

Supplementary material

for

pH-controlled regioselective nucleophilic ring opening of epoxide: Improved process for the preparation of (*R*)-(-)- or (*S*)-(+)-3-hydroxytetrahydrofuran.

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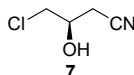
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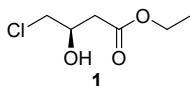
1. General information

^1H NMR spectra were determined in CDCl_3 using a Bruker AC 400 spectrometer at operating at 400 MHz, using TMS as an internal standard. Splitting patterns are as follows: *s*, singlet; *d*, doublet; *m*, multiplet; *br*, broad signal. The ^{13}C NMR spectrum was obtained using the same apparatus described above at 100 MHz. Epichlorohydrin was purchased from Sigma-Aldrich and was utilized as the chromatographic standard in TLC analysis. The progress of the reactions was monitored by TLC and GC. The chromatograms were visualized using a solution of KMnO_4 in water.

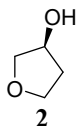
2. Characterisation data of synthesized compounds



4-chloro-3-hydroxybutanenitrile (7): Pale yellowish oil, IR (neat): ν 3422, 2963, 2257, 1632, 1419, 1302, 1088 cm^{-1} ; ^1H NMR (400MHz, CDCl_3): δ 4.24-4.17 (m, 1H), 3.71- 3.63 (m, 2H), 3.10 (d, $J=$ 4Hz, 1H), 2.78-2.67 (m, 2H), ^{13}C NMR (100MHz, CDCl_3): δ 117.1, 67.4, 47.3, 23.3.

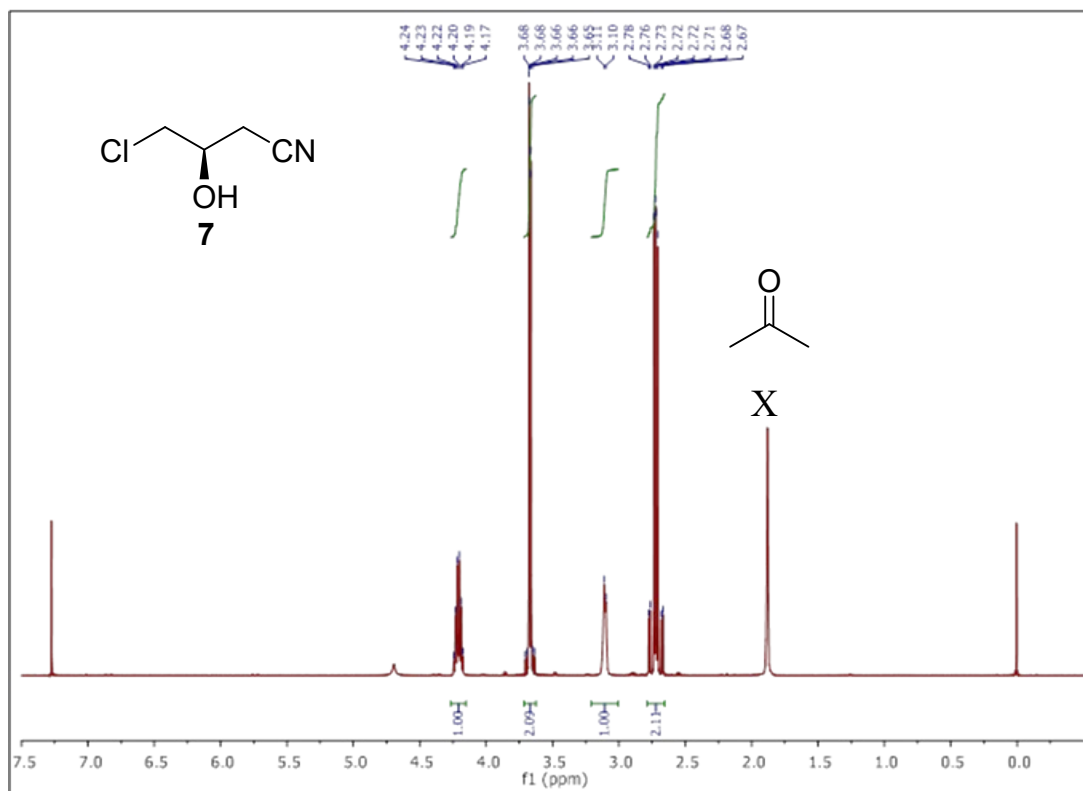


Ethyl- (R)-4-chloro-3-hydroxybutanoate (1) Yellowish oil, IR (neat): ν 3449, 2983, 1725, 1637, 1380, 1192, 1031 cm^{-1} ; ^1H NMR (400MHz, CDCl_3): δ 4.27-4.22 (m, 1H), 4.17 (q, $J=$ 8Hz, 2H), 3.63-3.56 (m, 2H), 3.17 (d, $J=$ 4 Hz, 1H), 2.66-2.57 (m, 2H), 1.26 (t, $J=$ 4Hz, 3H), ^{13}C NMR (100MHz, CDCl_3): δ 171.8, 68.0, 61.1, 48.2, 38.6, 14.2, MS (ESI) m/z ($\text{M}+\text{Na}$) $^+$ calculated for $\text{C}_6\text{H}_{11}\text{ClO}_3\text{Na}$: 189, observed: 189.

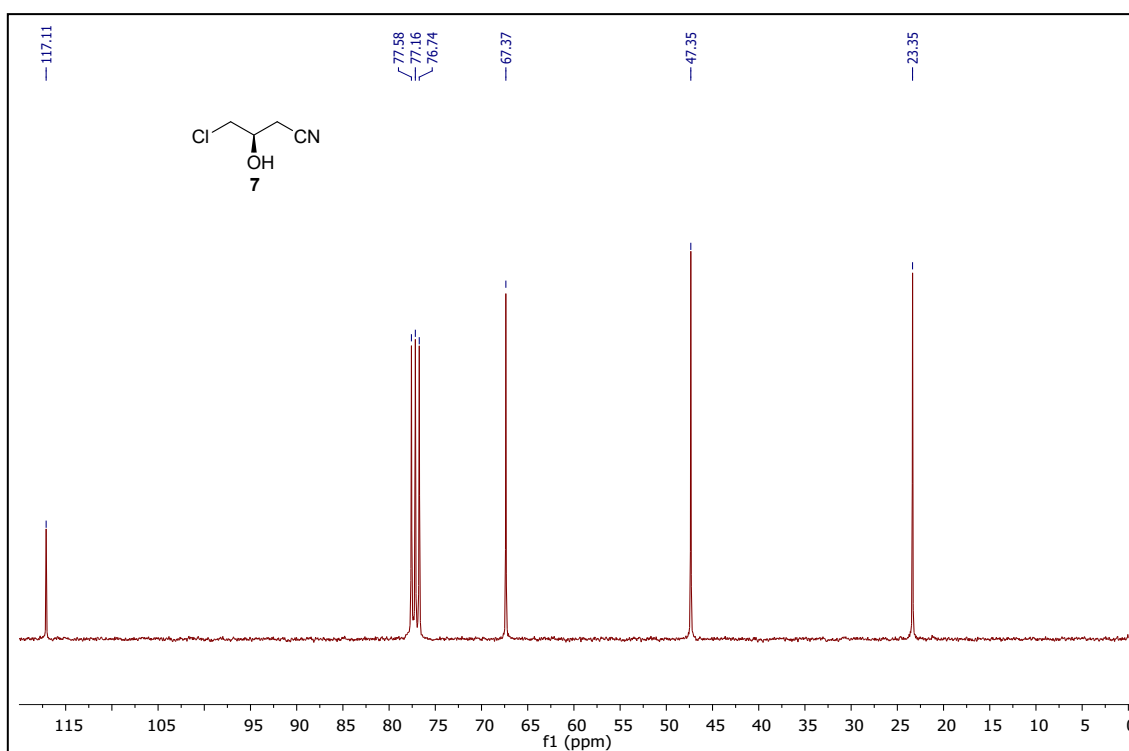


(R)-3-hydroxytetrahydrofuran (2): Colourless oil, IR (neat): ν 3410, 2952, 2882, 1764, 1647, 1054 cm^{-1} , ^1H NMR (400MHz, CDCl_3): 4.52-4.49 (m, 1H), 4.0- 3.95(m, 1H), 3.87-3.74 (m, 3H), 2.16-2.04 (m, 1H), 1.95-1.87 (m, 1H), ^{13}C NMR (100MHz, CDCl_3): 75.8, 72.3, 66.8, 35.8. $[\alpha]_{\text{D}}^{23.9} = -16.99$ ($c=$ 2.45, CH_3OH), lit¹⁸ $[\alpha]_{\text{D}}^{23} = -17.3$ ($c=$ 2.4, CH_3OH).

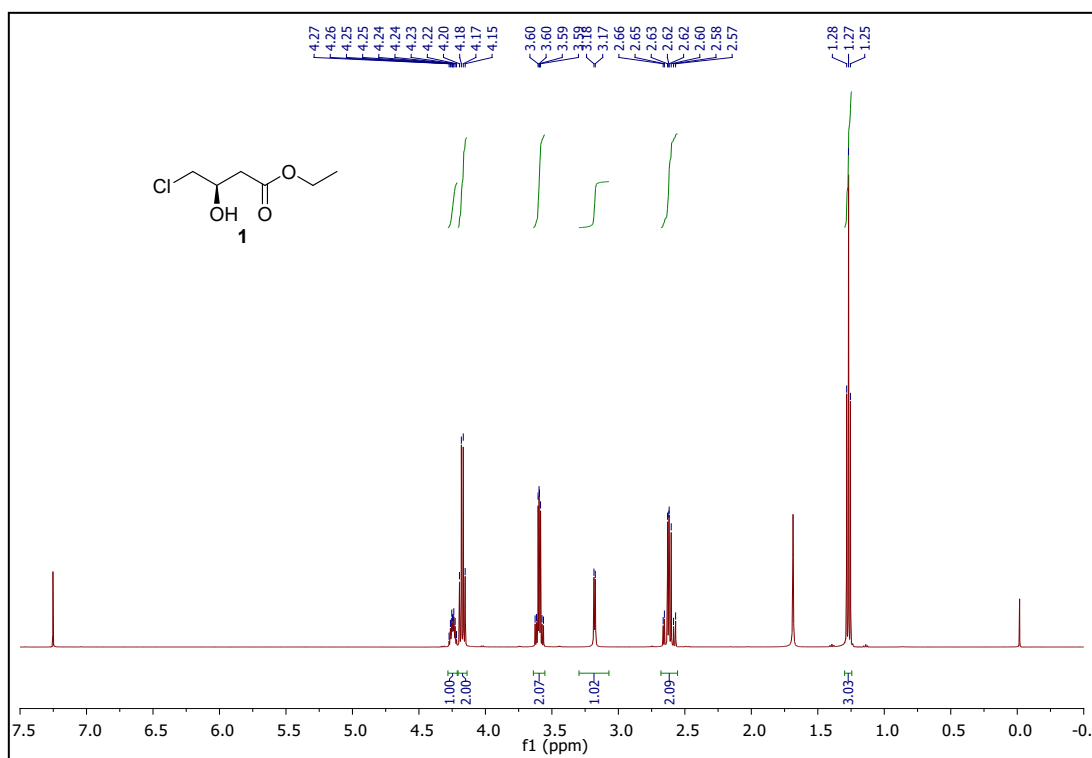
¹H NMR of 7 (400 MHz, CDCl₃)



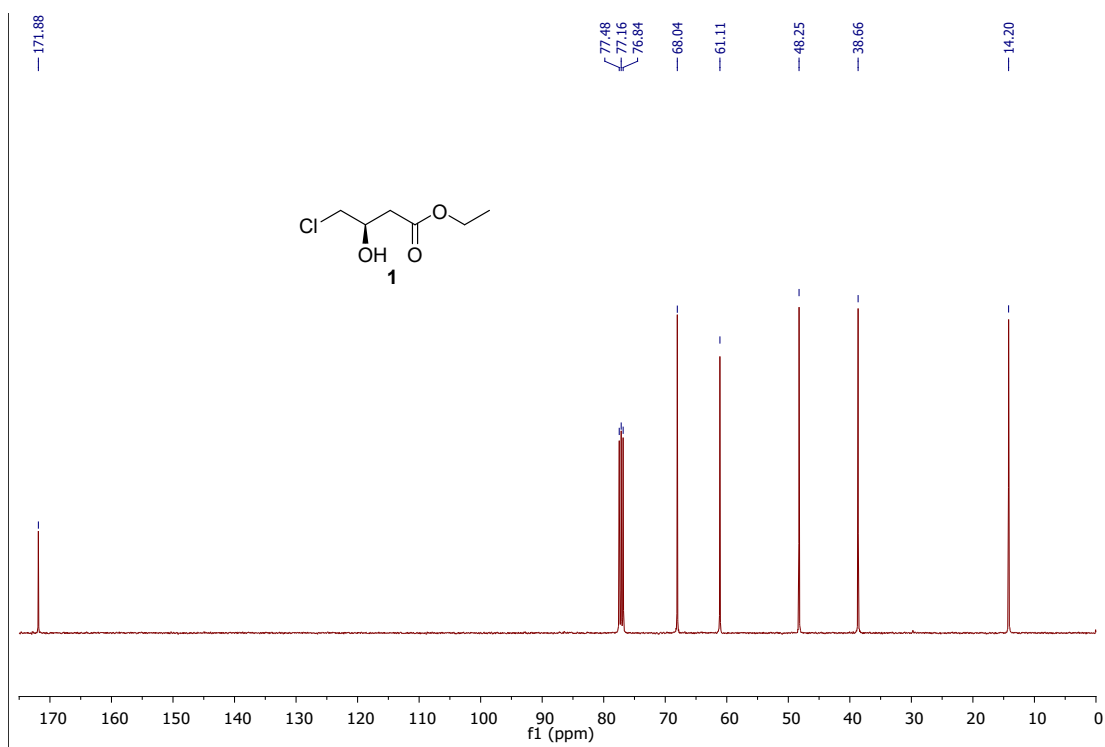
¹³C NMR of 7 (100 MHz, CDCl₃)



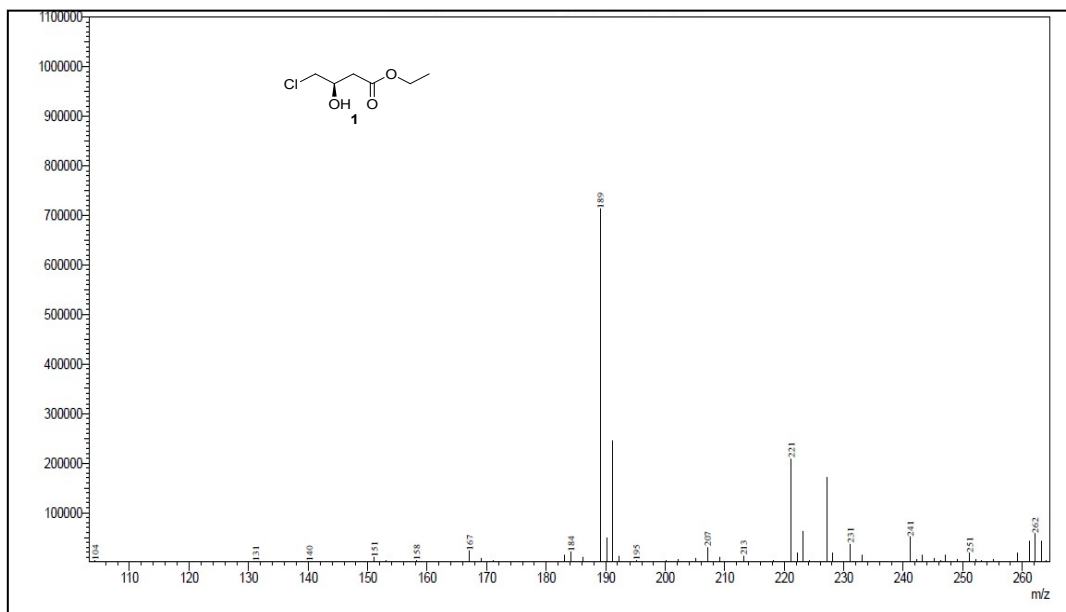
¹H NMR of 1 (400MHz, CDCl₃)



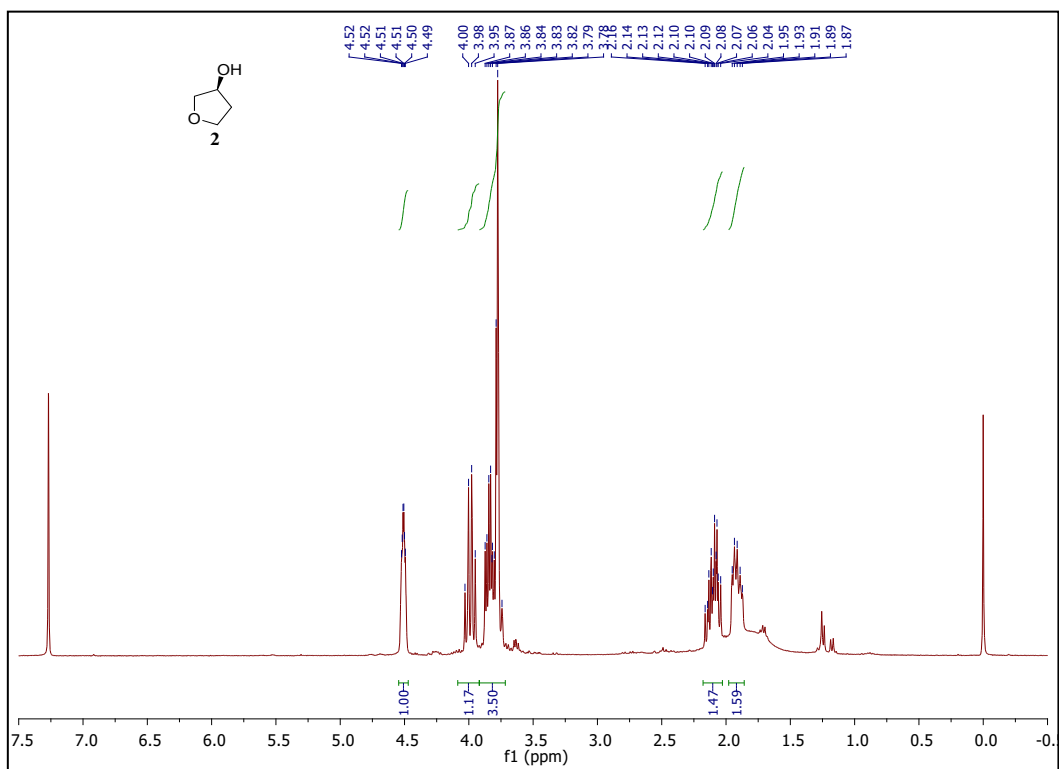
¹³C NMR of 1 (100 MHz, CDCl₃)



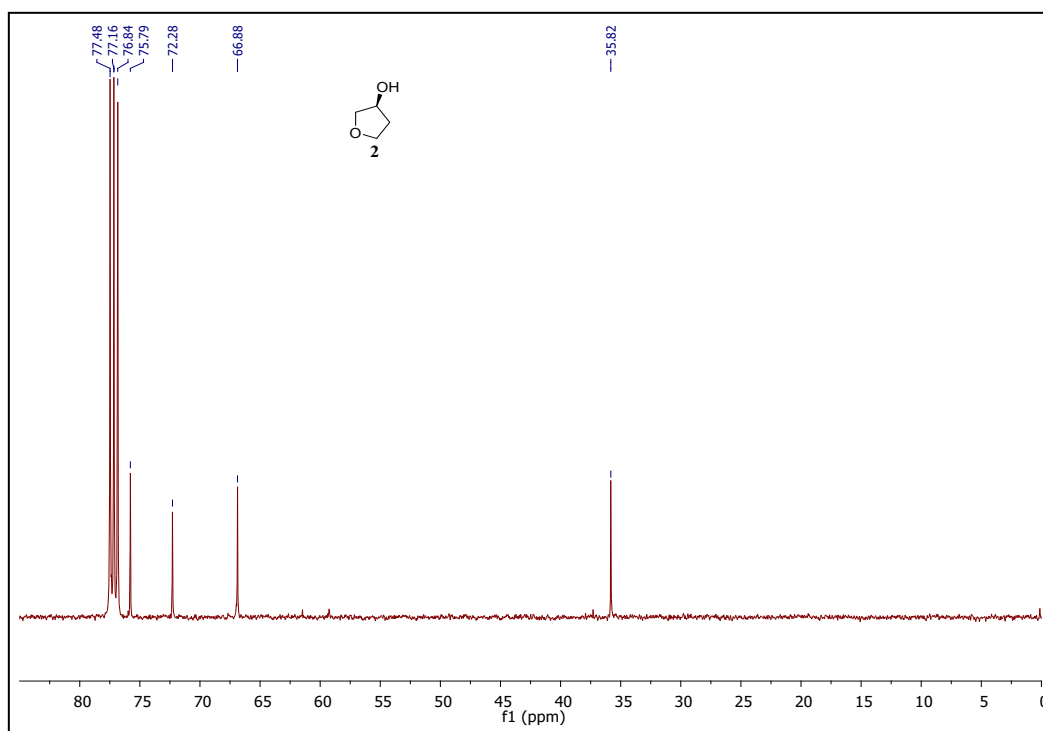
Mass spectra (ESI) of 1



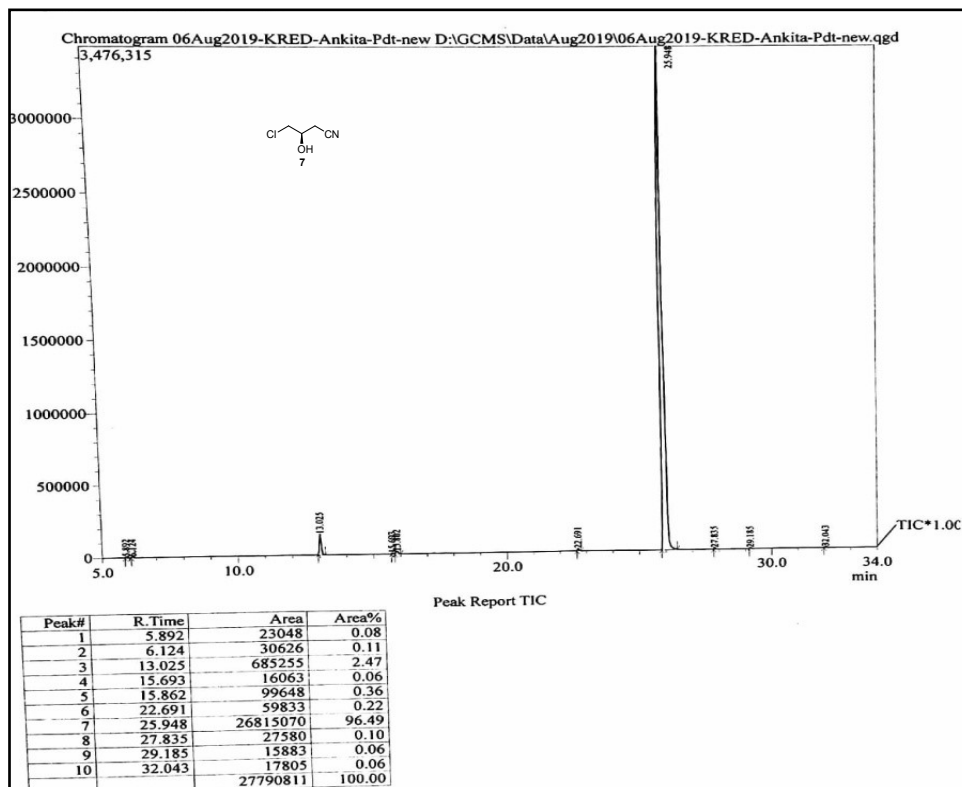
¹H NMR of 2 (400MHz, CDCl₃)

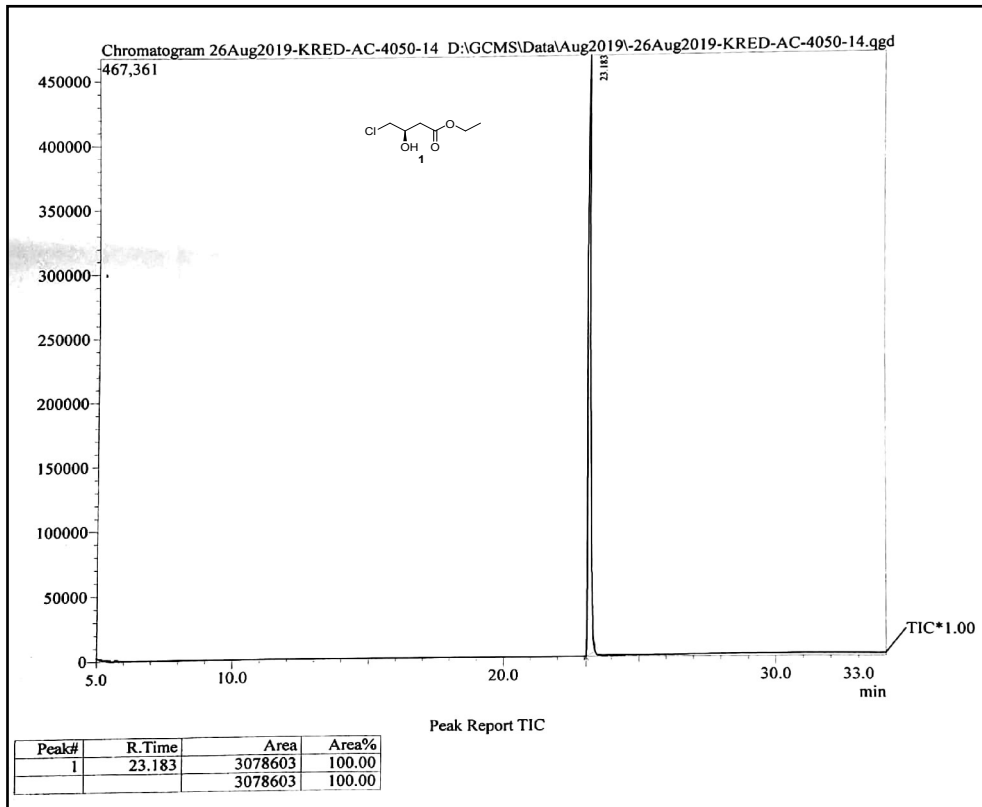
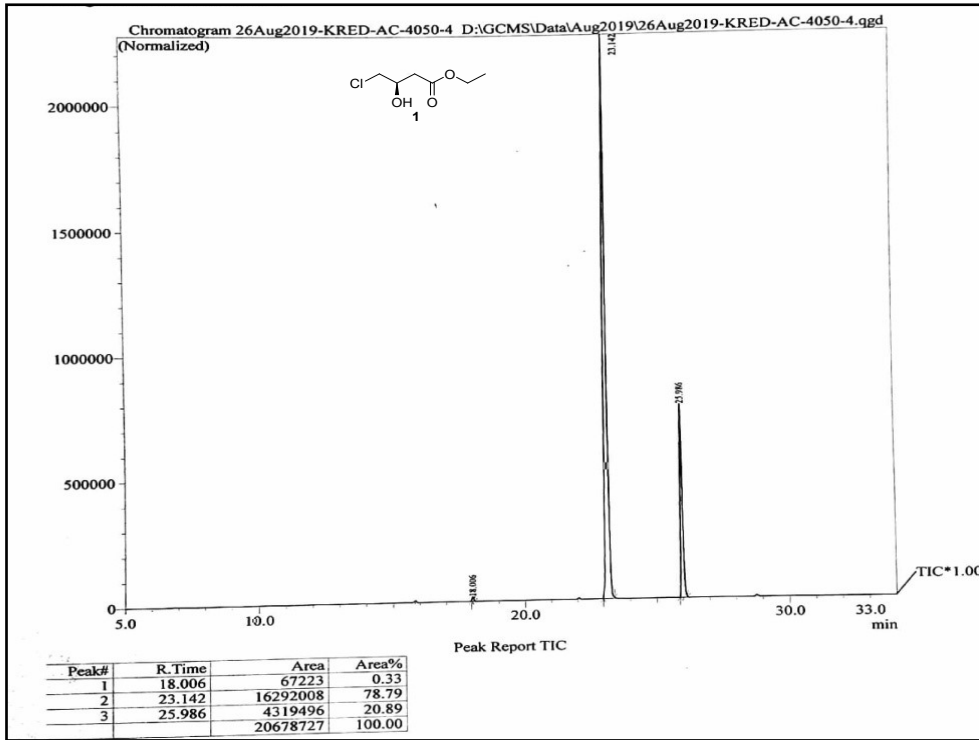


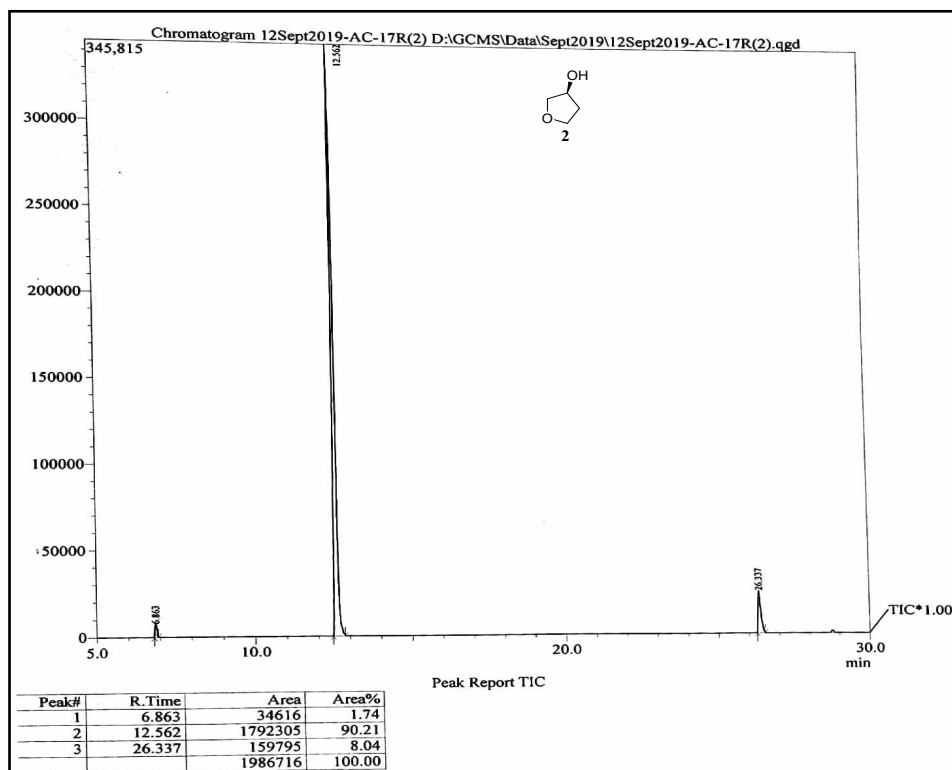
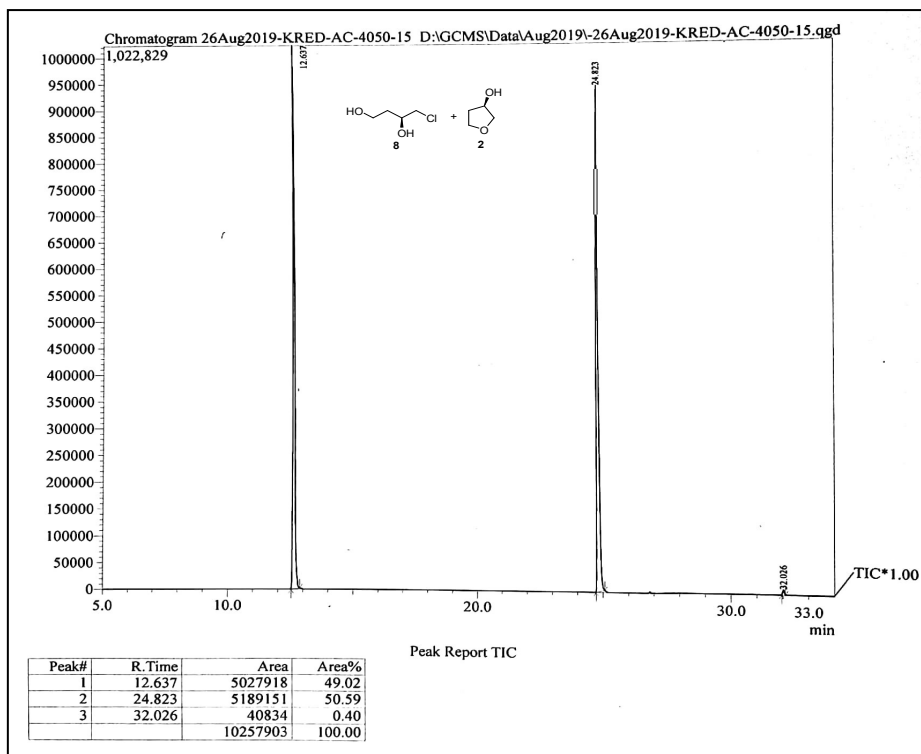
^{13}C NMR of 2 (100 MHz, CDCl_3)



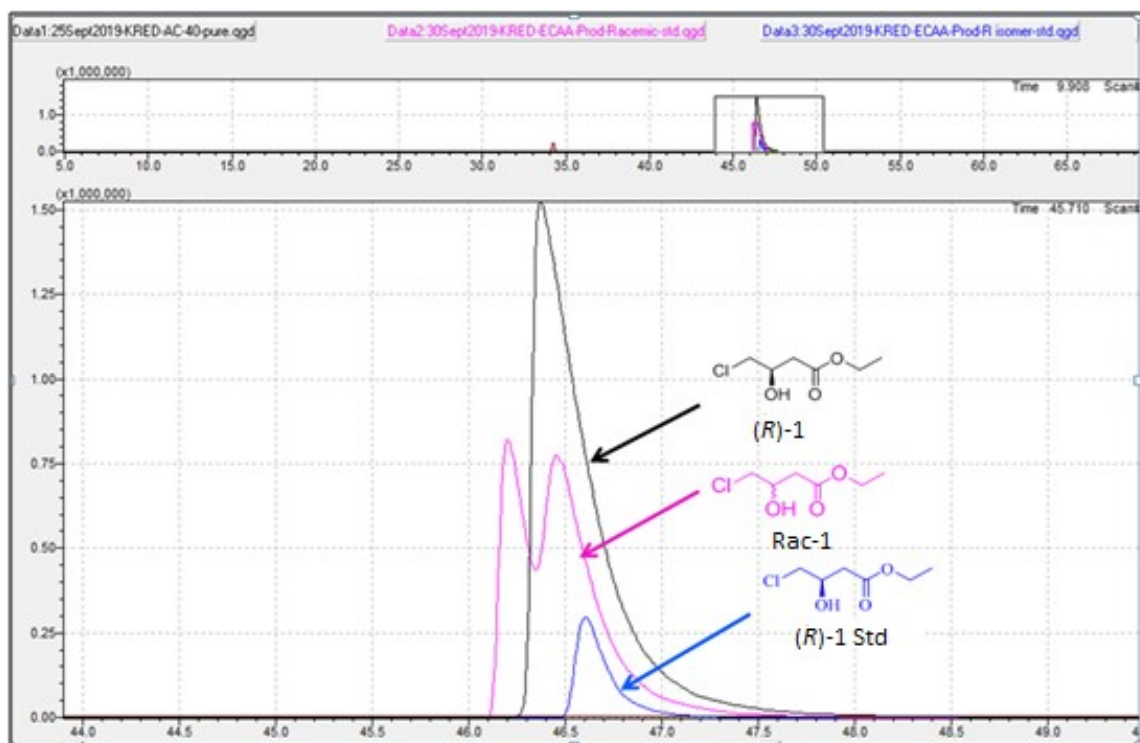
GC CHROMATOGRAMS







Comparison of (*R*)-Ethyl-4-chloro-3-hydroxybutanoate (1) with standard sample.



Overlap of all products in GC:

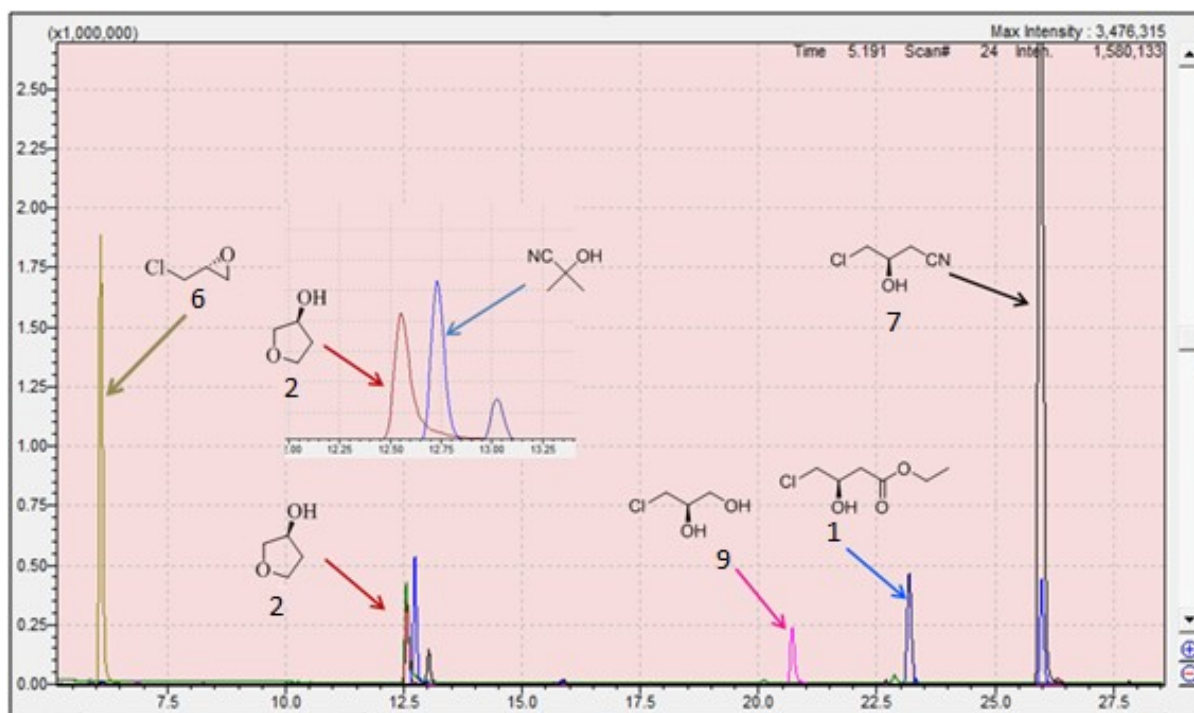


Table S1. Comparison of present work with reported methods

S.No	Reaction conditions	Remarks	References
1.	HCN	HCN is toxic.	1
2.	NaCN, Acetic acid, pH 8 – 10.	Using salts.	2
3.	Cyanide salt, inorganic acid. pH 8-10.	Simultaneous adjustment of pH. Racemization issues.	3
4.	NaCN, inorganic acid, pH 7 – 8.	No simultaneous pH adjustment	4
5.	NaCN, Citric acid, pH 7.8 – 8.3	Replaced H ₂ SO ₄ .	5
6.	HCN (aq. solution), H ₂ SO ₄ , pH 8-10 & enzymatic resolution of ester.	Process for L-Carnitine and acetyl L-Carnitine Hydrochloride.	6
7.	Liq. HCN, several days in sealed container.	Highly toxic to handle	7
8.	HCN, KCN (cat).	Highly toxic to handle	8
9.	Aq. solution of HCN, KCN, Acetic acid.	-	9
10	Ce(OTf) ₄ , 10 mole% NaCN, Neat	6h, 85 % yield.	10
11	Ce(OTf) ₄ , NaCN, Micellar media, rt	7 h, 85 % yield.	11
12	i). KCN, cat. TBAB, 1N KOH/DCE. ii). Actone cyanohydrin, Et ₃ N, reflux.	Product extraction issues in the biphasic system. 4 h, 67 % yield.	12
13	i). Acetone cyanohydrin, 0.1 M KPB, pH 7.5–8.5 ii). Water: DCM (1:1), TBAB (0.1% w/w). iii). Acetone cyanohydrins, 0.1 eq K ₂ CO ₃ .	12h, 93% yield & 94.5% GC purity. 24h, 88% yield & 83.28% GC purity 24h, 90% yield & > 99% GC purity	Present study

References.

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