SUPPORTING INFORMATION

Efficient Synthesis of Pentasubstituted Pyrroles via Intramolecular C-arylation

Barbora Lemrová¹, Michal Maloň², Miroslav Soural^{1,3}

¹ Department of Organic Chemistry, Faculty of Science, Palacký University, 17. Listopadu 12, 77146 Olomouc, Czech Republic

² JEOL (U.K.) Ltd., JEOL House, Silver Court, Watchmead, Welwyn Garden City, Hertfordshire AL7 1LT, United Kingdom

³ Institute of Molecular and Translational Medicine, Faculty of Medicine and Dentistry, Palacký University, Hněvotínská 5, 77900 Olomouc, Czech Republic

Table of Contents

General information	S-2
Experimental procedures	S-4
NMR spectra of the final compounds	S-13

General information

Solvents and chemicals were purchased from Sigma-Aldrich (Milwaukee, USA, www.sigmaaldrich.com), Acros Organic (Geel, Belgium, www.acros.com) and Fluorochem (Derbyshire, UK, www.fluorochem.co.uk). Rink Amide resin (100-200 mesh, 1% DVB, 0.6 mmol/g) and Wang resin (100-200 mesh, 1% DVB, 1.4 mmol/g) were obtained from AAPPTec (Louisville, USA, www.aapptec.com). Solid-phase synthesis was carried out in plastic reaction vessels (syringes, each equipped with a porous disk) using a manually operated synthesizer (Torviq, Niles, USA, www.torviq.com). Dry solvents were dried over 4Å molecular sieves or stored as received from commercial suppliers.

All reactions were carried out at ambient temperature (23 °C) unless stated otherwise. For the LC/MS analyses, a sample of resin (~5 mg) was treated with 50% TFA in DCM, the cleavage cocktail was evaporated under a stream of nitrogen, the cleaved compounds dissolved in CH_3CN / H_2O (20% or 50%; 1 mL) and the resin was removed by filtration.

LC/MS analyses were carried out using UPLC-MS system consisting of UPLC chromatograph Acquity with photodiode array detector and single quadrupole mass spectrometer (Waters), using C18 X-Select HSS T3 column (2.5 μ m, 3.0 mm x 50 mm) at 30 °C and flow rate of 0.6 mL/min. Mobile phase was (A) 0.01M ammonium acetate (AmAc) in H₂O, and (B) CH₃CN, linearly programmed gradient elution. The ESI source operated at capillary voltage of 3 kV, desolvation temperature of 350 °C and source temperature of 120 °C.

Purification was carried out using semipreparative HPLC chromatograph with MS detector (YMC-Actus Pro C18 column, 20 x 100 mm, 5 μ m, MP 0.01M aqueous AmAc/ CH₃CN, flow rate 15 mL/min, gradient elution) or by normal phase (silica gel chromatography). Residual solvents (H₂O and AmAc buffer) were lyophilized by the ScanVac Coolsafe 110-4 operating at -110 °C.

HRMS analysis was performed using LC-MS (Dionex Ultimate 3000, Thermo Fischer Scientific, USA) with Exactive Plus Orbitrap high-resolution mass spectrometer (Thermo Exactive plus, Thermo Fischer Scientific, USA) operating at positive or negative full scan mode (120 000 FWMH) in the range of 100-1000 *m/z* with electrospray ionization operating at 150 °C and the source voltage of 3.6 kV. Chromatographic separation was performed on Phenomenex Gemini column (C18, 50 x 2 mm, 3 μ m particle size) with isocratic elution and mobile phase (MP) containing MeOH/H₂O/formic acid 95:5:0.1. The samples were dissolved in CH₃CN or MeOH/H₂O (95:5 v/v).

NMR experiments were performed with the use of ECX500 spectrometer (JEOL RESONANCE, Tokyo, Japan) at magnetic field strength of 11.75 T corresponding to ¹H, ¹³C and ¹⁵N resonance

S-2

frequencies of 500.16 MHz, 125.77 MHz and 50.7 MHz at 25 °C. The assignment of ¹H, ¹³C and ¹⁵N signals was done by the combination of ¹H, ¹³C{¹H}, ¹H-¹H COSY, ¹H-¹H NOESY, ¹H-¹³C HMQC, ¹H-¹³C HMQC, ¹H-¹³C HMBC, ¹H-¹⁵N HMQC and ¹H-¹⁵N HMBC experiments. In case of **14** and **15**, the proton measurements were performed at 25°C, 35°C, 45°C, 55°C, 85°C and 115°C. Chemical shifts (δ) are reported in parts per million (ppm) and coupling constants (*J*) are reported in Hertz (Hz). The signals of DMSO-*d*₆ were set at 2.52 ppm in ¹H NMR spectra and at 39.52 ppm in ¹³C NMR spectra. ¹⁵N chemical shifts were referenced to external 90% formamide in DMSO-*d*₆ (112.00 ppm).¹ Abbreviations in NMR spectra: br. s – broad singlet, s – singlet, d – doublet, dd – doublet of doublets, ddd – doublet of doublets of doublets, dt – doublet of triplets, t - triplet, m – multiplet.

<u>General Method for Calculation of Yields Using ¹H NMR</u>: ¹H NMR spectra of external standard in DMSO- d_6 at three different concentrations were measured. In each spectrum solvent signal was integrated followed by the integration of selected H_{Ar} signals of external standard. Ratios of solvent/standard signal areas along with a known quantity of the standard were used to construct a calibration curve. Then ¹H NMR spectra of the studied sample were measured, and the ratio of solvent/sample (selected H_{Ar} signal) areas was determined. Using the calibration curve, the quantity of compound in the sample was calculated.

<u>Quantification of the resin loading</u>: Quantification of loading of Rink or Wang resin with immobilized Fmoc-Asp(OMe)-OH, Fmoc-Ala-OH, Fmoc-Pro-OH, Fmoc-Sar-OH or 2-(Fmoc-amino)ethanol: After the immobilization, the sample of resin **2**, **2e**, **1b**, **1c**, **1d**, **1f** (~30 mg) was washed with DCM (3x), with MeOH (5x), dried and divided into two samples (2 × 10 mg). Both samples were cleaved from the resin using TFA in DCM (0.5 mL, 50%) for 2 h at ambient temperature. The cleavage cocktail was evaporated by a stream of nitrogen and oily residue was extracted into 1 mL of MeOH and analyzed by UPLC-UV-MS. Loading of resin was calculated with the use of an external standard (Fmoc-Ala-OH, 0.5 mg/mL).

Quantification of resin with immobilized 1,3-diaminopropane: After the immobilization, the resin **1g** (~30 mg) was reacted with Fmoc-OSu (65 mg, 0.2 mmol) in DCM (0.5 mL) for 30 min at room temperature and subsequently washed with DCM (3x), with MeOH (5x), dried and divided into two samples (2×10 mg). The quantification was carried out according to the procedure described in the previous paragraph.

Experimental procedures

Immobilization of Fmoc-Asp(OMe)-OH on Wang resin and Rink amide resin (2a, 2e)

Rink amide resin (1 g, 0.6 mmol/g) was washed with DCM (3x) and with DMF (3x) and then subjected to Fmoc-deprotection using 50% solution of piperidine in DMF (10 mL) at room temperature. After shaking for 20 min, the resin was washed with DMF (5x). Wang resin (1 g, 1.4 mmol/g) was washed with DCM (3x) and DMF (3x). A solution of Fmoc-Asp(OMe)-OH (739 mg; 2 mmol), HOBt (306 mg, 2.0 mmol), DMAP (61 mg, 0.5 mmol, used only for Wang resin), and DIC (312 μ L, 2.0 mmol) in DMF/DCM (10 mL, 50%) was added to the swollen Wang resin and deprotected Rink amide resin, and the reaction mixtures were shaken for 24 h at ambient temperature. Both resins were washed with DMF (3x) and with DCM (3x). The quantification was carried out as described in the paragraph "Quantification of the resin loading". Calculated loadings of immobilized amino acids: 0.6 mmol/g for Wang resin and 0.3 mmol/g for Rink amide resin.

Immobilization of Fmoc-Sar-OH, Fmoc-Ala-OH and Fmo-Pro-OH on Rink amide resin (1b, 1c, 1d)

Rink amide resin (1 g, 0.6 mmol/g) was washed with DCM (3x) and with DMF (3x) and then subjected to Fmoc-deprotection using 50% solution of piperidine in DMF (10 mL) at room temperature. After shaking for 20 min, the resin was washed with DMF (5x). A solution of Fmoc-AA-OH (2 mmol), HOBt (306 mg, 2.0 mmol) and DIC (312 μ L, 2.0 mmol) in DMF/DCM (10 mL, 50%) was added and the resin slurry was shaken for 24 h at ambient temperature. After the reaction, the resin was washed with DMF (3x) and with DCM (3x) and quantified as described above. Calculated loadings of immobilized amino acids were 0.3 mmol/g.

Immobilization of 2-(Fmoc-amino)ethanol to Wang resin (1f)

Wang resin (1g; 1.4 mmol/g) was washed with anhydrous DCM (3x) and a solution of trichloroacetonitrile (15 mmol; 1.5 mL) in 10 mL anhydrous DCM was added. The resin was kept in a freezer for 30 min. Then solution of DBU (0.67 mmol; 100 μ L) in 2 mL anhydrous DCM was added, the resin slurry was shaken for 1 h and subsequently washed with anhydrous DCM (3x) and with anhydrous THF (3x). Solution of 2-(Fmoc-amino)ethanol (3 mmol; 849 mg) in 10 mL anhydrous THF was added to the resin followed by dropwise addition of BF₃·Et₂O (0.3 mmol; 63 μ L) and the resin slurry was shaken for 30 min. Resin was washed with THF (3x), MeOH (3x) and with DCM (3x). The quantification was carried out as described in the paragraph "Quantification of the resin loading". Calculated loading was 0.33 mmol/g.

Immobilization of 1,3-diaminopropane (1g)

Wang resin (1 g, 1.4 mmol/g) was washed with DCM (3x). A solution of CDI (810 mg, 5.0 mmol) and pyridine (400 μ L, 5.0 mmol) in 10 mL DCM was added and the resin slurry was shaken for 2 h at ambient temperature. The resin was then washed with DCM (3x) and a solution of 1,3-diaminopropane (421 μ L, 5.0 mmol) in 10 mL DCM was added. The resin was shaken for 16 h and washed with DCM (5x). Sample of resulting resin was derivatized with FmocOSu according to procedure described in the paragraph "Quantification of the resin loading". Calculated loading was 0.6 mmol/g.

Acylation with Fmoc-Asp(OMe)-OH (2b-d, 2f-g)

Resin **1b**, **1c**, **1d** and **1f** were washed with DCM (3x) and with DMF (3x) and then subjected to Fmocdeprotection using 20% solution of piperidine in DMF (10 mL) at room temperature. After shaking for 20 min, the resins were washed with DMF (5x). Subsequently, resins **1b**, **1c**, **1d** and **1f**, **1g** were shaken with solution of Fmoc-Asp(OMe)-OH (739 mg; 2 mmol), HOBt (306 mg, 2.0 mmol), and DIC (312 μ L, 2.0 mmol) in DMF/DCM (10 mL, 50%) for 16 h at ambient temperature. All resins were washed with DMF (3x) and with DCM (3x) and analyzed by the standard procedure.

Nosylation with 4-nitrobenzenesulfonyl chloride (3)

Resins **2** (1 g) were treated with 20% piperidine in DMF (10 ml) for 20 min and subsequently washed with DMF (3x) and DCM (5x). A solution of 4-NsCl (640 mg; 3 mmol) and 2,6-lutidine (380 μ L, 3 mmol) in DCM (10 mL) was added to the resin and the resin slurry was shaken for 16 h. Resins were then washed with DCM (5x) and analyzed by the standard procedure.

Alkylation with α -bromoketones (4)

Resins **3** (300 mg) were washed with DMF (3x) and a solution of α -bromoketone (0.6 mmol) and DIPEA (209 μ L, 1.2 mmol) in DMF (3 mL) was added. The reaction mixtures were shaken for 16 h at ambient temperature. The resin was washed with DMF (3x) and with DCM (3x) and analyzed by the standard procedure.

TMSOK-based cyclization and cleavage from the resin (6-9; 14-16)

Resins **4** (300 mg) were washed with DCM (3x), DMF (3x) and a 0.3 M solution of TMSOK (116 mg; 0.9 mmol) in 3 mL DMF was added. The reaction mixtures were shaken at room temperature for 30 min. After the reaction, the resins were washed with DMF (3x) and DCM (3x). Subsequently, the resins were treated with 50% TFA in DCM for 1 h. The cleavage cocktails were separated and each resin was washed with 50% TFA in DCM (2x). The washes were collected and evaporated by a stream of nitrogen. The crude compounds **6**, **7** and **9** were purified using column chromatography in EtOAc as

mobile phase. In cases of **8**, **14**- **16**, the crude compounds were dissolved in CH₃CN and purified by semipreparative HPLC.

Alkylation with 1-chloropropan-2-one with thermal pyrrole cyclization (10-13)

Resins **3a**, **3c**, **3f** and **3g** (500 mg) were washed with DCM (3x) and DMSO (3x). Then, the swollen resins were shaken with solution of 1-chloropropan-2-one (207 μ L; 2.5 mmol), DIPEA (875 μ L; 5 mmol) in 5 mL DMSO for 16 h at 80°C. The resins were washed with DMSO (3x) and DCM (3x) and treated with 50% TFA in DCM for 1 h. The cleavage cocktails were separated and each resin was washed with 50% TFA in DCM (2x). The washes were collected and evaporated by a stream of nitrogen. The crude compounds were dissolved in CH₃CN and purified by semipreparative HPLC.

Pyrrole on-resin alkylation (19, 20)

Resin **5** (500 mg) was washed with DCM (3x) and DMF (3x). For methyl derivative: 5 mL of 10% solution of methyl iodide in DMF and TMSOK (320 mg; 2.5 mmol) was added to the resin and the resin slurry was shaken at room temperature for 16 h. For benzyl derivative: 5 mL of 10% solution of benzyl bromide in *N*-methyl pyrrolidone (NMP) and TMSOK (320 mg; 2.5 mmol) was added and the resin slurry was shaken at 110°C for 16 h. After the reaction, the resin was washed with DMF (3x), DCM (3x). In both cases, the alkylated products were cleaved from resin and purified by semipreparative HPLC.

Amide post-cleavage formation (23, 24)

Resin **5** (300 mg) was washed with DCM (3x) and shaken with cleavage cocktail (3 mL) at room temperature for 1 h. The cleavage cocktail was then collected, and the combined washes were evaporated using a stream of nitrogen. The oil residue was dissolved in 2 mL of DCM and evaporated again using a stream of nitrogen. Resulting crude compound **6** was dissolved in 1 mL of propan-1-amine or 3-aminopropan-1-ol and heated at 80°C for 24 h (for 3-aminopropan-1-ol) or 48 h (for propan-1-amine). In the latter, additional 0.5 mL of propan-1-amine was added after 24 h. The reaction mixture was diluted with 3 ml DCM and 3 mL water. Conc. HCl was added until the acidic *p*H of the aqueous layer was obtained. The DCM layer was separated, the water layer was extracted 2x with 2 mL of DCM, combined DCM layers were extracted with 2 mL of 10% solution of HCl, then with brine and dried with magnesium sulfate. Resulting solution was adsorbed to silica gel and purified by column chromatography using mobile phase DCM/MeOH (9/1).

Reduction with sodium dithionite (22)

Resin **5** (300 mg) was washed with DCM (3x) and 4 mL of DCM was added. Then a solution of sodium dithionite (840 mg; 4 mmol), potassium carbonate (760 mg; 5.6 mmol) and tetrabutylammonium hydrogen sulfate (TBAHS) (136 mg; 0.4 mmol) in 8 mL of water was prepared and immediately added to the reaction mixture. The resin slurry was shaken for 16 h, then washed with mixture water/DCM (1/1) (3x), DMF (3x) and with DCM (3x). The final compound was cleaved from resin and purified by semipreparative HPLC. The NMR measurement revealed the presence of TBAHS and for this reason, the final compound **22** was re-purified by using column chromatography in EtOAc as mobile phase.

Preparation of indazole-oxide (27)

Resin **2** (0.5 g) was treated with 20% piperidine in DMF (5 ml) for 20 min and subsequently washed with DMF (3x) and DCM (5x). A solution of 2,4-NsCl (400 mg; 1.5 mmol) and 2,6-lutidine (190 μ L, 1,5 mmol) in DCM (10 mL) was added and the reaction slurry was shaken for 16 h. Resin was washed with DCM (5x). DMF (3x) and a solution of 2-bromoacetophenone (210 μ L; 1 mmol) and DIPEA (348 μ L, 2 mmol) in DMF (5 mL) was added. The resin was shaken for 16 h, washed with DMF (3x) and with DCM (3x). The product was cleaved from the resin and purified by using semipreparative HPLC as described above.

1. G. E. Martin and C. E. Hadden, J. Nat. Prod., 2000, 63, 543–585.

Methyl 2-carbamoyl-5-(4-nitrophenyl)-4-(p-tolyl)-1H-pyrrole-3-carboxylate (6)



Yellow solid 22 mg (77% yield). ¹H-NMR (500 MHz, DMSO-d₆) δ 12.52 (s, 1H; N¹-H), 8.45 (bs, 1H; N⁷-H^α), 8.09 – 8.11 (m, 2H; C¹⁹-H, C²¹-H), 7.74 (bs, 1H; N⁷-H^β), 7.48-7.45 (m, 2H; C¹⁸-H), 7.16 (d, *J* = 8.0 Hz, 2H; C¹²-H, C¹⁴-H), 7.04 (d, *J* = 8.0 Hz, 2H; C¹¹-H, C¹⁵-H), 3.53 (s, 3H; C⁹-H₃), 2.34 (s, 3H; C¹⁶-H). ¹³C-NMR (126 MHz, DMSO-d₆) δ 166.2 (C⁸), 160.7 (C⁶), 145.9 (C²⁰), 137.5 (C¹⁷), 136.1 (C¹³), 131.2 (C¹⁰), 129.9 (C¹¹, C¹⁵), 128.9 (C¹⁸, C²²), 128.8 (C¹², C¹⁴), 128.6 (C⁵), 125.7 (C⁴), 123.2 (C¹⁹, C²¹), 115.8 (C³), 51.6 (C⁹), 20.8 (C¹⁶) ppm. ¹⁵N NMR (51 MHz, DMSO-d₆) δ 369.6 (N²³), 162.8 ('*J* = 96.4 Hz; N¹), 105.0 ('*J* = 90.0 Hz; N⁷) ppm. HRMS (ESI): m/z calcd

 $C_{20}H_{17}N_3O_5$ for $[M+H]^+ = 380.1241$, found $[M+H]^+ = 380.1240$.

Methyl 2-carbamoyl-4-(4-methoxyphenyl)-5-(4-nitrophenyl)-1H-pyrrole-3-carboxylate (7)

Yellow solid 16 mg (54% yield). ¹H-NMR (500 MHz, DMSO-*d*₆) δ 12.48 (s, 1H;N¹-H), 8.45 (s, 1H; N⁷-H^α),



8.11-8.08 (m, 2H; C¹⁹-H, C²¹-H), 7.72 (s, 1H; N⁷-H^β), 7.47-7.44 (m, 2H; C¹⁸-H, C²²-H), 7.07-7.04 (m, 2H; C¹¹-H, C¹⁵-H), 6.91-6.88 (m, 2H; C¹²-H, C¹⁴-H), 3.76 (s, 3H; C¹⁶-H₃), 3.52 (s, 3H; C⁹-H₃) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 166.2 (C⁸), 160.7 (C⁶), 158.3 (C¹³), 145.8 (C²⁰), 137.6 (C¹⁷), 131.2 (C¹¹, C¹⁵), 129.9 (C²), 128.9 (C¹⁸, C²²), 128.6 (C⁵), 126.3 (C¹⁰), 125.5 (C⁴), 123.2 (C¹⁹, C²¹), 115.8 (C³), 113.6 (C¹², C¹⁴), 55.0 (C¹⁶), 51.6 (C⁹) ppm. ¹⁵N NMR (51 MHz, DMSO- d_6) δ 369.6 (N²³), 162.7 ('J = 96.3 Hz; N¹), 105.1 ('J = 90.0 Hz; N⁷) ppm. HRMS (ESI): m/z calcd C₂₀H₁₇N₃O₆ for [M+H]⁺ =

396.1190, found [M+H]⁺ = 396.1191.

Methyl 2-carbamoyl-4-(4-fluorophenyl)-5-(4-nitrophenyl)-1H-pyrrole-3-carboxylate (8)



Yellow solid 14.3 mg (50% yield). ¹H NMR (500 MHz, DMSO- d_6) δ 12.62 (s, 1H), 8.58 (s, 1H), 8.13-8.10 (m, 2H), 7.79 (s, 1H), 7.47-7.44 (m, 2H), 7.22-7.16 (m, 4H), 3.53 (s, 3H) ppm. ¹³C NMR (126 MHz, DMSO- d_6) δ 166.0, 161.3 (d, ¹ J_{C-F} = 243.8 Hz), 160.7, 146.0, 137.2, 132,1 (d, ³ J_{C-F} = 8.2 Hz), 130.8 (d, ⁴ J_{C-F} = 3.3 Hz), 130.3, 129.1, 128.9, 124.8, 123.3, 115.3, 115.0 (d, ² J_{C-F} = 21.4 Hz), 51.6 ppm. HRMS (ESI): m/z calcd C₁₉H₁₄FN₃O₅ for [M+H]⁺ = 384.0989.

Methyl 2-carbamoyl-5-(4-nitrophenyl)-4-(thiophen-3-yl)-1H-pyrrole-3-carboxylate (9)



Yellow solid 10.4 mg (55% yield). ¹H NMR (500 MHz, DMSO- d_6) δ 12.54 (s, 1H), 8.49 (s, 1H), 8.13 – 8.10 (m, 2H), 7.74 (s, 1H), 7.54 (dd, J = 4.9, 2.9 Hz, 1H), 7.50 – 7.47 (m, 2H), 7.21 (dd, J = 2.9, 1.2 Hz, 1H), 6.94 (dd, J = 5.0, 1.3 Hz, 1H), 3.56 (s, 3H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 166.1, 160.6, 146.0, 137.4, 133.8, 130.1, 129.6, 129.1, 128.8, 125.5, 124.1, 123.2, 120.5, 115.8, 51.6 ppm. HRMS (ESI): m/z calcd C₁₇H₁₃N₃O₅S for [M+H]⁺ = 372.0649, found [M+H]⁺ = 372.0648.

Methyl 2-carbamoyl-4-methyl-5-(4-nitrophenyl)-1H-pyrrole-3-carboxylate (10)



Yellow solid 6.7 mg (23% yield). ¹H NMR (500 MHz, DMSO- d_6) δ 12.37 (s, 1H), 8.85 (s, 1H), 8.29 – 8.26 (m, 2H), 7.78 – 7.75 (m, 2H), 7.71 (s, 1H), 3.84 (s, 3H), 2.30 (s, 3H). ¹³C NMR (126 MHz, DMSO- d_6) δ 166.7, 160.7, 146.0, 137.6, 130.8, 129.4, 129.2, 123.4, 121.0, 114.4, 51.8, 12.3. HRMS (ESI): m/z calcd C₁₄H₁₃N₃O₅ for [M+H]⁺ = 304.0928, found [M+H]+ = 304.0934.

Methyl (*S*)-2-((1-amino-1-oxopropan-2-yl)carbamoyl)-4-methyl-5-(4-nitrophenyl)-1*H*-pyrrole-3-carboxylate (11)



Yellow solid 5 mg (9% yield). ¹H-NMR (500 MHz, DMSO- d_6) δ 12.41 (s, 1H), 9.65 (d, J = 7.2 Hz, 1H), 8.29-8.26 (m, 2H), 7.78-7.76 (m, 2H), 7.47 (s, 1H), 7.18-7.00 (m, 2H), 4.48-4.42 (m, 1H), 3.86 (s, 3H), 2.31 (s, 3H), 1.34 (d, J = 7.0 Hz, 3H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 173.8, 166.4, 158.7, 146.0, 137.6, 130.6, 129.4, 129.2, 123.4, 120.9, 114.3, 51.9, 48.6, 18.6, 12.2 ppm. HRMS (ESI): m/z calcd C₁₇H₁₈N₄O₆ for [M+H]⁺ = 375.1299, found [M+H]+ = 375.1302.

Methyl 2-((2-hydroxyethyl)carbamoyl)-4-methyl-5-(4-nitrophenyl)-1H-pyrrole-3-carboxylate (12)



Yellow solid 10.6 mg (20% yield). ¹H-NMR (500 MHz, DMSOd₆) δ 12.40 (s, 1H), 9.53 (t, *J* = 5.4 Hz, 1H), 8.31-8.28 (m, 2H), 7.80-7.77 (m, 2H), 4.79 (t, *J* = 5.0 Hz, 1H), 3.86 (s, 3H), 3.56 (dd, *J* = 10.5, 5.5 Hz, 2H), 3.41 (q, *J* = 5.8 Hz, 2H), 2.33 (s, 3H) ppm. ¹³C-NMR (126 MHz, DMSO-d₆) δ 166.5, 159.4, 145.9, 137.7, 131.1, 129.3, 129.0, 123.4, 120.9, 114.0, 59.7, 51.8, 41.8, 12.3 ppm. HRMS (ESI): m/z calcd C₁₆H₁₇N₃O₆ for [M+H]⁺ = 348.1190, found [M+H]+ = 348.1194.

2-(3-(Methoxycarbonyl)-4-methyl-5-(4-nitrophenyl)-1*H*-pyrrole-2-carboxamido)ethan-1-aminium acetate (13)



Brown solid 11.8 mg (19% yield). ¹H-NMR (500 MHz, DMSO- d_6) δ 9.38 (t, J = 5.6 Hz, 1H), 8.28-8.25 (m, 2H), 7.77 (m, 2H), 4.43 (s, 3H), 3.81 (s, 3H), 3.36 (dd, J = 12.5, 6.6 Hz, 2H), 2.73 (t, J = 7.0 Hz, 2H), 2.31 (s, 3H), 1.81 (s, 3H), 1.70 (q, J = 6.9 Hz, 2H) ppm. 13C-NMR (126 MHz, DMSO d_6) δ 173.1, 166.4, 159.8, 145.8, 137.9, 131.3, 129.0, 128.8, 123.5, 120.6, 114.5, 51.7, 37.8, 36.3, 30.2, 22.5, 12.1 ppm. HRMS (ESI): m/z calcd C₁₇H₂₀N₄O₅ for [M+H]⁺ = 361.1506, found [M+H]+ = 361.1508.

Methyl 2-((2-amino-2-oxoethyl)(methyl)carbamoyl)-5-(4-nitrophenyl)-4-(*p*-tolyl)-1*H*-pyrrole-3-carboxylate (14)



Yellow solid 20 mg (30%). Measured at 25°C: ¹H-NMR (500 MHz, DMSO- d_6) δ 12.66 (s, 1H), 8.10 (d, J = 8.4 Hz, 2H), 7.40 (d, J = 8.3 Hz, 2H), 7.38-7.23 (multiplet for NH₂ rotamers, 2H), 7.18 (d, J = 7.9 Hz, 2H), 7.13-7.11 (m, 2H), 4.1 and 3.9 (two singlets of CH₂ rotamers, 2H), 3.55 (s, 3H), 3.05 and 2.99 (two singlets of CH₃ rotamers, 3H), 2.36 (s, 3H) ppm. Measured at 115°C: ¹H-NMR (500 MHz, DMSO- d_6) δ 12.04 (s, 1H), 8.07-8.05 (m, 2H), 7.47-7.44 (m, 2H), 7.18 (d, J = 7.7 Hz,

2H), 7.14 (d, J = 8.2 Hz, 2H), 6.97 (s, 2H), 4.05 (s, 2H), 3.57 (s, 3H), 3.04 (s, 3H), 2.38 (s, 3H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 169.7, 163.8, 163.3, 145.5, 138.0, 136.2, 132.5, 131.1, 130.3, 128.7, 127.7, 127.4, 124.6, 123.6, 113.1, 51.1, 49.8, 37.1, 20.9 ppm. HRMS (ESI): m/z calcd C₂₃H₂₂N₄O₆ for [M+H]⁺ = 451.1612, found [M+H]+ = 451.1609.

Methyl 2-(2-carbamoylpyrrolidine-1-carbonyl)-5-(4-nitrophenyl)-4-(*p*-tolyl)-1*H*-pyrrole-3carboxylate (15)



Yellow solid 7.2 mg (17% yield). Measured at 25°C: ¹H-NMR (500 MHz, DMSO- d_6) δ 12.43 (s, 1H), 8.10-8.04 (two doublets of C_{Ar}-H_{Ar}, 2H), 7.40-7.34 (two doublets of C_{Ar}-H_{Ar}, 2H), 7.24 (s, 2H), 7.16 (d, *J* = 7.9 Hz, 2H), 7.10 (d, *J* = 8.0 Hz, 2H), 4.41 (dd, *J* = 8.5, 4.5 Hz, 1H), 3.52 (s, 3H), 3.46-3.37 (m, 2H), 2.31-2.38 (3H), 2.21-2.13 (m, 1H), 2.00-1.94 (m, 1H), 1.92-1.80 (m, 2H) ppm. Measured at 115°C. ¹H-NMR (500 MHz, DMSO- d_6) δ 11.85 (s, 1H), 8.07-8.04 (m, 2H), 7.47-7.45 (m, 2H), 7.18 (d, J = 7.9 Hz, 2H), 7.13 (dd, J = 6.2, 2.0

Hz, 2H), 6.87 (s, 2H), 4.48 (q, J = 4.2 Hz, 1H), 3.56 (s, 3H), 3.55-3.48 (m, 2H), 2.38 (s, 3H), 2.23-2.15 (m, 1H), 2.11-2.05 (m, 1H), 2.00-1.85 (m, 2H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 173.1, 164.0, 161.7, 145.5, 138.0, 136.2, 133.4, 131.0, 130.3, 128.7, 127.7, 127.3, 124.5, 123.6, 113.0, 59.7, 51.1, 48.3, 29.6, 24.2, 20.9 ppm. HRMS (ESI): m/z calcd C₂₅H₂₄N₄O₆ for [M+H]⁺ = 477.1769, found [M+H]⁺ = 477.1771.

3-(methoxycarbonyl)-5-(4-nitrophenyl)-4-(p-tolyl)-1H-pyrrole-2-carboxylic acid (16)



Yellow solid 14.4 mg (22% yield). ¹H-NMR (500 MHz, DMSO- d_6) δ 8.11 (d, *J* = 8.9 Hz, 2H), 7.52 (d, *J* = 9.0 Hz, 2H), 7.13 (d, *J* = 7.9 Hz, 2H), 7.03 (d, *J* = 8.0 Hz, 2H), 3.61 (s, 3H), 2.30 (s, 3H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 166.2, 161.2, 146.0, 137.5, 136.4, 130.3, 129.3, 129.2, 129.1, 124.3, 123.4, 123.3, 122.0, 51.8, 20.7 ppm. HRMS (ESI): m/z calcd C₂₀H₁₆N₂O₆ for [M+H]⁺ = 381.1081, found [M+H]⁺ = 381.1082.

Methyl 2-carbamoyl-1-methyl-5-(4-nitrophenyl)-4-(*p*-tolyl)-1*H*-pyrrole-3-carboxylate (19)



Yellow solid 9.6 mg (16% yield). ¹H-NMR (500 MHz, DMSO- d_6) δ 8.20-8.17 (m, 2H), 8.06 (s, 1H), 7.79 (s, 1H), 7.48-7.46 (m, 2H), 7.03 (d, J = 7.9 Hz, 2H), 6.95 (d, J = 8.0 Hz, 2H), 3.56 (s, 3H), 3.54 (s, 3H), 2.26 (s, 3H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 164.0, 163.2, 146.7, 137.4, 135.5, 134.1, 132.2, 130.7, 130.4, 130.3, 128.2, 123.9, 123.3, 112.3, 50.9, 33.5, 20.7 ppm. HRMS (ESI): m/z calcd C₂₁H₁₉N₃O₅ for [M+H]⁺ = 394.1397, found [M+H]+ = 394.1398.

Methyl 1-benzyl-2-carbamoyl-5-(4-nitrophenyl)-4-(p-tolyl)-1H-pyrrole-3-carboxylate (20)



Light yellow solid 19.7 mg (30% yield). ¹H-NMR (500 MHz, DMSO- d_6) δ 8.09-8.06 (m, 3H), 7.72 (s, 1H), 7.30-7.28 (m, 2H), 7.23-7.17 (m, 3H), 7.00 (d, J = 8.0 Hz, 2H), 6.97-9.95 (m, 2H), 6.85-6.83 (m, 2H), 5.29 (s, 2H), 3.58 (s, 3H), 2.24 (s, 3H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 164.1, 163.2, 146.8, 137.6, 137.1, 135.5, 133.5, 132.3, 130.8, 130.4, 130.2, 128.3, 128.2, 127.2, 126.3, 124.3, 123.2, 113.2, 51.1, 48.5, 20.7 ppm. HRMS (ESI): m/z calcd C₂₇H₂₃N₃O₅ for [M+H]⁺ = 470.1710, found [M+H]+ = 470.1712.

Methyl 5-(4-aminophenyl)-2-carbamoyl-4-(p-tolyl)-1H-pyrrole-3-carboxylate (22)



White solid 7 mg (22%). ¹H-NMR (500 MHz, DMSO- d_6) δ 11.69 (s, 1H), 8.25 (s, 1H), 7.44 (s, 1H), 7.08-7.06 (m, 2H), 6.98-6.96 (m, 2H), 6.89-6.86 (m, 2H), 6.42-6.39 (m, 2H), 5.15 (s, 2H), 3.49 (s, 3H), 2.29 (s, 3H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 167.1, 160.8, 148.1, 135.1, 132.4, 132.3, 129.9, 129.1, 128.4, 126.3, 121.4, 118.2, 116.1, 113.3, 51.4, 40.1, 40.0, 39.9, 39.9, 39.8, 39.7, 39.6, 39.5, 39.4, 39.4, 39.2, 39.0, 20.8 ppm. HRMS (ESI): m/z calcd C₂₀H₁₉N₃O₃ for [M+H]⁺ = 350.1499, found [M+H]+ = 350.1500.

5-(4-nitrophenyl)-N³-propyl-4-(p-tolyl)-1H-pyrrole-2,3-dicarboxamide (23)



Yellow solid 10.7 mg (29% Yield). ¹H-NMR (500 MHz, DMSO- d_6) δ 12.32 (s, 1H), 8.27 (s, 1H), 8.12-8.10 (m, 2H), 7.51-7.48 (m, 3H), 7.45 (t, *J* = 5.7 Hz, 1H), 7.17 (d, *J* = 7.9 Hz, 2H), 7.09 (dd, *J* = 6.3, 1.7 Hz, 2H), 2.99 (dd, *J* = 12.7, 6.8 Hz, 2H), 2.33 (s, 3H), 1.19 (td, *J* = 14.3, 7.3 Hz, 2H), 0.59 (t, *J* = 7.4 Hz, 3H) ppm. ¹³C-NMR (126 MHz, DMSO-D6) δ 165.8, 161.3, 145.8, 137.8, 136.4, 130.6, 130.1, 129.0, 128.8, 128.2, 126.6, 123.9, 123.3, 122.0, 40.7, 21.8, 20.8, 11.1 ppm. HRMS (ESI): m/z calcd C₂₂H₂₂N₄O₄ for [M+H]⁺ = 407.1714, found [M+H]+ = 407.1713.

*N*³-(3-hydroxypropyl)-5-(4-nitrophenyl)-4-(*p*-tolyl)-1*H*-pyrrole-2,3-dicarboxamide (24)



Yellow solid 25 mg (66% Yield). ¹H-NMR (500 MHz, DMSO- d_6) δ 12.32 (s, 1H), 8.29 (s, 1H), 8.11 (dt, J = 9.4, 2.3 Hz, 2H), 7.51-7.48 (m, 3H), 7.45 (t, J = 5.7 Hz, 1H), 7.17 (dd, J = 8.3, 0.6 Hz, 2H), 7.09-7.07 (m, 2H), 4.29 (t, J = 5.2 Hz, 1H), 3.18 (dd, J = 11.4, 6.1 Hz, 2H), 3.09 (dd, J = 12.8, 6.8 Hz, 2H), 2.34 (s, 3H), 1.36-1.30 (m, 2H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 165.8, 161.3, 145.8, 137.9, 136.4, 130.6, 130.0, 129.1, 128.8, 128.2, 126.7, 123.9, 123.3, 122.0, 58.2, 36.3, 31.8, 20.8 ppm. HRMS (ESI): m/z calcd C₂₂H₂₂N₄O₅ for [M+H]⁺ = 423.1663, found

[M+H]+ = 423.1659.

(*S*)-2-(1-amino-4-methoxy-1,4-dioxobutan-2-yl)-3-(4-methylbenzoyl)-6-nitro-2*H*-indazole 1-oxide (27)



Yellow solid 25mg (65% yield). ¹H-NMR (500 MHz, DMSO- d_6) δ 8.64 (dd, J = 2.1, 0.6 Hz, 1H), 7.98 (dd, J = 9.5, 2.0 Hz, 1H), 7.77 (d, J = 8.2 Hz, 2H), 7.48-7.46 (m, 3H), 7.44 (br.s, 1H), 7.10 (dd, J = 9.5, 0.7 Hz, 1H), 6.48 (dd, J = 7.0 Hz, J = 6.4 Hz, 1H), 3.59 (dd, J = 17.0, 6.4 Hz, 1H), 3.55 (s, 3H), 3.22 (dd, J = 17.0, 7.0 Hz, 1H), 2.49 (s, 3H) ppm. ¹³C-NMR (126 MHz, DMSO- d_6) δ 181.6, 170.4, 166.2, 145.3, 144.1, 135.7, 129.5, 129.5, 127.2, 122.3, 120.9, 120.9, 118.7, 111.5, 56.2, 51.8, 32.6, 21.3 ppm. NMR data were compared with analogical 2*H*-Indazole 1-oxides reported in Bouillon, I. et al. J. Org. Chem 2008, 73, 9027-9032. HRMS

(ESI): m/z calcd $C_{20}H_{18}N_4O_7$ for $[M+H]^+ = 427.1248$, found [M+H] + = 427.1247.

NMR spectra of the final compounds



Methyl 2-carbamoyl-5-(4-nitrophenyl)-4-(p-tolyl)-1H-pyrrole-3-carboxylate (6)





















Methyl 2-carbamoyl-4-(4-methoxyphenyl)-5-(4-nitrophenyl)-1*H*-pyrrole-3-carboxylate (7)













¹H-¹³C HMBC Correlations

¹H -¹H NOESY Correlations







Methyl 2-carbamoyl-4-(4-fluorophenyl)-5-(4-nitrophenyl)-1H-pyrrole-3-carboxylate (8)





Methyl 2-carbamoyl-5-(4-nitrophenyl)-4-(thiophen-3-yl)-1H-pyrrole-3-carboxylate (9)





Methyl 2-carbamoyl-4-methyl-5-(4-nitrophenyl)-1*H*-pyrrole-3-carboxylate (10)





Methyl (S)-2-((1-amino-1-oxopropan-2-yl)carbamoyl)-4-methyl-5-(4-nitrophenyl)-1H-pyrrole-3-carboxylate (11)







2-(3-(Methoxycarbonyl)-4-methyl-5-(4-nitrophenyl)-1*H*-pyrrole-2-carboxamido)ethan-1-aminium acetate (13)

Methyl 2-((2-amino-2-oxoethyl)(methyl)carbamoyl)-5-(4-nitrophenyl)-4-(p-tolyl)-1H-pyrrole-3-carboxylate (14)

The presence of rotamers was confirmed by temperature-dependent proton measurements at 25°C, 35°C, 45°C, 55°C, 85°C and 115°C. We present only proton spectra at standard 25°C and the highest temperature 115°C.

¹H Proton spectra of compound **14** at 25°C - Selected area

¹H Proton spectra of compound **14** at 115°C - Selected area

¹H Proton spectra of compound **14** at 25°C - Selected area

¹H Proton spectra of compound **14** at 115°C - Selected area

Methyl 2-(2-carbamoylpyrrolidine-1-carbonyl)-5-(4-nitrophenyl)-4-(*p*-tolyl)-1*H*-pyrrole-3-carboxylate (15)

The presence of rotamers was confirmed by temperature-dependent proton measurements at 25°C, 35°C, 45°C, 55°C, 85°C and 115°C. We present only proton spectra at standard 25°C and the highest temperature 115°C.

¹H Proton spectra of compound **15** at 25°C - Selected area

¹H Proton spectra of compound **15** at 115°C - Selected area

3-(methoxycarbonyl)-5-(4-nitrophenyl)-4-(p-tolyl)-1H-pyrrole-2-carboxylic acid (16)

Methyl 2-carbamoyl-1-methyl-5-(4-nitrophenyl)-4-(*p*-tolyl)-1*H*-pyrrole-3-carboxylate (19)

Methyl 1-benzyl-2-carbamoyl-5-(4-nitrophenyl)-4-(*p*-tolyl)-1*H*-pyrrole-3-carboxylate (20)

Methyl 5-(4-aminophenyl)-2-carbamoyl-4-(*p*-tolyl)-1*H*-pyrrole-3-carboxylate (22)

5-(4-nitrophenyl)-*N*³-propyl-4-(*p*-tolyl)-1*H*-pyrrole-2,3-dicarboxamide (23)

N³-(3-hydroxypropyl)-5-(4-nitrophenyl)-4-(*p*-tolyl)-1*H*-pyrrole-2,3-dicarboxamide (24)

S)-2-(1-amino-4-methoxy-1,4-dioxobutan-2-yl)-3-(4-methylbenzoyl)-6-nitro-2*H*-indazole 1-oxide (27)

