## **Electronic Supplementary Information**

# for

# **Bioinspired Total Synthesis of Boneratamides A-C**

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#### **General Information**

Melting points (Mp) were recorded on a Yanaco MP-S3 melting point apparatus and are not corrected. Optical rotations were measured on a JASCO P-2200 digital polarimeter at the sodium D line with a 100 mm path length cell, and are reported as follows:  $[\alpha]_D^T$ , concentration (g/100 mL), and solvent. Infrared spectra (IR), recorded on a JASCO FT/IR-460 spectrophotometer and Thermo Fisher Scientific Nicolet 6700 FT-IR spectrometer, are reported in wave number (cm<sup>-1</sup>). Proton nuclear magnetic resonance (<sup>1</sup>H NMR) spectra were recorded on JEOL LA 500 (500 MHz), JEOL ECS-400 (400 MHz) and JEOL ECZ 500 (500 MHz) spectrometers. <sup>1</sup>H NMR chemical shifts ( $\delta_{\rm H}$ ) are reported in parts per million (ppm) with the solvent resonance as the internal standard relative to chloroform ( $\delta = 7.26$ ), CHD<sub>2</sub>OD ( $\delta = 3.31$ ), benzene ( $\delta = 7.15$ ) and DMSO ( $\delta = 2.50$ ). Data are reported as follows: chemical shift ( $\delta$ ), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, br = broadened, m = multiplet), coupling constants (J, given in Hz) and number of protons. <sup>13</sup>C NMR chemical shifts ( $\delta$ ) are recorded in parts per million (ppm) relative to the residual solvent peaks of CDCl<sub>3</sub> ( $\delta$  = 77.16), CD<sub>3</sub>OD ( $\delta$  = 49.9), C<sub>6</sub>D<sub>6</sub> ( $\delta$  = 128.06) and DMSO-d<sub>6</sub> ( $\delta$  = 39.5) as internal standards. High-resolution mass spectra (HRMS), measured on a BRUKER, FT-ICRMS, solariX XR, Thermo Fisher Scientific Exactive<sup>™</sup> Plus, JEOL JMS-GCMATEII and JEOL JMS-T100CS AccuTOF spectrometer, are reported in m/z. Reactions were monitored by thin-layer chromatography on glass plates 0.25 mm coated with silica gel 60 F<sub>254</sub> (MERCK 1.05715). Open-column chromatography was carried out with Silica gel 60 (particle size 0.063-0.200 mm, 70-230 mesh ASTM). Reactions were run under atmosphere of argon when the reactions were sensitive to moisture or oxygen. THF was distilled from LiAlH<sub>4</sub> before use. Dichloromethane was dried over molecular sieves 3 Å. Pyridine and triethylamine were stocked over anhydrous KOH. All other commercially available reagents were used as CH<sub>2</sub>Cl<sub>2</sub>

#### **Experimental Section**



(-)-(2S,5S)-5-Isopropyl-2-methylcyclohexanone [(-)-carvomenthone] (10). A solution of (S)-(+)-carvone (9) (60.0 g, 26.6 mmol) dissolved in methanol (240 mL) in the presence of platinum oxide (120 mg, 0.13 mol%) was vigorously stirred at room temperature under hydrogen atmosphere for 6.5 h. The solution was filtered on the pad of Celite and concentrated under reduced pressure to afford a mixture of (-)-carvomenthone (10) (57.4 g). This mixture was treated with a solution of sodium methoxide (5.0 M solution in methanol, 3.4 mL). After standing at room temperature for 3.5 h, the reaction mixture was diluted with water. The aqueous layer was extracted with ether (×2). The combined organic layers were washed with water and brine, and dried (Na<sub>2</sub>SO<sub>4</sub>). Concentration under reduced pressure gave the residue (55.75 g), which was subjected to Kugelrohr distillation (oven temperature 85 °C/0.1 mmHg) to afford (-)-carvomenthone (10) (53.5 g, trans:cis = 8:1, 87%). A portion of this product (25.0 g) was separated by silica-gel column chromatography (Et<sub>2</sub>O/hexane 1:40 $\rightarrow$ 1:20) to afford the trans-isomer 10 as a colorless liquid (15.0 g). The <sup>1</sup>H NMR data obtained for 10 were in agreement with values previously reported for *ent*-(+)-10.<sup>1</sup>  $[\alpha]_D^{22} = -11.9$  (c = 3.17, EtOH) {Literature:  $[\alpha]_D^{22} = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (\text{ddd}, J = -15.5 \ (c = 3, \text{ EtOH})^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (d dd)^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (d dd)^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (d dd)^1$ }; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 2.40 \ (d$ 13.0, 3.5, 2.5 Hz, 1H), 2.33 (dqd, J = 13.0, 6.5, 1.0 Hz, 1H), 2.13–2.01 (m, 2H), 1.85 (dddd, J = 12.5, 5.5, 2.5, 2.5 Hz, 1H), 1.61–1.51 (m, 2H), 1.44 (dddd, J = 13.0, 13.0, 11.5, 3.5 Hz, 1H), 1.30 (qd, J = 13.0, 3.5 Hz, 1H), 1.01 (d, J = 6.5 Hz, 3H), 0.90 (d, J = 6.5 Hz, 3H), 0.89 (d, J = 13.0, 3.5 Hz, 1H), 1.01 (d, J = 6.5 Hz, 3H), 0.89 (d, J = 13.0, 3.5 Hz, 1H), 1.01 (d, J = 13.0, 3.5 6.5 Hz, 3H).



(*E*)-Ethyl 2-((2*S*,5*S*)-5-isopropyl-2-methylcyclohexylidene)acetate (11). A solution of (–)-carvomenthone (10) (9.90 g, 64.2 mmol) in THF (15 mL) was added to a solution of sodium trimethylphosphonoacetate {prepared from trimethylphosphonoacetate (43.2 g, 193 mmol) and sodium hydride (5.13 g; 60% in oil, washed with hexane, 128 mmol) in THF 80 mL)}. After heating at reflux for 3 h, The reaction mixture was cooled to room temperature and diluted with saturated aqueous NH<sub>4</sub>Cl. The separated aqueous layer was extracted with Et<sub>2</sub>O (×2), and the combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to afford the product (18.4 g). A portion of this crude product (11.4 g) was purified by column chromatography on silica gel eluting with EtOAc/hexane (1:40) to afford **11** (7.95 g, 89%) as a colorless oil. The <sup>1</sup>H NMR data obtained for **11** were in agreement with values previously reported for *ent*-**11**.<sup>3</sup> <sup>-1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz):  $\delta = 5.54$  (brs, 1H), 4.16 (qd, *J* = 7.0, 2.0 Hz, 2H), 3.94 (brddd, *J* = 13.0, 2.5, 2.0 Hz, 1H), 2.18–2.04 (m, 1H), 1.94 (ddd, *J* = 12.5, 7.0, 2.0 Hz, 1H), 1.80–1.70 (m, 1H), 1.55–1.45 (m, 2H), 1.29 (t, *J* = 7.0 Hz, 3H), 0.90 (d, *J* = 7.0 Hz, 3H).



**2-((3***R***,6***S***)-3-Isopropyl-6-methylcyclohex-1-enyl)ethanol (14).** To a solution of ester **11** (2.50 g, 11.1 mmol) dissolved in a mixture of methanol (16 mL) and water (12 mL) was added potassium hydroxide (4.06 g, 72.4 mmol). The solution was heated at 60 °C for 3 h and washed with  $Et_2O$  (×3). The aqueous layer was acidified with 6 N HCl, and extracted with EtOAc (×3). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to afford the crude product **12** (2.44 g), which was used for the subsequent reactions without purification.

A solution of 2,2,6,6-tetramethylpiperidine (15 mL, 89.1 mmol) in THF (46 mL) was degassed by three freeze-thaw cycles. To this solution cooled to -20 °C (a sodium chloride/ice bath) under argon atmosphere was added a solution of *n*-butyllithium (1.15 M in hexane, 39 mL, 44.6 mmol) dropwise. The solution was warmed to 0 °C and stirred at 0 °C for 50 minutes. To this solution was added 12 (2.44 g) in THF (10 mL) at 0 °C. The cooling bath was removed and stirring continued at room temperature for 5 h. The reaction mixture was acidified with aqueous 6 N HCl, and the separated aqueous layer was extracted with ether ( $\times$ 3). The combined organic layer was extracted with aqueous 1 N NaOH. The separated aqueous layer was washed with Et<sub>2</sub>O (×3), acidified with 6 M HCl, and extracted with EtOAc (×3). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to afford the crude product 13 (2.29 g), which was used for the subsequent reactions without purification. To a suspension of lithium aluminum hydride (1.27 g, 33.4 mmol) in Et<sub>2</sub>O (45 mL) cooled to 0 °C under nitrogen atmosphere was added a solution of 13 (2.29 g) in Et<sub>2</sub>O (15 mL). The cooling bath was removed and stirring was continued for 5 h at room temperature. The reaction mixture was cooled to 0 °C and treated with several drops of ethanol to quench the reaction and aqueous potassium sodium (+)-tartrate tetrahydrate solution (12.6 g). After stirring at room temperature for 1 h, the solution was filtered on the pad of Celite. The separated aqueous layer was extracted with Et<sub>2</sub>O (×3). The combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and concentrated under reduced pressure to afford crude residue, which was purified by silica-gel column chromatography (EtOAc/hexane 1:7) to provide 14 (1.54 g, 12.5 mmol, 76% for three steps) as a pale yellow oil. The <sup>1</sup>H and <sup>13</sup>C NMR data obtained for 14 are in good agreement with values previously reported.<sup>1</sup>  $[\alpha]_D^{23} = -39.6$  (c = 1.15, CH<sub>2</sub>Cl<sub>2</sub>) {Literature reported for ent-(+)-14:  $[\alpha]_D^{25} = +39.3$  (c = 1, CH<sub>2</sub>Cl<sub>2</sub>)<sup>1</sup>}{Literature reported for *ent*-(+)-14:  $[\alpha]_D^{20} = +34$  (*c* = 2.20, CHCl<sub>3</sub>)<sup>2</sup>}; IR (KBr):  $v_{max} = 3327, 2956, 2930, 2871, 1463,$ 1446, 1385, 1367, 1043 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta = 5.39$  (brs, 1H), 3.71–3.57 (m, 2H), 2.47–2.39 (m, 1H), 2.17 (dq, J = 8.5, 7.0 Hz, 1H), 2.14–2.06 (m, 1H), 1.97–1.90 (m, 1H), 1.84 (dddd, *J* = 9.5, 5.0, 4.5, 2.0 Hz, 1H), 1.66 (dddd, *J* = 10.0, 5.0, 5.0, 1.0 Hz, 1H), 1.61–1.51 (m, 1H), 1.23 (dddd, J = 11.0, 9.5, 6.5, 2.0 Hz, 1H), 1.19 (dddd, J = 10.5, 9.5, 6.5, 2.0 Hz, 1H), 0.99 (d, J = 7.0 Hz, 3H), 0.88 (d, J = 7.0 Hz, 3H), 0.85 (d, J = 7.0 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta = 138.2, 129.0, 60.4, 42.3, 38.1, 32.3, 32.2, 32.0, 24.7, 19.9, 19.7, 19.4;$  HRMS (ESI): m/z calcd for C<sub>12</sub>H<sub>23</sub>O [M+H]<sup>+</sup>, 183.1743, found 183. 1745.





3-((3R,6S)-3-Isopropyl-6-methylcyclohex-1-enyl)propanenitrile (17). To a solution of p-toluenesulfonyl chloride (3.92 g, 20.6 mmol), 4-dimethylaminopyridine (251 mg, 2.05 mmol) and triethylamine (4.4 mL, 32 mmol) dissolved in CH<sub>2</sub>Cl<sub>2</sub> (75 mL) cooled to 0 °C was added a solution of 14 (2.50 g, 13.7 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (5.0 mL). After stirring at room temperature for 2 h, saturated aqueous NaHCO<sub>3</sub> was added. The separated aqueous layer was extracted with Et<sub>2</sub>O (×3). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>. Concentration under reduced pressure gave 16, which was successively dissolved in DMSO (75 mL). This solution was treated with sodium cyanide (1.21 g, 24.7 mmol), and heated at 80 °C for 2 h. After cooling to room temperature, the reaction mixture was diluted with H<sub>2</sub>O and extracted with EtOAc (×3). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to afford a residue, which was purified by silica-gel column chromatography (EtOAc/hexane 1:10) to provide 17 (2.39 g, 12.5 mmol, 91% for two steps) as a pale yellow oil. 2-((3*R*,6*S*)-3-Isopropyl-6-methylcyclohex-1-enyl)ethyl 4-methylbenzenesulfonate (16):  $[\alpha]_D^{24} =$ -23.4 (*c* = 2.66, CHCl<sub>3</sub>); IR (KBr): *v*<sub>max</sub> = 2957, 2929, 2871, 1599, 1463, 1362, 1177, 961, 909, 815 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  = 7.79 (d, J = 8.0 Hz, 2H), 7.34 (d, J = 8.0 Hz, 2H), 5.27 (brs, 1H), 4.07 (ddd, J = 14.5, 9.5, 8.0 Hz, 1H), 4.06 (ddd, J = 14.5, 9.5, 7.5 Hz, 1H), 2.45 (s, 1H), 2.44–2.36 (m, 1H), 2.28 (ddd, J = 14.0, 8.0, 7.5 Hz, 1H), 2.01–1.92 (m, 1H), 1.88–1.81 (m, 1H), 1.77 (dddd, *J* = 10.0, 5.0, 4.5, 2.0 Hz, 1H), 1.60 (dddd, *J* = 10.0, 5.0, 4.5, 1.0 Hz, 1H), 1.50 (qd, J = 6.5, 1.5 Hz, 1H), 1.15 (dddd, J = 11.0, 10.0, 6.5, 2.0 Hz, 1H), 1.13 (dddd, J = 11.5, 10.0, 6.5, 1.0 Hz, 1H), 0.90 (d, J = 7.0 Hz, 3H), 0.84 (d, J = 6.5 Hz, 3H), 0.81 (d, J = 6.5 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  = 144.7, 136.7, 133.4, 129.8, 129.1, 127.9, 69.4, 42.2, 34.2, 32.3, 32.2, 24.3, 21.7, 19.8, 19.7, 19.3; HRMS (ESI): m/z calcd for C<sub>19</sub>H<sub>28</sub>O<sub>3</sub>NaS [M+Na]<sup>+</sup>, 359.1655, found 359.1650. Compound 17:  $[\alpha]_D^{25} = -26.1$  (*c* = 2.20, CHCl<sub>3</sub>); IR (KBr):  $v_{max} =$ 

2957, 2931, 2871, 2246, 1463, 1445, 1385, 1368 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  = 5.41 (brs, 1H), 2.51–2.26 (m, 4H), 2.16–2.06 (m, 1H), 1.96–1.89 (m, 1H), 1.86 (dddd, *J* = 10.0, 5.5, 5.0, 2.0 Hz, 1H), 1.66 (dddd, *J* = 10.0, 5.5, 5.0, 1.5 Hz, 1H), 1.59 (qd, *J* = 7.0, 1.5 Hz, 1H), 1.25 (dddd, *J* = 11.5, 10.0, 6.0, 2.0 Hz, 1H), 1.21 (dddd, *J* = 12.0, 10.0, 6.0, 1.5 Hz, 1H), 0.99 (d, *J* = 7.0 Hz, 3H), 0.91 (d, *J* = 7.0 Hz, 3H), 0.87 (d, *J* = 7.0 Hz, 3H); <sup>13</sup>C NMR (C<sub>6</sub>D<sub>6</sub>, 100 MHz)  $\delta$  = 138.4, 128.4, 119.7, 42.2, 32.3, 32.0, 31.9, 30.6, 21.3, 19.8, 19.6, 19.3, 16.5; HRMS (ESI): m/z calcd for C<sub>13</sub>H<sub>22</sub>N [M+H]<sup>+</sup>, 192.1747, found 192.1748.



Ethyl 3-((3R,6S)-3-isopropyl-6-methylcyclohex-1-enyl)propanoate (19). A solution of 17 (1.80 g, 9.40 mmol) and potassium hydroxide (2.64g, 47.0 mmol) dissolved in a mixture of EtOH (25 mL) and H<sub>2</sub>O (25 mL) was heated at 100 °C for 22 h. After cooling to room temperature, the reaction mixture was extracted with  $Et_2O$  (×3). The aqueous layer was acidified with 1 M HCl (10 mL), and then extracted with EtOAc (×3). The combined organic layers were dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure to afford crude carboxylic acid 18, which was dissolved in DMF (62 mL). To this solution was added solid K<sub>2</sub>CO<sub>3</sub> (6.49 g, 47.0 mmol). After stirring at room temperature for 1 h, ethyl bromide (0.84 mL, 11.3 mmol) was added. The reaction mixture was stirred at room temperature for 2 h, diluted with H<sub>2</sub>O, neutralized with 1 M HCl, and then extracted with a 3:1 mixture of hexane and EtOAc (×3). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure. The crude product was purified by silica-gel column chromatography (EtOAc/hexane 1:30) to provide 19 (1.84 g, 97% calculated based on consumed 18) and recovered 18 (275 mg, 15%). 3-((3R,6S)-3-Isopropyl-6-methylcyclohex-1-en-1-yl)propanoic acid (18):  $[\alpha]_D^{25} = -3.9$  (c = 4.30, CHCl<sub>3</sub>); IR (KBr):  $v_{max} = 2957$ , 2930, 2872, 2660, 1713, 1445, 1297, 934 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  = 5.32 (brs, 1H), 2.55–2.27 (m, 4H), 2.16–2.07 (m, 1H), 1.94–1.87 (m, 1H), 1.87–1.79 (m, 1H), 1.67–1.59 (m, 1H), 1.55 (qd, *J* = 7.0, 1.5 Hz, 1H), 1.21 (dddd, J = 12.0, 10.0, 6.0, 2.0 Hz, 1H),1.18 (dddd, J = 12.5, 10.0, 6.0, 2.0 Hz, 1H), 0.99 (d, J = 7.0 Hz, 3H), 0.86 (d, J = 7.0 Hz, 3H), 0.83 (d, J = 7.0 Hz, 3H); <sup>13</sup>C NMR  $(CDCl_3, 100 \text{ MHz}) \delta = 180.2, 140.1, 126.5, 77.4, 77.1, 76.8, 42.1, 33.1, 32.4, 32.3, 32.3, 30.0,$ 24.5, 19.7, 19.7, 19.3; HRMS (ESI): m/z calcd for C13H21O2 [M-H]-, 209.1547, found 209.1538. The characterization data obtained for compound 19 are in agreement with values

previously reported.<sup>4</sup> Compound **19**:  $[\alpha]_D^{25} = -10.9 (c = 3.18, CHCl_3)$ ; IR (KBr):  $\nu_{max} = 2957$ , 2931, 2872, 1739, 1463, 1445, 1369, 1162, 1045 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta = 5.30$  (brs, 1H), 4.12 (q, J = 7.0 Hz, 2H), 2.50–2.27 (m, 4H), 2.15–2.06 (m, 1H), 1.92–1.78 (m, 2H), 1.66–1.59 (m, 1H), 1.57–1.49 (m, 1H), 1.25 (t, J = 7.0 Hz, 3H), 1.24–1.15 (m, 2H), 0.99 (d, J = 7.0 Hz, 3H), 0.86 (d, J = 7.0 Hz, 3H), 0.83 (d, J = 7.0 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta = 173.7$ , 140.5, 126.0, 60.3, 42.1, 33.4, 32.4, 32.3, 30.2, 24.6, 19.7, 19.4, 14.3; HRMS (ESI): m/z calcd for C<sub>15</sub>H<sub>27</sub>O<sub>2</sub> [M+H]<sup>+</sup>, 239.2006, found 239.2004.



4-((3R,6S)-3-Isopropyl-6-methylcyclohex-1-enyl)-1-(phenylperoxythio)butan-2-one (20). Methyl phenyl sulfone (4.22 g, 27.0 mmol) was dissolved in toluene and the solvent is removed azeotropically by rotary evaporation ( $\times$ 3). After this procedure followed by drying under vacuum, methyl phenyl sulfone was dissolved in THF (85 mL). To this solution cooled to -78 °C under nitrogen atmosphere, n-butyllithium (1.5 M solution in hexane, 15.4 mL, 23.2 mmol) was added. The solution was warmed to 0 °C and stirred at 0 °C for 30 min. To this solution cooled to -78 °C was added a solution of ester 19 (1.84 g, 7.72 mmol) in THF (5.0 The reaction mixture was stirred at -78 °C for 30 min and warmed at 0 °C. After mL). stirring at 0 °C for 30 min, saturated aqueous ammonium chloride was added. The separated aqueous layer was extracted with EtOAc (×3). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure. The resulting residue was purified by column chromatography on silica gel eluting with EtOAc/hexane (1:4) to afford 20 (2.72g, quant.) as a colorless oil. The characterization data obtained for 20 are in agreement with values previously reported.4



1-Diazo-4-((3R,6S)-3-isopropyl-6-methylcyclohex-1-enyl)-1-(phenylperoxythio)butan-2-one (7). To a solution of 20 (4.22 g, 12.1 mmol) and *p*-acetamidobenzenesulfonyl azide (3.20 g, 13.3 mmol) dissolved in acetonitrile (120 mL) cooled to 0 °C was added triethylamine (5.10 mL, 36.3 mmol) dropwise under nitrogen atmosphere. The reaction mixture was stirred at room temperature for 16 h and then filtered through a pad of Celite. The filtered cake was washed with a 1:5 mixture of EtOAc and hexane. The combined filtrate was concentrated under reduced pressure, and the resulting crude product was purified by silica-gel column chromatography (EtOAc/hexane, 1:5) to afford 7 (4.04 g, 89%) as a yellow oil. The characterization data obtained for compound 7 are in agreement with values previously reported.<sup>4</sup>



(3b*R*,4*R*,7*S*,7a*S*)-4-Isopropyl-7-methyl-3a-(phenylsulfonyl)octahydro-3*H*-cyclopenta[1,3]cy clopropa[1,2]benzen-3-one (8). To a solution of bis-(5-fluoro-(*N*-methylsalicyl aldiminato))copper (II) (21) (63 mg, 0.17 mmol) in toluene (60 mL) was added a solution of diazo compound 7 (800 mg, 2.14 mmol) dissolved in toluene (20 mL). The solution was heated at 80 °C for 15 h under nitrogen atmosphere, cooled to room temperature and concentrated under reduced pressure. The crude product was purified by silica-gel column chromatography (EtOAc/hexane 1:8 and EtOAc/ toluene 1:30) to afford 8 (344 mg, 46%) as a white solid. The characterization data obtained for compound 8 are in agreement with values previously reported.<sup>4</sup>

### Separation of cyclopropane intermediate 8 and its ent-epimer

ent-Epimer of 8 is an intermediate in our previously reported synthesis of exigurin.<sup>4</sup>



A solution of the substance to be examined was spotted on a 0.25 mm E. Merck silica plate (60F-254). The plate was inserted into a developing chamber containing a mixture of v/v 8:1 hexane/ethyl acetate. After the solvent had risen to near the top of the plate, the plate was removed from the developing chamber and the solvent front was marked with a pencil. The resulting plate was returned to the chamber, developed, removed from the chamber, and allow the solvent to evaporate. After one more cycle of this process, we observed two cleanly separated spots (line 2).



Lane 1: Cyclopropane intermediate **8** Lane 2: Co-spot. Lane 3: *ent*-epimer

Visualizing agent: *p*-anisaldehyde



(1R,5S,6S,7R,10S)-6-Azido-7-isopropyl-10-methyl-1-(phenylsulfonyl)spiro[4.5]decan-2-one (22). To a solution of 8 (610 mg, 1.77 mmol) dissolved in DMF (24 mL) were added sodium 38.9 mmol), magnesium perchlorate azide 5.30 (2.53)g, (1.18 g, mmol) and tetrabutylammonium hydrogen sulfate (7.20 g, 21.2 mmol). The mixture was heated under nitrogen atmosphere at 100 °C for 14 h. After cooling to room temperature, the reaction mixture was diluted with water (125 mL) and then extracted with EtOAc (×3). The combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure. The crude product was purified by silica-gel column chromatography (EtOAc/hexane 1:6) to afford 22 (528 mg, 88%, calculated based on consumed 8) as a white solid and recovered 8 (76 mg, 12%). Mp 40–41 °C;  $[\alpha]_D^{25} = -36.3$  (*c* = 1.54, CHCl<sub>3</sub>); IR (KBr):  $v_{max} = 2961, 2928, 2881, 2101, 1747,$ 1448, 1311, 1141, 712 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  = 7.80–7.76 (m, 2H), 7.72–7.67 (m, 1H), 7.60–7.54 (m, 2H), 4.80 (s, 1H), 3.61 (s, 1H), 2.87–2.66 (m, 2H), 2.41–2.29 (m, 1H), 2.20–2.12 (m, 2H), 2.04–1.94 (m, 2H), 1.76–1.63 (m, 2H), 1.50 (ddd, *J* = 14.0, 6.0, 3.0 Hz, 1H), 1.34–1.22 (m, 1H), 1.14 (d, J = 6.5 Hz, 3H), 1.11–1.03 (m, 1H), 1.01 (d, J = 6.5 Hz, 3H), 0.61 (d, J = 6.5 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta = 209.0$ , 137.8, 134.4, 129.1, 128.9, 75.5, 68.9, 54.0, 37.9, 36.7, 33.4, 31.5, 30.5, 24.2, 20.9, 20.4, 17.1; HRMS (ESI): m/z calcd for C<sub>20</sub>H<sub>27</sub>O<sub>3</sub>N<sub>3</sub>NaS [M+Na]<sup>+</sup>, 412.1665, found 412.1664.



(5S,6S,7R,10S)-6-Azido-7-isopropyl-10-methyl-1-(phenylsulfonyl)spiro[4.5]dec-1-en-2-yl trifluoromethanesulfonate (23). To a solution of 22 (519 mg, 1.33 mmol) in 1,2-dimethoxyethane (24 mL) was added sodium hydride (55% dispersion in mineral oil, 240 mg, 6.65 mmol). The mixture was heated under nitrogen atmosphere at 40 °C for 30 min. After cooling to 0 °C, trifluoromethanesulfonic anhydride (0.75mL, 2.66 mmol) was added dropwise. After stirring at 0 °C for 3 h, saturated aqueous NH<sub>4</sub>Cl was added. The separated aqueous layer was extracted with EtOAc (×3), and the combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure. The crude product was purified by silica-gel column chromatography (EtOAc/hexane 1:7) to furnish 23 (643 mg, 93%). Mp 93–94 °C;  $[\alpha]_D^{25} = +15.3$  (*c* = 1.39, CHCl<sub>3</sub>); IR (KBr):  $v_{max} = 2959$ , 2925, 2872, 2102, 1607, 1434, 1331, 1217, 1152, 950, 812 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  = 7.99–7.94 (m, 2H), 7.68–7.62 (m, 1H), 7.59–7.54 (m, 2H), 4.06 (brs, 1H), 2.81 (ddd, J = 13.0, 9.0, 6.5 Hz, 1H), 2.66 (ddd, J = 18.0, 9.0, 5.5 Hz, 1H), 2.25 (ddd, J = 13.5, 9.5, 6.5 Hz, 1H), 2.14–2.04 (m, 2H), 2.00–1.53 (m, 5H), 1.49–1.38 (m, 1H), 1.17 (d, J = 6.5 Hz, 3H), 1.00 (d, J = 7.0 Hz, 3H), 0.98 (d, J = 7.0 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta = 157.2$ , 142.2, 138.1, 133.9, 129.2, 127.8, 119.8, 116.1, 68.6, 61.8, 44.3, 36.2, 34.0, 29.6, 29.3, 23.3, 21.5, 21.0, 17.5, 14.2; HRMS (ESI): m/z calcd for  $C_{21}H_{26}O_5N_3F_3NaS_2$  [M+Na]<sup>+</sup>, 544.1158, found 544.1159.



(5S,6S,7R,10S)-6-Azido-7-isopropyl-2,10-dimethyl-1-(phenylsulfonyl)spiro[4.5]dec-1-ene

(24). To a solution of 23 (420 mg, 0.805 mmol) in 1,4-dioxane (7.0 mL, argon was bubbled through the solution for 60 min) was added tetrakis(triphenylphosphine)palladium (93 mg, 0.81 mmol). After stirring at room temperature for 5 min under argon atmosphere, potassium carbonate (333 mg, 2.42 mmol) and trimethylboroxine (0.12 mL, 0.81 mmol) were added. The reaction mixture was heated at 110 °C for 1 h, diluted with THF and filtered through a pad of silica-gel. Concentration of the filtrate under reduced pressure followed by purification by silica-gel column chromatography (EtOAc/hexane 1:12) provided 24 (304 mg, 98%) as a white solid. Mp 123–124 °C;  $[\alpha]_D^{25} = +15.1$  (c = 1.74, CHCl<sub>3</sub>); IR (KBr):  $v_{max} = 2960$ , 2931, 2871, 2098, 1463, 1446, 1302, 1146, 725, 689 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta = 7.91-7.87$  (m, 2H), 7.60–7.49 (m, 3H), 4.00 (brs, 1H), 2.54 (ddd, J = 18.0, 7.5, 7.5 Hz, 1H), 2.45 (ddd, J = 17.5, 7.5, 6.0 Hz, 1H), 2.17 (ddd, J = 13.0, 8.0, 6.5 Hz, 1H), 2.04 (ddd, J = 13.0, 8.0, 6.0 Hz, 1H), 1.95 (s, 3H), 1.92–1.78 (m, 3H), 1.72–1.31 (m, 4H), 1.13 (d, J = 7.0 Hz, 3H), 0.88 (d, J = 6.5 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta = 162.1$ , 144.3, 140.7, 132.7, 129.2, 126.3, 69.0, 64.9, 44.0, 37.8, 36.2, 35.9, 29.5, 29.1, 23.4, 21.4, 20.8, 18.6, 17.5; HRMS (ESI): m/z calcd for C<sub>21</sub>H<sub>29</sub>O<sub>2</sub>N<sub>3</sub>NaS [M+Na]<sup>+</sup>, 410.1873, found 410.1871.



N-((5R,6S,7R,10S)-7-isopropyl-2,10-dimethylspiro[4.5]dec-1-en-6-yl)formamide (26). To a suspension of magnesium powder (941 mg, 38.7 mmol, activated by washing with 1N HCl and acetone before use) in MeOH (7.5 mL) under nitrogen atmosphere was added a solution of 24 (300 mg, 0.77 mmol) in THF (2.1 mL). The solution was heated to reflux for 30 min, diluted with EtOAc and then filtered through a pad of Celite. The filtrate was concentrated under reduced pressure to give crude amine 25, which was dissolved in EtOAc (15 mL). The solution was treated with acetic formic anhydride (0.15 mL, 1.94 mmol). After stirring at room temperature for 60 min, saturated aqueous NaHCO<sub>3</sub> was added. The separated aqueous layer was extracted with EtOAc (×3), and the combined organic layer was dried (Na<sub>2</sub>SO<sub>4</sub>). Concentration under reduced pressure gave a residue, which was purified by silica-gel column chromatography (EtOAc/hexane 1:3  $\rightarrow$  1:2) to provide 26 (167 mg, 86% for two steps). Formamide 26 exists as a mixture of amide rotamers on the <sup>1</sup>H and <sup>13</sup>C NMR time scale. When two rotamers are observed, signals for the minor rotamers are given in braces. (5R, 6S, 7R, 10S)-7-Isopropyl-2,10-dimethylspiro[4.5]dec-1-en-6-amine (25):  $[\alpha]_D^{25} = +14.2 \ (c = 1.5)$ 1.18, CHCl<sub>3</sub>); IR (KBr):  $v_{max} = 2955$ , 2925, 2870, 1613, 1462, 1375, 846, 819 cm<sup>-1</sup>; <sup>1</sup>H NMR  $(CDCl_3, 400 \text{ MHz}) \delta = 5.30 \text{ (brs, 1H)}, 2.72 \text{ (brs, 1H)}, 2.22-2.15 \text{ (m, 1H)}, 1.83-1.77 \text{ (m, 1H)},$ 1.74 (d, J = 1.0 Hz, 3H), 1.72–1.64 (m, 2H), 1.51–1.38 (m, 2H), 1.37–1.16 (m, 6H), 1.14–1.03 (m, 1H), 0.91 (d, J = 6.5 Hz, 3H), 0.89 (d, J = 6.5 Hz, 3H), 0.74 (d, J = 6.5 Hz, 3H); <sup>13</sup>C NMR  $(CDCl_3, 100 \text{ MHz}) \delta = 141.8, 126.8, 58.7, 58.0, 45.3, 36.6, 35.2, 34.1, 31.8, 29.4, 24.4, 21.5,$ 20.7, 17.1, 16.4; HRMS (ESI): m/z calcd for  $C_{15}H_{28}N$  [M+H]<sup>+</sup>, 222.2216, found 222.2216. Compound **26**: Mp 96–98 °C;  $[\alpha]_D^{25} = -13.9$  (c = 1.55, CHCl<sub>3</sub>); IR (KBr):  $v_{max} = 3306$ , 3044, 2957, 2929, 2870, 1659, 1538, 1463, 1384, 739 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta = 8.27$  (d,

 $J = 1.5 \text{ Hz}, 1\text{H}, \{8.05 \text{ (d, } J = 12.0 \text{ Hz}, 1\text{H})\}, \{5.72-5.64 \text{ (m, 1H)}\}, 5.57-5.49 \text{ (m, 1H)}, 5.26 \text{ (brs}, 1\text{H}), 4.23 \text{ (brd, } J = 11.5 \text{ Hz}, 1\text{H}), \{3.30 \text{ (brdd, } J = 11.5, 2.5 \text{ Hz}, 1\text{H})\}, 2.32-2.08 \text{ (m, 2H)}, \{2.32-2.08 \text{ (m, 2H)}\}, 1.89-1.81 \text{ (m, 2H)}, \{1.89-1.81 \text{ (m, 2H)}\}, \{1.75 \text{ (brs, 3H)}\}, 1.73 \text{ (brs, 3H)}, \{1.72-1.61 \text{ (m, 1H)}\}, 1.64 \text{ (ddd, } J = 13.5, 9.5, 4.0 \text{ Hz}, 1\text{H}), 1.53-1.47 \text{ (m, 1H)}, \{1.53-1.47 \text{ (m, 1H)}\}, 1.43-1.25 \text{ (m, 4H)}, \{1.43-1.25 \text{ (m, 4H)}\}, 1.22-1.10 \text{ (m, 1H)}, \{1.22-1.10 \text{ (m, 1H)}\}, 0.92 \text{ (d, } J = 6.0 \text{ Hz}, 3\text{H}), \{0.90 \text{ (d, } J = 6.5 \text{ Hz}, 3\text{H})\}, 0.88 \text{ (d, } J = 6.0 \text{ Hz}, 3\text{H}), \{0.85 \text{ (d, } J = 6.5 \text{ Hz}, 3\text{H})\}, 0.77 \text{ (d, } J = 6.5 \text{ Hz}, 3\text{H}); {}^{13}\text{C} \text{ NMR} \text{ (CDCl}_3, 100 \text{ MHz}) \delta = \{164.4\}, 161.1, \{143.8\}, 143.6, \{125.2\}, 125.1, \{59.5\}, 58.1, \{57.9\}, 54.0, 44.3, \{44.1\}, \{36.2\}, 36.1, \{36.0\}, 35.7, \{34.7\}, 33.7, 31.6, \{31.3\}, 29.6, \{28.8\}, 25.4, \{25.1\}, 21.3, 21.0, \{20.8\}, \{20.7\}, 17.1, \{17.0\}, 16.3, \{16.2\}; \text{HRMS} \text{ (ESI): m/z calcd for C}_{16}\text{H}_{28}\text{ON} \text{ [M+H]}^+, 250.2165, \text{found } 250.2165.$ 



(+)-Axisonitrile- 3 (4). To a solution of 26 (165 mg, 0.66 mol) and triethylamine (0.65 mL, 4.63 mmol) dissolved in CH<sub>2</sub>Cl<sub>2</sub> (2.4 mL) cooled to -20 °C was added triphosgene (118 mg, 0.40 mmol). After stirring at -20 °C for 10 min, the reaction mixture was warmed to 0 °C and then treated with saturated aqueous of NaHCO<sub>3</sub> and CH<sub>2</sub>Cl<sub>2</sub>. After vigorous stirring at 0 °C for 1 h, the separated aqueous layer was extracted with EtOAc (×3). The combined organic layers were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure. The crude product was purified by silica-gel column chromatography (EtOAc/hexane 1:10) to afford (+)-axisonitrile-3 (4) (133 mg, 87%) as a white crystalline mass. Mp 99–100 °C (Literature: 101–103 °C<sup>5</sup>) (Literature: 99–102 °C<sup>6</sup>) (Literature: 94–95°C<sup>7</sup>);  $[\alpha]_D^{22} = +89.2$  (c = 1.0, CHCl<sub>3</sub>) {Literature:  $[\alpha]_D^{22} =$ +68.4 (c = 1, CHCl<sub>3</sub>)<sup>5</sup>} {Literature:  $[\alpha]_D^{22} = +43.4$  (c = 0.006, CHCl<sub>3</sub>)<sup>6</sup>} {Literature:  $[\alpha]_D^{22} = -43.4$ +54.4 (c = 0.10, CHCl<sub>3</sub>)<sup>7</sup>}; IR (KBr):  $v_{max} = 2959$ , 2920, 2893, 2853, 2133, 1655, 1458, 1442, 1373, 1328 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  = 5.13 (q, J = 1.0 Hz, 1H), 3.59 (brs, 1H), 2.28–2.20 (m, 2H), 1.98–1.92 (m, 2H), 1.84–1.76 (m, 2H), 1.74 (brs, 3H), 1.59 (qd, J = 6.5, 2.5 Hz, 1H), 1.54-1.48 (m, 1H), 1.40-1.27 (m, 1H), 1.21-1.10 (m, 2H), 0.94 (d, J = 6.5 Hz, 3H), 0.91 (d, J = 6.5 Hz, 3H), 0.76 (d, J = 7.0 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta = 155.7$  (t), 144.8, 123.7, 64.6 (t), 57.1, 43.8, 35.9, 35.0, 34.4, 31.2, 29.8, 24.9, 20.8, 20.4, 17.0, 16.1; HRMS (ESI): m/z calcd for  $C_{16}H_{29}N_2$  [M+NH<sub>4</sub>]<sup>+</sup>, 249.2325, found 249.2325.



Boneratamide A methyl ester (1b). A solution of L-glutamic acid 1-methyl ester (27) (10 mg, 0.065 mmol) and acetone (0.030 mL, 0.43 mmol) in MeOH (0.60 mL) under nitrogen atmosphere was heated at 50 °C for 60 min. To this mixture was added (+)-axisonitrile-3 (4) (10 mg, 0.043 mmol). After stirring at 50 °C for 60 min and at 80 °C for 120 min, the reaction mixture was cooled to room temperature and diluted with H<sub>2</sub>O. The aqueous layer was extracted with EtOAc (×3), and the combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure. The resulting crude mixture of 1a and 1b was dissolved in a mixture of benzene (0.40 mL) and methanol (0.20 mL), and then treated with trimethylsilyldiazomethane (2.0 M solution in ether, 0.21mL, 0.42 mmol). After stirring at room temperature for 16 h, the solution was concentrated under reduced pressure to afford a crude product, which was purified by silica-gel chromatography (EtOAc/hexane 1:3) to furnish boneratamide A methyl ester (1b) (14 mg, 73%) as a white solid. Mp 165-166 °C (recrystallized from AcOEt/hexane);  $[\alpha]_{D}^{28} = +2.6$  (c = 1.31, CH<sub>2</sub>Cl<sub>2</sub>),  $[\alpha]_{D}^{20} = +8.8$  (c = 1.73, MeOH); IR (KBr): v<sub>max</sub> = 3366, 2956, 2929, 2871, 1732, 1708, 1673, 1530, 1463, 1439, 1393, 1220, 1180 cm<sup>-1</sup>; <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz)  $\delta$  = 7.51 (d, *J* = 10.5 Hz, 1H), 5.41 (q, *J* = 1.0 Hz, 1H) 1H), 4.66 (dd, J = 10.5, 2.5 Hz, 1H), 3.76 (dd, J = 10.0, 1.5 Hz, 1H), 3.16 (s, 3H), 2.66 (ddd, J = 10.0, 1.5 Hz, 1H), 3.16 (s, 3H), 2.66 (ddd, J = 10.0, 1.5 Hz, 1H), 3.16 (s, 3H), 2.66 (ddd, J = 10.0, 1.5 Hz, 1H), 3.16 (s, 3H), 2.66 (ddd, J = 10.0, 1.5 Hz, 1H), 3.16 (s, 3H), 2.66 (ddd, J = 10.0, 1.5 Hz, 1H), 3.16 (s, 3H), 2.66 (ddd, J = 10.0, 1.5 Hz, 1H), 3.16 (s, 3H), 2.66 (ddd, J = 10.0, 1.5 Hz, 1H), 3.16 (s, 3H), 3.16 (s, 3 13.5, 9.5, 5.5 Hz, 1H), 2.47–2.37 (m, 1H), 2.21–2.04 (m, 3H), 1.95–1.88 (m, 1H), 1.86–1.68 (m, 3H), 1.79 (ddd, J = 16.5, 9.5, 1.5 Hz, 1H), 1.66 (s, 3H), 1.62 (d, J = 1.0 Hz, 3H), 1.51–1.39 (m, 3H), 1.36–1.25 (m, 2H), 1.24 (s, 3H), 1.17 (d, J = 6.5 Hz, 3H), 0.98 (d, J = 6.5 Hz, 6H); <sup>13</sup>C NMR (C<sub>6</sub>D<sub>6</sub>, 100 MHz)  $\delta$  = 175.5, 175.3, 175.1, 143.7, 126.3, 60.5, 58.9, 56.3, 45.6, 36.6, 35.6, 34.1, 32.4, 32.0, 30.2, 30.1, 25.6, 23.3, 23.1, 21.8, 17.2, 17.1, 14.4; HRMS (ESI): m/z calcd for  $C_{25}H_{41}O_4N_2$  [M+H]<sup>+</sup>, 433.3061, found 433.3061.



Boneratamide A (1a). A suspension of L-glutamic acid (5) (11 mg, 0.072 mmol), acetone (0.10 mL, 1.30 mmol) and MeOH (0.60 mL) in a sealed test tube under nitrogen atmosphere was heated at 50 °C for 2 h. To this mixture was added (+)-axisonitrile-3 (4) (10 mg, 0.043 mmol) at room temperature. The reaction mixture was heated at 50 °C for 90 min and then at 80 °C for 12 h . After cooling at room temperature, concentration under reduced pressure afforded a residue, which was purified by silica-gel chromatography (CHCl<sub>3</sub>/MeOH 7:1) to furnish boneratamide A (1a) (14 mg, 76%) as a white solid. Mp 222–223 °C;  $[\alpha]_D^{28} = +13.8$  (c = 0.54, CH<sub>2</sub>Cl<sub>2</sub>); IR (KBr):  $v_{max} = 3330$ , 2956, 2927, 2870, 1716, 1654, 1541, 1463, 1386, 1229 cm<sup>-1</sup>; <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz)  $\delta$  = 7.36–7.22 (m, 1H), 5.31 (brs, 1H), 4.50 (brd, J = 10.0 Hz, 1H), 4.04 (brd, J = 9.5 Hz, 1H), 2.37–2.06 (m, 5H), 1.91–1.65 (m, 5H), 1.63 (brs, 3H), 1.49 (brs, 3H), 1.40–1.25 (m, 5H), 1.23 (brs, 3H), 1.15 (d, J = 6.5 Hz, 3H), 0.97 (d, J = 6.5 Hz, 3H), 0.91 (d, J = 6.5 Hz, 3H); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta = 6.31$  (brd, J = 10.5 Hz, 1H), 5.24 (brs, 1H), 4.41 (d, J = 8.5 Hz, 1H), 4.16 (dd, J = 10.5, 2.5 Hz, 1H), 2.55–2.08 (m, 6H), 1.96–1.87 (m, 1H), 1.79-1.71 (m, 1H), 1.74 (brs, 3H), 1.68 (s, 3H), 1.65-1.50 (m, 3H), 1.48 (s, 3H), 1.38-1.26 (m, 2H), 1.24–1.12 (m, 1H), 1.04–0.92 (m, 1H), 0.90 (d, *J* = 6.5 Hz, 3H), 0.89 (d, *J* = 6.5 Hz, 3H), 0.78 (d, J = 6.5 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta = 176.3$ , 175.4, 175.3, 143.6, 125.4, 60.2, 58.5, 58.4, 56.1, 44.9, 36.3, 35.6, 33.6, 31.8, 30.0, 29.8, 29.5, 25.9, 25.7, 25.2, 23.1, 21.4, 17.1, 16.5; HRMS (ESI): m/z calcd for C<sub>24</sub>H<sub>39</sub>O<sub>4</sub>N<sub>2</sub> [M+H]<sup>+</sup>, 419.2904, found 419.2904.



Methyl esters of boneratamide B (2b) and C (3b). A solution of L-glutamic acid 1-methyl ester (27) (8 mg, 0.048 mmol) and acetaldehyde (0.075 mL, 1.30 mmol) in MeOH (0.30 mL) was stirred at room temperature for 60 min. To this mixture was added a solution of axisonitrile-3 (4) (10 mg, 0.043 mmol) dissolved in a mixture of benzene (0.025 mL) and methanol (0.10 mL). The reaction mixture was stirred at room temperature for 3 h, and then diluted with water. The aqueous layer was extracted with EtOAc (×3), and the combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to afford a residue, which was dissolved in a mixture of benzene (0.44 mL) and methanol (0.21 mL). The solution was treated with trimethylsilyldiazomethane (2.0 M solution in ether, 0.21mL, 0.42 mmol), stirred at room temperature for 12 h and then concentrated under reduced pressure. The crude product was purified by silica-gel chromatography (EtOAc/hexane 1:2) to furnish boneratamide B methyl ester (2b) (7.1 mg, 39%) as a colorless oil and boneratamide C methyl ester (3b) (2.8 mg, 15%) as a white solid. Boneratamide B methyl ester (2b):  $[\alpha]_D^{26} = +15.2$  $(c = 2.15, \text{CH}_2\text{Cl}_2), \ [\alpha]_D^{26} = +19.9 \ (c = 2.07, \text{CHCl}_3); \text{IR (KBr): } \nu_{\text{max}} = 3338, 2955, 2930, 2871,$ 1746, 1667, 1537, 1438, 1416, 1204, 1179 cm<sup>-1</sup>; <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz)  $\delta = 6.92$  (d, J =10.5 Hz, 1H), 5.30 (q, J = 1.5 Hz, 1H), 4.48 (dd, J = 10.5, 2.5 Hz, 1H), 4.46 (q, J = 7.0 Hz, 1H), 4.11 (dd, J = 9.5, 2.0 Hz, 1H), 3.22 (s, 3H), 2.31–2.21 (m, 2H), 2.16–2.08 (m, 2H), 1.87 (ddd, J = 17.0, 10.0, 2.5 Hz, 1H, 1.75-1.62 (m, 4H), 1.61 (d, J = 1.0 Hz, 3H), 1.53-1.40 (m, 3H), 1.37 (m, 3H)(d, J = 7.0 Hz, 3H), 1.32 - 1.27 (m, 1H), 1.20 - 1.12 (m, 2H), 1.11 (d, J = 7.0 Hz, 3H), 0.90 (d, J = 7.0 Hz, 3H)6.5 Hz, 3H), 0.86 (d, J = 6.5 Hz, 3H); <sup>13</sup>C NMR (C<sub>6</sub>D<sub>6</sub>, 100 MHz)  $\delta = 175.4$  173.2, 170.5, 142.8, 126.0, 58.7, 58.6, 55.0, 52.9, 51.6, 44.8, 36.0, 35.2, 33.4, 32.0, 29.7, 29.5, 25.3, 24.1, 21.4, 21.0,

16.7, 16.5, 14.2; HRMS (ESI): m/z calcd for C<sub>24</sub>H<sub>39</sub>O<sub>4</sub>N<sub>2</sub> [M+H]<sup>+</sup>, 419.2904, found 419.2904. Boneratamide C methyl ester (**3b**): Mp 103–104 °C (recrystallized from benzene/hexane);  $[\alpha]_D^{26}$ = -29.5 (*c* = 1.00, CH<sub>2</sub>Cl<sub>2</sub>),  $[\alpha]_D^{27}$  = -35.7 (*c* = 1.00, CHCl<sub>3</sub>); IR (KBr): v<sub>max</sub> = 3339, 2954, 2927, 2870, 1746, 1666, 1541, 1438, 1208, 1178 cm<sup>-1</sup>; <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 400 MHz) δ = 7.21 (d, *J* = 11.0 Hz, 1H), 5.32 (q, *J* = 1.5 Hz, 1H), 4.53 (q, *J* = 7.0 Hz, 1H), 4.46 (brd, *J* = 11.0 Hz, 1H), 3.86 (dd, *J* = 9.0, 3.0 Hz, 1H), 3.34 (s, 3H), 2.34–2.03 (m, 4H), 1.84 (ddd, *J* = 17.0, 9.5, 3.5 Hz, 1H), 1.82–1.76 (m, 1H), 1.77 (ddd, *J* = 13.0, 9.0, 4.0 Hz, 1H), 1.64–1.60 (m, 2H), 1.59 (d, *J* = 1.0 Hz, 3H), 1.53–1.50 (m, 1H), 1.46–1.35 (m, 2H), 1.34 (d, *J* = 7.0 Hz, 3H), 1.33–1.27 (m, 2H), 1.17 (d, *J* = 6.5 Hz, 3H), 1.15–1.10 (m, 1H), 0.98 (d, *J* = 6.5 Hz, 3H), 0.84 (d, *J* = 6.5 Hz, 3H); <sup>13</sup>C NMR (C<sub>6</sub>D<sub>6</sub>, 100 MHz) δ = 175.8 172.3, 170.0, 143.1, 126.3, 59.2, 58.0, 55.3, 53.3, 51.9, 45.2, 36.2, 35.4, 33.7, 32.3, 29.8, 29.4, 25.5, 23.8, 22.3, 21.4, 17.0, 16.8, 14.7; HRMS (ESI): m/z calcd for C<sub>24</sub>H<sub>39</sub>O<sub>4</sub>N<sub>2</sub> [M+H]<sup>+</sup>, 419.2904, found 419.2904.



(R)-1-((S)-1-(((5R,6S,7R,10S)-7-isopropyl-2,10-dimethylspiro[4.5]dec-1-en-6-yl) Methyl amino)-1-oxopropan-2-yl)-5-oxopyrrolidine-2-carboxylate (28) and methyl (R)-1-((R)-1 -(((5R,6S,7R,10S)-7-isopropyl-2,10-dimethylspiro[4.5]dec-1-en-6-yl)amino)-1-oxopropan-2yl)-5-oxopyrrolidine-2-carboxylate (29). A solution of D-glutamic acid (ent-5) (9.5 mg, 0.065 mmol) and acetaldehyde (0.075 mL, 1.30 mmol) in MeOH (0.30 mL) was stirred at room temperature for 3 h. To this mixture was added a solution of (+)-axisonitrile-3 (4) (10 mg, 0.043 mmol) dissolved in a mixture of benzene (0.050 mL) and methanol (0.10 mL). The reaction mixture was stirred at room temperature for 19 h and diluted with water. The aqueous layer was extracted with EtOAc (×3), and the combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>) and concentrated under reduced pressure to afford a residue, which was dissolved in a mixture of benzene (0.44 mL) and methanol (0.22 mL). The solution was treated with trimethylsilyldiazomethane (2.0 M solution in ether, 0.21 mL, 0.42 mmol), stirred at room temperature for 20 h and then concentrated under reduced pressure. The crude product was purified by silica-gel chromatography (EtOAc/hexane 1:2) to furnish 28 (9.2 mg, 51%) as a white solid and 29 (3.8 mg, 21%) as a colorless oil.

Methyl (*R*)-1-(((*S*)-1-(((*SR*,6*S*,7*R*,10*S*)-7-isopropyl-2,10-dimethylspiro[4.5]dec-1-en-6-yl)amino) -1-oxopropan-2-yl)-5-oxopyrrolidine-2-carboxylate (**28**): Mp 70–71 °C;  $[\alpha]_D^{25} = -51.1$  (*c* = 1.77, CH<sub>2</sub>Cl<sub>2</sub>); IR (KBr):  $v_{max} = 3341$ , 2955, 2930, 2871, 1746, 1667, 1538, 1438, 1415, 1205, 1179 cm<sup>-1</sup>; <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz)  $\delta = 6.78$  (brd, *J* = 10.5 Hz, 1H), 5.28 (q, *J* = 1.5 Hz, 1H), 4.59 (q, *J* = 7.0 Hz, 1H), 4.44 (dd, *J* = 11.0, 2.0 Hz, 1H), 4.19 (dd, *J* = 9.5, 1.5 Hz, 1H), 3.22 (s, 3H), 2.34–2.25 (m, 2H), 2.18–2.04 (m, 2H), 1.87 (ddd, *J* = 17.0, 9.5, 2,5 Hz, 1H), 1.78–1.63 (m, 3H), 1.53–1.41 (m, 3H), 1.36 (d, J = 7.0 Hz, 3H), 1.35–1.16 (m, 3H), 1.14–1.07 (m, 1H), 1.04 (d, J = 6.5 Hz, 3H), 0.87 (d, J = 6.5 Hz, 3H), 0.81 (d, J = 6.5 Hz, 3H); <sup>13</sup>C NMR (C<sub>6</sub>D<sub>6</sub>, 100 MHz)  $\delta = 175.8$ , 173.7, 170.6, 170.5, 143.3, 126.2, 58.9, 58.5, 55.3, 52.7, 51.9, 45.1, 36.3, 35.4, 33.7, 32.2, 30.1, 29.7, 25.6, 24.4, 21.6, 21.4, 17.1, 16.8, 14.6, 14.4; HRMS (ESI): m/z calcd for C<sub>24</sub>H<sub>39</sub>O<sub>4</sub>N<sub>2</sub> [M+H]<sup>+</sup>, 419.2904 , found 419.2904.

Methyl (*R*)-1-(((*R*)-1-(((5R,6S,7R,10S)-7-isopropyl-2,10-dimethylspiro[4.5]dec-1-en-6-yl)amino) -1-oxopropan-2-yl)-5-oxopyrrolidine-2-carboxylate (**29**):  $[\alpha]_{D}^{26} = +8.4$  (*c* = 0.60, CH<sub>2</sub>Cl<sub>2</sub>); IR (KBr):  $\nu_{max} = 3336$ , 2955, 2928, 2871, 1748, 1689, 1664, 1541, 1438, 1419, 1209, 1178 cm<sup>-1</sup>; <sup>1</sup>H NMR (C<sub>6</sub>D<sub>6</sub>, 500 MHz)  $\delta$  = 7.22 (brd, *J* = 10.5 Hz, 1H), 5.33 (q, *J* = 1.5 Hz, 1H), 4.50 (q, *J* = 7.0 Hz, 1H), 4.42 (dd, *J* = 10.5, 2.5 Hz, 1H), 3.89 (dd, *J* = 8.5, 3.0 Hz, 1H), 3.36 (s, 3H), 2.40–2.33 (m, 1H), 2.28 (ddd, *J* = 17.0, 10.0, 10.0 Hz, 1H), 2.22–2.14 (m, 2H), 1.84 (ddd, *J* = 17.0, 9.5, 3.5 Hz, 1H), 1.83 (ddd, *J* = 17.0, 4.5, 4.5 Hz, 1H), 1.79–1.70 (m, 2H), 1.64 (s, 3H), 1.55–1.38 (m, 4H), 1.33 (d, *J* = 7.0 Hz, 3H), 1.31–1.15 (m, 3H), 1.13 (d, *J* = 6.5 Hz, 3H), 0.91 (d, *J* = 6.5 Hz, 3H), 0.88 (d, *J* = 6.5 Hz, 3H);<sup>13</sup>C NMR (C<sub>6</sub>D<sub>6</sub>, 100 MHz)  $\delta$  = 175.8, 172.3, 170.0, 143.0, 126.6, 58.7, 58.3, 55.5, 53.3, 52.0, 45.4, 36.6, 35.4, 33.8, 32.4, 30.1, 29.9, 25.7, 23.9, 21.8, 21.4, 17.1, 17.0, 14.8, 14.4; HRMS (ESI): m/z calcd for C<sub>24</sub>H<sub>39</sub>O<sub>4</sub>N<sub>2</sub> [M+H]<sup>+</sup>, 419.2904 , found 419.2904.



(+)-axisonitrile-3 (4)

<sup>1</sup> H NMR (δ) Synthetic Sample (400 MHz, CDCl <sub>3</sub> )	<sup>1</sup> H NMR (δ) Natural Sample <sup>6</sup> (400 MHz, CDCl <sub>3</sub> )
5.13 (1H, q, <i>J</i> = 1.0)	5.11 (1H, br s, H-1)
3.59 (1H, brs)	3.57 (1H, br s, H-6)
2.20–2.28 (2H, m)	2.21 (2H, m, H-3a, H-3b)
1.92–1.98 (2H, m)	1.93 (2H, m, H-4a, H-4b)
1.76–1.84 (2H, m)	1.77 (2H, m, H-8a, H-10)
1.74 (3H, brs)	1.72 (3H, br s, H-14)
1.59 (1H, qd, <i>J</i> = 6.5, 2.5)	1.57 (1H, m, H-11)
1.48–1.54 (1H, m)	1.48 (1H, m, H-9a)
1.27–1.40 (1H, m)	1.33 (1H, ddd, <i>J</i> = 13.0, 13.0, 4.0)
1.10–1.21 (2H, m)	1.13 (1H, m, H-7)
_	1.04 (1H, m, H-9b)
0.94 (3H, d, <i>J</i> = 6.5)	0.92 (3H, d, <i>J</i> = 6.6, H-12)
0.91 (3H, d, <i>J</i> = 6.5)	0.89 (3H, d, <i>J</i> = 6.6, H-13)
0.76 (3H, d, <i>J</i> = 7.0)	0.74 (3H, d, <i>J</i> = 7.0, H-15)

Coupling constants,  $J_{H-H}$  (in Hz), are given in parentheses.



(+)-axisonitrile-3 (4)

	<sup>13</sup> C NMR (δ)	$^{13}$ C NMR ( $\delta$ )	
Position	(100 MHz, CDCl <sub>3</sub> )	(100 MHz, CDCl <sub>3</sub> )	Δδ
1	123.7	123.6	+0.1
2	144.8	144.8	0
3	35.9	35.8	+0.1
4	35.0	34.9	+0.1
5	57.1	57.0	+0.1
6	64.6	64.5	+0.1
7	43.8	43.8	0
8	24.9	24.9	0
9	31.2	31.2	0
10	34.4	34.3	+0.1
11	29.8	29.7	+0.1
12	20.8	20.7	+0.1
13	20.4	20.3	+0.1
14	17.0	16.9	+0.1
15	16.1	16.0	+0.1
16	156.4	155.6	-0.2

Comparison of <sup>1</sup>H NMR data of synthetic and isolated boneratamide A methyl ester



methyl ester of boneratamide A (1b)			
Position	<sup>1</sup> H NMR (δ) Synthetic Sample (400 MHz, C <sub>6</sub> D <sub>6</sub> )	<sup>1</sup> H NMR (δ) Natural Sample <sup>8</sup> (500 MHz, C <sub>6</sub> D <sub>6</sub> )	
1	4.66 (dd, <i>J</i> = 10.5, 2.5)	4.65 (dd, <i>J</i> = 10.4, 2.3)	
2	***	1.42–1.50 (m)	
3a	**	1.68 (m)	
3b	1.88–1.95 (m)	1.91 (dm, $J = 12.1$ )	
4a	****	1.28 (m)	
4b	**	1.70 (m)	
5	**	1.81 (m)	
6			
7a	*	2.07 (ddd, <i>J</i> = 13.7, 9.3, 4.3)	
7b	2.66 (ddd, <i>J</i> = 13.5, 9.5, 5.5)	2.65 (ddd, <i>J</i> = 13.7, 9.8, 5.9)	
8a	*	2.14 (m)	
8b	2.37–2.47 (m)	2.42 (m)	
10	5.41 (q, J = 1.0)	5.41 (brs)	
11	***	1.42–1.50	
12	1.17 (d, <i>J</i> = 6.5)	1.17 (d, <i>J</i> = 6.4)	
13	0.98 (d, $J = 6.5$ )	0.98 (d, <i>J</i> = 6.4)	
14	0.98 (d, J = 6.5)	0,97 (d, <i>J</i> = 6.5)	
15	1.62 (d, $J = 1.0$ )	1.62 (brs)	
16 NH	7.51 (d, <i>J</i> = 10.5)	7.49 (d, <i>J</i> = 10.4)	
18			
21a	1.79 (ddd, <i>J</i> = 16.5, 9.5, 1.5)	1.79 (ddd, <i>J</i> = 16.4, 9.5, 1.9)	
21b	*	2.14 (m)	
22a	****	1.30 (m)	
22b	***	1.42–1.50	
23	3.76 (dd, <i>J</i> = 10.0, 1.5)	$3.78 (\mathrm{dd}, J = 9.8, 1.5)$	
25	1.66 (s)	1.66 (s)	
26	1.24 (s)	1.25 (s)	
OMe	3.16 (s)	3.17 (s)	

\*2.04–2.21(m), 3H; \*\*1.68–1.86 (m), 3H; \*\*\*1.39–1.51 (m), 3H; \*\*\*\*1.25–1.36 (m), 2H.

Comparison of <sup>13</sup>C NMR data of synthetic and isolated boneratamide A methyl ester



$\frac{13 \text{ C NMR } (\delta)}{13 \text{ C NMR } (\delta)}$				
Position	Synthetic Sample	Natural Sample <sup>8</sup>	$\Delta\delta$ (nnm)	
1	55.4	55.3	+0.1	
2	45.5	45.6	-0.1	
3	25.6	25.7	-0.1	
4	32.6	32.7	-0.1	
5	35.6	35.6	0	
6	59.1	59.2	-0.1	
7	33.9	33.9	0	
8	36.6	36.6	0	
9	143.6	143.6	0	
10	126.4	126.4	0	
11	30.3	30.3	0	
12	21.7	21.7	0	
13	21.6	21.6	0	
14	17.1	17.1	0	
15	17.1	17.2	-0.1	
16	-	_		
17	172.8	172.7	+0.1	
18	60.9	60.9	0	
19	_	_		
20	174.1	174.0	+0.1	
21	30.0	30.0	0	
22	24.5	24.5	0	
23	57.7	57.8	-0.1	
24	175.7	175.5	+0.2	
25	23.7	23.8	-0.1	
26	25.6	25.6	0	
OMe	52.3	52.2	+0.1	



methyl ester of boneratamide B (2b) <sup>1</sup>H NMR ( $\delta$ ) <sup>1</sup>H NMR ( $\delta$ ) Synthetic Sample Natural Sample<sup>8</sup> Position (400 MHz, C<sub>6</sub>D<sub>6</sub>) (500 MHz, C<sub>6</sub>D<sub>6</sub>) 1 4.48 (dd, *J* = 10.5, 2.5) 4.48 (dd, J = 10.8, 2.6)2 1.27-1.32 (m) 1.28 (m) \*\*\*\* 1.14 (m) 3a 3b \*\*\* 1.73 (m) \*\*\*\* 4a 1.12 (m) \*\*\*\* 4b 1.49 (m) \*\*\* 5 1.61 (m) 6 7a \*\*\* 1.71 (m) \*\* 7b 2.12 (m) 8a \*\* 2.11 (m) \* 8b 2.28 (m) 10 5.30 (q, J = 1.5)5.30 (brs) \*\*\*\* 11 1.47 (m) 1.11 (d, J = 7.0) 12 1.11 (d, J = 6.6)13 0.90 (d, J = 6.5)0.90 (d, J = 6.7)14 0.86 (d, J = 6.5)0.86 (d, J = 6.6)15 1.61 (d, J = 1.0)1.61 (brs) 16 NH 6.92 (d, *J* 10.5) 6.91 (d, J = 10.8)18 4.46 (q, J = 7.0)4.46 (q, J = 7.2)1.87 (ddd, J = 17.0, 10.0, 2.5)1.87 (ddd, J = 16.6, 9.6, 2.4)21a \* 21b 2.26 (m) \*\*\*\* 22a 1.42 (m) \*\*\* 22b 1.67 (m) 23 4.11 (dd, J = 9.5, 2.0) 4.11 (dd, *J* = 9.5, 1.8) 25 1.37 (d, J = 7.0)1.37 (d, J = 7.2)OMe 3.22 (s) 3.22 (s)

\*2.21–2.31(m), 2H; \*\*2.08–2.16 (m), 2H; \*\*\*1.62–1.75 (m), 4H; \*\*\*\*1.40–1.53 (m), 3H; \*\*\*\*1.12–1.20 (m), 2H. 31



methyl ester of boneratamide B (2b)

	$^{13}C$ NMR ( $\delta$ )	$^{13}$ C NMR ( $\delta$ )	
Position	(100 MHz, C <sub>6</sub> D <sub>6</sub> )	(125 MHz, C <sub>6</sub> D <sub>6</sub> )	Δδ (ppm)
1	55.0	55.2	-0.2
2	44.8	45.1	-0.3
3	25.3	25.7	-0.4
4	32.0	32.2	-0.2
5	35.2	35.6	-0.4
6	58.7	59.0	-0.3
7	33.4	33.6	-0.2
8	36.0	36.3	-0.3
9	142.8	143.1	-0.3
10	126.0	126.2	-0.2
11	29.7	30.0	-0.3
12	21.4	21.7	-0.3
13	21.0	21.3	-0.3
14	16.5	16.8	-0.3
15	16.7	17.0	-0.3
16	_	_	
17	170.3	170.2	+0.1
18	52.9	53.3	-0.4
19	_	_	
20	175.4	175.6	-0.2
21	29.5	29.7	-0.2
22	24.1	24.3	-0.2
23	58.6	58.8	-0.2
24	173.3	173.4	-0.1
25	14.2	14.1	+0.1
OMe	51.6	51.9	-0.3

Comparison of <sup>1</sup>H NMR data of synthetic and isolated boneratamide C methyl ester



methyl ester of boneratamide C ( <b>3b</b> )			
Position	<sup>1</sup> H NMR (δ) Synthetic Sample (400 MHz, C <sub>6</sub> D <sub>6</sub> )	<sup>1</sup> H NMR (δ) Natural Sample <sup>8</sup> (500 MHz, C <sub>6</sub> D <sub>6</sub> )	
1	4.46 (brd $J = 10.8$ )	4.46 (bd, <i>J</i> = 10.9)	
2	****	1.32 (m)	
3a	****	1.30 (m)	
3b	1.76–1.82 (m)	1.78 (m)	
4a	1.10–1.15 (m)	1.14 (m)	
4b	1.50–1.53 (m)	1.51 (m)	
5	**	1.61 (m)	
6			
7a	1.77 (ddd, <i>J</i> = 13.0, 9.0, 4.0)	1.76 (ddd, <i>J</i> = 13.2, 9.2, 3.9)	
7b	*	2.18 (m)	
8a	*	2.10 (m)	
8b	*	2.31 (m)	
10	5.32 (q, J = 1.5)	5.32 (br)	
11	**	1.61 (m)	
12	1.17 (d, $J = 6.5$ )	1.17 (d, J=6.4)	
13	0.98 (d, $J = 6.5$ )	0.98 (d, <i>J</i> = 6.7)	
14	0.84 (d, J = 6.5)	0.84 (d, <i>J</i> = 6.6)	
15	1.59 (d, <i>J</i> = 1.0)	1.59 (brs)	
16 NH	7.21 (d, J = 11.0)	7.20 (d, <i>J</i> = 10.9)	
18	4.53 (q, <i>J</i> = 7.0)	4.52 (q, <i>J</i> = 7.3)	
21a	1.84 (ddd, <i>J</i> = 17.0, 9.5, 3.5)	1.84 (ddd, $J = 16.8, 9.6, 3.3$ )	
21b	*	2.27 (m)	
22a	***	1.42 (m)	
22b	***	1.45 (m)	
23	3.86 (dd, J=9.0, 3.0)	3.86 (dd, <i>J</i> = 8.8, 2.8)	
25	1.34 (d, J = 7.0)	1.34 (d, <i>J</i> = 7.2)	
OMe	3.34 (s)	3.34 (s)	

 $*2.03-2.34\ (m),\ 4H;\ **1.60-1.64\ (m),\ 2H;\ ***1.35-1.46\ (m),\ 2H;\ ****1.27-1.33\ (m),\ 2H.$ 

Comparison of <sup>13</sup>C NMR data of synthetic and isolated boneratamide C methyl ester



methyl ester of boneratamide C (3b)

	<sup>13</sup> C NMR (δ) Synthetic Sample	$^{13}$ C NMR ( $\delta$ ) Natural Sample <sup>8</sup>	
Position	(100 MHz, C <sub>6</sub> D <sub>6</sub> )	(125 MHz, C <sub>6</sub> D <sub>6</sub> )	Δδ (ppm)
1	55.3	55.4	-0.1
2	45.2	45.3	-0.1
3	25.5	25.5	0
4	32.3	32.3	0
5	35.4	35.5	-0.1
6	59.2	58.9	+0.3
7	33.7	33.7	0
8	36.2	36.3	-0.1
9	143.1	143.5	-0.4
10	126.3	126.5	-0.2
11	29.4	29.4	0
12	22.3	22.3	0
13	21.4	21.4	0
14	16.8	17.1	-0.3
15	17.0	17.2	-0.2
16	_	-	
17	170.0	170.0	0
18	53.3	53.5	-0.2
19	_	-	
20	175.8	175.9	-0.1
21	29.8	29.9	-0.1
22	23.8	23.7	+0.1
23	58.0	58.1	-0.1
24	172.3	172.4	-0.1
25	14.7	14.4	+0.3
OMe	51.9	52.0	-0.1










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#### X-ray crystallographic structure of 1b





CCDC 2144949 Empirical Formula Formula Weight Crystal Color, Habit Crystal Dimensions Crystal System Lattice Type Lattice Parameters

Space Group Z value

D<sub>calc</sub>

F<sub>000</sub> μ(CuKα) C<sub>25</sub>H<sub>40</sub>N<sub>2</sub>O<sub>4</sub> 432.60 colorless, block 0.700 X 0.350 X 0.300 mm orthorhombic Primitive a = 7.9942(2) Åb = 10.5274(3) Åc = 29.6819(8) Å $V = 2497.98(12) \text{ Å}^3$ P2<sub>1</sub>2<sub>1</sub>2<sub>1</sub> (#19) 4 1.150 g/cm<sup>3</sup> 944.00

6.154 cm<sup>-1</sup>

## B. Intensity Measurements

Diffractometer	R-AXIS RAPID
Radiation	CuK $\alpha$ ( $\lambda$ = 1.54187 Å) graphite monochromated
Voltage, Current	50kV, 100mA
Temperature	-180.0 <sup>0</sup> C
Detector Aperture	460.0 x 256.0 mm
Data Images	450 exposures
ω oscillation Range (χ=54.0, φ=0.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>
ω oscillation Range (χ=54.0, φ=90.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>
ω oscillation Range (χ=54.0, φ=180.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>
ω oscillation Range (χ=54.0, φ=270.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>
ω oscillation Range (χ=10.0, φ=60.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>

Detector Position	127.40 mm
Pixel Size	0.100 mm
2θ <sub>max</sub>	136.4 <sup>0</sup>
No. of Reflections Measured	Total: 28338 Unique: 4564 (R <sub>int</sub> = 0.0711) Parsons quotients (Flack x
parameter). 1004	
Corrections	Lorentz-polarization Absorption (trans. factors: 0.710 - 0.831)

#### C. Structure Solution and Refinement

Structure Solution Version 2014/4)	Direct Methods (SHELXT
Refinement	Full-matrix least-squares on F <sup>2</sup>
Function Minimized	$\Sigma \text{ w } (\text{Fo}^2 - \text{Fc}^2)^2$
Least Squares Weights	w = 1/ [ $\sigma^2(Fo^2)$ + (0.0758 · P) <sup>2</sup> + 1.0018 · P]
2Fc <sup>2</sup> )/3	where P = (Max(Fo <sup>2</sup> ,0) +
$2\theta_{max}$ cutoff	136.4 <sup>0</sup>
Anomalous Dispersion	All non-hydrogen atoms
No. Observations (All reflections)	4564
No. Variables	280
Reflection/Parameter Ratio	16.30
Residuals: R1 (I>2.00 $\sigma$ (I))	0.0468
Residuals: R (All reflections)	0.0479
Residuals: wR2 (All reflections)	0.1305
Goodness of Fit Indicator	1.051
Flack parameter (Parsons' quotients = 1834)	0.09(7)

Max Shift/Error in Final Cycle	0.000
Maximum peak in Final Diff. Map	0.62 e <sup>-</sup> /Å <sup>3</sup>
Minimum peak in Final Diff. Map	-0.40 e <sup>-</sup> /Å <sup>3</sup>

#### X-ray crystallographic structure of 3b





CCDC 2144950 Empirical Formula Formula Weight Crystal Color, Habit Crystal Dimensions Crystal System Lattice Type Lattice Parameters

Space Group Z value

D<sub>calc</sub>

F<sub>000</sub> μ(CuKα) C<sub>27</sub>H<sub>41</sub>N<sub>2</sub>O<sub>4</sub> 457.63 colorless, block 0.600 X 0.500 X 0.300 mm monoclinic C-centered a = 17.0655(8) Å b = 10.6435(5) Å c = 16.6001(8) Å $\beta = 116.597(8)$  <sup>O</sup>  $V = 2696.1(3) Å^3$ C2 (#5) 4 1.127 g/cm<sup>3</sup> 996.00 5.969 cm<sup>-1</sup>

## B. Intensity Measurements

Diffractometer	R-AXIS RAPID
Radiation	CuK $\alpha$ ( $\lambda$ = 1.54187 Å) graphite monochromated
Voltage, Current	50kV, 100mA
Temperature	-100.0 <sup>0</sup> C
Detector Aperture	460.0 x 256.0 mm
Data Images	180 exposures
ω oscillation Range (χ=54.0, φ=0.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>
ω oscillation Range (χ=54.0, φ=90.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>
ω oscillation Range (χ=54.0, φ=180.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>
ω oscillation Range (χ=54.0, φ=270.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>
ω oscillation Range (χ=0.0, φ=0.0)	80.0 - 260.0 <sup>0</sup>
Exposure Rate	10.0 sec./ <sup>0</sup>

Detector Position	127.40 mm
Pixel Size	0.100 mm
20 <sub>max</sub>	136.5 <sup>0</sup>
No. of Reflections Measured	Total: 15545 Unique: 4797 (R <sub>int</sub> = 0.0333) Parsons quotients (Flack x
parameter): 2053	
Corrections	Lorentz-polarization Absorption (trans. factors: 0.671 - 0.836)

#### C. Structure Solution and Refinement

Structure Solution Version 2014/4)	Direct Methods (SHELXT
Refinement	Full-matrix least-squares on F <sup>2</sup>
Function Minimized	$\Sigma \text{ w } (\text{Fo}^2 - \text{Fc}^2)^2$
Least Squares Weights	w = 1/ [ $\sigma^2(Fo^2)$ + (0.1032 · P) <sup>2</sup> + 1.5528 · P]
2Fc <sup>2</sup> )/3	where P = (Max(Fo <sup>2</sup> ,0) +
2θ <sub>max</sub> cutoff	136.5 <sup>0</sup>
Anomalous Dispersion	All non-hydrogen atoms
No. Observations (All reflections)	4797
No. Variables	299
Reflection/Parameter Ratio	16.04
Residuals: R1 (I>2.00 $\sigma$ (I))	0.0543
Residuals: R (All reflections)	0.0559
Residuals: wR2 (All reflections)	0.1571
Goodness of Fit Indicator	1.064
Flack parameter (Parsons' quotients = 2053)	0.11(5)

Max Shift/Error in Final Cycle	0.000
Maximum peak in Final Diff. Map	0.63 e⁻/Å <sup>3</sup>
Minimum peak in Final Diff. Map	-0.39 e <sup>-</sup> /Å <sup>3</sup>

# X-ray crystallographic structure of 28



CCDC 2183045	
Empirical formula	$C_{24}H_{40}N_2O_5$
Formula weight	436.58
Temperature/K	100.01(10)
Crystal system	monoclinic
Space group	P21
a/Å	13.2329(2)
b/Å	6.99870(10)
c/Å	14.9696(3)
a/°	90
β/°	115.326(2)
γ/°	90
Volume/ų	1253.13(4)
Z	2
$ ho_{calc}g/cm^3$	1.157
µ/mm <sup>-1</sup>	0.646
F(000)	476.0
Crystal size/mm <sup>3</sup>	0.361 × 0.137 × 0.047
Radiation	Cu Kα (λ = 1.54184)
20 range for data collection/°	6.532 to 149.21
Index ranges	-15 ≤ h ≤ 16, -7 ≤ k ≤ 8, -18 ≤ l ≤ 18
Reflections collected	15311
Independent reflections	4609 [ $R_{int} = 0.0346$ , $R_{sigma} = 0.0347$ ]
Data/restraints/parameters	4609/1/290

Goodness-of-fit on  $F^2$ Final R indexes [I>= $2\sigma$  (I)] Final R indexes [all data] Largest diff. peak/hole / e Å<sup>-3</sup> Flack parameter 1.041  $R_1 = 0.0315$ ,  $wR_2 = 0.0815$   $R_1 = 0.0332$ ,  $wR_2 = 0.0826$ 0.24/-0.20 -0.08(8)

### Catalyst Screening for Cyclopropanation<sup>a</sup>



Entry	Catalyst (mol%)	Solvent	Temperature	Time	Yield (%) <sup>b</sup>
1	copper (II) catalyst 1 (8)	toluene	80 °C	15 h	46
2	copper ( $\rm I\!I$ ) catalyst 1 (8) - BF <sub>3</sub> $\cdot$ OEt <sub>2</sub> (1)	toluene	r.t. to 80 $^\circ \mathrm{C}$	4 h	20
3	$Cu(acac)_2(2)$	toluene	80 °C	6 h	33
4 <sup>9</sup>	$Cu(acac)_2(1) - BF_3 \cdot OEt_2(1)$	$\mathrm{CH}_2\mathrm{Cl}_2$	r.t.	6 h	49
5 <sup>9</sup>	$CuOTf(C_6H_6)_{0.5}$ (2)	$CH_2Cl_2$	r.t. to 40 $^\circ \rm C$	6 h	no reaction
6	$CuOTf(C_6H_6)_{0.5}$ (5)	toluene	80 °C	18 h	18
7	CuOAc (2) - $BF_3 \cdot OEt_2(1)$	$CH_2Cl_2$	r.t.	68 h	<5
8	$Cu(OTf)_2(2)$	$CH_2Cl_2$	r.t.	68 h	<5
9	$Cu(OTf)_2(2) - BF_3 \cdot OEt_2(1)$	$CH_2Cl_2$	r.t.	48 h	<5
10	$Cu(TFA)_2(2)$	toluene	80 °C	48 h	8
11	$Cu(TFA)_2$ (2) - BF <sub>3</sub> ·OEt <sub>2</sub> (1)	$\mathrm{CH}_2\mathrm{Cl}_2$	r.t.	24 h	22
12	Cu powder (60)	toluene	80 °C	3 h	<5
13	CuCl (100)	toluene	80 °C	3 h	45
14	CuBr (100)	toluene	80 °C	18 h	33
15	CuI (100)	toluene	80 °C	20 h	25
$16^{10}$	$[RuCl_2(C_6H_6)]_2$ (1)	$CH_2Cl_2$	r.t. to 40 $^\circ \rm C$	3 d	no reaction
$17^{11}$	ruthenium (II) catalyst $2(1)$	$CH_2Cl_2$	r.t. to 40 $^\circ \rm C$	3 d	<5
18	Rh <sub>2</sub> (OAc) <sub>4</sub> (5)	$CH_2Cl_2$	r.t.	5 min	42
19	$Rh_2(OAc)_4(5)$	$ClCH_2CH_2Cl$	r.t.	5 min	38
20	$Rh_2(OAc)_4(1)$	toluene	r.t.	30 min	46
21	$Rh_2(Opiv)_2$ (1)	$\mathrm{CH}_2\mathrm{Cl}_2$	r.t.	5 min	0
22	$Rh_2(Opiv)_2$ (1)	toluene	-40 °C	30 min	0
23	$Pd(OAc)_2$ (2)	$CH_2Cl_2$	r.t.	39 h	no reaction
24 <sup>12</sup>	$Pd(OAc)_2$ (5)	toluene	80 °C	21 h	15
25 <sup>12</sup>	$Pd(PPh_3)_4$ (5)	toluene	80 °C	21 h	13
26	PdCl <sub>2</sub> (10)	toluene	80 °C	24 h	27

<sup>a</sup>All reactions were performed on 10 mg scale and run under nitrogen atmosphere. <sup>b</sup>Isolated yields.



#### A Plausible reaction mechanism for formation of boneratamide A



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