Supplementary Information for

Electrochemical S-H and O-H Insertion Reactions from Thiols or

Salicylic Acids with Diazo Esters

Zhiqin He,^a Wei Zhao,^a Yufeng Li,^a Yang Yu,^c Fei Huang*,^{a,b}

^aSchool of Food Science and Pharmaceutical Engineering, Nanjing Normal University, Nanjing, 210023, P. R. China
^bSchool of Pharmaceutical Sciences, Nanjing Tech University, Nanjing, 211816, P. R. China
^cSchool of Environmental Science and Engineering, Nanjing Tech University, Nanjing, 211816, P. R. China
Email: huangfei0208@yeah.net

Table of contents

Contents:	page
1. General information	S2
2. Experimental procedures	S2
3. Characterization data for products	S15
4. References	S26
5. NMR spectra	S27

1. General information

The solvents were dried and distilled prior to use by the literature methods. Analytical TLC plates, Sigma-Aldrich silica gel 60F200 were viewed by UV light (254 nm). Column chromatographic purifications were performed on SDZF silica gel 160. The instrument for electrolysis was IKA ElectraSyn 2.0 and both cathode and anode electrodes were graphite electrodes (52 mm \times 8 mm \times 2 mm). ¹H and ¹³C NMR spectra were obtained on a Bruker NMR spectrometer at 400 MHz and 100 MHz, respectively, and referenced internally based on the residual solvent signal. The data reporting for ¹H NMR spectra is as follows: chemical shift (δ , ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet), coupling constant in hertz, and number of protons. The data reporting for ${}^{13}C$ spectra is given as chemical shift (δ , ppm). High-resolution mass spectra (HRMS) were measured on a Waters GC-TOF CA156 mass spectrometer. X-ray Crystallographic analysis was achieved by the Shanghai institute of Organic Chemistry, Chinese Academy of Sciences. All the chemical reagents were purchased from commercial sources and used as received unless otherwise indicated. Compounds **3a**,¹ **3b**,¹ **3c**,¹ **3d**,² **3e**,² **3g**,³ **3h**,⁴ **3i**,¹ **3j**,¹ **3k**,⁵ **31**,⁵ **3m**,¹ **3n**,² **3o**,¹ **3p**,² **3q**,⁶ **3w**,⁷ **3y**,⁸ **3z1**,⁹ **5a**,⁶ and **5f**¹⁰ are known and their spectroscopic feature is in good agreement with that reported in the literature.

2. Experimental procedures

2.1 Preparation of a-diazoesters¹



A typical procedure for the synthesis of a-diazoesters 1 - Synthesis of 1b: DBU (2.24 mL, 15 mmol) was added slowly to a stirred solution of ethyl 2phenylacetate (sm1a, 1.41 mL, 10.0 mmol) and tosylazide (sm2, 2.42 mL, 11.0 mmol) in the CH₃CN (20 mL) at 0 °C. After that, it was placed in microwave reactor that was heated to 40 °C (400 W, monitored by IR temperature sensor) and maintained at this temperature for 30 min. After cooling to room temperature, the reaction mixture was quenched with saturated aqueous solution of NH₄Cl (5 mL), extracted with DCM (3×30 mL), washed with brine (3×30 mL), dried over anhydrous Na₂SO₄, and concentrated under reduced pressure to give the product. The residue was purified by flash chromatography (petroleum ether (60-90 °C)/AcOEt, 10:1) to afford the corresponding ethyl-2-diazo-2-phenylacetate **1b** as a yellow oil (1.65 g, 87%).

2.2 Screening the optimum reaction conditions for the synthesis of 5a

	N ₂ CO ₂ Et +	СООН	C (+) C (-), I = 5 mA TBAI (0.9 equiv) DCE (10 mL) undivided cell, rt, 7 h	COOH	CO ₂ Et
	1b	4a		5	a
Entry	Variation f	rom the	standard conditio	ns	Yield ^b (%)
1		n	one		80
2	DCM instead of DCE				51
3	MeCN instead of DCE				40
4	C (+) Ni (-) instead of C (+) C (-)				49
5	C (+) Pt (-) instead of C (+) C (-)				37
6	3	mA inst	ead of 5 mA		60
7	7 mA instead of 5 mA				36
8 ^c	TBAB instead of TBAI				20
9	ⁿ Bu ₄ NBF ₄ instead of TBAI				23
10	0.6 mmol 4a				34
11	1.2 mmol 4a				64
12	No electric current				33

Table S1. Screening of reaction conditions.^a

^aReaction Conditions: **1b** (0.3 mmol), **4a** (0.9 mmol), TBAI (0.27 mmol) in DCE (10 mL), graphite electrodes (52 mm \times 8 mm \times 2 mm) as anode and cathode, constant current = 5 mA, undivided cell, room temperature, 7 h. ^{*b*}Isolated yield.

Among a set of representative solvents (Table S1,[†] entry 1-3), DCE performed better. On replacing the graphite cathode with a Ni cathode or a Pt cathode, the yields decreased strongly (Table S1,[†] entries 4 and 5). Decreasing the operating current to 3 mA or increasing the operating current to 7 mA delivered the product **5a** in low yield (Table S1,[†] entries 6 and 7). Other electrolytes, such as TBAB or ⁿBu₄NBF₄ gave a worse performance than TBAI (Table S1,[†] entries 8 and 9). The ratio of **4a:1b** was also evaluated, 1:2 and 1:4 gave inferior results (Table S1,† entries 10 and 11). At last, it was observed that this conversion reduced sharply without current, suggesting that electricity was critical for this reaction (Table S1,† entry 12).

2.3 General procedure for S-H insertion of thiols



A typical procedure for the synthesis of S-H insertion products of thiols (3) -Synthesis of ethyl 2-((4-chlorophenyl)thio)acetate (3a): An ElectraSyn vial (12 mL) with a stir bar was charged with TBAB (80.6 mg, 0.25 mmol), ethyl 2-diazoacetate (1a) (57.1 mg, 0.5 mmol) and 4-chlorobenzenethiol (2a) (144.6 mg, 1.0 mmol) and DCE (10 mL). [Liquid compounds were added after the addition of DCE. The order of the addition does not affect the result] The ElectraSyn vial cap equipped with graphite electrodes was inserted into the mixture. Then, the reacton mixture was electrolyzed under a constant current of 5 mA until complete consumption of the starting material as judged by TLC. After the reaction, the ElectraSyn vial cap was removed and the combined organic phase was concentrated under reduced pressure. The resultant mixture was subject to purification by column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **3a** as a colorless oil (86.5 mg, 75%).

Graphical guide for electrochemical S-H insertion:







Left: graphite electrodes, ElectraSyn 2.0 cap and ElectraSyn 2.0 vial (12 mL). Middle: ElectraSyn 2.0 cap equipped with graphite electrodes. Right: the cap was screwed into the vial and the electrodes were submerged into the solution.



Left: select New experiments. Middle: select Constant Current. Right: adjust the current value.



Left: select Time. Middle: define the "time. Right: indicate the "mmol" of the substrate.



Left: no alternate polarity. Middle: select the "start". Right: start the reaction on the ElectraSyn 2.0 with a stirring speed of 500 rpm.

2.4 General procedure for O-H insertion of salicylic acids



A typical procedure for the synthesis of O-H insertion products of salicylic acids (5) - Synthesis of (R)-2-(2-ethoxy-2-oxo-1-phenylethoxy)benzoic acid (5a): An ElectraSyn vial (12 mL) with a stir bar was charged with TBAI (99.7 mg, 0.27 mmol), ethyl 2-diazo-2-phenylacetate (1b) (57.1 mg, 0.3 mmol) and salicylic acid (4a) (124.3 mg, 0.9 mmol) and DCE (10 mL). [Liquid compounds were added after the addition of DCE. The order of the addition does not affect the result]. The ElectraSyn vial cap equipped with graphite electrodes were inserted into the mixture. Then, the mixture was electrolyzed under a constant current of 5 mA until complete consumption of the starting material as judged by TLC. After the reaction, the the ElectraSyn vial cap was removed and the combined organic phase was concentrated under reduced pressure. The resultant mixture was subject to purification by column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **5a** as a colorless oil (72.1 mg, 80%).

2.5 A gram-scale experiment





An ElectraSyn vial (20 mL) with a stir bar was charged with TBAB (161.2 mg, 0.5 mmol), ethyl 2-diazoacetate (1a) (1.00 g, 8.8 mmol) and 4-chlorobenzenethiol (2a) (1.90 g, 13.2 mmol) and DCE (15 mL). [Liquid compounds were added after the addition of DCE. The order of the addition does not affect the result]. The ElectraSyn vial cap equipped with graphite electrodes were inserted into the mixture. Then, the reacton mixture was electrolyzed under a constant current of 5 mA until complete set

consumption of the starting material as judged by TLC. After the reaction, the ElectraSyn vial cap was removed and the combined organic phase was concentrated under reduced pressure. The resultant mixture was subject to purification by column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **3a** as a colorless oil (1.67 g, 82%).

1 gram-scale experiment for the synthesis of 5a



An ElectraSyn vial (20 mL) with a stir bar was charged with TBAI (0.89 g, 2.4 mmol), ethyl 2-diazo-2-phenylacetate (**1b**) (1.01 g, 5.3 mmol) and salicylic acid (**4a**) (1.46 mg, 10.6 mmol) and DCE (15 mL). [Liquid compounds were added after the addition of DCE. The order of the addition does not affect the result]. The ElectraSyn vial cap equipped with anode (graphite) and cathode (graphite) were inserted into the mixture. Then, the mixture was electrolyzed under a constant current of 5 mA until complete consumption of the starting material as judged by TLC for 24 hours. After completion of the reaction, the ElectraSyn vial cap was removed and the combined organic phase was concentrated under reduced pressure. The resultant mixture was subject to purification by column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **5a** as a colorless oil (1.25 g, 79%).

2.6 Mechanism research

Radical-trapping experiment with TEMPO



An ElectraSyn vial (12 mL) with a stir bar was charged with TBAB (80.6 mg, 0.25 mmol), ethyl 2-diazoacetate (1a) (57.1 mg, 0.5 mmol), 4-chlorobenzenethiol (2a)

(144.6 mg, 1.0 mmol), TEMPO (156.3 mg, 1.0 mmol) and DCE (10 mL). [Liquid compounds were added after the addition of DCE. The order of the addition does not affect the result]. The ElectraSyn vial cap equipped with graphite electrodes were inserted into the mixture. Then, the reacton mixture was electrolyzed under a constant current of 5 mA until complete consumption of the starting material as judged by TLC. After the reaction, the ElectraSyn vial cap was removed and the combined organic phase was concentrated under reduced pressure. There, **3a** was not detected by TLC and HRMS. The resultant mixture was subject to purification by column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **6a** as a colorless oil (27.0 mg, 18%).

Radical-trapping experiment with DPE



An ElectraSyn vial (12 mL) with a stir bar was charged with TBAB (80.6 mg, 0.25 mmol), ethyl 2-diazoacetate (1a) (57.1 mg, 0.5 mmol), 4-chlorobenzenethiol (2a) (144.6 mg, 1.0 mmol), DPE (180.3 mg, 1.0 mmol) and DCE (10 mL). [Liquid compounds were added after the addition of DCE. The order of the addition does not affect the result]. The ElectraSyn vial cap equipped with graphite electrodes were inserted into the mixture. Then, the reacton mixture was electrolyzed under a constant current of 5 mA until complete consumption of the starting material as judged by TLC. After the reaction, the ElectraSyn vial cap was removed and the combined organic phase was concentrated under reduced pressure. The resultant mixture was subject to purification by column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **3a** as a colorless oil (62.3 mg, 54%). And the radical intermediated **3a'** was trapped by DPE were detected by HRMS (**Figure S1**).



Figure S1. HRMS analysis of the formation of product 3a'.

The experiment starting from disulfide 7a



An ElectraSyn vial (12 mL) with a stir bar was charged with TBAB (80.6 mg, 0.25 mmol), 1,2-bis(4-chlorophenyl)disulfane (7a) (287.3 mg, 1.0 mmol), ethyl 2diazoacetate (1a) (57.1 mg, 0.5 mmol), MeOH (0.5 mL) and DCE (10 mL). [Liquid compounds were added after the addition of DCE. The order of the addition does not affect the result]. The ElectraSyn vial cap equipped with graphite electrodes were inserted into the mixture. Then, the reacton mixture was electrolyzed under a constant current of 5 mA until complete consumption of the starting material as judged by TLC. After the reaction, the ElectraSyn vial cap was removed and the combined organic phase was concentrated under reduced pressure. The resultant mixture was subject to purification by column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **3a** as a colorless oil (28 mg, 24%).

The gas protection experiments



The gas protection test cannot be conducted for IKA ElectraSyn 2.0 in our laboratory (eqn (a)), thus we use the dual display potentiostat (DP-305) (made in China) for this control experiments. In an oven-dried undivided three-necked cell (25 mL) equipped with a stir bar, TBAB (80.6 mg, 0.25 mmol) and 4-chlorobenzenethiol (**2a**) (144.6 mg, 1.0 mmol) were combined and added. The cell was equipped with carbon cloth (60 mm × 6 mm) as the anode and cathode and then charged with nitrogen. Under the protection of N₂, DCE (10 mL) and ethyl 2-diazoacetate (**1a**) (57.1 mg, 0.5 mmol) was injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzsd at a constant current of 5.0 mA at room temperature for 3.0 h. After the reaction, the pure product was obtained by flash column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **3a** as a colorless oil (64.0 mg, 56%) (eqn (b)).

In an oven-dried undivided three-necked cell (25 mL) equipped with a stir bar, TBAB (80.6 mg, 0.25 mmol), 4-chlorobenzenethiol (**2a**) (144.6 mg, 1.0 mmol), DCE (10 mL) and ethyl 2-diazoacetate (**1a**) (57.1 mg, 0.5 mmol) were combined and added. The cell was equipped with carbon cloth (60 mm \times 6 mm) as the anode and cathode. The reaction mixture was stirred and electrolyzsd at a constant current of 5.0 mA at room temperature for 3.0 h. After the reaction, the pure product was obtained by flash column chromatography on silica gel (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **3a** as a colorless oil (57.0 mg, 50%) (eqn (c)).

In an oven-dried undivided three-necked cell (25 mL) equipped with a stir bar, TBAB (80.6 mg, 0.25 mmol) and 4-chlorobenzenethiol (**2a**) (144.6 mg, 1.0 mmol) were combined and added. The cell was equipped with carbon cloth (60 mm × 6 mm) as the anode and cathode and then charged with oxygen. Under the protection of O_2 , DCE (10 mL) and ethyl 2-diazoacetate (**1a**) (57.1 mg, 0.5 mmol) was injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzsd at a constant current of 5.0 mA at room temperature for 3.0 h. After the reaction, the pure product was obtained by flash column chromatography on silica gel. (eluent: petroleum ether (60-90 °C)/ethyl acetate = 10:1, v/v) to afford **3a** as a colorless oil (30.0 mg, 26%) (eqn (d)).

In an oven-dried undivided three-necked cell (25 mL) equipped with a stir bar, TBAB (80.6 mg, 0.25 mmol) was added. The cell was equipped with carbon cloth (60 mm \times 6 mm) as the anode and cathode and then charged with nitrogen. Under the protection of N₂, DCE (10 mL) and ethyl 2-diazo-2-phenylacetate (**1b**) (95.1 mg, 0.5 mmol) was injected respectively into the tubes via syringes. The reaction mixture was stirred and electrolyzsd at a constant current of 5.0 mA at room temperature (eqn (e)). And we found the formation of **1b'** (the dimerization of **1b**) detected by HRMS (**Figure S2**).



Figure S2. HRMS analysis of the formation of product 1b'.

2.7 CV experiments

Cyclic voltammetry was performed in a three electrode cell under air at room temperature. The working electrode was a 3 mm glassy carbon electrode while the counter electrode was a platinum mesh. The reference was an Ag/AgCl electrode submerged in saturated aqueous KCl solution. The solvent (DCE = 10 mL) containing TBAB (0.1 mmol) and "Bu₄NBF₄ (0.1 mmol) were poured into thr electrochemical cell in cyclic voltammetry experiments. The scan rate was 0.1 V/s, ranging from 0.0 V to 3.8 V. The peak potentials *vs*. Ag/AgCl for used. An obvious oxidation peak of TEMPO was observed at 2.70 V. The oxidation peak of 4-chlorobenzenethiol **2a** could also observed at 2.50 V. So, **2a** was oxidized preferentially at the anode than TEMPO in this system, which indicated that the feasibility of TEMPO as a free radical trapping agent. A reduction peak of 4,4'-dichlorodiphenyl disulfide **7a** was observed at -1.58 V under the reaction solvent system. Therefore, the **7a** may involve in reduction processes in the catalytic cycle.



Figure S3. Cyclic voltammograms of related compounds (0.1 mmol) in DCE with ${}^{n}Bu_{4}NBF_{4}$ (0.01 M) and TBAB (0.01 M) at a scan rate of v = 100 mV/s.



Figure S4. Cyclic voltammetry of 7a (0.1 mmol) in DCE with TBAB (0.01 M) at a scan rate of v = 100 mV/s.

2.8 Failed experiments

Table S2. Different substituent groups on the benzene ring of phenol.^a



^{*a*} Conditions: **1b** (0.3 mmol), **B** (0.9 mmol), TBAI (0.27 mmol) in DCE (10 mL), graphite electrodes (52 mm \times 8 mm \times 2 mm) as anode and cathode, constant current = 5 mA, undivided cell, room temperature, 7 h, isolated yield.

For O-H insertion, we have tried different phenols with diazo compounds for the construction of C-O bonds. However, no obvious target product was produced. As shown in the Table S2, 4-hydroxybenzoic acid, 3-hydroxybenzoic acid and 2,4-dihydroxybenzoic acid all contain carboxyl group, but the envisaged products were not generated. Therefore, we speculated that it may be related to hydrogen bond and the damage of this system to the electrode.¹¹

2.9 NMR experiments

No obvious target product was produced when the substrate contained carboxylic acids, such as 4-hydroxybenzoic acid 4d and 3-hydroxybenzoic acid 4e. Therefore, it cannot be ruled out whether acidity can promote the reaction. Based on the above results and the literature proceedings,¹² we speculate that hydrogen bond plays a vital role in this conversion. Then, hydrogen bond was further studied through NMR in (CD₃)₂SO. Comparing the chemical shifts of hydrogen in hydroxyl group in Fig S5, obvious hydrogen bond observed for 2-hydroxybenzoic acid was (13.8921>10.2144>9.7373), which agreed with our previously proposed hydrogenbond interaction.



Figure S5. ¹H NMR spectra of the benzoic acid signals in $(CD_3)_2SO$. (a) 2-hydroxybenzoic acid, (b) 3-hydroxybenzoic acid, (c) 4-hydroxybenzoic acid.

3. Characterization data for products



Ethyl 2-((4-chlorophenyl)thio)acetate (3a): 86.5 mg, 75% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.35 (d, J = 8.5 Hz, 2H), 7.27 (d, J = 8.5 Hz, 2H), 4.16 (q, J = 7.1 Hz, 2H), 3.60 (s, 2H), 1.22 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.6, 133.6, 133.3, 131.6, 129.3, 61.8, 37.0, 14.2. The analytical data are consistent with literature.¹



Ethyl 2-((4-methoxyphenyl)thio)acetate (3b): 84.9 mg, 75% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, J = 8.8 Hz, 2H), 6.84 (d, J = 8.8 Hz, 2H), 4.13 (q, J = 7.1 Hz, 2H), 3.79 (s, 3H), 3.50 (s, 2H), 1.21 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.1, 159.8, 134.4, 125.1, 114.8, 61.5, 55.5, 38.8, 14.2. The analytical data are consistent with literature.¹



Ethyl 2-(p-tolylthio)acetate (3c): 73.6 mg, 70% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, J = 8.1 Hz, 2H), 7.11 (d, J = 8.0 Hz, 2H), 4.15 (q, J = 7.1 Hz, 2H), 3.58 (s, 2H), 2.32 (s, 3H), 1.22 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 169.9, 137.4, 131.2, 131.0, 129.9, 61.6, 37.5, 21.2, 14.2. The analytical data are consistent with literature.¹



Ethyl 2-(m-tolylthio)acetate (3d): 67.3 mg, 64% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.25 – 7.14 (m, 3H), 7.03 (d, *J* = 7.0 Hz, 1H), 4.16 (q, *J* = 7.1 Hz, 2H), 3.62 (s, 2H), 2.32 (s, 3H), 1.22 (t, *J* = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.8, 138.8, 134.8, 130.6, 128.9, 127.8, 127.0, 61.5, 36.7, 21.3, 14.1. The analytical data are consistent with literature.²



Ethyl 2-(o-tolylthio)acetate (3e): 76.8 mg, 73% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.33 (m, 1H), 7.20 – 7.09 (m, 3H), 4.15 (q, *J* = 7.1 Hz, 2H), 3.61 (s, 2H), 2.42 (s, 3H), 1.22 (t, *J* = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.8, 138.5, 134.3, 130.4, 129.7, 127.0, 126.7, 61.7, 36.1, 20.5, 14.2. The analytical data are consistent with literature.²



Ethyl 2-((2-(trifluoromethyl)phenyl)thio)acetate (3f): 80.6 mg, 61% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.66 (d, J = 7.7 Hz, 1H), 7.60 (d, J = 7.9 Hz, 1H), 7.48 (t, J = 7.4 Hz, 1H), 7.33 (t, J = 7.6 Hz, 1H), 4.15 (q, J = 7.1 Hz, 2H), 3.65 (s, 2H), 1.20 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.2, 134.5, 132.7, 132.3, 130.7 (q, J = 30.2 Hz), 127.1, 127.0 (q, J = 5.7 Hz), 123.8 (q, J = 273.7 Hz), 61.8, 37.1, 14.1. HRMS (EI) calcd for C₁₁H₁₂F₃O₂S [M+H]⁺ 265.0505, found 265.0513.



Ethyl 2-((3,4-dichlorophenyl)thio)acetate (3g): 120.6 mg, 91% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.49 (d, J = 2.2 Hz, 1H), 7.35 (d, J = 8.4 Hz, 1H), 7.22 (dd, J = 8.4, 2.2 Hz, 1H), 4.18 (q, J = 7.1 Hz, 2H), 3.62 (s, 2H), 1.24 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.2, 135.4, 133.1, 131.3, 130.8, 129.0, 100.1, 62.0, 36.5, 14.2. The analytical data are consistent with literature.³



Ethyl 2-(naphthalen-1-ylthio)acetate (3h): 103.5 mg, 84% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 8.45 (d, *J* = 8.5 Hz, 1H), 7.86 (d, *J* = 7.6 Hz, 1H), 7.80 (d, *J* = 8.2 Hz, 1H), 7.71 (dd, *J* = 7.2, 1.0 Hz, 1H), 7.62 – 7.49 (m, 2H), 7.42 (dd, *J* = 8.1, 7.3 Hz, 1H), 4.09 (q, *J* = 7.1 Hz,

2H), 3.66 (s, 2H), 1.14 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.8, 134.1, 133.4, 131.9, 130.7, 128.8, 126.9, 126.5, 125.7, 125.2, 61.6, 37.4, 14.1. The analytical data are consistent with literature.⁴



Ethyl 2-(naphthalen-2-ylthio)acetate (3i): 80.1 mg, 65% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.86 (s, 1H), 7.78 (m, 3H), 7.52 – 7.42 (m, 3H), 4.17 (q, J = 7.1 Hz, 2H), 3.74 (s, 2H), 1.21 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.7, 133.7, 132.4, 132.1, 128.6, 128.2, 127.7, 127.6, 127.3, 126.6, 126.1, 61.6, 36.7, 14.1. The analytical data are consistent with literature.¹



Ethyl 2-(pyridin-2-ylthio)acetate (3j): 94.7 mg, 96% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 8.35 (ddd, J = 4.9, 1.7, 0.9 Hz, 1H), 7.45 (ddd, J = 8.0, 7.5, 1.8 Hz, 1H), 7.19 (dt, J = 8.1, 0.9 Hz, 1H), 6.95 (ddd, J = 7.3, 4.9, 1.0 Hz, 1H), 4.16 (q, J = 7.1 Hz, 2H), 3.94 (s, 2H), 1.22 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.8, 157.0, 149.3, 136.1, 122.0, 119.8, 61.5, 32.3, 14.1. The analytical data are consistent with literature.¹



Ethyl 2-(thiophen-2-ylthio)acetate (3k): 70.0 mg, 69% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.38 (dd, J = 5.4, 1.2 Hz, 1H), 7.20 (dd, J = 3.6, 1.2 Hz, 1H), 6.97 (dd, J = 5.4, 3.6 Hz, 1H), 4.16 (q, J = 7.1 Hz, 2H), 3.48 (s, 2H), 1.24 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.5, 135.2, 132.5, 130.6, 127.7, 61.6, 41.1, 14.2. The analytical data are consistent with literature.⁵

CO₂Et

Ethyl 2-(benzo[*d*]thiazol-2-ylthio)acetate (3l): 100.0 mg, 79% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.85 (d, *J* = 8.1 Hz, 1H), 7.75 (dd, *J* = 8.0, 0.5 Hz, 1H), 7.41 (t, *J* = 7.7 Hz, 1H),

7.30 (t, J = 7.6 Hz, 1H), 4.24 (q, J = 7.1 Hz, 2H), 4.17 (s, 2H), 1.29 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 168.4, 164.9, 153.0, 135.6, 126.2, 124.6, 121.8, 121.2, 62.2, 35.3, 14.3. The analytical data are consistent with literature.⁵

Ethyl 2-(benzylthio)acetate (3m): 65.2 mg, 62% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.27 (m, 4H), 7.27 – 7.21 (m, 1H), 4.17 (q, *J* = 7.1 Hz, 2H), 3.82 (s, 2H), 3.05 (s, 2H), 1.28 (t, *J* = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.5, 137.3, 129.3, 128.6, 127.4, 61.4, 36.4, 32.3, 14.3. The analytical data are consistent with literature.¹

(4-chlorobenzyl)(2-(ethylperoxy)- $2\lambda^2$ -ethyl)sulfane (3n): 74.6 mg, 61% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.28 (d, J = 0.8 Hz, 4H), 4.17 (q, J = 7.1 Hz, 2H), 3.79 (s, 2H), 3.04 (s, 2H), 1.28 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.4, 135.9, 133.2, 130.6, 128.8, 61.5, 35.7, 32.3, 14.3. The analytical data are consistent with literature.²

SCO₂Et

Ethyl 2-(cyclohexylthio)acetate (30): 40.5 mg, 40% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 4.18 (q, J = 7.2 Hz, 2H), 3.24 (s, 2H), 2.92 – 2.68 (m, 1H), 2.04 – 1.93 (m, 2H), 1.80 – 1.71 (m, 2H), 1.48 – 1.06 (m, 9H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 171.1, 61.4, 44.1, 33.2, 32.2, 26.1, 25.9, 14.3. The analytical data are consistent with literature.¹

S CO₂Et

Ethyl 2-(octylthio)acetate (3p): 70.9 mg, 61% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 4.18 (q, J = 7.1 Hz, 2H), 3.19 (s, 2H), 2.67 – 2.57 (m, 2H), 1.63 – 1.53 (m, 2H), 1.40 – 1.32 (m, 2H), 1.30 – 1.23 (m, 11H), 0.87 (t, J = 6.8 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.8, 61.4, 33.9, 32.8, 31.9, 29.4, 29.3, 29.1, 28.9, 22.8, 14.3, 14.2. The analytical data are consistent with literature.²



Ethyl (S)-2-((4-chlorophenyl)thio)-2-phenylacetate (3q): 98.2 mg, 64% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.42 (dd, J = 7.7, 1.8 Hz, 2H), 7.36 – 7.32 (m, 1H), 7.31 (m, 3H), 7.28 (m, 1H), 7.25 – 7.20 (m, 2H), 4.86 (s, 1H), 4.21 – 4.07 (m, 2H), 1.18 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.3, 135.5, 134.5, 134.4, 132.2, 129.2, 128.9, 128.7, 128.6, 62.0, 56.6, 14.1. The analytical data are consistent with literature.⁶



Ethyl (S)-2-(4-chlorophenyl)-2-((4-chlorophenyl)thio)acetate (3r): 117.7 mg, 69% yield, colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 7.34 (dd, J = 8.7, 2.1 Hz, 2H), 7.30 (m, 2H), 7.27 (t, J = 3.5 Hz, 2H), 7.24 (dd, J = 8.6, 2.1 Hz, 2H), 4.80 (s, 1H), 4.18 – 4.09 (m, 2H), 1.18 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.9, 134.8, 134.7, 134.5, 134.1, 131.7, 130.0, 129.3, 129.0, 62.2, 55.9, 14.1. HRMS (EI) calcd for C₁₆NH₁₈Cl₂O₂S [M+NH₄]⁺ 358.0430, found 358.0430.



Ethyl (S)-2-(4-bromophenyl)-2-((4-chlorophenyl)thio)acetate (3s): 129.2 mg, 67% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.44 (dd, J = 8.8, 2.2 Hz, 2H), 7.28 (dd, J = 8.6, 2.1 Hz, 4H), 7.23 (dd, J = 8.7, 2.1 Hz, 2H), 4.79 (s, 1H), 4.19 – 4.08 (m, 2H), 1.18 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.8, 134.8, 134.6, 132.0, 131.6, 130.3, 129.3, 122.7, 62.1, 56.0, 14.1. HRMS (EI) calcd for C₁₆H₁₅BrClO₂S [M+H]⁺ 384.9659, found 384.9654.



Ethyl (S)-2-((4-chlorophenyl)thio)-2-(4-methoxyphenyl)acetate (3t): 52.2 mg, 31% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.34 (dd, *J* =8.7, 2.5 Hz, 2H), 7.29 (dd, *J* = 8.8, 2.2 Hz, 2H), 7.23 (dd, *J* = 8.8, 2.2 Hz, 2H), 6.87 – 6.83 (m, 2H), 4.83 (s, 1H), 4.17 – 4.07 (m, 2H), 3.79 (s, 3H), 1.17 (t, *J* = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.4, 159.7, 134.3, 134.2, 132.4, 129.8, 129.2, 127.3, 114.2, 61.9, 55.9, 55.4, 14.1. HRMS (EI) calcd for C₁₇H₁₇ClO₃SNa [M+Na]⁺ 359.0479, found 359.0479.



Ethyl (S)-2-((4-chlorophenyl)thio)-2-(p-tolyl)acetate (3u): 83.4 mg, 52% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.31 (d, J = 8.3, 4H), 7.23 (dd, J = 8.8, 2.3 Hz, 2H), 7.13 (d, J = 7.9 Hz, 2H), 4.84 (s, 1H), 4.12 (m, 2H), 2.33 (s, 3H), 1.18 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.4, 138.4, 134.3, 134.1, 132.5, 132.4, 129.6, 129.2, 128.5, 61.9, 56.3, 21.3, 14.1. HRMS (EI) calcd for C₁₇H₁₇ClO₂SNa [M+Na]⁺ 343.0530, found 343.0532.



Ethyl (S)-2-((4-chlorophenyl)thio)-2-(naphthalen-1-yl)acetate (3v): 35.7 mg, 20% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 8.15 (d, *J* = 8.5 Hz, 1H), 7.89 (d, *J* = 7.9 Hz, 1H), 7.82 (d, *J* = 8.2 Hz, 1H), 7.66 (d, *J* = 7.2 Hz, 1H), 7.58 (ddd, *J* = 8.5, 6.9, 1.5 Hz, 1H), 7.52 (ddd, *J* = 8.0, 6.9, 1.2 Hz, 1H), 7.42 (t, *J* = 7.7 Hz, 1H), 7.30 (dd, *J* = 8.9, 2.3 Hz, 2H), 7.21 (dd, *J* = 8.9, 2.3 Hz, 2H), 5.62 (s, 1H), 4.24 – 4.06 (m, 2H), 1.16 (t, *J* = 7.1 Hz, 3H). ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 170.7, 134.4, 134.3, 134.1, 132.6, 131.1, 131.0, 129.3, 129.2, 129.1, 127.0, 126.9, 126.1,

125.5, 123.2, 62.2, 53.3, 14.2. HRMS (EI) calcd for $C_{20}H_{17}ClO_2SNa$ [M+Na]⁺ 379.0530, found 379.0525.



Methyl (S)-2-((4-chlorophenyl)thio)-2-phenylacetate (3w): 103.9 mg, 71% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.41 (dd, J = 7.7, 1.9 Hz, 2H), 7.34 – 7.27 (m, 5H), 7.23 (dd, J =8.8, 2.2 Hz, 2H), 4.88 (s, 1H), 3.69 (s, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.7, 135.4, 134.5, 134.4, 132.1, 129.2, 128.9, 128.6, 128.5, 56.5, 52.9. The analytical data are consistent with literature.⁷



Methyl (S)-2-(4-chlorophenyl)-2-((4-chlorophenyl)thio)acetate (3x): 99.8 mg, 61% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.34 (dd, J = 8.6, 2.1 Hz, 2H), 7.30 (dd, J = 8.7, 2.6 Hz, 2H), 7.27 (dd, J = 8.7, 2.5 Hz, 2H), 7.24 (dd, J = 8.6, 2.1 Hz, 2H), 4.82 (s, 1H), 3.69 (s, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.3, 134.9, 134.7, 134.5, 134.0, 131.5, 130.0, 129.4, 129.0, 55.8, 53.0. HRMS (EI) calcd for C₁₅H₁₃Cl₂O₂S [M+H]⁺ 327.0008, found 327.0024.



Methyl (S)-2-((4-chlorophenyl)thio)-2-(4-methoxyphenyl)acetate (3y): 90.38 mg, 56% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.40 (dd, J = 9.2, 2.5 Hz, 2H), 7.35 (dd, J = 8.7, 2.2 Hz, 2H), 7.32 – 7.27 (m, 2H), 6.92 (dd, J = 9.3, 2.6 Hz, 2H), 4.91 (s, 1H), 3.87 (s, 3H), 3.75 (s, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 171.0, 159.8, 134.5, 134.4, 132.2, 129.8, 129.3, 127.3, 114.3, 55.9, 55.4, 52.9. The analytical data are consistent with literature.⁸



Tert-butyl 2-((4-chlorophenyl)thio)acetate (3z1): 81.5 mg, 63% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.33 (dd, J = 8.9, 2.3 Hz, 2H), 7.25 (dd, J = 8.9, 2.3 Hz, 2H), 3.52 (s, 2H), 1.40 (s, 9H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 168.6, 133.9, 132.9, 131.3, 129.2, 82.2, 37.9, 28.0. The analytical data are consistent with literature.⁹



Benzyl 2-((4-chlorophenyl)thio)acetate (3z2): 93.69 mg, 64% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.35 (m, 3H), 7.29 (dd, *J* = 9.0, 2.4 Hz, 3H), 7.28 – 7.25 (m, 1H), 7.22 (dd, *J* = 8.9, 2.3 Hz, 2H), 5.13 (s, 2H), 3.64 (s, 2H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.4, 135.3, 133.4, 133.3, 131.8, 129.3, 128.7, 128.6, 128.5, 67.5, 37.0. HRMS (EI) calcd for C₁₅H₁₄ClO₂S [M+H]⁺ 293.0398, found 293.0406.



(**R**)-2-(2-Ethoxy-2-oxo-1-phenylethoxy)benzoic acid (5a): 72.1 mg, 80% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 10.45 (s, 1H), 8.00 (dd, J = 8.0, 1.7 Hz, 1H), 7.59 (dd, J = 7.5, 2.1 Hz, 2H), 7.51 – 7.41 (m, 4H), 7.00 (dd, J = 8.4, 0.8 Hz, 1H), 6.91 (ddd, J = 8.2, 7.2, 1.1 Hz, 1H), 6.15 (s, 1H), 4.24 (m, 2H), 1.24 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.4, 168.4, 161.9, 136.4, 133.6, 130.4, 129.6, 129.0, 127.7, 119.5, 117.7, 111.9, 75.3, 62.1, 14.1. The analytical data are consistent with literature.⁶



(**R**)-2-(2-Ethoxy-2-oxo-1-(**p**-tolyl)ethoxy)benzoic acid (5b): 62.3 mg, 65% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 10.45 (s, 1H), 7.98 (dd, J = 8.0, 1.7 Hz, 1H), 7.51 – 7.47 (m, 1H), 7.46 (d, J = 8.1 Hz, 2H), 7.25 (d, J = 8.4 Hz, 2H), 6.99 (dd, J = 8.4, 0.8 Hz, 1H), 6.90 (ddd, J = 8.2, 7.2, 1.1 Hz, 1H), 6.11 (s, 1H), 4.30 – 4.15 (m, 2H), 2.39 (s, 3H), 1.24 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.5, 168.6, 161.9, 139.7, 136.3, 130.7, 130.5, 129.7, 127.8, 119.5, 117.7, 112.0, 75.2, 62.0, 21.4, 14.1. HRMS (EI) calcd for C₁₈H₁₈O₅Na [M+Na]⁺ 337.1046, found 337.1045.



(**R**)-2-(2-Ethoxy-1-(4-methoxyphenyl)-2-oxoethoxy)benzoic acid (5c): 21.8 mg, 22% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 10.45 (s, 1H), 7.96 (dd, J = 8.0, 1.6 Hz, 1H), 7.49 (d, J = 8.7 Hz, 2H), 7.46 (m, 1H), 6.98 (d, J = 8.5 Hz, 1H), 6.96 (d, J = 8.8 Hz, 2H), 6.89 (t, J = 7.1 Hz, 1H), 6.08 (s, 1H), 4.30 – 4.16 (m, 2H), 3.83 (s, 3H), 1.24 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.5, 168.7, 161.9, 160.6, 136.3, 130.5, 129.3, 125.7, 119.5, 117.7, 114.5, 112.0, 75.0, 62.0, 55.5, 14.2. HRMS (EI) calcd for C₁₈H₁₈O₆Na [M+Na]⁺ 353.0996, found 353.0999.



(**R**)-2-(1-(4-Chlorophenyl)-2-ethoxy-2-oxoethoxy)benzoic acid (5d): 47.2 mg, 47% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 10.38 (s, 1H), 7.96 (dd, J = 8.0, 1.5 Hz, 1H), 7.52 (d, J = 8.5 Hz, 2H), 7.48 (d, J = 8.6 Hz, 1H), 7.42 (d, J = 8.5 Hz, 2H), 7.00 (d, J = 8.4 Hz, 1H), 6.91 (t, J = 7.5 Hz, 1H), 6.11 (s, 1H), 4.23 (m, 2H), 1.24 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.2, 168.1, 162.0, 136.5, 135.7, 132.1, 130.3, 129.3, 129.1, 119.6, 117.8, 111.7, 74.5, 62.3, 14.1. HRMS (EI) calcd for C₁₇H₁₅ClO₅Na [M+Na]⁺ 357.0500, found 357.0503.



(**R**)-2-(1-(4-Bromophenyl)-2-ethoxy-2-oxoethoxy)benzoic acid (5e): 83.0 mg, 73% yield, colorless oil. ¹H NMR (400 MHz, CDCl₃) δ 10.37 (s, 1H), 7.96 (dd, J = 8.0, 1.7 Hz, 1H), 7.57 (dd, J = 8.7, 2.0 Hz, 2H), 7.52 – 7.47 (m, 1H), 7.45 (d, J = 8.5 Hz, 2H), 7.00 (d, J = 8.4 Hz, 1H), 6.92 (ddd, J = 8.1, 7.2, 1.0 Hz, 1H), 6.10 (s, 1H), 4.32 – 4.16 (m, 2H), 1.24 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.2, 168.0, 162.0, 136.6, 132.7, 132.3, 130.3, 129.4, 123.9, 119.6, 117.9, 111.7, 74.6, 62.3, 14.1. HRMS (EI) calcd for C₁₇H₁₅BrO₅Na [M+Na]⁺ 400.9995, found 400.9992.



(**R**)-2-(2-Methoxy-2-oxo-1-phenylethoxy)benzoic acid (5f): 39.5 mg, 46% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 10.43 (s, 1H), 8.00 (dd, J = 8.0, 1.7 Hz, 1H), 7.58 (dd, J = 7.3, 2.2 Hz, 2H), 7.51 – 7.43 (m, 4H), 7.00 (dd, J = 8.4, 0.8 Hz, 1H), 6.91 (ddd, J = 8.1, 7.3, 1.0 Hz, 1H), 6.18 (s, 1H), 3.77 (s, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.4, 168.9, 162.0, 136.4, 133.5, 130.4, 129.7, 129.1, 127.8, 119.5, 117.8, 111.8, 75.1, 53.0. The analytical data are consistent with literature.¹⁰



(R)-2-(2-(Allyloxy)-2-oxo-1-phenylethoxy)benzoic acid (5g): 23.4 mg, 25% yield, colorless oil,
¹H NMR (400 MHz, CDCl₃) δ 10.42 (s, 1H), 7.99 (dd, J = 8.0, 1.7 Hz, 1H), 7.58 (dd, J = 7.3, 2.3 Hz, 2H), 7.49 (ddd, J = 8.8, 7.4, 1.8 Hz, 1H), 7.44 (m, 3H), 7.00 (dd, J = 8.4, 0.8 Hz, 1H), 6.91 (ddd, J = 8.2, 7.3, 1.1 Hz, 1H), 6.19 (s, 1H), 5.85 (ddt, J = 17.2, 10.6, 5.6 Hz, 1H), 5.23 (ddd, J = 17.2, 10.6, 5.6 Hz), 5.23 (ddd, J = 17.2, 10.6,

17.1, 2.9, 1.5 Hz, 1H), 5.20 (ddd, J = 10.1, 2.4, 1.4 Hz, 1H), 4.76 – 4.50 (m, 2H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.4, 168.2, 162.0, 136.5, 133.5, 131.3, 130.4, 129.7, 129.1, 127.8, 119.5, 118.8, 117.8, 111.9, 75.2, 66.4. HRMS (EI) calcd for C₁₈H₁₆O₅Na [M+Na]⁺ 335.0890, found 335.0890.

COOH O CO₂Et

(**R**)-2-(2-Ethoxy-2-oxo-1-phenylethoxy)-4-methylbenzoic acid (5h): 33.9 mg, 36% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 10.38 (s, 1H), 7.85 (d, J = 8.1 Hz, 1H), 7.57 (dd, J = 7.3, 2.1 Hz, 2H), 7.47 – 7.40 (m, 3H), 6.80 (s, 1H), 6.72 (dd, J = 8.1, 1.0 Hz, 1H), 6.12 (s, 1H), 4.29 – 4.16 (m, 2H), 2.35 (s, 3H), 1.23 (t, J = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 169.4, 168.6, 162.0, 147.9, 133.8, 130.2, 129.5, 129.0, 127.8, 120.8, 117.9, 109.3, 75.1, 62.1, 22.1, 14.1. HRMS (EI) calcd for C₁₈H₁₈O₅Na [M+Na]⁺ 337.1046, found 337.1045.



(**R**)-4-Chloro-2-(2-ethoxy-2-oxo-1-phenylethoxy)benzoic acid (5i): 30.1 mg, 30% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 10.53 (s, 1H), 7.90 (d, *J* = 8.6 Hz, 1H), 7.59 – 7.51 (m, 2H), 7.46 – 7.37 (m, 3H), 7.02 (d, *J* = 2.0 Hz, 1H), 6.89 (dd, *J* = 8.6, 2.0 Hz, 1H), 6.13 (s, 1H), 4.30 – 4.15 (m, 2H), 1.24 (t, *J* = 7.1 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 168.9, 168.3, 162.5, 142.3, 133.4, 131.5, 129.7, 129.1, 127.8, 120.3, 118.0, 110.6, 75.5, 62.2, 14.1. HRMS (EI) calcd for C₁₇H₁₆ClO₅ [M+H]⁺ 335.0681, found 335.0682.



1-(((4-Chlorophenyl)thio)oxy)-2,2,6,6-tetramethylpiperidine (6a): 27.0 mg, 18% yield, colorless oil, ¹H NMR (400 MHz, CDCl₃) δ 7.63 – 7.57 (m, 2H), 7.44 – 7.37 (m, 2H), 1.65 (s, 6H), S25

1.57 - 1.35 (m, 9H), 0.90 (s, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 149.0, 135.8, 128.9, 127.6, 61.6, 59.1, 43.6, 41.5, 35.5, 32.9, 28.9, 28.0, 17.3. HRMS (EI) calcd for C₁₅H₂₃ClNOS [M+H]⁺ 300.1183, found 300.1186.

4. References

- 1 A. Röske, I. Alt, B. Plietker, ChemCatChem., 2019, 11, 5260-5263.
- 2 V. Tyagi,; R.-B. Bonn,; R. Fasan, Chem. Sci., 2015, 6, 2488-2494.
- 3 L.-H. Zou, C. Zhao, P.-G. Li, Y. Wang, J. Li, J. Org. Chem., 2017, 82, 12892-12898.
- 4 T. Nakazawa, N. Hirose, K. Itabashi, Synthesis, 1989, 12, 955-957.
- 5 J.-Y. Yang, G.-G. Wang, S.-W. Chen, B. Ma, H.-Y. Zhou, M.-H. Song, C. Liu, C.-D. Huo, Org. Biomol. Chem., 2020, 18, 9494-9498.
- 6 Z.-P. Zhang, Z.-Q. He, Y.-X. Xie, T.-T. He, Y.-F. Fu, Y. Yu and F. Huang, *Org. Chem. Front.*, 2021, **8**, 1233-1242.
- 7 H. Keipour, A. Jalba, L. Delage-Laurin,; T. Ollevier, J. Org. Chem., 2017, 82, 3000-3010.
- 8 P. Saha, H. Jeon, P.-K. Mishra, H. Rhee, J.-H. Kwak, J. Mol. Catal. A-Chem., 2016, 417, 10-18.
- 9 L.-L Zong, C. Wang, A.-M.-P. Moeljadi, X.-Y. Ye, R. Ganguly, Y.-X. Li, H. Hirao, C.-H. Tan, *Nat. Commun.*, 2016, 7, 13455.
- 10 Z.-P. Zhang, Y. Yu, F. Huang, X.-Y. Yi, Y. Xu, Y.-D. He, J.-B. Baell, H. Huang, *Green Chem.*, 2020, **22**, 1594-1604.
- 11 T. Wirtanen, T. Prenzel, J.-P. Tessonnier, S.-R. Waldvogel, *Chem. Rev.*, 2021, **121**, 10241-10270.
- 12 C. Empel, S. Jana, C. Pei, T.-V. Nguyen, R.-M. Koenigs, Org. Lett., 2020, 22, 7225-7229.

5. Copies of NMR spectra











































210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)





























210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)



rr (ppm)



