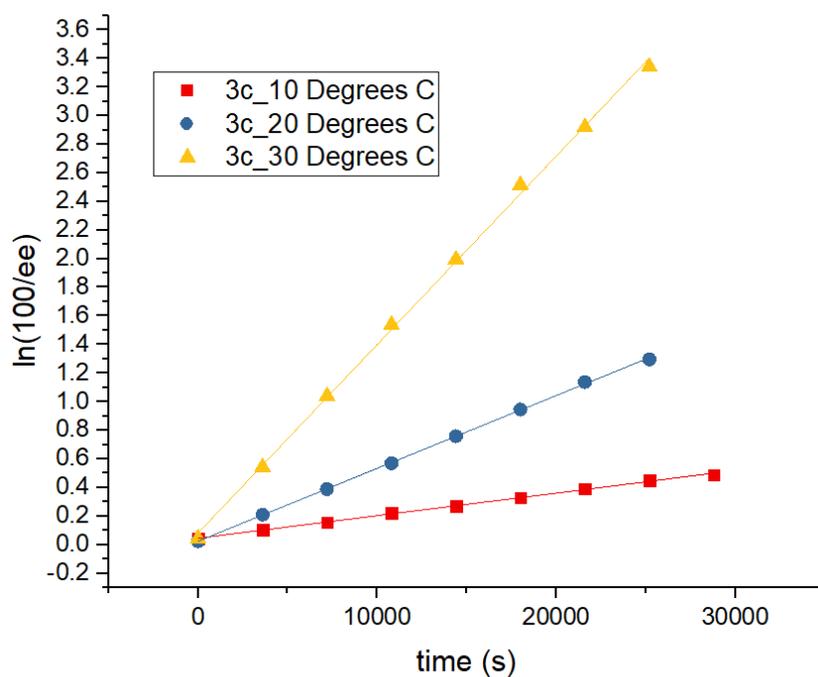
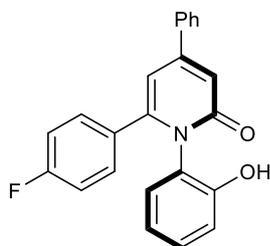
	T (K)		
	283	293	303
k_{rac} (s ⁻¹)	2.485E-05	8.015E-05	2.067E-04
k_{ent} (s ⁻¹)	1.242E-05	4.008E-05	1.034E-04
ΔG_T^\ddagger (kJ mol ⁻¹)	95.77	96.39	97.37
$t_{1/2}(rac)_T$ (min)	464.92	144.13	55.89



$$\Delta H_{ent}^\ddagger = 73.2 \text{ kJ mol}^{-1}$$

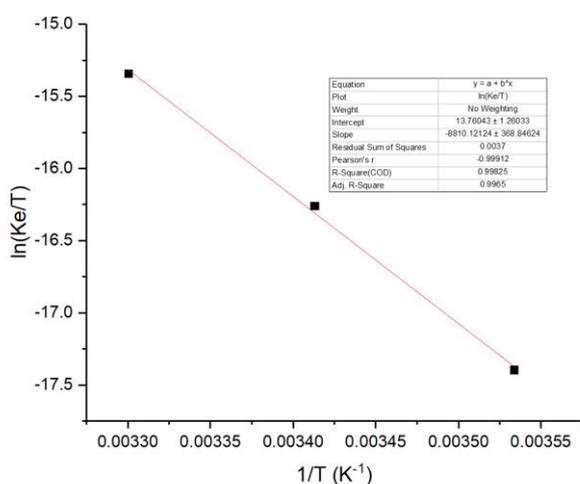
$$\Delta S_{ent}^\ddagger = -79.6 \text{ J K}^{-1} \text{ mol}^{-1}$$

3c



Equation	y = a + b*x		
Plot	ln(100/ee)		
Weight	No Weighting		
Intercept	0.04601 ± 0.00428	0.025 ± 0.00493	0.08217 ± 0.02716
Slope	1.57896E-5 ± 2.49711E-7	5.09242E-5 ± 3.27242E-7	1.31733E-4 ± 1.80372E-6
Residual Sum of Squares	3.38413E-4	3.4874E-4	0.01063
Pearson's r	0.99913	0.99988	0.99944
R-Square(COD)	0.99825	0.99975	0.99888
Adj. R-Square	0.998	0.99971	0.99869

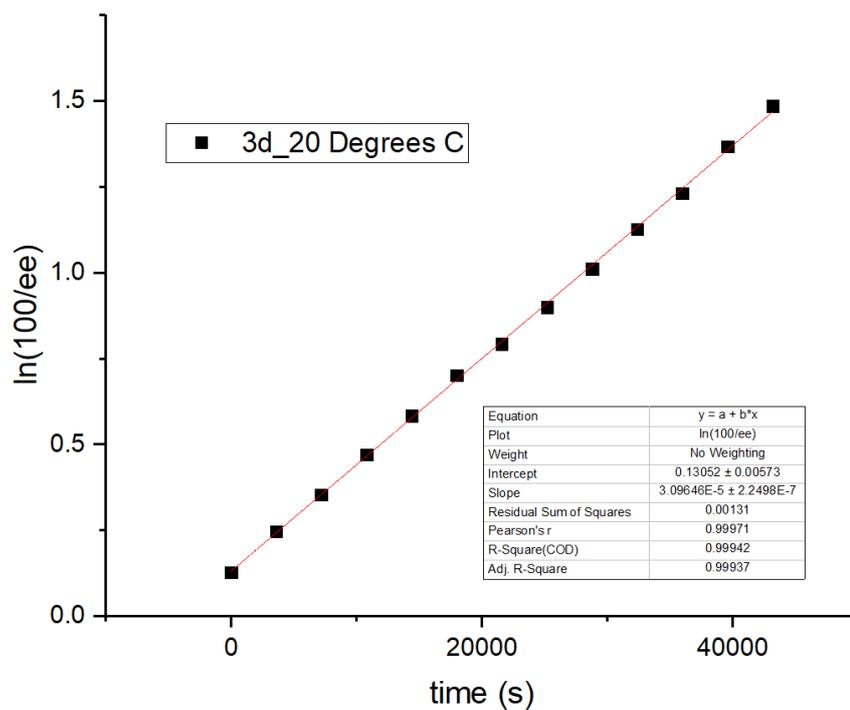
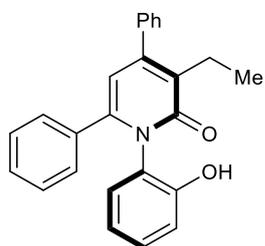
	T (K)		
	283	293	303
k_{rac} (s ⁻¹)	1.579E-05	5.092E-05	1.317E-04
k_{ent} (s ⁻¹)	7.895E-06	2.546E-05	6.587E-05
ΔG_T^\ddagger (kJ mol ⁻¹)	96.84	97.49	98.51
$t_{1/2}(rac)_T$ (min)	731.65	226.86	87.70



$$\Delta H_{ent}^\ddagger = 73.3 \text{ kJ mol}^{-1}$$

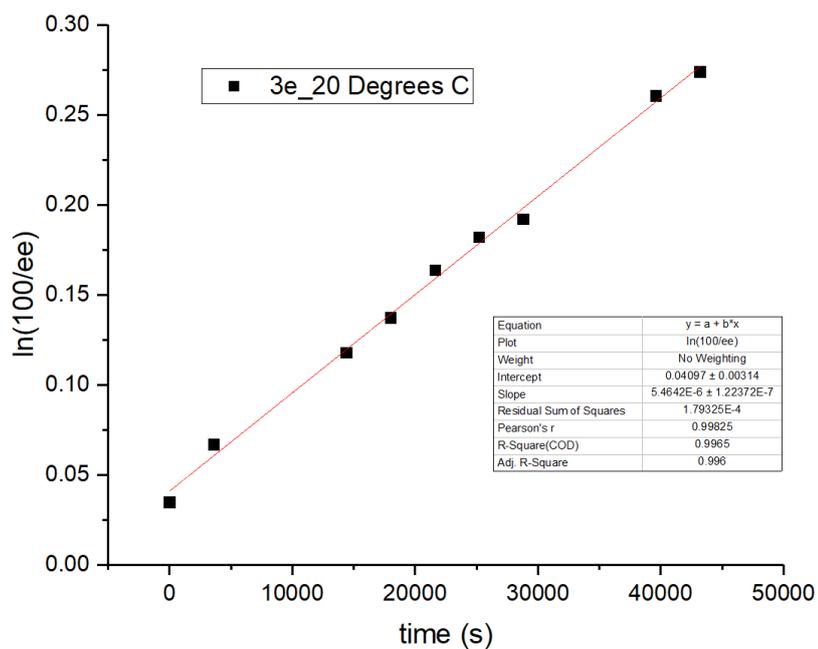
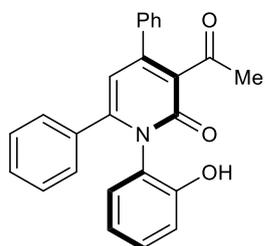
$$\Delta S_{ent}^\ddagger = -83.1 \text{ J K}^{-1} \text{ mol}^{-1}$$

3d



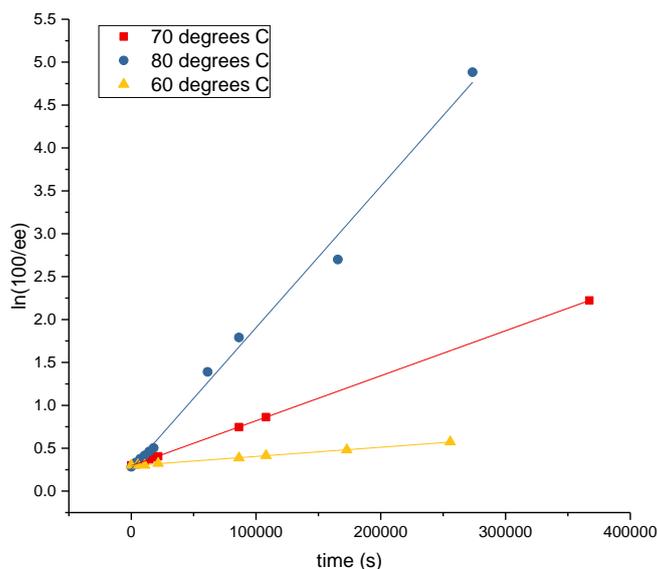
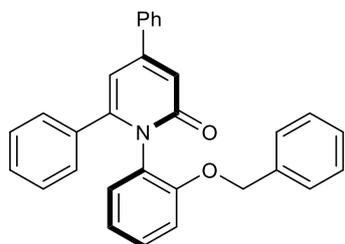
	T (K)
	393
k_{rac} (s ⁻¹)	3.096E-05
k_{ent} (s ⁻¹)	1.548E-05
ΔG_T^\ddagger (kJ mol ⁻¹)	98.70
$t_{1/2}(rac)_T$ (min)	373.09

3e



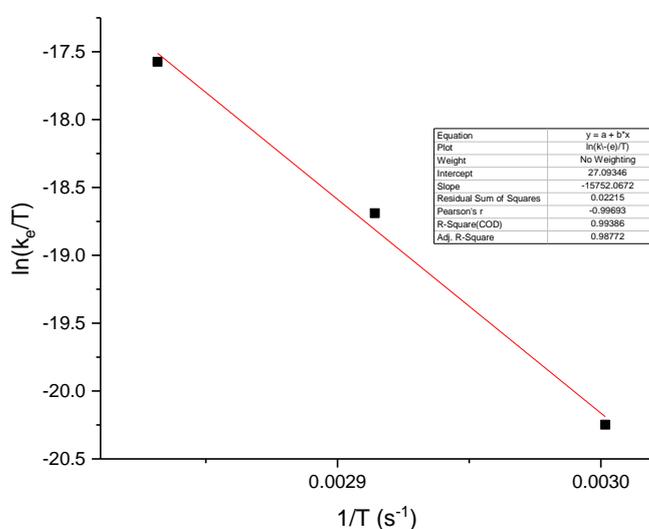
	T (K)
	393
k_{rac} (s ⁻¹)	5.464E-06
k_{ent} (s ⁻¹)	2.732E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	102.93
$t_{1/2}(rac)_T$ (min)	2114.21

4a



Equation	y = a + b*x		
Plot	ln(100/ee)	ln(100/ee)	ln(100/ee)
Weight	No Weighting		
Intercept	0.2954	0.25744	0.29711
Slope	5.24593E-6	1.64779E-5	1.0719E-6
Residual Sum of Square	7.70709E-5	0.12879	1.33402E-4
Pearson's r	0.99999	0.99677	0.99904
R-Square(COD)	0.99998	0.99356	0.99808
Adj. R-Square	0.99997	0.99275	0.99776

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	1.072E-06	5.246E-06	1.648E-05
k_{ent} (s ⁻¹)	5.360E-07	2.623E-06	8.239E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	121.84	121.06	121.31
$t_{1/2}(rac)_T$ (min)	10777.55	2202.17	701.09



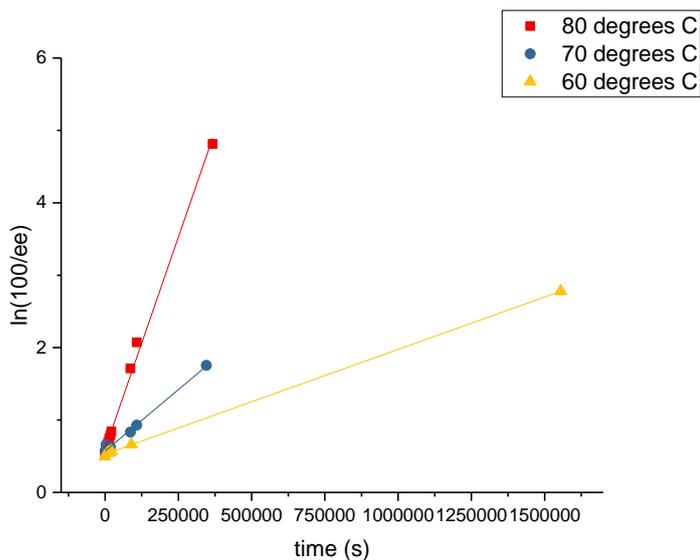
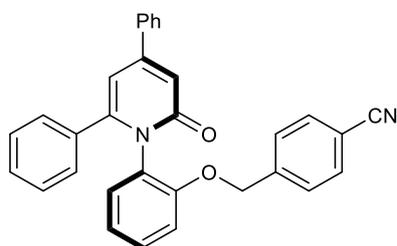
Equation	y = a + b*x
Plot	ln(k _e /T)
Weight	No Weighting
Intercept	27.09346
Slope	-15752.0672
Residual Sum of Squares	0.02215
Pearson's r	-0.99693
R-Square(COD)	0.99386
Adj. R-Square	0.98772

$$\Delta H_{ent}^\ddagger = 131.0 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = +27.7 \text{ J K}^{-1} \text{ mol}^{-1}$$

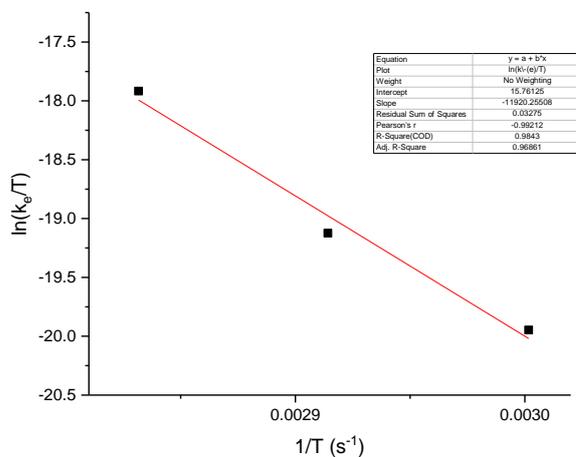
Extrapolated half-life of racemization at 20°C: 14.3 y

4b



Equation	y = a + b*x		
Plot	ln(100/ee)	ln(100/ee)	ln(100/ee)
Weight	No Weighting		
Intercept	0.60148	0.57021	0.52778
Slope	1.16899E-5	3.39693E-5	1.44743E-5
Residual Sum of Squares	0.0704	0.00759	0.00132
Pearson's r	0.99776	0.99673	0.99984
R-Square(COD)	0.99552	0.99347	0.99969
Adj. R-Square	0.99496	0.99254	0.99963

	T (K)		T (K)	
	333		333	
k_{rac} (s ⁻¹)	1.447E-06		1.447E-06	
k_{ent} (s ⁻¹)	7.237E-07		7.237E-07	
ΔG_T^\ddagger (kJ mol ⁻¹)	121.01		121.01	
$t_{1/2}(rac)_T$ (min)	7981.36		7981.36	



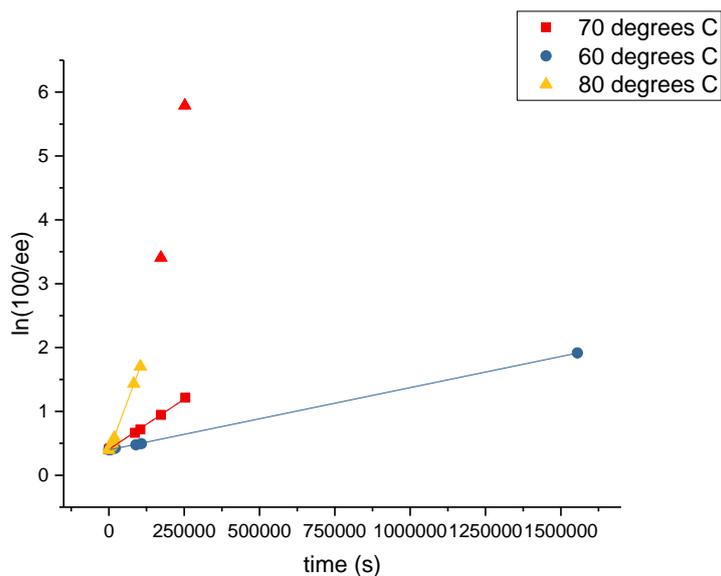
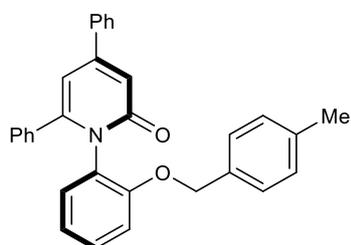
Equation	y = a + b*x
Plot	ln(k _r /T)
Weight	No Weighting
Intercept	15.78125
Slope	-11920.25508
Residual Sum of Squares	0.03275
Pearson's r	-0.99212
R-Square(COD)	0.9843
Adj. R-Square	0.98061

$$\Delta H_{ent}^\ddagger = 99.1 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -66.5 \text{ J K}^{-1} \text{ mol}^{-1}$$

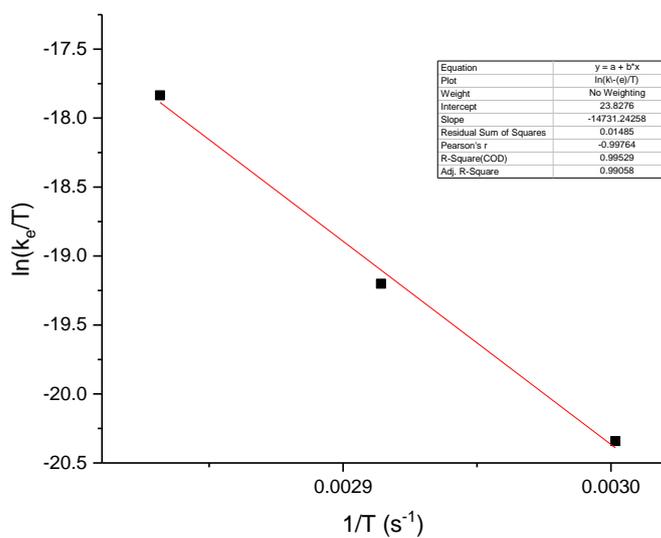
Extrapolated half-life of
racemization at 20°C: 2.5 y

4C



Equation	y = a + b*x		
Plot	ln(100/ee)	ln(100/ee)	ln(100/ee)
Weight	No Weighting		
Intercept	0.40612	0.39669	0.37596
Slope	3.14403E-6	9.75417E-7	1.28364E-5
Residual Sum of Squares	0.0012	1.63851E-4	0.00197
Pearson's r	0.99911	0.99996	0.99945
R-Square(COD)	0.99822	0.99992	0.99891
Adj. R-Square	0.998	0.9999	0.99872

	T (K)	
	333	333
k_{rac} (s^{-1})	9.754E-07	9.754E-07
k_{ent} (s^{-1})	4.877E-07	4.877E-07
ΔG_T^\ddagger ($kJ\ mol^{-1}$)	122.11	122.11
$t_{1/2}(rac)_T$ (min)	11843.60	11843.60

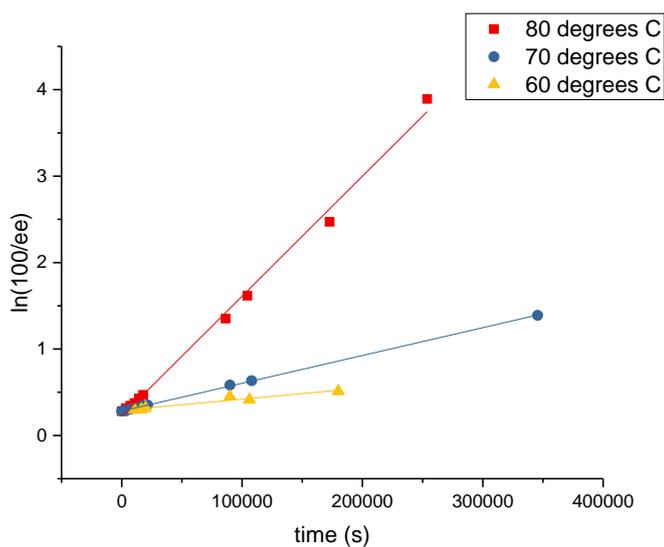
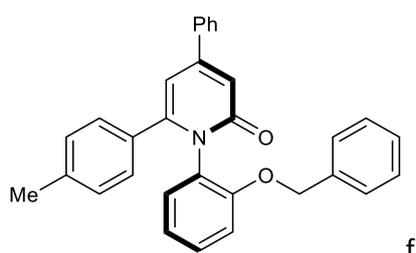


$$\Delta H_{ent}^\ddagger = 122.5\ kJ\ mol^{-1}$$

$$\Delta S_{ent}^\ddagger = +0.6\ J\ K^{-1}\ mol^{-1}$$

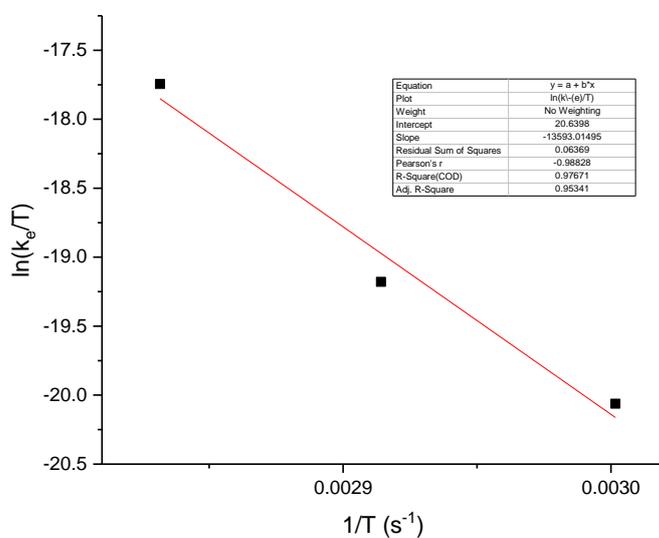
Extrapolated half-life of
racemization at 20°C: 11.5 y

4d



Equation	y = a + b*x		
	ln(100/ee)	ln(100/ee)	ln(100/ee)
Plot		No Weighting	
Weight			
Intercept	0.22285	0.28199	0.29059
Slope	1.38817E-5	3.21633E-6	1.29122E-6
Residual Sum of Squares	0.05757	2.68552E-4	0.00249
Pearson's r	0.99782	0.99987	0.96873
R-Square(COD)	0.99564	0.99975	0.93645
Adj. R-Square	0.9951	0.99971	0.92306

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	1.291E-06	3.216E-06	1.388E-05
k_{ent} (s ⁻¹)	6.456E-07	1.608E-06	6.941E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	121.33	122.45	121.82
$t_{1/2}(rac)_T$ (min)	8946.93	3591.81	832.21



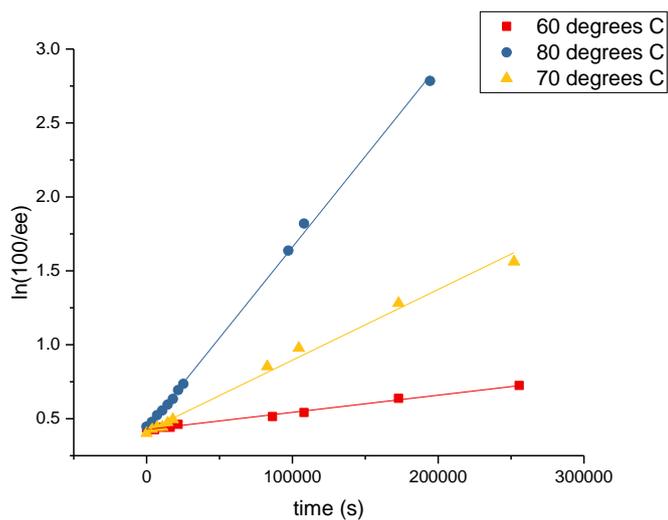
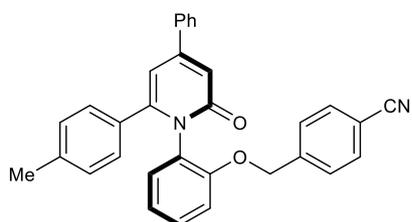
Equation	y = a + b*x
Plot	ln(k _e /T)
Weight	No Weighting
Intercept	20.6398
Slope	-13593.01495
Residual Sum of Squares	0.06369
Pearson's r	-0.98528
R-Square(COD)	0.97671
Adj. R-Square	0.95341

$$\Delta H_{ent}^\ddagger = 113.0 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -25.9 \text{ J K}^{-1} \text{ mol}^{-1}$$

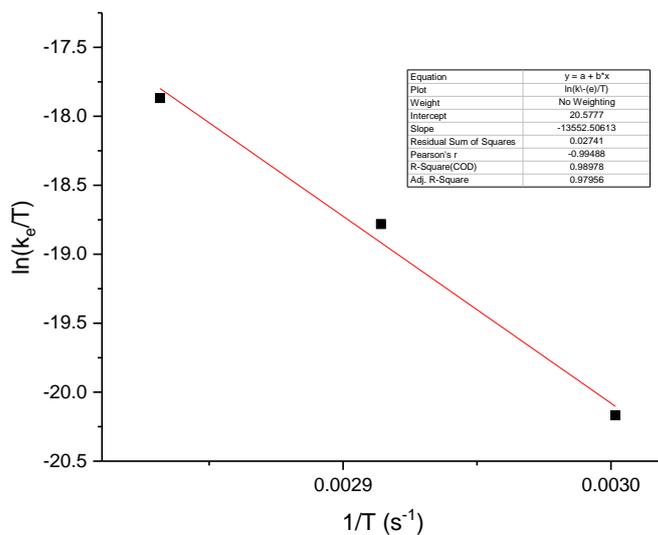
Extrapolated half-life of racemization at 20°C: 5.7 y

4e



Equation	y = a + b*x		
	ln(100/ee)	ln(100/ee)	ln(100/ee)
Plot		No Weighting	
Weight			
Intercept	0.42686	0.43267	0.41714
Slope	1.16124E-6	1.22767E-5	4.7849E-6
Residual Sum of Squares	6.6832E-4	0.00607	0.01167
Pearson's r	0.99617	0.99947	0.99816
R-Square(COD)	0.99235	0.99894	0.99238
Adj. R-Square	0.99126	0.99862	0.99142

	T (K)		
	333	343	353
k_{rac} (s^{-1})	1.161E-06	4.785E-06	1.228E-05
k_{ent} (s^{-1})	5.806E-07	2.392E-06	6.138E-06
ΔG_T^\ddagger (kJ mol $^{-1}$)	121.62	121.32	122.18
$t_{1/2}(rac)_T$ (min)	9948.38	2414.36	941.01



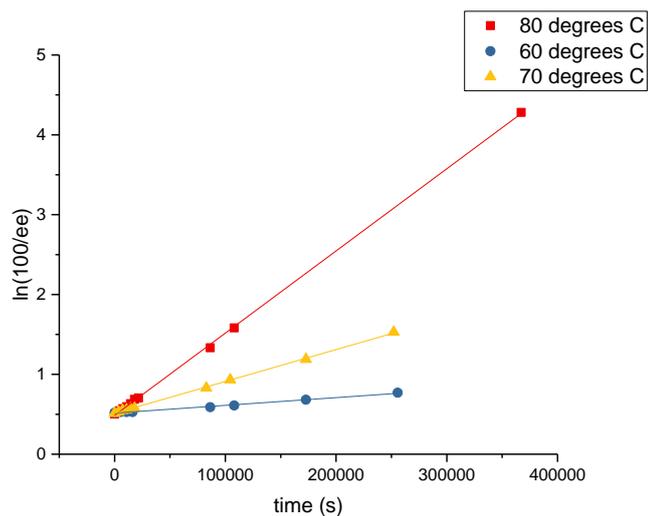
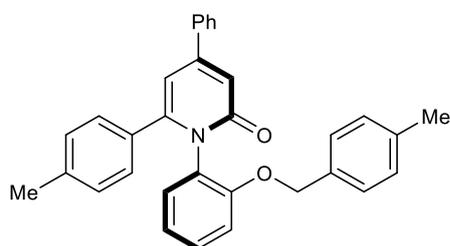
Equation	y = a + b*x	
	ln(k _e /T)	ln(k _e /T)
Plot		No Weighting
Weight		
Intercept	20.5777	
Slope	-13552.50613	
Residual Sum of Squares	0.02741	
Pearson's r	-0.99496	
R-Square(COD)	0.98978	
Adj. R-Square	0.97956	

$$\Delta H_{ent}^\ddagger = 112.7 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -26.5 \text{ J K}^{-1} \text{ mol}^{-1}$$

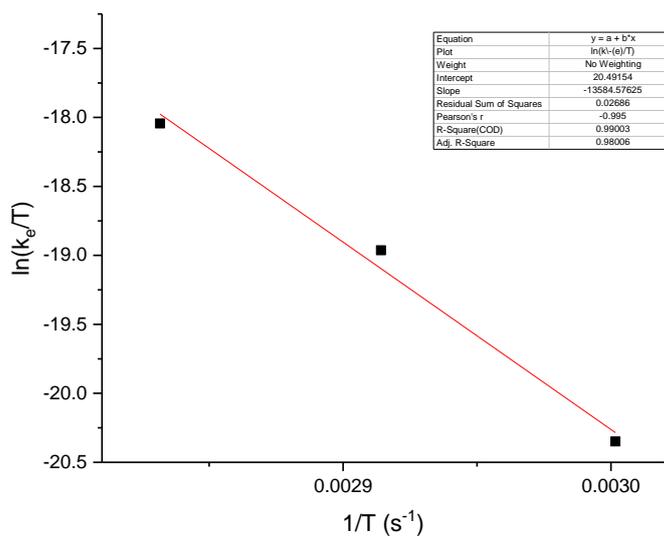
Extrapolated half-life of
racemization at 20°C: 5.3 y

4f



Equation	y = a + b*x		
Plot	ln(100/ee)	ln(100/ee)	ln(100/ee)
Weight	No Weighting		
Intercept	0.4873	0.51488	0.51182
Slope	1.02527E-5	9.6823E-7	3.99273E-6
Residual Sum of Squares	0.00328	3.03691E-4	6.12457E-4
Pearson's r	0.99986	0.99736	0.99972
R-Square(COD)	0.99973	0.99472	0.99943
Adj. R-Square	0.9997	0.99384	0.99836

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	9.682E-07	3.993E-06	1.029E-05
k_{ent} (s ⁻¹)	4.841E-07	1.996E-06	5.146E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	122.13	121.84	122.69
$t_{1/2}(rac)_T$ (min)	11931.48	2893.37	1122.39



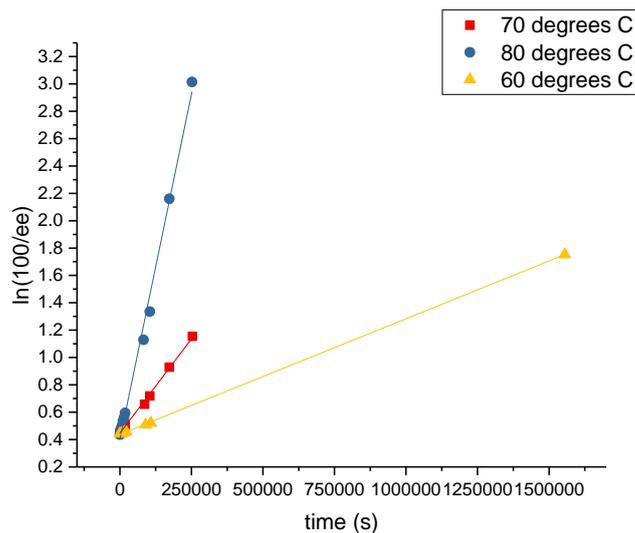
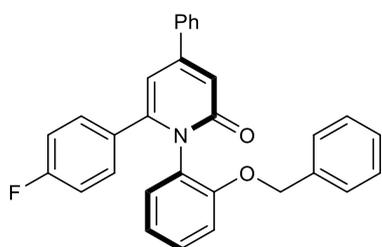
Equation	y = a + b*x
Plot	ln(k _e /T)
Weight	No Weighting
Intercept	20.49154
Slope	-13584.57625
Residual Sum of Squares	0.02686
Pearson's r	-0.995
R-Square(COD)	0.99003
Adj. R-Square	0.98006

$$\Delta H_{ent}^\ddagger = 112.9 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -27.2 \text{ J K}^{-1} \text{ mol}^{-1}$$

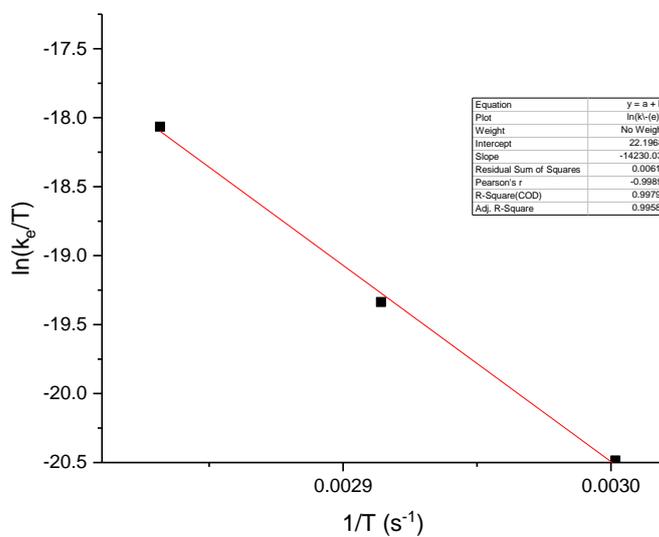
Extrapolated half-life of
racemization at 20°C: 6.5 y

4g



Equation	y = a + b*x		
Plot	ln(100/ee)	ln(100/ee)	ln(100/ee)
Weight	No Weighting		
Intercept	0.44743	0.40233	0.43808
Slope	2.74858E-6	1.00819E-5	8.44969E-7
Residual Sum of Squares	0.00133	0.03525	1.56816E-4
Pearson's r	0.99744	0.99744	0.99995
R-Square(COD)	0.99744	0.99489	0.99989
Adj. R-Square	0.99712	0.99425	0.99987

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	8.450E-07	2.749E-06	1.008E-05
k_{ent} (s ⁻¹)	4.225E-07	1.374E-06	5.041E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	122.50	122.90	122.70
$t_{1/2}(rac)_T$ (min)	13672.04	4203.06	1145.86



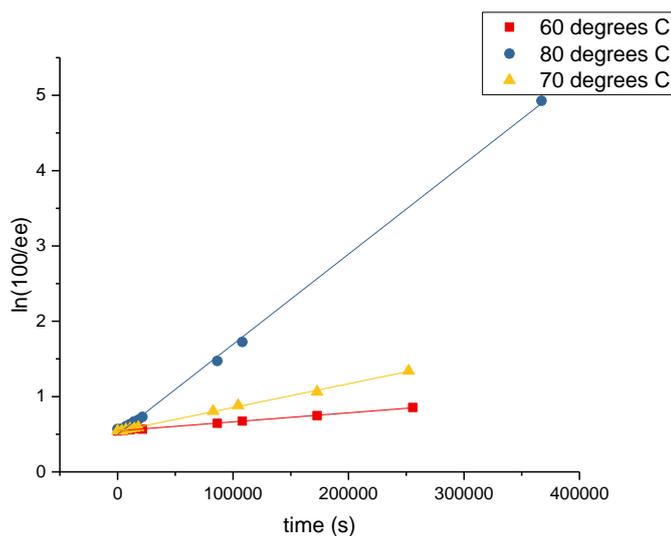
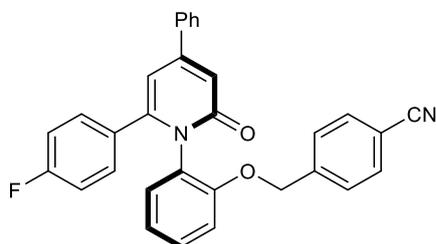
Equation	y = a + b*x
Plot	ln(k _e /T)
Weight	No Weighting
Intercept	22.19687
Slope	-14230.03448
Residual Sum of Squares	0.00614
Pearson's r	-0.99895
R-Square(COD)	0.99791
Adj. R-Square	0.99581

$$\Delta H_{ent}^\ddagger = 118.3 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -13.0 \text{ J K}^{-1} \text{ mol}^{-1}$$

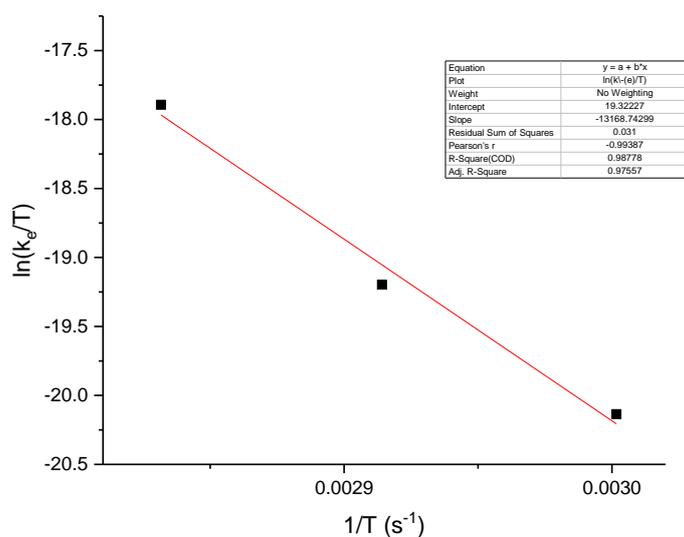
Extrapolated half-life of
racemization at 20°C:
10.6 y

4h



Equation	y = a + b*x		
Plot	ln(100/ee)	ln(100/ee)	ln(100/ee)
Weight	No Weighting		
Intercept	0.54455	0.49695	0.5388
Slope	1.19674E-6	1.19706E-5	3.15503E-6
Residual Sum of Squares	1.83607E-4	0.01693	7.72368E-4
Pearson's r	0.99901	0.99948	0.99943
R-Square(COD)	0.99802	0.99897	0.99885
Adj. R-Square	0.99773	0.99884	0.99871

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	1.199E-06	3.155E-06	1.197E-05
k_{ent} (s ⁻¹)	5.994E-07	1.578E-06	5.985E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	121.53	122.51	122.25
$t_{1/2}(rac)_T$ (min)	9637.16	3661.60	965.07

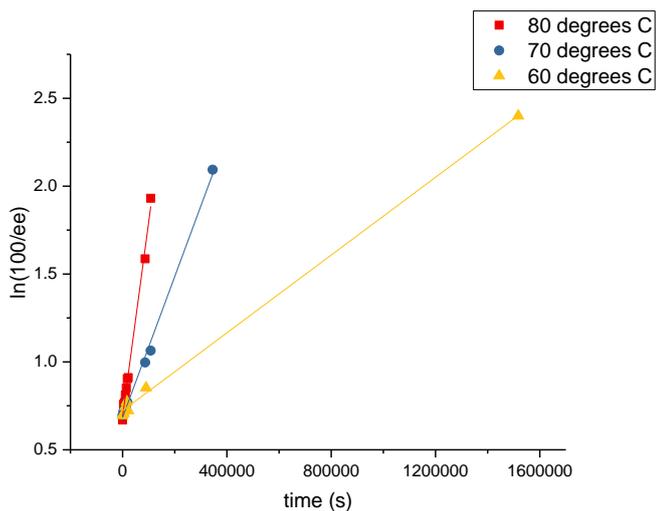
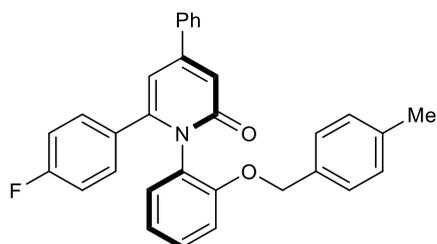


Equation	y = a + b*x
Plot	ln(k_e/T)
Weight	No Weighting
Intercept	19.32227
Slope	-13168.74299
Residual Sum of Squares	0.031
Pearson's r	-0.99387
R-Square(COD)	0.98778
Adj. R-Square	0.97557

$$\Delta H_{ent}^\ddagger = 109.5 \text{ kJ mol}^{-1}$$

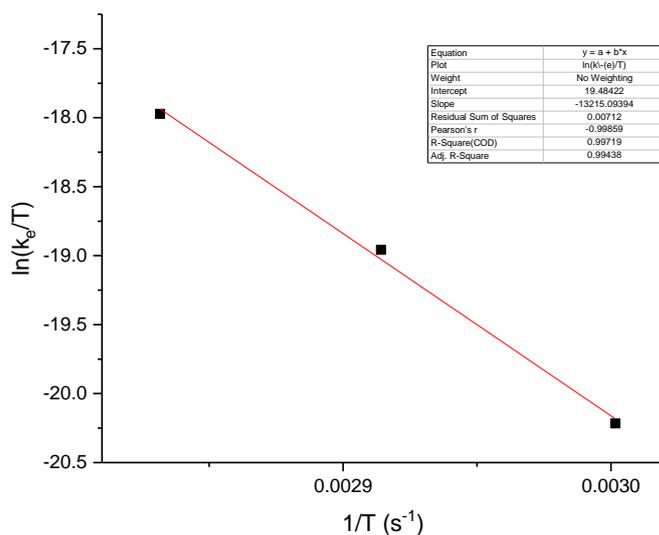
$$\Delta S_{ent}^\ddagger = -36.9 \text{ J K}^{-1} \text{ mol}^{-1}$$

Extrapolated half-life of
racemization at 20°C: 5.0 y



Equation	y = a + b*x		
	ln(100/ee)	ln(100/ee)	ln(100/ee)
Plot		No Weighting	
Weight			
Intercept	0.68945	0.68403	0.7229
Slope	1.10648E-5	4.00925E-6	1.10622E-6
Residual Sum of Squares	0.00747	0.00574	0.004
Pearson's r	0.99751	0.99822	0.99915
R-Square(COD)	0.99503	0.99644	0.99653
Adj. R-Square	0.99432	0.99553	0.99796

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	1.106E-06	4.009E-06	1.106E-05
k_{ent} (s ⁻¹)	5.531E-07	2.005E-06	5.532E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	121.76	121.83	122.48
$t_{1/2}(rac)_T$ (min)	10443.18	2881.45	1044.07



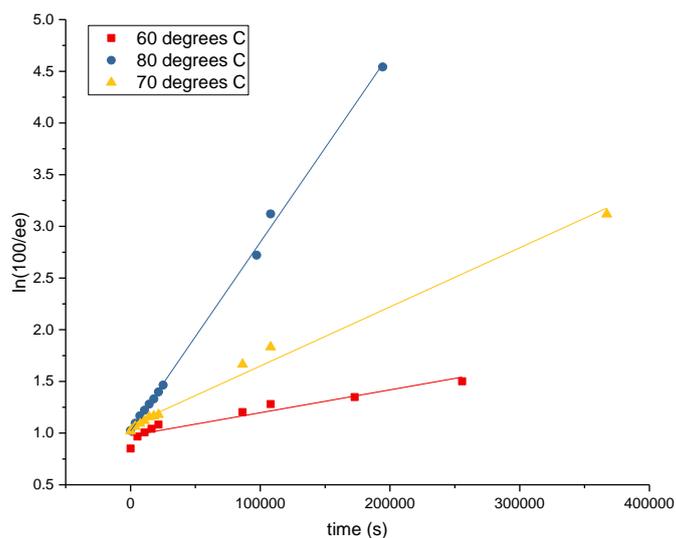
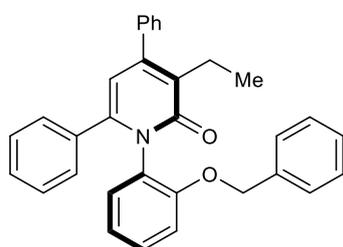
Equation	y = a + b*x	
	ln(k _e /T)	ln(k _e /T)
Plot		
Weight		No Weighting
Intercept	19.48422	
Slope	-13215.09394	
Residual Sum of Squares	0.00712	
Pearson's r	-0.99859	
R-Square(COD)	0.99719	
Adj. R-Square	0.99438	

$$\Delta H_{ent}^\ddagger = 109.9 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -35.6 \text{ J K}^{-1} \text{ mol}^{-1}$$

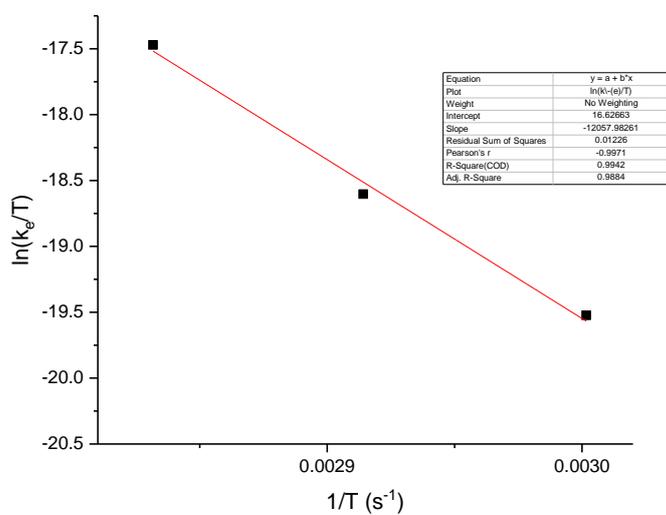
Extrapolated half-life of
racemization at 20°C: 5.0 y

4j



Equation	y = a + b*x		
Plot	ln(100/ee)	ln(100/ee)	ln(100/ee)
Weight	No Weighting	No Weighting	No Weighting
Intercept	0.97592	1.00445	1.07696
Slope	2.21257E-6	1.82572E-5	5.71937E-6
Residual Sum of Squares	0.02814	0.00291	0.03735
Pearson's r	0.9581	0.9926	0.9505
R-Square(COD)	0.91735	0.99811	0.90013
Adj. R-Square	0.90622	0.9973	0.8889

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	2.213E-06	5.719E-06	1.826E-05
k_{ent} (s ⁻¹)	1.106E-06	2.860E-06	9.129E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	119.84	120.81	121.01
$t_{1/2}(rac)_T$ (min)	5221.28	2019.92	632.76



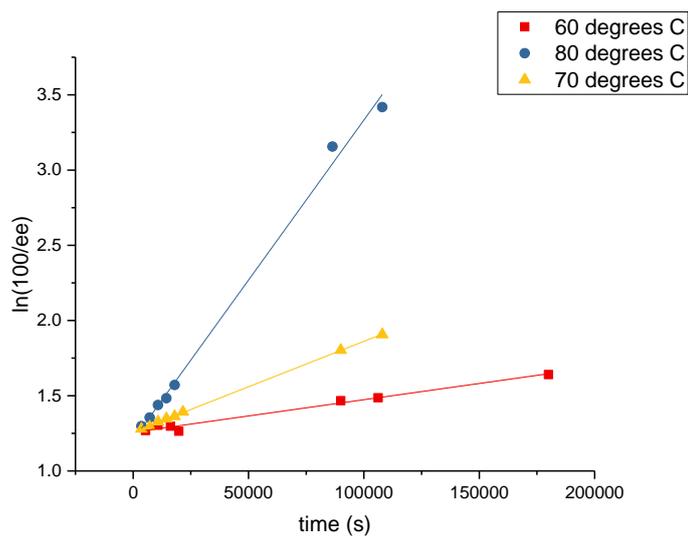
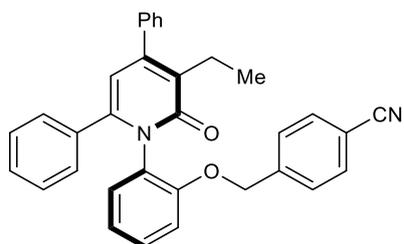
Equation	y = a + b*x
Plot	ln(k _e /T)
Weight	No Weighting
Intercept	16.62653
Slope	-12057.98261
Residual Sum of Squares	0.01226
Pearson's r	-0.9971
R-Square(COD)	0.9942
Adj. R-Square	0.9884

$$\Delta H_{ent}^\ddagger = 100.3 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -59.3 \text{ J K}^{-1} \text{ mol}^{-1}$$

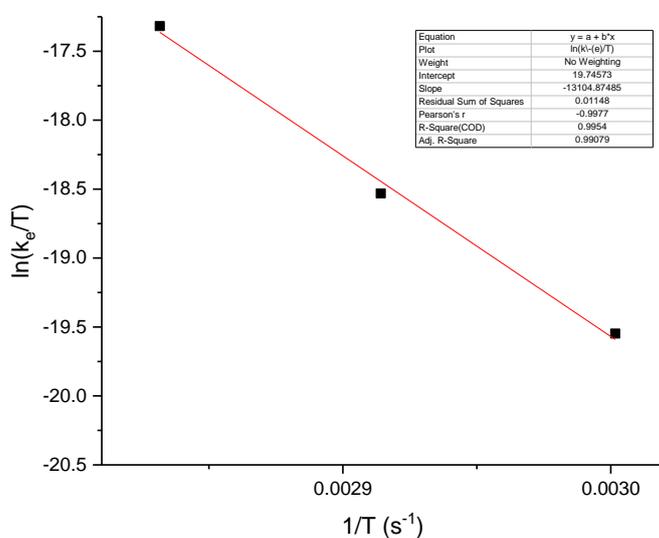
Extrapolated half-life of racemization at 20°C: 1.7 y

4k



Equation Plot	y = a + b*x		
	ln(100/ee)		ln(100/ee)
Weight	No Weighting		
Intercept	1.25769	1.20502	1.25774
Slope	2.15851E-6	2.12756E-5	6.03729E-6
Residual Sum of Squares	0.00214	0.02115	1.27596E-4
Pearson's r	0.99141	0.99789	0.99885
R-Square(COD)	0.9829	0.99678	0.9997
Adj. R-Square	0.97948	0.99494	0.99865

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	2.159E-06	6.143E-06	2.128E-05
k_{ent} (s ⁻¹)	1.079E-06	3.071E-06	1.064E-05
ΔG_T^\ddagger (kJ mol ⁻¹)	119.91	120.61	120.56
$t_{1/2}(rac)_T$ (min)	5352.05	1880.65	543.01

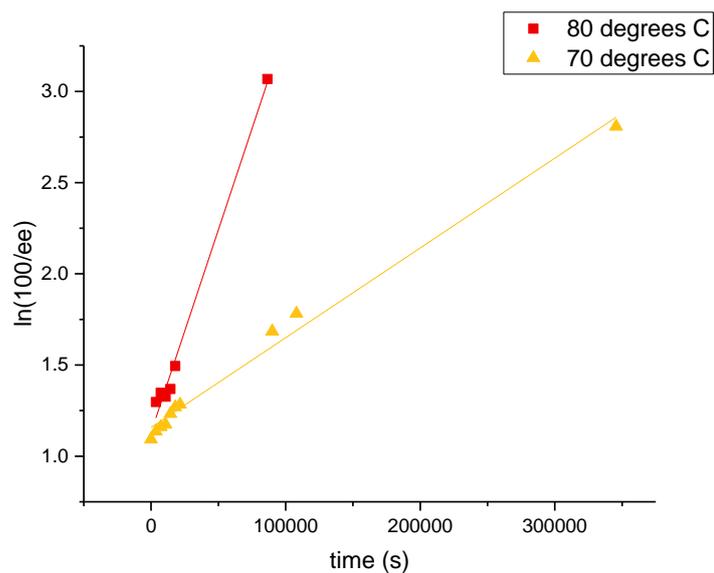
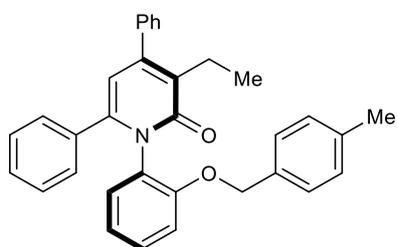


Equation Plot	y = a + b*x	
	ln(k _e /T)	1/T
Weight	No Weighting	
Intercept	19.74573	
Slope	-13104.87485	
Residual Sum of Squares	0.01148	
Pearson's r	-0.9977	
R-Square(COD)	0.9954	
Adj. R-Square	0.99079	

$$\Delta H_{ent}^\ddagger = 109.0 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -33.4 \text{ J K}^{-1} \text{ mol}^{-1}$$

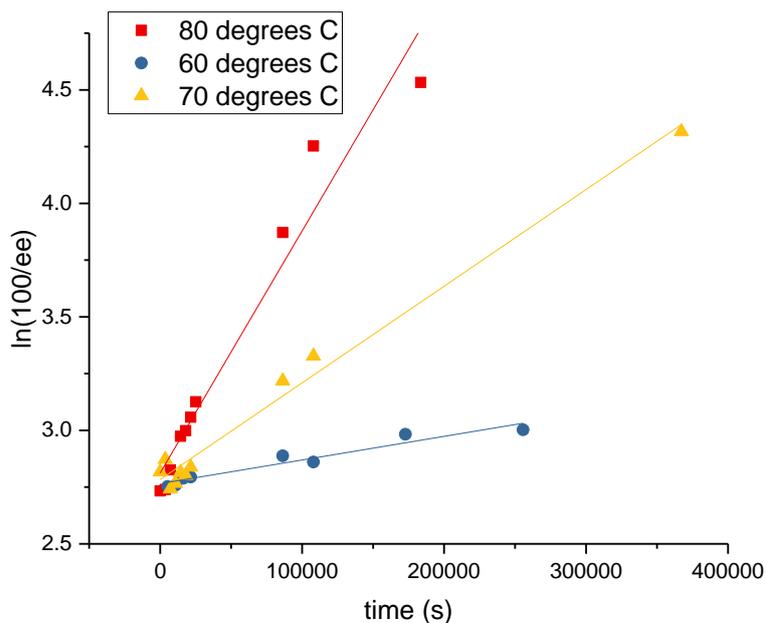
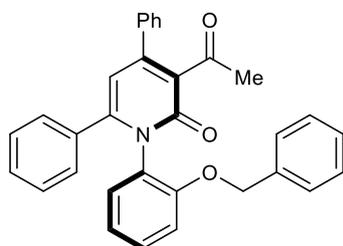
Extrapolated half-life of racemization at 20°C: 2.6 y



Equation	y = a + b*x	
Plot	ln(100/ee)	ln(100/ee)
Weight	No Weighting	
Intercept	1.13025	1.15755
Slope	2.22156E-5	4.92286E-6
Residual Sum of Squares	0.02095	0.02741
Pearson's r	0.99569	0.9945
R-Square(COD)	0.9914	0.98904
Adj. R-Square	0.98325	0.98767

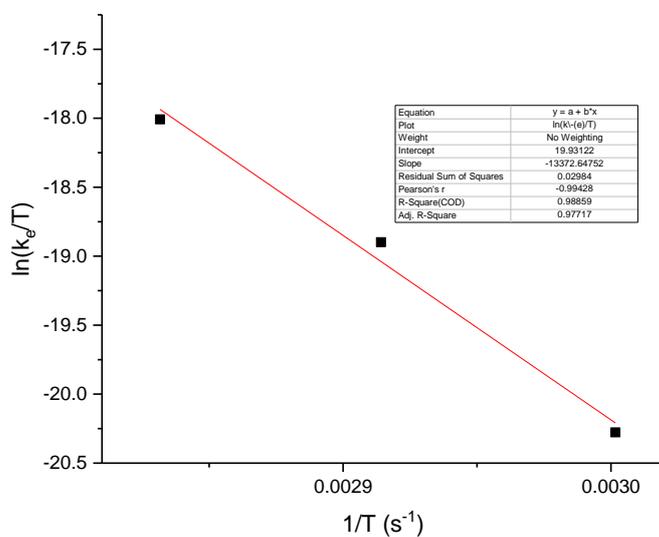
	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	<i>not measured</i>	4.923E-06	2.222E-05
k_{ent} (s ⁻¹)	-	2.461E-06	1.111E-05
ΔG_T^\ddagger (kJ mol ⁻¹)	-	121.24	120.44
$t_{1/2}(rac)_T$ (min)	-	2346.70	520.02

4m



Equation	y = a + b*x		
Plot	ln(100/ee)	ln(100/ee)	ln(100/ee)
Weight	No Weighting		
Intercept	2.81188	2.76545	2.78334
Slope	1.06692E-5	1.04072E-6	4.25857E-6
Residual Sum of Squares	0.18368	0.00434	0.03506
Pearson's r	0.97616	0.96709	0.99192
R-Square(COD)	0.95288	0.93526	0.98391
Adj. R-Square	0.94699	0.92447	0.98212

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	1.041E-06	4.259E-06	1.067E-05
k_{ent} (s ⁻¹)	5.204E-07	2.129E-06	5.335E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	121.93	121.65	122.59
$t_{1/2}(rac)_T$ (min)	11100.44	2712.75	1082.79



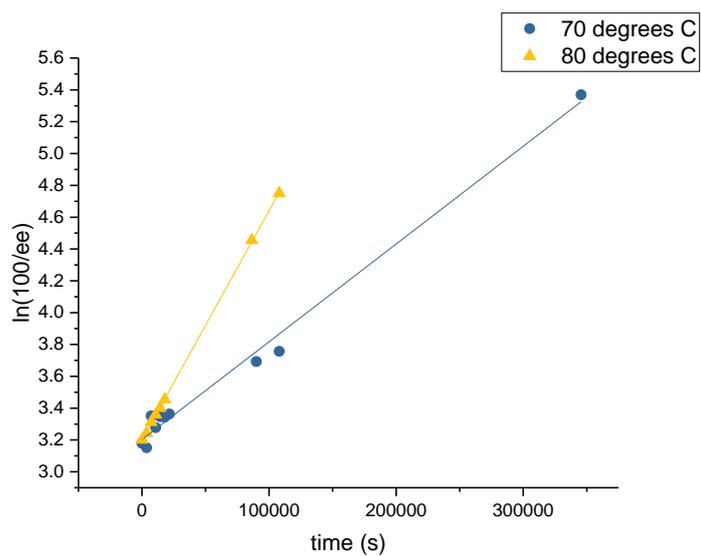
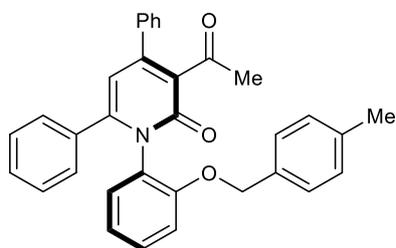
Equation	y = a + b*x
Plot	ln(k _e /T)
Weight	No Weighting
Intercept	19.53122
Slope	-13372.64752
Residual Sum of Squares	0.02984
Pearson's r	-0.99428
R-Square(COD)	0.98859
Adj. R-Square	0.97717

$$\Delta H_{ent}^\ddagger = 111.2 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -31.8 \text{ J K}^{-1} \text{ mol}^{-1}$$

Extrapolated half-life of racemization at 20°C: 5.5 y

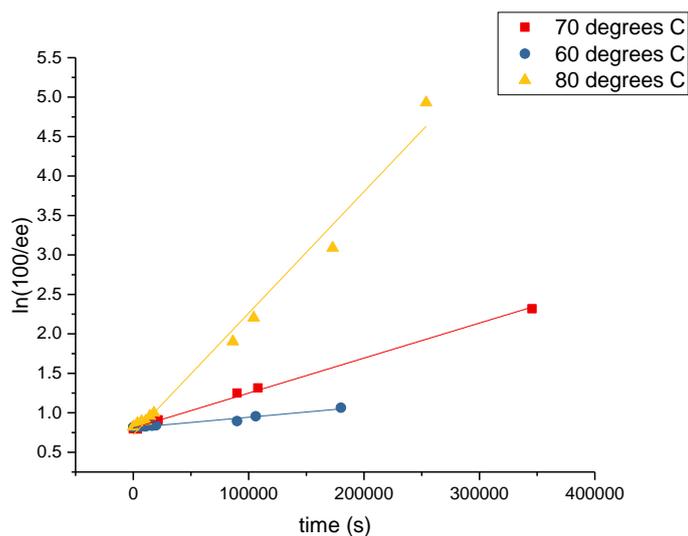
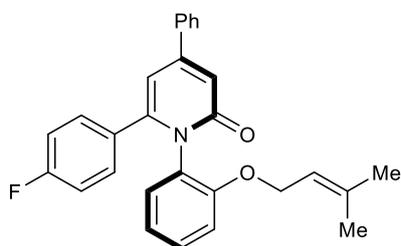
4o



Equation	ln(100/ee)	ln(100/ee)
Plot		
Weight		
Intercept	3.20285	3.19908
Slope	6.14279E-6	1.44216E-5
Residual Sum of Squares	0.03845	3.71975E-4
Pearson's r	0.99492	0.99993
R-Square(COD)	0.98986	0.99985
Adj. R-Square	0.98859	0.99983

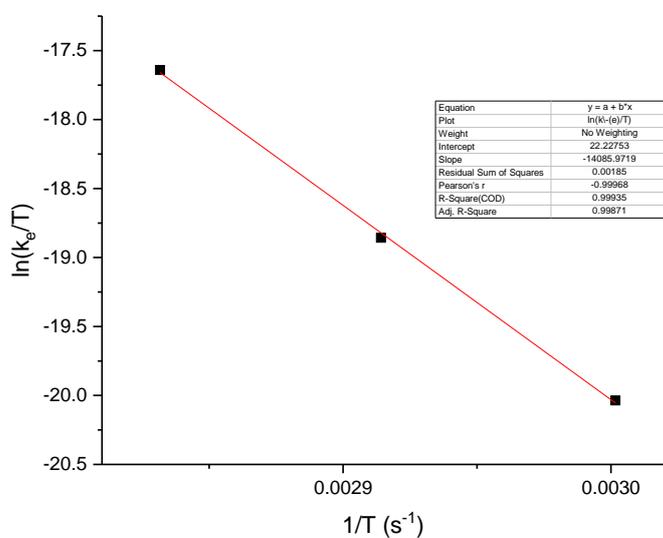
	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	<i>not measured</i>	6.143E-06	1.442E-05
k_{ent} (s ⁻¹)	-	3.071E-06	7.211E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	-	120.61	121.70
$t_{1/2}(rac)_T$ (min)	-	1880.65	801.05

4p



Equation	y = a + b*x		
	ln(100/ee)	ln(100/ee)	ln(100/ee)
Plot		No Weighting	
Weight			
Intercept	0.80576	0.81113	0.72364
Slope	4.43721E-6	1.32408E-6	1.54021E-5
Residual Sum of Squares	0.00534	0.00159	0.23766
Pearson's r	0.99867	0.98503	0.99274
R-Square(COD)	0.99735	0.97029	0.98554
Adj. R-Square	0.99702	0.96534	0.98373

	T (K)		
	333	343	353
k_{rac} (s ⁻¹)	1.324E-06	4.437E-06	1.540E-05
k_{ent} (s ⁻¹)	6.620E-07	2.219E-06	7.701E-06
ΔG_T^\ddagger (kJ mol ⁻¹)	121.26	121.54	121.51
$t_{1/2}(rac)_T$ (min)	8724.89	2603.54	750.06



Equation	y = a + b*x	
	ln(k_e/T)	ln(k_e/T)
Plot		
Weight	No Weighting	
Intercept	22.22753	
Slope	-14085.9719	
Residual Sum of Squares	0.00185	
Pearson's r	-0.99968	
R-Square(COD)	0.99935	
Adj. R-Square	0.99871	

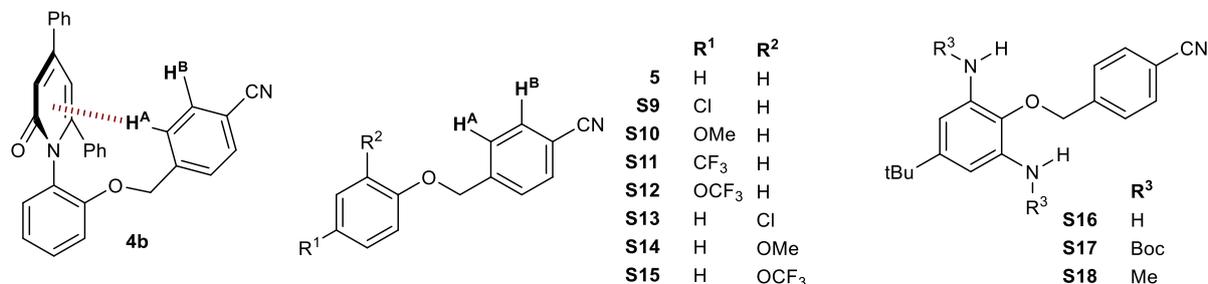
$$\Delta H_{ent}^\ddagger = 117.1 \text{ kJ mol}^{-1}$$

$$\Delta S_{ent}^\ddagger = -12.7 \text{ J K}^{-1} \text{ mol}^{-1}$$

5 NMR Analysis of CH \cdots π Interaction

Singe crystal data and quantum chemical calculations both indicate the presence of a CH \cdots π interaction present in the ground state of the benzylated products **4**. To demonstrate that this effect is present in solution as well as solid phase, the chemical shift values of **H^A** and **H^B** in 4-cyanobenzyl compound **4b** were compared to literature values for structurally related compounds (see manuscript, figure 2B). In addition to the comparison with *O*-(*para*-cyanobenzyl)phenol provided in the manuscript, below is given comparison with a large number of literature compounds.

Table S2. Comparison of ¹H chemical shift values of **4b** with structurally similar compounds previously reported. All ¹H signals are reported in CDCl₃.



Compound	H ^A (ppm)	H ^B (ppm)	Reference
4b	7.42	7.60	
5	7.56	7.69	19
S9	7.53	7.68	20
S10	7.54	7.67	20
S11	7.54	7.69	20
S12	7.56	7.7	20
S13	7.60	7.68	20
S14	7.56	7.66	20
S15	7.55	7.69	20
S16	7.58	7.68	21
S17	7.54	-*	21
S18	7.55	7.69	21

* Co-incident with another ¹H signal.

6 Density Functional Theory Calculations

6.1 Methods

All optimisation, frequency, and single-point calculations were performed with *Gaussian 16*, rev C.01.²² Structures were generated with CYLview.²³ All non-covalent interaction surfaces were calculated with and displayed using Jmol.²⁴ The B3LYP functional was used for all geometry optimisations with the 6-31G(d,p) basis set on all atoms. Grimme's DFT-D3²⁵ correction was included in the optimisation procedure as well as single point calculations. All optimised structures were confirmed as minima by the absence of imaginary frequencies while transition states were characterised by the presence of a single imaginary frequency. All transition states were further analysed via intrinsic reaction coordinate calculations. Further refinements to the electronic energies were made through single point calculations on the optimised geometries using the B3LYP functional with the 6-311+G(d,p) basis set for all atoms. Grimme's DFT-D3 correction and further corrections for bulk solvation through a polarisable continuum model (CPCM) were also incorporated. Free energies were determined from thermochemical corrections of the geometries applied to electronic energies.

6.2 Summary of Results

Three starting materials, and their corresponding products, were chosen for study each with a different (R^3) substituent at the pyridine 3-position: **3a**, **3d**, and **3e** leading to **4a**, **4j**, and **4m** respectively (addition of a benzyl group). Barriers were calculated for starting material, **3**, the deprotonated intermediate, **3(int)**, and the reaction products, **4**. Calculated rotational barriers are summarised in Table S3, showing good agreement with experimentally determined barriers. Computational barriers for **3** and **3(int)** were calculated in acetonitrile but for **4** were calculated in DMSO to reflect experimental procedures (see section 4 above).

Table S3: Calculated vs. experimental barriers for **3**, **3int**, and **4**. Calculated at B3LYP-D3-CPCM/6-311+G(d,p)//B3LYP-D3/6-31G(d,p).

	dG_{DFT} (kJ mol ⁻¹)	dG_{Exp} (kJ mol ⁻¹)
3a	95.4	96.6
3d	97.1	98.7
3e	97.5	102.9
3(int)a	123.0	-
3(int)d	125.9	-
3(int)e	123.4	-
4a	123.0	121.3
4j	124.7	121.0
4m	121.3	122.6

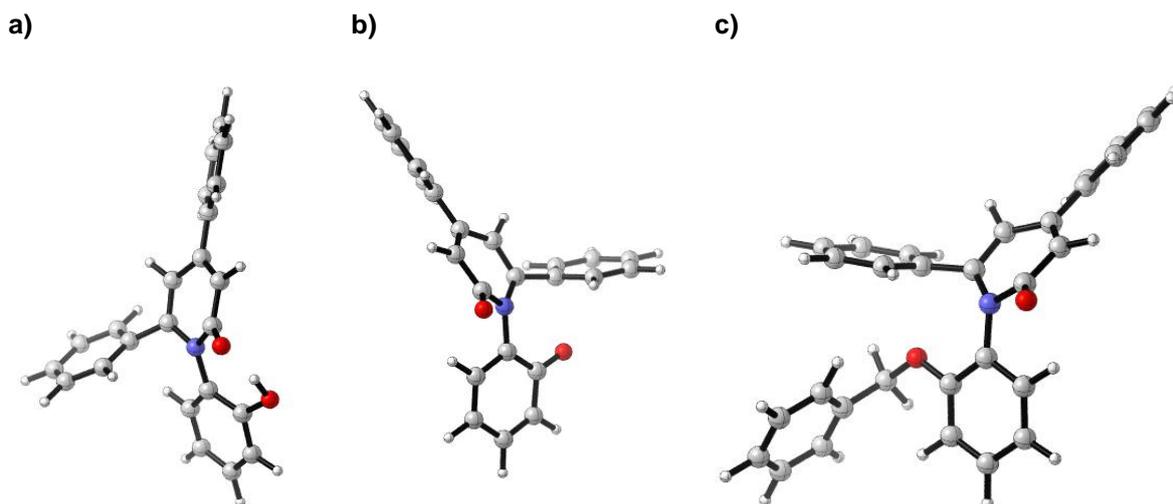


Figure S1. Representative enantiomerisation transition state geometries for: a) **3a**; b) **3(int)a**; c) **4a**. Calculated at B3LYP-D3-CPCM_{DMSO}/6-311+G(d,p)//B3LYP-D3/6-31G(d,p).

For starting material **3a**, the lowest barrier transition state was found with the phenol and carbonyl groups *syn* to allow for the favourable OH...O hydrogen bond, Figure S1a. The alternative transition state, with phenol and carbonyl groups *anti* was found to be *ca.* 10 kcal mol⁻¹ higher in energy.

A non-covalent interaction descriptor based on electron density, developed by the Johnson and Contreras-Garcia groups,²⁶ allows a qualitative visual analysis of NCIs and is shown here to be a useful tool. The coloration of the NCI surface allows identification and characterization of attractive and repulsive interactions; a strong attractive interaction is in blue, van der Waals and dispersion interactions are in green, and destabilizing steric interactions are in red. In the enantiomerisation transition state of **3a**, the strong hydrogen bond between the phenol and 2-pyridone oxygens is observed as a deep blue surface, Figure S2a. The stabilising OH...O hydrogen bond is also observed in the minimum, S2b. Rotating the phenol group out of reach of an OH...O hydrogen bond and reoptimizing the structure results in a large 6 kJ mol⁻¹ energy penalty

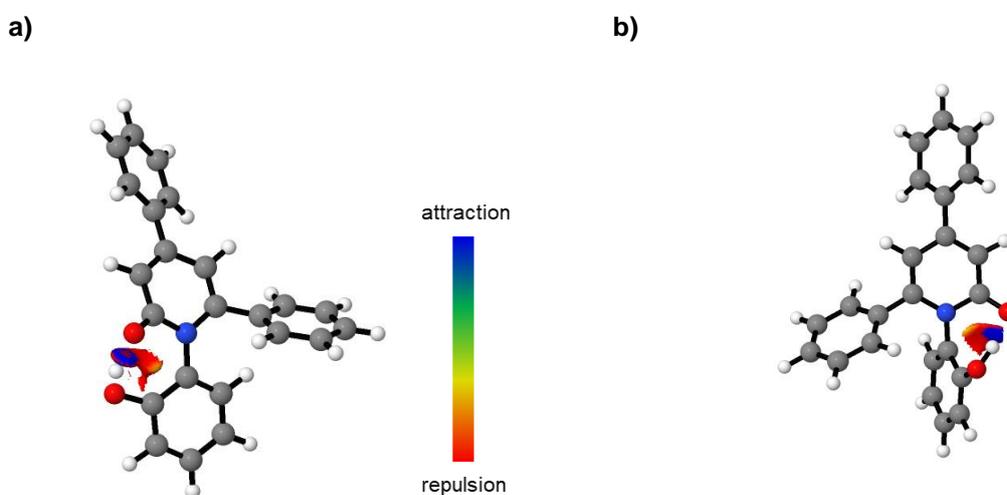


Figure S2. Non-covalent interaction surface of the starting material enantiomerisation transition state of **3a** showing a strong hydrogen bond in deep blue. The colour spectrum ranges from blue (strongly attractive) to green (weakly attractive) to yellow (mildly repulsive) to red (strongly repulsive), geometries optimised at B3LYP-D3/6-31G(d,p).

Without the proton to participate in the hydrogen bond, the enantiomerisation transition state barrier for **3(int)a** is significantly higher, by 28 kJ mol⁻¹ (Table S3) with the lowest energy transition state found with the phenoxide and carbonyl groups *anti*, Figure S1b. The *syn* transition state could not be found.

Addition of the benzyl group raises the transition state barrier for **4a** similarly to that of the deprotonated intermediate, Table S3. The lowest barrier to enantiomerisation was found with the benzyl ether and carbonyl group *anti*, Figure S1c.

The crystal structures and NMR shifts of the products, **4**, showed a CH- π non-covalent interaction between the *ortho*-CH of the benzyl group and the π -system of the 2-pyridone which was replicated in the solution phase *via* DFT. A non-covalent interaction surface shows this CH- π interaction as an area of light blue, Figure S3. The non-planar geometry around the 2-pyridone nitrogen, and tilt of this group, is also replicated.

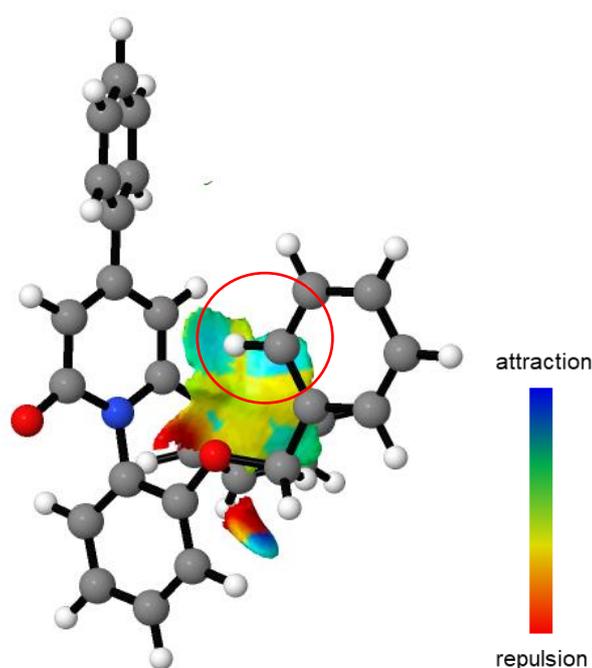


Figure S3. Non-covalent interaction surface of the minima of **4a** showing a stabilising CH- π interaction in light blue (circled). The colour spectrum ranges from blue (strongly attractive) to green (weakly attractive) to yellow (mildly repulsive) to red (strongly repulsive), geometries optimised at B3LYP-D3/6-31G(d,p).

In all structures examined, the minima that included the CH- π interaction were lower in energy than those without, Table S4, in which the benzyl group lies face on to the 2-pyridone.

Table S4. Relative energies of product minima with and without CH- π interaction as identified in the crystal structures of products **4**. B3LYP-D3-CPCM_{DMSO}/6-311+G(d,p)//B3LYP-D3/6-31G(d,p).

	With CH- π interaction (kJ mol ⁻¹)	Without CH- π interaction (kJ mol ⁻¹)
4a	0.0	+7.1
4j	0.0	+4.2
4m	0.0	+2.1

6.3 Cartesian Coordinates

3a – Minimum

C	4.67967	-1.22837	1.09847
C	4.20245	-1.88348	-0.03237
C	2.848	-1.81327	-0.38189
C	1.98287	-1.04515	0.41937
C	2.46119	-0.42349	1.5761
C	3.8067	-0.50727	1.91848
H	5.73348	-1.29568	1.35219
H	4.85516	-2.47125	-0.66876
H	1.76992	0.14693	2.18779
H	4.16939	-0.01467	2.81448
C	0.10808	0.46504	-0.05847
C	-0.25331	-1.95559	-0.06958
C	-1.24546	0.68217	-0.05975
C	-1.66084	-1.68486	-0.01702
C	-2.16557	-0.4066	0.00469
H	-1.60486	1.69103	-0.21407
H	-2.29805	-2.55978	0.01945
C	1.05447	1.58649	-0.27623
C	2.06283	1.48731	-1.24902
C	0.90711	2.78289	0.43791
C	2.90656	2.56638	-1.49383
H	2.17471	0.56615	-1.81149
C	1.75678	3.86119	0.19149
H	0.13688	2.85608	1.19988
C	2.75916	3.75489	-0.77306
H	3.68079	2.4801	-2.25005
H	1.63943	4.78	0.75821
H	3.42347	4.59257	-0.963
C	-3.62346	-0.14453	0.05103
C	-4.12475	0.96748	0.74808
C	-4.53404	-0.99869	-0.59288
C	-5.49535	1.2118	0.80868
H	-3.4374	1.62711	1.26894
C	-5.90381	-0.75126	-0.5354
H	-4.16256	-1.84349	-1.16414
C	-6.38985	0.35372	0.16658
H	-5.86507	2.07026	1.36153
H	-6.59216	-1.41748	-1.04679
H	-7.45764	0.54607	0.21026
O	2.41968	-2.44818	-1.49876
H	1.55557	-2.86152	-1.2472
O	0.23449	-3.09478	-0.17486
N	0.60131	-0.82991	0.04191

3a – Transition State

C	4.65174	-1.66841	1.30736
C	3.87256	-2.6547	0.73423
C	2.58312	-2.40997	0.22045
C	2.0861	-1.06694	0.1992
C	2.88581	-0.09917	0.84471
C	4.13503	-0.37494	1.38026
H	5.63283	-1.90533	1.7074
H	4.20484	-3.68597	0.68469
H	2.52771	0.90728	0.96018
H	4.69015	0.42499	1.85929
C	0.19976	0.58855	-0.06205
C	-0.08933	-1.60706	-0.97724
C	-1.15512	0.72688	0.06096
C	-1.49999	-1.48846	-0.72156
C	-2.0499	-0.3653	-0.16458
H	-1.5552	1.7267	0.17248
H	-2.08959	-2.34446	-1.02605
C	1.00195	1.82987	-0.22434
C	1.85414	1.96211	-1.3337
C	0.83638	2.91818	0.64416
C	2.52804	3.15816	-1.56121
H	1.98582	1.11984	-2.00529
C	1.51588	4.11407	0.41299
H	0.19283	2.8132	1.51226
C	2.36342	4.23781	-0.68875
H	3.1825	3.24928	-2.42279
H	1.38734	4.94624	1.09859
H	2.89451	5.16792	-0.86641
C	-3.497	-0.2195	0.09906
C	-3.94067	0.57313	1.17126
C	-4.45541	-0.86455	-0.70072
C	-5.30061	0.70636	1.4436
H	-3.21205	1.0643	1.80926
C	-5.81472	-0.72769	-0.42969
H	-4.13365	-1.45218	-1.55461
C	-6.24254	0.05656	0.64395
H	-5.62464	1.3141	2.28316
H	-6.54194	-1.22666	-1.06319
H	-7.30266	0.1631	0.85332
O	1.92146	-3.52738	-0.12812
H	1.30071	-3.31629	-0.87729
O	0.36877	-2.48662	-1.72771
N	0.76098	-0.68629	-0.29548

3(int)a – Intermediate Minimum

C	-4.76149	-1.7555	-0.20808
C	-4.13721	-1.52897	1.00783
C	-2.73795	-1.19417	1.11914
C	-2.06786	-1.12472	-0.16162
C	-2.69951	-1.36702	-1.37467
C	-4.06042	-1.67239	-1.42364
H	-5.8247	-1.99918	-0.22209
H	-4.69168	-1.58883	1.94187
H	-2.11034	-1.31196	-2.28849
H	-4.55598	-1.85345	-2.3733
C	-0.14255	0.40845	-0.10973
C	0.21343	-2.01474	-0.13541
C	1.21505	0.64246	-0.0718
C	1.63136	-1.72312	-0.06639
C	2.1306	-0.44598	-0.04388
H	1.56543	1.66634	-0.07255
H	2.2762	-2.59256	-0.00648
C	-1.06254	1.58147	-0.14651
C	-1.99519	1.81709	0.87507
C	-0.9239	2.51563	-1.18252
C	-2.77678	2.97079	0.84098
H	-2.08974	1.0839	1.6756
C	-1.71259	3.66716	-1.21115
H	-0.20154	2.32638	-1.97153
C	-2.64198	3.89758	-0.19665
H	-3.50094	3.1443	1.63272
H	-1.60198	4.37823	-2.02615
H	-3.25946	4.79257	-0.21521
C	3.59206	-0.19001	0.02249
C	4.10487	0.87202	0.78647
C	4.50173	-1.00183	-0.6758
C	5.47711	1.10931	0.8552
H	3.41838	1.49771	1.34833
C	5.87376	-0.76529	-0.60784
H	4.11938	-1.81166	-1.28935
C	6.36921	0.29222	0.15803
H	5.85069	1.93117	1.46053
H	6.55746	-1.40371	-1.16144
H	7.43865	0.47818	0.2098
O	-2.15822	-0.92555	2.21609
O	-0.24563	-3.15129	-0.19936
N	-0.64202	-0.86649	-0.15332

C	4.40566	-2.60492	-1.11183
C	4.3608	-1.23712	-0.96767
C	3.20769	-0.51813	-0.46745
C	2.09072	-1.35329	-0.06097
C	2.15884	-2.74566	-0.253
C	3.28508	-3.37935	-0.76125
H	5.30344	-3.08492	-1.50171
H	5.2013	-0.60612	-1.24574
H	1.3135	-3.36451	0.01428
H	3.28491	-4.4586	-0.88776
C	0.28434	0.44422	0.06136
C	-0.05454	-1.70334	1.13391
C	-1.08074	0.5821	-0.08829
C	-1.46184	-1.57692	0.83627
C	-1.9846	-0.47428	0.20802
H	-1.46735	1.57554	-0.27695
H	-2.07342	-2.40946	1.16516
C	1.03084	1.73423	0.11589
C	0.85374	2.70202	-0.88014
C	1.65994	2.10956	1.31728
C	1.33834	3.99934	-0.70613
H	0.35891	2.42071	-1.80502
C	2.1272	3.40445	1.49351
H	1.78699	1.36396	2.09494
C	1.97489	4.35986	0.48055
H	1.21429	4.72977	-1.5025
H	2.62063	3.67387	2.42435
H	2.34789	5.37153	0.61951
C	-3.42925	-0.32957	-0.07942
C	-3.86309	0.40832	-1.19567
C	-4.4084	-0.91899	0.74027
C	-5.21945	0.54494	-1.4853
H	-3.12089	0.8575	-1.84845
C	-5.76457	-0.78514	0.44957
H	-4.09743	-1.46571	1.62489
C	-6.17912	-0.05245	-0.6652
H	-5.52767	1.11454	-2.35831
H	-6.50166	-1.246	1.10231
H	-7.23698	0.05395	-0.88997
O	3.21713	0.74528	-0.42418
O	0.38841	-2.56664	1.89163
N	0.82204	-0.80207	0.40649

4a – Product Minimum**3(int)a – Intermediate Transition State**

O	-0.21381	-2.69452	-2.53727
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H	6.24876	0.38631	-1.91419
H	7.07576	1.66074	0.05408
O	-2.43766	-2.2859	-1.71724
H	-1.55415	-2.65971	-1.47502
O	-0.24267	-2.85595	-0.37154
N	-0.75676	-0.64337	-0.05103
C	2.51968	-2.53797	-0.17108
H	2.18852	-3.25199	-0.9316
H	3.53416	-2.22613	-0.42335
C	2.52269	-3.23814	1.20058
H	2.8614	-2.55322	1.98492
H	1.51784	-3.58763	1.45061
H	3.19374	-4.10293	1.18999

3d – Transition State

C	4.61507	-1.81562	-1.42665
C	3.77671	-2.72869	-0.81469
C	2.53661	-2.36942	-0.25344
C	2.15645	-0.99013	-0.22846
C	3.01024	-0.09692	-0.90982
C	4.21022	-0.48286	-1.48932
H	5.55528	-2.13869	-1.86278
H	4.02165	-3.78431	-0.76857
H	2.73294	0.93609	-1.0186
H	4.81397	0.26343	-1.99536
C	0.4385	0.83027	0.08513
C	-0.03281	-1.33659	0.97499
C	-0.90102	1.06581	-0.0248
C	-1.45077	-1.15993	0.67912
C	-1.87733	0.03367	0.14973
H	-1.23612	2.09084	-0.12465
C	1.34793	1.99203	0.2564
C	1.25174	3.11406	-0.57964
C	2.24662	2.01943	1.33642
C	2.04067	4.23957	-0.34346
H	0.57308	3.08969	-1.42672
C	3.03165	3.14494	1.56801
H	2.32546	1.15149	1.98316
C	2.93309	4.25878	0.72936
H	1.96262	5.09833	-1.00347
H	3.72098	3.15443	2.40692
H	3.54988	5.13378	0.91025
C	-3.29219	0.36062	-0.16955
C	-3.61786	0.83578	-1.44911
C	-4.3088	0.26159	0.79278

C	-4.93204	1.17609	-1.76779
H	-2.83422	0.92382	-2.19616
C	-5.62143	0.60842	0.47564
H	-4.06072	-0.07017	1.79618
C	-5.93797	1.0616	-0.80659
H	-5.16997	1.53089	-2.76628
H	-6.39607	0.53102	1.23282
H	-6.96107	1.32933	-1.05311
O	1.79608	-3.42496	0.14374
H	1.23925	-3.1519	0.92057
O	0.36013	-2.23072	1.74467
N	0.88724	-0.49195	0.3022
C	-2.30329	-2.38079	0.93784
H	-2.10439	-2.74111	1.95161
H	-3.36123	-2.12254	0.87808
C	-1.99692	-3.50411	-0.07237
H	-2.25054	-3.18632	-1.08878
H	-0.93789	-3.7751	-0.05697
H	-2.58141	-4.39961	0.16071

3(int)d – Intermediate Minimum

C	-4.84668	-1.80445	-0.43445
C	-4.23048	-1.68634	0.8011
C	-2.85617	-1.27392	0.95324
C	-2.20132	-1.00848	-0.30898
C	-2.82372	-1.13922	-1.54324
C	-4.16211	-1.52649	-1.63027
H	-5.89137	-2.1153	-0.47926
H	-4.77367	-1.89603	1.72004
H	-2.24618	-0.93268	-2.4429
H	-4.65195	-1.62208	-2.59521
C	-0.3853	0.64667	-0.13272
C	0.12762	-1.73333	-0.30729
C	0.95432	0.95373	-0.09939
C	1.5502	-1.38909	-0.24008
C	1.94047	-0.07157	-0.15674
H	1.25297	1.99033	-0.00774
C	-1.38118	1.74947	-0.03833
C	-2.31885	1.79961	1.00529
C	-1.31727	2.80491	-0.95855
C	-3.17865	2.89174	1.10657
H	-2.3532	0.97709	1.71904
C	-2.18317	3.89478	-0.85113
H	-0.59175	2.7585	-1.76578
C	-3.11739	3.94016	0.18377

H	-3.90539	2.92143	1.91421
H	-2.12918	4.70197	-1.57751
H	-3.79538	4.78599	0.27027
C	3.36785	0.35618	-0.09438
C	3.832	1.10043	1.00103
C	4.26474	0.07548	-1.13669
C	5.15619	1.53545	1.06172
H	3.14255	1.3265	1.80909
C	5.58844	0.51122	-1.07896
H	3.90919	-0.48176	-1.99813
C	6.04071	1.24134	0.02227
H	5.49765	2.10352	1.92319
H	6.2663	0.28482	-1.89796
H	7.0722	1.58045	0.06815
O	-2.28667	-1.09602	2.07355
O	-0.26642	-2.89444	-0.42175
N	-0.79559	-0.65471	-0.26618
C	2.47684	-2.57966	-0.18427
H	2.16003	-3.29573	-0.95059
H	3.50656	-2.28439	-0.40109
C	2.42111	-3.27872	1.18694
H	2.75338	-2.60087	1.98083
H	1.39576	-3.5885	1.40005
H	3.06589	-4.16588	1.20135

3(int)d – Intermediate Transition State

C	4.39952	-2.6446	-1.21085
C	4.44645	-1.28028	-1.03371
C	3.34316	-0.49686	-0.51936
C	2.17292	-1.2642	-0.13193
C	2.14586	-2.65144	-0.36091
C	3.22785	-3.34903	-0.88292
H	5.26458	-3.17453	-1.61
H	5.32869	-0.70125	-1.29518
H	1.25952	-3.21817	-0.11328
H	3.15406	-4.42241	-1.03522
C	0.50985	0.66349	0.02922
C	-0.0083	-1.47649	1.02281
C	-0.83735	0.89761	-0.12983
C	-1.42303	-1.27127	0.71589
C	-1.82415	-0.09685	0.1178
H	-1.15787	1.91767	-0.30106
C	1.34788	1.89044	0.14784
C	1.24945	2.91459	-0.80208
C	1.99066	2.16389	1.36966

C	1.8233	4.16418	-0.56362
H	0.74481	2.71326	-1.74234
C	2.54798	3.41212	1.60945
H	2.05671	1.37579	2.11233
C	2.47341	4.42267	0.64231
H	1.75844	4.93814	-1.32518
H	3.05063	3.60151	2.55504
H	2.91621	5.39745	0.83111
C	-3.23492	0.24955	-0.20559
C	-3.5655	0.67192	-1.50361
C	-4.25122	0.23139	0.76372
C	-4.87122	1.03813	-1.82934
H	-2.78246	0.70359	-2.25549
C	-5.55725	0.59857	0.44052
H	-4.00256	-0.05683	1.78034
C	-5.87465	0.99957	-0.85904
H	-5.1053	1.35401	-2.84272
H	-6.3268	0.58055	1.20803
H	-6.89236	1.28565	-1.11101
O	3.43564	0.76258	-0.45076
O	0.36381	-2.39217	1.76172
N	0.94154	-0.63155	0.34355
C	-2.32706	-2.42654	1.08324
H	-2.1812	-2.66266	2.14368
H	-3.37424	-2.14721	0.9478
C	-2.03738	-3.6932	0.25755
H	-2.16576	-3.49706	-0.81238
H	-1.0123	-4.02757	0.42978
H	-2.71678	-4.50644	0.53986

4j – Product Minimum

C	2.35700	3.43200	-1.54800
C	2.66400	2.09000	-1.36500
C	1.76400	1.21300	-0.74300
C	0.52400	1.68400	-0.24300
C	0.23500	3.04200	-0.47600
C	1.12500	3.90300	-1.10800
H	3.07400	4.09200	-2.02600
H	3.62500	1.71100	-1.68900
H	-0.70200	3.45500	-0.13900
H	0.84200	4.94100	-1.25000
C	-0.72100	-0.52600	0.15000
C	-1.60200	1.56100	0.99600
C	-1.99600	-0.95200	-0.06900
C	-2.95900	1.14200	0.61500

C	-3.14000	-0.08200	0.02400
H	-2.17400	-2.01800	-0.14800
C	0.29300	-1.55500	0.49300
C	1.01100	-1.44000	1.69400
C	0.44300	-2.71400	-0.27900
C	1.87000	-2.45600	2.10200
H	0.89100	-0.54400	2.29600
C	1.30700	-3.73100	0.13200
H	-0.10200	-2.80400	-1.21300
C	2.02700	-3.60500	1.32100
H	2.42000	-2.35300	3.03300
H	1.42000	-4.62000	-0.48300
H	2.70200	-4.39400	1.63800
C	-4.43800	-0.62000	-0.45900
C	-5.61200	-0.58200	0.31000
C	-4.48800	-1.23200	-1.72400
C	-6.80200	-1.12000	-0.18000
H	-5.58600	-0.15000	1.30300
C	-5.67800	-1.76300	-2.21600
H	-3.58400	-1.27300	-2.32500
C	-6.84100	-1.70700	-1.44500
H	-7.69800	-1.08600	0.43300
H	-5.69800	-2.22000	-3.20100
H	-7.76900	-2.12400	-1.82500
O	-1.39200	2.53900	1.70400
N	-0.50300	0.85400	0.38300
O	2.03900	-0.11200	-0.61400
C	3.13700	-0.71200	-1.29500
H	2.89200	-1.77800	-1.27300
H	3.15900	-0.39100	-2.34600
C	4.47500	-0.47200	-0.62700
C	5.64900	-0.46400	-1.38800
C	4.55500	-0.29600	0.75800
C	6.89000	-0.29200	-0.77200

4j – Product Transition State

C	2.35700	3.43200	-1.54800
C	2.66400	2.09000	-1.36500
C	1.76400	1.21300	-0.74300
C	0.52400	1.68400	-0.24300
C	0.23500	3.04200	-0.47600
C	1.12500	3.90300	-1.10800
H	3.07400	4.09200	-2.02600
H	3.62500	1.71100	-1.68900
H	-0.70200	3.45500	-0.13900

H	0.84200	4.94100	-1.25000
C	-0.72100	-0.52600	0.15000
C	-1.60200	1.56100	0.99600
C	-1.99600	-0.95200	-0.06900
C	-2.95900	1.14200	0.61500
C	-3.14000	-0.08200	0.02400
H	-2.17400	-2.01800	-0.14800
C	0.29300	-1.55500	0.49300
C	1.01100	-1.44000	1.69400
C	0.44300	-2.71400	-0.27900
C	1.87000	-2.45600	2.10200
H	0.89100	-0.54400	2.29600
C	1.30700	-3.73100	0.13200
H	-0.10200	-2.80400	-1.21300
C	2.02700	-3.60500	1.32100
H	2.42000	-2.35300	3.03300
H	1.42000	-4.62000	-0.48300
H	2.70200	-4.39400	1.63800
C	-4.43800	-0.62000	-0.45900
C	-5.61200	-0.58200	0.31000
H	2.42000	-2.35300	3.03300
H	1.42000	-4.62000	-0.48300
H	2.70200	-4.39400	1.63800
C	-4.43800	-0.62000	-0.45900
C	-5.61200	-0.58200	0.31000
C	-4.48800	-1.23200	-1.72400
C	-6.80200	-1.12000	-0.18000
H	-5.58600	-0.15000	1.30300
C	-5.67800	-1.76300	-2.21600
H	-3.58400	-1.27300	-2.32500
C	-6.84100	-1.70700	-1.44500
H	-7.69800	-1.08600	0.43300
H	-5.69800	-2.22000	-3.20100
H	-7.76900	-2.12400	-1.82500
O	-1.39200	2.53900	1.70400
C	-6.84100	-1.70700	-1.44500
H	-7.69800	-1.08600	0.43300
H	-5.69800	-2.22000	-3.20100
H	-7.76900	-2.12400	-1.82500
O	-1.39200	2.53900	1.70400
N	-0.50300	0.85400	0.38300
O	2.03900	-0.11200	-0.61400
C	3.13700	-0.71200	-1.29500
H	2.89200	-1.77800	-1.27300
H	3.15900	-0.39100	-2.34600
C	4.47500	-0.47200	-0.62700
C	5.64900	-0.46400	-1.38800
C	4.55500	-0.29600	0.75800
C	6.89000	-0.29200	-0.77200

3e – Minimum

C	4.8619	-1.52496	0.98473
C	4.28561	-2.05889	-0.16374
C	2.94096	-1.81429	-0.4674
C	2.19141	-0.9968	0.39725

C	2.76391	-0.49742	1.5698	C	4.62371	-1.91083	-1.53454
C	4.09829	-0.75442	1.86659	C	3.72972	-2.81027	-0.98419
H	5.9063	-1.72653	1.20345	C	2.50898	-2.41401	-0.40685
H	4.85003	-2.68343	-0.84767	C	2.20706	-1.01998	-0.30318
H	2.1582	0.11459	2.22993	C	3.11653	-0.13579	-0.92094
H	4.53705	-0.35671	2.77552	C	4.29745	-0.55519	-1.51581
C	0.50319	0.74268	0.00943	H	5.54755	-2.26001	-1.98503
C	-0.14573	-1.60847	-0.05791	H	3.91404	-3.87894	-1.00089
C	-0.81679	1.11095	0.01756	H	2.90076	0.91652	-0.96504
C	-1.52481	-1.19294	0.01676	H	4.94888	0.18421	-1.97001
C	-1.86398	0.14716	0.07752	C	0.57715	0.86377	0.10979
H	-1.06058	2.16324	-0.04813	C	-0.01657	-1.31564	0.85461
C	1.57403	1.75531	-0.15255	C	-0.74766	1.17874	-0.00134
C	2.55865	1.59526	-1.14136	C	-1.4106	-1.03617	0.56618
C	1.57138	2.91572	0.63262	C	-1.79111	0.20367	0.10154
C	3.52356	2.57953	-1.33171	H	-1.01968	2.22558	-0.05011
H	2.55863	0.70195	-1.75736	C	1.54411	1.96499	0.35025
C	2.54301	3.89793	0.44074	C	1.51821	3.13244	-0.42636
H	0.81727	3.03494	1.40469	C	2.42735	1.88567	1.44022
C	3.52145	3.73135	-0.53973	C	2.36395	4.19887	-0.12277
H	4.27876	2.44799	-2.10049	H	0.85051	3.18982	-1.28049
H	2.53816	4.78857	1.06178	C	3.26764	2.95361	1.74009
H	4.27988	4.49425	-0.68729	H	2.44968	0.98269	2.04177
C	-3.26344	0.65294	0.10272	C	3.24074	4.11286	0.95965
C	-3.661	1.61483	-0.84208	H	2.3415	5.09401	-0.7369
C	-4.18682	0.2308	1.07211	H	3.94377	2.88255	2.5866
C	-4.95569	2.12925	-0.83004	H	3.90078	4.94247	1.19363
H	-2.95601	1.94242	-1.60075	C	-3.16453	0.62317	-0.27182
C	-5.47795	0.75752	1.08702	C	-3.33673	1.3442	-1.46787
H	-3.88768	-0.51087	1.80154	C	-4.28566	0.37607	0.53722
C	-5.8677	1.7033	0.13781	C	-4.599	1.78245	-1.85983
H	-5.25182	2.86092	-1.57589	H	-2.47492	1.54052	-2.09938
H	-6.18055	0.42523	1.84529	C	-5.54562	0.82745	0.14533
H	-6.87589	2.10679	0.15131	H	-4.16072	-0.16494	1.46573
O	2.41111	-2.33609	-1.60089	C	-5.70886	1.52556	-1.05183
H	1.51896	-2.67062	-1.33824	H	-4.71614	2.32378	-2.79396
O	0.20931	-2.79528	-0.2066	H	-6.4025	0.63379	0.78358
N	0.83531	-0.60218	0.07021	H	-6.69354	1.87102	-1.35274
C	-2.53945	-2.31777	-0.03386	O	1.70557	-3.44586	-0.07134
O	-3.23732	-2.57966	0.92767	H	1.17151	-3.18966	0.72675
C	-2.65248	-3.05783	-1.34602	O	0.2993	-2.27739	1.57776
H	-1.66985	-3.43135	-1.6441	N	0.95902	-0.48789	0.24507
H	-3.36225	-3.88013	-1.24396	C	-2.31721	-2.22731	0.79488
H	-2.99948	-2.36223	-2.1201	O	-3.24305	-2.19729	1.58364
				C	-2.04853	-3.43767	-0.07698
				H	-0.98407	-3.60562	-0.25078
				H	-2.50923	-4.3201	0.36994
				H	-2.51899	-3.25212	-1.05138

3e – Transition State

3(int)e – Intermediate Minimum

C	-4.8972	-1.885	-0.27486
C	-4.26268	-1.71273	0.94467
C	-2.8989	-1.25649	1.05741
C	-2.2785	-1.00865	-0.22495
C	-2.91651	-1.19537	-1.44412
C	-4.24367	-1.62331	-1.49195
H	-5.9326	-2.22724	-0.29046
H	-4.78217	-1.91103	1.87947
H	-2.36205	-1.0005	-2.36049
H	-4.74878	-1.76226	-2.4434
C	-0.51064	0.68798	-0.11789
C	0.07414	-1.68327	-0.24406
C	0.82894	1.03595	-0.09678
C	1.48581	-1.29299	-0.20585
C	1.83813	0.05287	-0.13016
H	1.09207	2.07855	0.02422
C	-1.52959	1.77024	-0.03964
C	-2.45743	1.82711	1.01156
C	-1.49388	2.79974	-0.99009
C	-3.34097	2.9022	1.08879
H	-2.46386	1.02605	1.74996
C	-2.38698	3.8691	-0.90857
H	-0.77158	2.74891	-1.79992
C	-3.31313	3.92209	0.13347
H	-4.0603	2.93984	1.90241
H	-2.35882	4.65544	-1.65848
H	-4.01036	4.75366	0.20094
C	3.23755	0.55253	0.01036
C	4.04171	0.15853	1.08974
C	3.72859	1.52142	-0.87367
C	5.30788	0.70743	1.26913
H	3.66212	-0.58361	1.78459
C	5.00268	2.06471	-0.70131
H	3.10751	1.83743	-1.7067
C	5.79692	1.66112	0.37163
H	5.91627	0.39126	2.11233
H	5.37178	2.80606	-1.40534
H	6.78716	2.08666	0.51103
O	-2.31032	-1.02864	2.15855
O	-0.32957	-2.83747	-0.34763
N	-0.88287	-0.61398	-0.21434
C	2.52754	-2.35428	-0.33334
O	3.63444	-2.10272	-0.80362
C	2.22162	-3.76438	0.13793

H	1.63245	-3.76923	1.05813
H	1.60447	-4.27202	-0.6075
H	3.17188	-4.28879	0.2664

3(int)e – Intermediate Transition State

C	4.39291	-2.74668	-1.25331
C	4.53928	-1.40659	-0.98603
C	3.46491	-0.5559	-0.5111
C	2.21216	-1.24678	-0.24426
C	2.08813	-2.61231	-0.57066
C	3.14181	-3.36598	-1.06163
H	5.23755	-3.32582	-1.6254
H	5.48379	-0.89295	-1.14565
H	1.132	-3.10557	-0.4476
H	2.98887	-4.41264	-1.30838
C	0.61275	0.72557	0.0345
C	0.01724	-1.45194	0.87352
C	-0.74784	0.99486	-0.09816
C	-1.36292	-1.26845	0.43746
C	-1.7398	0.00464	-0.02108
H	-1.04597	2.03231	-0.17626
C	1.4837	1.91522	0.20009
C	1.36604	3.01104	-0.66279
C	2.24853	2.05278	1.37239
C	2.02455	4.20942	-0.38256
H	0.77744	2.90944	-1.56969
C	2.88264	3.25358	1.66105
H	2.33612	1.20442	2.04159
C	2.78047	4.33802	0.78185
H	1.94253	5.04284	-1.07593
H	3.47391	3.34292	2.56867
H	3.28761	5.27323	1.0054
C	-3.1362	0.45371	-0.27698
C	-3.42458	1.19336	-1.43268
C	-4.15944	0.26644	0.66615
C	-4.70169	1.71324	-1.65263
H	-2.63804	1.34833	-2.16524
C	-5.42887	0.79314	0.45443
H	-3.94588	-0.29967	1.56616
C	-5.70911	1.51736	-0.70919
H	-4.90544	2.27345	-2.56164
H	-6.20568	0.63863	1.19887
H	-6.70317	1.92452	-0.87481
O	3.65191	0.68405	-0.37344
O	0.40306	-2.29483	1.67424

N	1.00991	-0.58547	0.2238	H	-5.8235	-1.228	-1.7779
C	-2.29669	-2.4183	0.454	H	-0.7286	1.908	1.5637
O	-3.36438	-2.38298	-0.1599	H	-5.0567	-3.3548	-0.7426
C	-1.93201	-3.68817	1.20905	H	-2.7167	-1.8191	2.3501
H	-0.99201	-4.11249	0.84466	H	3.2102	-1.1115	2.9325
H	-1.76296	-3.48151	2.26858	H	-2.4704	3.111	2.8518
H	-2.75054	-4.39869	1.07435	H	0.506	1.3026	-0.529

4m – Product Minimum

O	-0.2949	-3.0466	-1.5741
O	-1.6507	0.5954	-1.2491
N	-0.8719	-1.5713	0.078
C	-2.2075	-1.5351	-0.459
C	-2.615	-0.3541	-1.1059
C	0.1021	-2.2726	-0.7087
C	-0.5337	-0.7276	1.1119
C	1.4979	-1.9644	-0.4117
C	-1.6019	-0.0363	1.875
C	1.8193	-1.0715	0.6
C	-3.931	-0.2401	-1.5594
C	-3.0711	-2.6169	-0.3528
C	0.7884	-0.5032	1.3956
C	3.1923	-0.5539	0.8538
C	-0.7517	2.7956	-1.4314
C	-2.015	1.9647	-1.3987
C	-4.8038	-1.3211	-1.4165
C	-1.5378	1.3565	2.0307
C	-4.3777	-2.5135	-0.8317
C	-2.663	-0.7421	2.4618
C	3.7487	-0.6042	2.1374
C	-2.5255	2.0315	2.7481
C	0.4549	2.3138	-0.9108
C	-3.6424	-0.0658	3.1845
C	3.8924	0.1068	-0.167
C	-3.58	1.3228	3.3254
C	5.686	0.6238	1.3703
C	4.9944	-0.0295	2.3907
C	5.1285	0.6939	0.0909
C	1.5961	3.118	-0.9096
C	1.5423	4.4132	-1.426
H	-4.2694	0.6736	-2.0334
H	-2.712	-3.5262	0.116
H	1.0431	0.17	2.2036
H	-2.65	2.2537	-0.5477
H	-2.5917	2.123	-2.3202

H	-5.8235	-1.228	-1.7779
H	-0.7286	1.908	1.5637
H	-5.0567	-3.3548	-0.7426
H	-2.7167	-1.8191	2.3501
H	3.2102	-1.1115	2.9325
H	-2.4704	3.111	2.8518
H	0.506	1.3026	-0.529
H	-4.4564	-0.6236	3.6375
H	3.457	0.1616	-1.16
H	-4.3496	1.8481	3.883
H	6.6525	1.0772	1.5695
H	5.4225	-0.0907	3.3869
H	5.6583	1.2045	-0.7079
H	2.5244	2.7213	-0.5078
H	2.4293	5.0399	-1.4253
C	-0.7951	4.0898	-1.9619
C	0.3428	4.8966	-1.9541
H	-1.723	4.47	-2.3834
H	0.2949	5.8993	-2.369
C	2.3074	-3.0595	-2.6359
H	1.695	-3.9644	-2.6595
H	1.7356	-2.2942	-3.1676
H	3.2683	-3.2426	-3.1204
C	2.5732	-2.6651	-1.198
O	3.654	-2.9001	-0.6781

4m – Product Transition State

O	-0.29498	-3.04641	-1.57447
O	-1.65068	0.59558	-1.24904
N	-0.87192	-1.57132	0.07782
C	-2.20753	-1.53498	-0.45912
C	-2.61501	-0.35394	-1.1059
C	0.10199	-2.27244	-0.70888
C	-0.53377	-0.72767	1.11184
C	1.49783	-1.96438	-0.41188
C	-1.60197	-0.0365	1.87501
C	1.81922	-1.07164	0.59995
C	-3.93101	-0.2398	-1.55945
C	-3.07119	-2.61675	-0.35302
C	0.78837	-0.50338	1.39563
C	3.19225	-0.55409	0.8538
C	-0.75157	2.79576	-1.43123
C	-2.01495	1.96489	-1.39852
C	-4.80387	-1.32084	-1.41665
C	-1.53778	1.35635	2.03095

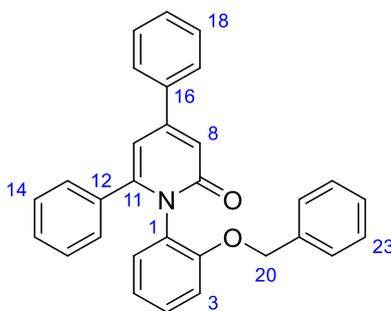
C	-4.37781	-2.51326	-0.83198
C	-2.66305	-0.74232	2.46172
C	3.74864	-0.60452	2.13746
C	-2.5255	2.03123	2.74837
C	0.45499	2.31389	-0.91044
C	-3.64247	-0.06613	3.18453
C	3.89241	0.10676	-0.16683
C	-3.58004	1.32244	3.32565
C	5.68602	0.6235	1.37049
C	4.9944	-0.02994	2.39078
C	5.12849	0.69379	0.09112
C	1.59618	3.11804	-0.90928
C	1.54254	4.41322	-1.42592
H	-4.2694	0.67394	-2.03337
H	-2.71215	-3.5261	0.11568
H	1.04308	0.1697	2.2037
H	-2.6498	2.25387	-0.54736
H	-2.59173	2.12333	-2.31987
H	-5.82355	-1.22761	-1.77805
H	-0.72863	1.90781	1.56397
H	-5.05685	-3.35454	-0.74293
H	-2.71681	-1.81928	2.34995
H	3.21017	-1.11198	2.93243
H	-2.47034	3.11076	2.8522
H	0.50595	1.30278	-0.5286
H	-4.45641	-0.62391	3.63751
H	3.45701	0.16176	-1.15983
H	-4.34963	1.84777	3.88327
H	6.65246	1.07683	1.56974
H	5.42242	-0.09127	3.38704
H	5.65831	1.2045	-0.70761
H	2.52447	2.72137	-0.50743
H	2.42953	5.03986	-1.42526
C	-0.7949	4.08986	-1.96185
C	0.34314	4.89665	-1.95407
H	-1.7227	4.47012	-2.38353
H	0.29527	5.89921	-2.36918
C	2.30738	-3.05918	-2.63618
H	1.69492	-3.964	-2.65998
H	1.73556	-2.29372	-3.16781
H	3.26827	-3.24222	-3.12075
C	2.57317	-2.66502	-1.1983
O	3.65397	-2.9001	-0.67842

7 HPLC data

7.1 O-Benzylated N-(2-phenoxy)pyridones (4)

Racemic traces were generated *via* thermal racemisation of the enantio-enriched materials, using the end-point trace generated according to General Procedure F (see page S37 below).

1-(2-(Benzyloxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4a)



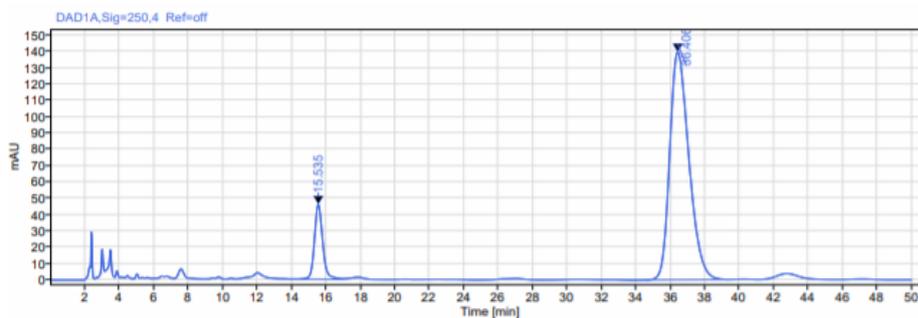
Conditions: HPLC: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 36.4 min, t_R (minor) = 15.5 min.

Asymmetric trace: 75% ee

Single Injection Report



Data file: JS-4-289 rot. bar. 80 oC t=0h 60% MeCN-H2O20210423 122213.dx
Sequence Name: SingleSample **Project Name:** JS
Sample name: JS-4-289 rot. bar. 80 oC t=0h 60% MeCN/H2O **Operator:** SYSTEM
Instrument: 1100HPLC **Injection date:** 2021-04-23 12:23:21+01:00
Inj. volume: 10.000 **Location:** 43
Acq. method: 60% MeCN-H2O 45 MIN.amx **Type:** Sample
Processing method: 3D UV Quantitative_DefaultMethod.pmx **Sample amount:** 0.00
Manually modified: Manual Integration



Signal: DAD1A,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
15.535	BV	2.53	1522.20	45.83	12.35	
36.406	BB	4.83	10804.04	139.60	87.65	
	Sum		12326.23			

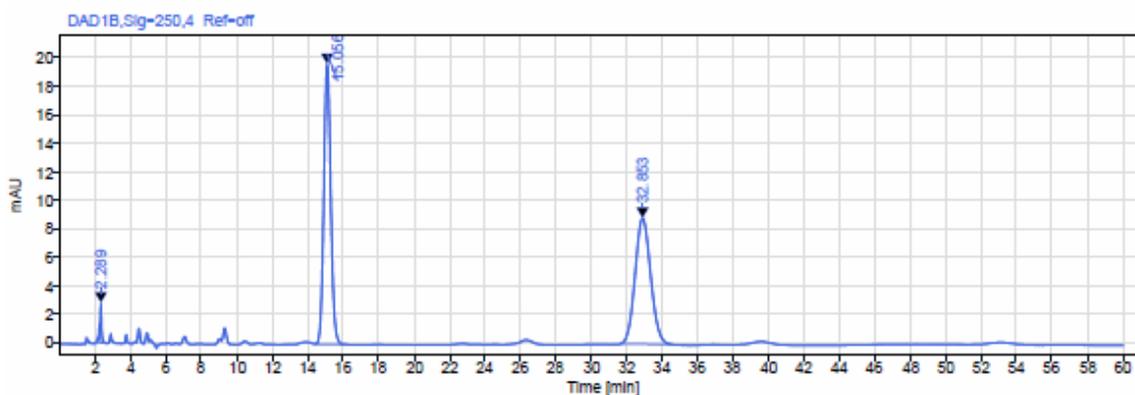
Racemic trace

Single Injection Report



Agilent Technologies

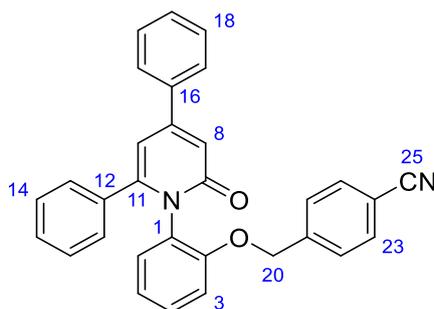
Data file:	JS #289 S10 76H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #289 S10 76H	Injection date:	2021-07-08 17:43:02+01:00
Instrument:	1220LC	Location:	2
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	None		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.289	VB	0.23	12.81	2.80	1.18	
15.056	BB	1.69	531.33	19.66	49.03	
32.853	BB	2.87	539.44	8.83	49.78	
		Sum	1083.58			

4-((2-(2-Oxo-4,6-diphenylpyridin-1(2H)-yl)phenoxy)methyl)benzonitrile (4b)

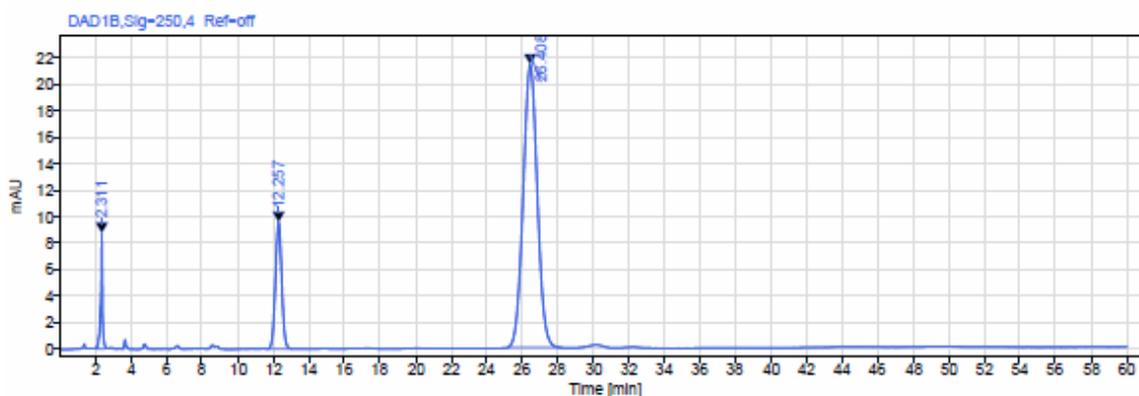


Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 26.4 min, t_R (minor) = 12.3 min.

Asymmetric trace: 67% ee

Single Injection Report 

Data file:	JS #302 80C Origin.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #302 80C Origin	Injection date:	2021-07-23 23:39:50+01:00
Instrument:	1220LC	Location:	21
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	None		



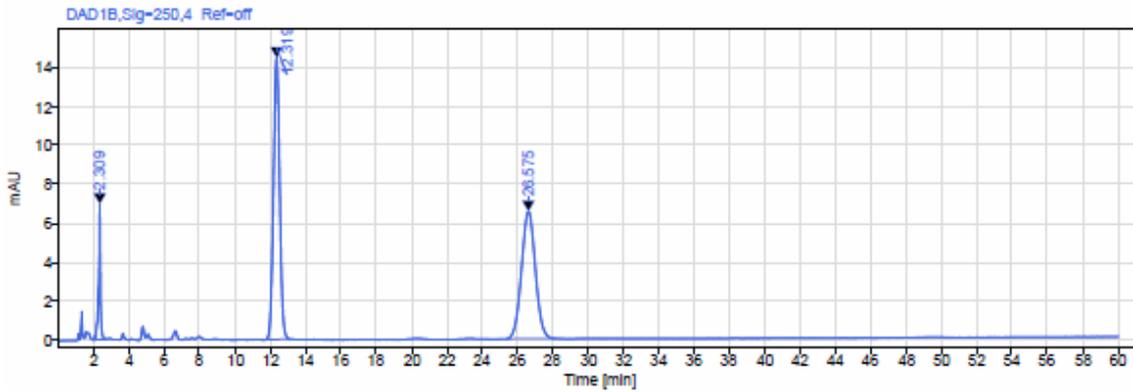
Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.311	VB	0.61	55.44	8.71	3.77	
12.257	BB	1.37	234.75	9.55	15.96	
26.408	BB	2.96	1180.71	21.35	80.27	
	Sum		1470.89			

Racemic trace

Single Injection Report 

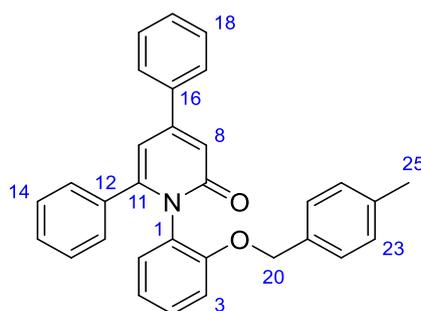
Data file: JS #298 80C S9 102H.dx
Sequence Name: SingleSample **Project Name:** Agilent
Sample name: JS #298 80C S9 102H **Operator:** Agilent
Instrument: 1220LC **Injection date:** 2021-07-22 20:27:57+01:00
Inj. volume: 5.000 **Location:** 64
Acq. method: Atropisomer barrier measurement.amx **Type:** Sample
Processing method: 3D UV Quantitative_DefaultMethod.pmx **Sample amount:** 1.00
Manually modified: None



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.309	VB	0.63	44.97	6.96	5.99	
12.319	BB	1.34	349.81	14.45	46.62	
26.575	BB	2.70	355.54	6.53	47.39	
		Sum	750.31			

1-(2-((4-Methylbenzyl)oxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4c)



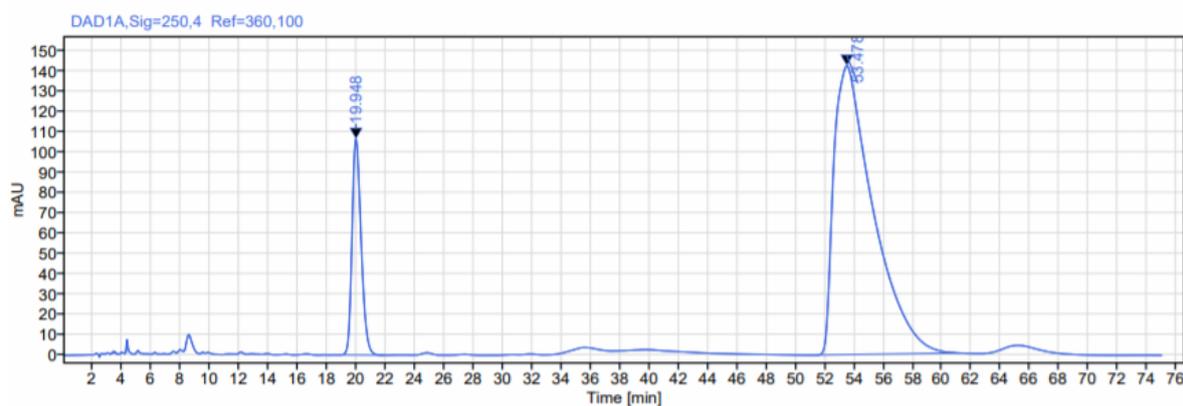
Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, $\lambda = 250$ nm, t_R (major) = 53.5 min, t_R (minor) = 19.9 min.

Asymmetric trace: 70% ee

Single Injection Report



Data file:	JS-4-303 col 60% MeCN-H2O20210504 142035.dx		
Sequence Name:	SingleSample	Project Name:	JS
Sample name:	JS-4-303 col 60% MeCN/H2O	Operator:	SYSTEM
Instrument:	1100HPLC	Injection date:	2021-05-04 14:21:44+01:00
Inj. volume:	5.000	Location:	17
Acq. method:	60% MeCN-H2O 45 MIN.amx	Type:	Sample
Processing method:	3D UV Quantitative_DefaultMethod.pmx	Sample amount:	0.00
Manually modified:	Manual Integration		



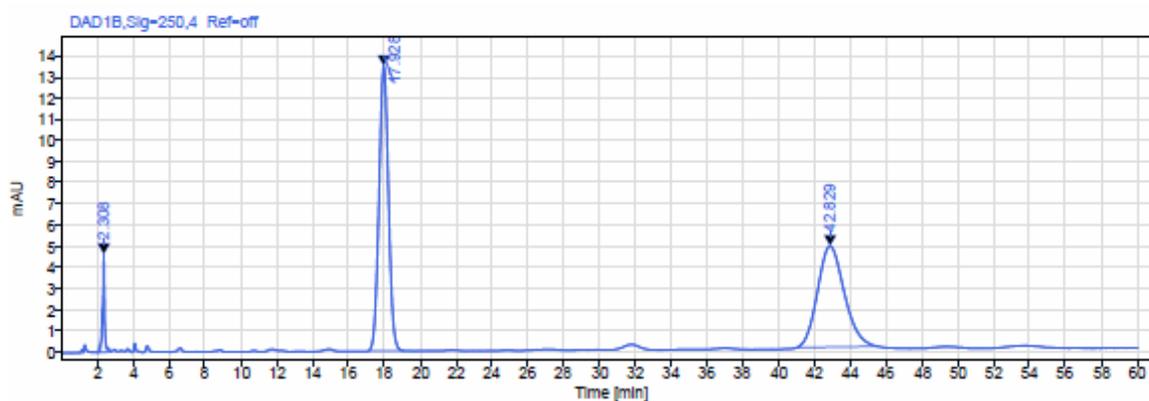
Signal: DAD1A,Sig=250,4 Ref=360,100

RT [min]	Type	Width [min]	Area	Height	Area%	Name
19.948	BB	3.00	4592.01	106.31	14.78	
53.478	BB	9.53	26471.31	142.11	85.22	
		Sum	31063.32			

Racemic trace

Single Injection Report Agilent Technologies

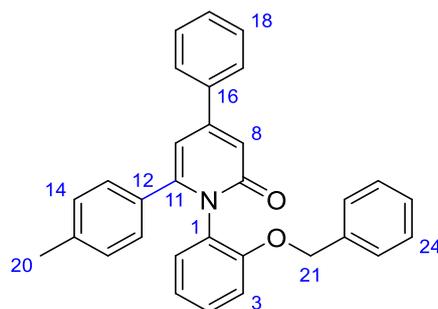
Data file:	JS #303 80C S9 70H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #303 80C S9 70H	Injection date:	2021-07-24 18:59:09+01:00
Instrument:	1220LC	Location:	80
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	None		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.308	VV	0.40	32.21	4.63	3.16	
17.928	BB	1.97	491.37	13.45	48.27	
42.829	BB	4.44	494.39	4.78	48.57	
	Sum		1017.96			

1-(2-(Benzyloxy)phenyl)-4-phenyl-6-(*p*-tolyl)pyridin-2(1*H*)-one (4d)

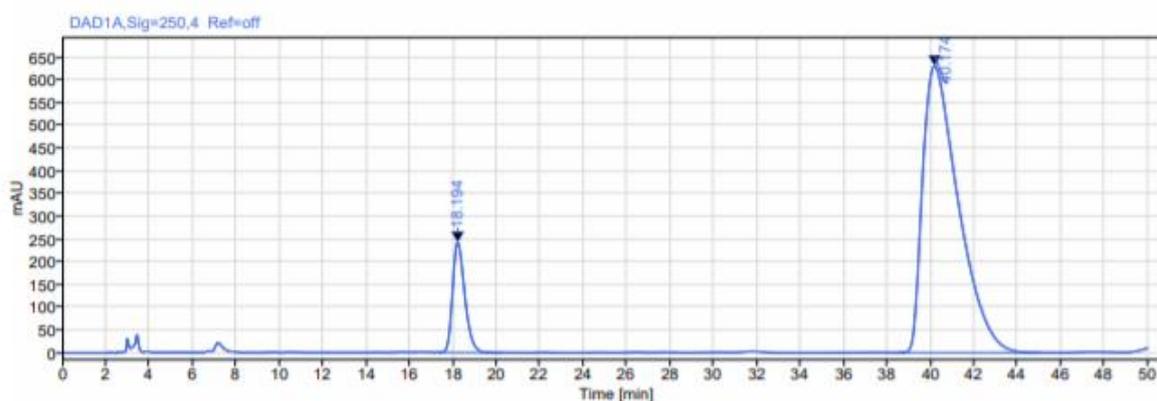


Conditions: HPLC: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 40.2 min, t_R (minor) = 18.2 min.

Asymmetric trace: 77% ee

Single Injection Report 

Data file: JS-5-308 col 60% MeCN-H2O20210430 153237.dx
Sequence Name: SingleSample **Project Name:** JS
Sample name: JS-5-308 col 60% MeCN/H2O **Operator:** SYSTEM
Instrument: 1100HPLC **Injection date:** 2021-04-30 15:49:26+01:00
Inj. volume: 10.000 **Location:** 15
Acq. method: 60% MeCN-H2O 45 MIN.amx **Type:** Sample
Processing method: 3D UV Quantitative_DefaultMethod.pmx **Sample amount:** 0.00
Manually modified: Manual Integration



Signal: DAD1A,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
18.194	BB	2.89	9504.28	242.00	11.63	
40.174	BB	6.55	72200.43	630.27	88.37	
	Sum		81704.70			

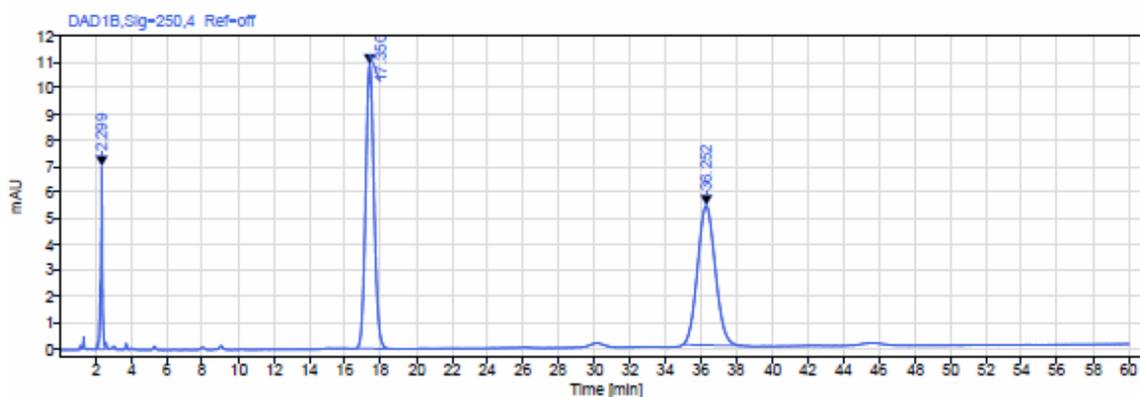
Racemic trace

Single Injection Report



Agilent Technologies

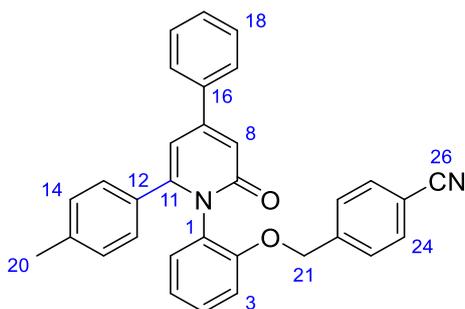
Data file:	JS #308 80C S9 70.5H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #308 80C S9 70.5H	Injection date:	2021-07-31 22:02:06+01:00
Instrument:	1220LC	Location:	55
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	None		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.299	VV	0.37	40.37	7.01	5.28	
17.350	BB	1.85	354.39	10.88	48.39	
36.252	BB	3.27	369.14	5.34	48.32	
	Sum		763.89			

4-((2-(2-Oxo-4-phenyl-6-(*p*-tolyl)pyridin-1(2*H*)-yl)phenoxy)methyl)benzonitrile (4e)

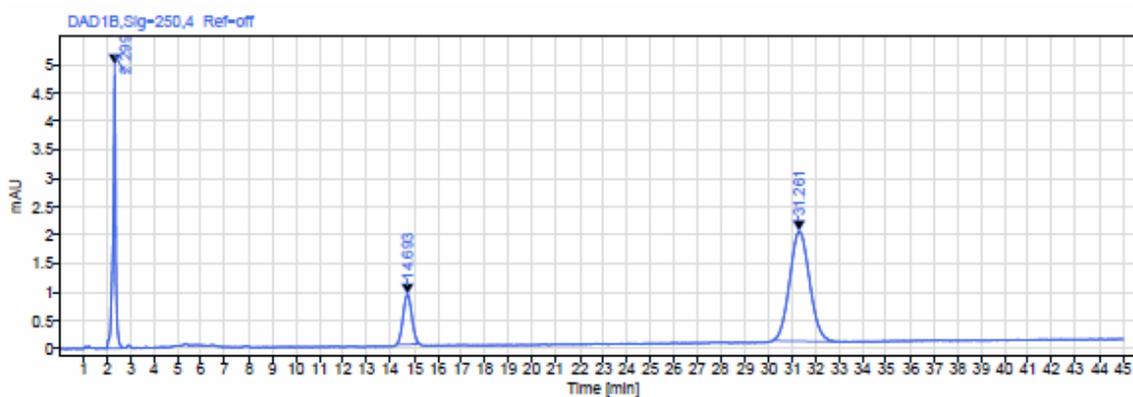


Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 31.3 min, t_R (minor) = 14.7 min.

Asymmetric trace: 67% ee

Single Injection Report 

Data file:	JS #292 70C Origin.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #292 70C Origin	Injection date:	2021-07-26 20:40:36+01:00
Instrument:	1220LC	Location:	1
Inj. volume:	5.000	Type:	Sample
Acq. method:	(45min) Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	Manual Integration		



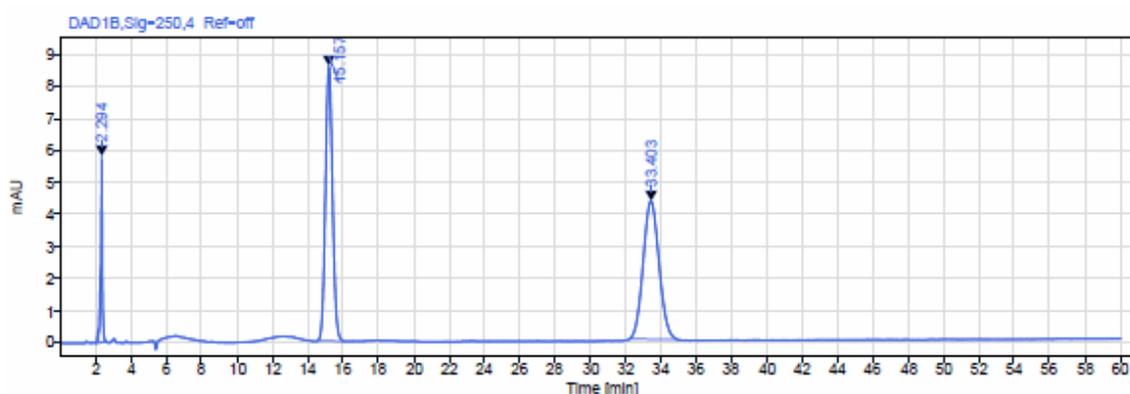
Signal: DAD1B, Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.299	BB	0.76	36.37	4.99	21.12	
14.693	MM m	0.34	22.46	0.88	13.04	
31.261	BB	2.69	113.35	1.93	65.83	
	Sum		172.18			

Racemic trace

Single Injection Report

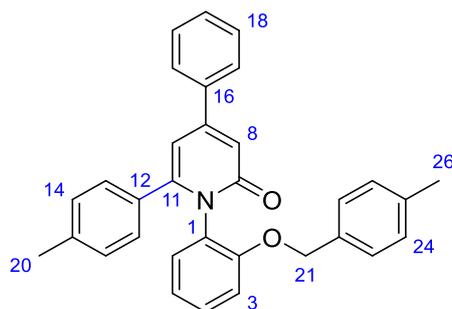
Data file:	JS #292 80C S10 54H (07.09).dx		
Sequence Name:	SingleSample	Project Name:	Agilent
Sample name:	JS #292 80C S10 54H (07.09)	Operator:	Agilent
Instrument:	1220LC	Injection date:	2021-07-09 20:40:05+01:00
Inj. volume:	5.000	Location:	32
Acq. method:	Atropisomer barrier measurement.amx	Type:	Sample
Processing method:	3D UV Quantitative_DefaultMethod.pmx	Sample amount:	1.00
Manually modified:	None		



Signal: DAD1B, Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.294	VB	0.39	32.98	5.81	6.11	
15.157	BB	1.44	237.55	8.57	44.04	
33.403	BB	2.70	268.82	4.31	49.84	
	Sum		539.35			

1-(2-((4-methylbenzyl)oxy)phenyl)-4-phenyl-6-(*p*-tolyl)pyridin-2(1*H*)-one (4f)

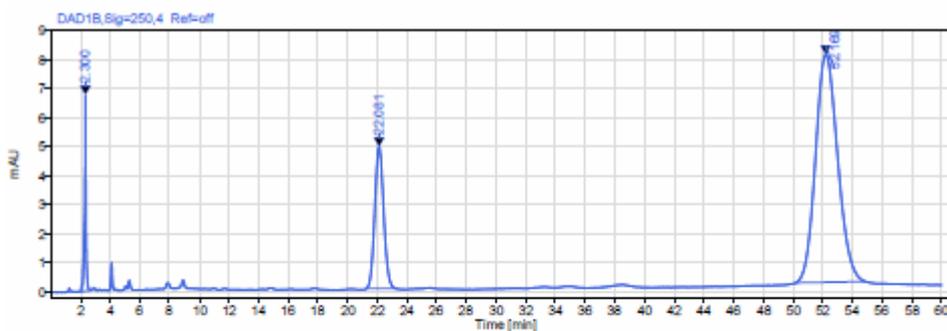


Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 52.8 min, t_R (minor) = 22.1 min.

Asymmetric trace: 58% ee

Single Injection Report 

Data file: JS #296 70C Origin.dx
 Sequence Name: SingleSample
 Sample name: JS #296 70C Origin
 Instrument: 1220LC
 Inj. volume: 5.000
 Acq. method: Atropisomer barrier measurement.amx
 Processing method: 3D UV Quantitative_DefaultMethod.pmx
 Manually modified: Manual Integration
 Project Name: Agilent
 Operator: Agilent
 Injection date: 2021-07-27 12:02:34+01:00
 Location: 21
 Type: Sample
 Sample amount: 1.00



Signal: DAD1B,Sig=250,4 Ref-off

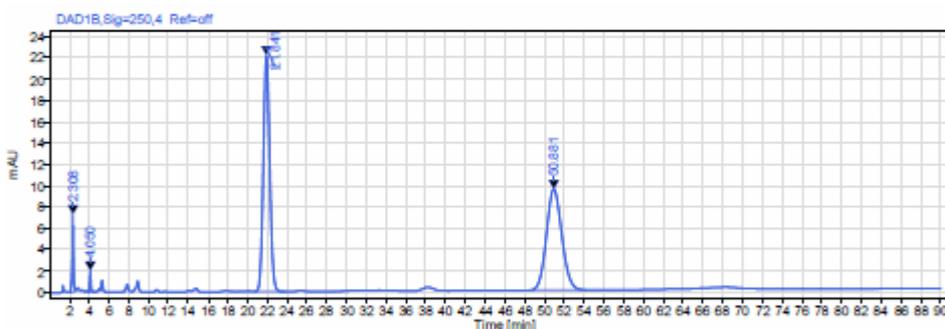
RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.300	VB	0.54	40.96	6.77	3.87	
22.081	BB	2.00	204.03	4.90	19.28	
52.169	MM m	1.21	813.43	7.85	76.85	
	Sum		1058.42			

Racemic Trace

Single Injection Report



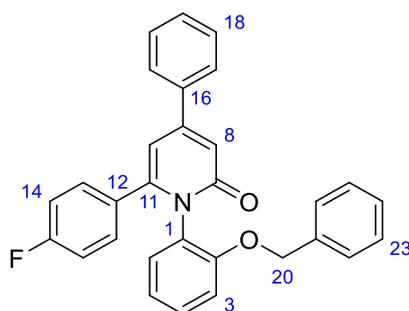
Data file:	JS #296 80C S9 102H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #296 80C S9 102H	Injection date:	2021-07-22 17:25:41+01:00
Instrument:	1220LC	Location:	62
Inj. volume:	5.000	Type:	Sample
Acq. method:	(90min) atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmh		
Manually modified:	None		



Signal: DAD1B,Sig=250,4 Ref-off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.308	VV	0.46	52.32	7.29	2.52	
4.050	BB	0.43	13.59	1.99	0.65	
21.841	BB	3.05	993.05	22.23	47.75	
50.881	BB	5.47	1020.69	9.48	49.08	
	Sum		2079.64			

1-(2-(Benzyloxy)phenyl)-6-(4-fluorophenyl)-4-phenylpyridin-2(1H)-one (4g)

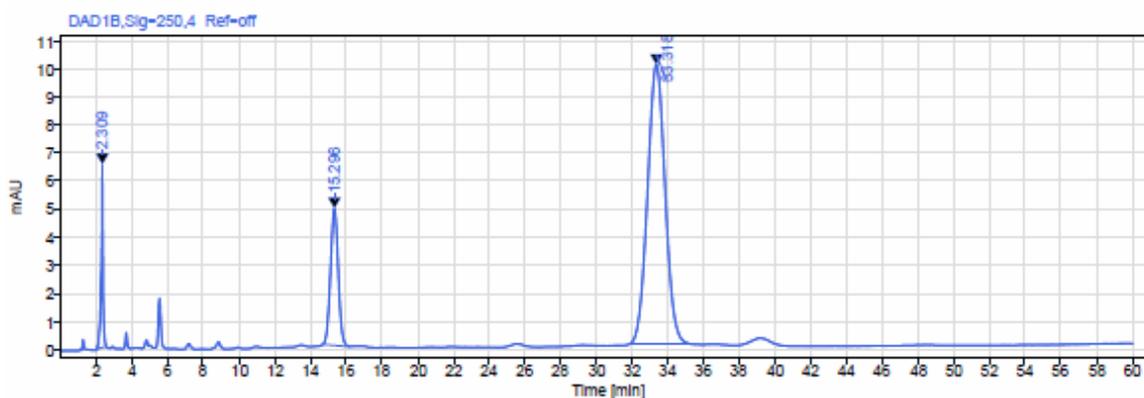


Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 33.3 min, t_R (minor) = 15.3 min.

Asymmetric trace: 65% ee

Single Injection Report 

Data file:	JS #301 80C Origin.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #301 80C Origin	Injection date:	2021-07-23 13:29:44+01:00
Instrument:	1220LC	Location:	11
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	Manual Integration		



Signal: DAD1B,Sig=250,4 Ref=off

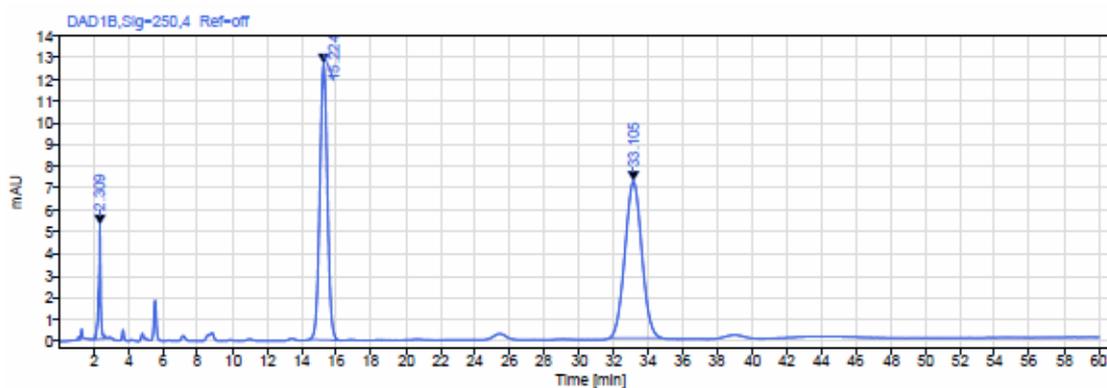
RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.309	MM m	0.09	42.16	6.50	4.86	
15.296	BB	1.50	145.65	4.86	16.80	
33.318	BB	3.31	678.99	9.91	78.33	
	Sum		866.80			

Racemic trace

Single Injection Report



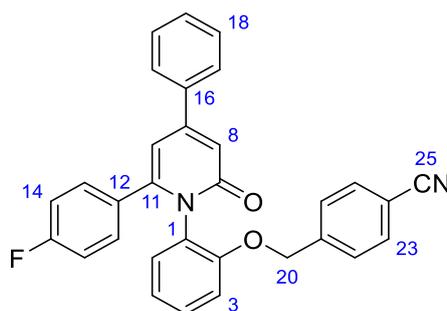
Data file:	JS #301 80C S8 48H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #301 80C S8 48H	Injection date:	2021-07-23 21:37:46+01:00
Instrument:	1220LC	Location:	19
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	Manual Integration		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.309	VV	0.44	36.47	5.23	4.02	
15.224	BB	1.78	384.75	12.65	42.46	
33.105	BB	3.00	485.02	7.17	53.52	
	Sum		906.24			

4-((2-(6-(4-Fluorophenyl)-2-oxo-4-phenylpyridin-1(2H)-yl)phenoxy)methyl)benzonitrile (4h)

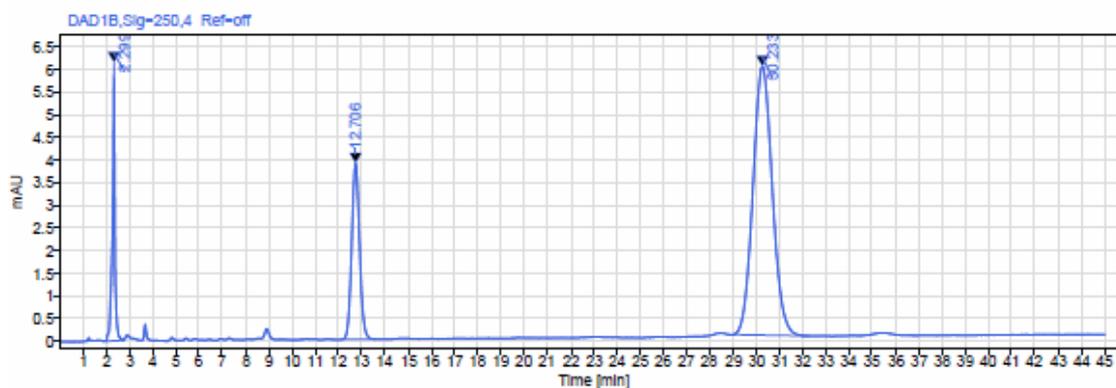


Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 30.3 min, t_R (minor) = 12.7 min.

Asymmetric trace: 58% ee

Single Injection Report 

Data file:	JS #293 70C Origin.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #293 70C Origin	Injection date:	2021-07-27 04:21:37+01:00
Instrument:	1220LC	Location:	11
Inj. volume:	5.000	Type:	Sample
Acq. method:	(45min) Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pm		
Manually modified:	None		



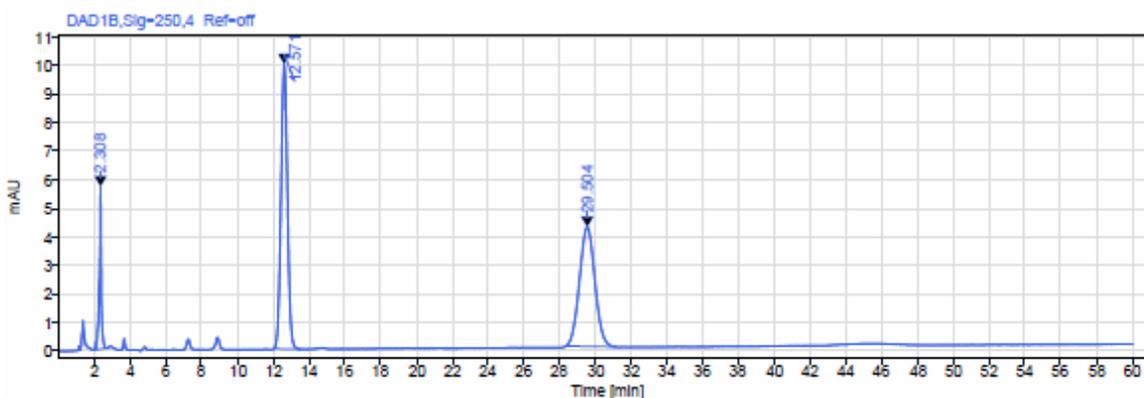
Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.299	BB	0.78	43.74	6.13	9.14	
12.706	BB	1.36	91.24	3.87	19.07	
30.233	BB	3.24	343.41	5.92	71.79	
	Sum		478.39			

Racemic trace

Single Injection Report

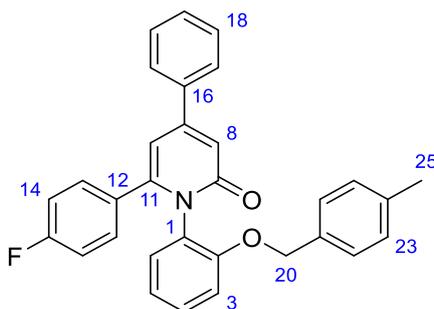
Data file:	JS #293 80C S9 102H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #293 80C S9 102H	Injection date:	2021-07-22 16:24:22+01:00
Instrument:	1220LC	Location:	61
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	None		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.308	VB	0.57	39.42	5.71	7.31	
12.571	BB	1.48	247.95	10.01	48.01	
29.504	BB	2.81	251.57	4.19	48.68	
	Sum		538.94			

6-(4-Fluorophenyl)-1-(2-((4-methylbenzyl)oxy)phenyl)-4-phenylpyridin-2(1H)-one (4i)

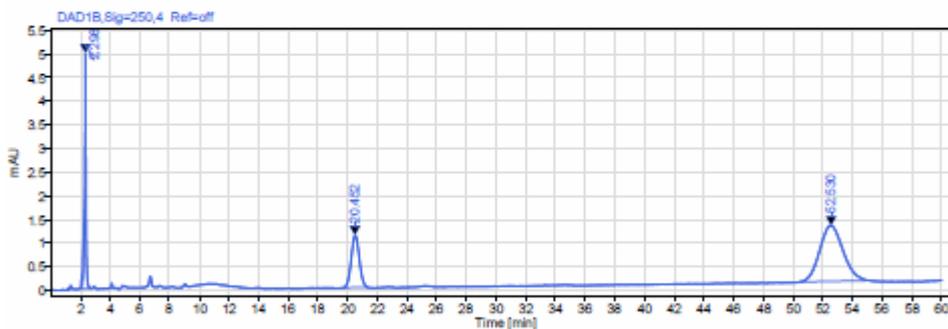


Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 52.3 min, t_R (minor) = 20.5 min.

Asymmetric trace: 51% ee

Single Injection Report 

Data file:	JS #297 80C Origin.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #297 80C Origin	Injection date:	2021-07-17 01:22:17+01:00
Instrument:	1220LC	Location:	17
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	Manual Integration		



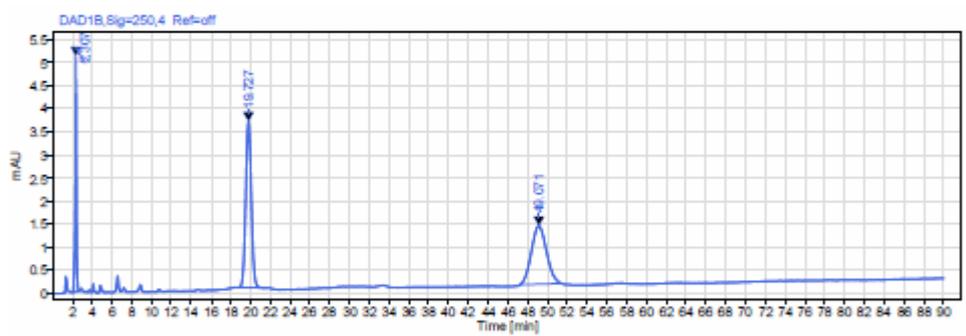
Signal: DAD1B,Sig=250,4 Ref-off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.298	VB	0.60	30.51	5.01	15.15	
20.452	MM m	0.44	41.71	1.11	20.70	
52.530	MM m	1.27	129.23	1.19	64.15	
	Sum		201.45			

Racemic Trace

Single Injection Report

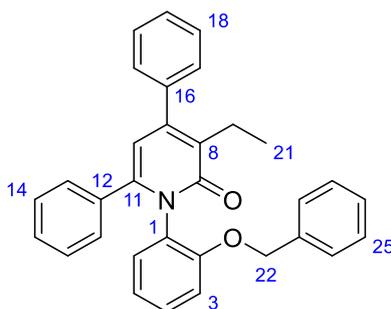
Data file:	JS #297 80C S9 102H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #297 80C S9 102H	Injection date:	2021-07-22 18:56:47+01:00
Instrument:	1220LC	Location:	63
Inj. volume:	5.000	Type:	Sample
Acq. method:	(90min) atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	Manual Integration		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.307	VB	0.53	38.58	5.11	12.30	
19.727	BB	2.30	140.96	3.61	44.94	
49.071	MM m	1.24	134.16	1.27	42.77	
	Sum		313.69			

1-(2-(Benzyloxy)phenyl)-3-ethyl-4,6-diphenylpyridin-2(1H)-one (4j)



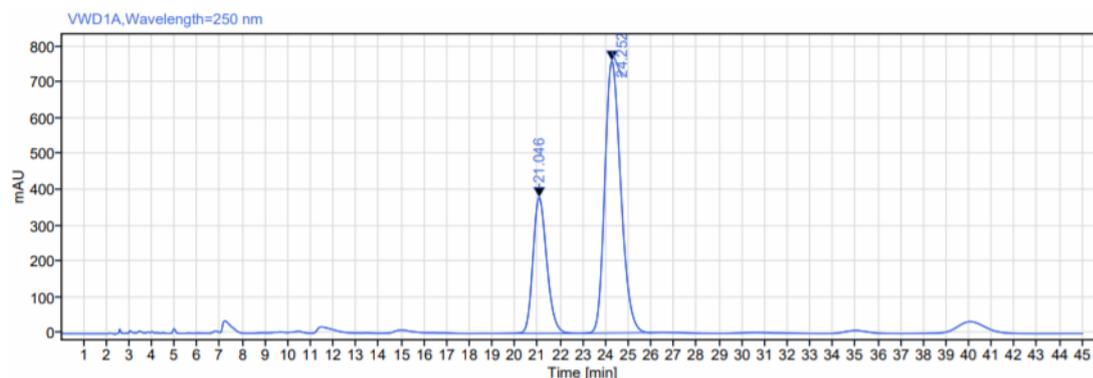
Conditions: HPLC: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 69.9 min, t_R (minor) = 30.2 min.

Asymmetric trace: 40% ee

Single Injection Report



Data file:	JS-4-291 col 2 60% MeCN-H2O20210330 115119.dx		
Sequence Name:	SingleSample	Project Name:	Walkup
Sample name:	JS-4-291 col 2 60% MeCN:H2O	Operator:	admin account
Instrument:	LCMS	Injection date:	2021-03-30 12:33:57+01:00
Inj. volume:	5.000	Location:	P1-a3
Acq. method:	60% MeCN in H2O OD-RH column 40 mins MS.amx	Type:	Sample
Processing method:	LCMS Purity.pmx	Sample amount:	0.00
Manually modified:	Manual Integration		



Signal: VWD1A,Wavelength=250 nm

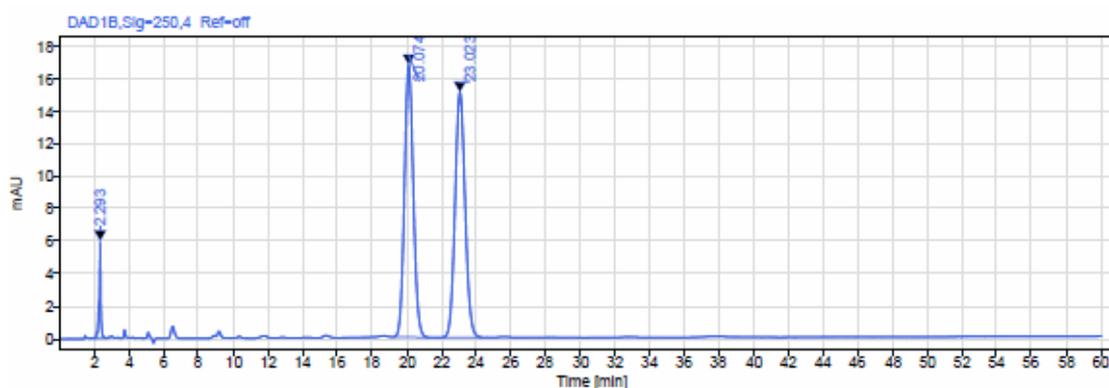
RT [min]	Type	Width [min]	Area	Height	Area%	Name
21.046	BB	3.08	16050.69	378.81	30.15	
24.252	BB	3.14	37179.46	758.83	69.85	
	Sum		53230.15			

Racemic trace

Single Injection Report



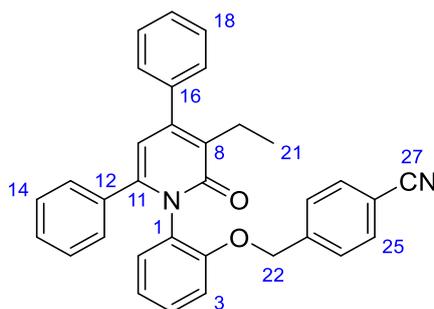
Data file:	JS #291 80C S10 54H (07.09).dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #291 80C S10 54H (07.09)	Injection date:	2021-07-09 19:39:03+01:00
Instrument:	1220LC	Location:	31
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmh		
Manually modified:	None		



Signal: DAD1B,Sig=250,4 Ref-off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.293	VB	0.59	35.16	6.04	2.84	
20.074	BB	1.97	595.66	16.83	48.06	
23.023	BB	2.26	608.50	15.18	49.10	
	Sum		1239.32			

4-((2-(3-Ethyl-2-oxo-4,6-diphenylpyridin-1(2H)-yl)phenoxy)methyl)benzonitrile (4k)



Conditions: CHIRALPAK OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = min, t_R (minor) = min.

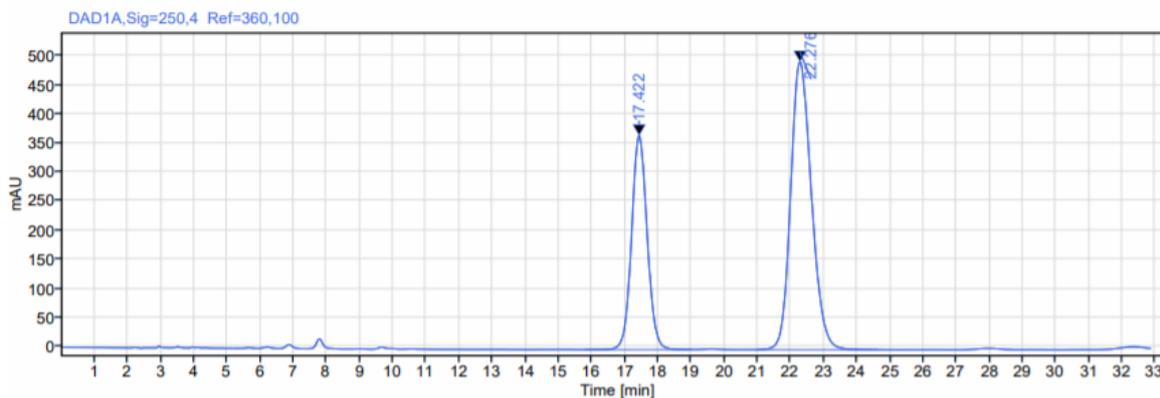
Asymmetric trace: 28% ee

Single Injection Report



Agilent Technologies

Data file:	JS-4-306 col 60% MeCN-H2O20210520 211751.dx		
Sequence Name:	SingleSample	Project Name:	JS
Sample name:	JS-4-306 col 60% MeCN/H2O	Operator:	SYSTEM
Instrument:	1100HPLC	Injection date:	2021-05-20 21:18:55+01:00
Inj. volume:	2.000	Location:	37
Acq. method:	60% MeCN-H2O 45 MIN.amx	Type:	Sample
Processing method:	3D UV Quantitative_DefaultMethod.pmx	Sample amount:	0.00
Manually modified:	Manual Integration		



Signal: DAD1A, Sig=250,4 Ref=360,100

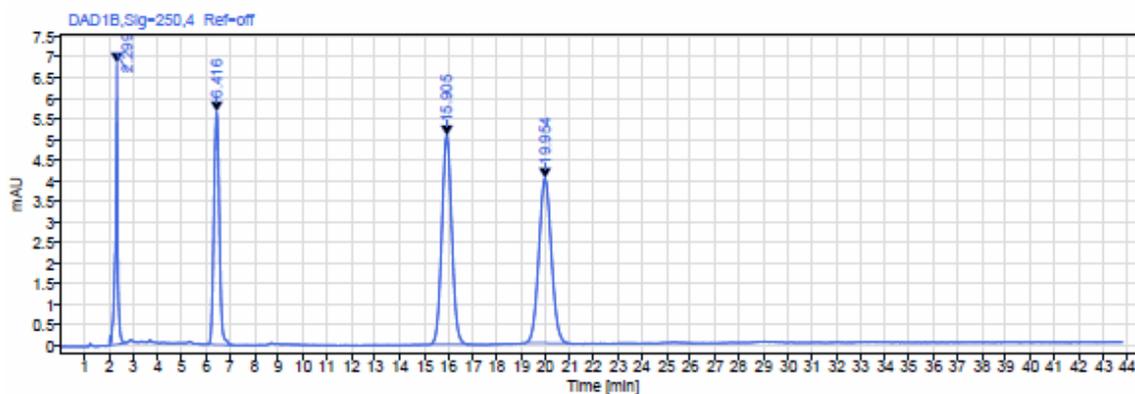
RT [min]	Type	Width [min]	Area	Height	Area%	Name
17.422	BB	3.23	12011.16	366.40	35.82	
22.276	BB	3.66	21520.34	494.41	64.18	
		Sum	33531.50			

Racemic trace*

Single Injection Report


Agilent Technologies

Data file:	JS #306 80C S8 96H (07.26).dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #306 80C S8 96H (07.26)	Injection date:	2021-07-26 15:51:27+01:00
Instrument:	1220LC	Location:	83
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	None		

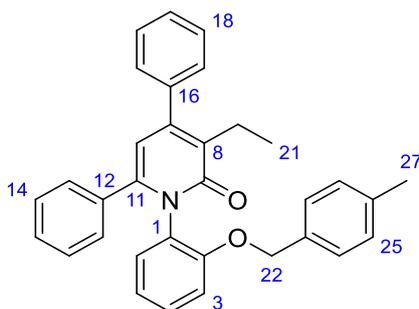


Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.299	VB	0.83	40.68	6.82	9.80	
6.416	BB	1.01	88.00	5.64	21.20	
15.905	BB	1.56	143.69	5.05	34.61	
19.954	BB	1.73	142.80	3.99	34.40	
		Sum	415.18			

* Peaks at 2.3 and 6.4 min are an impurity carried-over from a previous sample.

3-Ethyl-1-(2-((4-methylbenzyl)oxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4I)



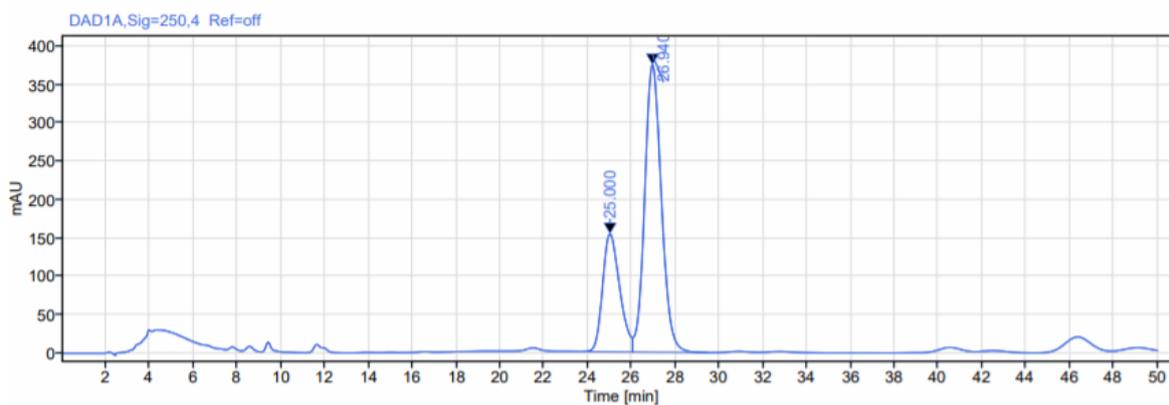
Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 26.9 min, t_R (minor) = 25.0 min.

Asymmetric trace: 41% ee

Single Injection Report



Data file:	JS-4-307 col 60% MeCN-H2O20210423 183713.dx		
Sequence Name:	SingleSample	Project Name:	JS
Sample name:	JS-4-307 col 60% MeCN/H2O	Operator:	SYSTEM
Instrument:	1100HPLC	Injection date:	2021-04-24 09:33:53+01:00
Inj. volume:	10.000	Location:	54
Acq. method:	60% MeCN-H2O 45 MIN.amx	Type:	Sample
Processing method:	3D UV Quantitative_DefaultMethod.pmx	Sample amount:	0.00
Manually modified:	Manual Integration		



Signal: DAD1A,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
25.000	BV	2.28	8345.52	153.68	29.57	
26.940	VB	3.42	19874.06	374.57	70.43	
	Sum		28219.58			

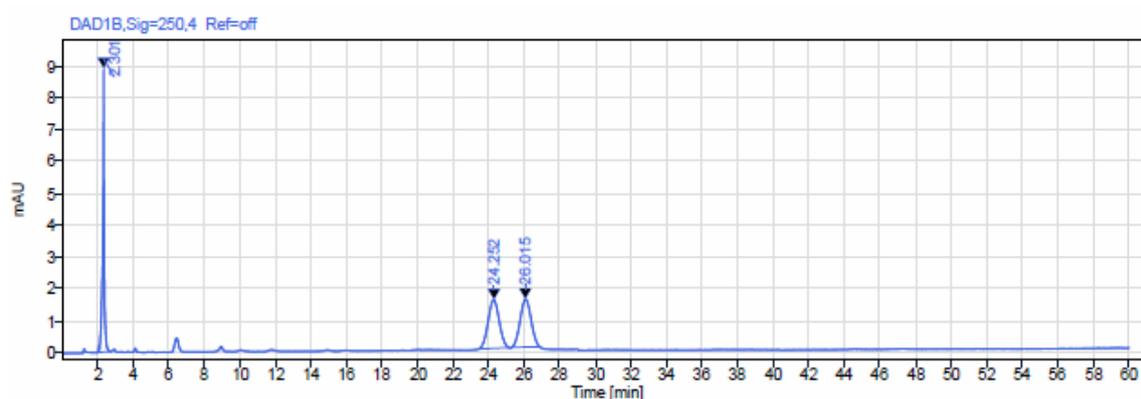
Racemic Trace

Single Injection Report



Agilent Technologies

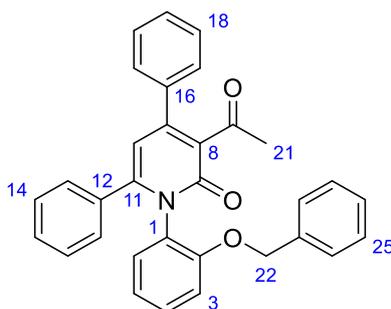
Data file:	JS #307 80C S8 96H (07.26).dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #307 80C S8 96H (07.26)	Injection date:	2021-07-26 16:36:16+01:00
Instrument:	1220LC	Location:	84
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmh		
Manually modified:	Manual Integration		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.301	VB	0.59	50.01	8.92	27.78	
24.252	MM m	0.51	65.80	1.52	36.55	
26.015	MM m	0.50	64.22	1.51	35.67	
	Sum		180.04			

3-Acetyl-1-(2-(benzyloxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4m)

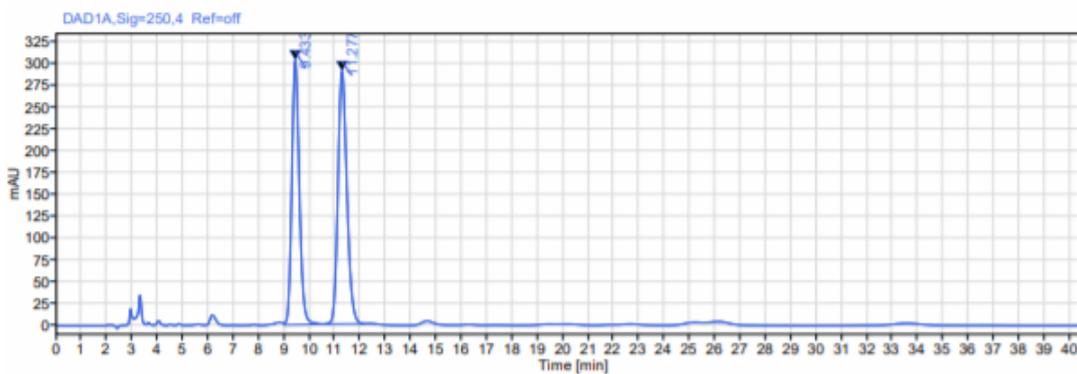


Conditions: HPLC: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 11.3 min, t_R (minor) = 9.4 min.

Asymmetric trace: 6% ee

Single Injection Report

Data file: JS-4-290 col 60% MeCN-H2O20210329 164450.dx
Sequence Name: SingleSample **Project Name:** JS
Sample name: JS-4-290 col 60% MeCN/H2O **Operator:** SYSTEM
Instrument: 1100HPLC **Injection date:** 2021-03-29 16:46:43+01:00
Inj. volume: 10.000 **Location:** 13
Acq. method: 60% MeCN-H2O 45 MIN.amx **Type:** Sample
Processing method: 3D UV Quantitative_DefaultMethod.pmx **Sample amount:** 0.00
Manually modified: Manual Integration



Signal: DAD1A,Sig=250,4 Ref=off

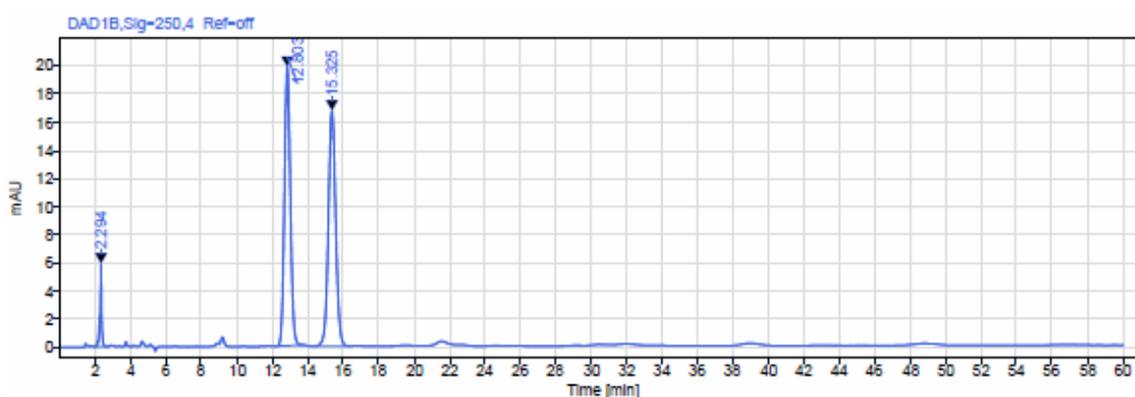
RT [min]	Type	Width [min]	Area	Height	Area%	Name
9.433	VB	1.50	6239.89	302.83	46.76	
11.277	BB	1.63	7104.91	289.41	53.24	
	Sum		13344.80			

Racemic trace

Single Injection Report



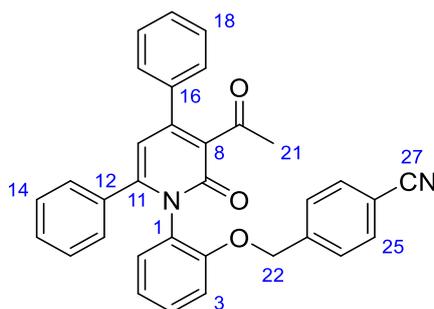
Data file:	JS #290 S11 76H (07.09).dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #290 S11 76H (07.09)	Injection date:	2021-07-09 18:38:04+01:00
Instrument:	1220LC	Location:	6
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	None		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.294	VB	0.48	34.18	5.91	3.52	
12.803	BB	1.44	480.14	19.85	47.35	
15.325	BB	1.81	477.52	16.76	49.14	
		Sum	971.83			

4-((2-(3-Acetyl-2-oxo-4,6-diphenylpyridin-1(2H)-yl)phenoxy)methyl)benzonitrile (4n)



Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 13.7 min, t_R (minor) = 10.9 min.

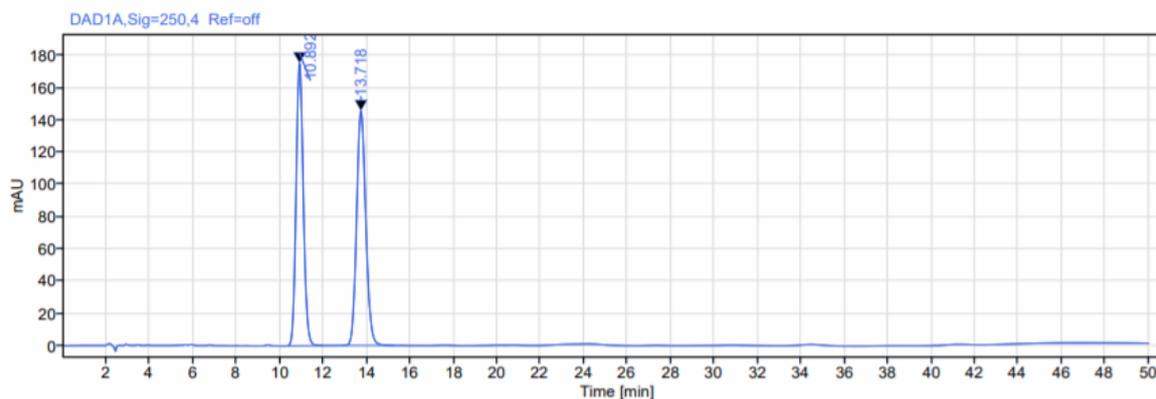
Asymmetric trace: 3% ee

Single Injection Report



Agilent Technologies

Data file:	JS-4-304 col 60% MeCN-H2O20210422 135035.dx		
Sequence Name:	SingleSample	Project Name:	JS
Sample name:	JS-4-304 col 60% MeCN/H2O	Operator:	SYSTEM
Instrument:	1100HPLC	Injection date:	2021-04-22 15:34:32+01:00
Inj. volume:	10.000	Location:	83
Acq. method:	60% MeCN-H2O 45 MIN.amx	Type:	Sample
Processing method:	3D UV Quantitative_DefaultMethod.pmx	Sample amount:	0.00
Manually modified:	Manual Integration		



Signal: DAD1A,Sig=250,4 Ref=off

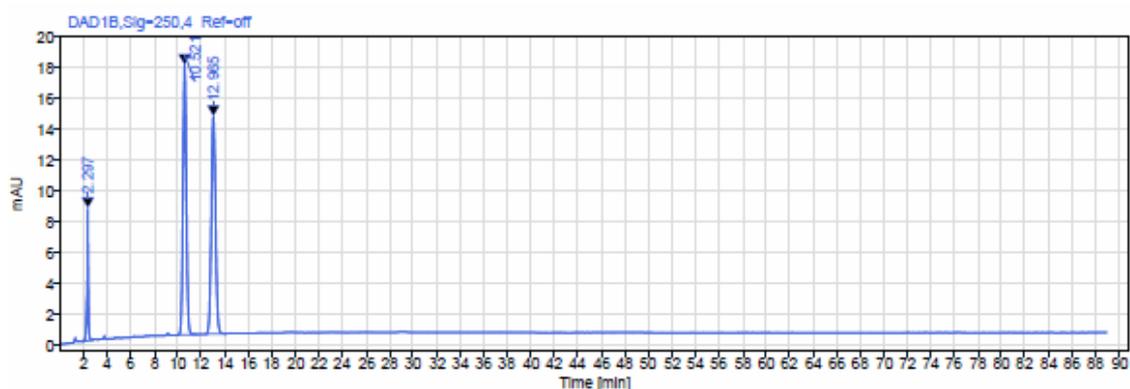
RT [min]	Type	Width [min]	Area	Height	Area%	Name
10.892	BB	1.72	4046.90	175.25	48.65	
13.718	BB	2.30	4272.14	145.40	51.35	
		Sum	8319.04			

Racemic trace

Single Injection Report



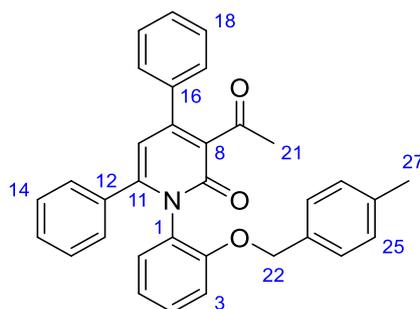
Data file:	JS #304 80C S8 96H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #304 80C S8 96H	Injection date:	2021-07-26 13:39:25+01:00
Instrument:	1220LC	Location:	81
Inj. volume:	5.000	Type:	Sample
Acq. method:	(90min) atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmw		
Manually modified:	None		



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.297	BB	0.73	54.92	8.63	7.35	
10.521	BB	1.60	345.12	17.55	46.22	
12.965	BB	1.74	346.71	14.17	46.43	
		Sum	746.76			

3-Acetyl-1-(2-((4-methylbenzyl)oxy)phenyl)-4,6-diphenylpyridin-2(1H)-one (4o)



Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 17.5 min, t_R (minor) = 15.6 min.

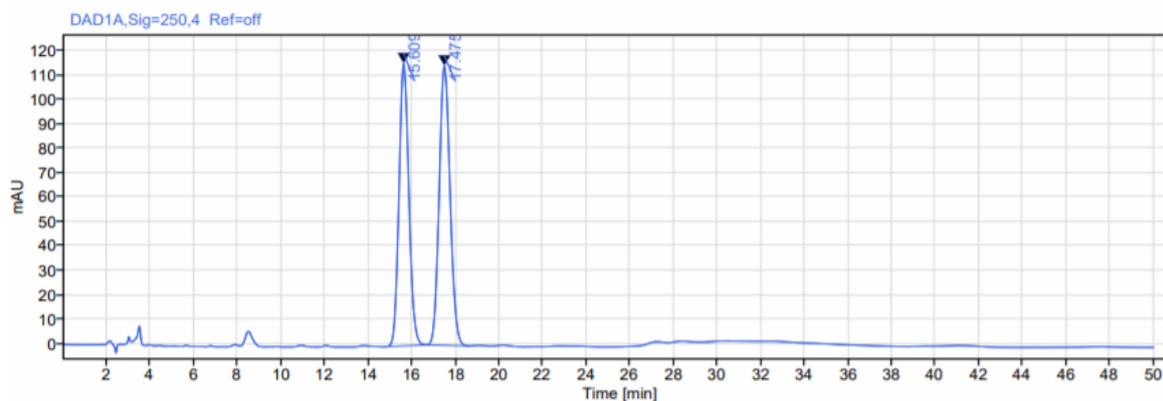
Asymmetric trace: 5% ee

Single Injection Report



Agilent Technologies

Data file: JS-4-305 col 60% MeCN-H2O20210422 135043.dx
Sequence Name: SingleSample **Project Name:** JS
Sample name: JS-4-305 col 60% MeCN/H2O **Operator:** SYSTEM
Instrument: 1100HPLC **Injection date:** 2021-04-22 16:25:40+01:00
Inj. volume: 10.000 **Location:** 84
Acq. method: 60% MeCN-H2O 45 MIN.amx **Type:** Sample
Processing method: 3D UV Quantitative_DefaultMethod.pmx **Sample amount:** 0.00
Manually modified: Manual Integration



Signal: DAD1A,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
15.609	BB	1.89	3621.20	115.52	47.71	
17.475	BB	1.95	3968.93	114.33	52.29	
	Sum		7590.13			

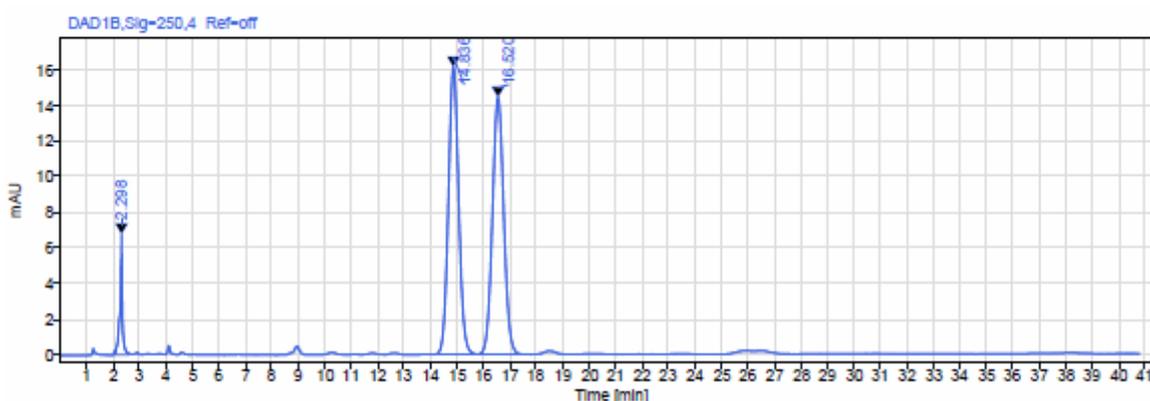
Racemic Trace

Single Injection Report



Agilent Technologies

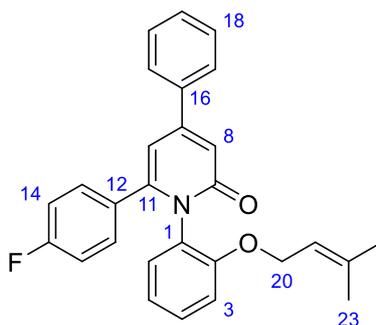
Data file: JS #305 80C S8 96H (07.26).dx
Sequence Name: SingleSample **Project Name:** Agilent
Sample name: JS #305 80C S8 96H (07.26) **Operator:** Agilent
Instrument: 1220LC **Injection date:** 2021-07-26 15:09:40+01:00
Inj. volume: 5.000 **Location:** 82
Acq. method: Atropisomer barrier measurement.amx **Type:** Sample
Processing method: 3D UV Quantitative_DefaultMethod.pmx **Sample amount:** 1.00
Manually modified: None



Signal: DAD1B,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.298	VB	0.67	40.74	6.78	4.52	
14.836	BB	1.56	432.90	16.11	47.99	
16.520	BB	1.65	428.50	14.42	47.50	
		Sum	902.14			

6-(4-Fluorophenyl)-1-(2-((3-methylbut-2-en-1-yl)oxy)phenyl)-4-phenylpyridin-2(1H)-one (4p)



Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 29.5 min, t_R (minor) = 13.7 min.

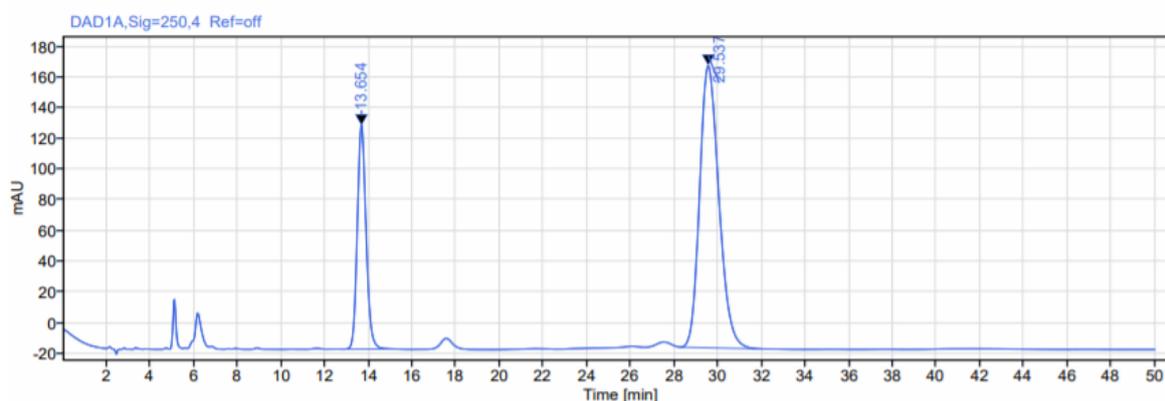
Asymmetric trace: 47% ee

Single Injection Report



Agilent Technologies

Data file: JS-5-310 col 60% MeCN-H2O20210430 153951.dx
Sequence Name: SingleSample **Project Name:** JS
Sample name: JS-5-310 col 60% MeCN/H2O **Operator:** SYSTEM
Instrument: 1100HPLC **Injection date:** 2021-04-30 16:40:37+01:00
Inj. volume: 10.000 **Location:** 16
Acq. method: 60% MeCN-H2O 45 MIN.amx **Type:** Sample
Processing method: 3D UV Quantitative_DefaultMethod.pmx **Sample amount:** 0.00
Manually modified: Manual Integration



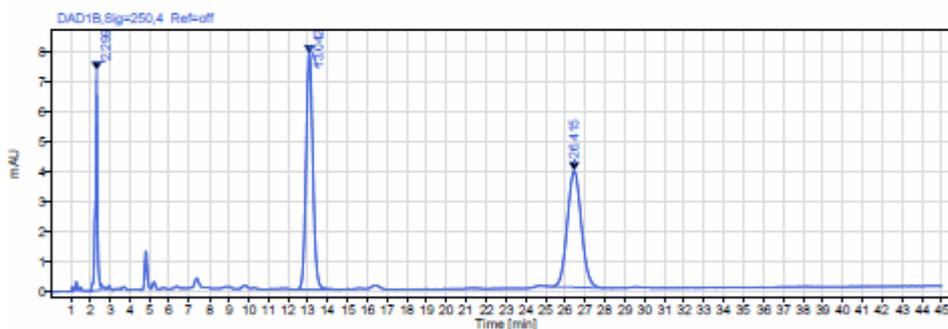
Signal: DAD1A,Sig=250,4 Ref=off

RT [min]	Type	Width [min]	Area	Height	Area%	Name
13.654	BB	2.22	4059.51	145.79	26.52	
29.537	BB	3.78	11249.32	183.73	73.48	
		Sum	15308.84			

Racemic Trace

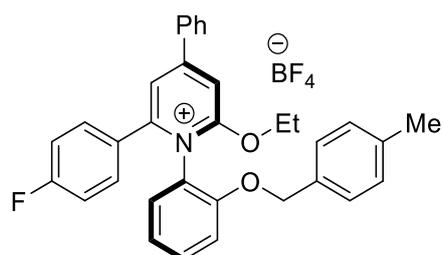
Single Injection Report

Data file:	JS #310 80C S9 70.5H.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	JS #310 80C S9 70.5H	Injection date:	2021-07-31 23:03:26+01:00
Instrument:	1220LC	Location:	56
Inj. volume:	5.000	Type:	Sample
Acq. method:	(45min) Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	None		



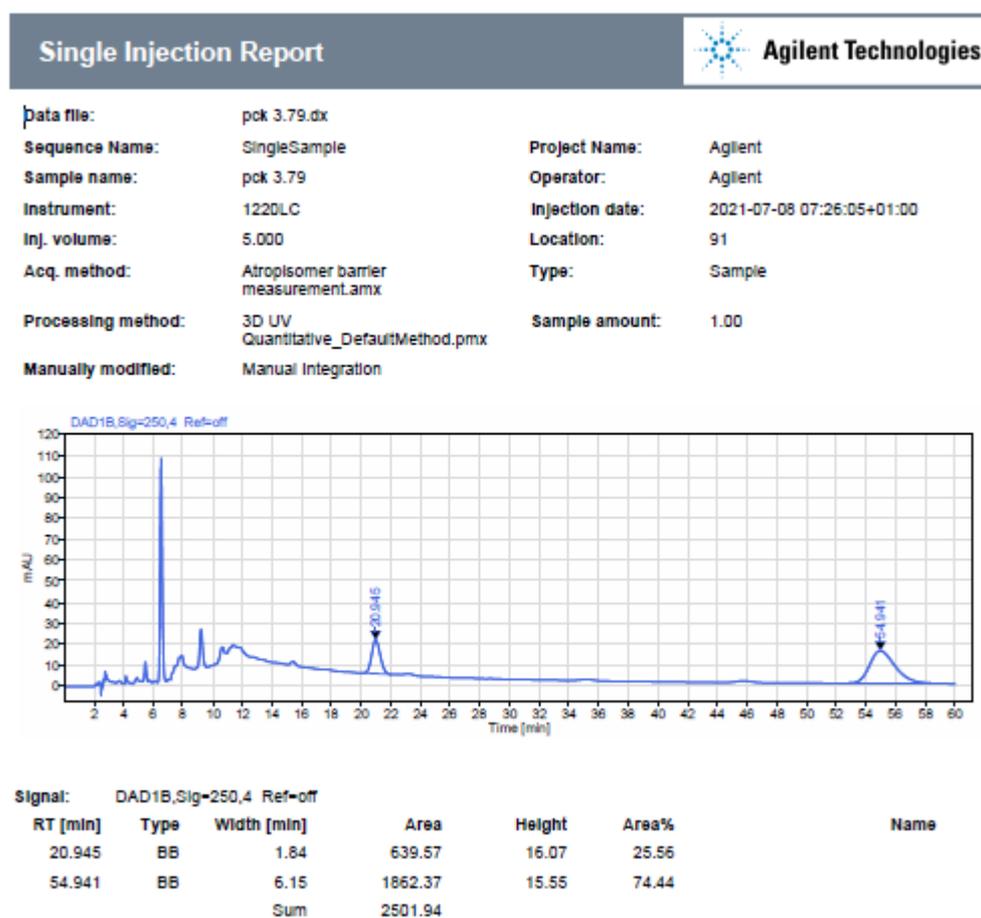
Signal: DAD1B,Sig=250,4 Ref-off						
RT [min]	Type	Width [min]	Area	Height	Area%	Name
2.299	BV	0.51	48.37	7.34	11.30	
13.042	BB	1.71	188.50	7.86	44.03	
26.415	BB	2.63	191.24	3.88	44.67	
	Sum		428.11			

7.2 Pyridinium salt (7)



Conditions: Chiralpak OD-RH, 60% MeCN/H₂O, 0.75 mL/min, λ = 250 nm, t_R (major) = 54.9 min, t_R (minor) = 20.9 min.

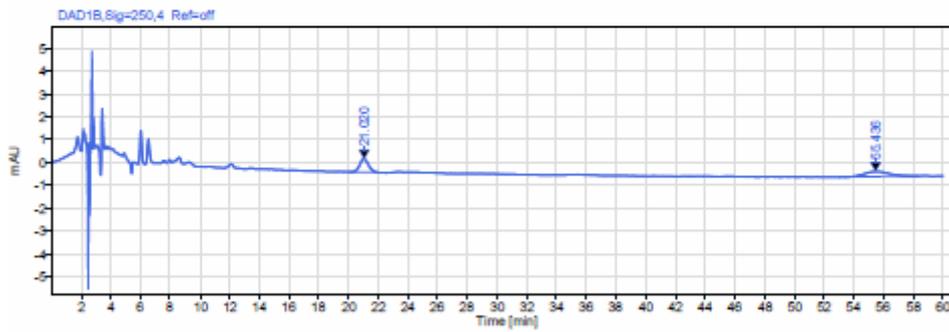
Asymmetric trace: 49% ee



Racemic Trace

Single Injection Report Agilent Technologies

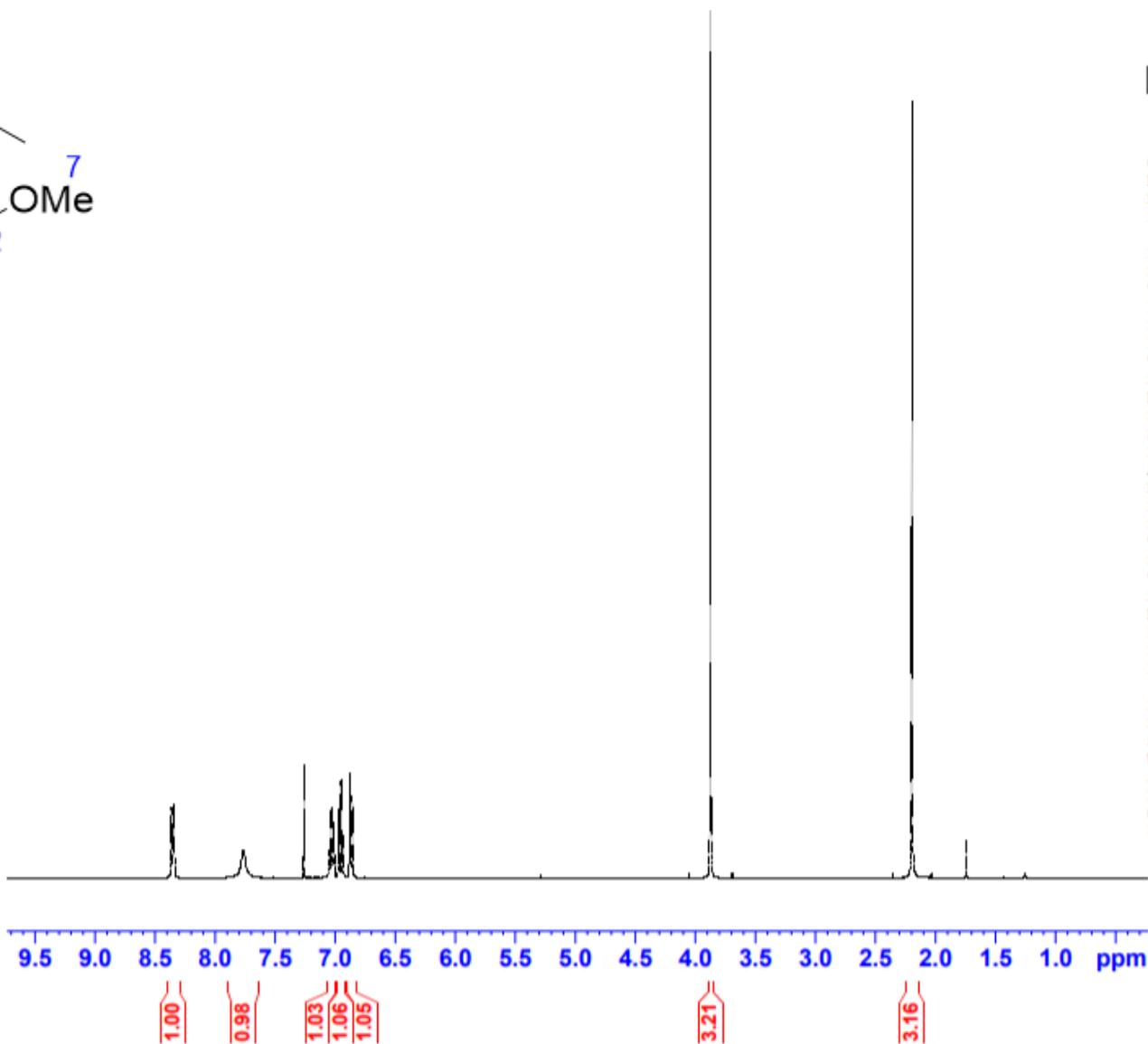
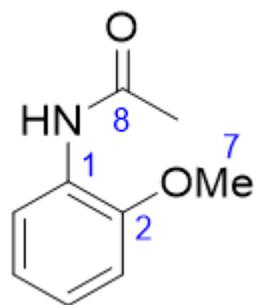
Data file:	pck 3.80.dx	Project Name:	Agilent
Sequence Name:	SingleSample	Operator:	Agilent
Sample name:	pck 3.80	Injection date:	2021-07-08 08:27:03+01:00
Instrument:	1220LC	Location:	92
Inj. volume:	5.000	Type:	Sample
Acq. method:	Atropisomer barrier measurement.amx	Sample amount:	1.00
Processing method:	3D UV Quantitative_DefaultMethod.pmx		
Manually modified:	Manual Integration		



Signal: DAD1B,Sig=250,4 Ref=off						
RT [min]	Type	Width [min]	Area	Height	Area%	Name
21.020	MM m	0.49	24.36	0.59	49.70	
55.436	MM m	1.29	24.65	0.22	50.30	
		Sum	49.01			

8 NMR data

1a - ¹H NMR, 400 MHz, CDCl₃

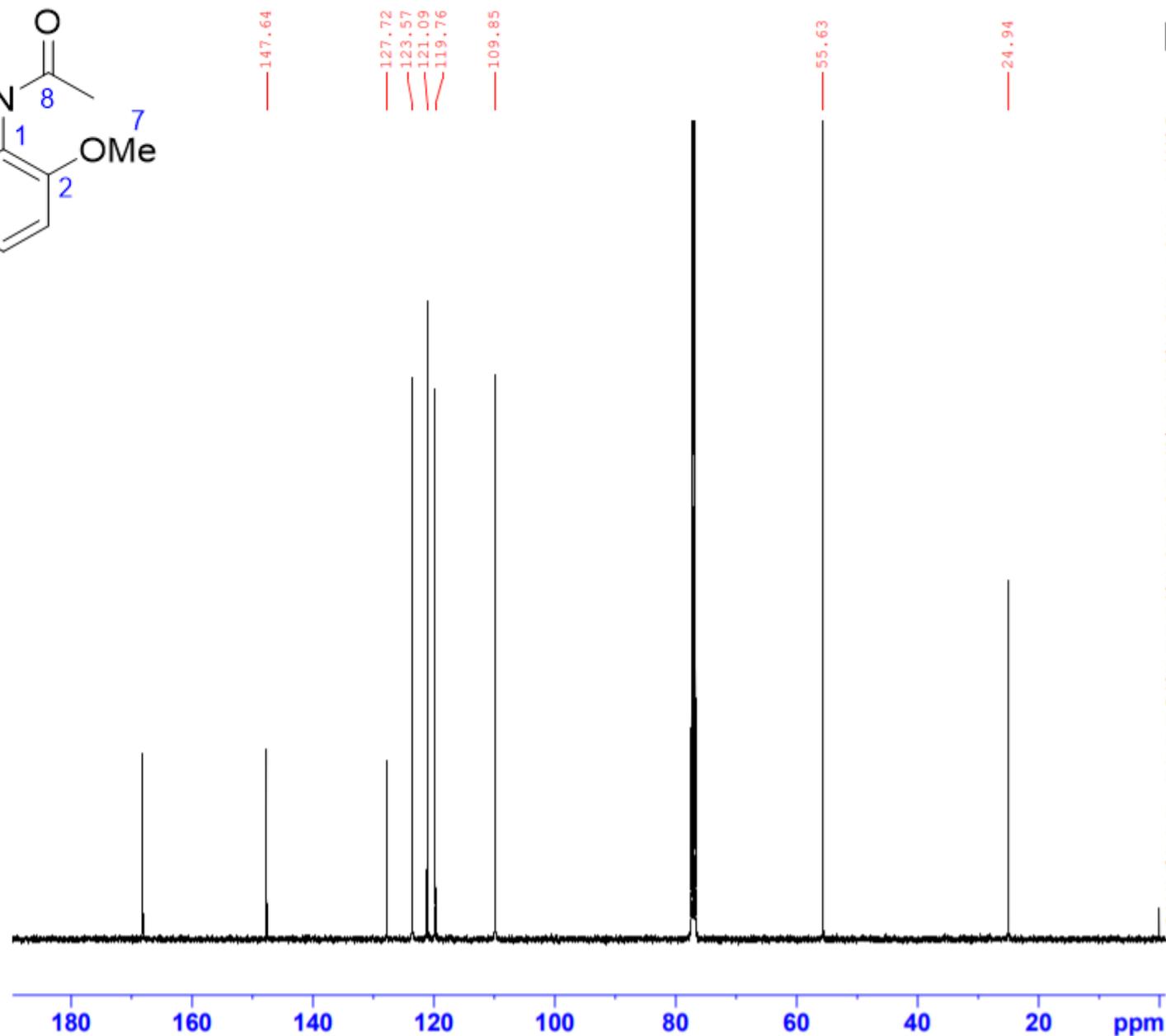
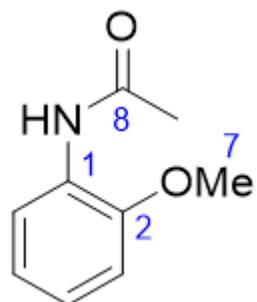


Current Data Parameters
NAME JS-3-180
EXPNO 14
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210506
Time_ 23.30 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 144
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100098 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1a - ¹³C NMR, 101 MHz, CDCl₃



Current Data Parameters
NAME JS-3-180
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters

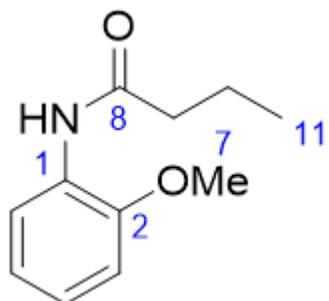
Date_ 20210506
Time_ 23.11 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters

SI 131072
SF 100.6077407 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

S109

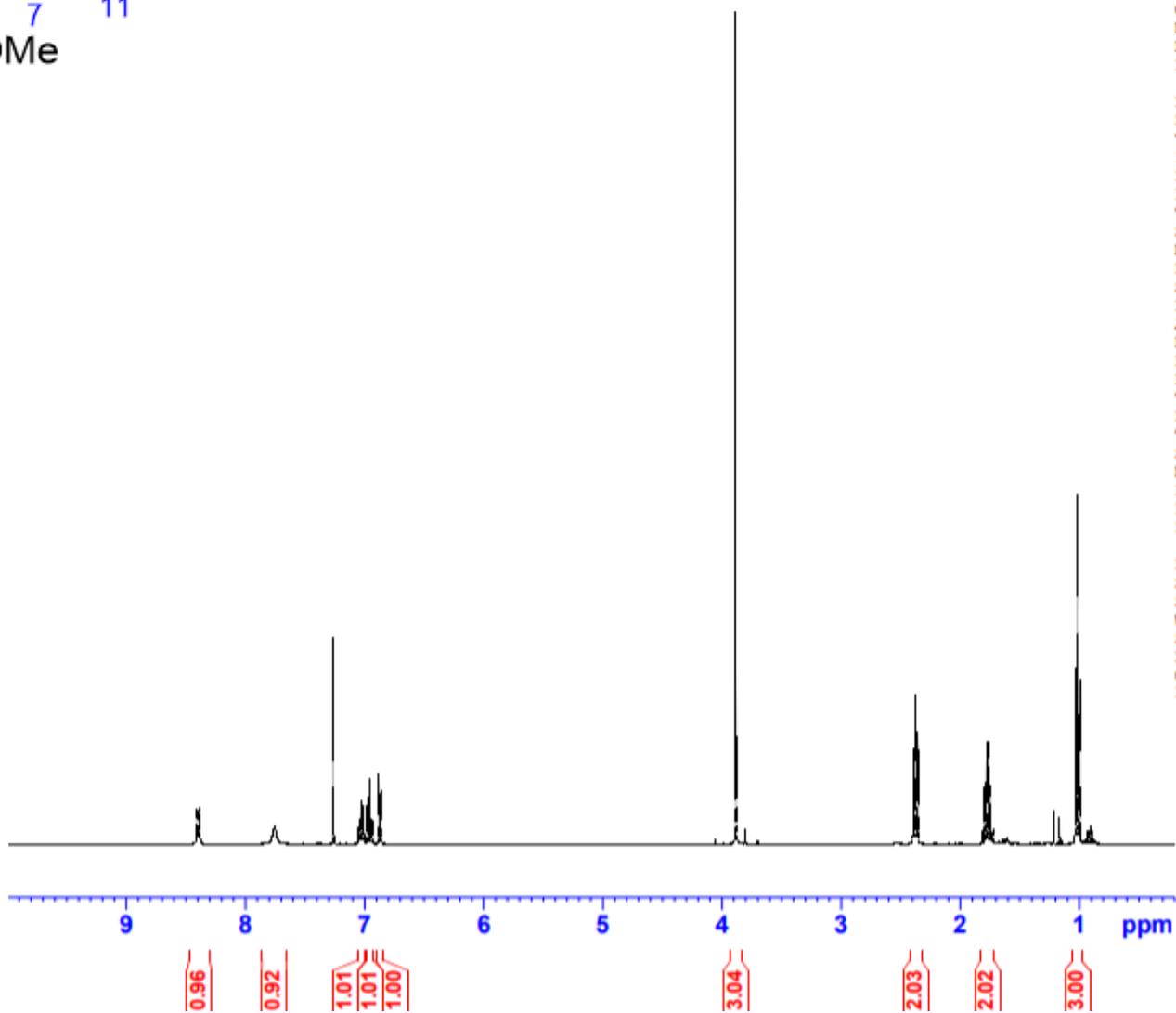
1b - ¹H NMR, 400 MHz, CDCl₃



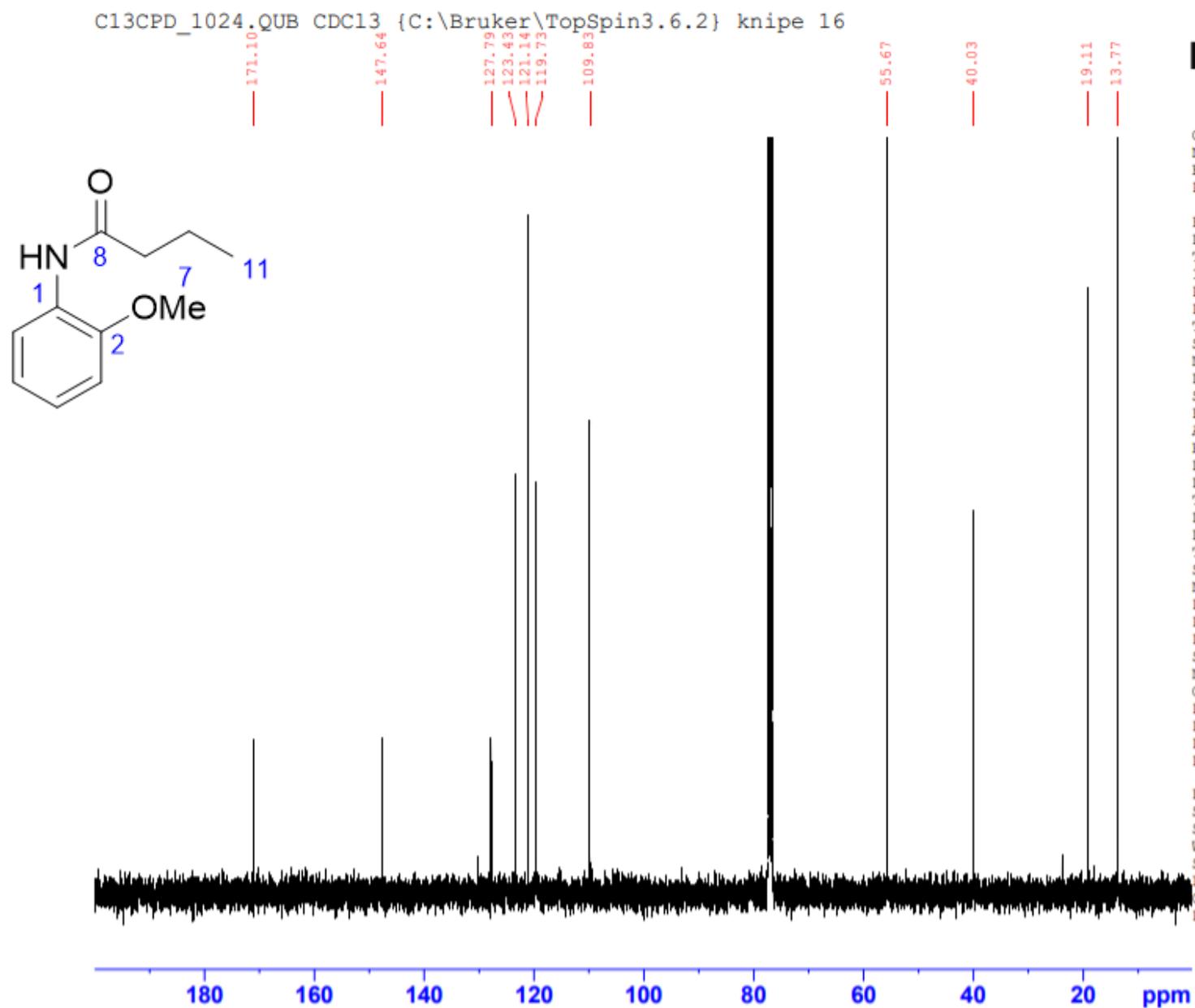
Current Data Parameters
NAME JS-4-272
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210304
Time_ 11.43 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (zg30)
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TDO 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100099 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



1b - ¹³C NMR, 101 MHz, CDCl₃

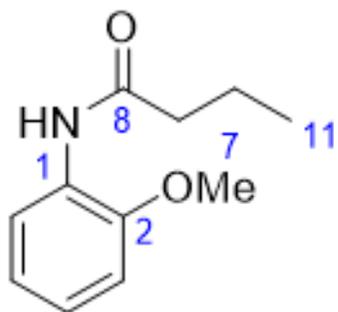


Current Data Parameters
NAME JS-4-272
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210304
Time_ 14.36 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl₃
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

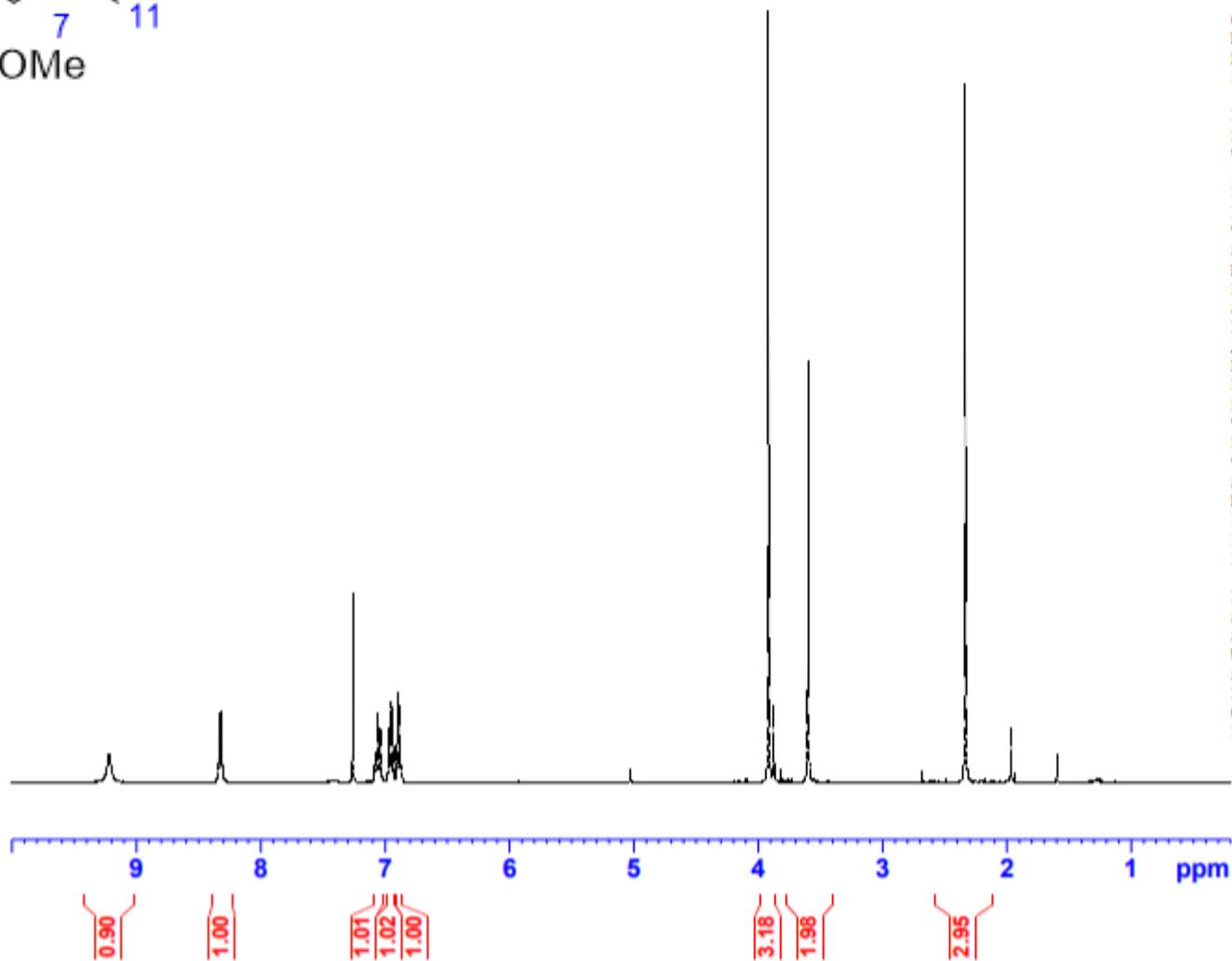
1c - ¹H NMR, 400 MHz, CDCl₃



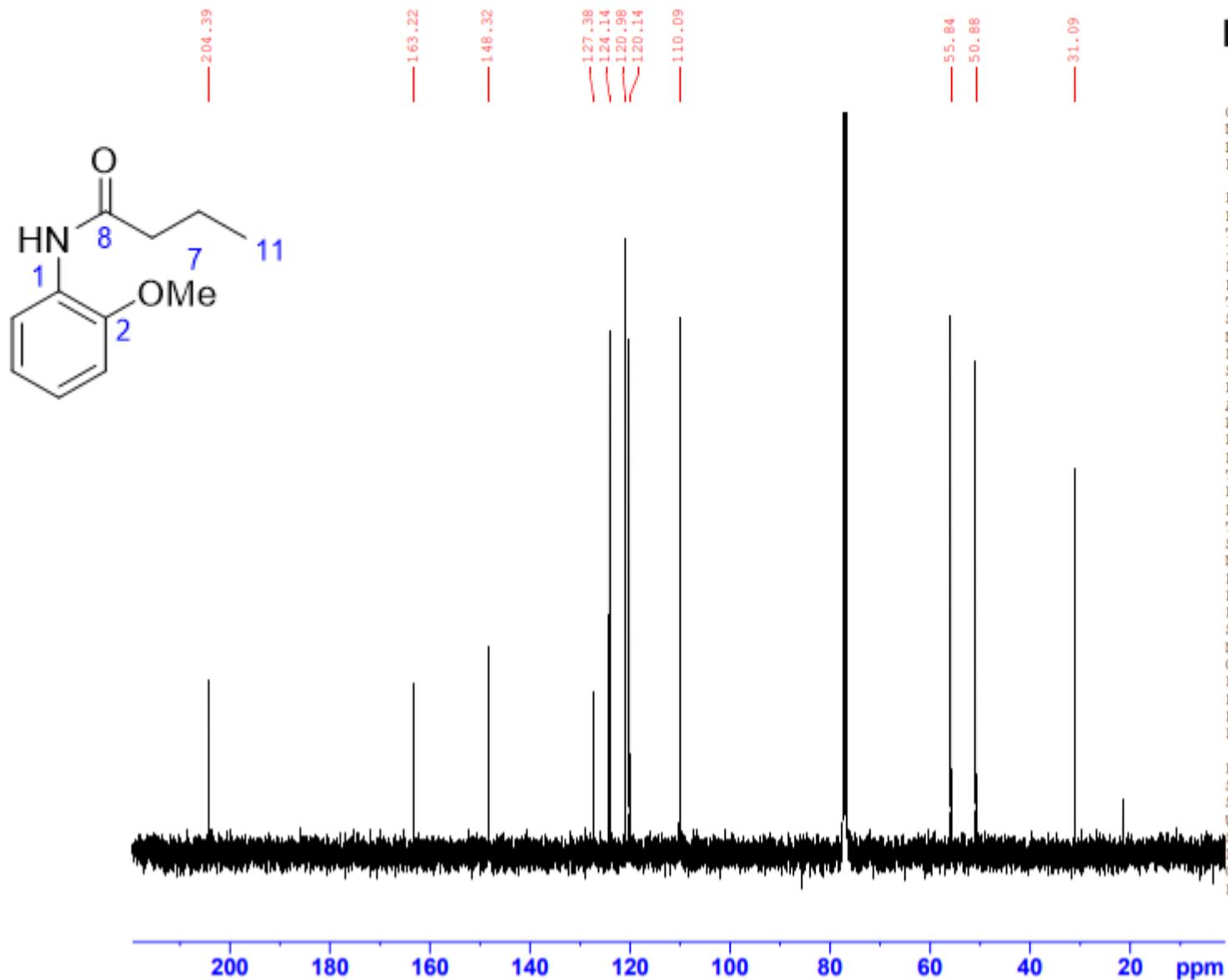
Current Data Parameters
NAME PCK-3-69
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210304
Time_ 16.07 h
INSTRUM AVIII 400
PROBHD z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 287
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



1c - ¹³C NMR, 101 MHz, CDCl₃



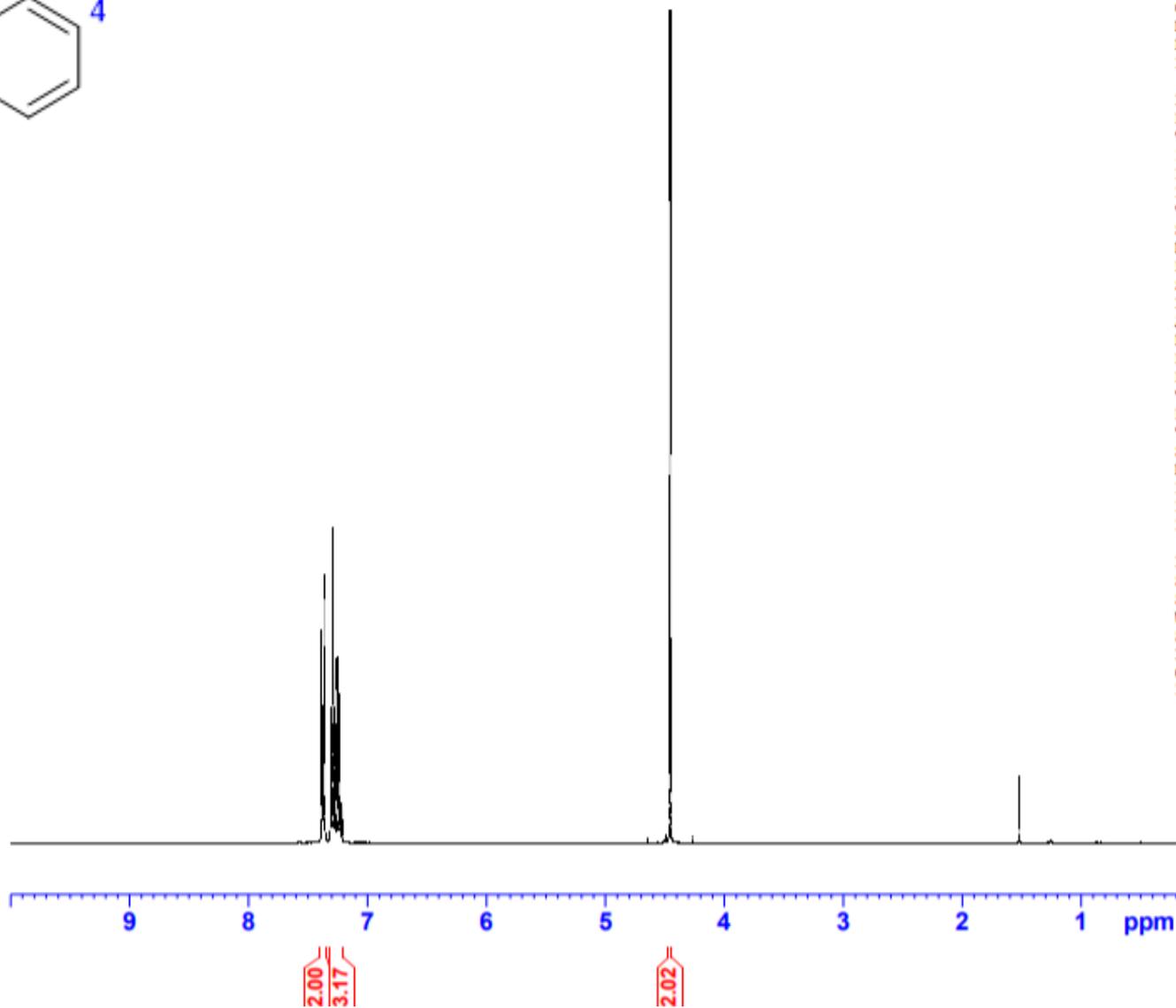
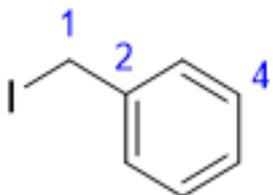
Current Data Parameters
NAME PCK-3-69
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210304
Time_ 16.02 h
INSTRUM AVIII 400
PROBHD Z108618_0146 ()
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

S113

S1 - ¹H NMR, 400 MHz, CDCl₃

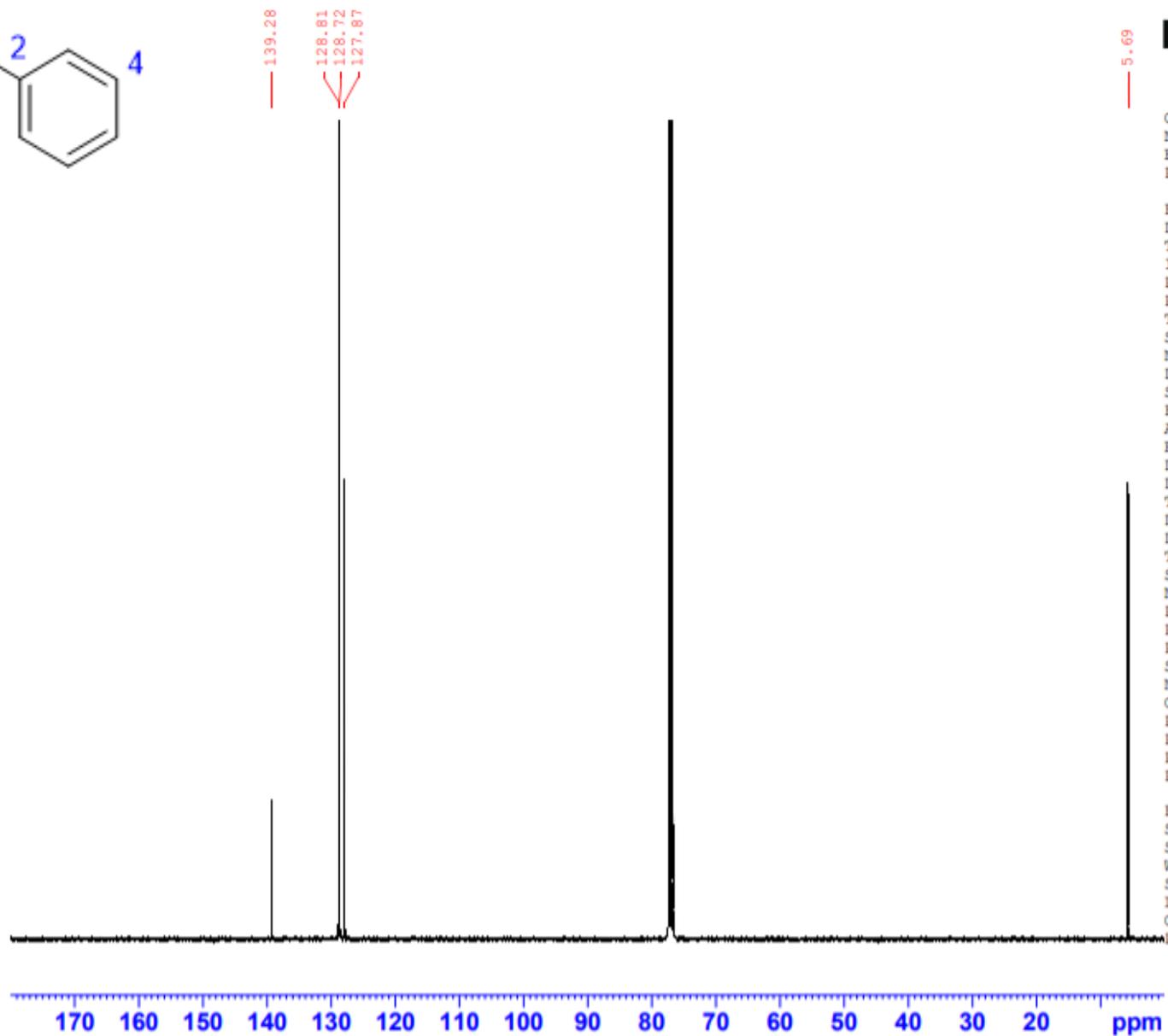
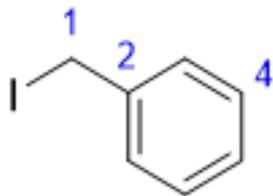


Current Data Parameters
NAME JS-4-294 recol
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210505
Time_ 14.38 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100135 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

S1 - ¹³C NMR, 101 MHz, CDCl₃

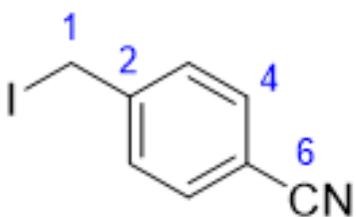


Current Data Parameters
NAME JS-4-294 recol
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210505
Time_ 23.11 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077428 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

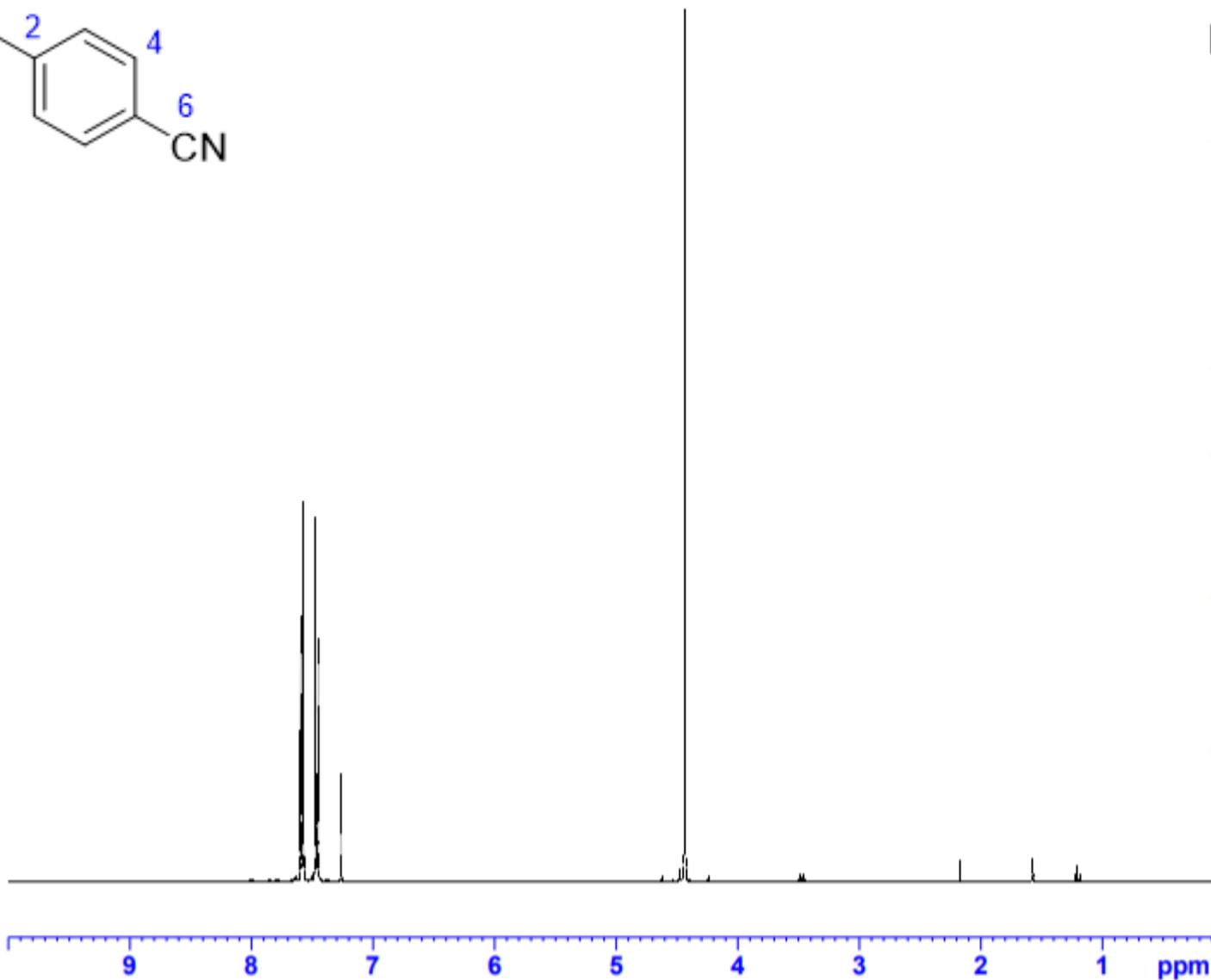
S2 - ¹H NMR, 400 MHz, CDCl₃



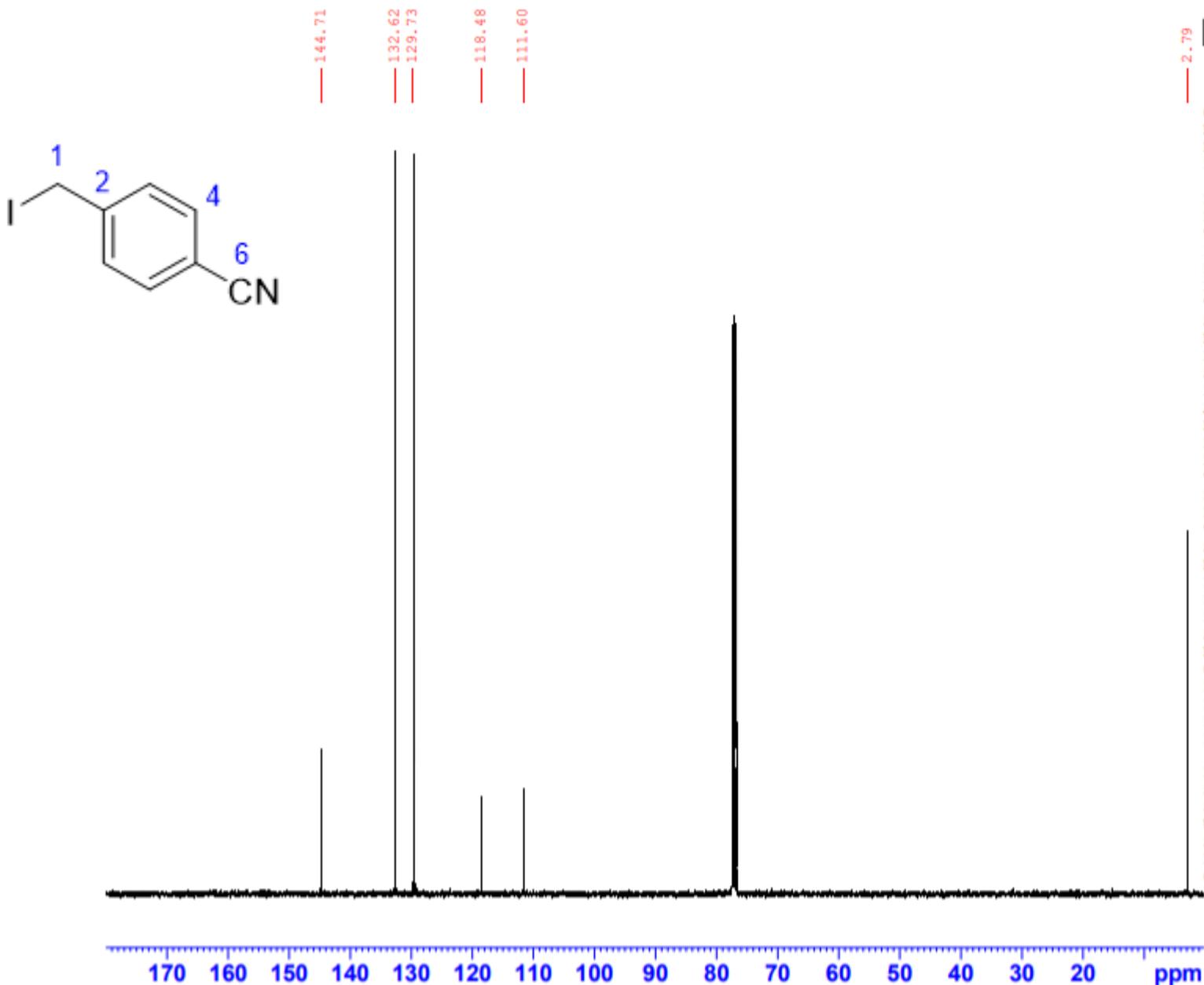
Current Data Parameters
NAME PCK-3-70
EXPNO 13
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210504
Time 21.57 h
INSTRUM AVIII 400
PROBHD z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 228
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TDO 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



S2 - ¹³C NMR, 101 MHz, CDCl₃

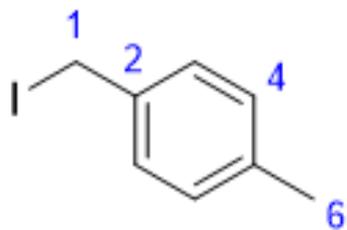


Current Data Parameters
NAME PCK-3-70
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210504
Time 21.39 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

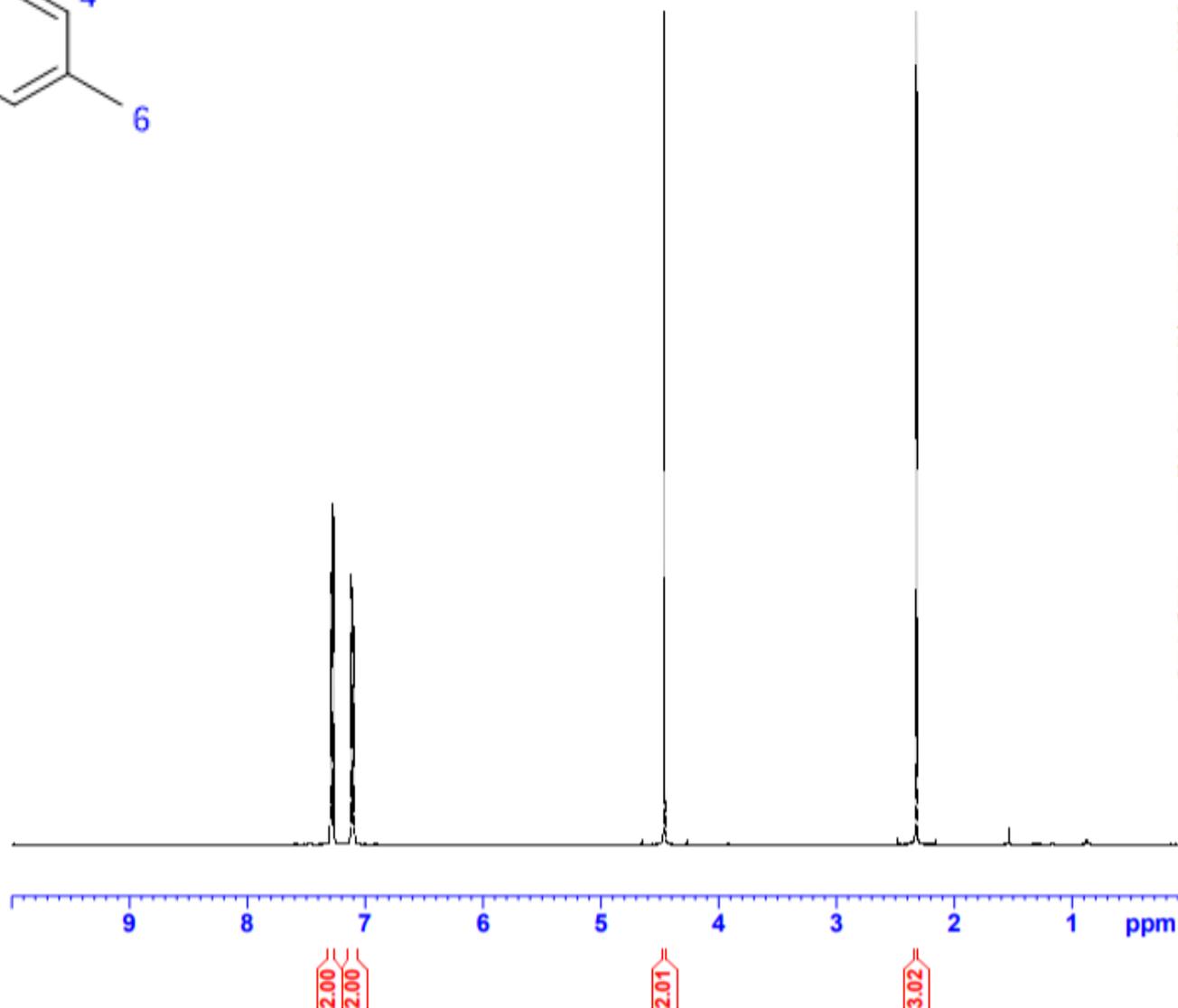
S3 - ¹H NMR, 400 MHz, CDCl₃



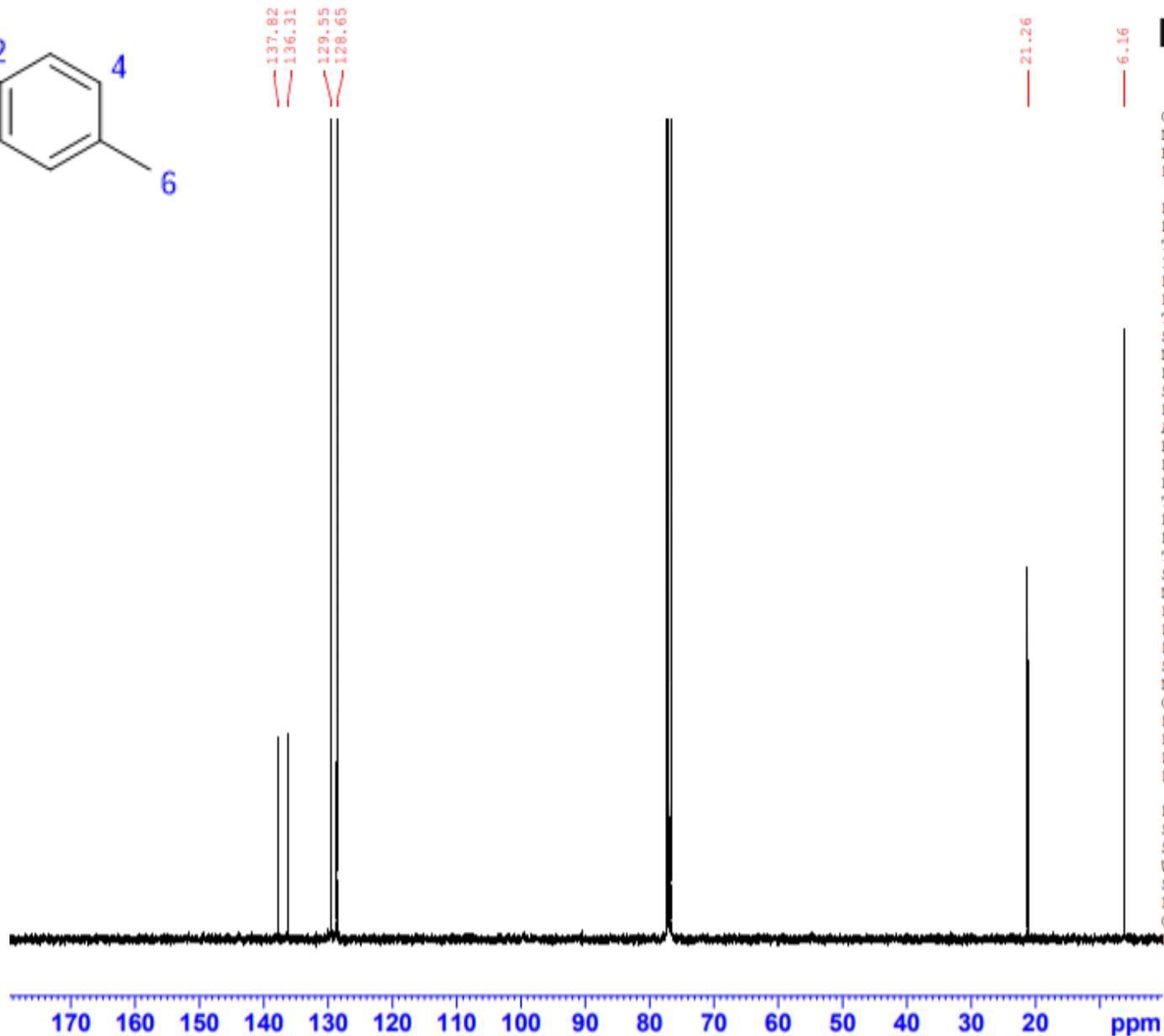
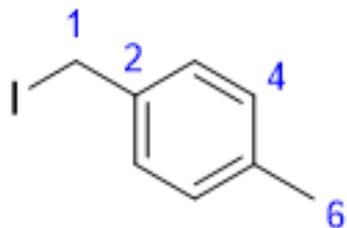
Current Data Parameters
NAME PCK-3-71
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210504
Time_ 16.25 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 161
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



S3 - ¹³C NMR, 101 MHz, CDCl₃

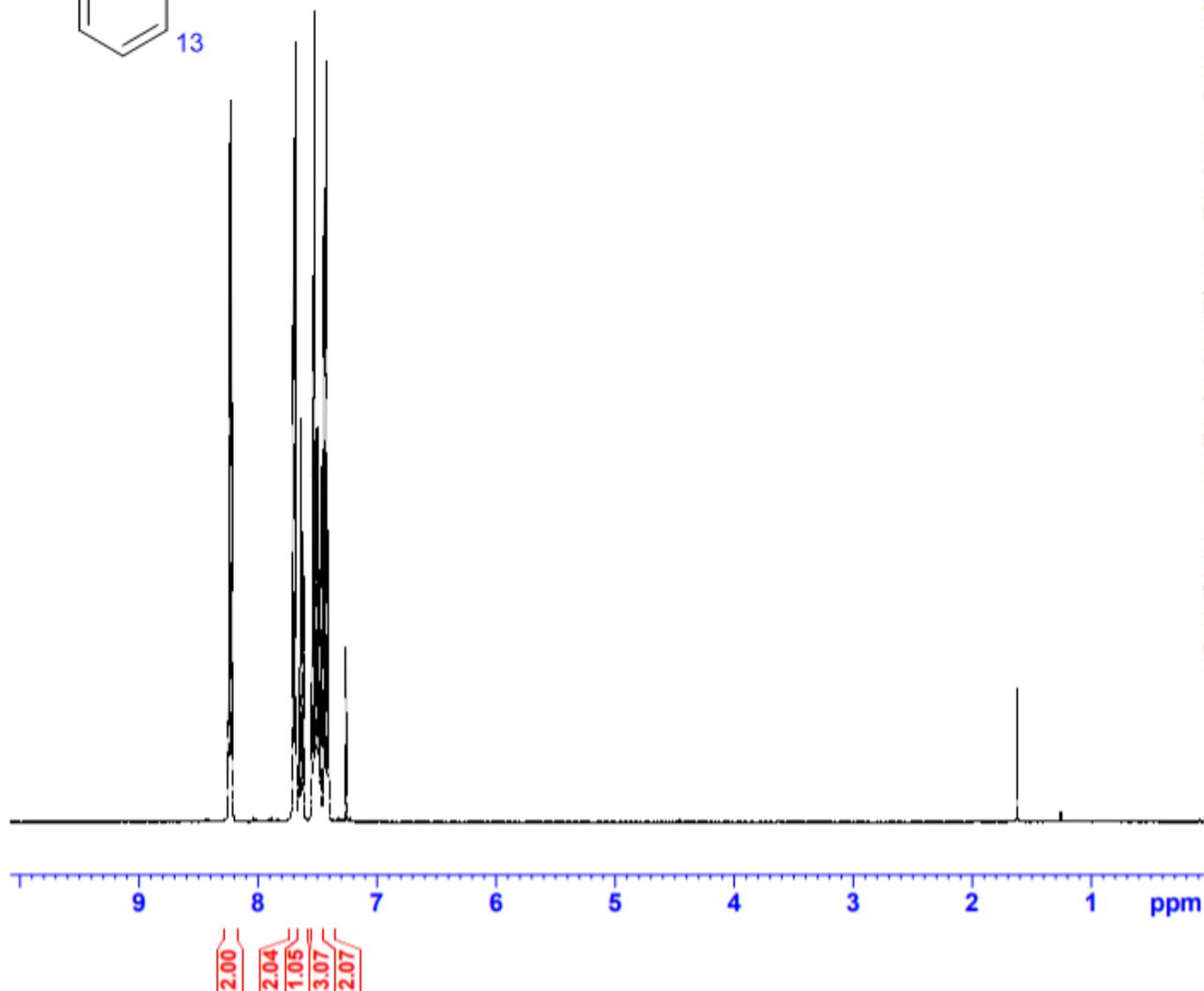
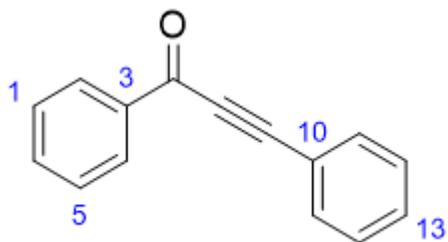


Current Data Parameters
NAME PCK-3-71
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date 20210504
Time 23.05 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

2a - ¹H NMR, 400 MHz, CDCl₃

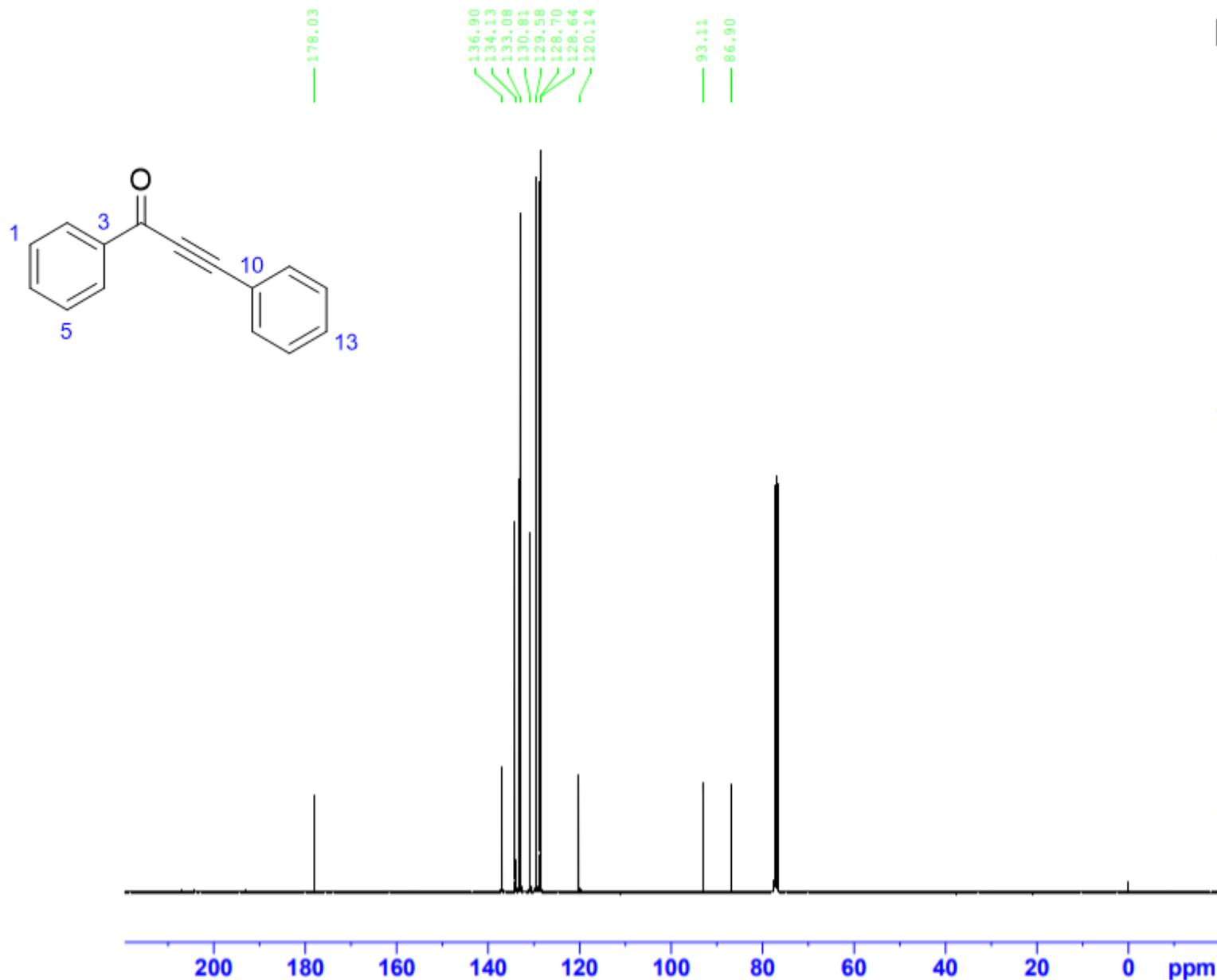


Current Data Parameters
NAME JS-4-243 recol
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210505
Time 16.17 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (zg30)
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 128
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100099 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

2a - ¹³C NMR, 101 MHz, CDCl₃

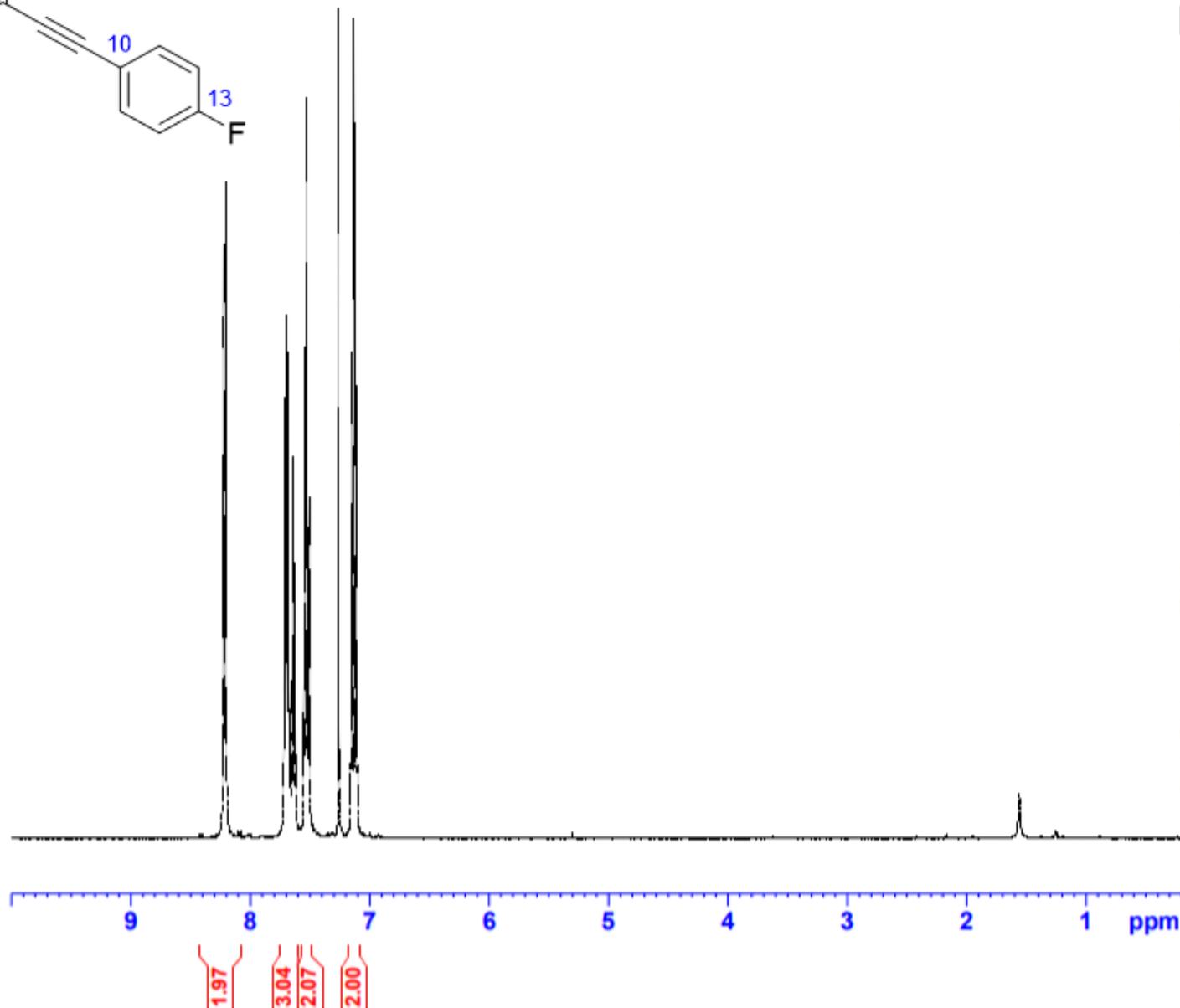
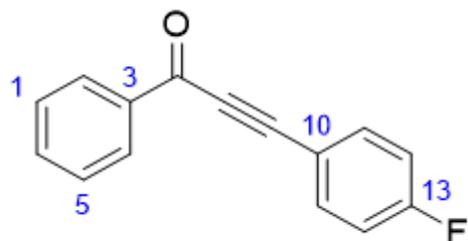


Current Data Parameters
NAME JS-4-243 recol
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210506
Time 3.17 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077420 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

2b - ¹H NMR, 400 MHz, CDCl₃



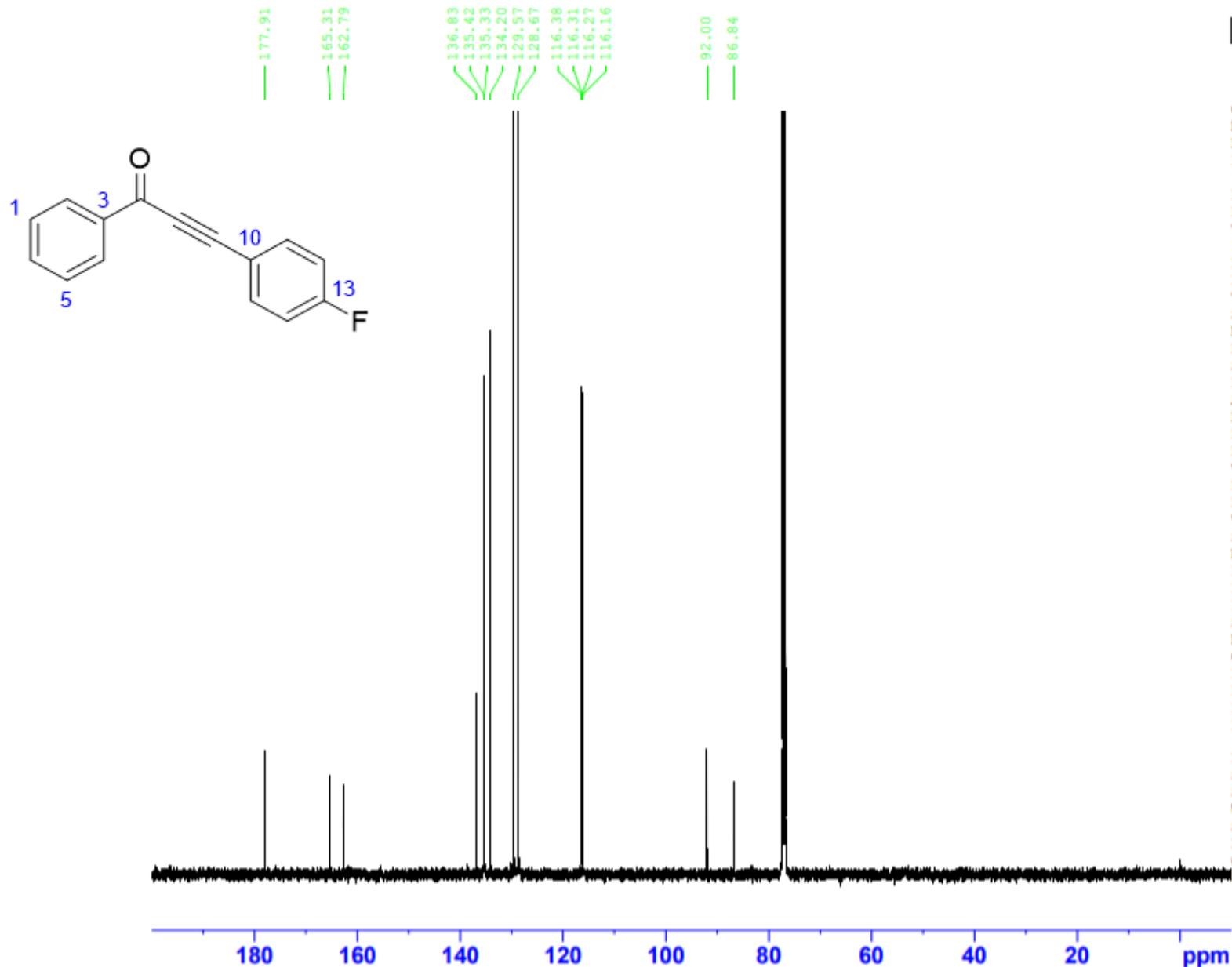
Current Data Parameters
NAME PCK-3-68
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210309
Time_ 20.01
INSTRUM AVIII_400
PROBHD 5 mm PABBO_BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 80.6
DW 60.800 usec
DE 6.50 usec
TE 296.2 K
D1 1.00000000 sec
TD0 1

----- CHANNEL f1 -----
SF01 399.9124696 MHz
NUC1 1H
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 399.9100097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

2b - ¹³C NMR, 101 MHz, CDCl₃



Current Data Parameters
NAME PCK-3-68
EXPNO 18
PROCNO 1

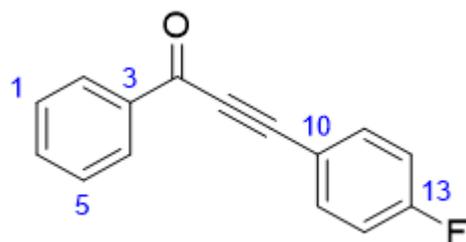
F2 - Acquisition Parameters

Date_ 20210325
Time_ 22.41 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters

SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

2b - ^{19}F NMR, 376 MHz, CDCl_3

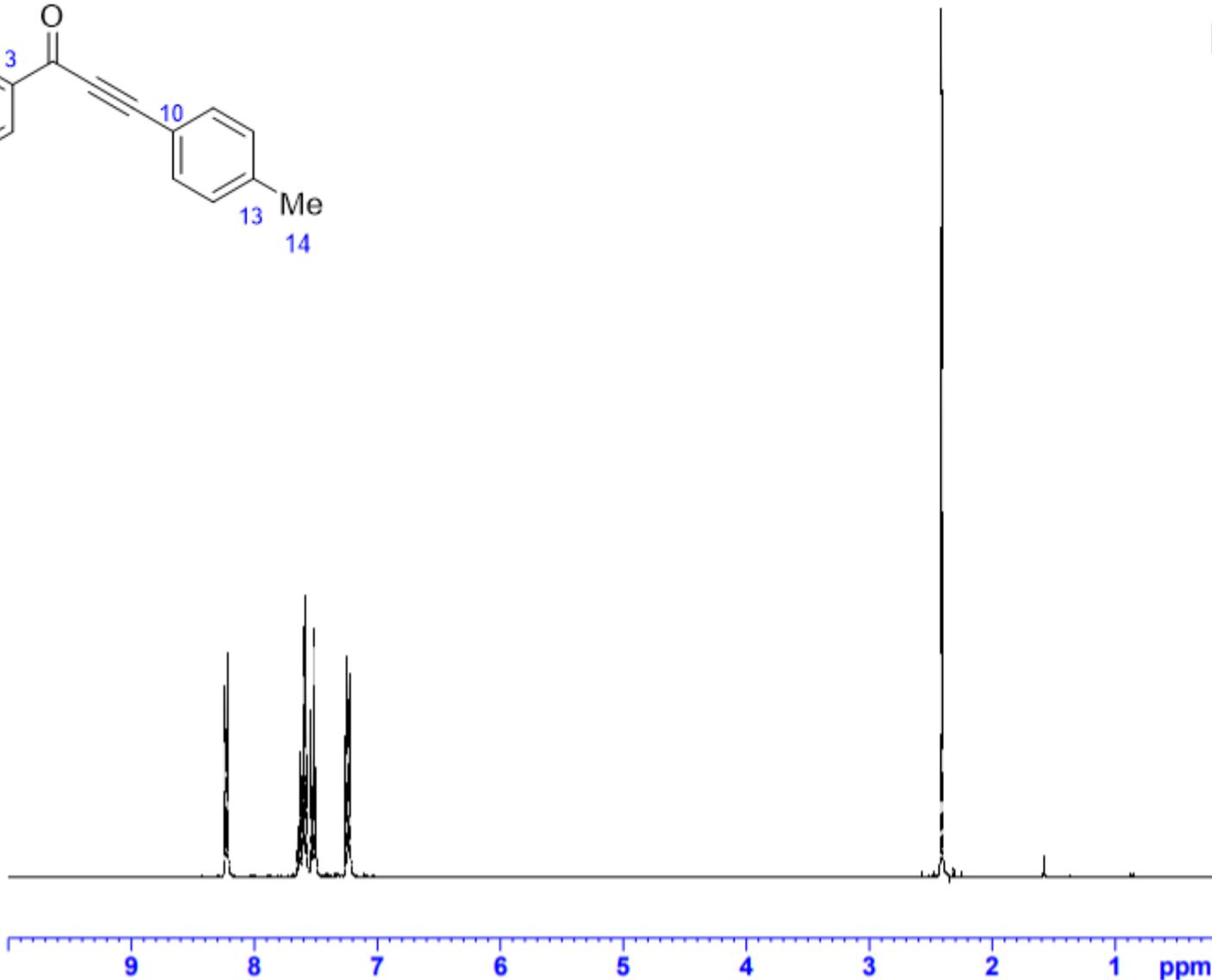
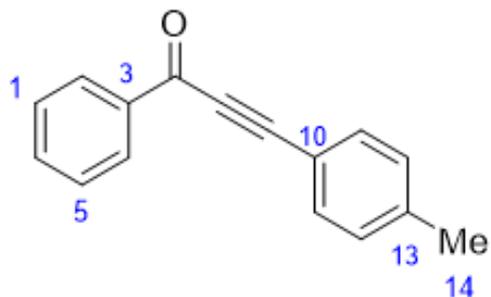


Current Data Parameters
NAME PCK-3-68
EXPNO 31
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210326
Time_ 10.55 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zg
TD 261992
SOLVENT CDCl3
NS 16
DS 4
SWH 89285.711 Hz
FIDRES 0.681591 Hz
AQ 1.4671552 sec
RG 575
DW 5.600 usec
DE 7.11 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 376.4418995 MHz
NUC1 19F
P1 11.80 usec
PLW1 32.96500015 W

F2 - Processing parameters
SI 262144
SF 376.4795470 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

2c - ¹H NMR, 400 MHz, CDCl₃



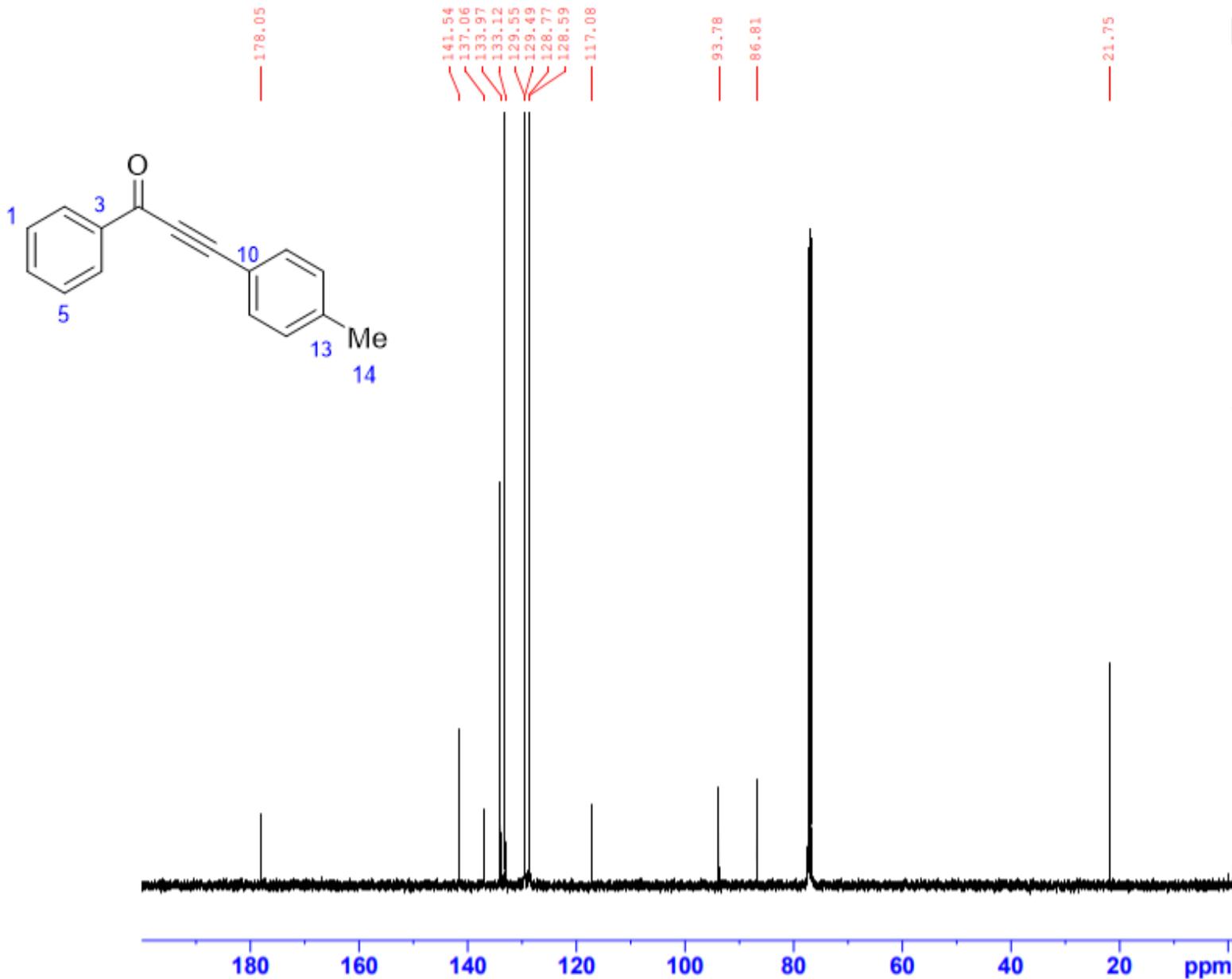
Current Data Parameters
NAME PCK-3-67
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210309
Time_ 18.42
INSTRUM AVIII 400
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 80.6
DW 60.800 usec
DE 6.50 usec
TE 297.0 K
D1 1.00000000 sec
TD0 1

----- CHANNEL f1 -----
SF01 399.9124696 MHz
NUC1 1H
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 399.9100098 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

2c - ¹³C NMR, 101 MHz, CDCl₃



Current Data Parameters
NAME PCK-3-67
EXPNO 11
PROCNO 1

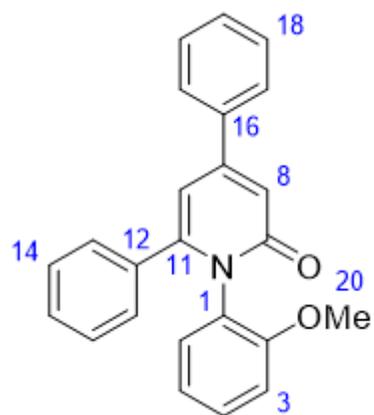
F2 - Acquisition Parameters
Date_ 20210309
Time_ 18.39
INSTRUM AVIII 400
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 1024
DS 4
SWH 24038.461 Hz
FIDRES 0.250010 Hz
AQ 1.9999200 sec
RG 128
DW 20.800 usec
DE 6.50 usec
TE 301.5 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

----- CHANNEL f1 -----
SFO1 100.5675047 MHz
NUC1 13C
P1 9.00 usec
PLW1 96.68000031 W

----- CHANNEL f2 -----
SFO2 399.9115996 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.38907000 W

F2 - Processing parameters
SI 131072
SF 100.5574500 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

S4 - ¹H NMR, 400 MHz, CDCl₃

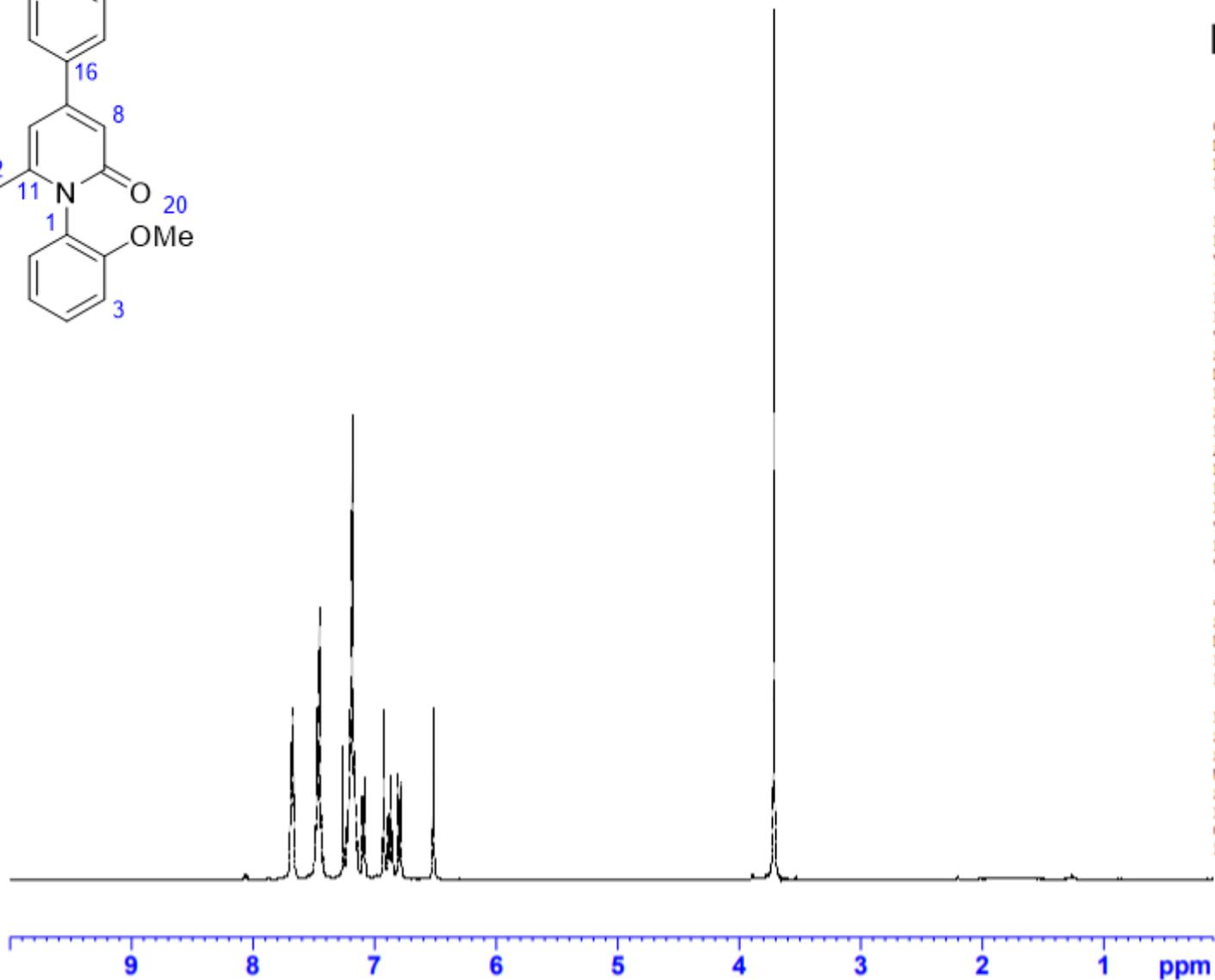


Current Data Parameters
NAME JS-4-266 xtal
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210227
Time 8.18
INSTRUM AVIII_400
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 80.6
DW 60.800 usec
DE 6.50 usec
TE 296.5 K
D1 1.00000000 sec
TD0 1

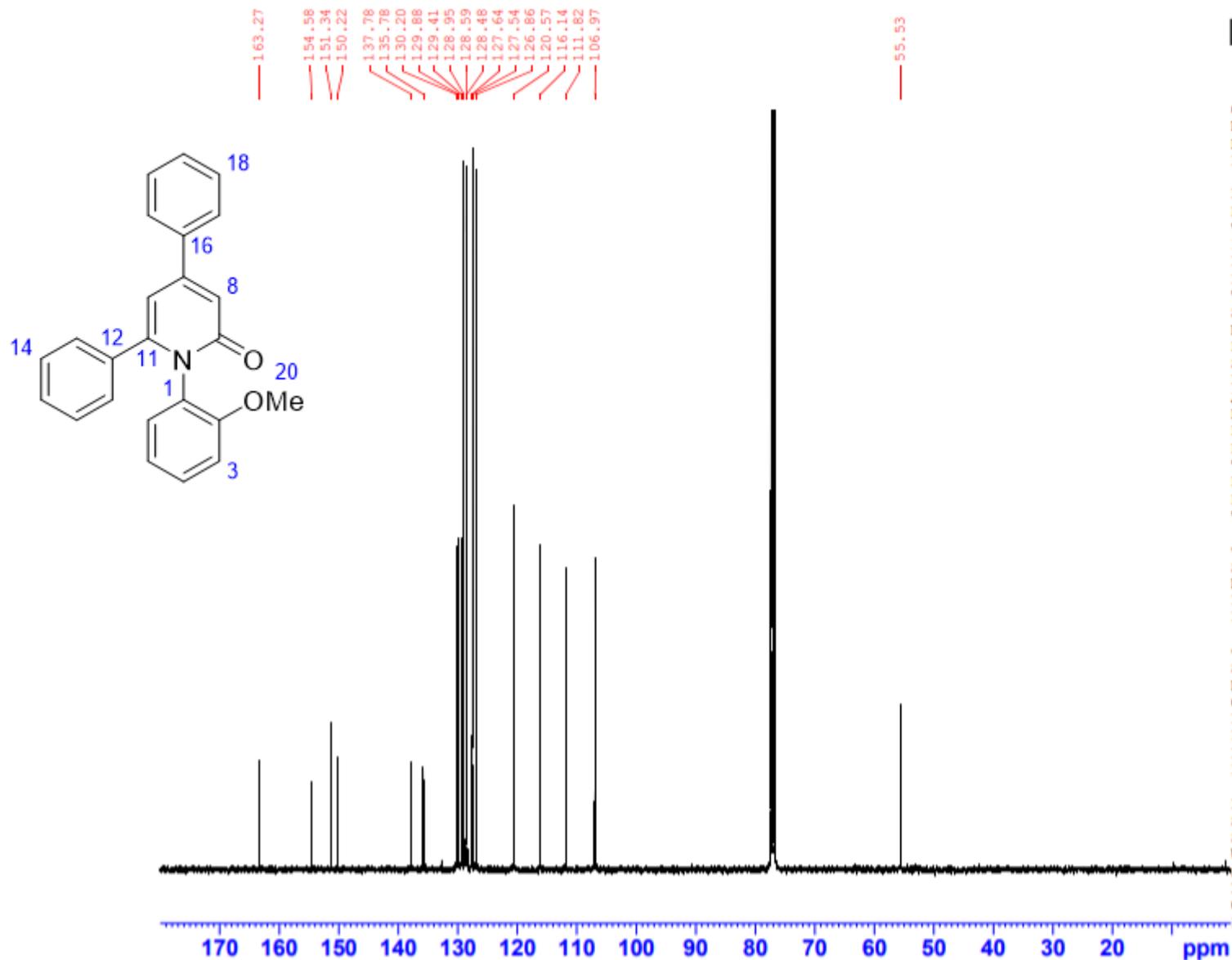
===== CHANNEL f1 =====
SFO1 399.9124696 MHz
NUC1 1H
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 399.9100097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



2.00
3.06
5.86
1.00
1.00
1.00
1.00
0.99

3.02



Current Data Parameters
 NAME JS-4-266 xtal
 EXPNO 11
 PROCNO 1

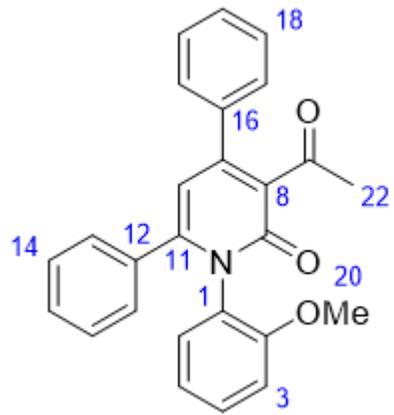
F2 - Acquisition Parameters
 Date_ 20210227
 Time_ 8.14
 INSTRUM AVIII_400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 96150
 SOLVENT CDCl3
 NS 4096
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.250010 Hz
 AQ 1.9999200 sec
 RG 80.6
 DW 20.800 usec
 DE 6.50 usec
 TE 301.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

----- CHANNEL f1 -----
 SFO1 100.5675047 MHz
 NUC1 13C
 P1 9.00 usec
 PLW1 96.68000031 W

----- CHANNEL f2 -----
 SFO2 399.9115996 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 17.29199982 W
 PLW12 0.48032999 W
 PLW13 0.38907000 W

F2 - Processing parameters
 SI 131072
 SF 100.5574500 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

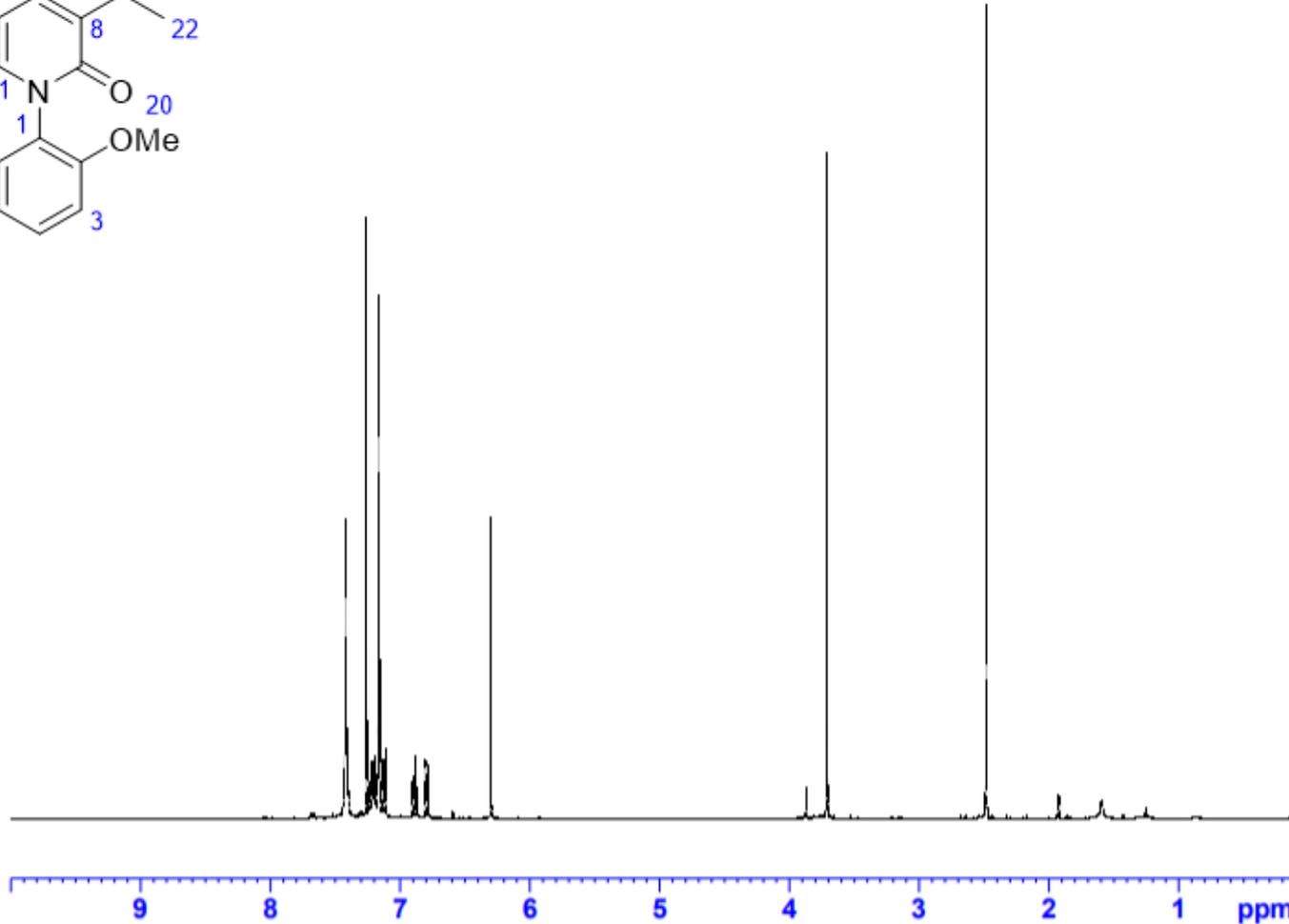
S5 - ¹H NMR, 400 MHz, CDCl₃



Current Data Parameters
NAME JS-4-282 conc
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210317
Time_ 22.36 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100099 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4.96
6.08
1.02
1.02
1.00
0.96

3.01

2.97

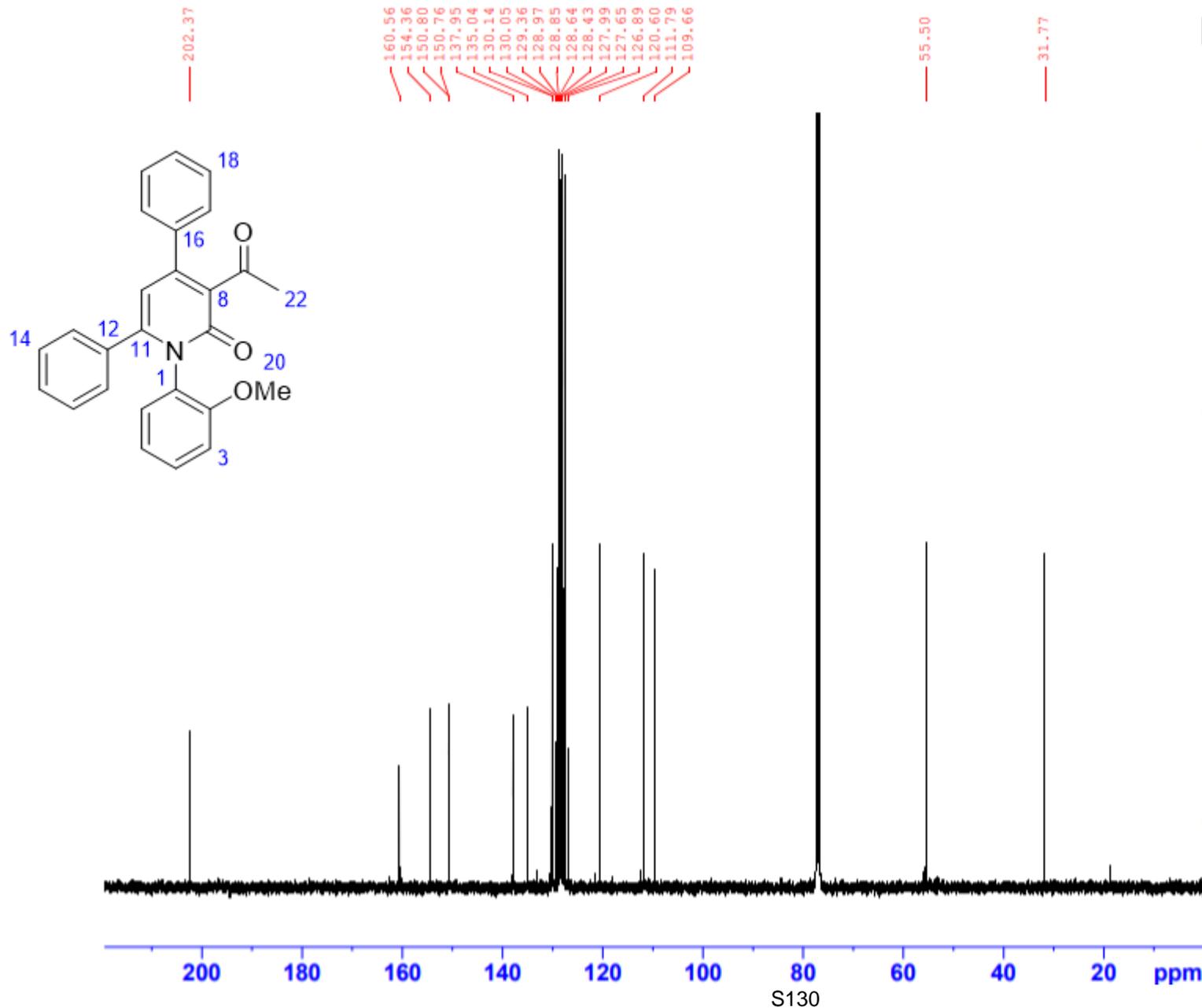
S129



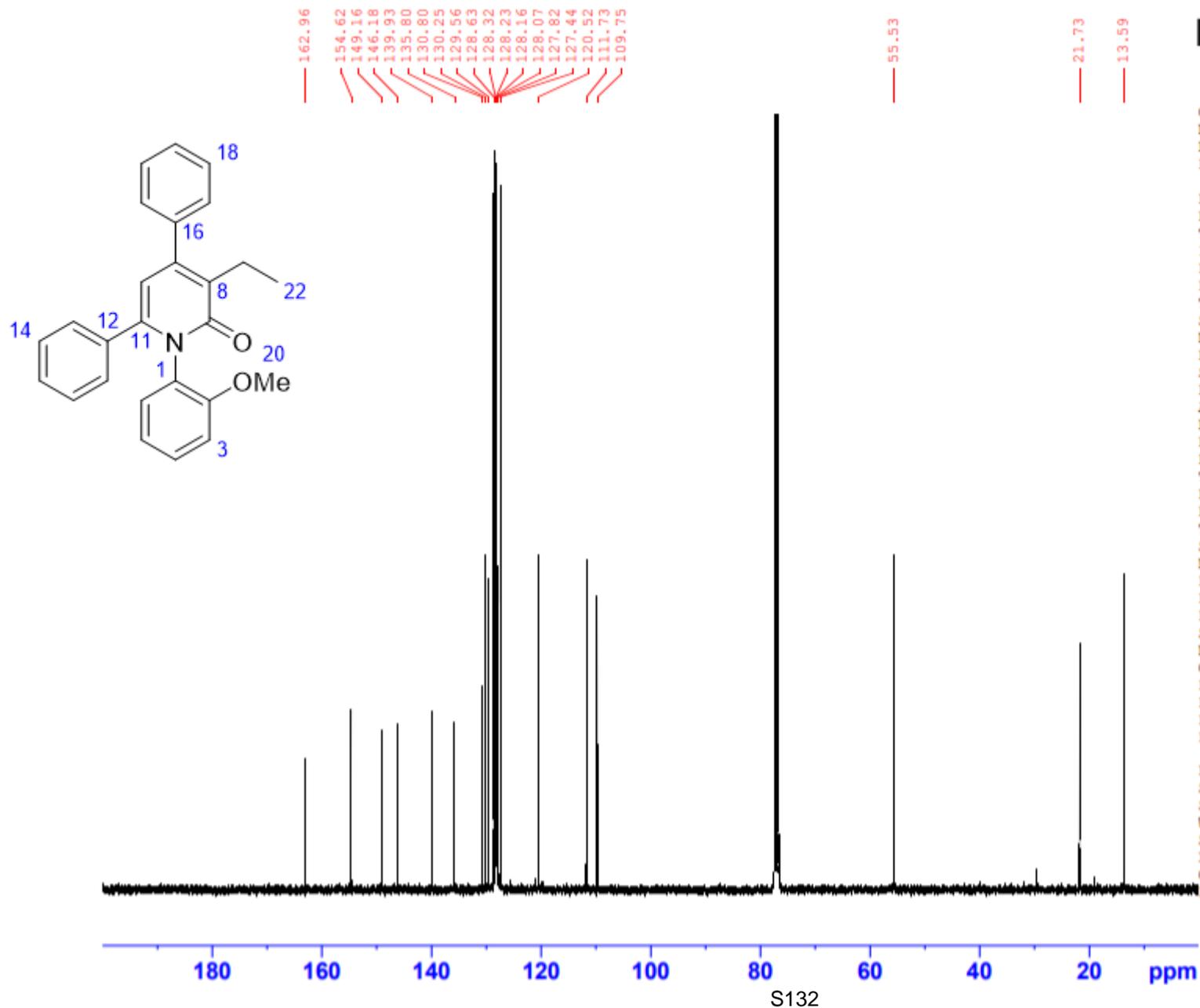
Current Data Parameters
 NAME JS-4-282 conc
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210317
 Time_ 22.33 h
 INSTRUM AVIII 400
 PROBHD Z108618_0146 (
 PULPROG zgpg30
 TD 96150
 SOLVENT CDCl3
 NS 4096
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.500020 Hz
 AQ 1.9999200 sec
 RG 2050
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1
 SFO1 100.6178003 MHz
 NUC1 13C
 P0 3.00 usec
 P1 9.00 usec
 PLW1 96.68000031 W
 SFO2 400.1116004 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 17.29199982 W
 PLW12 0.48032999 W
 PLW13 0.24160001 W

F2 - Processing parameters
 SI 131072
 SF 100.6077409 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



S6 - ¹³C NMR, 101 MHz, CDCl₃

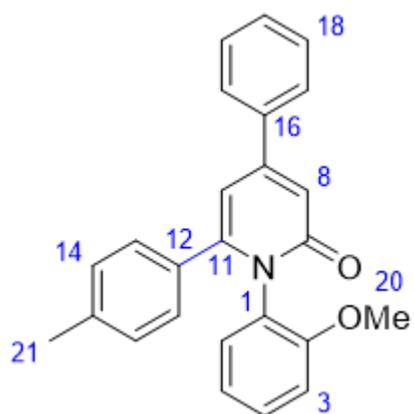


Current Data Parameters
NAME JS-4-281 recol
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210326
Time_ 2.17 h
INSTRUM AVIII 400
PROBHD z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077411 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

S7 - ¹H NMR, 400 MHz, CDCl₃

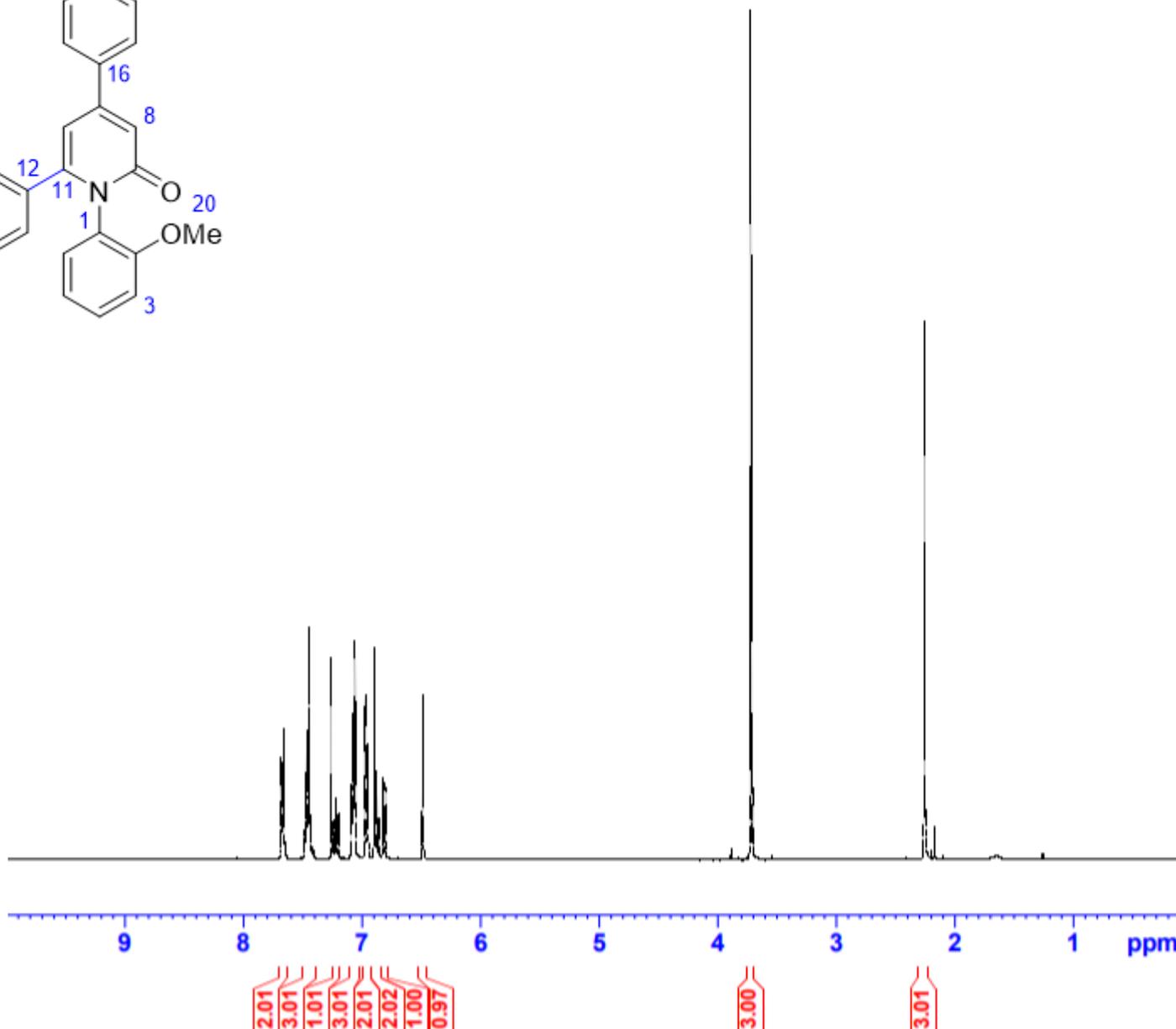


Current Data Parameters
NAME JS-4-278
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210312
Time 22.44
INSTRUM AVIII_400
PROBHD 5 mm PABBO_BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 80.6
DW 60.800 usec
DE 6.50 usec
TE 296.8 K
D1 1.00000000 sec
TD0 1

----- CHANNEL f1 -----
SFO1 399.9124696 MHz
NUC1 1H
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 399.9100097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



S133



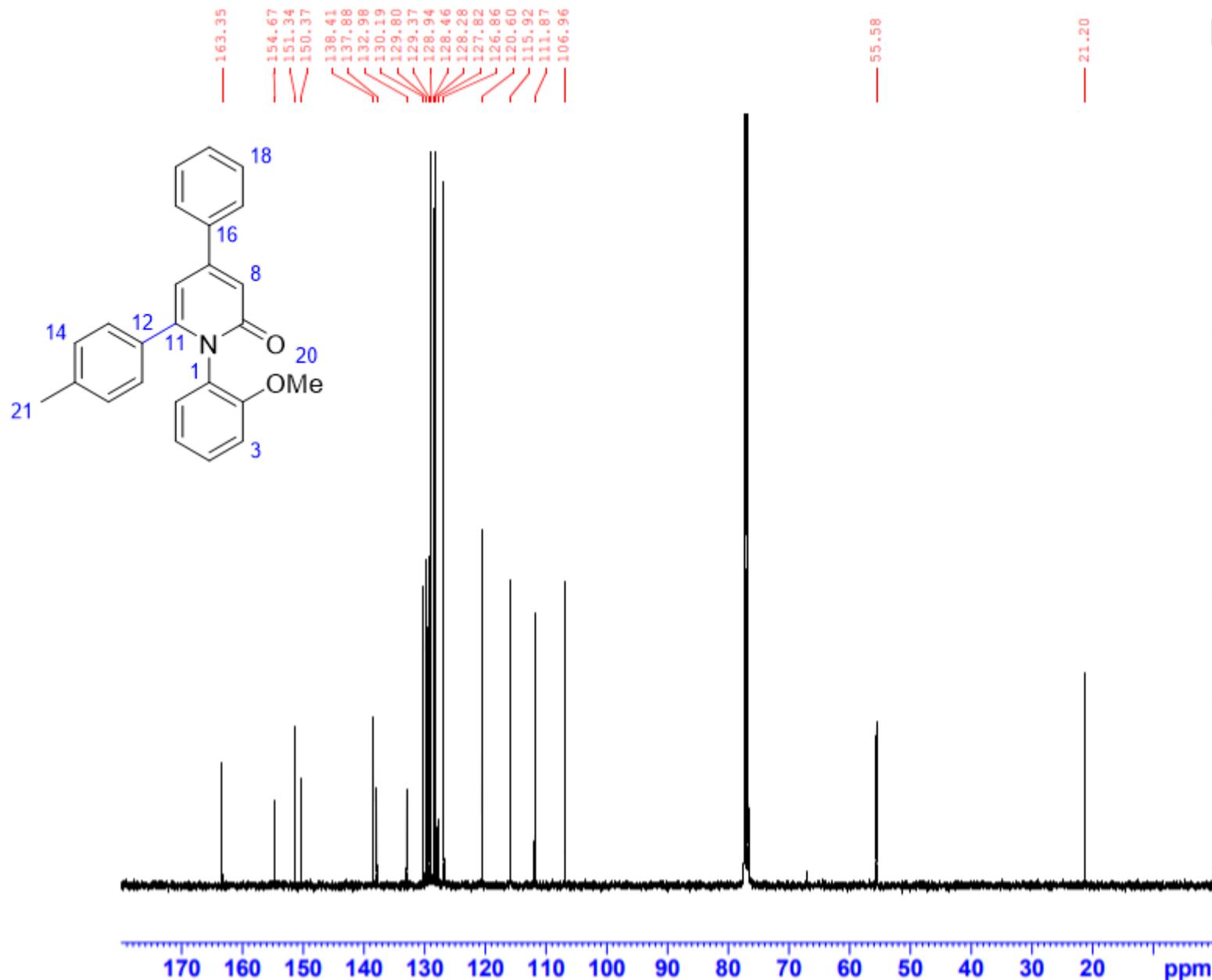
Current Data Parameters
 NAME JS-4-278
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210312
 Time_ 22.40
 INSTRUM AVIII_400
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 96150
 SOLVENT CDCl3
 NS 4096
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.250010 Hz
 AQ 1.9999200 sec
 RG 181
 DW 20.800 usec
 DE 6.50 usec
 TE 301.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

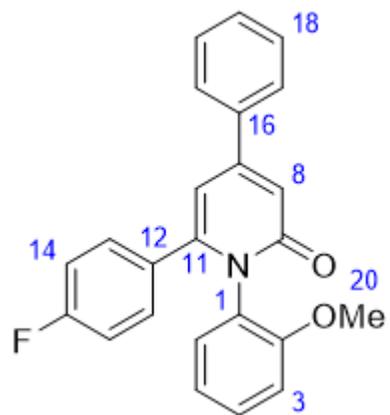
----- CHANNEL f1 -----
 SFO1 100.5675047 MHz
 NUC1 13C
 P1 9.00 usec
 PLW1 96.68000031 W

----- CHANNEL f2 -----
 SFO2 399.9115996 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 17.29199982 W
 PLW12 0.48032999 W
 PLW13 0.38907000 W

F2 - Processing parameters
 SI 131072
 SF 100.5574483 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



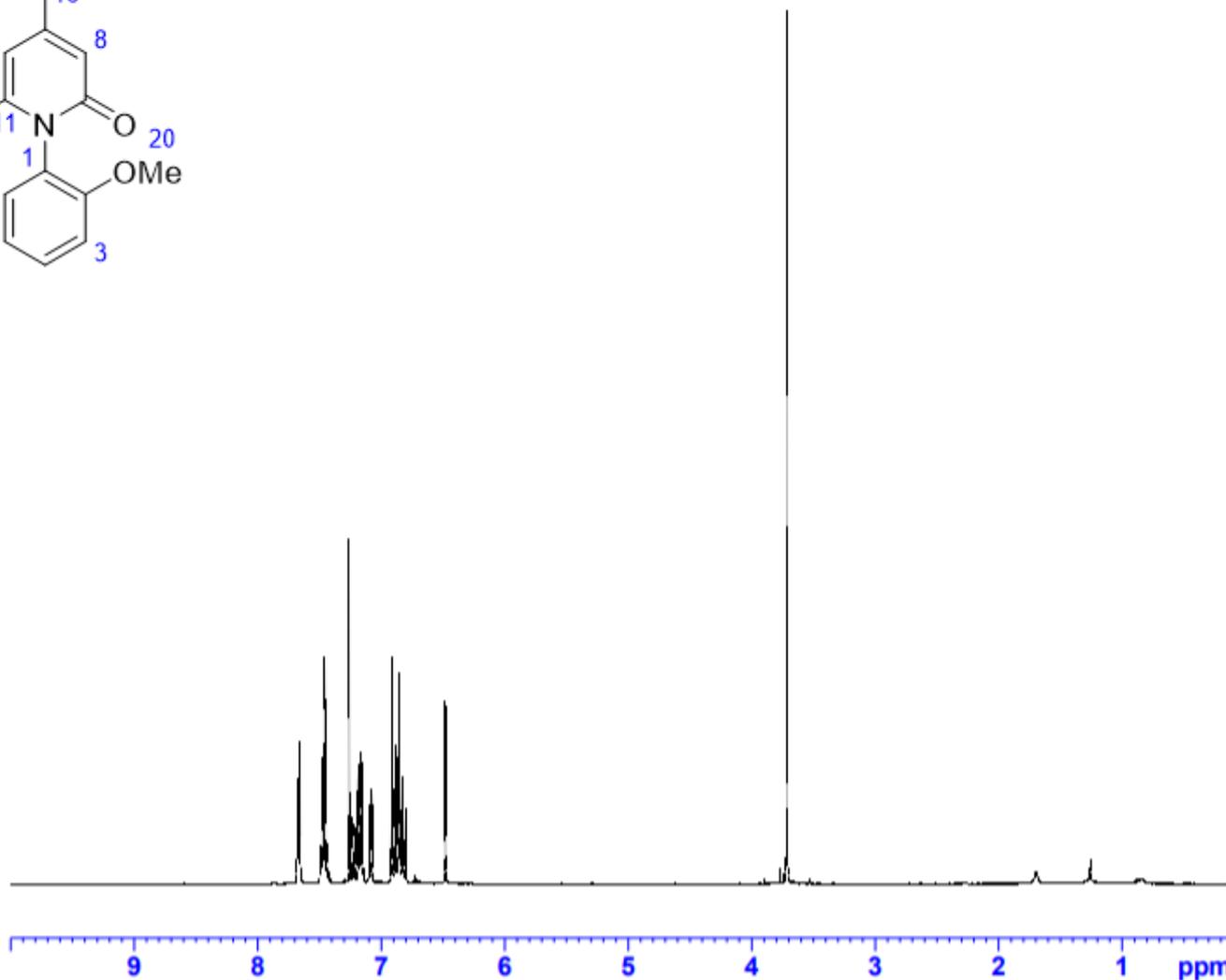
S8 - ¹H NMR, 400 MHz, CDCl₃



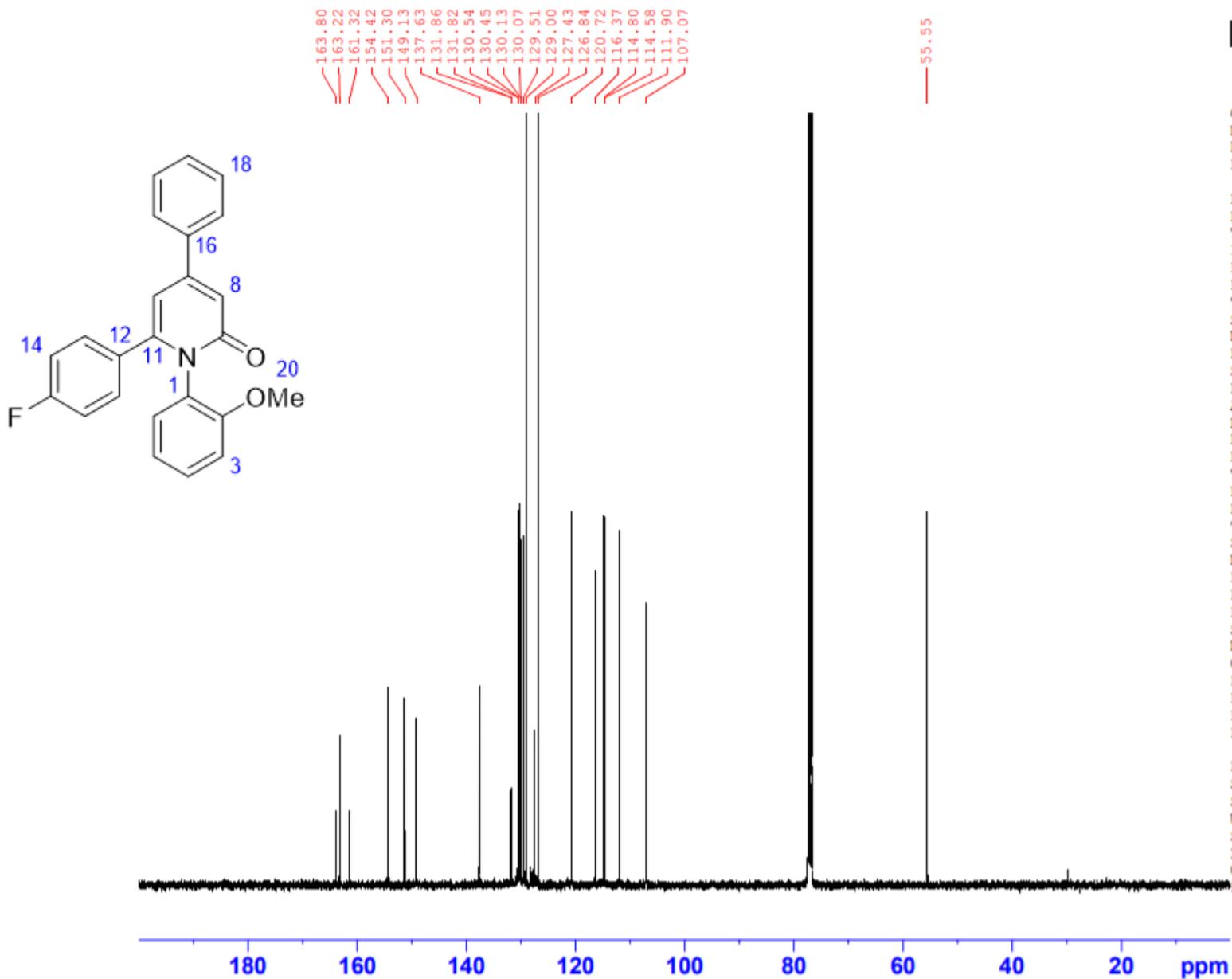
Current Data Parameters
NAME JS-4-275 recol
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210326
Time_ 10.44 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 161
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TDO 1
SFO1 400.1124708 MHz
NUC1 1H
PO 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



S135

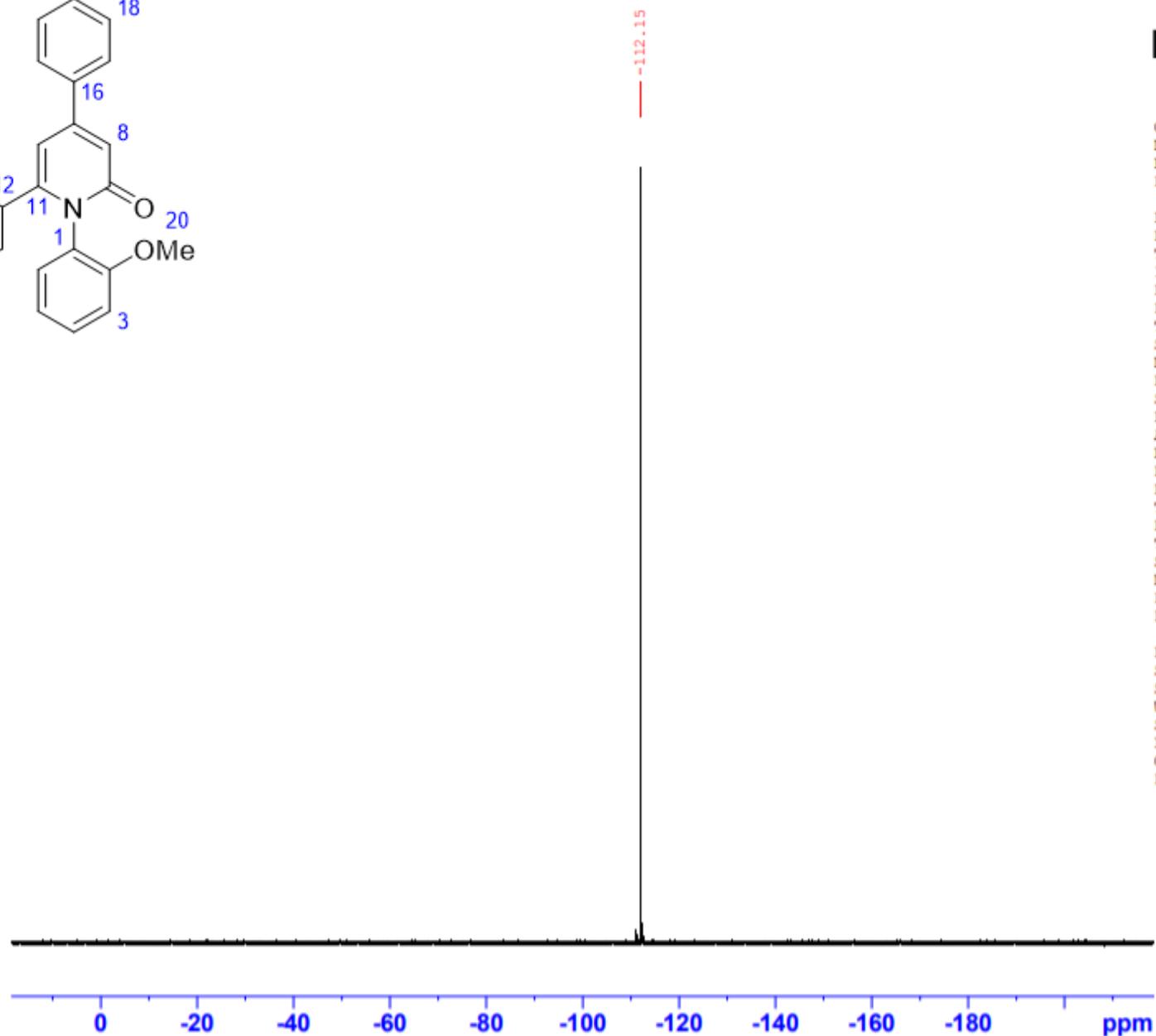
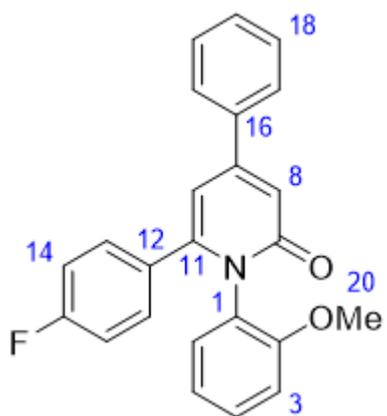


Current Data Parameters
 NAME JS-4-275
 EXPNO 12
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210419
 Time_ 23.30 h
 INSTRUM AVIII 400
 PROBHD Z108618_0146 ()
 PULPROG zgpg30
 TD 96150
 SOLVENT CDCl3
 NS 4096
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.500020 Hz
 AQ 1.9999200 sec
 RG 2050
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1
 SFO1 100.6178003 MHz
 NUC1 13C
 P0 3.00 usec
 P1 9.00 usec
 PLW1 96.68000031 W
 SFO2 400.1116004 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 17.29199982 W
 PLW12 0.48032999 W
 PLW13 0.24160001 W

F2 - Processing parameters
 SI 131072
 SF 100.6077400 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

S8 - ^{19}F NMR, 376 MHz, CDCl_3

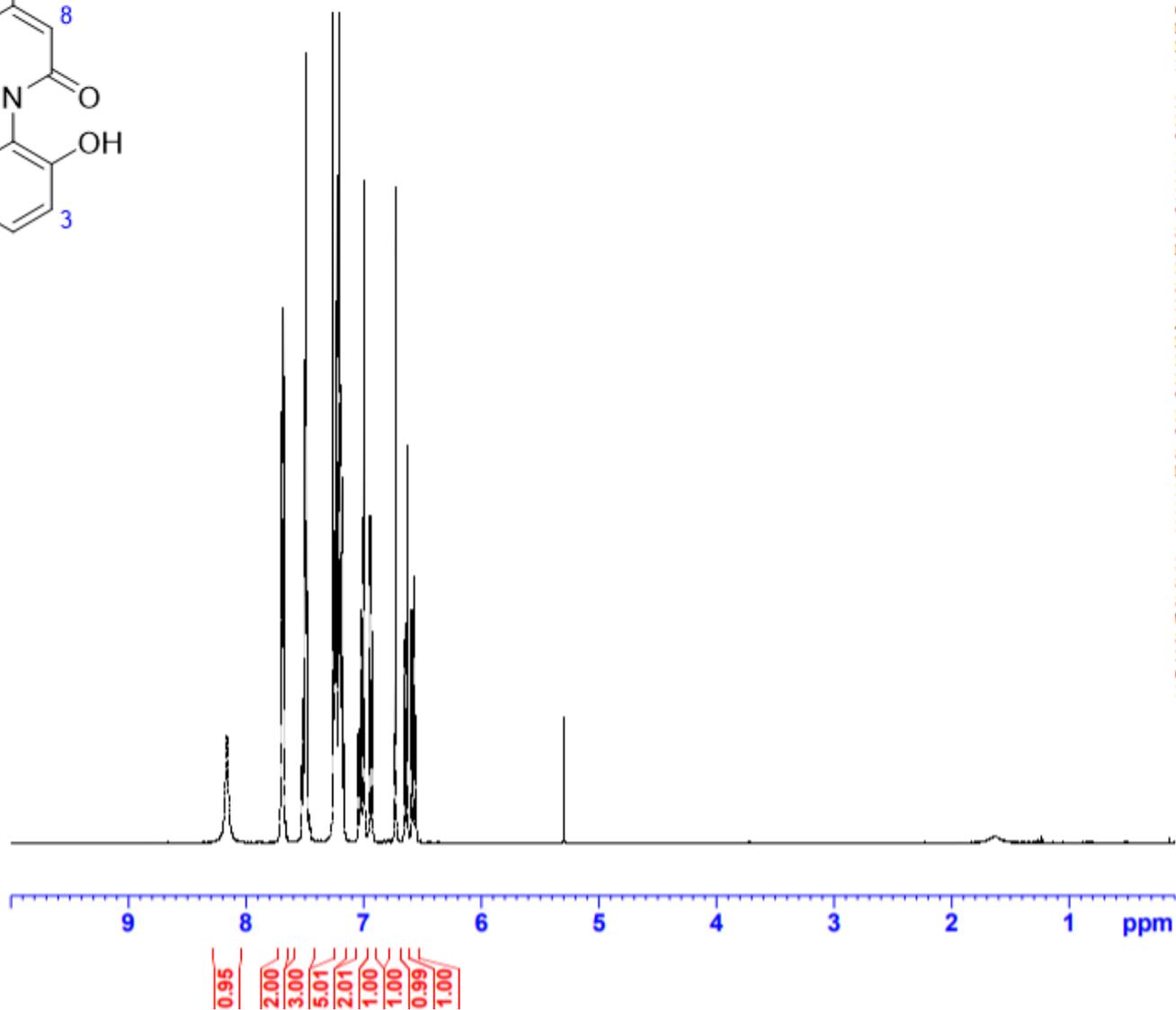
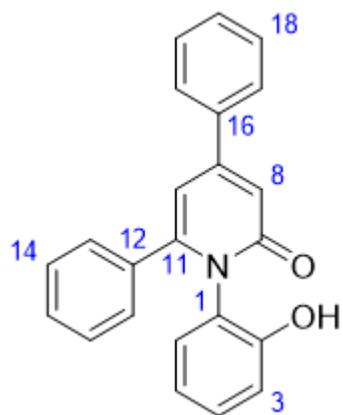


Current Data Parameters
NAME JS-4-275
EXPNO 16
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210420
Time_ 0.01 h
INSTRUM AVIII 400
PROBHD Z108618_0146 ()
PULPROG zg
TD 261992
SOLVENT CDCl_3
NS 16
DS 4
SWH 89285.711 Hz
FIDRES 0.681591 Hz
AQ 1.4671552 sec
RG 575
DW 5.600 usec
DE 7.11 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 376.4418995 MHz
NUC1 ^{19}F
P1 11.80 usec
PLW1 32.96500015 W

F2 - Processing parameters
SI 262144
SF 376.4795470 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

3a - ¹H NMR, 400 MHz, CDCl₃

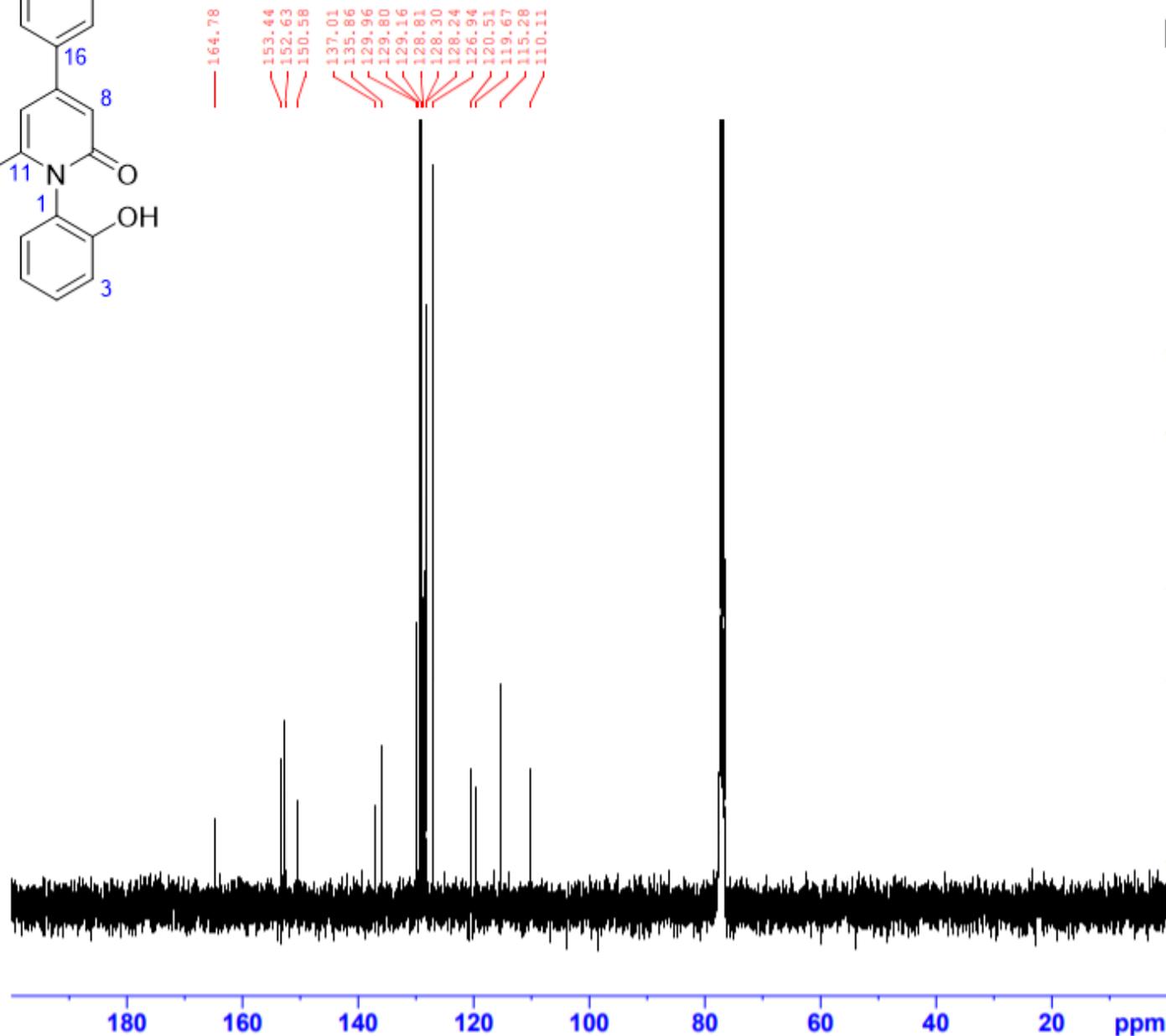
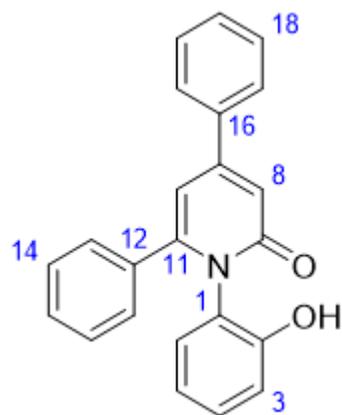


Current Data Parameters
NAME JS-4-267 trit
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210506
Time_ 12.51 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100099 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

3a - ¹³C NMR, 101 MHz, CDCl₃

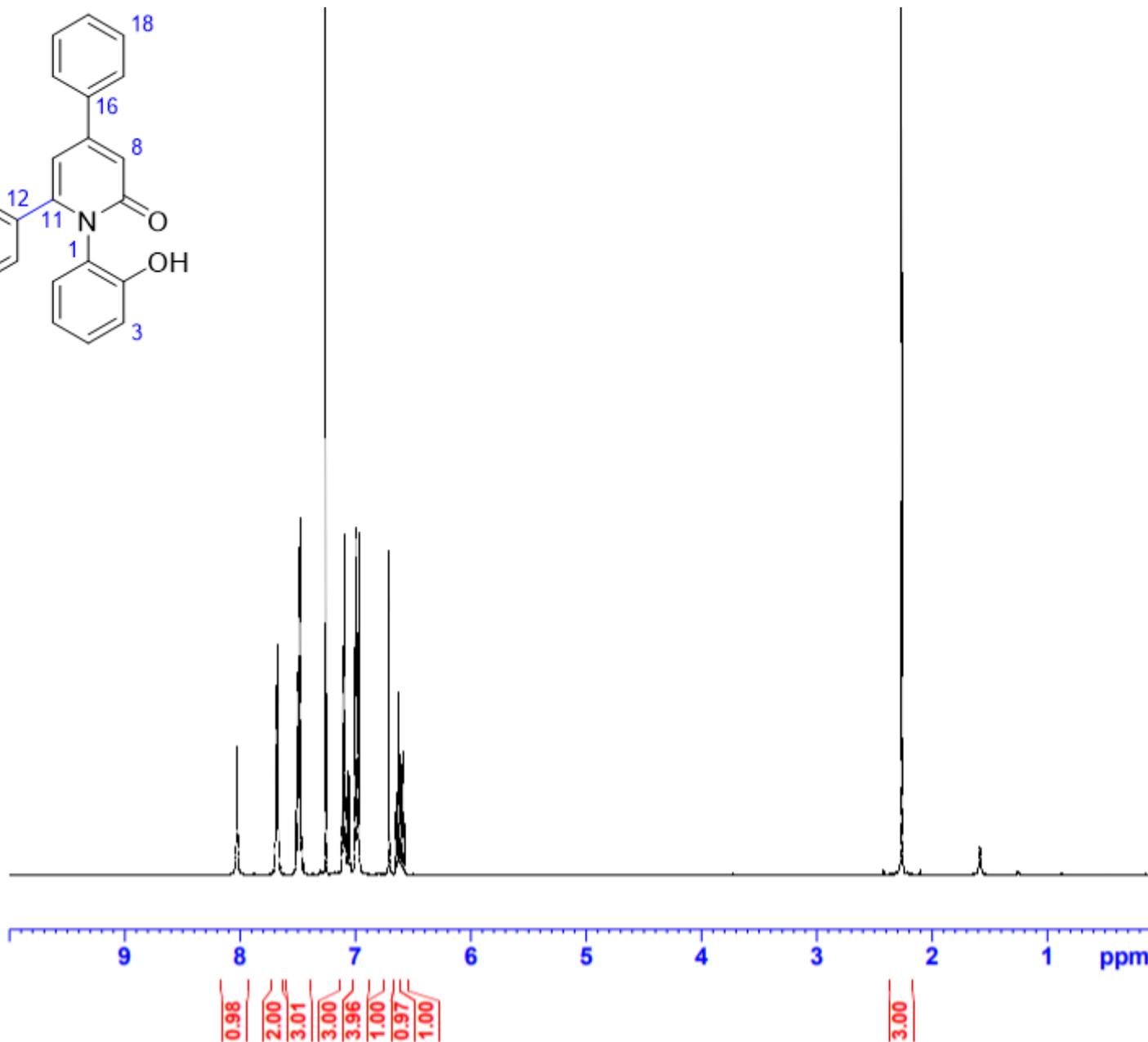
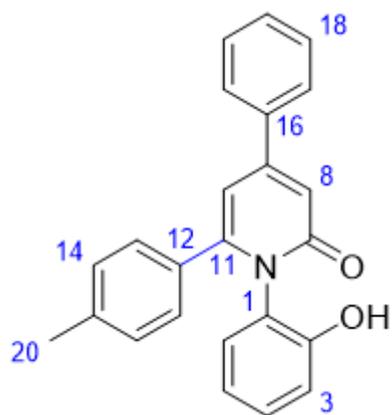


Current Data Parameters
NAME JS-4-267 trit
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210507
Time_ 3.16 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.6800031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077403 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

3b - ¹H NMR, 400 MHz, CDCl₃

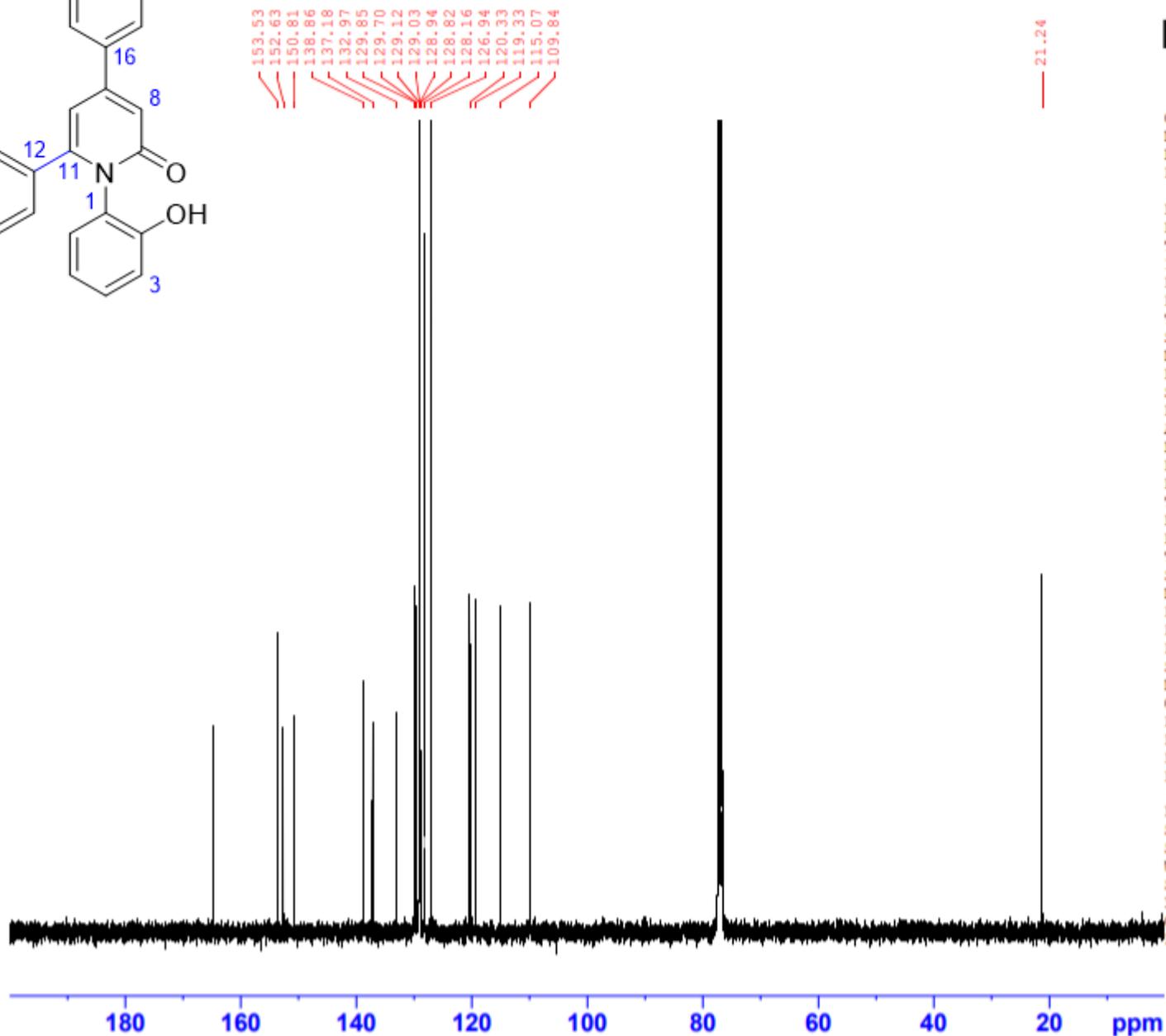
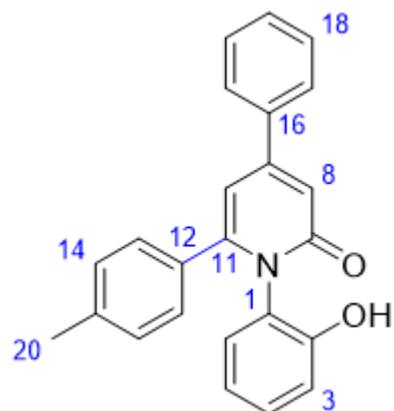


Current Data Parameters
NAME JS-4-283 col 1
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210317
Time 14.02 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (zg30)
PULPROG _zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 322
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100105 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

3b - ¹³C NMR, 101 MHz, CDCl₃



S141

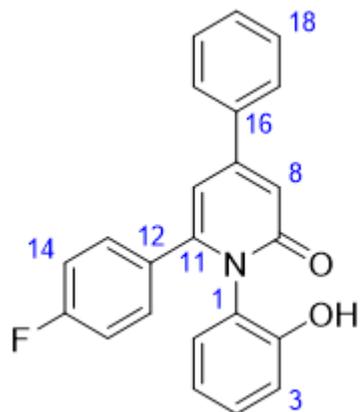


Current Data Parameters
NAME JS-4-283 col 1
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210318
Time 2.36 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077407 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

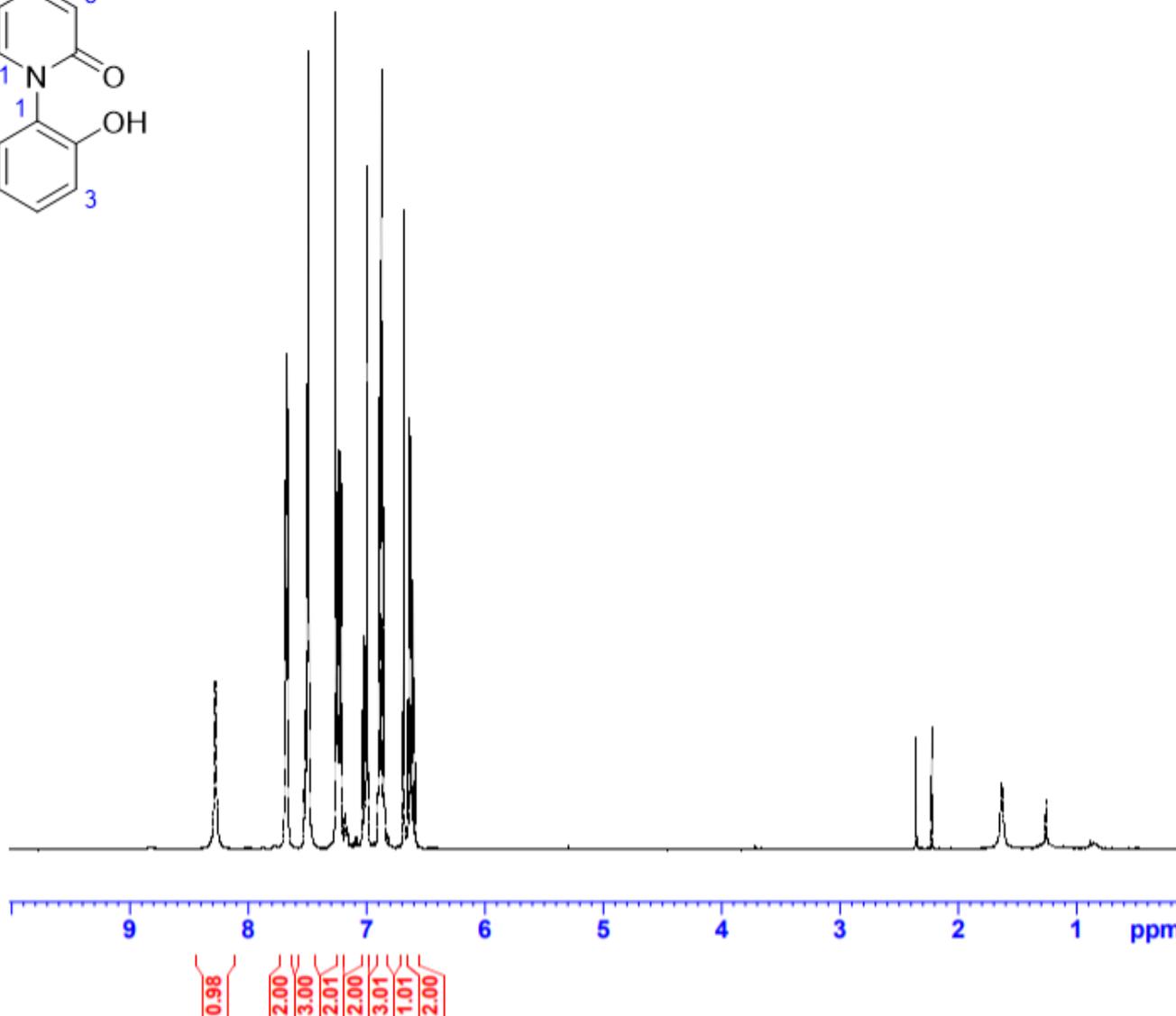
3c - ¹H NMR, 400 MHz, CDCl₃



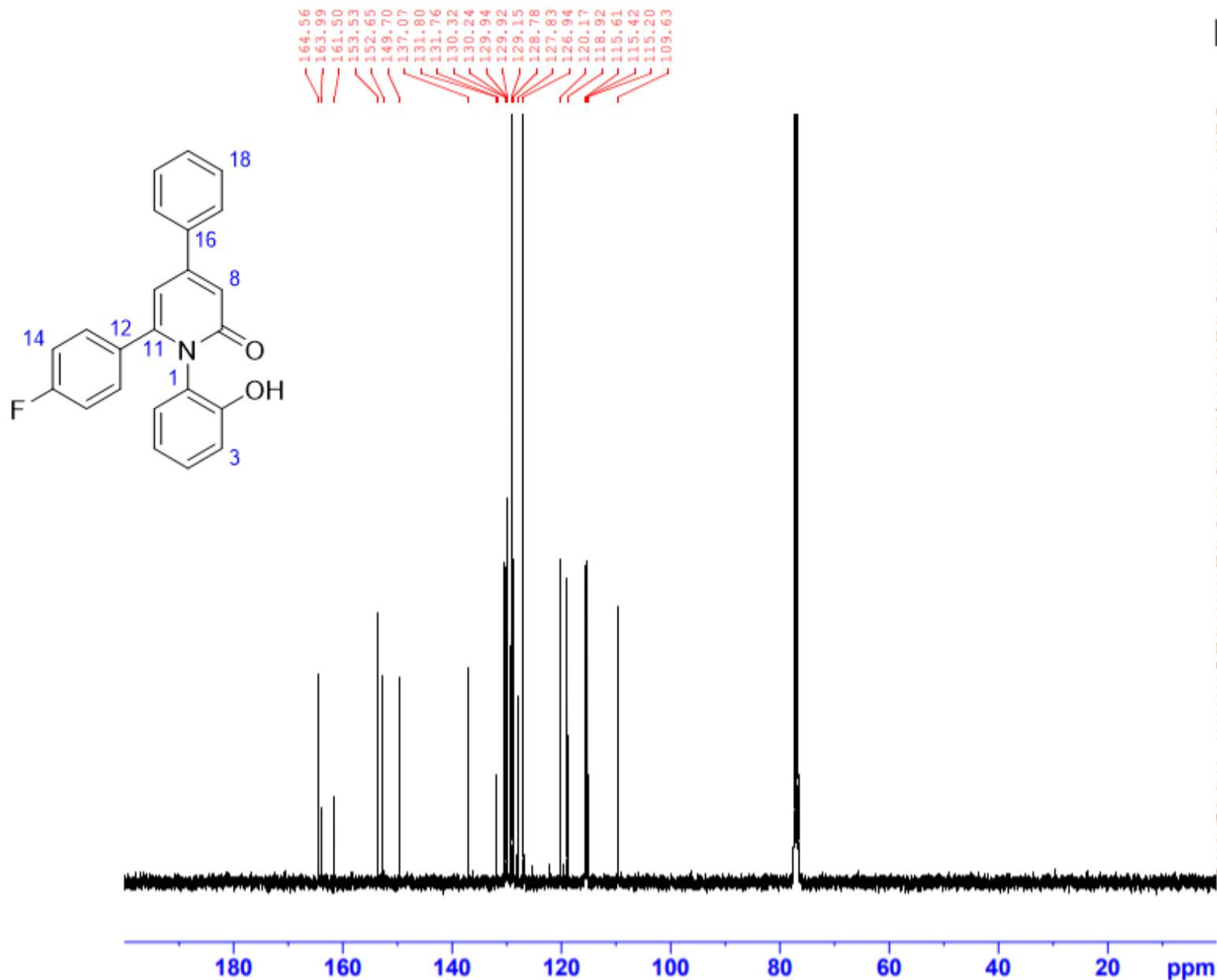
Current Data Parameters
NAME JS-4-280 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210317
Time_ 17.30 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
PO 5.00 usec
PL 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



3c - ¹³C NMR, 101 MHz, CDCl₃



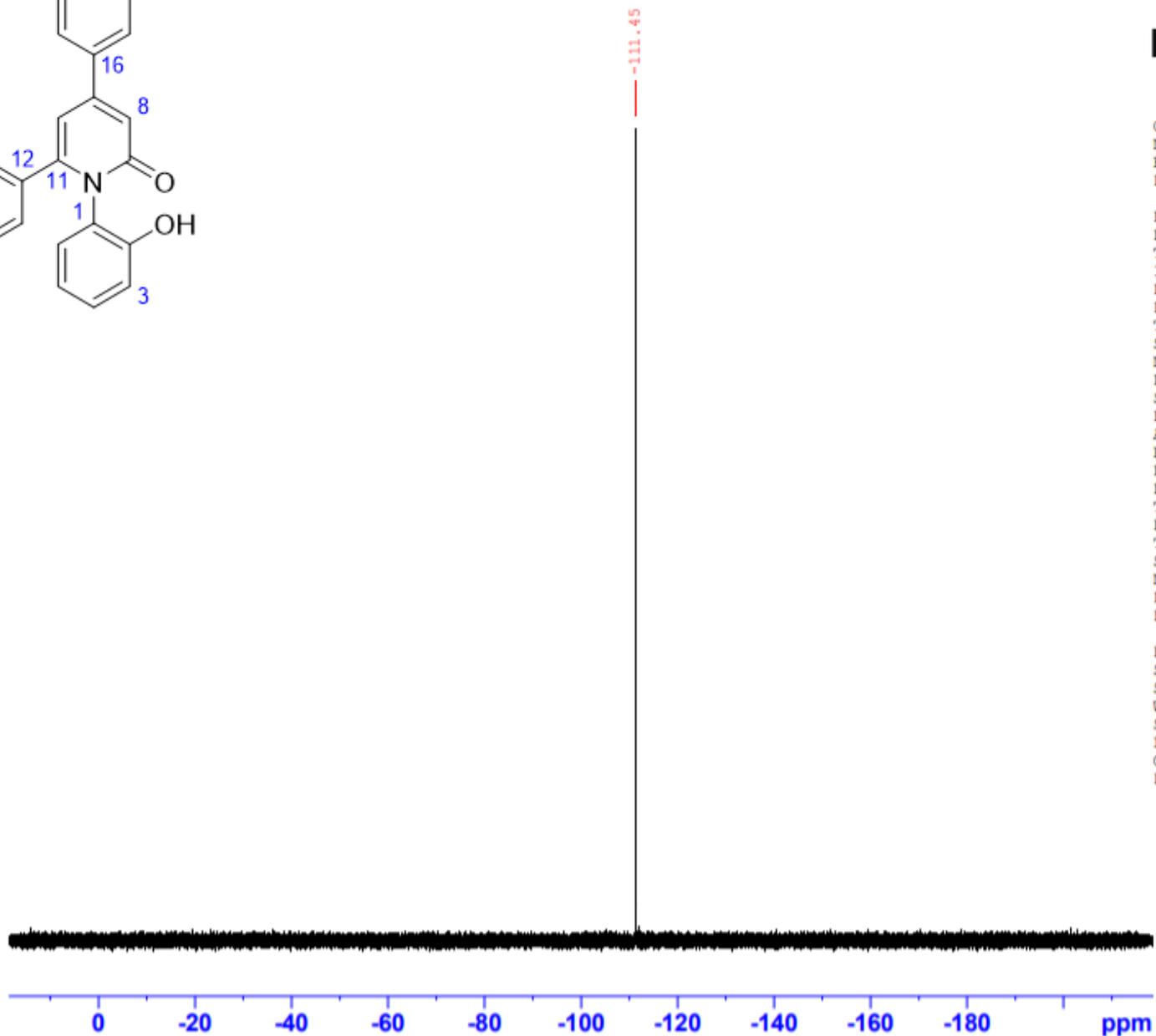
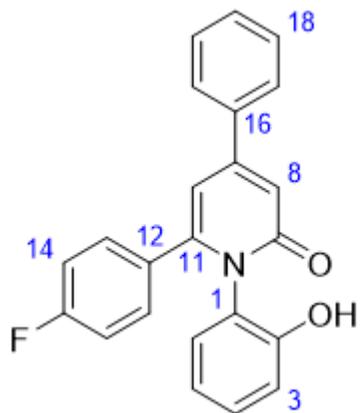
Current Data Parameters
NAME JS-4-280 col
EXPNO 20
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210318
Time 23.13 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.6800031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077405 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

S143

3c - ^{19}F NMR, 376 MHz, CDCl_3

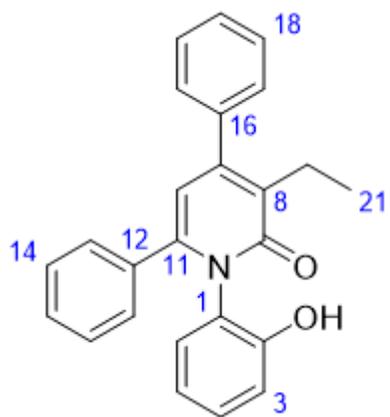


Current Data Parameters
NAME JS-4-280 col
EXPNO 47
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210505
Time 13.55 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zg
TD 261992
SOLVENT CDCl_3
NS 16
DS 4
SWH 89285.711 Hz
FIDRES 0.681591 Hz
AQ 1.4671552 sec
RG 575
DW 5.600 usec
DE 7.11 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 376.4418995 MHz
NUC1 ^{19}F
P1 11.80 usec
PLW1 32.96500015 W

F2 - Processing parameters
SI 262144
SF 376.4795470 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

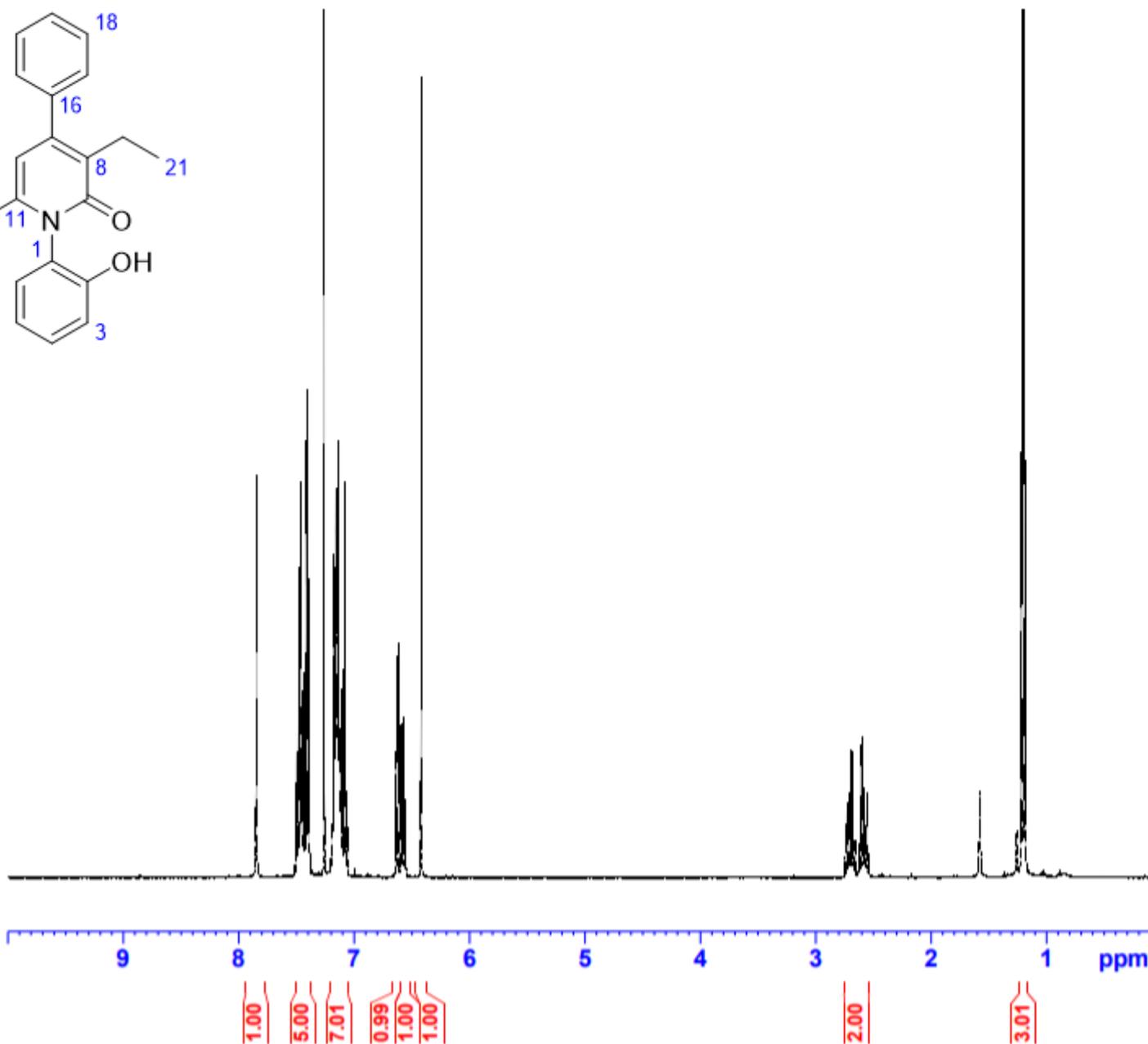
3d - ¹H NMR, 400 MHz, CDCl₃



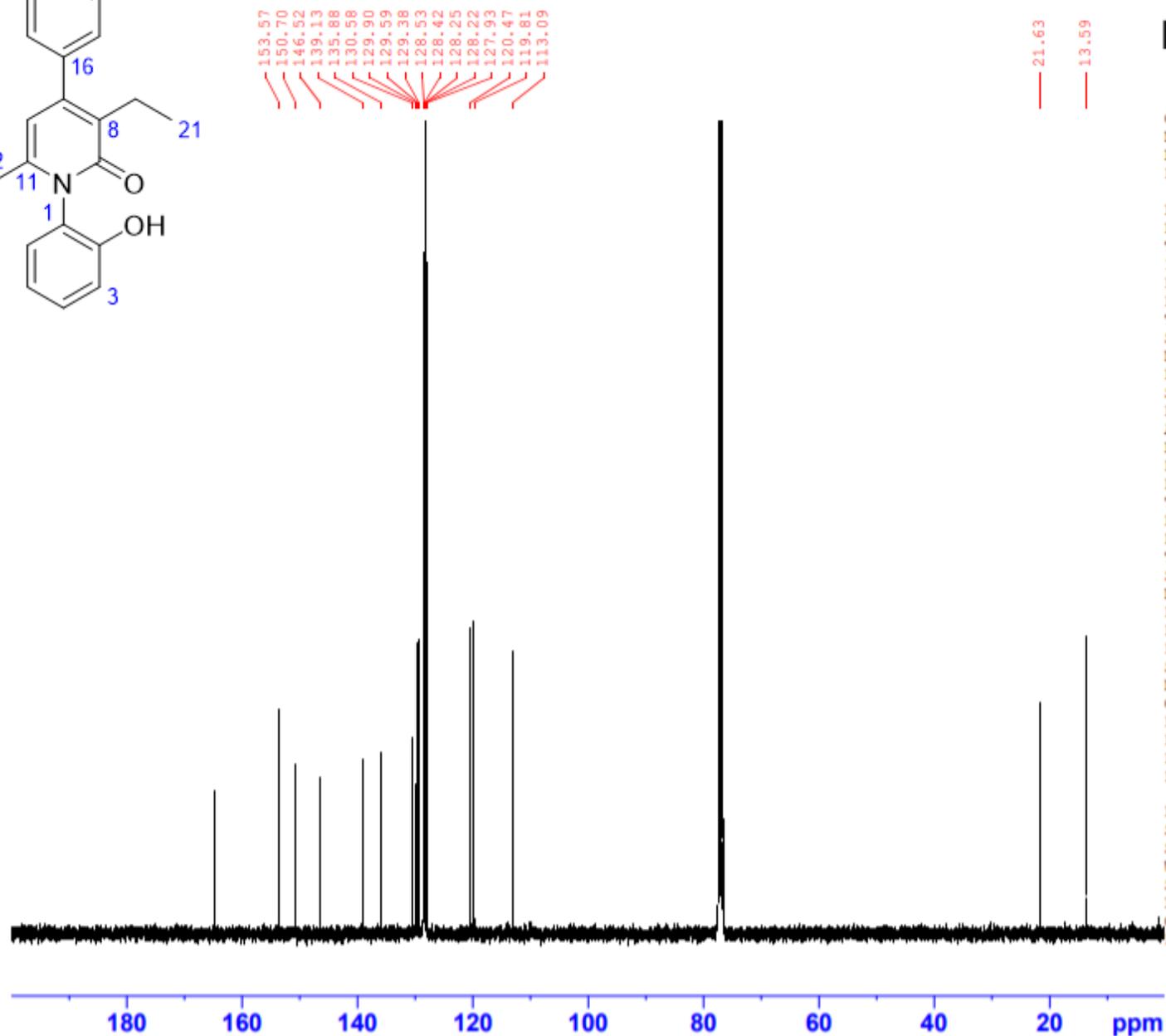
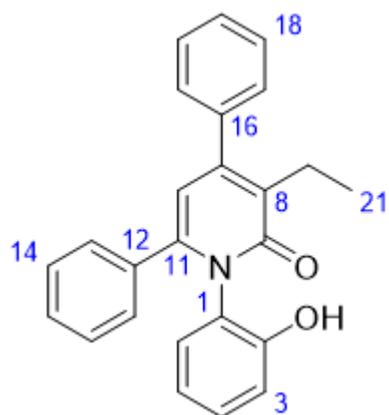
Current Data Parameters
NAME JS-4-285 col
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210323
Time_ 22.50 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 322
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



3d - ¹³C NMR, 101 MHz, CDCl₃



153.57
150.70
146.52
139.13
135.88
130.58
129.90
129.59
129.38
128.93
128.42
128.25
128.22
127.93
120.47
119.81
113.09

21.63

13.59

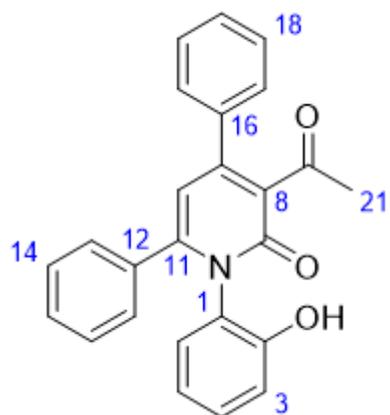


Current Data Parameters
NAME JS-4-285 col
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210323
Time 22.47 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077405 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

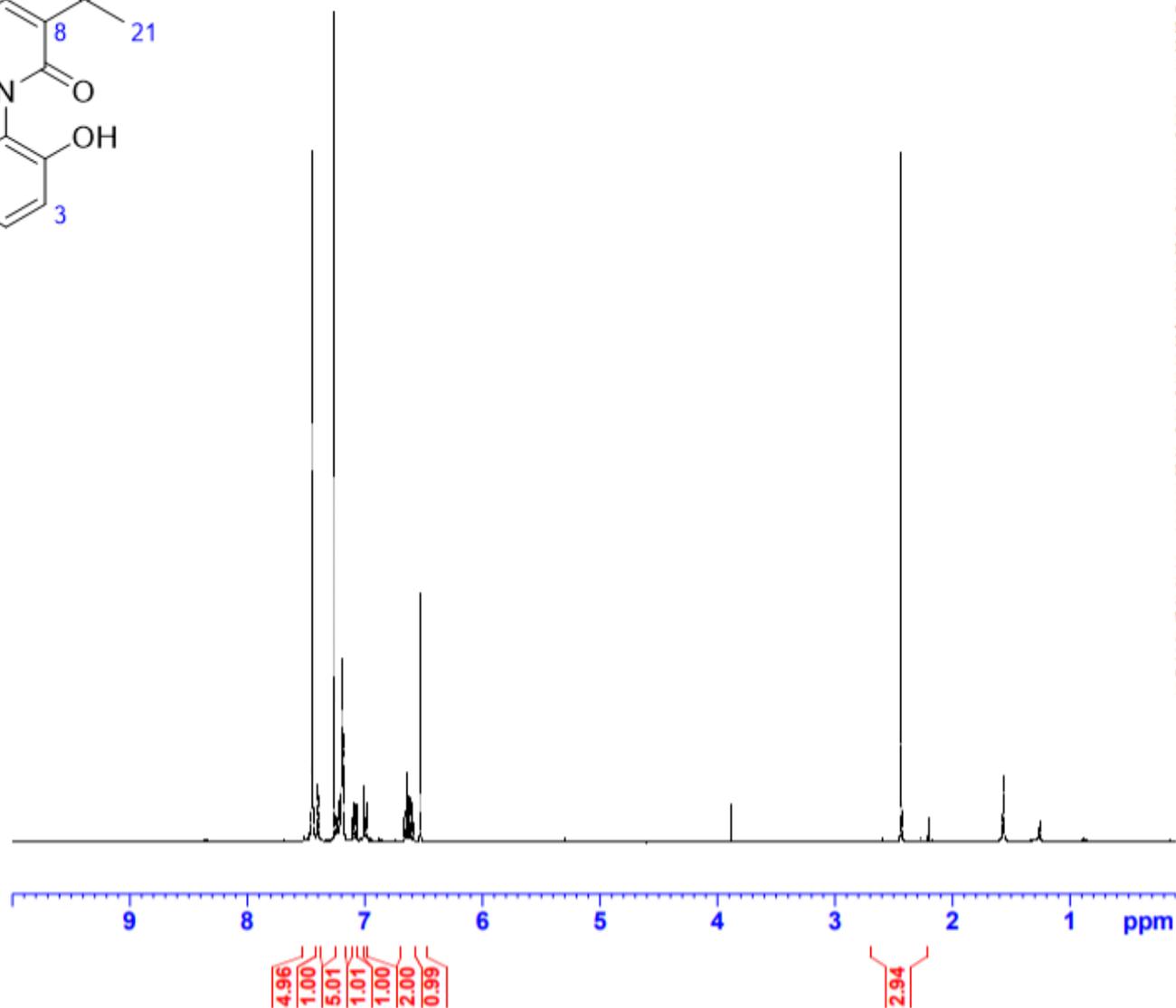
3e - ¹H NMR, 400 MHz, CDCl₃



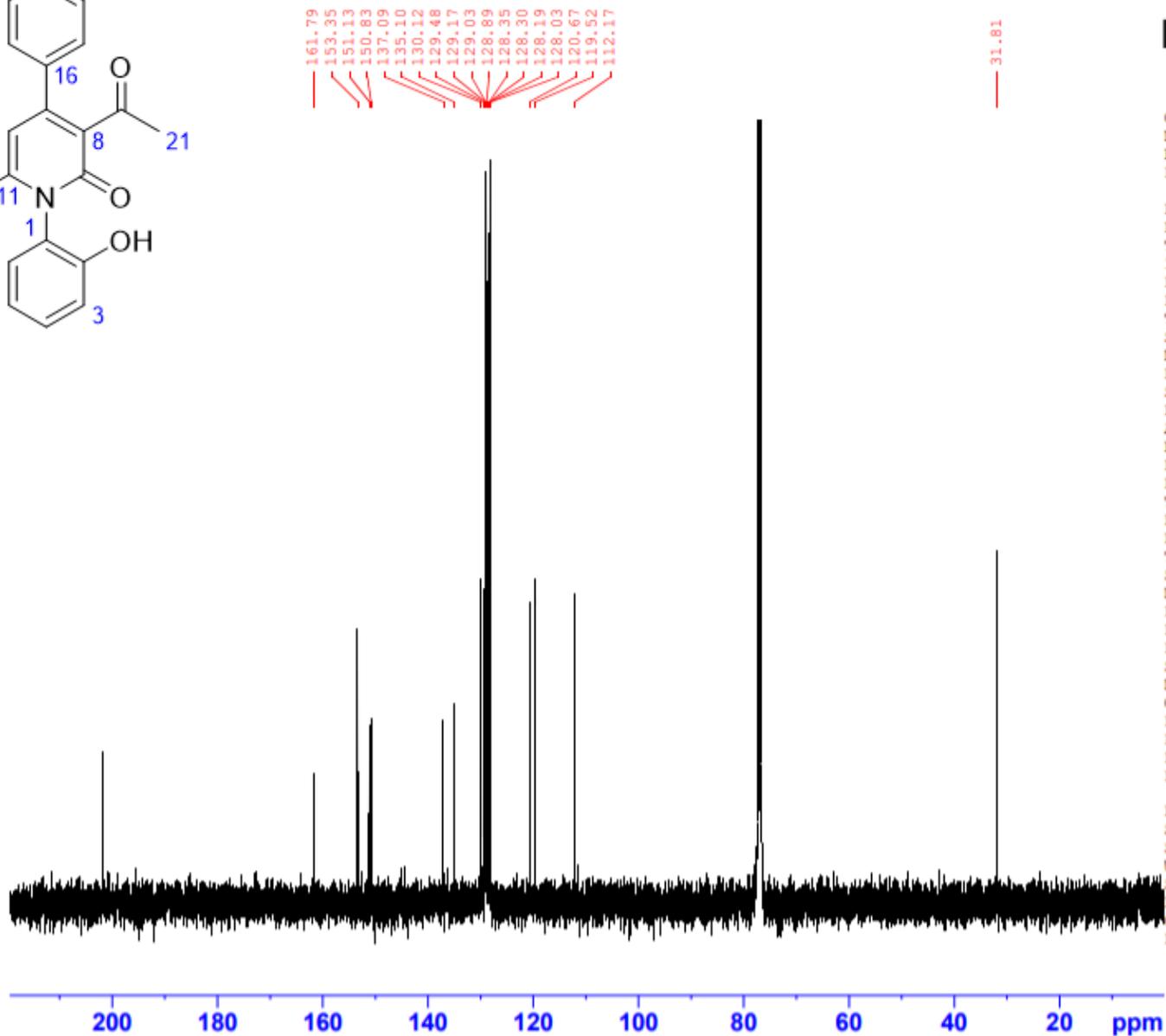
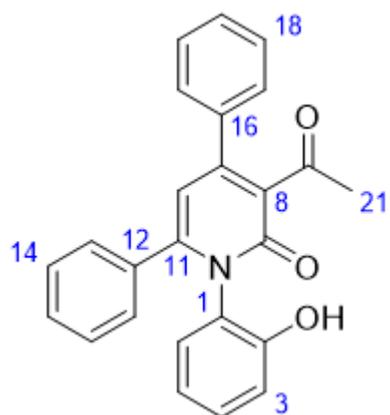
Current Data Parameters
NAME JS-4-286 RECOL
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210507
Time_ 17.10 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100095 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



3e - ¹³C NMR, 101 MHz, CDCl₃

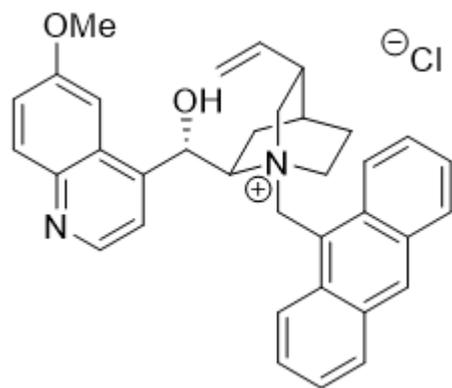


Current Data Parameters
NAME JS-4-286 RECOL
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210508
Time_ 7.08 h
INSTRUM AVIII 400
PROBHD Z108618_0146 ()
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077403 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

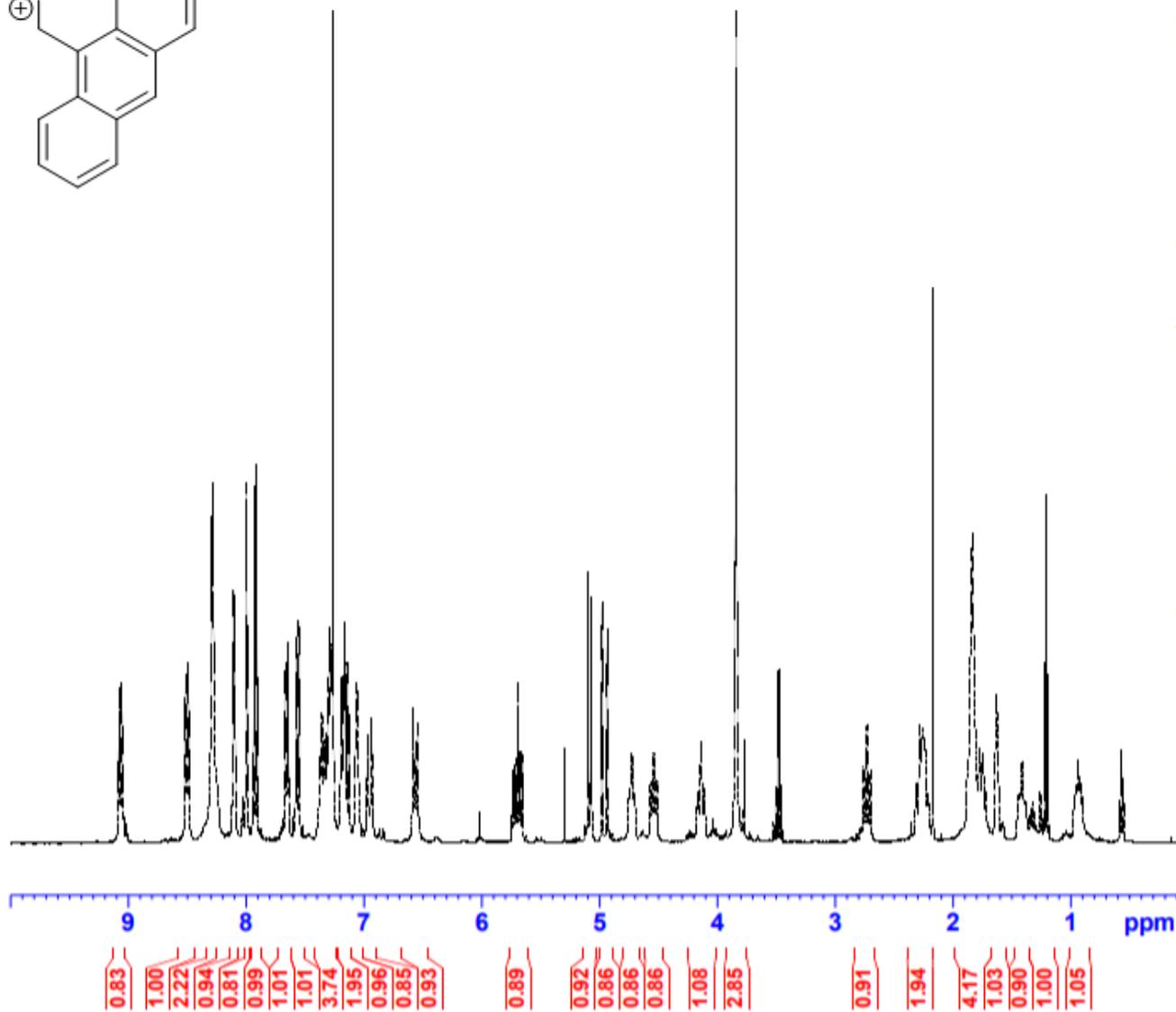
QD-1 - ¹H NMR, 400 MHz, CDCl₃



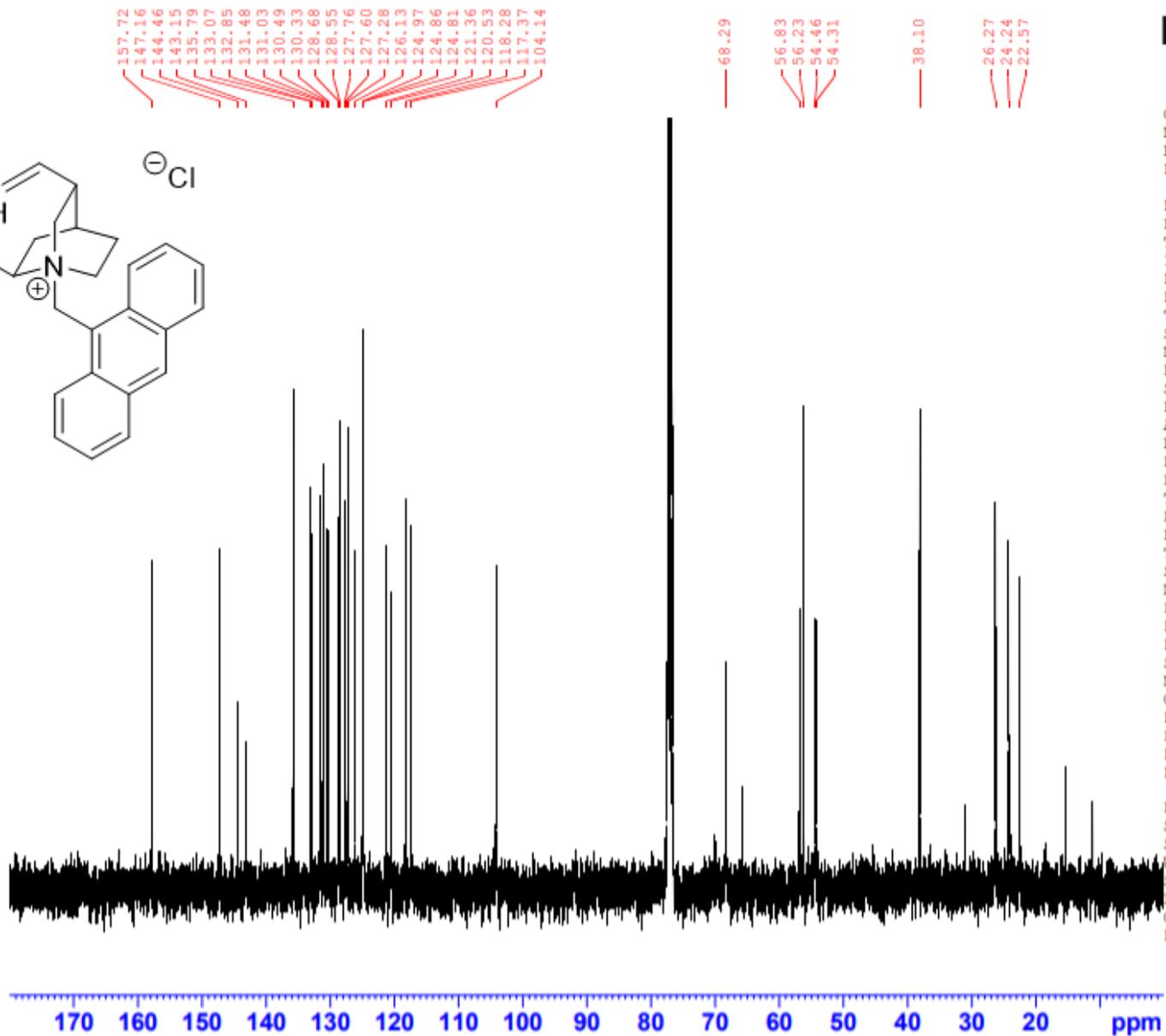
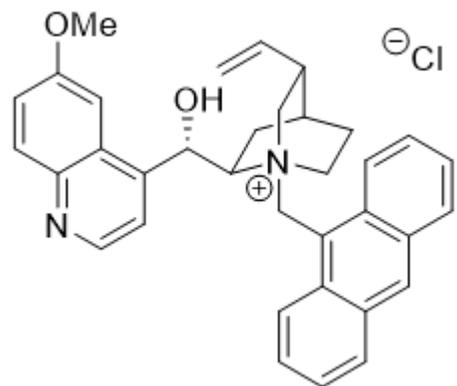
Current Data Parameters
NAME PCK-3-77
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210520
Time_ 17.06 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 128
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100067 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



S149

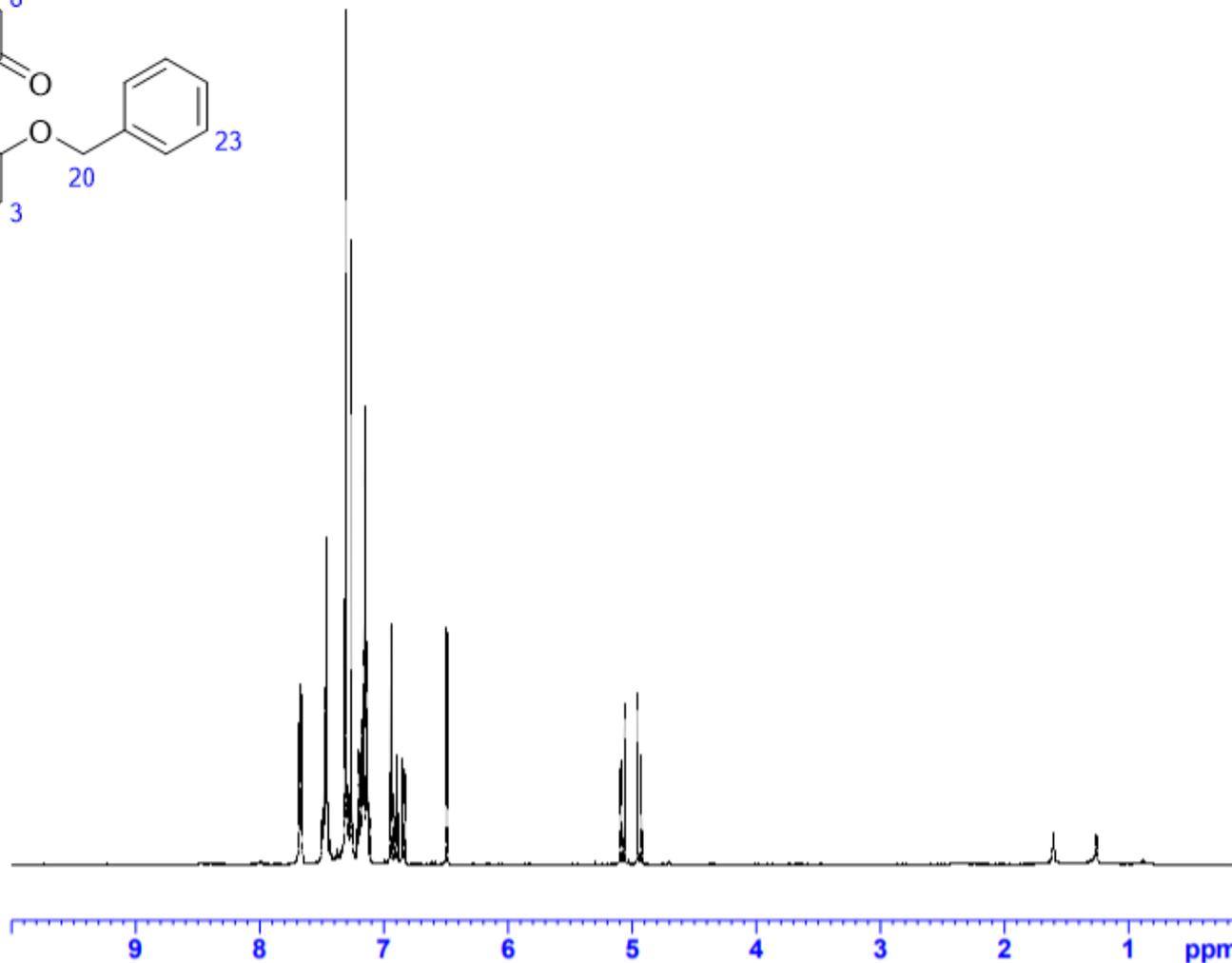
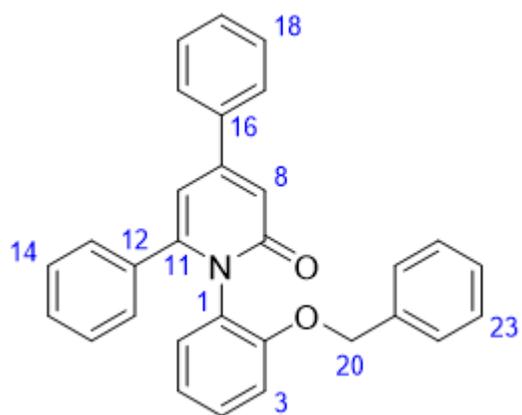


Current Data Parameters
 NAME PCK-3-77
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210520
 Time_ 22.35 h
 INSTRUM AVIII_400
 PROBHD Z108618_0146 ()
 PULPROG zgpg30
 TD 96150
 SOLVENT CDCl3
 NS 4096
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.500020 Hz
 AQ 1.9999200 sec
 RG 2050
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1
 SFO1 100.6178003 MHz
 NUC1 13C
 P0 3.00 usec
 P1 9.00 usec
 PLW1 96.68000031 W
 SFO2 400.1116004 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 17.29199982 W
 PLW12 0.48032999 W
 PLW13 0.24160001 W

F2 - Processing parameters
 SI 131072
 SF 100.6077398 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

4a - ¹H NMR, 400 MHz, CDCl₃



2.03
3.11
6.17
7.06
1.00
1.02
0.99
1.00

2.00

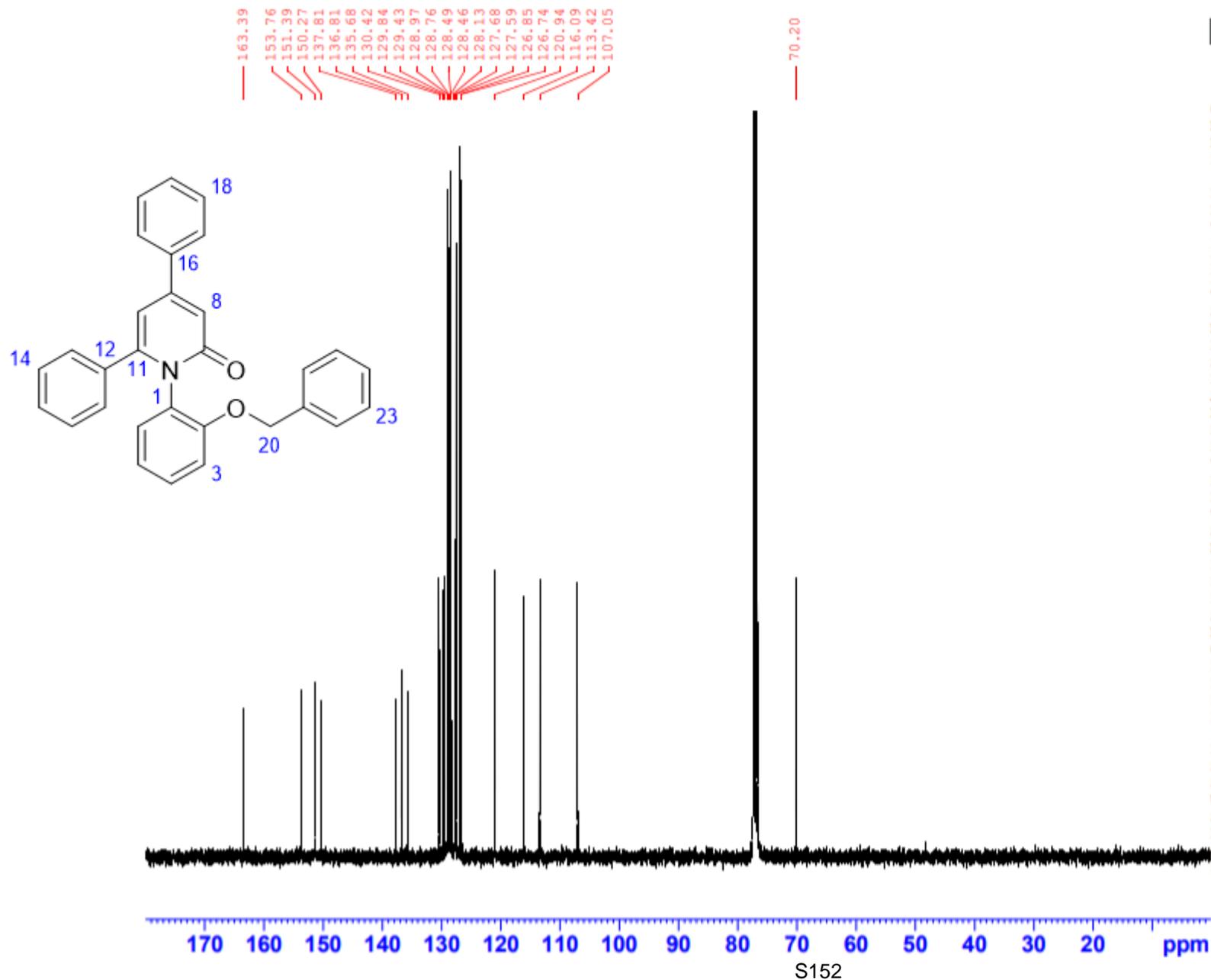


Current Data Parameters
NAME JS-4-289 recol
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210507
Time_ 17.16 h
INSTRUM AVIII_400
PROBHD z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100095 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

4a - ¹³C NMR, 101 MHz, CDCl₃

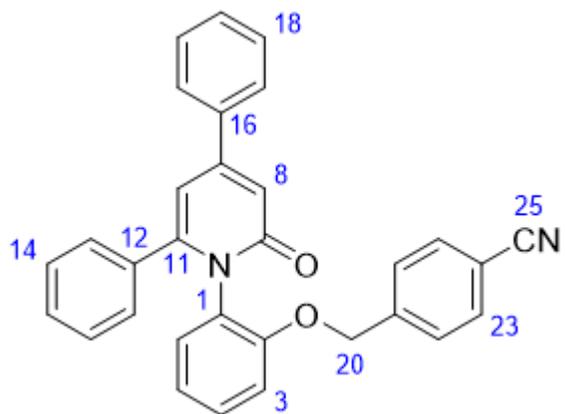


Current Data Parameters
NAME JS-4-289 recol
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210508
Time_ 11.11 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077411 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

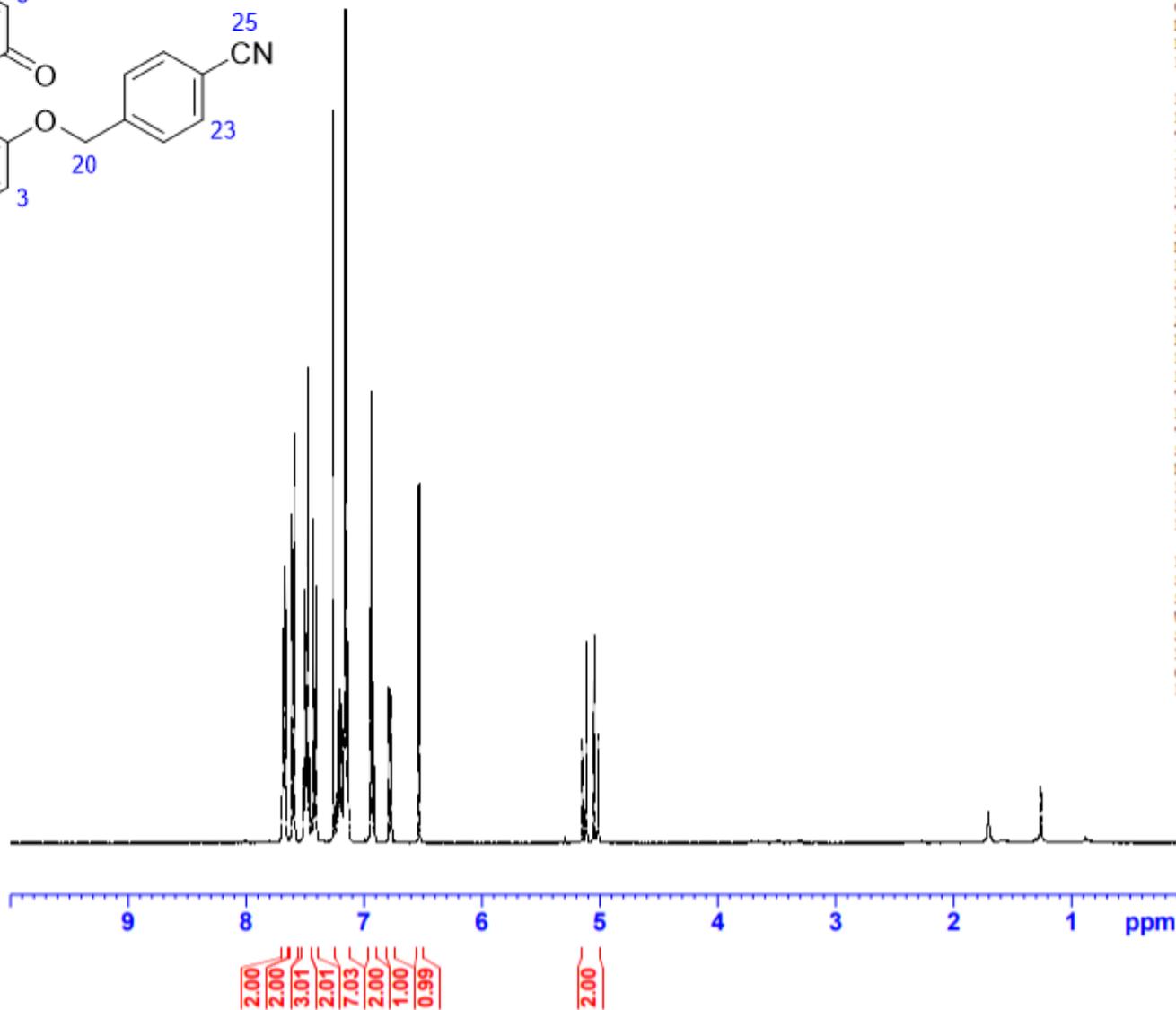
4b - ¹H NMR, 400 MHz, CDCl₃



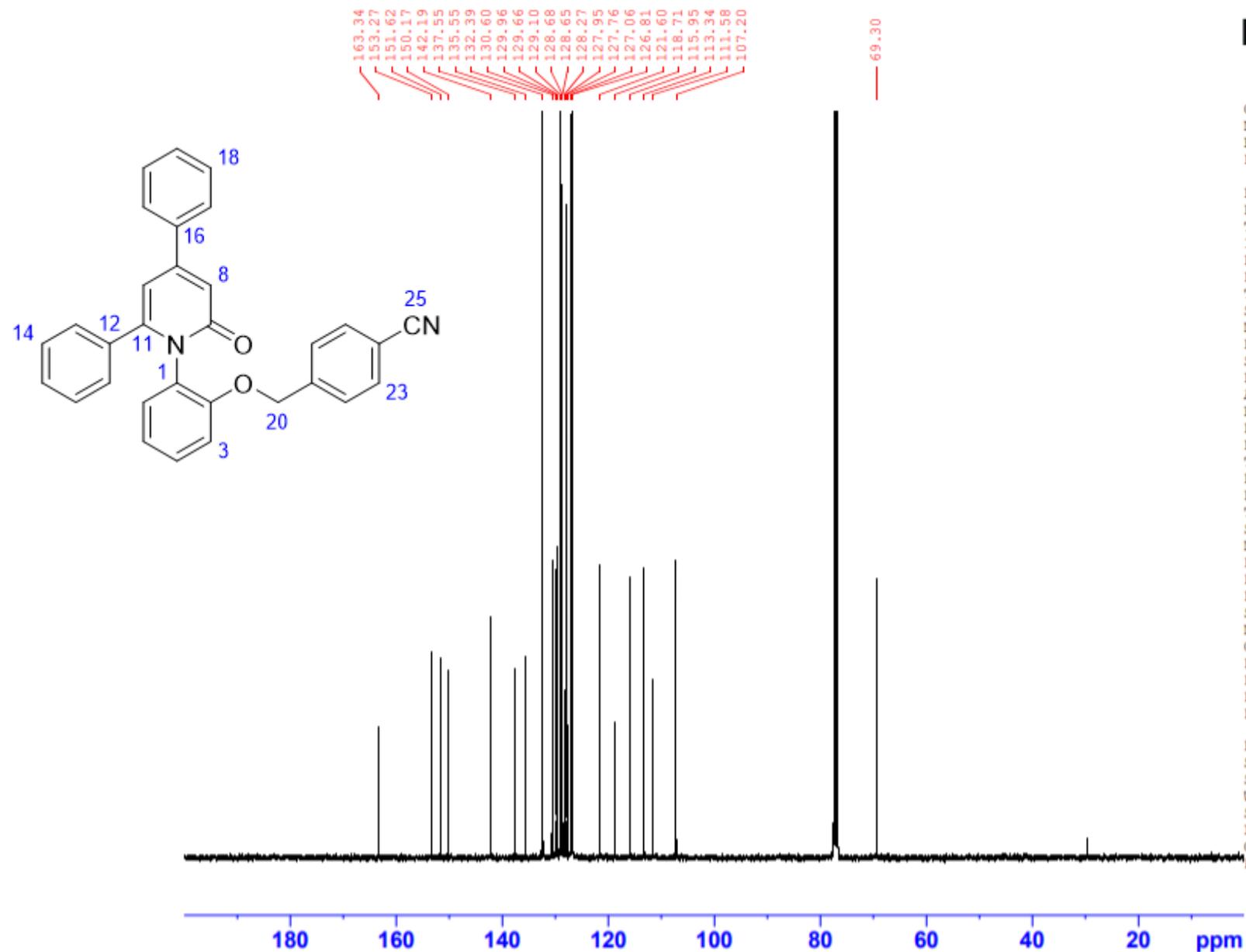
Current Data Parameters
NAME JS-4-298 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210413
Time_ 15.47 h
INSTRUM AVIII 400
PROBHD z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 144
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100099 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4b - ¹³C NMR, 101 MHz, CDCl₃

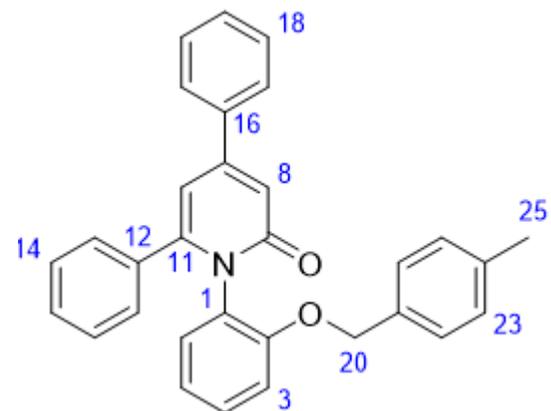


Current Data Parameters
NAME JS-4-298 col
EXPNO 16
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210420
Time_ 3.36 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TDO 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

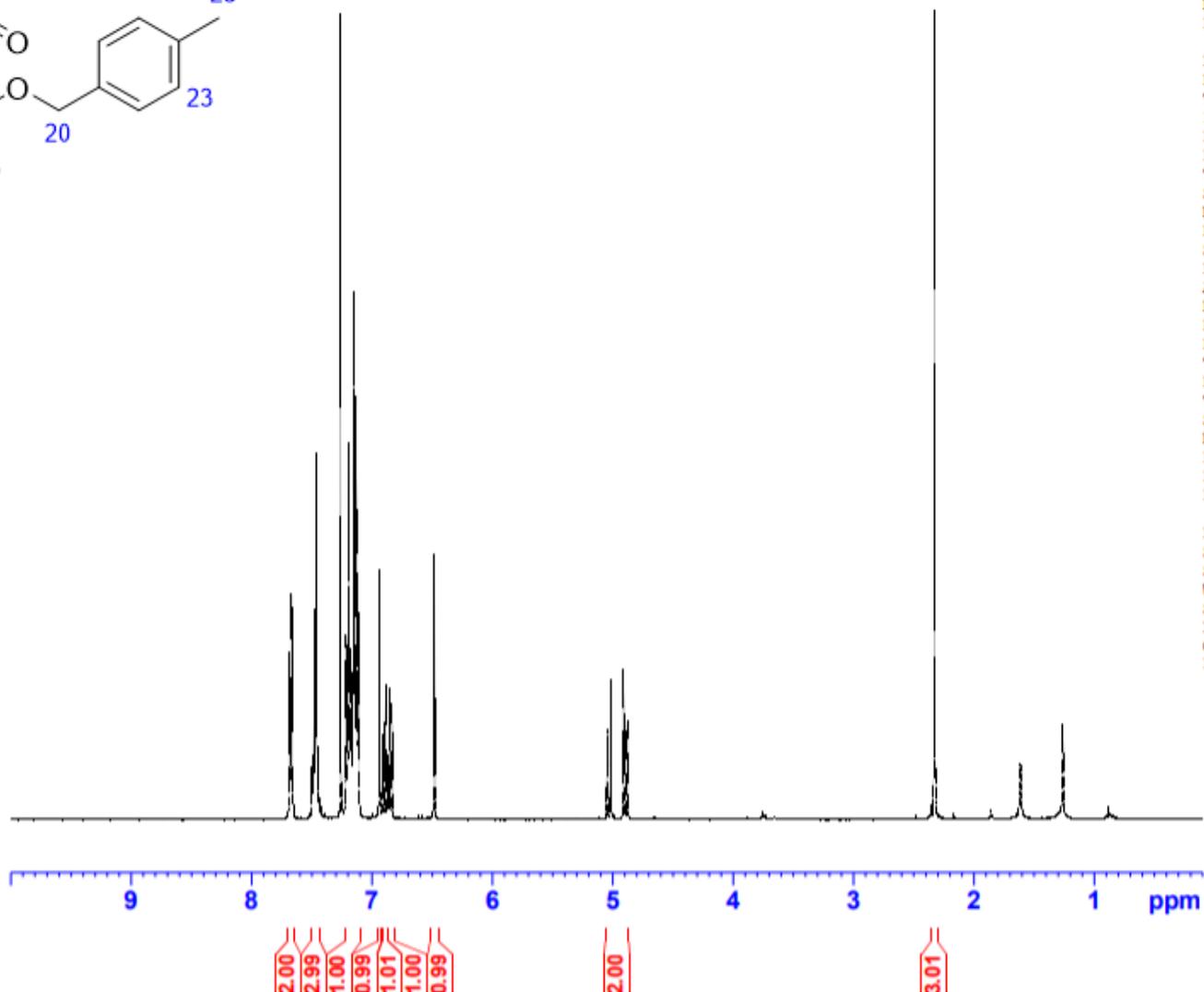
4c - ¹H NMR, 400 MHz, CDCl₃



Current Data Parameters
NAME JS-4-303 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210416
Time 10.29 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4c - ¹³C NMR, 101 MHz, CDCl₃



Current Data Parameters
NAME JS-4-303 col
EXPNO 11
PROCNO 1

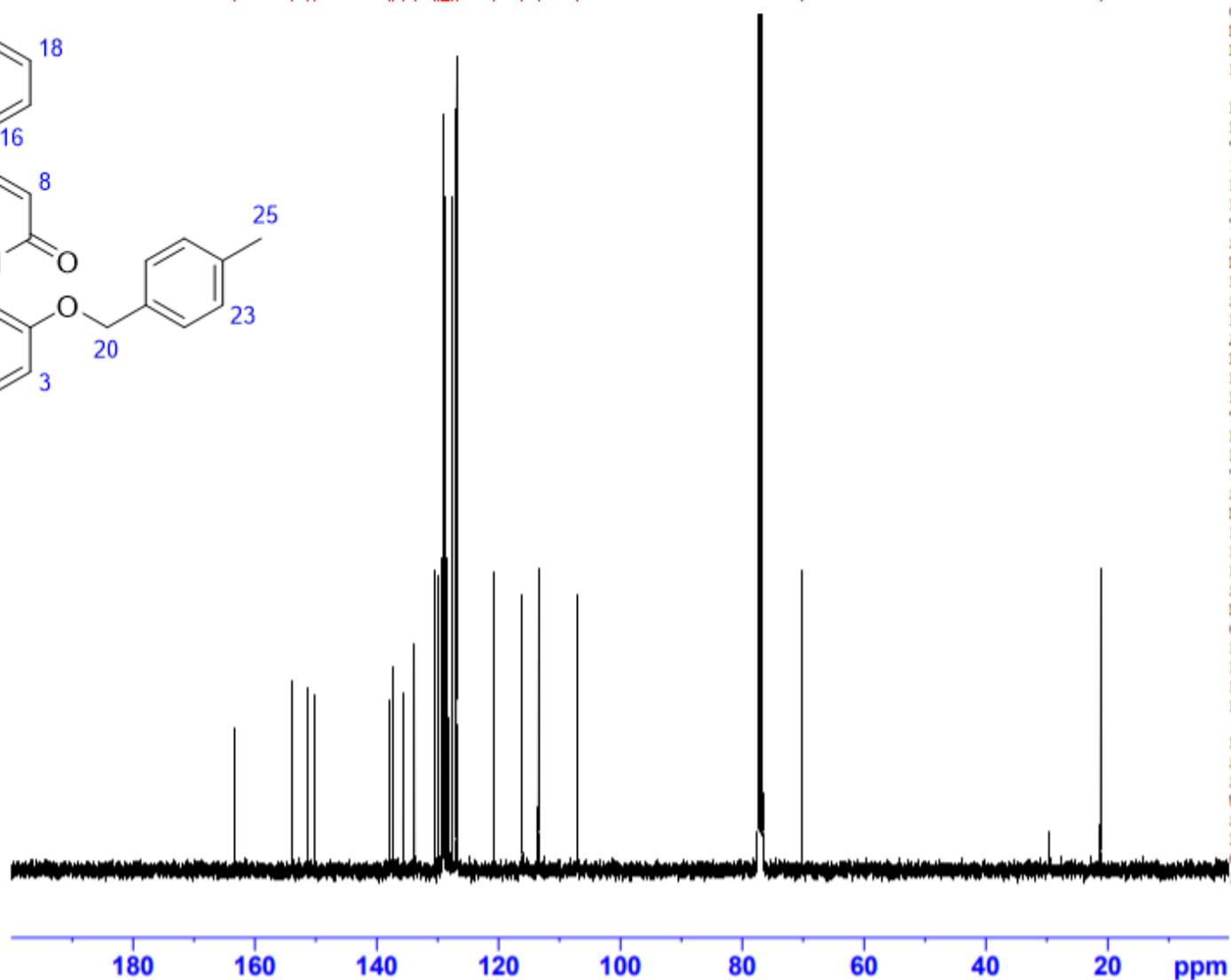
F2 - Acquisition Parameters
Date_ 20210416
Time_ 22.35 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

163.39
153.89
151.34
150.27
137.83
137.40
135.71
133.77
130.38
129.83
129.42
129.14
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128.48
128.11
127.58
126.90
126.86
120.85
116.09
113.45
107.03

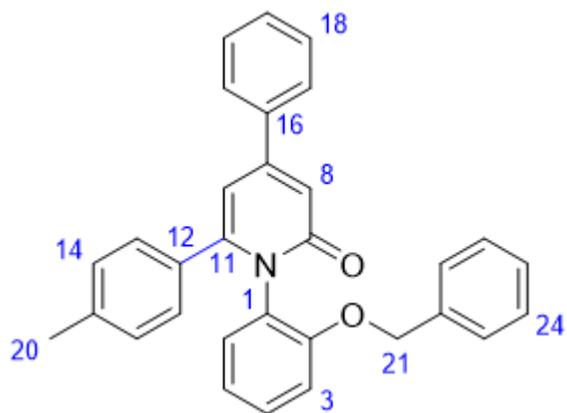
70.22

21.18



S156

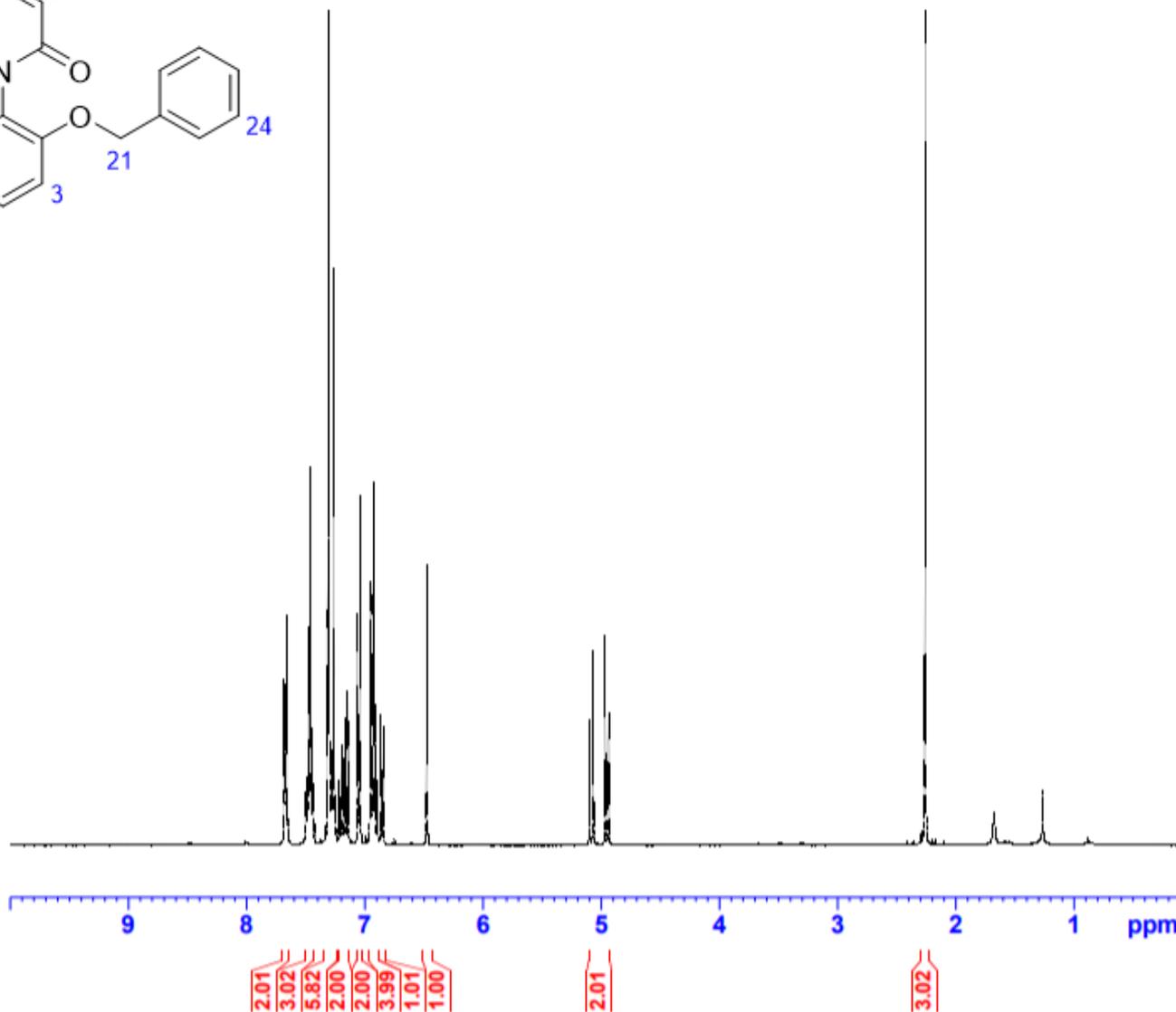
4d - ¹H NMR, 400 MHz, CDCl₃



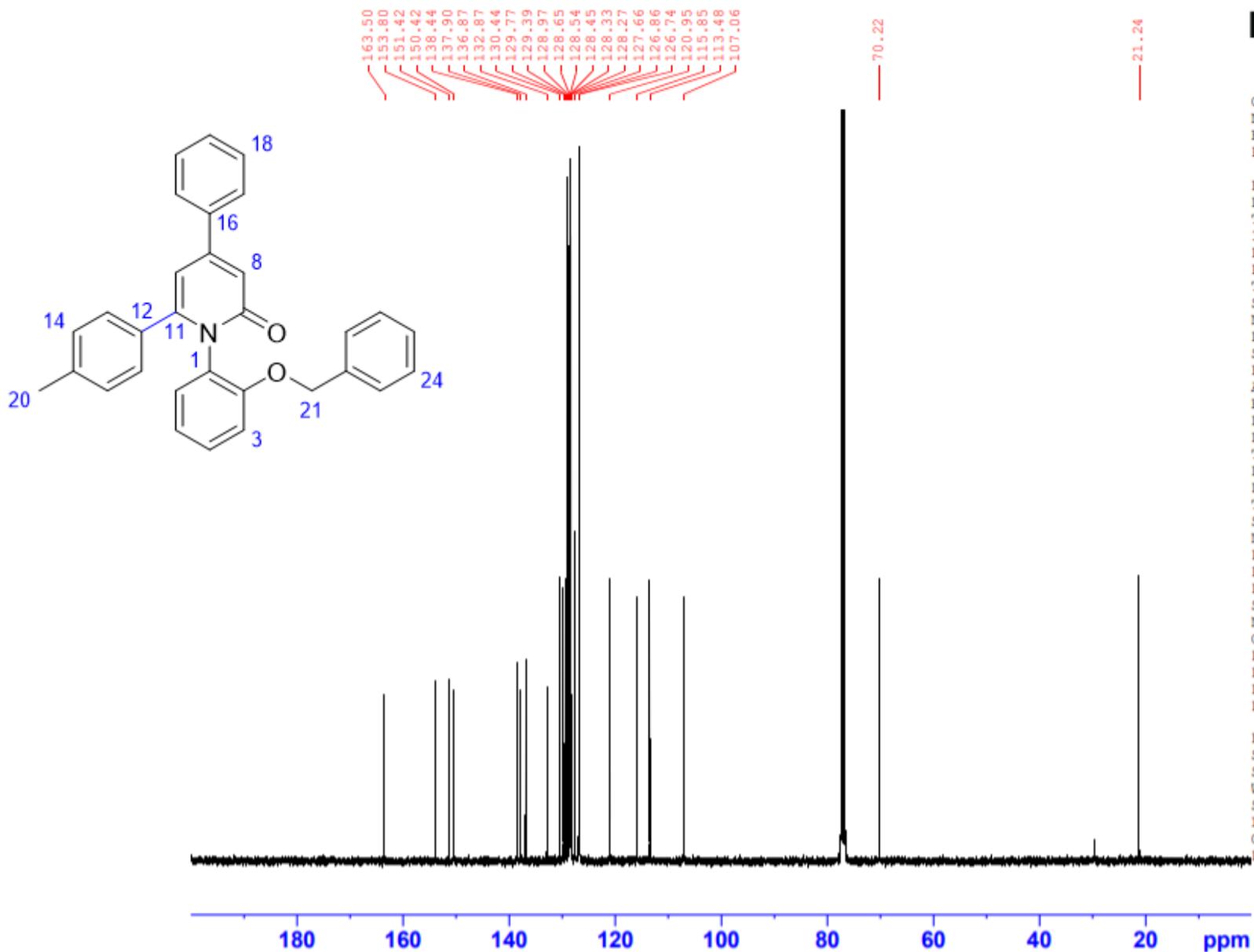
Current Data Parameters
NAME JS-4-300 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210414
Time 15.25 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 161
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.112478 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.110098 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4d - ¹³C NMR, 101 MHz, CDCl₃

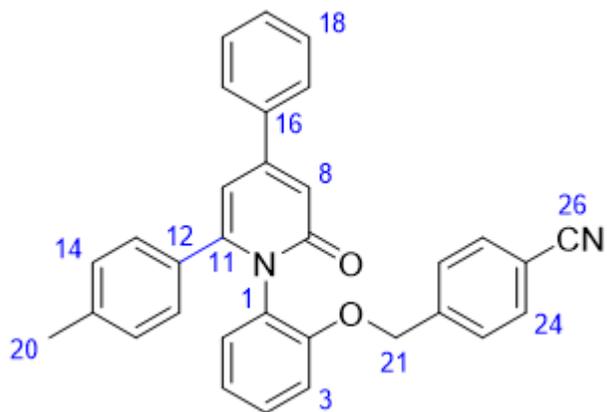


Current Data Parameters
NAME JS-4-300 col
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210414
Time 22.44 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

4e - ¹H NMR, 400 MHz, CDCl₃

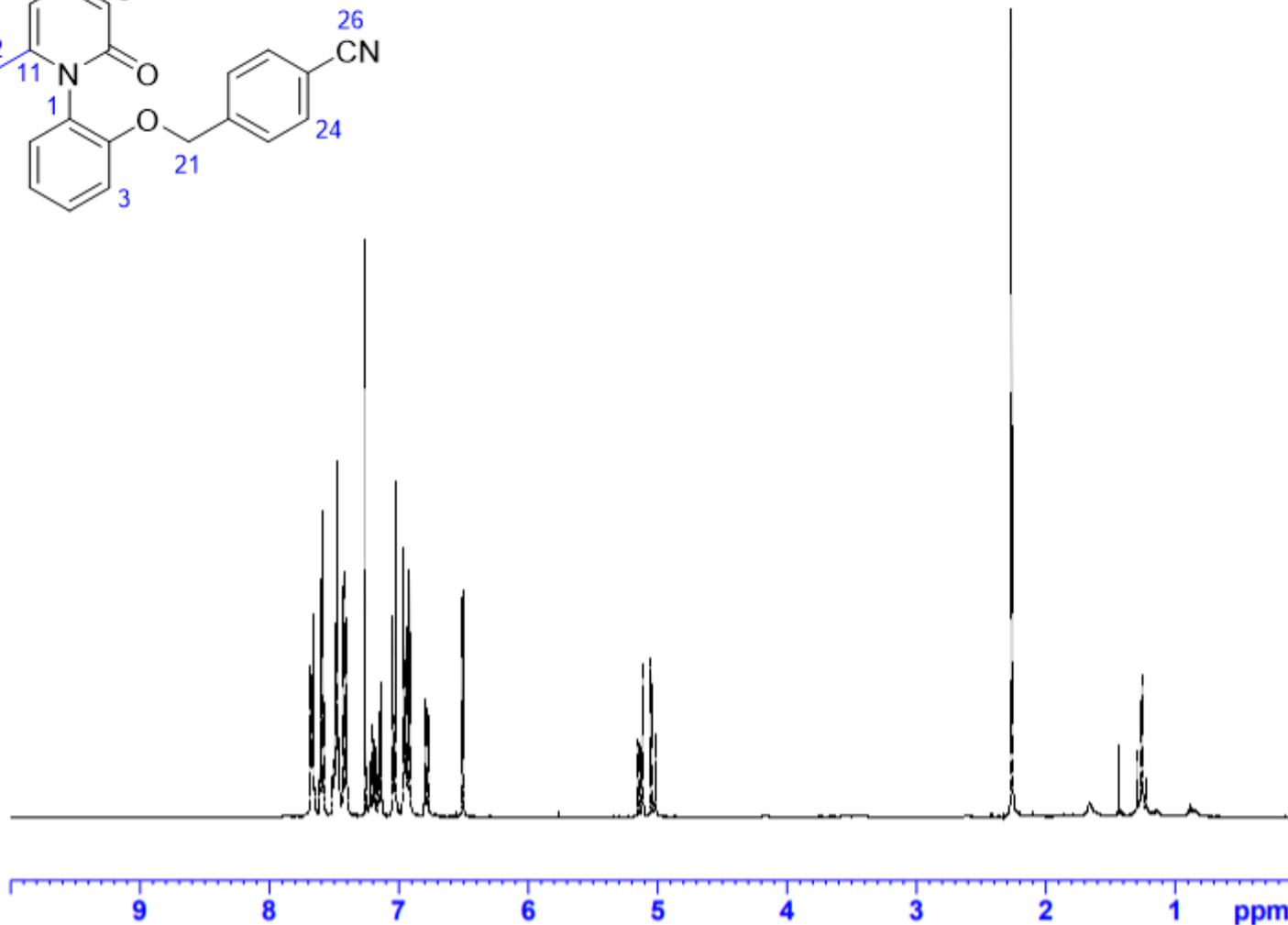


Current Data Parameters
NAME JS-4-292 hivac
EXPNO 12
PROCNO 1

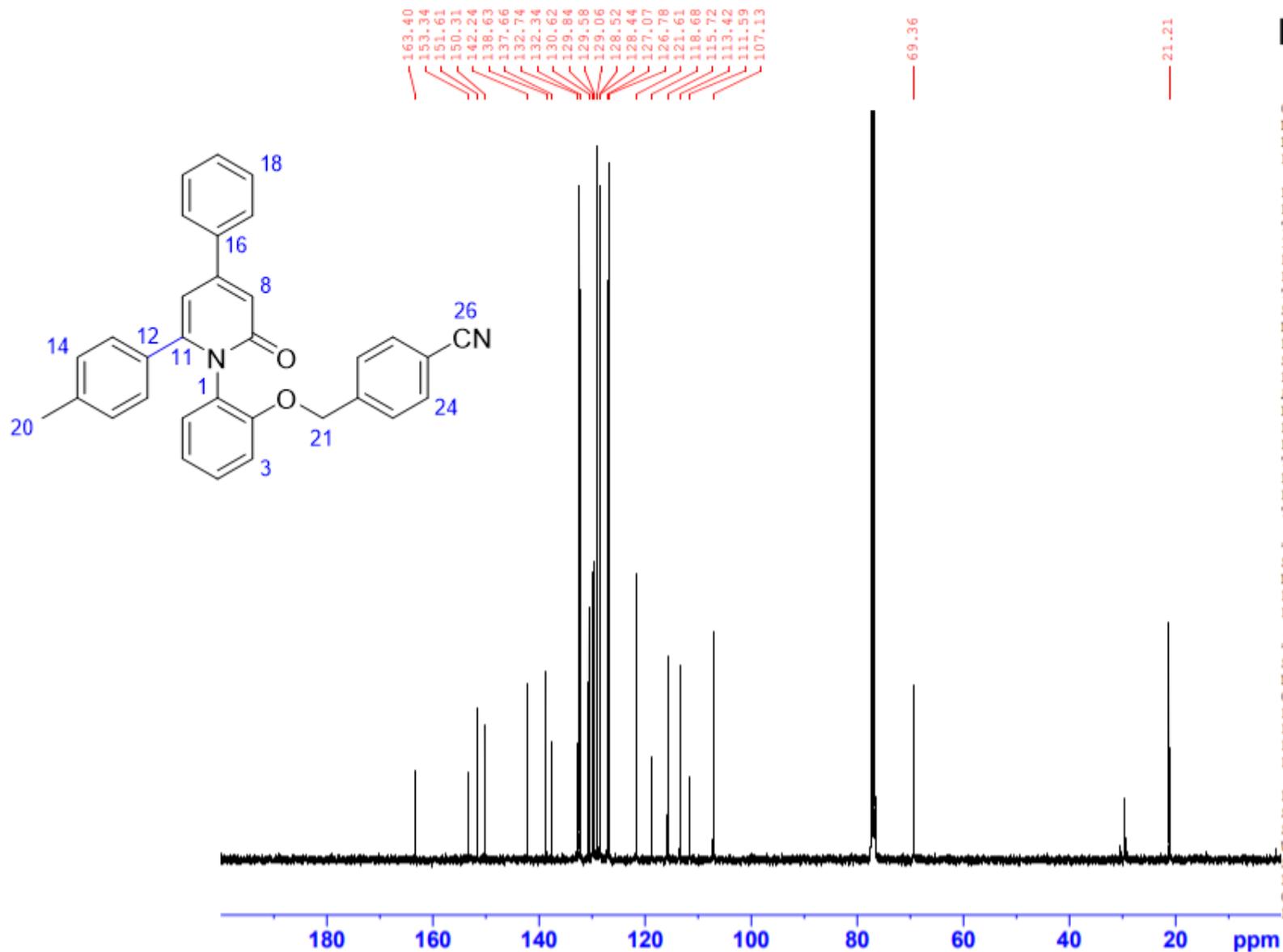
F2 - Acquisition Parameters
Date_ 20210408
Time_ 23.38
INSTRUM AVIII 400
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 80.6
DW 60.800 usec
DE 6.50 usec
TE 295.9 K
D1 1.00000000 sec
TD0 1

----- CHANNEL f1 -----
SF01 399.9124696 MHz
NUC1 1H
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 399.9100098 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4e - ¹³C NMR, 101 MHz, CDCl₃



Current Data Parameters
NAME JS-4-292 hivac
EXPNO 11
PROCNO 1

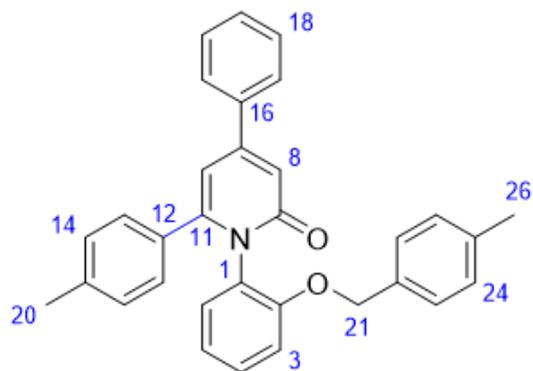
F2 - Acquisition Parameters
Date_ 20210408
Time 23.34
INSTRUM AVIII_400
PROBHD 5 mm PABBO_BB/
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.250010 Hz
AQ 1.9999200 sec
RG 80.6
DW 20.800 usec
DE 6.50 usec
TE 300.1 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

----- CHANNEL f1 -----
SFO1 100.5675047 MHz
NUC1 13C
P1 9.00 usec
PLW1 96.68000031 W

----- CHANNEL f2 -----
SFO2 399.9115996 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.38907000 W

F2 - Processing parameters
SI 131072
SF 100.5574500 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

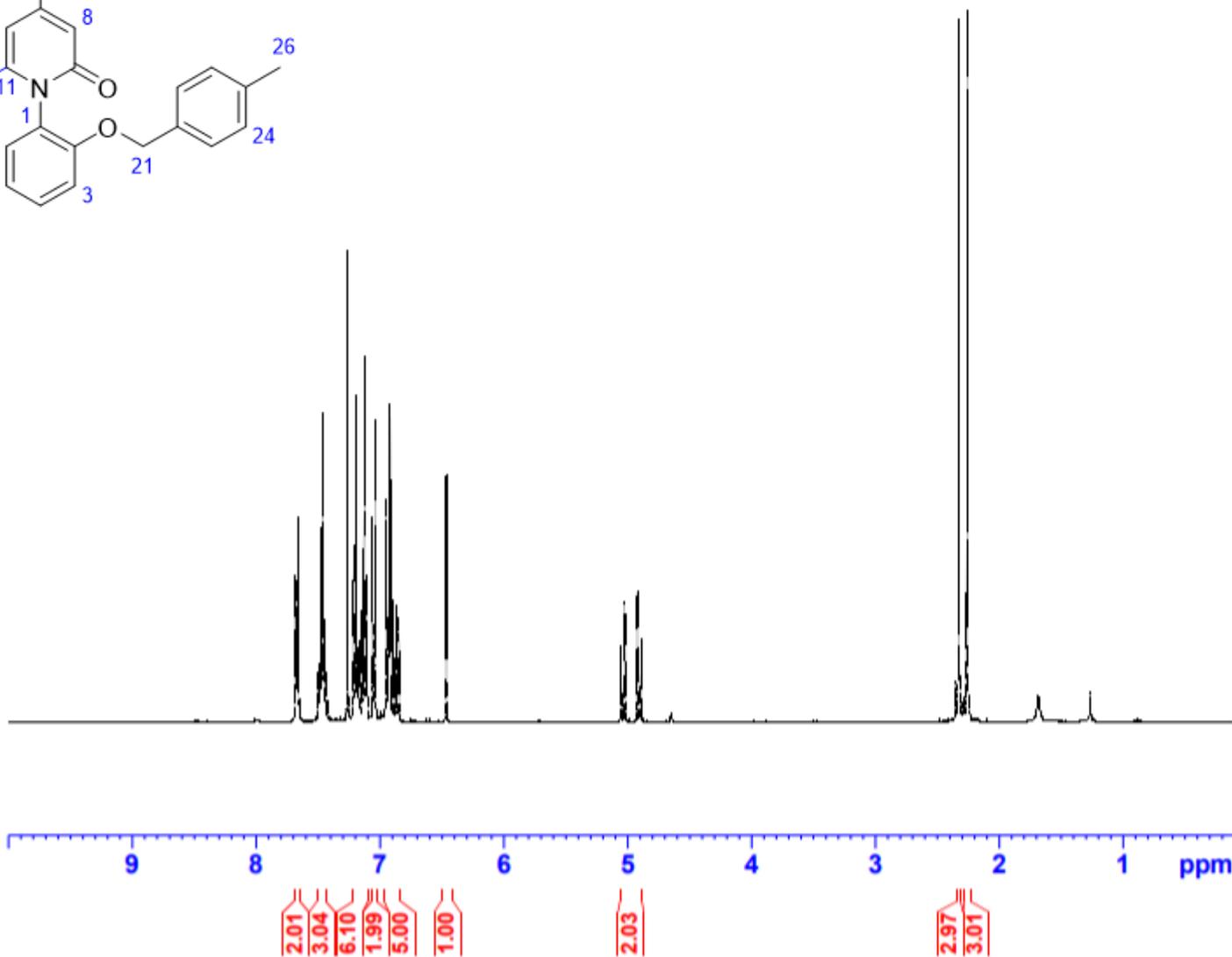
4f - ¹H NMR, 400 MHz, CDCl₃



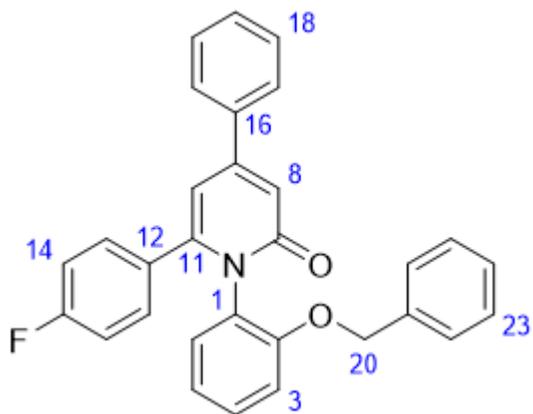
Current Data Parameters
NAME JS-4-296
EXPNO 14
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210413
Time 13.15 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 181
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100099 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



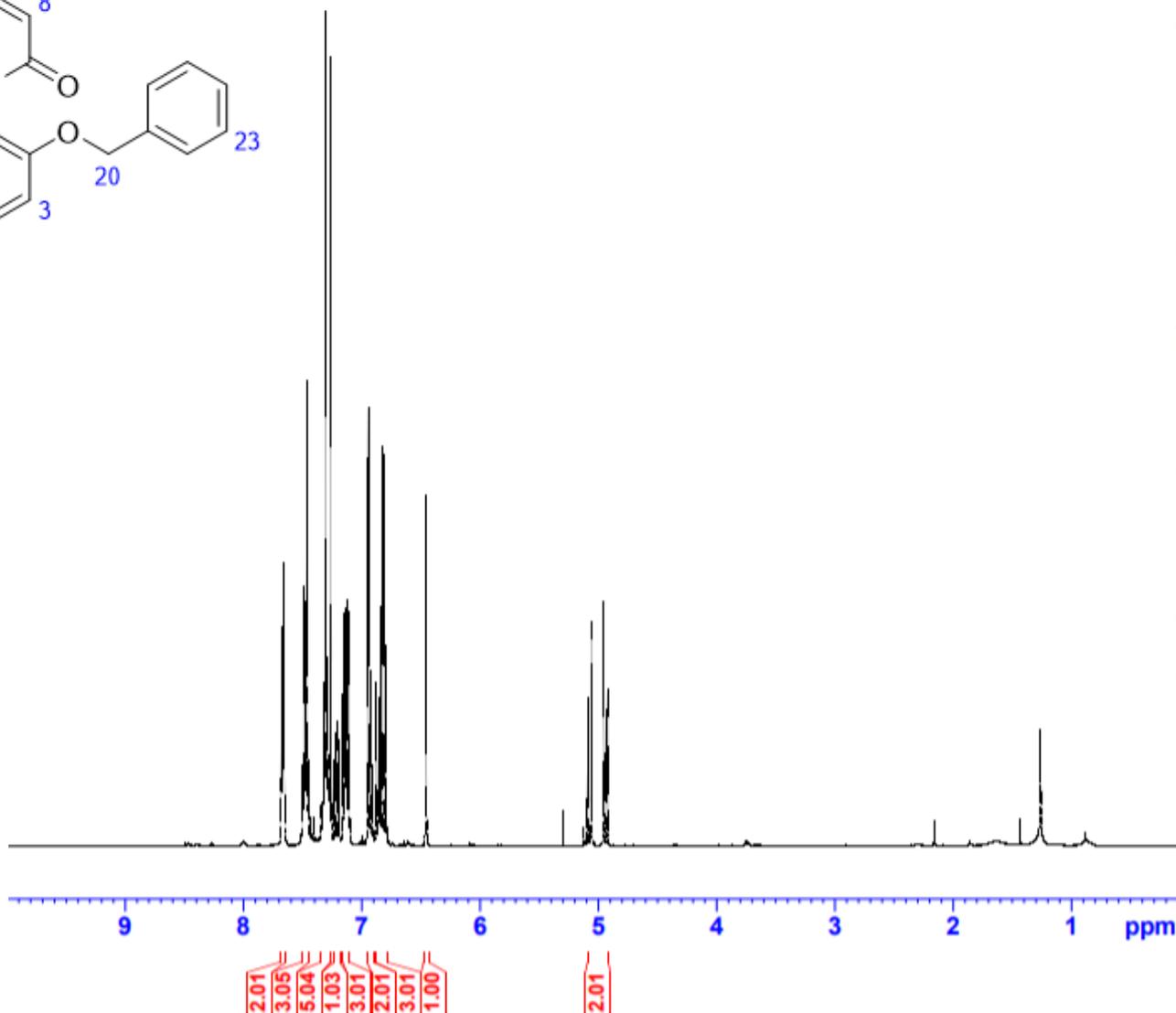
4g - ¹H NMR, 400 MHz, CDCl₃



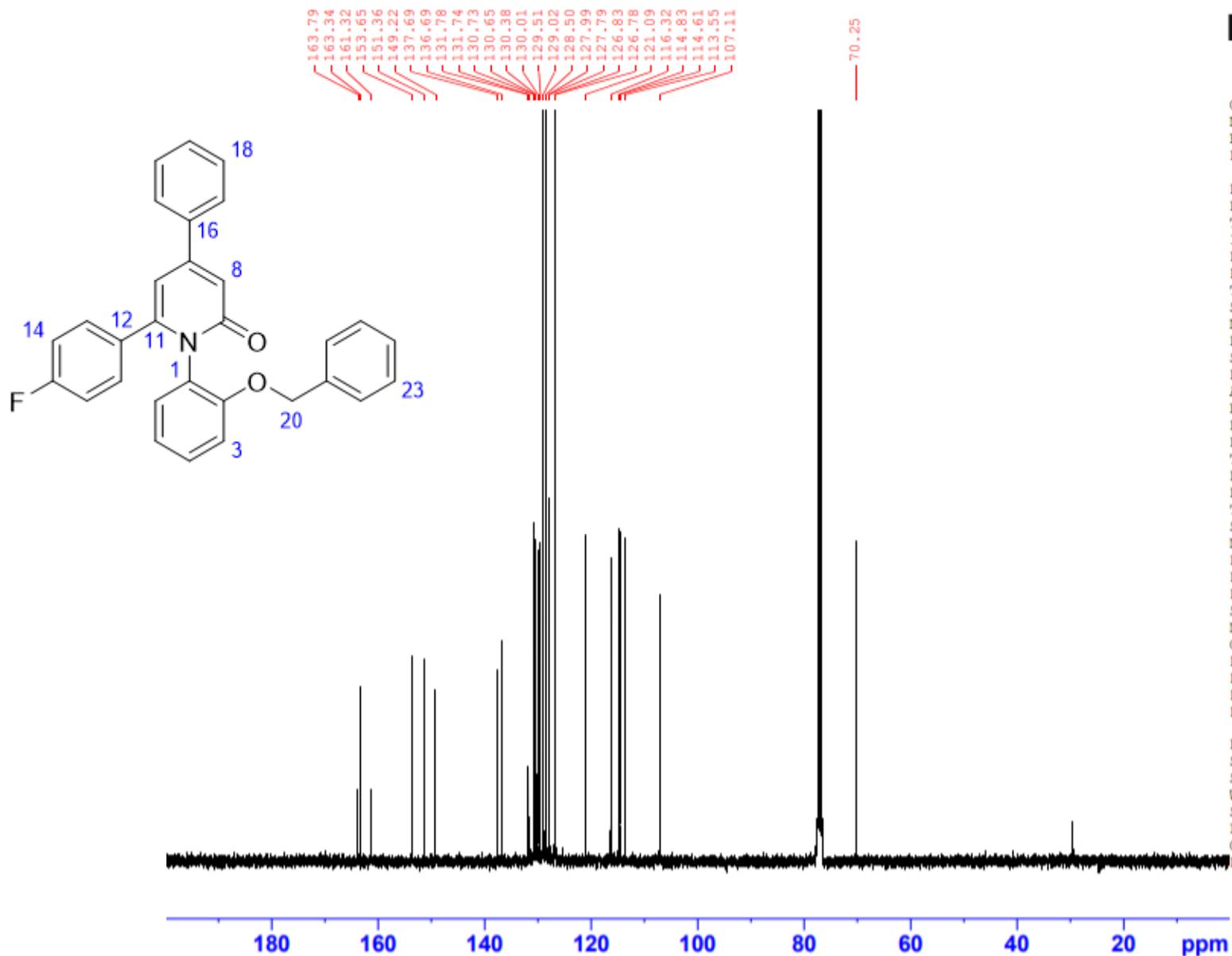
Current Data Parameters
NAME JS-4-301 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210415
Time 14.07 h
INSTRUM AVIII_400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4g - ¹³C NMR, 101 MHz, CDCl₃

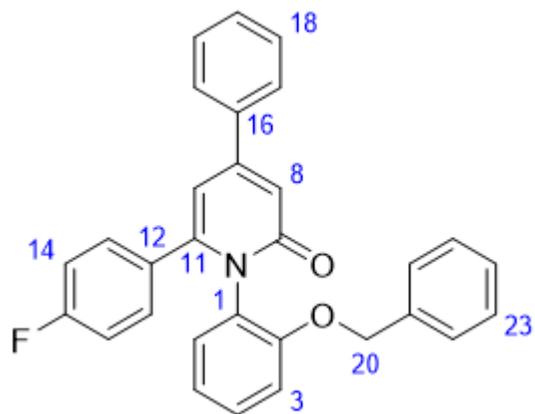


Current Data Parameters
NAME JS-4-301 col
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210415
Time 22.34 h
INSTRUM AVIII 400
PROBHD Z108618_0146 ()
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

4g - ¹⁹F NMR, 376 MHz, CDCl₃



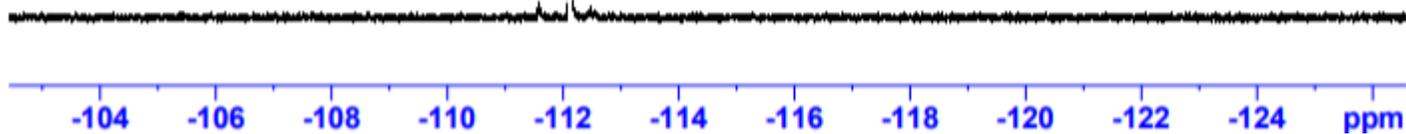
-112.13



Current Data Parameters
NAME JS-4-301 col
EXPNO 25
PROCNO 1

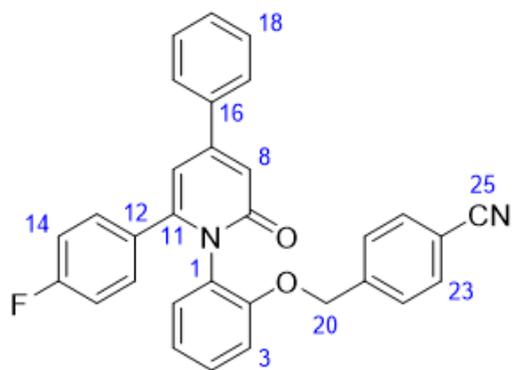
F2 - Acquisition Parameters
Date_ 20210505
Time_ 14.13 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg
TD 261992
SOLVENT CDCl3
NS 16
DS 4
SWH 89285.711 Hz
FIDRES 0.681591 Hz
AQ 1.4671552 sec
RG 575
DW 5.600 usec
DE 7.11 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 376.4418995 MHz
NUC1 19F
P1 11.80 usec
PLW1 32.96500015 W

F2 - Processing parameters
SI 262144
SF 376.4795470 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



S165

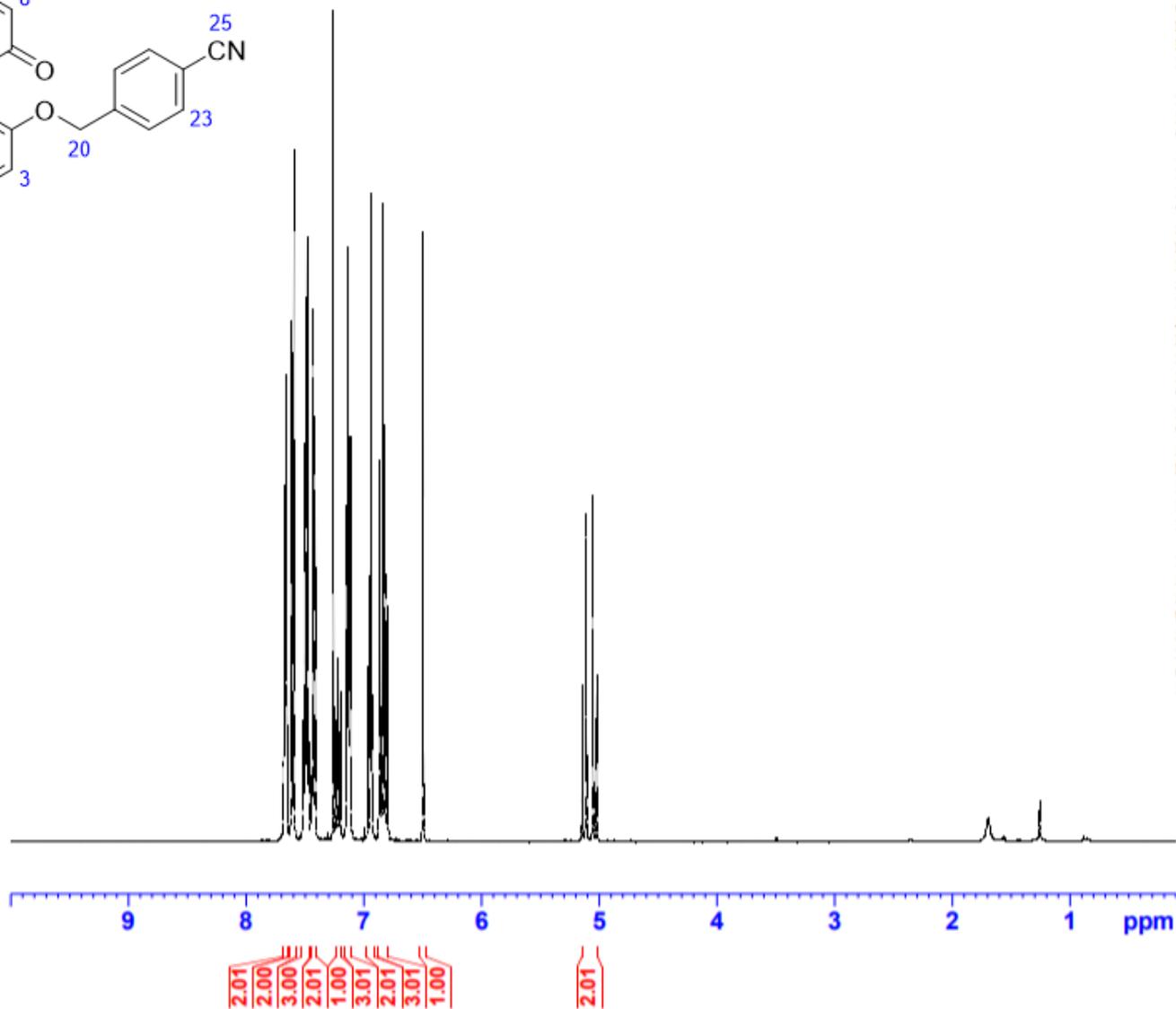
4h - ¹H NMR, 400 MHz, CDCl₃



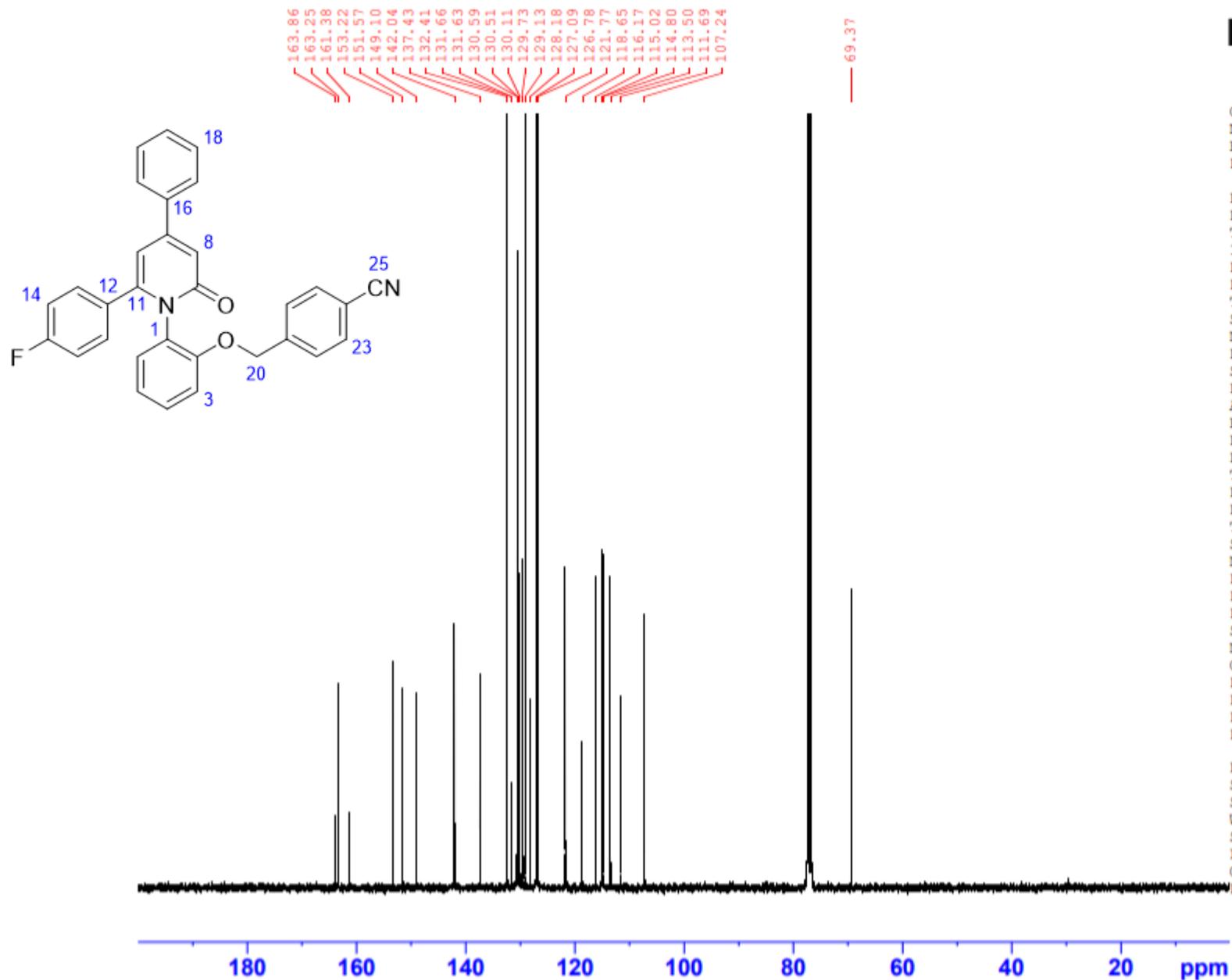
Current Data Parameters
NAME JS-4-293 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210331
Time_ 15.06 h
INSTRUM AVIII 400
PROBHD z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 161
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100104 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4h - ¹³C NMR, 101 MHz, CDCl₃



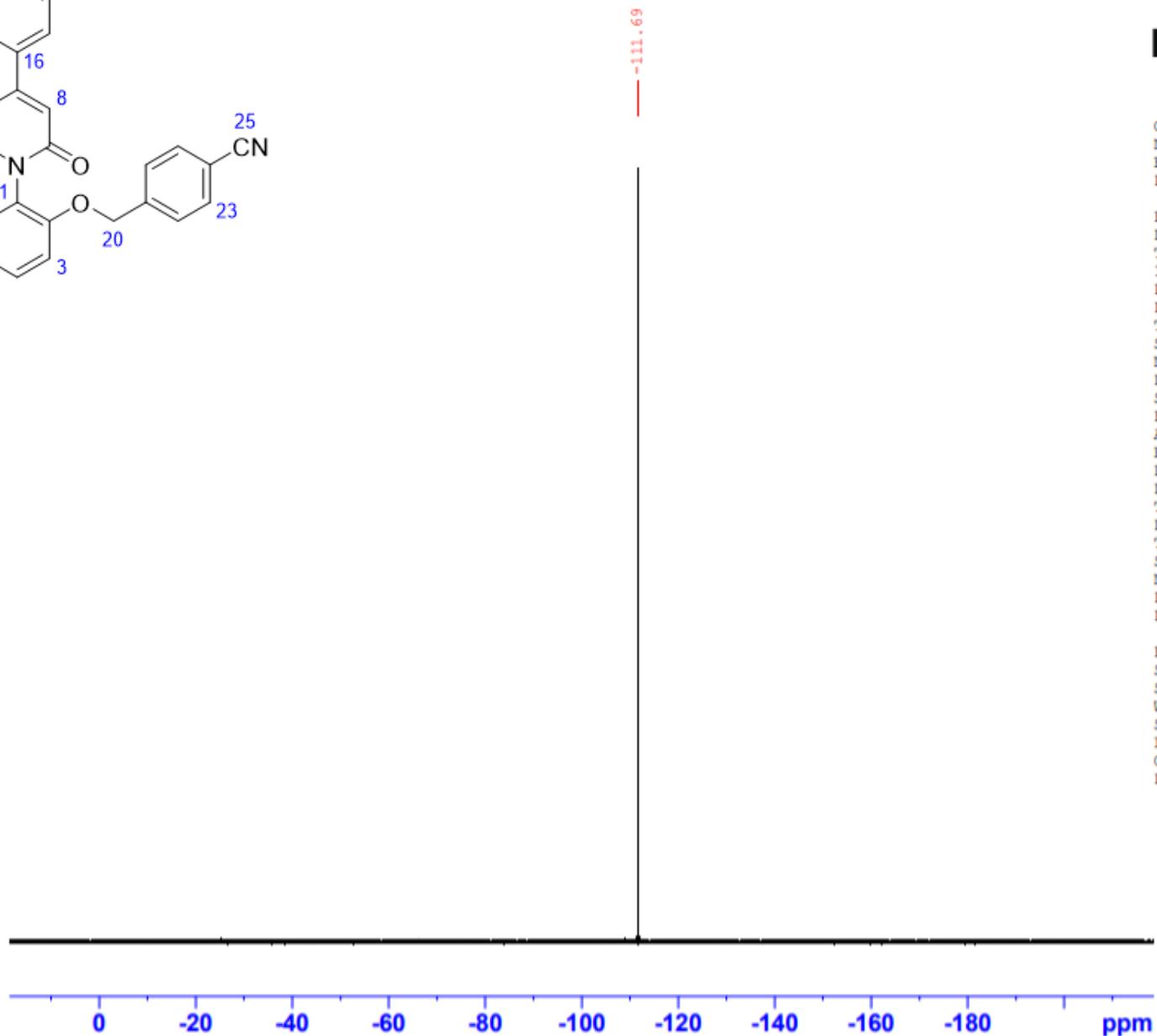
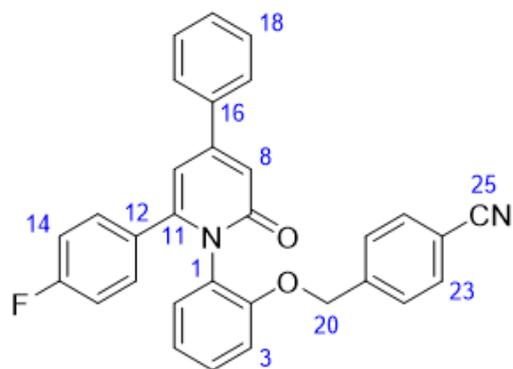
Current Data Parameters
NAME JS-4-293 col
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210331
Time_ 22.35 h
INSTRUM AVIII 400
PROBHD z108618_0146 ()
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

S167

4h - ¹⁹F NMR, 376 MHz, CDCl₃



Current Data Parameters
NAME JS-4-293 col
EXPNO 18
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210505
Time_ 14.07 h
INSTRUM AVIII 400
PROBHD Z108618_0146 ()
PULPROG zg
TD 261992
SOLVENT CDCl3
NS 16
DS 4
SWH 89285.711 Hz
FIDRES 0.681591 Hz
AQ 1.4671552 sec
RG 645
DW 5.600 usec
DE 7.11 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 376.4418995 MHz
NUC1 19F
P1 11.80 usec
PLW1 32.96500015 W

F2 - Processing parameters
SI 262144
SF 376.4795470 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

4i - ¹H NMR, 400 MHz, CDCl₃

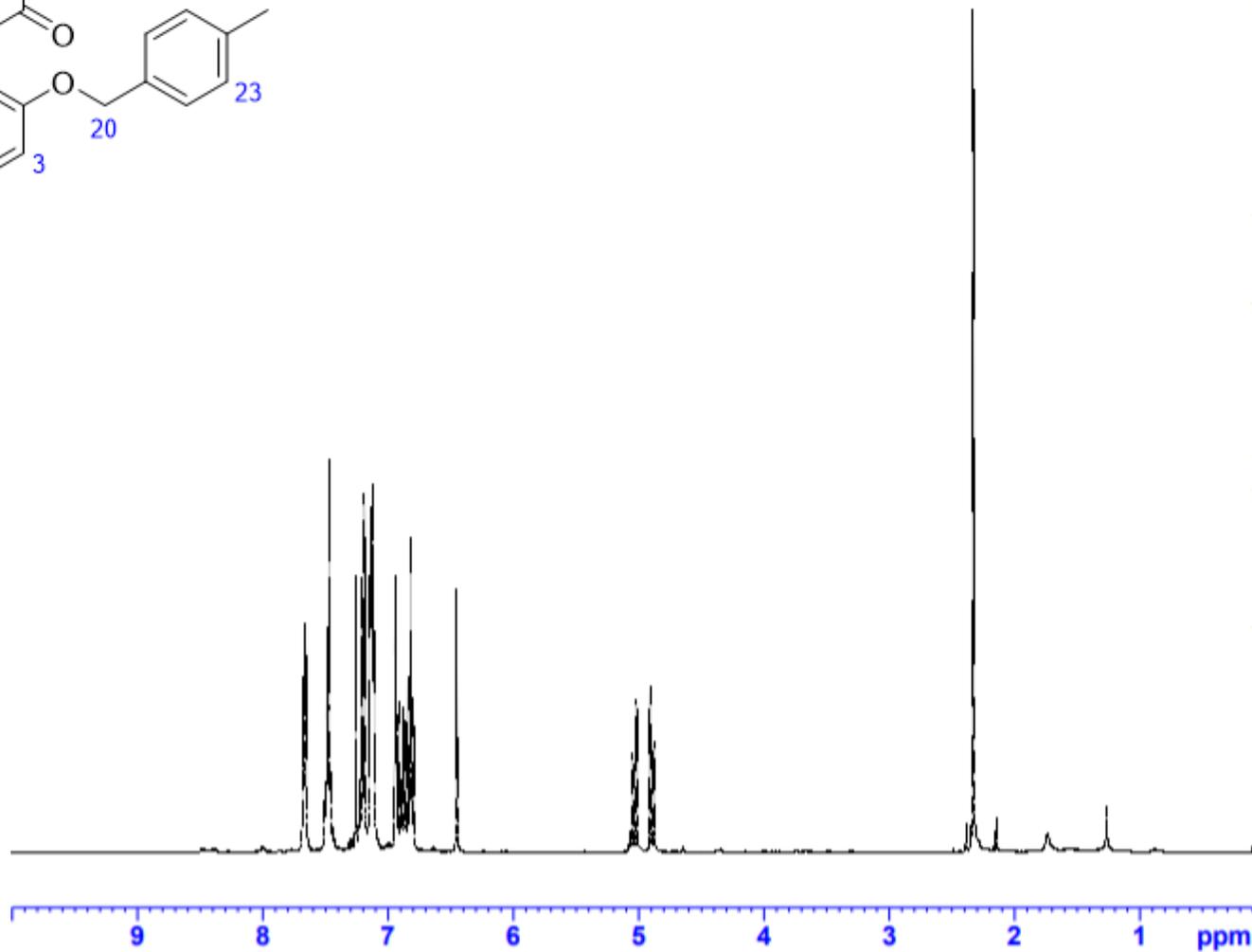
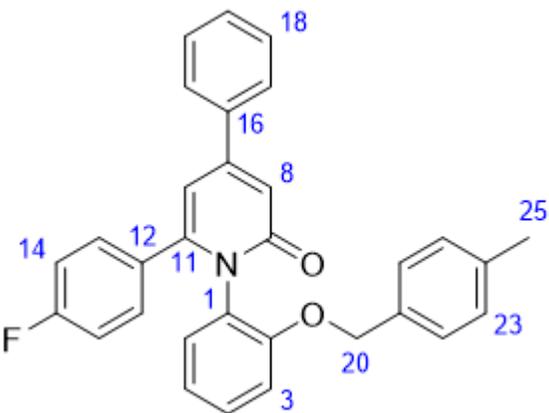


Current Data Parameters
NAME JS-4-297
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210411
Time_ 18.10
INSTRUM AVIII_400
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 57
DW 60.800 usec
DE 6.50 usec
TE 294.9 K
D1 1.00000000 sec
TD0 1

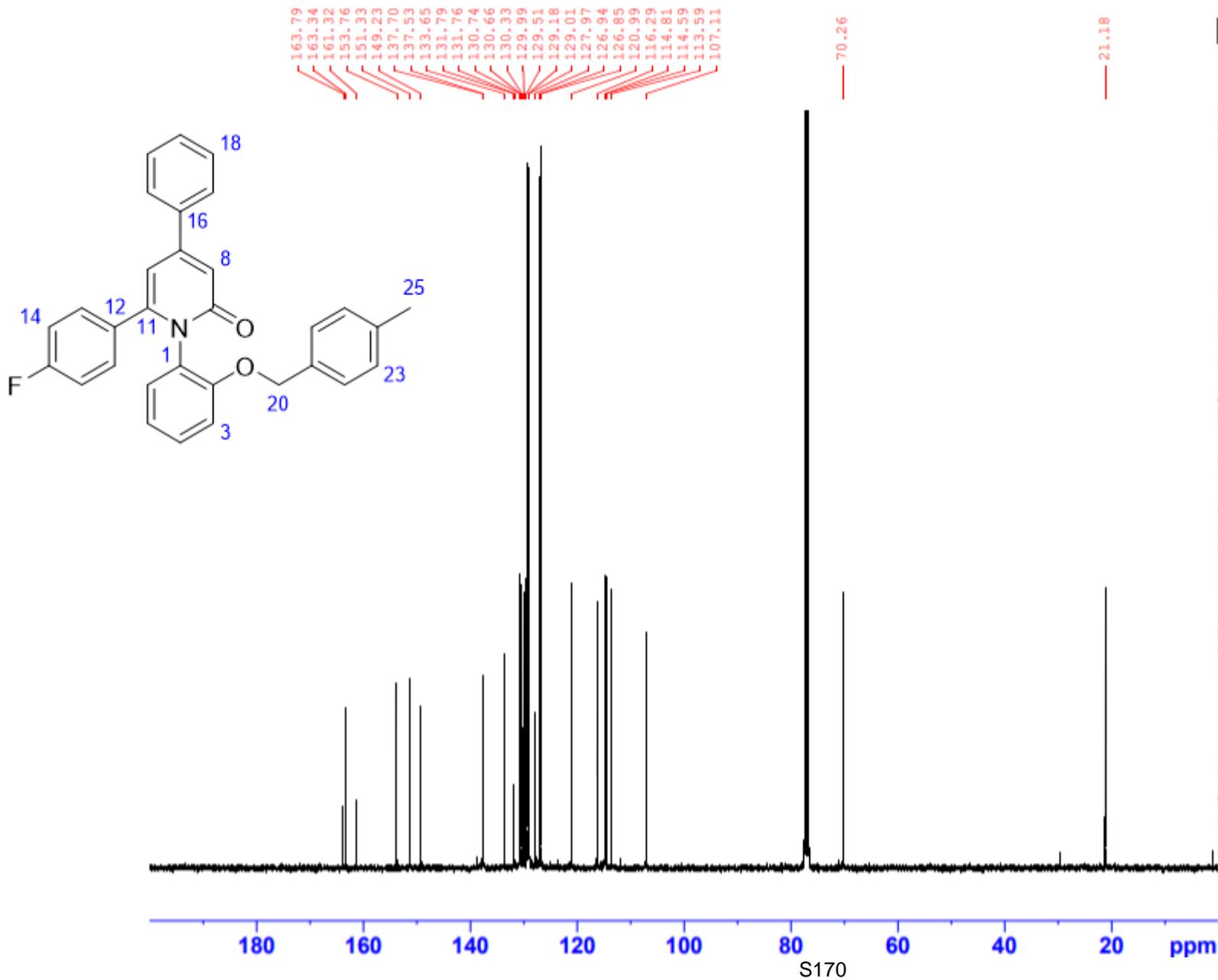
----- CHANNEL f1 -----
SFO1 399.9124696 MHz
NUC1 1H
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 399.9100096 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



2.00
3.00
3.00
5.00
5.01
0.98
2.02
3.00

4i - ¹³C NMR, 101 MHz, CDCl₃

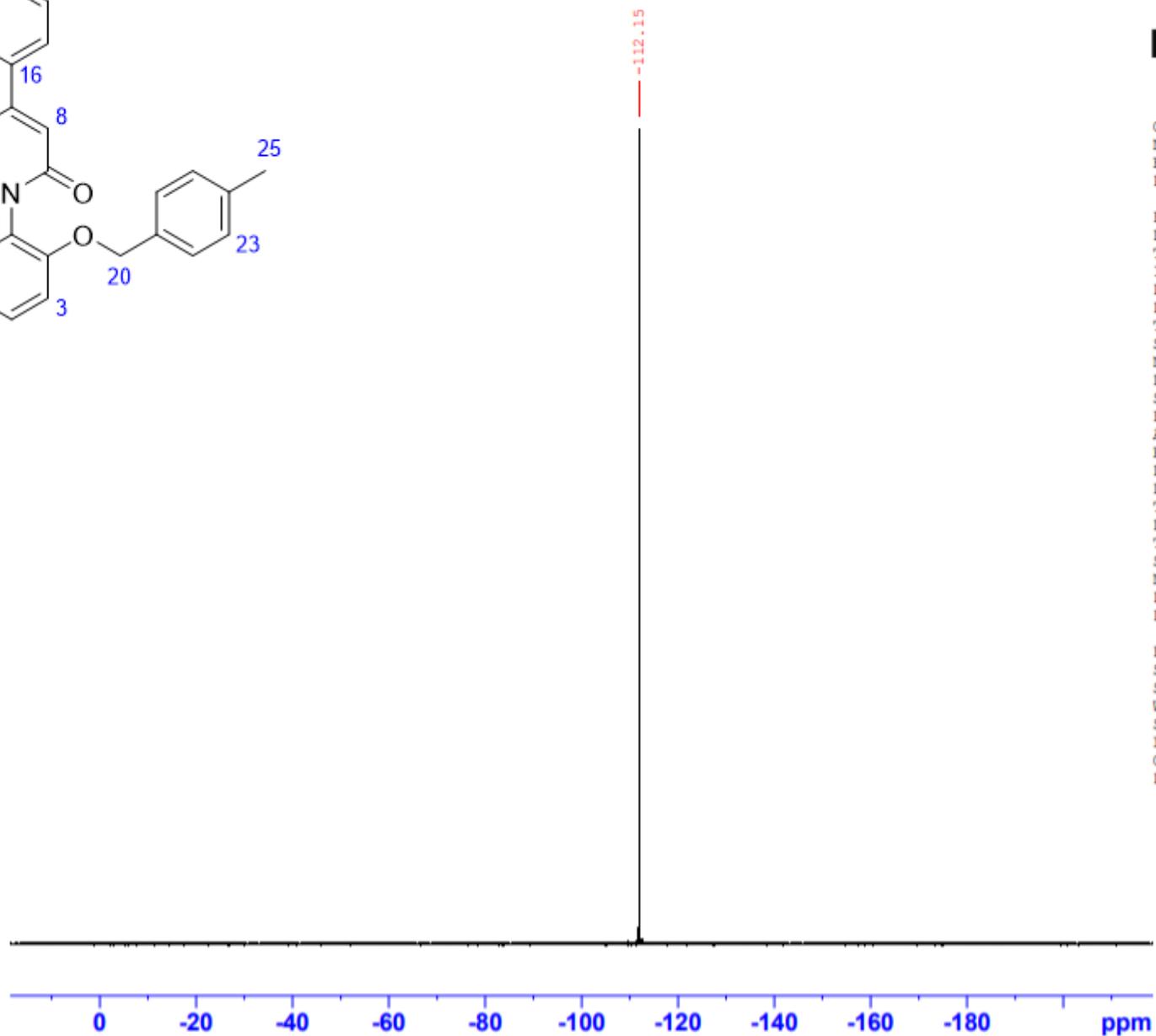
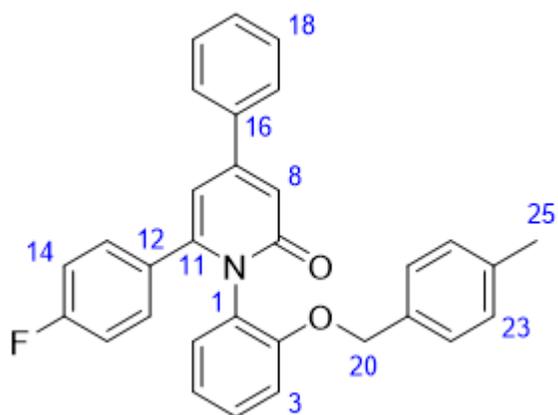


Current Data Parameters
NAME JS-4-297
EXPNO 47
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210420
Time_ 22.34 h
INSTRUM AVIII 400
PROBHD z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077400 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

4i - ¹⁹F NMR, 376 MHz, CDCl₃

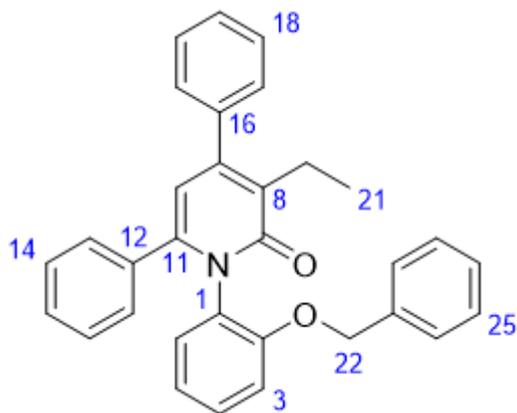


Current Data Parameters
NAME JS-4-297
EXPNO 14
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210420
Time 23.06 h
INSTRUM AVIII 400
PROBHD z108618_0146 (zg)
PULPROG zg
TD 261992
SOLVENT CDCl3
NS 16
DS 4
SWH 89285.711 Hz
FIDRES 0.681591 Hz
AQ 1.4671552 sec
RG 575
DW 5.600 usec
DE 7.11 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 376.4418995 MHz
NUC1 19F
P1 11.80 usec
PLW1 32.96500015 W

F2 - Processing parameters
SI 262144
SF 376.4795470 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

4j - ¹H NMR, 400 MHz, CDCl₃

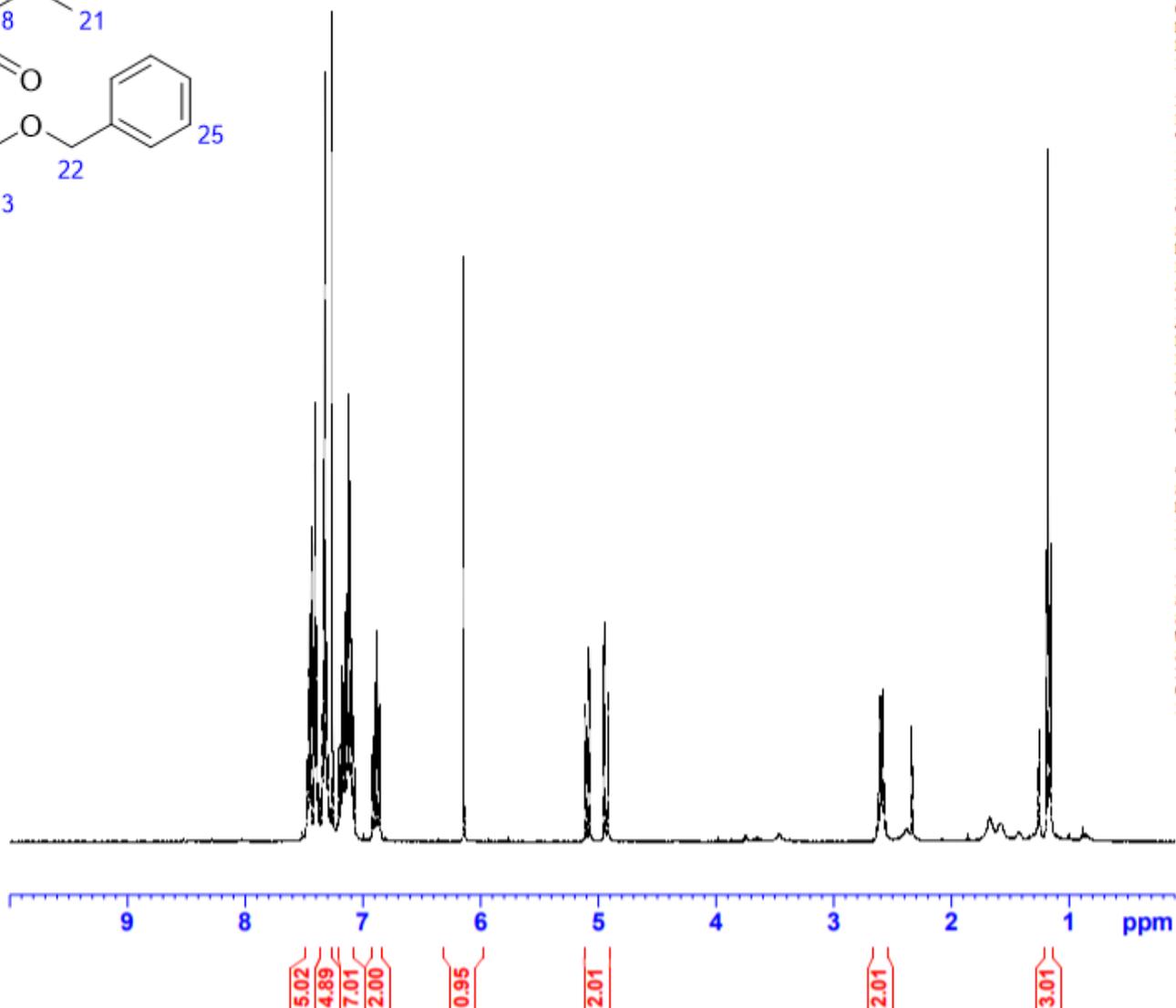


Current Data Parameters
NAME JS-4-291 recol
EXPNO 10
PROCNO 1

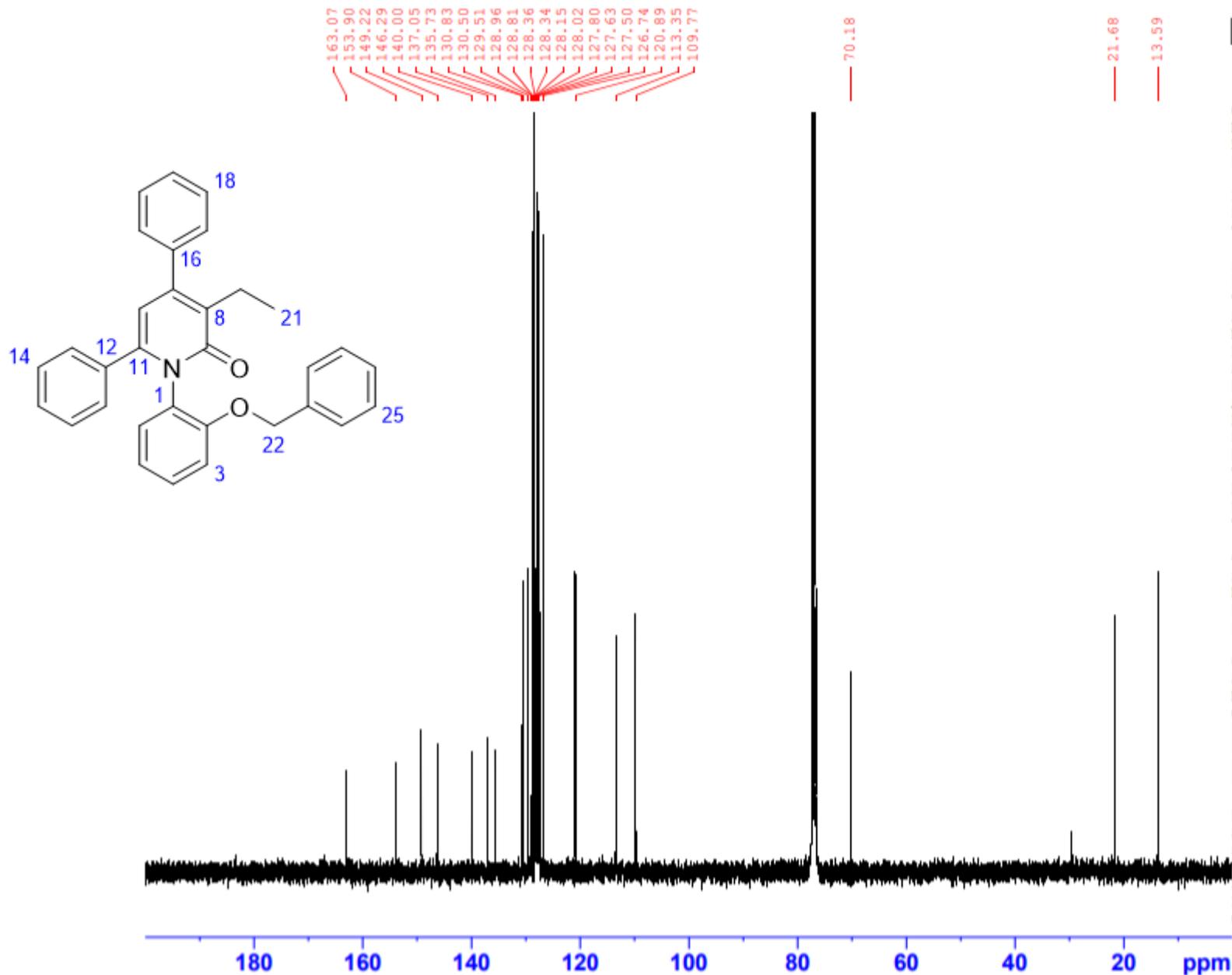
F2 - Acquisition Parameters
Date_ 20210409
Time_ 13.12
INSTRUM AVIII_400
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 114
DW 60.800 usec
DE 6.50 usec
TE 294.8 K
D1 1.00000000 sec
TD0 1

----- CHANNEL f1 -----
SFO1 399.9124696 MHz
NUC1 1H
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 399.9100097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4j - ¹³C NMR, 101 MHz, CDCl₃



Current Data Parameters
NAME JS-4-291 recol
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210411
Time 17.28
INSTRUM AVIII_400
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.250010 Hz
AQ 1.9999200 sec
RG 64
DW 20.800 usec
DE 6.50 usec
TE 300.6 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

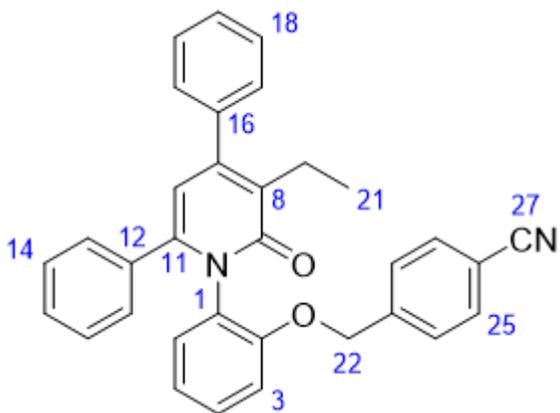
----- CHANNEL f1 -----
SFO1 100.5675047 MHz
NUC1 13C
P1 9.00 usec
PLW1 96.68000031 W

----- CHANNEL f2 -----
SFO2 399.9115996 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.38907000 W

F2 - Processing parameters
SI 131072
SF 100.5574500 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

S173

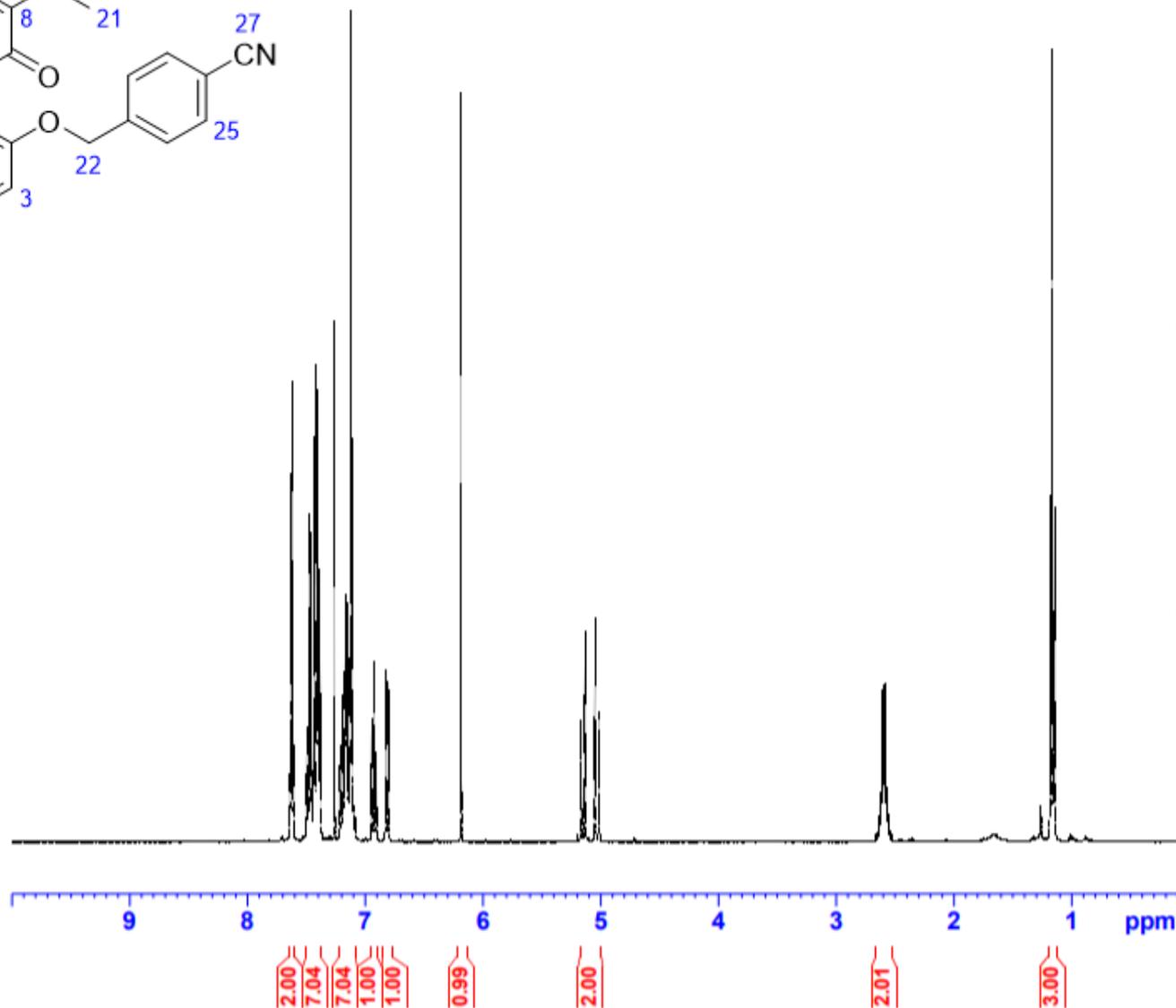
4k - ¹H NMR, 400 MHz, CDCl₃



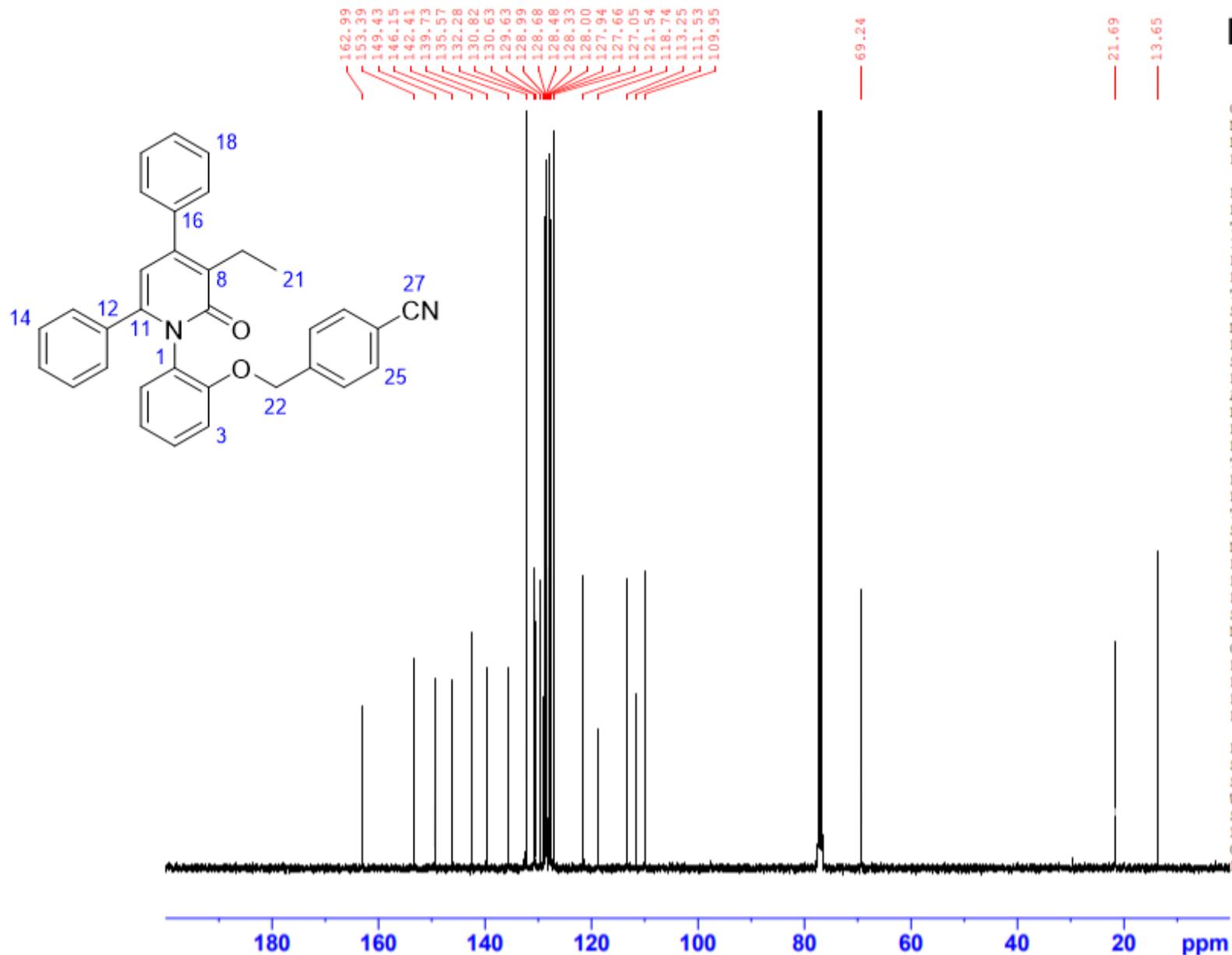
Current Data Parameters
NAME JS-4-306 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210422
Time 17.26 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 161
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4k - ¹³C NMR, 101 MHz, CDCl₃



Current Data Parameters
NAME JS-4-306 col
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210423
Time_ 22.34 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077414 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

4I - ¹H NMR, 400 MHz, CDCl₃

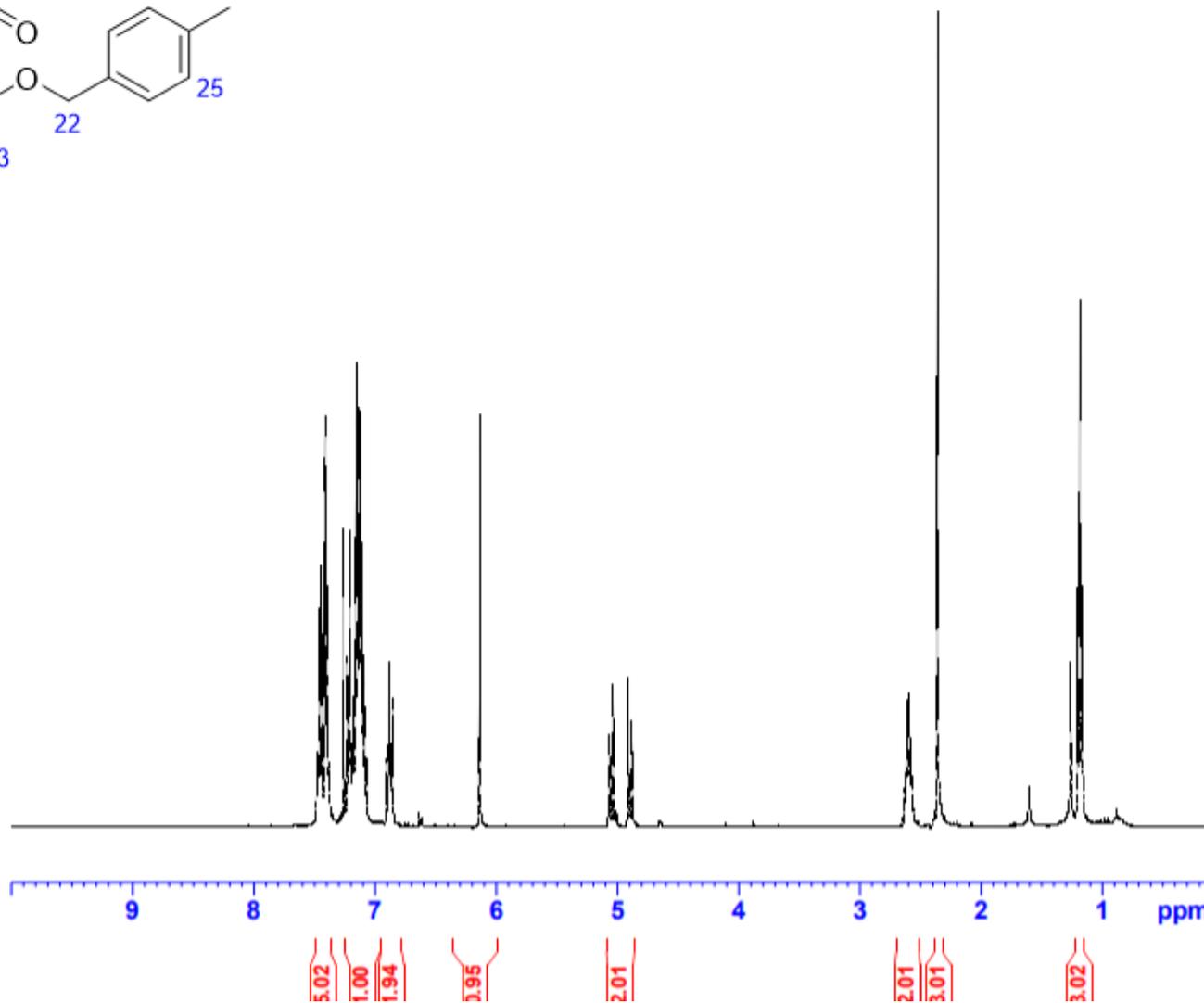
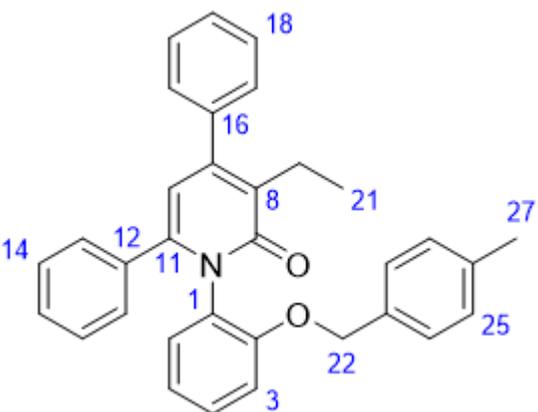


Current Data Parameters
NAME JS-4-307 col
EXPNO 10
PROCNO 1

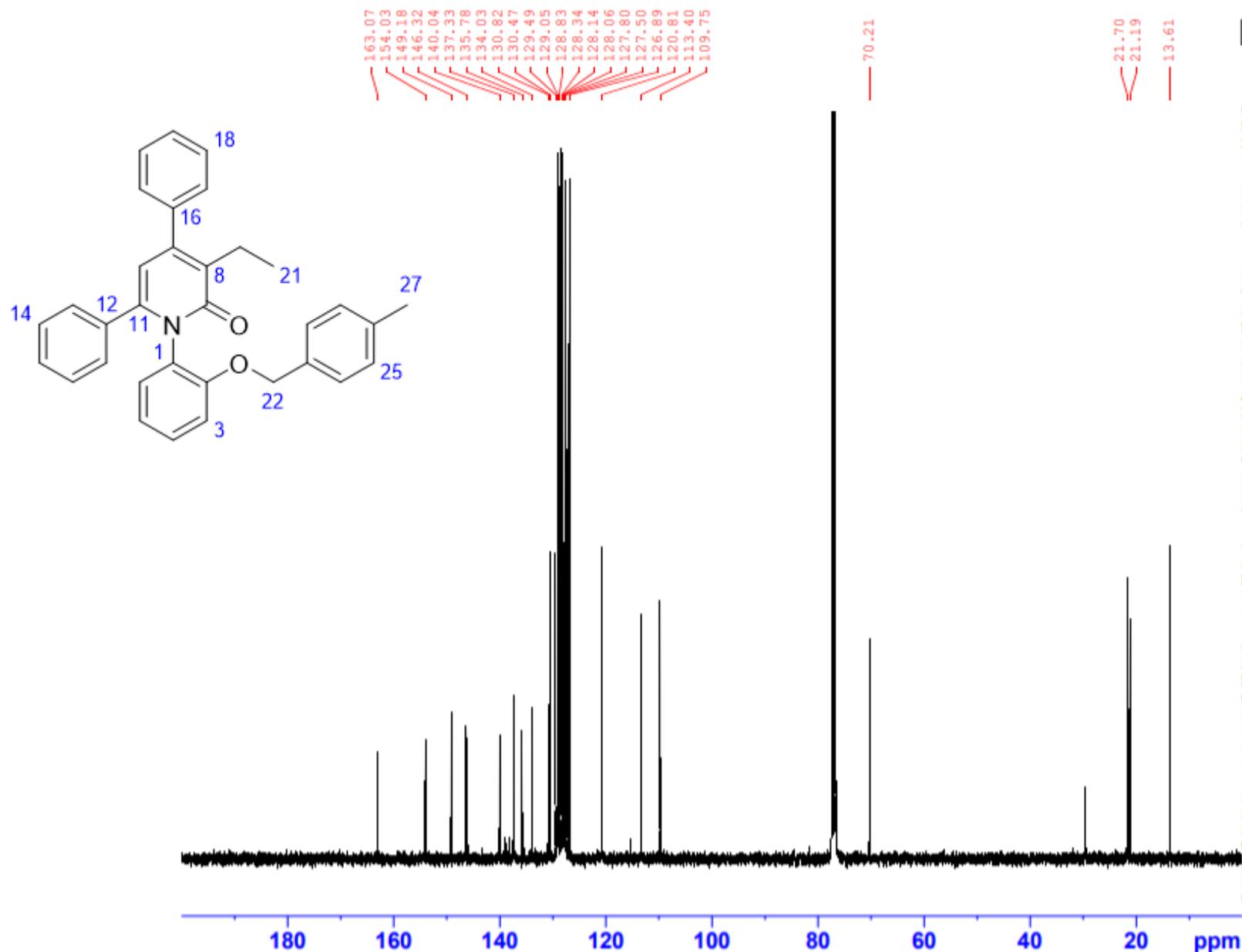
F2 - Acquisition Parameters
Date_ 20210422
Time_ 17.25
INSTRUM AVIII_400
PROBHD 5 mm PABBO_BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9845889 sec
RG 80.6
DW 60.800 usec
DE 6.50 usec
TE 296.0 K
D1 1.0000000 sec
TDO 1

----- CHANNEL f1 -----
SFO1 399.9124696 MHz
NUC1 1H
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 399.9100098 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4I - ¹³C NMR, 101 MHz, CDCl₃



Current Data Parameters
NAME JS-4-307 col
EXPNO 11
PROCNO 1

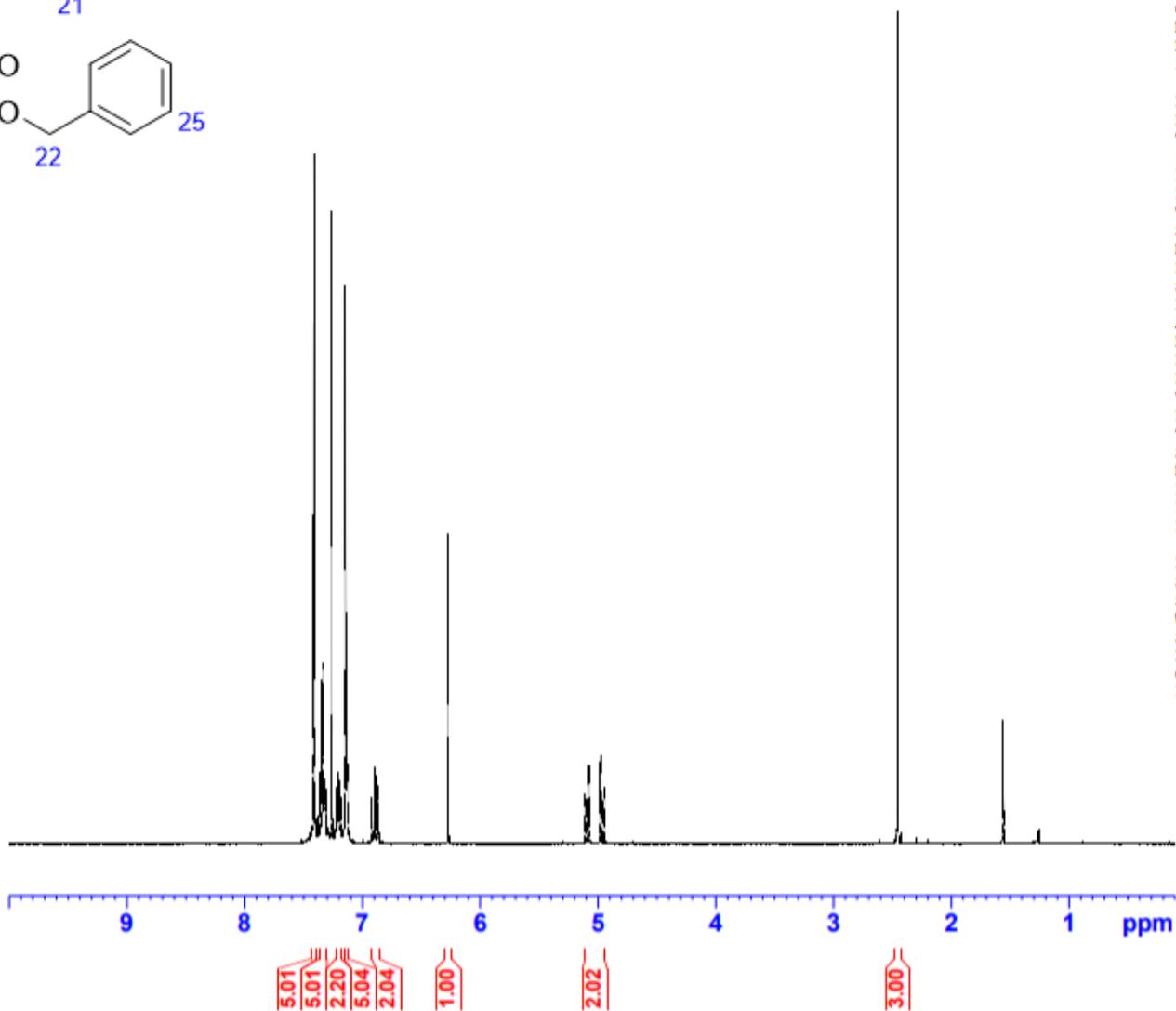
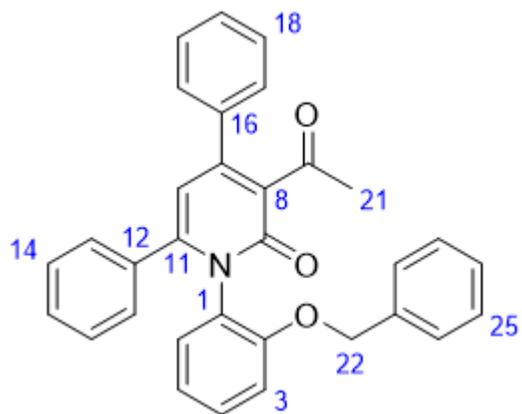
F2 - Acquisition Parameters
Date_ 20210422
Time_ 22.34
INSTRUM AVIII_400
PROBHD 5 mm PABBO_BB/
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.250010 Hz
AQ 1.9999200 sec
RG 80.6
DW 20.800 usec
DE 6.50 usec
TE 301.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

----- CHANNEL f1 -----
SFO1 100.5675047 MHz
NUC1 13C
P1 9.00 usec
PLW1 96.68000031 W

----- CHANNEL f2 -----
SFO2 399.9115996 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.38907000 W

F2 - Processing parameters
SI 131072
SF 100.5574487 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

4m - ¹H NMR, 400 MHz, CDCl₃

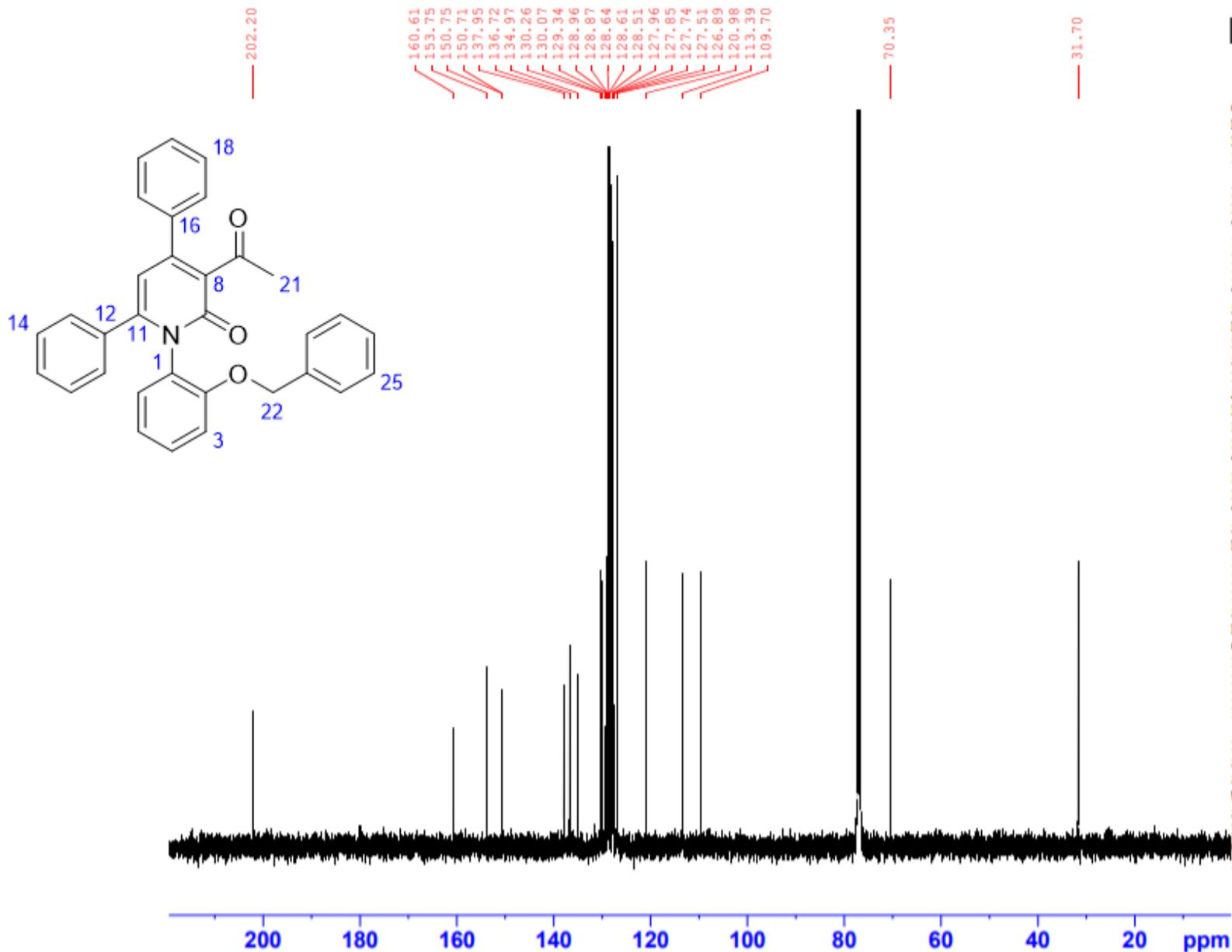


Current Data Parameters
NAME JS-4-290 recol
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210507
Time_ 17.23 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (zg30)
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TDO 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100095 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

4m - ¹³C NMR, 101 MHz, CDCl₃

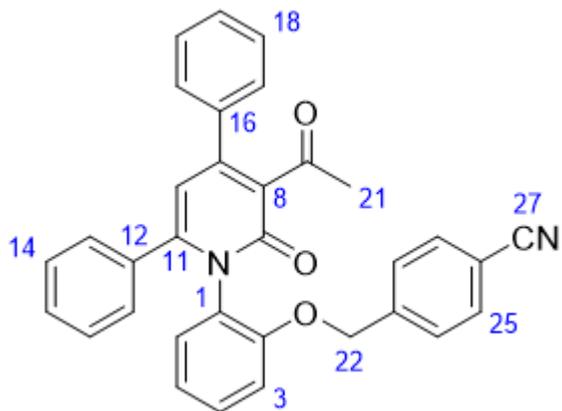


Current Data Parameters
NAME JS-4-290 recol
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date 20210508
Time 15.15 h
INSTRUM AVIII 400
PROBHD z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077407 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

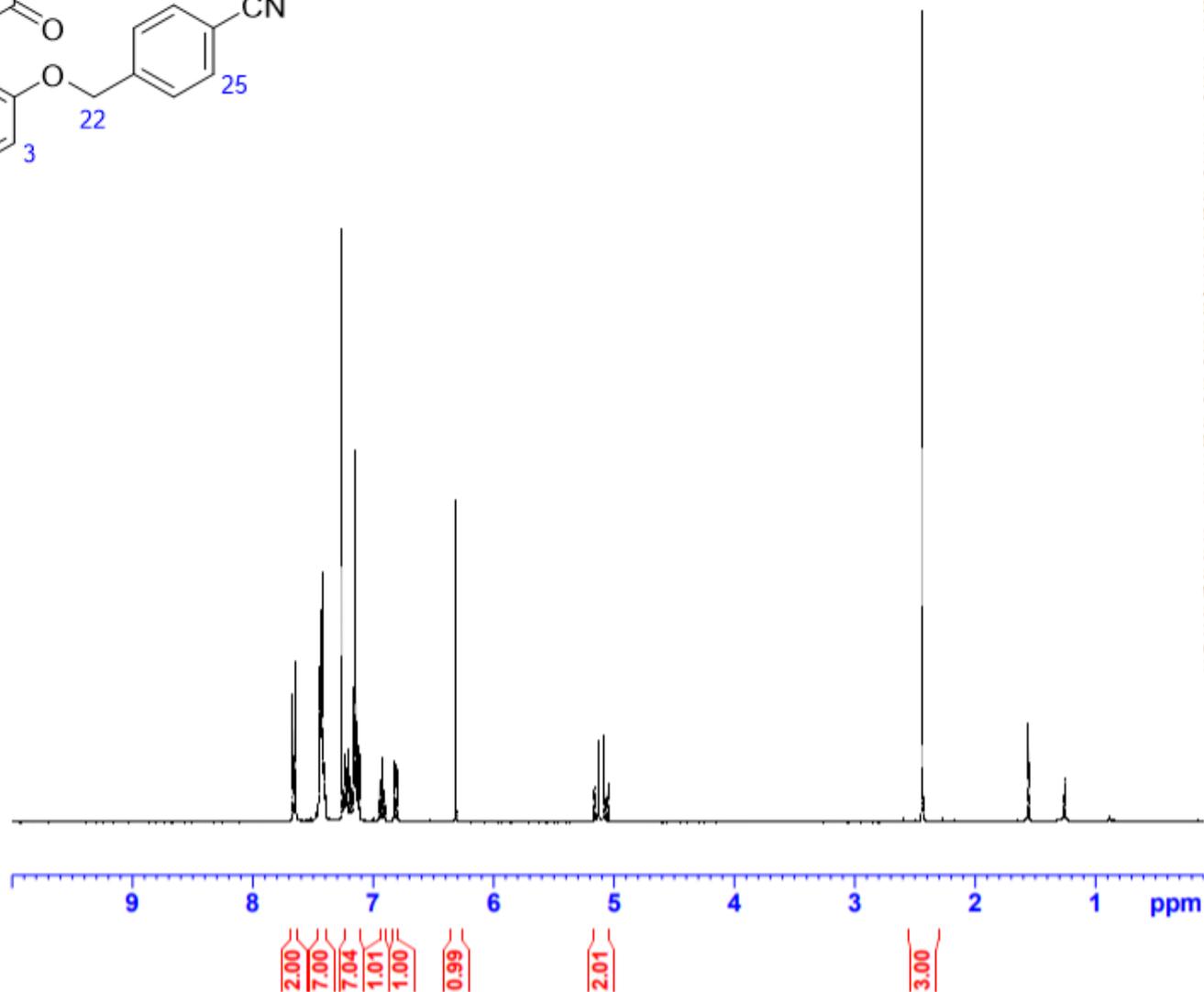
4n - ¹H NMR, 400 MHz, CDCl₃



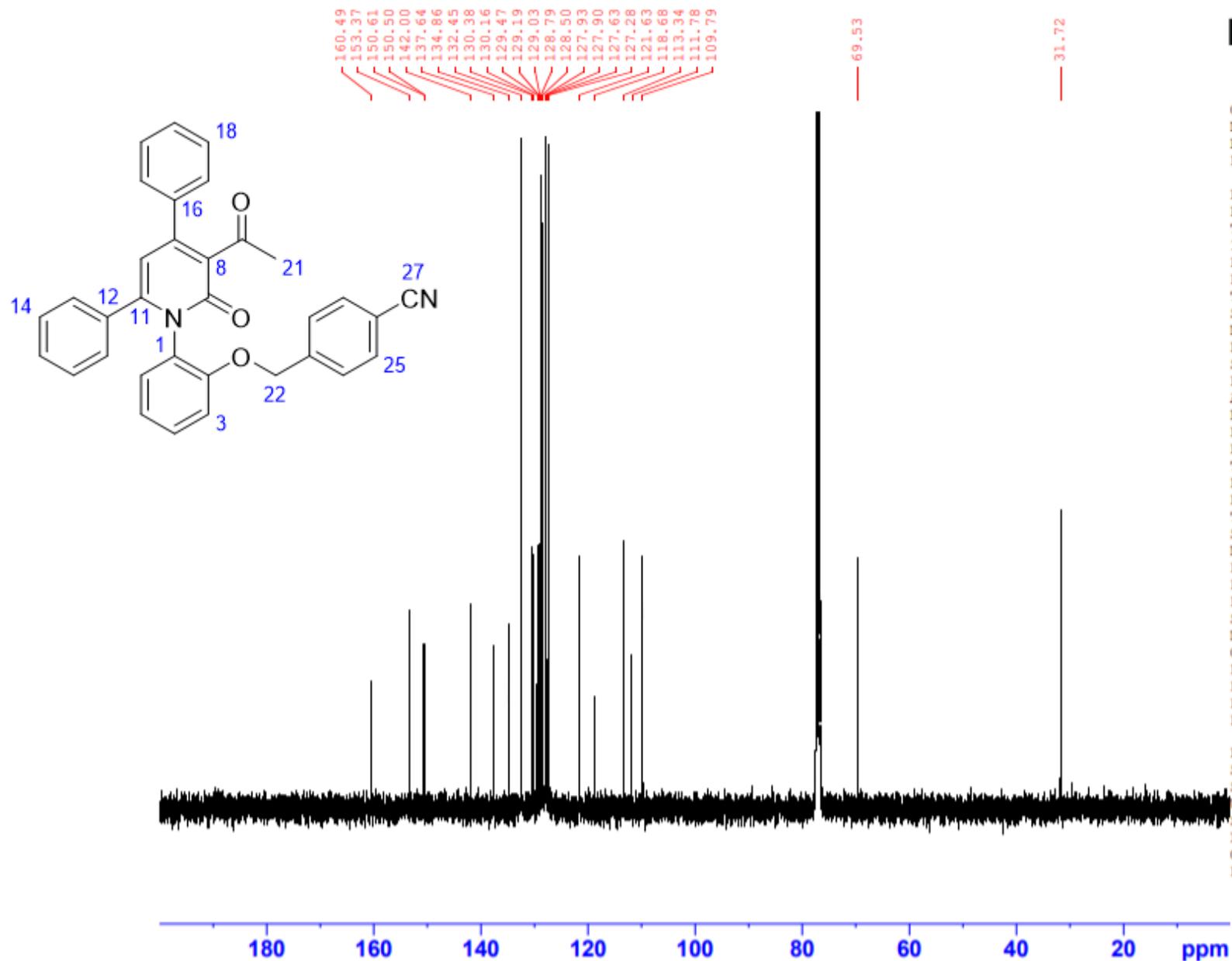
Current Data Parameters
NAME JS-4-304 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210421
Time 16.56 h
INSTRUM AVIII 400
PROBHD z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
PO 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4n - ¹³C NMR, 101 MHz, CDCl₃

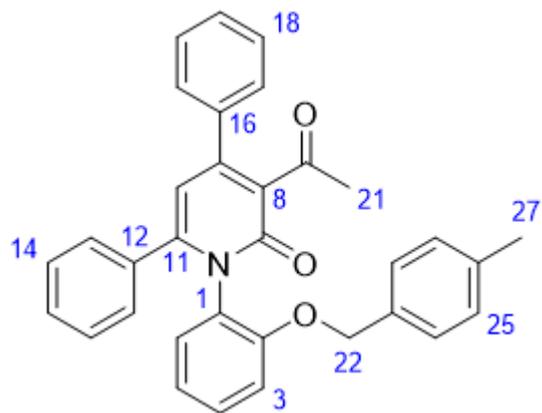


Current Data Parameters
NAME JS-4-304 col
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210421
Time_ 22.35 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077403 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

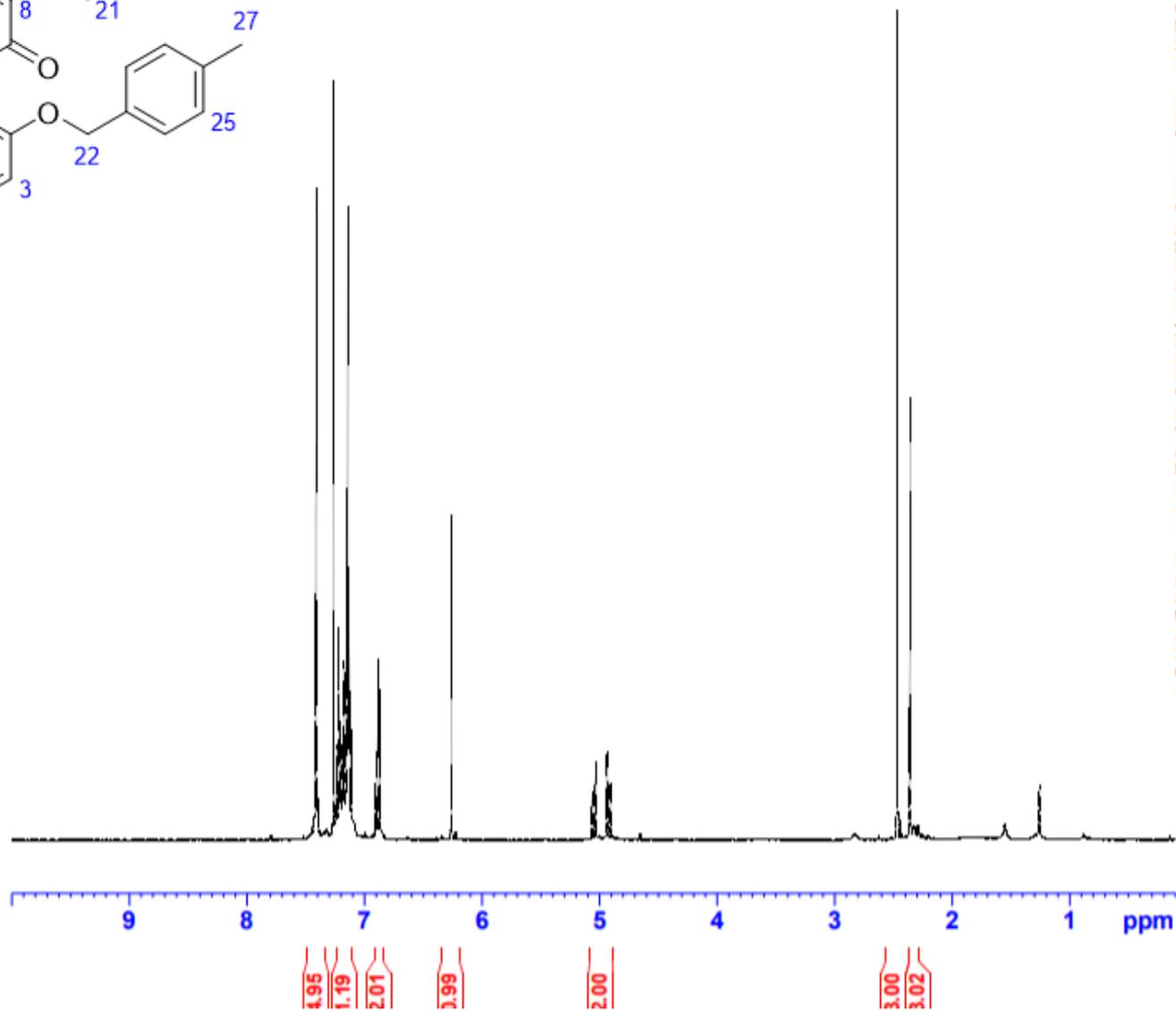
4o - ¹H NMR, 400 MHz, CDCl₃



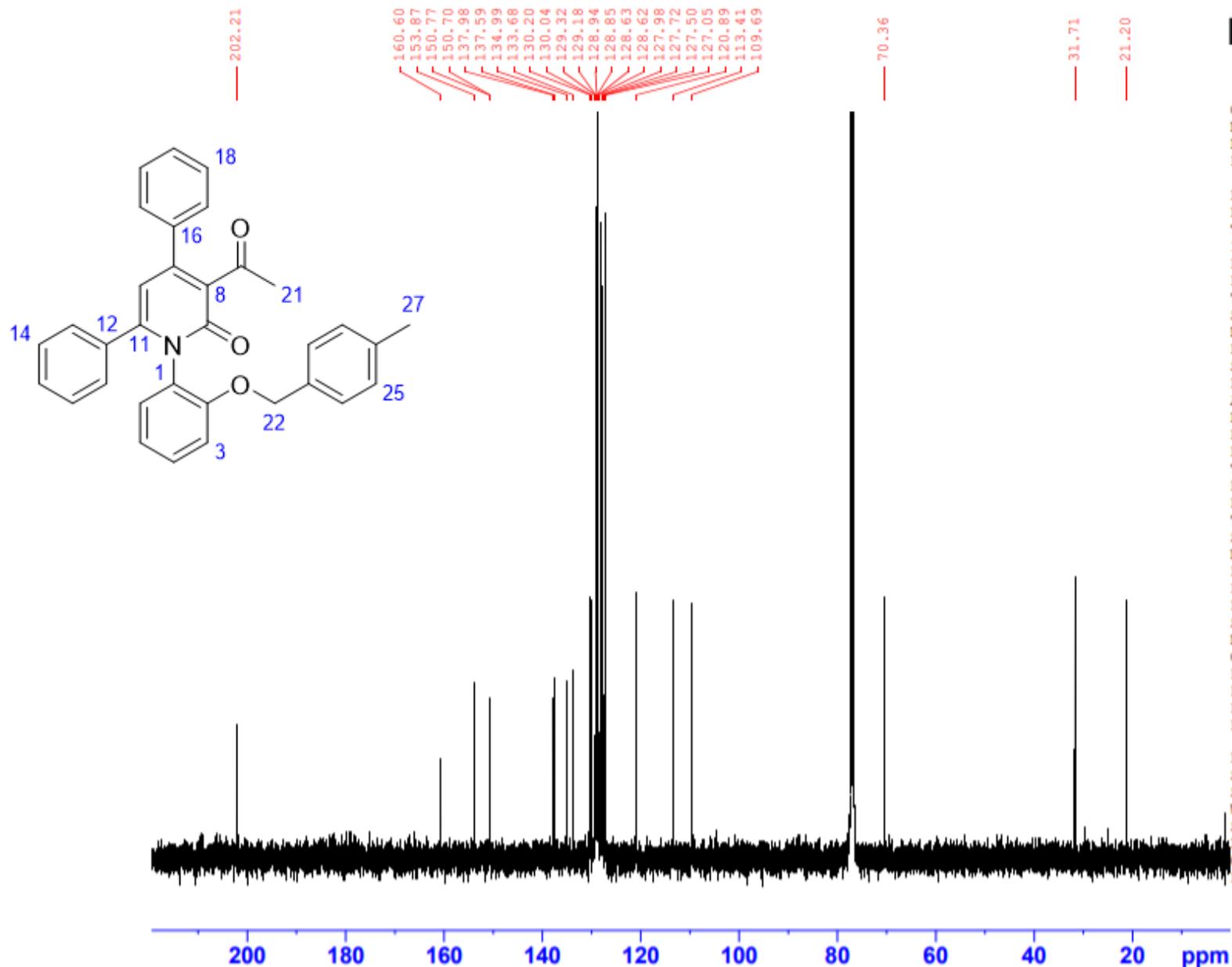
Current Data Parameters
NAME JS-4-305 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210422
Time_ 11.38 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TDO 1
SFO1 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4o - ¹³C NMR, 101 MHz, CDCl₃



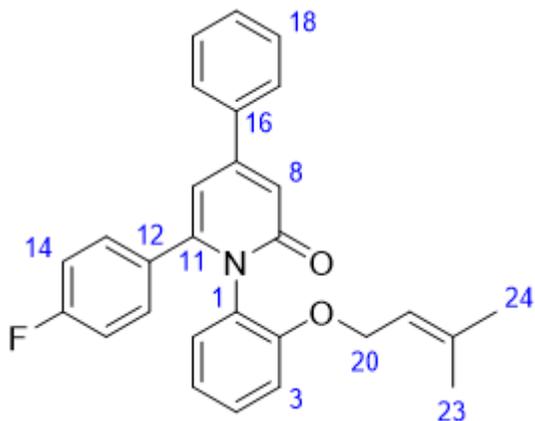
Current Data Parameters
NAME JS-4-305 col
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210422
Time_ 23.16 h
INSTRUM AVIII 400
PROBHD Z108618_0146 ()
PULPROG zgpg30
TD 96150
SOLVENT CDCl3
NS 4096
DS 4
SWH 24038.461 Hz
FIDRES 0.500020 Hz
AQ 1.9999200 sec
RG 2050
DW 20.800 usec
DE 6.50 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1
SFO1 100.6178003 MHz
NUC1 13C
P0 3.00 usec
P1 9.00 usec
PLW1 96.68000031 W
SFO2 400.1116004 MHz
NUC2 1H
CPDPRG[2] waltz64
PCPD2 90.00 usec
PLW2 17.29199982 W
PLW12 0.48032999 W
PLW13 0.24160001 W

F2 - Processing parameters
SI 131072
SF 100.6077404 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

S183

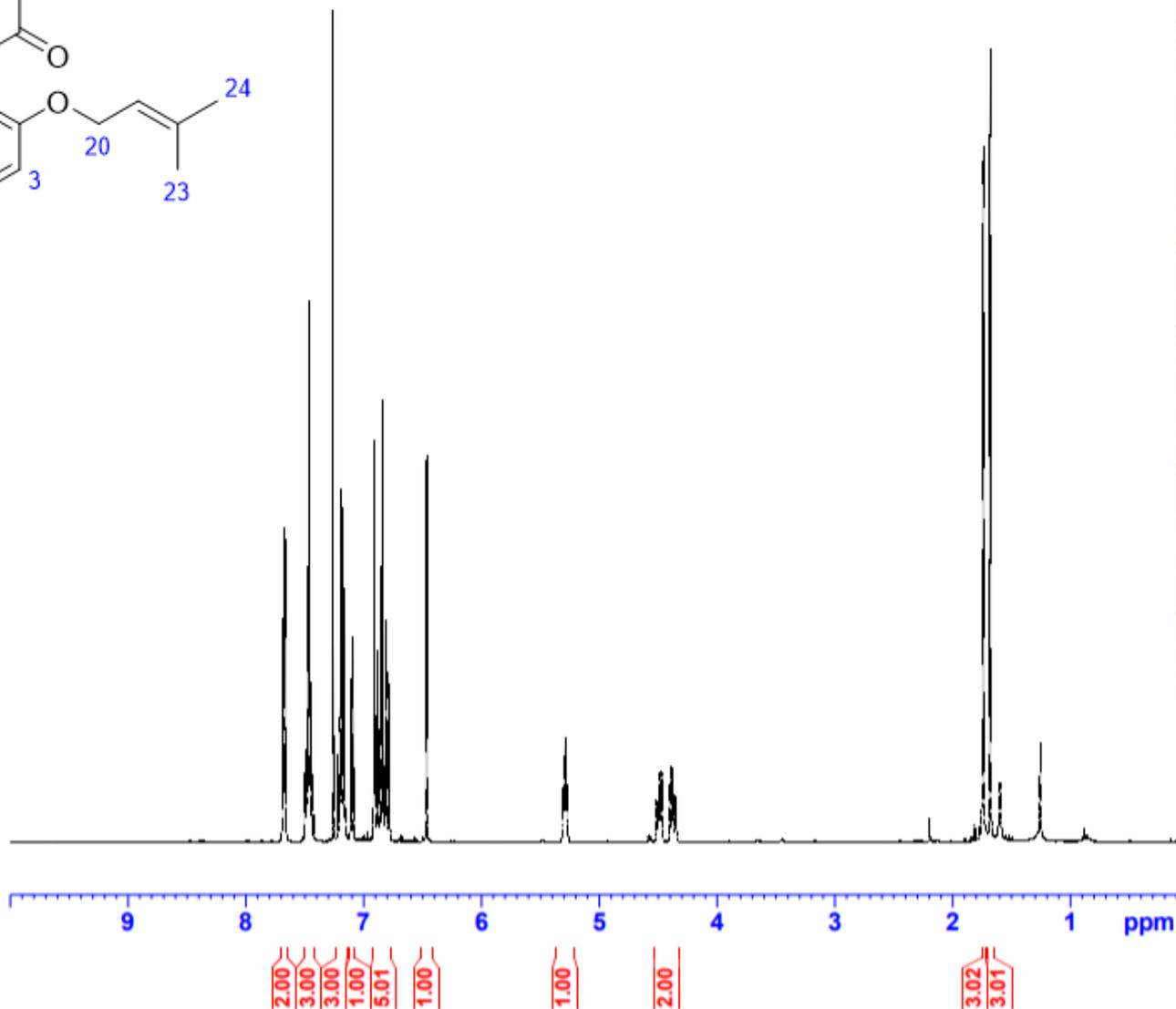
4p - ¹H NMR, 400 MHz, CDCl₃



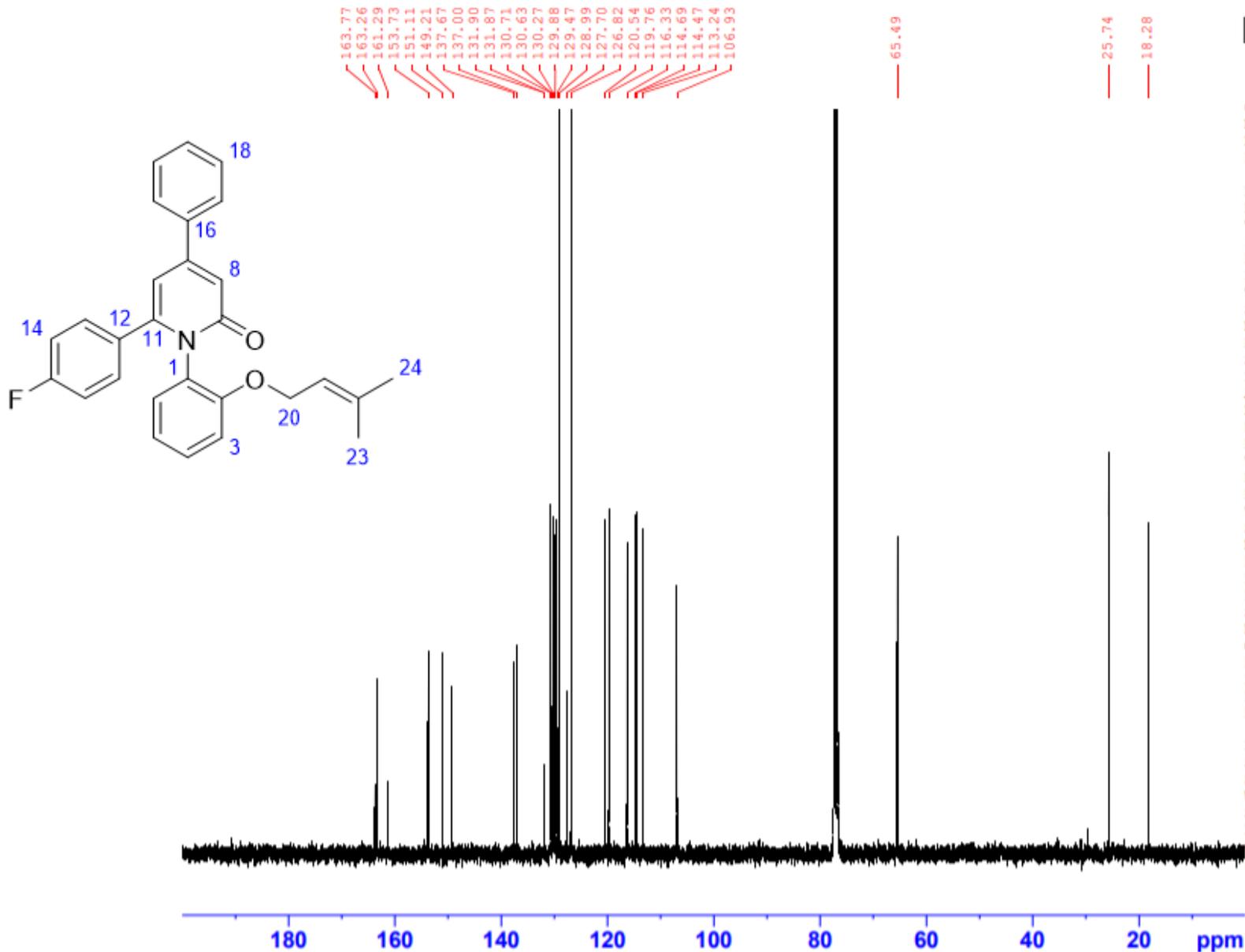
Current Data Parameters
NAME JS-4-310 col
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210430
Time 14.54 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.250967 Hz
AQ 3.9845889 sec
RG 256
DW 60.800 usec
DE 17.42 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SF01 400.1124708 MHz
NUC1 1H
P0 5.00 usec
P1 15.00 usec
PLW1 17.29199982 W

F2 - Processing parameters
SI 32768
SF 400.1100102 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



4p - ¹³C NMR, 101 MHz, CDCl₃

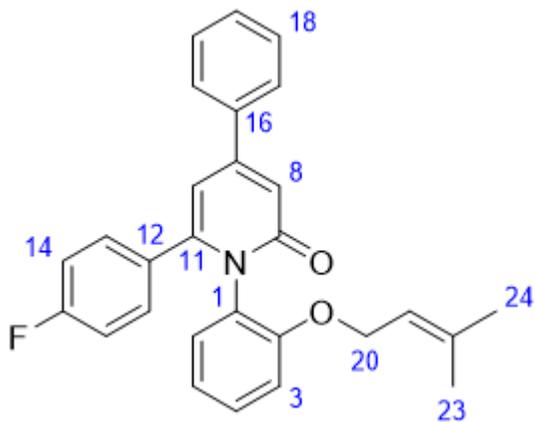


Current Data Parameters
 NAME JS-4-310 col
 EXPNO 11
 PROCNO 1

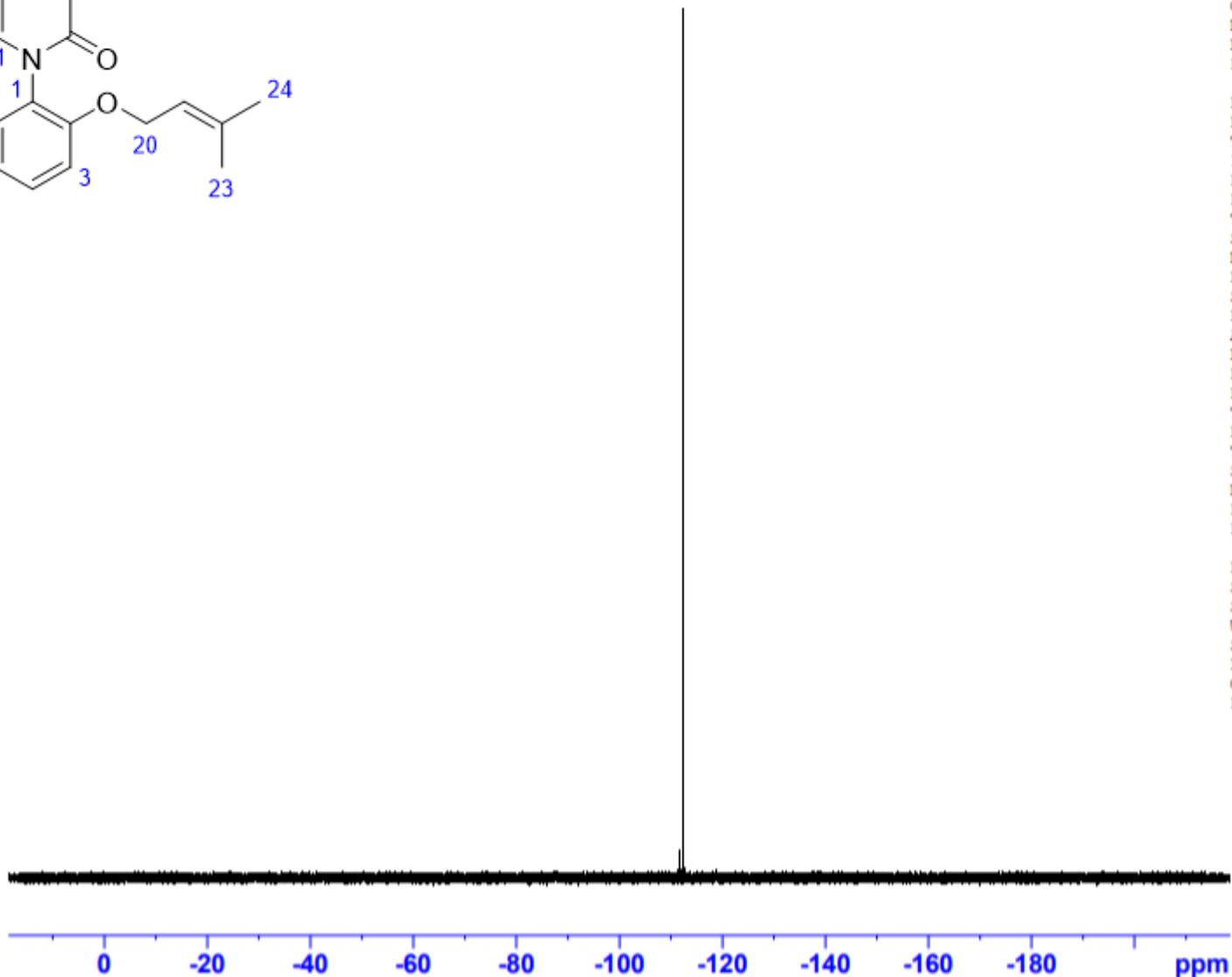
F2 - Acquisition Parameters
 Date_ 20210430
 Time 22.48 h
 INSTRUM AVIII 400
 PROBHD Z108618_0146 ()
 PULPROG zgpg30
 TD 96150
 SOLVENT CDCl3
 NS 4096
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.500020 Hz
 AQ 1.9999200 sec
 RG 2050
 DW 20.800 usec
 DE 6.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1
 SFO1 100.6178003 MHz
 NUC1 13C
 P0 3.00 usec
 P1 9.00 usec
 PLW1 96.68000031 W
 SFO2 400.1116004 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 17.29199982 W
 PLW12 0.48032999 W
 PLW13 0.24160001 W

F2 - Processing parameters
 SI 131072
 SF 100.6077401 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

4p - ¹⁹F NMR, 376 MHz, CDCl₃



-112.32



S186

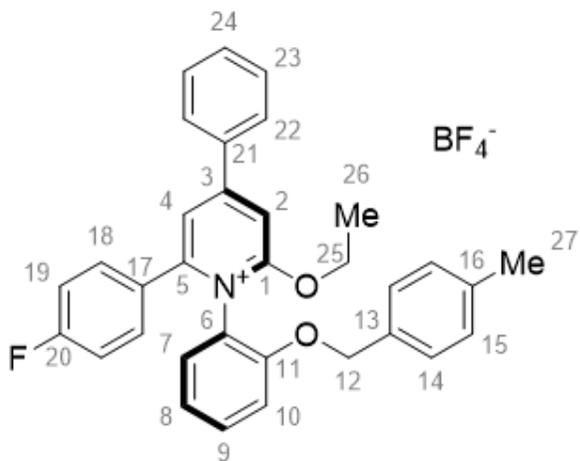


Current Data Parameters
NAME JS-4-310 col
EXPNO 19
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210505
Time_ 14.01 h
INSTRUM AVIII 400
PROBHD Z108618_0146 (
PULPROG zg
TD 261992
SOLVENT CDCl3
NS 16
DS 4
SWH 89285.711 Hz
FIDRES 0.681591 Hz
AQ 1.4671552 sec
RG 575
DW 5.600 usec
DE 7.11 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1
SFO1 376.4418995 MHz
NUC1 19F
P1 11.80 usec
PLW1 32.96500015 W

F2 - Processing parameters
SI 262144
SF 376.4795470 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

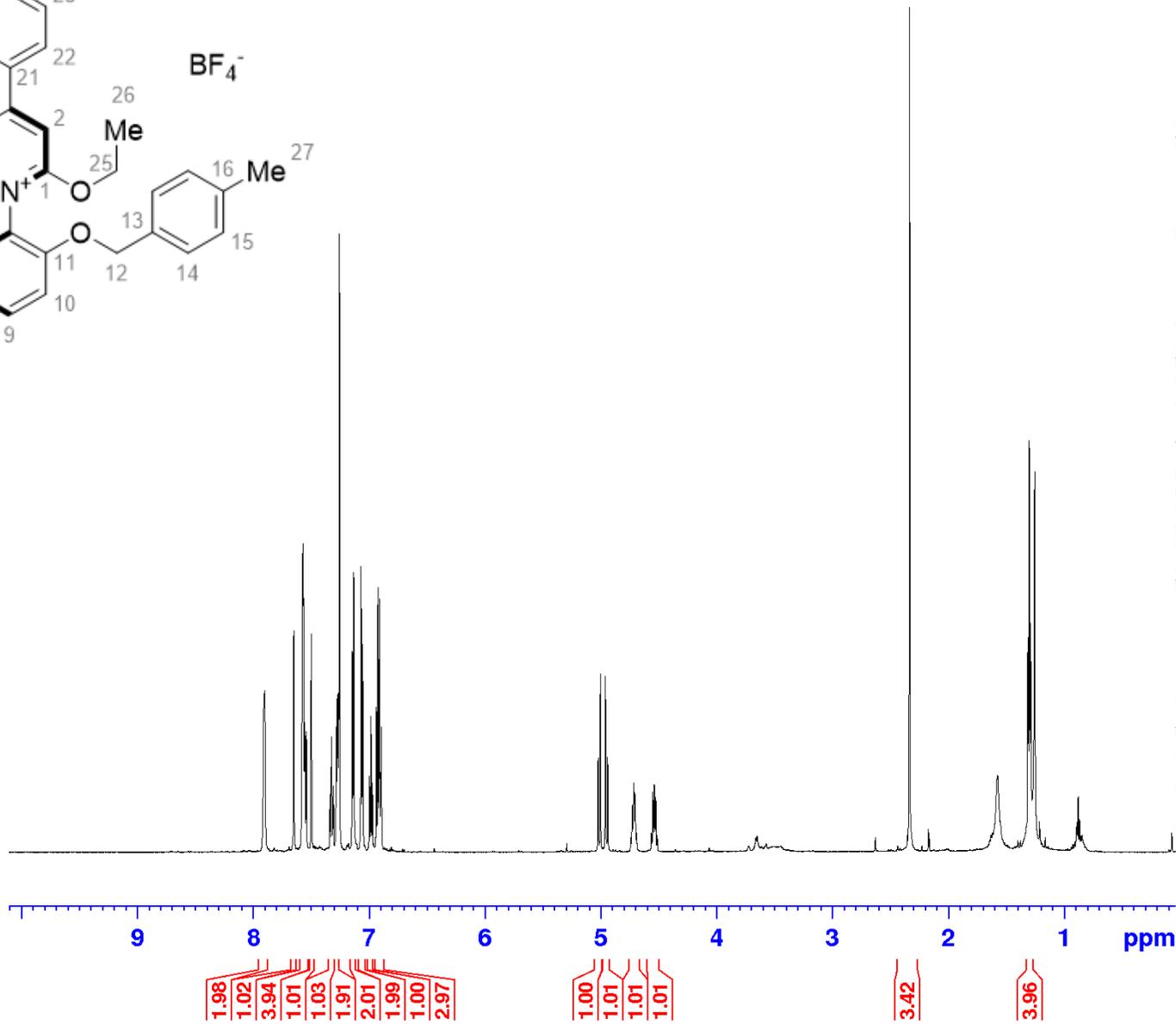
7 - ¹H NMR, 600 MHz, CDCl₃



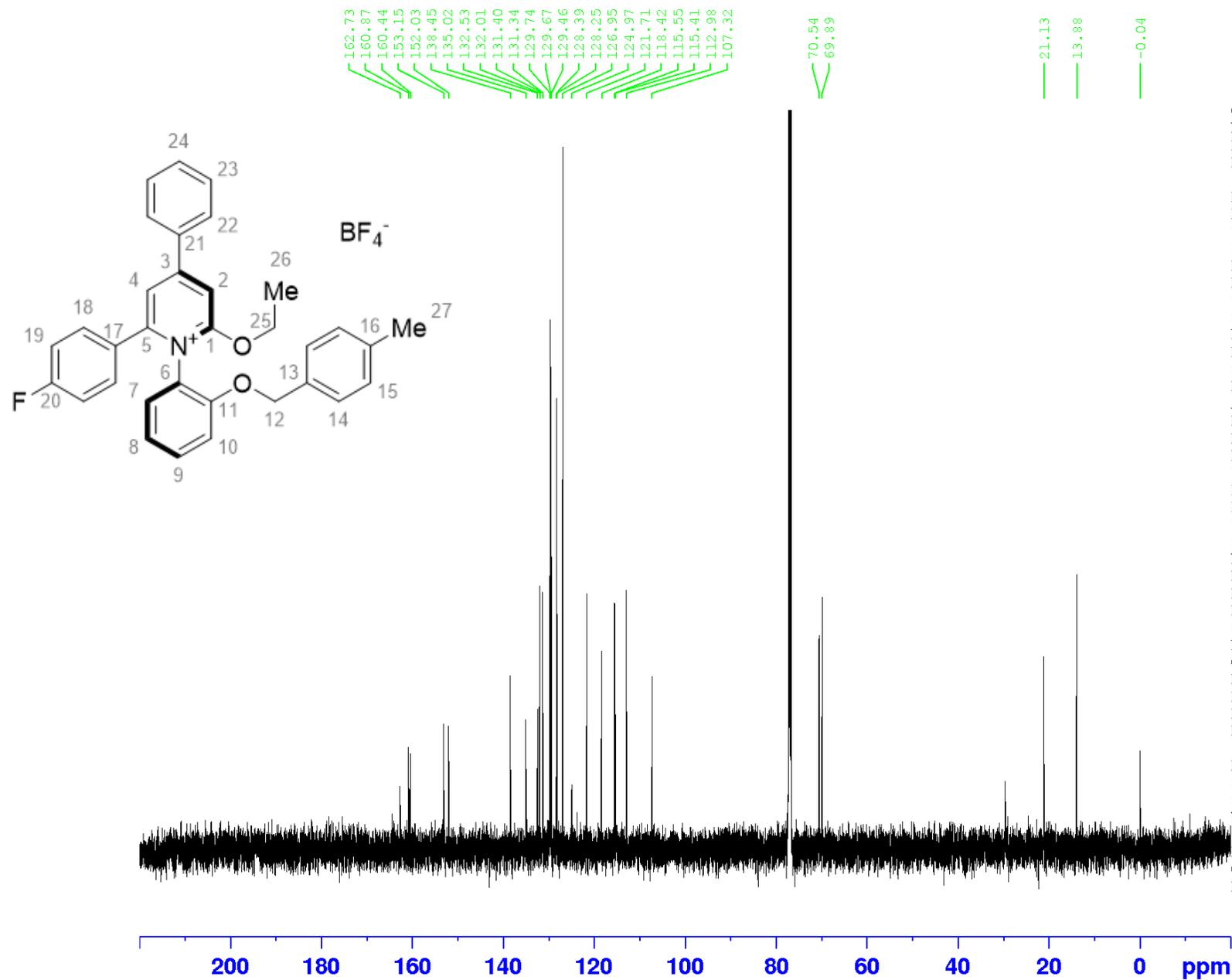
Current Data Parameters
 NAME PCK 3.79 re-col
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210624
 Time 18.25 h
 INSTRUM spect
 PROBHD Z114607_0188 (
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.366798 Hz
 AQ 2.7262976 sec
 RG 105.21
 DW 41.600 usec
 DE 12.10 usec
 TE 307.4 K
 D1 1.00000000 sec
 TD0 1
 SFO1 600.1337058 MHz
 NUC1 1H
 P0 3.33 usec
 P1 10.00 usec
 PLW1 26.60000038 W

F2 - Processing parameters
 SI 65536
 SF 600.1300153 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



7 - ¹³C NMR, 151 MHz, CDCl₃

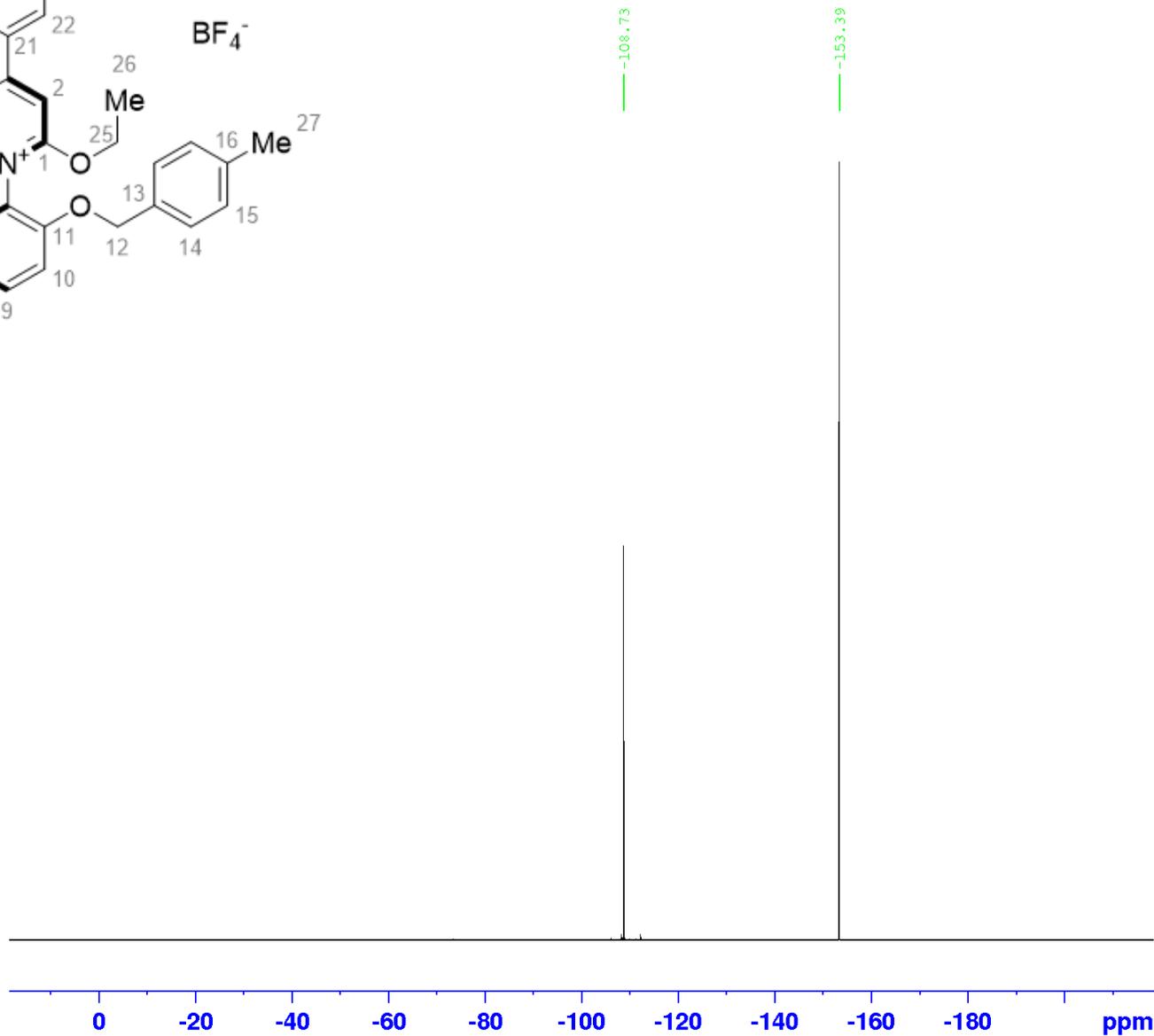
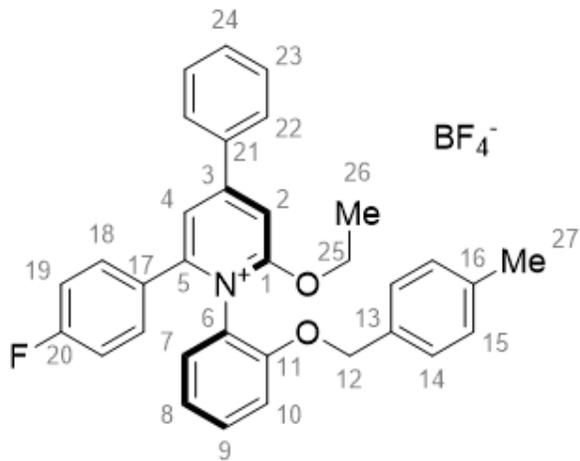


Current Data Parameters
 NAME PCK 3.79 re-col
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210624
 Time 19.17 h
 INSTRUM spect
 PROBHD Z114607_0188 ()
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1024
 DS 4
 SWH 36231.883 Hz
 FIDRES 1.105709 Hz
 AQ 0.9043968 sec
 RG 186.92
 DW 13.800 usec
 DE 6.50 usec
 TE 305.5 K
 D1 2.0000000 sec
 D11 0.03000000 sec
 TD0 1
 SFO1 150.9178988 MHz
 NUC1 13C
 P0 3.93 usec
 P1 11.80 usec
 PLW1 85.00000000 W
 SFO2 600.1324005 MHz
 NUC2 1H
 CPDPRG[2] waltz65
 PCPD2 70.00 usec
 PLW2 27.00000000 W
 PLW12 0.57327998 W
 PLW13 0.28836000 W

F2 - Processing parameters
 SI 32768
 SF 150.9028085 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

7 - ¹⁹F NMR, 564 MHz, CDCl₃



Current Data Parameters
 NAME PCK 3.79
 EXPNO 18
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210612
 Time 4.14 h
 INSTRUM spect
 PROBHD Z114607_0188 (
 PULPROG zgfhigqn.b
 TD 261896
 SOLVENT CDCl₃
 NS 16
 DS 4
 SWH 133928.578 Hz
 FIDRES 1.022761 Hz
 AQ 0.9777451 sec
 RG 186.92
 DW 3.733 usec
 DE 6.70 usec
 TE 302.0 K
 D1 4.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1
 SFO1 564.6299217 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 49.00000000 W
 SFO2 600.1324005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 27.00000000 W
 PLW12 0.57327998 W

F2 - Processing parameters
 SI 262144
 SF 564.6863882 MHz
 WDW EM
 SSB 0
 LB 0.50 Hz
 GB 0
 PC 2.00

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