

Supporting Information

***De novo* Synthesis of Dimerization-Ready Flavan Unit via Intramolecular Pummerer/Friedel–Crafts Cascade**

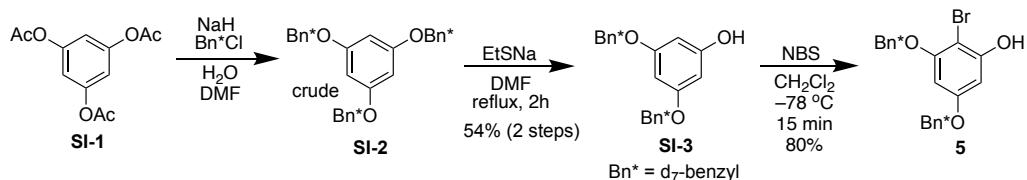
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General

All reactions utilizing air- or moisture-sensitive reagents were performed in flame-dried glasswares under an atmosphere of dry argon. Ethereal solvents and dichloromethane (anhydrous; Kanto Chemical Co., Inc.) were purified under argon, using an Organic Solvent Pure Unit (Wako Pure Chemical Industries, Ltd.). For thin-layer chromatography (TLC) analysis, Merck pre-coated plates (TLC silica gel 60 F254, Art 5715, 0.25 mm) were used. Silica-gel preparative thin-layer chromatography (PTLC) was performed using plates prepared from Merck Silica gel 60 PF254 (Art 7747). For flash column chromatography, silica gel 60N (Spherical, neutral, 63–210 µm) from Kanto Chemical was used. Melting point (mp) determinations were performed by using a METTLER TOLEDO MP70 melting point system and are uncorrected. ¹H-, and ¹³C-NMR were measured on a Bruker Avance III (600 MHz) spectrometer equipped with the cold probe (CryoProbe ProdigyTM) and in the solvent indicated; Chemical shifts (*d*) are expressed in parts per million (ppm) downfield from internal standard (tetramethylsilane 0.00 ppm) or referenced to residual undeuterated solvents as internal standard. All coupling constants (*J*) are reported as hertz (Hz). Splitting patterns are indicated as follows: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad. Infrared (IR) spectra were recorded on a Thermo SCIENTIFIC NICOLET iS5 FT-IR spectrometer. Attenuated total reflectance Fourier transform infrared (ATR-FTIR) spectra were recorded by using Thermo SCIENTIFIC NICOLET iS5 FTIR spectrometer equipped iD5 ATR accessory. High-resolution mass spectra (HRMS) were obtained with Bruker Daltonics micrOTOF-Q II. Optical rotations ([α]_D) were measured on a JASCO P-3000 polarimeter. High performance liquid chromatography (HPLC) analyses were performed on a LC-Net II/ADC controller (JASCO) equipped with a Jasco PU-2080 Plus Intelligent Pump, a Jasco MD-2010 Plus Multiwavelength Detector, a Jasco DG-2080-54 degasser and LG-2080-02 Ternary Gradient Unit. Preparative HPLC separation was performed on a LC-Net II/ADC controller (JASCO) equipped with a Jasco PU-2086 Plus Intelligent Prep Pump, a Jasco UV-1575 UV/Vis Detector and a Jasco DG-2080-54 degasser.

Preparation of bromophenol 5

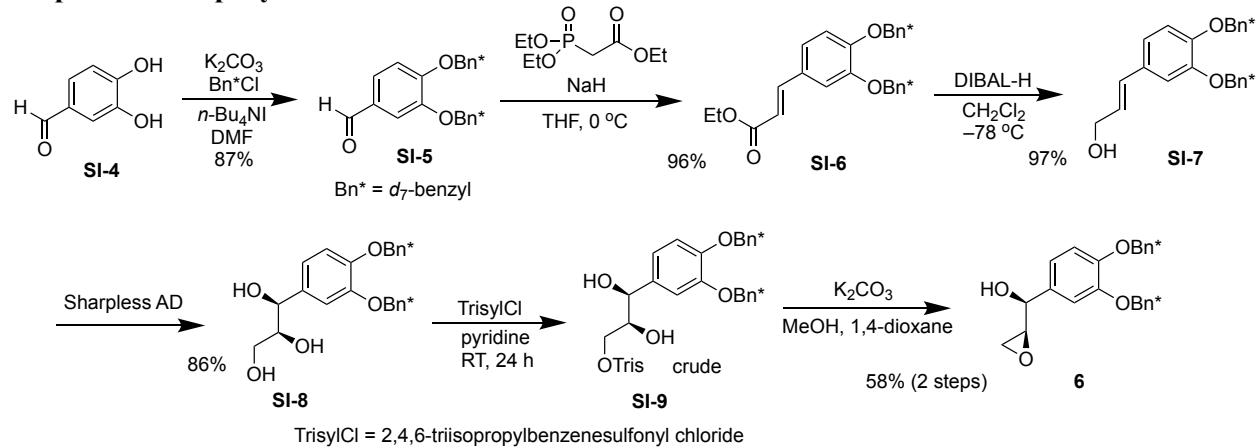


Phenol SI-3^{1,2}: To a suspension of NaH (1.1 g, 63% dispersion in mineral oil, 29 mmol) in DMF (10 mL) at 0 °C was added a solution of 1,3,5-triacetoxy benzene **SI-1** (1.0 g, 4.0 mmol) in DMF (10 mL), tetrabutyl ammonium iodide (TBAI) (0.4 g, 1 mmol) and stirred for 10 min. Bn^*Cl (*d*₇-benzyl chloride, 1.5 mL, 13 mmol) and H_2O (215 μ L, 11.9 mmol) was successively added and stirred at 15 °C for 10 h. The reaction was quenched at 0 °C by adding Et₂NH followed by H_2O , and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with H_2O , brine, dried (Na_2SO_4) and concentrated in vacuo to afford crude 1,3,5-tribenzyloxy benzene **SI-2** (1.5 g) which was used for the next reaction without further purification. At 0 °C, ethanethiol (1.1 mL, 15 mmol) in DMF (5 mL) was added to a suspension of NaH (0.25 g, 63 % dispersion in mineral oil, 6.6 mmol) in DMF (10 mL) over a period of 10 min. After 30 min, solution of crude **SI-2** (1.5 g) in DMF (10 mL) was added over a period of 5 min. The reaction mixture was stirred at 150 °C for 2 h. The reaction was quenched at 0 °C by adding H_2O and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with H_2O , brine, dried (Na_2SO_4) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, EtOAc/hexane = 1/7) to afford phenol **SI-3** (0.68 g, 54%, 2 steps) as a white solid: R_f = 0.55 (EtOAc/hexane = 3/1); mp = 93–94 °C; ¹H NMR (600 MHz, CDCl₃) δ 4.80 (s, 1H), 6.10 (d, J = 1.8 Hz, 2H), 6.24 (t, J = 1.8 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 69.6 (quint, J_{C-d} = 22.5 Hz), 95.1, 95.5, 127.3 (t, J_{C-d} = 24.0 Hz), 127.7 (t, J_{C-d} = 24.0 Hz), 128.3 (t, J_{C-d} = 24.0 Hz), 136.6, 157.4, 161.0; IR (neat) 3295, 1608, 1591, 1503, 1360, 1203, 1165, 1148, 1026, 972, 819 cm⁻¹; HRMS (ESI) calcd for C₂₀H₂₅D₁₄O₃ [(M+H)⁺] *m/z* 321.2208, found *m/z* 321.2211.

Bromophenol 5³: To a solution of phenol **SI-3** (0.51 g, 1.6 mmol) in CH_2Cl_2 (30 mL) at –78 °C was added *N*-bromosuccinimide (0.30 g, 1.7 mmol) and stirred for 15 min. The reaction was quenched by adding 10% aqueous K_2CO_3 solution and the mixture was extracted with CH_2Cl_2 (x3). The combined organic extracts were washed with H_2O , brine, dried (Na_2SO_4) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, EtOAc/hexane = 1/7) to afford bromophenol **5** (0.51 g, 80%) as a white solid. R_f = 0.65 (EtOAc/Hexane = 2/1); mp = 81–82°C; ¹H NMR (600 MHz, CDCl₃) δ 5.67 (s), 6.24 (d, J = 3.0 Hz, 1H), 6.35 (d, J = 3.0 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 69.8 (quint, J_{C-d} = 22.5 Hz), 70.3 (quint, J_{C-d} = 22.5 Hz), 92.2, 94.7, 94.9, 126.8 (t, J_{C-d} = 24.0 Hz), 127.3 (t, J_{C-d} = 24.0 Hz), 127.7 (t, J_{C-d} =

24.0 Hz), 127.8 (t, J_{C-d} = 24.0 Hz), 128.26 (t, J_{C-d} = 24.0 Hz), 128.32 (t, J_{C-d} = 24.0 Hz), 136.2, 136.3, 154.1, 156.1, 159.9; IR (neat) 3481, 1580, 1493, 1479, 1370, 1200, 1188, 1105, 1009, 975, 800, 654, 632 cm⁻¹; HRMS (ESI) calcd for C₂₀H₄D₁₄BrO₃ [(M+H)⁺] *m/z* 401.1293, found *m/z* 401.1276.

Preparation of epoxy alcohol **6**⁴



Aldehyde SI-5: To a solution of aldehyde **SI-4** (5.0 g, 36 mmol) in DMF (50 mL) was added K₂CO₃ (20.0 g, 147 mmol), Bn*Cl (*d*₇-benzyl chloride, 14.0 mL, 116 mmol), tetrabutylammonium iodide (2.7 g, 7.2 mmol) and stirred at 70 °C for 3 h. It was cooled to room temperature, quenched by adding Et₂NH followed by H₂O and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed successively with aqueous HCl (1 M), H₂O, brine and dried (Na₂SO₄) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, EtOAc/hexane = 1/1) to afford **SI-5** (10.5 g, 87%) as a white solid. R_f = 0.65 (EtOAc/Hexane = 1/1); mp = 92–93 °C; ¹H NMR (600 MHz, CDCl₃) δ 7.02 (d, J = 8.4 Hz, 1H), 7.41 (dd, J = 1.8, 8.4 Hz, 1H), 7.48 (d, J = 1.8 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 70.0–70.1 (m), 112.6, 113.3, 126.8 (t, J_{C-d} = 24.0 Hz), 127.1 (t, J_{C-d} = 24.0 Hz), 127.5 (t, J_{C-d} = 24.0 Hz), 128.3 (t, J_{C-d} = 24.0 Hz), 128.4 (t, J_{C-d} = 24.0 Hz), 136.1, 136.4, 149.4, 154.5, 191.0; IR (neat) 1677, 1596, 1598, 1350, 1278, 1169, 1182, 1137, 1077, 982, 953, 922, 871, 750, 725, 630 cm⁻¹; HRMS (ESI) calcd for C₂₁H₅D₁₄O₃ [(M+H)⁺] *m/z* 333.2207, found *m/z* 333.2214.

Ester SI-6: To a suspension of NaH (0.29 g, 63% dispersion in mineral oil, 7.6 mmol) in THF (20 mL) at 0 °C was added triethyl phosphonoacetate (1.4 mL, 7.1 mmol) in THF (5 mL) and stirred for 1 h. A solution of **SI-5** (2.0 g, 6.0 mmol) in THF (10 mL) was added dropwise to the mixture, and the reaction mixture was additionally stirred for 1 h. The reaction was quenched by adding H₂O, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with H₂O, brine, dried (Na₂SO₄) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, EtOAc/hexane = 1/1) to afford **SI-6** (2.3 g, 96%) as a white solid. R_f = 0.55 (EtOAc/Hexane = 2/1); mp = 80–81 °C; ¹H NMR (600

MHz, CDCl₃) δ 1.32 (t, *J* = 7.2 Hz, 1H), 4.24 (q, *J* = 7.2 Hz, 1H), 6.22 (d, *J* = 16.2 Hz, 1H), 6.90 (d, *J* = 8.4 Hz, 1H), 7.06 (dd, *J* = 2.4, 8.4 Hz, 1H), 7.12 (d, *J* = 1.8 Hz, 1H), 7.56 (d, *J* = 16.2 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 14.5, 60.6, 70.0–70.2 (m), 113.9, 114.5, 116.4, 123.0, 126.9 (t, *J*_{C-d} = 22.5 Hz), 127.1 (t, *J*_{C-d} = 24.0 Hz), 127.6 (t, *J*_{C-d} = 24.0 Hz), 128.2 (t, *J*_{C-d} = 24.0 Hz), 136.7, 136.8, 144.5, 149.1, 151.2, 167.4; IR (neat) 2982, 2184, 1697, 1684, 1594, 1427, 1394, 1367, 1049, 1035, 1022, 991, 835, 820, 800, 751, 732, 648 cm⁻¹; HRMS (ESI) calcd for C₂₅H₁₀D₁₄O₄Na [(M+Na)⁺] *m/z* 425.2445, found *m/z* 425.2445.

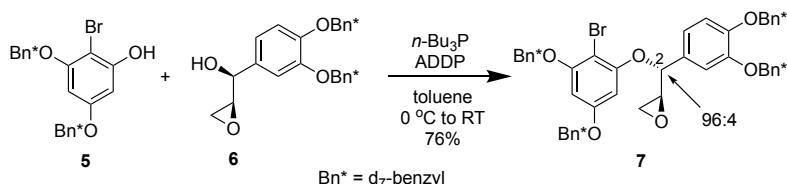
Alcohol SI-7: To a solution of **SI-6** (2.0 g, 5.0 mmol) in THF (25 mL) at -78 °C was added in DIBAL-H (1.0 M in toluene, 13 mL, 13 mmol) and stirred for 2 h. Reaction was quenched by slowly adding saturated solution of Rochelle salt. It was additionally stirred for 2 h and layers were separated. The aqueous layer was further extracted with EtOAc (x3). The combined organic extracts were washed with H₂O, brine, dried (Na₂SO₄) and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, EtOAc/hexane = 1/1) to afford **SI-7** (1.73 g, 97%) as a white solid. *R*_f = 0.45 (EtOAc/hexane = 1/1); mp = 76–77 °C; ¹H NMR (600 MHz, CDCl₃) δ 1.41 (brm, 1H), 4.28 (d, *J* = 4.8 Hz, 1H), 6.18 (dt, *J* = 4.8, 15.6 Hz, 1H), 6.49 (d, *J* = 15.6 Hz, 1H), 6.87 (d, *J* = 8.4 Hz, 1H), 6.89 (dd, *J* = 1.8, 8.4 Hz, 1H), 7.01 (d, *J* = 1.8 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 64.0, 70.1–70.2 (m), 113.1, 115.1, 120.5, 126.9, 127.05 (t, *J*_{C-d} = 24.0 Hz), 127.11 (t, *J*_{C-d} = 24.0 Hz), 127.4–127.6 (m), 128.16 (t, *J*_{C-d} = 24.0 Hz), 128.17 (t, *J*_{C-d} = 24.0 Hz), 130.6, 131.2, 137.08, 137.11, 149.0, 149.2; IR (neat) 3357, 2192, 1507, 1461, 1425, 1271, 1238, 1137, 1080, 1053, 1034, 956, 840, 819, 775 cm⁻¹; HRMS (ESI) calcd for C₂₃H₈D₁₄O₃Na [(M+Na)⁺] *m/z* 383.2340, found *m/z* 383.2332.

Triol SI-8: To a solution of potassium hexacyanoferrate(III) (5.7 g, 17 mmol) in the mixed solvent (140 mL, *t*-BuOH/H₂O = 1/1), was added K₂CO₃ (2.4 g, 17 mmol), methanesulfonamide (0.55 g, 0.58 mmol), (DHQ)₂PHAL (0.23 g, 0.29 mmol), and K₂OsO₂(OH)₄ (22 mg, 0.059 mmol) at 0 °C and stirred for 30 min. To this, **SI-7** (2.00 g, 5.55 mmol) was added to rection mixture and stirred for 72 h. The reaction was quenched by adding solid sodium sulfite (5.7 g) and additionally stirred for 24 h at room temperature. It was diluted with H₂O and mixture was extracted with EtOAc (x3). The combined organic extracts were washed with aqueous KOH (2 M), H₂O, brine, dried (Na₂SO₄), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, EtOAc/hexane = 4/1) to afford **SI-8** (1.84 g, 86%) as a white solid. *R*_f = 0.25 (EtOAc/hexane = 4/1); mp = 81–82 °C; [α]_D²⁰ = +16 (*c* 0.40, CHCl₃); ¹H NMR (600 MHz, CDCl₃) 2.04 (brs, 1H), 2.75 (brs), 2.90 (brs), 3.39 (dd, *J* = 5.4, 11.4 Hz, 1H), 3.50 (dd, *J* = 3.0, 11.4 Hz, 1H), 3.64–3.66 (1H), 4.57 (d, *J* = 7.2 Hz, 1H), 6.85 (dd, *J* = 1.8, 8.4 Hz, 1H), 6.89 (d, *J* = 8.4 Hz, 1H), 6.95 (d, *J*=1.8 Hz, 1H); ¹³C NMR (150 MHz, CDCl₃) δ 63.4, 70.7 (quint., *J*_{C-d} = 21.0 Hz), 74.8, 75.9, 113.8, 115.1, 119.9, 127.1 (t, *J*_{C-d} = 24.0 Hz), 127.2 (t, *J*_{C-d} = 24.0 Hz), 127.5 (t, *J*_{C-d} = 24.0 Hz), 128.1 (t, *J*_{C-d} =

24.0 Hz), 128.2 (t, J_{C-d} = 24.0 Hz), 133.8, 136.98, 137.02, 149.0, 149.1; IR (neat) 3385, 2882, 1590, 1528, 1490, 1328, 1277, 1165, 1139, 1084, 1051, 1032, 909, 732, 648 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{10}\text{D}_{14}\text{O}_5\text{Na}$ [(M+Na)⁺] m/z 417.2395, found m/z 417.2396.

Epoxy alcohol 6: To a solution of 2,4,6-triisopropylbenzenesulfonyl chloride (TrisylCl, 3.7 g, 12 mmol) in pyridine (10 mL) was added triol **SI-8** (1.8 g, 4.6 mmol) at 0 °C. It was slowly warmed to room temperature and stirred for 24 h. The reaction was quenched by adding aqueous HCl (1 M). The mixture was extracted with EtOAc (x3), and the combined organic extracts were washed with H_2O , brine, dried (Na_2SO_4) and concentrated in vacuo. The residue was eluted through a silica gel pad and washed with hexane/EtOAc (2/1) to remove unreacted TrisylCl, giving crude **SI-9**. The crude material was dissolved in a mixed solvent (methanol/1,4-dioxane = 2/1, 24 mL), and K_2CO_3 (1.2 g, 8.7 mmol) was added at 0 °C. After stirring for 2 h, the reaction was stopped by adding H_2O , and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with H_2O , brine, dried (Na_2SO_4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, EtOAc/hexane = 1/2) to afford epoxy alcohol **6** (1.0 g, 58%, 2 steps) as a colorless viscous oil. R_f = 0.25 (EtOAc/Hexane = 1/1); $[\alpha]_D^{20}$ = +0.34 (c 0.87, CHCl_3); ¹H NMR (600 MHz, CDCl_3) δ 2.38 (d, J = 4.8 Hz, 1H), 2.74 (dd, J = 3.0, 4.8 Hz, 1H), 2.79 (t, J = 4.8 Hz, 1H), 3.14 (ddd, J = 3.0, 4.8, 5.4 Hz, 1H), 4.36 (dd, J = 4.8, 5.4 Hz, 1H), 6.91 (dd, J = 1.8, 8.4 Hz, 1H), 6.93 (d, J = 8.4 Hz, 1H), 7.04 (d, J = 1.8 Hz, 1H); ¹³C NMR (150 MHz, CDCl_3) δ 45.5, 56.0, 70.7 (quint, J_{C-d} = 22.5 Hz), 70.8 (quint, J_{C-d} = 22.5 Hz), 74.3, 113.5, 115.2, 119.6, 127.0 (t, J_{C-d} = 24.0 Hz), 127.2 (t, J_{C-d} = 24.0 Hz), 127.4–127.6 (m), 128.1 (t, J_{C-d} = 24.0 Hz), 133.6, 137.0, 137.1, 149.1, 149.2; IR (neat) 3438, 2994, 2205, 1605, 1590, 1509, 1426, 1328, 1271, 1183, 1166, 1031, 993, 919, 852, 819, 801, 748, 542 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{8}\text{D}_{14}\text{O}_4\text{Na}$ [(M+Na)⁺] m/z 399.2289, found m/z 399.2292.

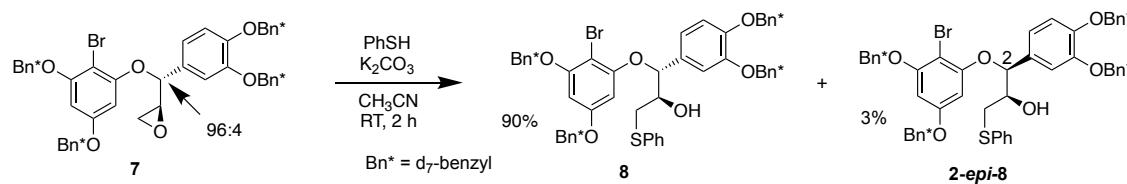
Preparation of epoxy ether **7**



To a solution of bromophenol **5** (0.67 g, 1.7 mmol), epoxy-alcohol **6** (0.95 g, 2.5 mmol) and ADDP (1.3 g, 5.2 mmol) in toluene (17 mL) was added *n*-Bu₃P (1.2 mL, 4.8 mmol) at 0 °C. It was warmed to room temperature and stirred for 2 h. The reaction was quenched by adding phosphate buffer (pH = 7) and the mixture extracted with EtOAc (x3). The combined organic extracts were washed with H_2O , brine, dried (Na_2SO_4), and concentrated in vacuo. The residue was purified by flash column chromatography (silica gel, EtOAc/hexane = 1/3) to afford **7** as a diastereomeric mixture (0.96 g, 76%, d.r. = 96:4) as an ivory foam.

$R_f = 0.75$ (EtOAc/hexane = 1/2); $[\alpha]_D^{20} = -5.8$ (c 0.38, CHCl_3); ^1H NMR (600 MHz, CDCl_3 , signals of minor diastereomers are omitted) δ 2.76 (dd, $J = 3.6, 4.8$ Hz, 1H), 2.97 (dd, $J = 2.4, 4.8$ Hz, 1H), 3.23–3.24 (m, 1H), 5.01 (d, $J = 3.6$ Hz, 1H), 6.05 (d, $J = 2.4$ Hz, 1H), 6.22 (d, $J = 2.4$ Hz, 1H), 6.85 (dd, $J = 1.8, 8.4$ Hz, 1H), 6.88 (d, $J = 8.4$ Hz, 1H), 7.02 (d, $J = 1.8$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3 , signals of minor diastereomers are omitted) δ 45.1, 54.6, 69.9–70.2 (m), 70.6 (quint, $J_{\text{C}-\text{d}} = 22.5$ Hz), 79.4, 95.0, 95.5, 96.2, 113.4, 114.9, 120.1, 126.6, 126.8–128.5 (m), 136.2, 136.4, 136.9, 137.1, 149.2, 149.3, 155.5, 156.7, 159.2; IR (neat) 2118, 1585, 1491, 1478, 1327, 1272, 1178, 1086, 1050, 1032, 988, 730, 610, 574, 542, 513 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{43}\text{H}_{10}\text{D}_{28}\text{BrO}_6$ [(M+H) $^+$] m/z 757.3604, found m/z 757.3609.

Opening of epoxide 7 to give sulfide 8



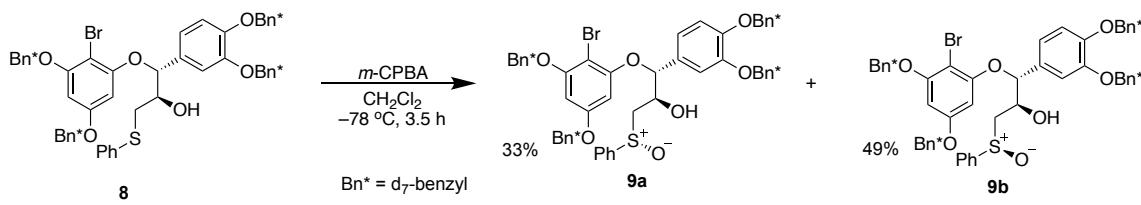
To a solution of epoxide 7 (500 mg, 0.660 mmol) and K_2CO_3 (182 mg, 1.32 mmol) in CH_3CN (6.5 mL) was added PhSH (135 μL , 1.32 mmol) at room temperature. It was stirred for 2 h and diluted with diethyl ether. Reaction mixture was filtered through a Celite® pad and washed with diethyl ether (x3). The combined filtrate was concentrated in vacuo. The residue was purified by flash column chromatography (silica-gel, EtOAc/hexane = 1/5) to afford 8 (514 mg, 90%) as an ivory foam and 2-*epi*-8 (16 mg, 3%) as an colorless oil.

Sulfide 8: $R_f = 0.23$ (hexane/EtOAc = 7/2); $[\alpha]_D^{20} = +20$ (c 0.75, CHCl_3); ^1H NMR (600 MHz, CDCl_3) δ 2.46 (d, $J = 5.4$ Hz, 1H), 3.12 (dd, $J = 8.4, 13.8$ Hz, 1H), 3.27 (dd, $J = 3.6, 13.8$ Hz, 1H), 3.96–4.00 (m, 1H), 5.08 (d, $J = 4.8$ Hz, 1H), 5.95 (d, $J = 2.4$ Hz, 1H), 6.21 (d, $J = 2.4$ Hz, 1H), 6.80 (dd, $J = 1.8, 8.4$ Hz, 1H), 6.87 (d, $J = 8.4$ Hz, 1H), 6.95 (d, $J = 1.8$ Hz, 1H), 7.11–7.14 (m, 1H), 7.18–7.21 (m, 2H), 7.26–7.27 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 36.3, 69.4–70.8 (m), 73.4, 82.6, 94.4, 95.3, 95.7, 113.4, 114.8, 120.2, 126.6, 126.6–128.4 (m), 129.1, 129.2, 129.9, 135.6, 136.2, 136.3, 136.7, 137.0, 149.06, 149.12, 155.4, 156.6, 159.2; IR (neat) 3475, 2981, 2119, 1584, 1509, 1327, 1270, 1176, 1085, 1032, 909, 818, 738, 539 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{49}\text{H}_{16}\text{D}_{28}\text{O}_6\text{BrS}$ [(M+H) $^+$] m/z 867.3794, found m/z 867.3789.

Sulfide 2-*epi*-8: $R_f = 0.25$ (hexane/EtOAc = 7/2); $[\alpha]_D^{20} = -1.31$ (c 0.130, CHCl_3); ^1H NMR (600 MHz, CDCl_3) δ 2.78 (dd, $J = 7.2, 13.8$ Hz, 1H), 2.96 (d, $J = 4.2$ Hz, 1H), 3.18 (dd, $J = 4.2, 13.8$ Hz, 1H), 4.00–4.03 (m, 1H), 5.09 (d, $J = 5.4$ Hz, 1H), 5.95 (d, $J = 2.4$ Hz, 1H), 6.22 (d, $J = 2.4$ Hz, 1H), 6.80 (dd, $J = 1.8,$

8.4 Hz, 1H), 6.87 (d, J = 8.4 Hz, 1H), 6.92 (d, J = 1.8 Hz, 1H), 7.12 (t, J = 7.2 Hz, 1H), 7.19 (t, J = 7.8 Hz, 1H), 7.24–7.26 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 36.1, 53.6, 69.4–70.9 (m), 74.0, 82.8, 94.5, 95.4, 95.9, 113.5, 114.9, 120.2, 126.3, 126.6–128.4 (m), 129.1, 129.2, 130.1, 135.9, 136.1, 136.3, 136.7, 137.0, 149.1, 149.3, 155.5, 156.6, 159.2; IR (neat) 3552, 2921, 1584, 1508, 1363, 1271, 1230, 1175, 1085, 1031, 818, 739, 540 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{49}\text{H}_{16}\text{D}_{28}\text{O}_6\text{BrS}$ [(M+H) $^+$] m/z 869.3776, found m/z 869.3784.

Oxidation of sulfide 8 to hydroxy-sulfoxides 9a and 9b



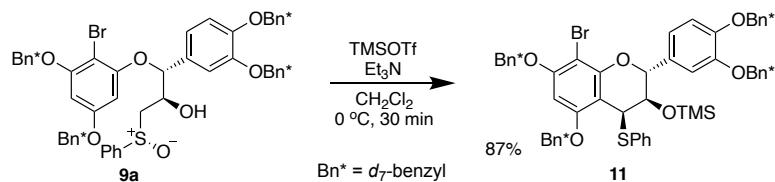
To a solution of sulfide **8** (30.0 mg, 0.0346 mmol) in CH_2Cl_2 (0.7 mL) at -78°C was added *m*-CPBA (77%, 8.5 mg, 0.038 mmol). After stirring for 3.5 h, the reaction was quenched by adding saturated aqueous Na_2SO_3 solution. The crude products were extracted with EtOAc (x3). Combined organic extracts were washed with water, brine, dried (Na_2SO_4) and concentrated in vacuo. The residue was purified by preparative TLC (silica-gel, THF/hexane = 2/7, developed 7 times) to afford less polar hydroxy-sulfoxide **9a** (10.1 mg, 33%) as a viscous oil and more polar hydroxy-sulfoxide **9b** (15.0 mg, 49%) as a viscous oil.

anti-isomer 9a: $R_f = 0.30$ (EtOAc/hexane = 3/2); $[\alpha]_D^{20} = -22$ (c 0.87, CHCl_3); ^1H NMR (600 MHz, CDCl_3) δ 2.91 (dd, J = 1.8, 14.4 Hz, 1H), 3.27 (dd, J = 9.6, 14.4 Hz, 1H), 3.71 (br, 1H), 4.30–4.33 (m, 1H), 5.04 (d, J = 5.4 Hz, 1H), 5.94 (d, J = 2.4 Hz, 1H), 6.21 (d, J = 2.4 Hz, 1H), 6.67 (dd, J = 1.8, 7.8 Hz, 1H), 6.80 (d, J = 7.8 Hz, 1H), 6.85 (d, J = 1.8 Hz, 1H), 7.44–7.49 (m, 5H); ^{13}C NMR (150 MHz, CDCl_3) δ 56.7, 69.8–70.9 (m), 71.3, 83.2, 94.2, 95.4, 95.6, 113.0, 114.8, 119.9, 124.2, 126.6–129.4 (m), 129.5, 131.1, 136.1, 136.3, 136.8, 137.0, 142.8, 149.0, 149.1, 155.4, 156.6, 159.3; IR (neat) 3322, 1585, 1510, 1427, 1468, 1365, 1271, 1231, 1202, 1176, 1112, 1085, 996, 818, 731, 690 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{49}\text{H}_{15}\text{D}_{28}\text{BrO}_7\text{SNa}$ [(M+Na) $^+$] m/z 907.3545, found m/z 907.3543.

syn-isomer 9b: $R_f = 0.30$ (EtOAc/hexane = 3/2); $[\alpha]_D^{20} = +31$ (c 0.67, CHCl_3); ^1H NMR (600 MHz, CDCl_3) δ 3.17 (dd, J = 3.0, 13.2 Hz, 1H), 3.20 (dd, J = 8.4, 13.2, 1H), 3.64 (d, J = 3.6 Hz, 1H), 4.37–4.40 (m, 1H), 5.01 (d, J = 5.4 Hz, 1H), 5.95 (d, J = 2.4 Hz, 1H), 6.21 (d, J = 2.4 Hz, 1H), 6.80 (dd, J = 1.8, 14.4 Hz, 1H), 6.87 (d, J = 14.4 Hz, 1H), 6.94 (d, J = 1.8 Hz, 1H), 7.50–7.53 (m, 3H), 7.63–7.64 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 53.6, 58.1, 68.9–70.8 (m), 72.6, 83.3, 94.3, 95.4, 95.7, 113.2, 114.9, 120.1, 124.3, 126.6–

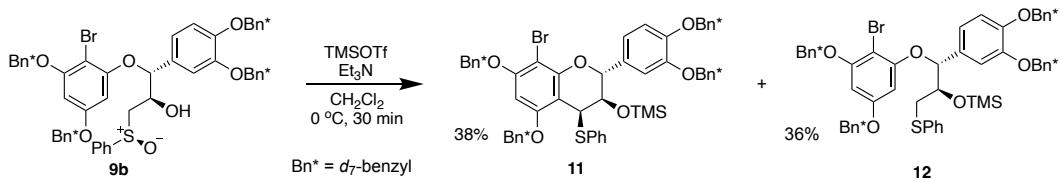
128.4 (m), 129.6, 131.6, 136.1, 136.3, 136.7, 137.0, 143.8, 149.2, 149.3, 155.4, 155.6, 156.6, 159.3; IR (neat) 3330, 1584, 1508, 1427, 1468, 1327, 1271, 1231, 1202, 1176, 1112, 1086, 1032, 818, 747 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{49}\text{H}_{15}\text{D}_{28}\text{BrO}_7\text{SNa}^+[(\text{M}+\text{Na})^+] m/z$ 905.3563, found m/z 905.3561.

Pummerer/Friedel–Crafts cascade of 9a for the preparation of flavan 11



To the solution of **9a** (20.0 mg, 0.0226 mmol) in CH₂Cl₂ (1 mL) at 0 °C, was added Et₃N (26 µL, 0.18 mmol) followed by TMSOTf (30 µL, 0.16 mmol) and stirred for 30 min. The reaction was quenched by adding aqueous saturated NaHCO₃ solution and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with H₂O, brine, dried (Na₂SO₄) and concentrated in vacuo. The residue was purified by preparative TLC (silica gel, EtOAc/hexane = 1/4) to afford flavan **11** (18.3 mg, 87%) as a colorless oil.

Pummerer/Friedel-Crafts cascade of 9b to give flavan 11 and reduced product 12



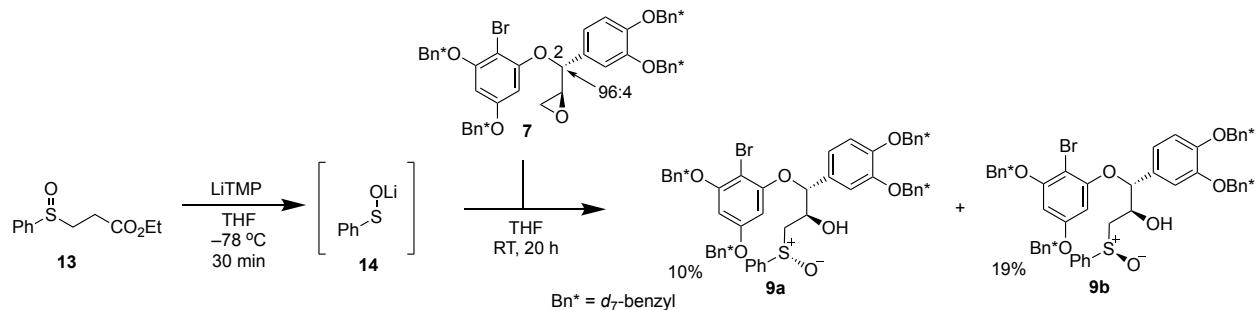
To the solution of **9b** (20.0 mg, 0.0226 mmol) in CH₂Cl₂ (1 mL) at 0 °C, was added Et₃N (26 µL, 0.18 mmol) followed by TMSOTf (30 µL, 0.16 mmol) and stirred for 30 min. The reaction was quenched by adding aqueous saturated NaHCO₃ solution and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with H₂O, brine, dried (Na₂SO₄) and concentrated in vacuo. The residue was purified by preparative TLC (silica gel, EtOAc/hexane = 1/9, developed 4 times) to afford flavan **11** (8.1 mg, 38%) as a colorless oil and sulfide **12** (7.6 mg, 36%) as a colorless oil.

Flavan 11: $R_f = 0.70$ (EtOAc/hexane = 1/4); $[\alpha]_D^{20} = +63$ (c 0.23, CHCl_3); ^1H NMR (600 MHz, CDCl_3) δ 0.64 (s, 9H), 3.87 (dd, $J = 9.0, 4.2$ Hz, 1H), 4.53 (d, $J = 4.2$ Hz, 1H), 5.42 (d, $J = 9.0$ Hz, 1H), 6.26 (s, 1H), 6.93 (d, $J = 8.4$ Hz, 1H), 7.02–7.06 (m, 3H), 7.12 (dd, $J = 8.4, 1.8$ Hz, 1H), 7.17 (d, $J = 1.8$ Hz, 1H), 7.42–7.44 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3) δ 0.1, 50.4, 71.1–72.1 (m), 73.6, 79.0, 93.5, 93.6, 105.9, 115.6,

122.3, 127.6–128.1 (m), 128.2, 128.4–129.4 (m), 129.5, 133.2, 135.2, 137.1, 137.5, 138.2, 138.5, 149.8, 150.0, 152.8, 157.2, 157.4; IR (neat) 2918, 2277, 1600, 1511, 1502, 1479, 1414, 1365, 1270, 1184, 1113, 1084, 1051, 1029, 841 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{52}\text{H}_{22}\text{D}_{28}\text{BrO}_6\text{SSi}[(\text{M}+\text{H})^+]$ m/z 937.4033, found m/z 937.4029.

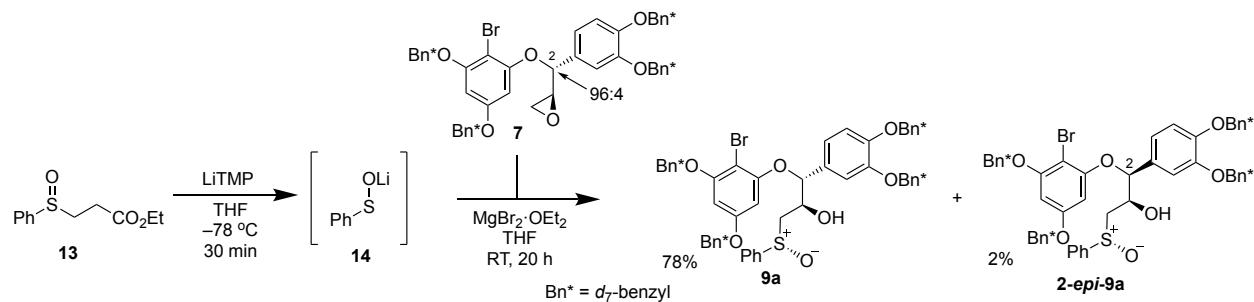
Sulfide 12: $R_f = 0.72$ (EtOAc/hexane = 1/4); $[\alpha]_D^{20} = +11.5$ (c 1.07, CHCl_3); ^1H NMR (600 MHz, CDCl_3) δ –0.18 (s, 9H), 3.20 (dd, $J = 6.6, 13.8$ Hz, 1H), 3.46 (dd, $J = 3.0, 13.8$ Hz, 1H), 4.11 (ddd, $J = 3.0, 5.4, 6.6$ Hz, 1H), 5.01 (d, $J = 5.4$ Hz, 1H), 6.00 (d, $J = 2.4$ Hz, 1H), 6.19 (d, $J = 2.4$ Hz, 1H), 6.81–6.85 (m, 2H), 7.01 (brs, 1H), 7.11 (t, $J = 7.8$ Hz, 1H), 7.22 (t, $J = 7.8$ Hz, 1H), 7.36 (d, $J = 7.8$ Hz, 1H); ^{13}C NMR (150 MHz, C_6D_6) δ 0.3, 38.8, 69.4–70.7 (m), 75.6, 83.0, 94.8, 95.3, 96.0, 114.5, 115.1, 121.3, 126.9, 127.0–128.5 (m), 129.2, 131.7, 136.8, 137.4, 137.5, 137.7, 149.6, 149.9, 156.2, 157.2, 159.7; IR (neat) 2954, 2119, 1584, 1480, 1469, 1365, 1230, 1177, 1085, 1031, 841, 819, 739, 690, 541 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{52}\text{H}_{24}\text{D}_{28}\text{BrO}_6\text{SSi}[(\text{M}+\text{H})^+]$ m/z 939.4189, found m/z 939.4193.

Ring-opening of epoxide 7 by phenyl sulfinate 14 to provide 9a and 9b (without $\text{MgBr}_2 \cdot \text{OEt}_2$)



To the solution of 2,2,6,6-tetramethylpipiridine (0.17 mL, 0.99 mmol) in THF (3 mL) at 0 °C, was added *n*-BuLi (0.56 mL, 0.89 mmol) and stirred for 1 h. To this, a solution of sulfoxide **13** (0.18 g, 0.80 mmol) in THF (2 mL) was added at –78 °C. After stirring for 30 min, a solution of **7** (40 mg, 0.053 mmol) in THF (2 mL) was added and stirred for 20 h at room temperature. Reaction was quenched by adding saturated aqueous NH₄Cl solution and the mixture was extracted with EtOAc (x3). Combined organic extracts were washed with H₂O, brine, dried (Na₂SO₄) and concentrated in vacuo. The residue was purified by preparative TLC (silica gel, EtOAc/hexane = 1/1) to obtain a mixture of **9a** and **9b** (13.4 mg in total) and starting epoxy-ether **7** (24.5 mg, 61%). **9a** and **9b** was separated by preparative TLC (silica gel, THF/hexane = 1/4, developed 6 times) to provide less polar *anti*-isomer **9a** (4.5 mg, 10%) as a viscous oil and more polar *syn*-isomer **9b** (8.8 mg, 19%) as a viscous oil.

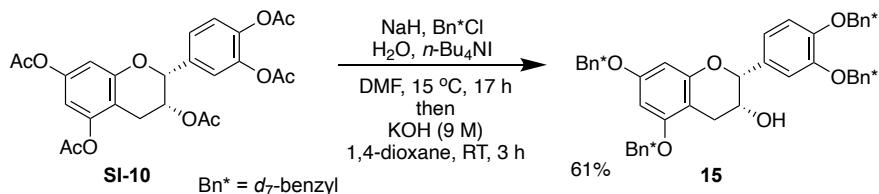
Conversion of epoxide 7 to 9a (with MgBr₂·OEt₂)



To a solution of 2,2,6,6-tetramethylpipiridine (0.17 mL, 0.998 mmol) in THF (3 mL) at 0 °C, was added *n*-BuLi (0.56 mL, 0.89 mmol) and stirred for 1 h. To this, a solution of sulfoxide **13** (0.18 g, 0.80 mmol) in THF (2 mL) was added at –78 °C. After stirring for 30 min, MgBr₂·OEt₂ (0.21 g, 0.80 mmol) and a solution of **7** (40 mg, 0.053 mmol) in THF (2 mL) were successively added, and the mixture was stirred for 20 h at room temperature. The reaction was quenched by adding saturated aqueous NH₄Cl solution, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with H₂O, brine, dried (Na₂SO₄) and concentrated in vacuo. The residue was purified by preparative TLC (silica gel, EtOAc/hexane = 1/1) to obtain **9a** (36.3 mg, 78%) as an ivory foam and **2-epi-9a** (0.8 mg, 2%) as a colorless viscous oil.

2-epi-9a: R_f = 0.32 (EtOAc/hexane = 3/2); $[\alpha]_D^{20} = -44$ (*c* 0.27, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 2.64 (d, *J* = 12.6 Hz, 1H), 2.86 (dd, *J* = 10.2, 12.6 Hz, 1H), 3.69 (d, *J* = 1.8 Hz, 1H), 4.46–4.48 (m, 1H), 4.87 (d, *J* = 5.4 Hz, 1H), 5.87 (d, *J* = 2.4 Hz, 1H), 6.19 (d, *J* = 2.4 Hz, 1H), 6.75 (dd, *J* = 1.8, 8.4 Hz, 1H), 6.85 (d, *J* = 8.4 Hz, 1H), 6.87 (d, *J* = 1.8 Hz, 1H), 7.44–7.51 (m, 3H), 7.57–7.58 (m, 2H); ¹³C NMR (150 MHz, CDCl₃) δ 29.9, 57.2, 69.8–71.2 (m), 69.9, 82.9, 94.4, 95.4, 95.6, 113.5, 114.8, 120.4, 124.2, 126.6–128.4 (m), 129.0, 131.1, 136.1, 136.3, 136.8, 137.0, 143.0, 148.9, 149.3, 155.2, 156.6, 159.2; IR (neat) 3329, 1580, 1510, 1425, 1464, 1269, 1230, 1200, 1173, 1109, 1030, 819, 746 cm^{–1}; HRMS (ESI) calcd for C₄₉H₁₆D₂₈BrO₇S [(M+H)⁺] *m/z* 883.3743, found *m/z* 883.3707.

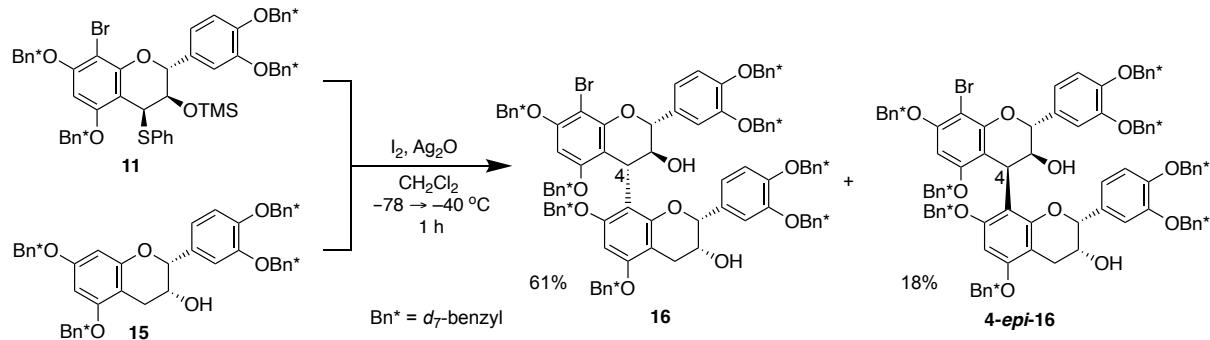
Synthesis of tetra-*O*-*d*₇-benzyl epicatechin **15**



To a suspension of NaH (0.19 g, 63% dispersion in mineral oil, 5.0 mmol) in DMF (5 mL) at 0 °C was added **SI-10**⁵ (0.3 g, 0.6 mmol) in DMF (5 mL) followed by tetrabutylammonium iodide (TBAI) (55 mg,

0.15 mmol). After stirring for 10 min, $Bn^*\text{Cl}$ (d_7 -benzyl chloride, 0.30 mL, 2.5 mmol) and H_2O (43 μL , 2.4 mmol) was successively added and stirred at 15 °C for 17 h. The reaction was quenched by successively adding Et_2NH and H_2O at 0 °C, and the mixture was extracted with EtOAc (x3). The combined organic extracts were washed with water, brine, dried (Na_2SO_4) and concentrated in vacuo. The residue was re-dissolved in 1,4-dioxane (12 mL), and aqueous KOH (9 M, 3 mL) was added at 0 °C. After stirring for 3 h, the reaction was quenched by adding H_2O . The mixture was extracted with EtOAc (x3), and the combined organic extracts were successively washed with aqueous 1 M HCl, H_2O , brine, dried (Na_2SO_4) and concentrated in vacuo. The residue was purified by column chromatography (silica gel, $\text{EtOAc}/\text{hexane} = 1/4$) to afford **15** (0.25 g, 61%) as an ivory foam. $R_f = 0.55$ ($\text{EtOAc}/\text{hexane} = 1/3$); $[\alpha]_D^{20} = -13$ (c 0.91, CHCl_3); ^1H NMR (600 MHz, CDCl_3) δ 2.93 (dd, $J = 4.2, 17.4$ Hz, 1H), 2.99 (dd, $J = 1.8, 17.4$ Hz, 1H), 4.22 (s, 1H), 4.92 (s, 1H), 6.26 (d, $J = 2.4$ Hz, 1H), 6.28 (d, $J = 2.4$ Hz, 1H), 6.97 (d, $J = 8.4$ Hz, 1H), 7.01 (dd, $J = 1.8, 8.4$ Hz, 1H), 7.14 (d, $J = 1.8$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3) δ 14.4, 21.2, 28.4, 60.6, 66.5, 67.8–69.9 (m), 70.1–71.8 (m), 78.6, 94.2, 94.8, 101.1, 113.7, 115.3, 119.6, 126.8–128.4 (m), 131.6, 136.8, 136.9, 137.0, 137.1, 149.0, 149.2, 155.4, 158.5, 159.0, 171.3; IR (neat) 3539, 2269, 1737, 1616, 1593, 1509, 1493, 1327, 1200, 1158, 1141, 1084, 1038, 995, 838, 807, 765, 654 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{43}\text{H}_{11}\text{D}_{28}\text{O}_6$ $[(\text{M}+\text{H})^+]$ m/z 679.4499, found m/z 679.4473.

Attachment of bottom epicatechin unit **15** to sulfide **11** to give **16**



To a solution of sulfide **11** (40 mg, 0.043 mmol) and epicatechin derivative **15** (43 mg, 0.063 mmol) in CH_2Cl_2 (3 mL) was added Ag_2O (15 mg, 0.065 mmol) followed by dropwise addition of a solution of I_2 (33 mg, 0.13 mmol) in CH_2Cl_2 (2 mL) at -78 °C. It was stirred for 30 min before warming up to -40 °C over 30 min. The reaction was quenched by adding 10% aqueous $\text{Na}_2\text{S}_2\text{O}_3$ solution and saturated NaHCO_3 solution. The mixture was extracted with EtOAc (x3). The combined organic extracts were washed with H_2O , brine, dried (Na_2SO_4) and concentrated in vacuo. The residue was purified by preparative TLC (silica gel, $\text{CHCl}_3/\text{EtOAc}/\text{CH}_2\text{Cl}_2$ /hexane = 4/1/1/2, developed 2 times) to afford dimer **16** (mixture of rotamers, 37.4 mg, 61%) as an ivory foam and **4-epi-16** (mixture of rotamers, 11.0 mg, 18%) as a colorless viscous oil.

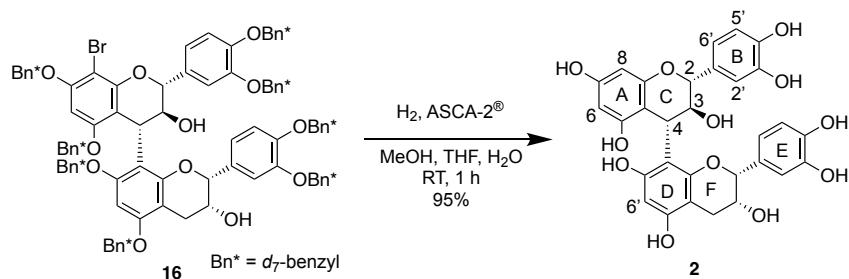
16: $R_f = 0.27$ ($\text{CHCl}_3/\text{EtOAc}/\text{CH}_2\text{Cl}_2/\text{hexane} = 4/1/1/2$); $[\alpha]_D^{20} = -83$ (c 0.49, CHCl_3); ^1H NMR (600 MHz, CDCl_3 , rotamer ratio is 1.0:0.55, signals of the minor rotamer are marked with an asterisk) δ *1.52 (br, 0.55H), 1.66 (br, 1H), 2.58 (dd, $J = 4.2, 16.8$ Hz, 1H), *2.87 (dd, $J = 4.2, 17.4$ Hz, 0.55H), 2.88 (d, $J = 17.4$ Hz, 1H), *3.01 (d, $J = 17.4$ Hz, 0.55H), 3.79 (s, 1H), 3.88 (brs, 1H), *4.06 (brm, 0.55H), *4.16 (dt, $J = 3.6, 9.6$ Hz, 0.55H), 4.19 (t, $J = 9.6$ Hz, 1H), 4.58 (d, $J = 9.6$ Hz, 1H), *4.68 (d, $J = 9.6$ Hz, 0.55H), *4.75 (brs, 0.55H), 4.83 (d, $J = 9.6$ Hz, 1H), *4.85 (d, $J = 9.6$ Hz, 0.55H), *5.95 (s, 0.55H), 6.17 (s, 1H), *6.18 (s, 0.55H), *6.21 (dd, $J = 1.8, 8.4$ Hz, 0.55H), 6.22 (s, 1H), *6.68 (d, $J = 1.8$ Hz, 0.55H), 6.75 (d, $J = 8.4$ Hz, 1H), 6.76 (d, $J = 1.8$ Hz, 1H), *6.77–6.91 (m, 1.10H), *6.84 (d, $J = 8.4$ Hz, 0.55H), *6.88 (dd, $J = 1.8, 8.4$ Hz, 0.55H), 6.92 (d, $J = 8.4$ Hz, 1H), 7.01–7.02 (m, 2H), 7.14 (d, $J = 1.8$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3 signals of the minor rotamer are marked with an asterisk) δ 28.3, *29.9, 37.3, *37.6, 66.2, *66.3, *69.0–70.01 (m), 70.03–71.8 (m), *72.5, 73.1, 77.5, *78.3, *82.0, 82.4, 91.3, *92.1, 93.5, *93.8, 93.9, *94.0, 100.9, *102.3, 110.3, *110.9, *111.2, 111.4, *112.7, 113.1, *113.5, 113.8, 114.7, *114.9, *115.1, 115.4, 118.9, *119.7, *120.8, 121.2, *126.6–128.4 (m), *130.4, 131.5, *131.5, 132.2, *136.2–137.5, 148.5, *148.7, *148.96, *149.00, *149.04, 149.1, 149.15, 149.17, *152.8, *153.4, 153.6, 153.8, 154.3, *154.4, 155.6, *156.3, 156.4, 156.5, *156.9, *157.1; IR (neat) 3441, 2921, 2119, 1598, 1510, 1416, 1357, 1327, 1270, 1201, 1114, 1052, 1030, 909, 819, 731, 544 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{86}\text{H}_{28}\text{D}_{56}\text{BrO}_{12}$ [(M+H) $^+$] m/z 1433.7873, found m/z 1433.7857.

^a ^{13}C signals of both rotamers are overlapping.

4-*epi*-16: $R_f = 0.30$ ($\text{CHCl}_3/\text{EtOAc}/\text{CH}_2\text{Cl}_2/\text{hexane} = 4/1/1/2$); $[\alpha]_D^{20} = +26$ (c 0.55, CHCl_3); ^1H NMR (600 MHz, CDCl_3 , rotamer ratio = 1.0:0.50, signals of the minor rotamer are marked with an asterisk) δ *2.61 (brd, $J = 4.2$ Hz, 0.5H), 2.86 (br, 1H), 2.92 (dd, $J = 9.0, 18.0$ Hz, 1H), *2.96–3.00 (m, 1H), *2.96–3.00 (m, 1H), 3.89 (d, $J = 3.0$ Hz, 0.5H), 4.07 (s, 1H), *4.09–4.11 (m, 0.5H), 4.15–4.19 (m, 1H), *4.15–4.19 (m, 0.5H), *4.88 (s, 0.5H), 5.00 (1H), *5.01 (d, $J = 6.0$ Hz, 0.5H), *5.21 (d, $J = 7.2$ Hz, 0.5H), 5.37 (d, $J = 7.8$ Hz, 1H), 6.01 (s, 1H), *6.16 (s, 0.5H), *6.21 (s, 0.5H), 6.30 (dd, $J = 1.8, 7.8$ Hz, 1H), 6.32 (s, 1H), *6.72 (d, $J = 7.8$ Hz, 0.5H), 6.78–6.87 (m, 3H), *6.78–6.87 (m, 1.5H), 6.89 (d, $J = 1.8$ Hz, 1H), 6.98 (d, $J = 7.2$ Hz, 1H), *7.04 (d, $J = 1.8$ Hz, 0.5H), *7.10 (d, $J = 1.8$ Hz, 0.5H), 7.20 (d, $J = 1.8$ Hz, 1H); ^{13}C NMR (150 MHz, CDCl_3 , signals of the minor rotamer are marked with an asterisk) δ 28.86, *28.92, *32.0, 33.4, *66.0, 66.8, *68.0–69.3 (m), 69.7–72.1 (m), *71.0, 71.7, 78.7, *78.8, *79.1, 79.3, 91.8, *92.5, *92.7, 92.9, *93.5, 94.0, *101.6, 103.2, *108.2, 108.5, 108.8, *109.0, 111.5, *113.4, *113.5, 113.6, 114.7, *114.8, *115.0, 115.5, 118.2, *119.5, *119.8, 120.3, *126.8–128.3 (m), 130.9, *131.4, *132.1, 132.2, *136.36, *136.40, 136.5, 136.6, *136.8, *136.9, *137.0, *137.1, 137.2, *137.3, 137.4, 137.6, 148.5, *148.7, *148.8, *148.9, 149.2, 149.7, 152.1, *152.3, *153.5, 154.7, *155.2, *156.2, 156.6, 156.9, *157.1; IR (neat) 3532, 2922, 1601, 1510, 1583, 1416, 1358, 1272, 1230, 1202, 1189, 1107, 1052, 909, 839, 731 cm^{-1} ; HRMS (ESI) calcd for $\text{C}_{86}\text{H}_{28}\text{D}_{56}\text{BrO}_{12}$ [(M+H) $^+$] m/z 1433.7873, found m/z 1433.7877.

^a ^{13}C signals of both rotamers are overlapping

Synthesis of procyanidin B4 (2)



A solution of tetramer **16** (40.0 mg, 0.0279 mmol) in the presence of ASCA-2[®] (92.0 mg, 0.0328 mmol) in a mixture of THF, MeOH and H₂O (v/v/v = 2/2/1, 15 mL) was stirred under H₂ atmosphere at room temperature. After stirring for 1 h, it was carefully filtered under argon atmosphere through a glass fiber filter (MeOH) and the filtrate was evaporated to remove organic solvents and lyophilized to give a crude material, which was purified by preparative HPLC [Inertsustain[®] C18, 20 mm ϕ \times 250 mm, MeOH, H₂O (35/65) containing 0.1% TFA, flow rate 8 mL/min, 20 \times 250 mm] and lyophilization gave **2**⁶ (15.3 mg, 95%) as an ivory amorphous solid. $[\alpha]_D^{25} = -198$ (*c* 0.560, EtOH); ^{13}C -NMR (150 MHz, CD₃OD, signals of the minor rotamer are marked with an asterisk) δ *30.1, 30.9, *39.6, 39.7, 68.3, *68.6, *74.6, 74.7, *80.7, 80.9, 84.7, *84.9, *97.2–98.0 (m), 98.1–98.9 (m), 100.3, *100.9, 108.1, 108.2, *109.1, *109.6, *115.6, 116.1, 116.7, *116.78, *116.82, 116.9, *117.2, 120.0, *121.1, *121.3, 122.1, *132.6, 133.1, *133.3, 133.4, *146.41, *146.44, 146.5, *146.6, 146.8, *146.9, 147.0, 147.3, *156.2, *156.6, 156.7, *157.1, 157.2, 158.1, *158.0, *158.2, 158.3, *159.3, 159.5; IR(neat) 3295 (br), 1605, 1519, 1446, 1281, 1260, 1197, 1143, 1062, 800, 609 cm⁻¹; HRMS (ESI) calcd for C₃₀H₂₅O₁₂ [(M+H)⁺] *m/z* 577.1351, found *m/z* 577.1338.

^a ^{13}C signals of both rotamers are overlapping.

Note: Because of deuterium exchange with CD₃OD, the corresponding signals for C4, C8 and C6' are observed as multiplets with decreased intensity levels.

Comparison of ^1H NMR data of 2 (600 MHz, CD₃OD, rotamer ratio = 1.0:0.77)

Major rotamer			
Ring	Carbon number	Literature ⁶	Synthetic
		^1H (400 MHz)	^1H (600MHz)
Top unit			
C	C-2	4.53 (d, J = 8.0 Hz) ⁴	4.67 (d, J = 7.8 Hz) ⁴
	C-3	4.48 (dd, J = 8.0, 9.5 Hz)	4.61 (dd, J = 7.8, 10.2 Hz)
	C-4	4.32 (d, J = 9.5 Hz) ⁴	4.45 (d, J = 10.2 Hz) ⁴
A	C-5	—	—
	C-6	5.74 (d, J = 2.3 Hz) ³	5.87 (d, J = 1.8 Hz) ³
	C-7	—	—
	C-8	5.69 (d, J = 2.3 Hz) ³	5.83 (d, J = 1.8 Hz) ³
	C-9	—	—
	C-10	—	—
B	C-1'	—	—
	C-2'	6.99 (d, J = 1.9 Hz) ¹	7.12 (d, J = 1.8 Hz) ¹
	C-3'	—	—
	C-4'	—	—
	C-5'	6.69 (d, J = 8.1 Hz) ²	6.82 (d, J = 8.4 Hz) ²
	C-6'	6.77 (dd, J = 1.9, 8.1 Hz) ^a	6.89 (dd, J = 1.8, 8.4 Hz) ^a
Bottom Unit			
F	C-2	4.83 (brs)	4.97 (brs)
	C-3	4.16–4.18 (m)	4.26 (brm)
	C-4	2.83 (dd, J = 4.3, 16.8) 2.72 (dd, J = 1.5, 16.8)	2.96 (dd, J = 4.8, 16.8 Hz) 2.87 (dd, J = 1.2, 16.8 Hz)
D	C-5	—	—
	C-6	5.85 (s)	5.99 (s)
	C-7	—	—
	C-8	—	—
	C-9	—	—
	C-10	—	—
E	C-1'	—	—
	C-2'	6.89 (d, J = 1.9 Hz) ¹	7.02 (d, J = 1.8 Hz) ¹
	C-3'	—	—
	C-4'	—	—
	C-5'	6.68 (d, J = 8.1 Hz) ²	6.80 (d, J = 8.4 Hz) ²
	C-6'	6.77 (dd, J = 1.9, 8.1 Hz) ^a	6.89 (dd, J = 1.8, 8.4 Hz) ^a

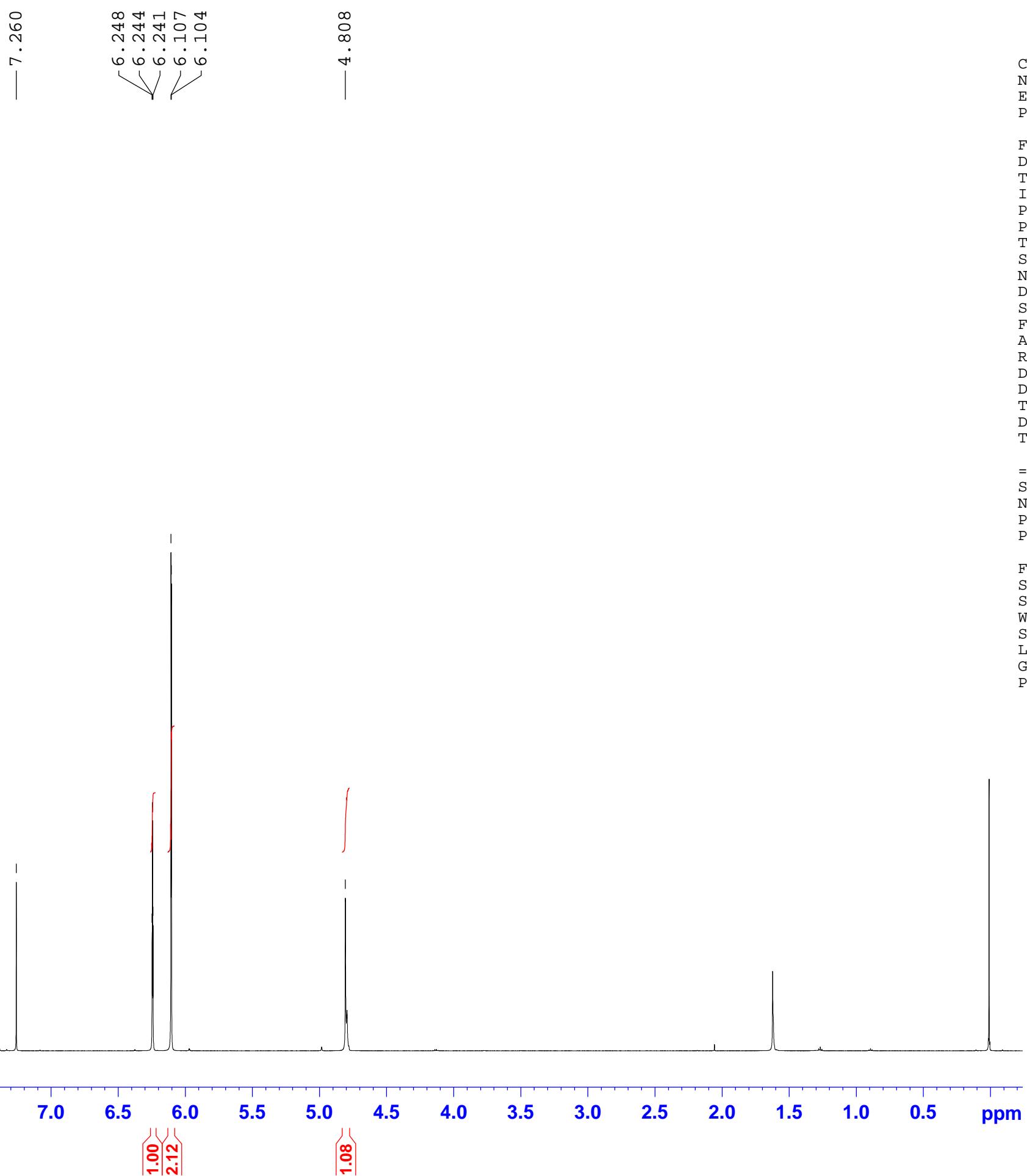
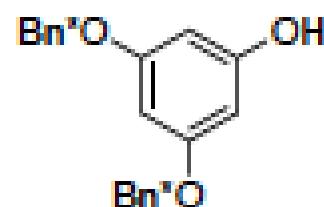
Minor rotamer			
Ring	Carbon number	Literature ⁶	Synthetic
		¹ H (400 MHz)	¹ H (600MHz)
Top unit			
C	C-2	4.22 (d, <i>J</i> = 6.1 Hz)	4.34–4.35 (m) ^a
	C-3	4.36 (dd, <i>J</i> = 2.2, 6.1 Hz)	4.50 (dd, <i>J</i> = 1.2, 6.0 Hz)
	C-4	4.21 (d, <i>J</i> = 2.2 Hz)	4.34–4.35 (m) ^a
A	C-5	—	—
	C-6	5.84 (d, <i>J</i> = 2.5 Hz) ⁴	5.97 (d, <i>J</i> = 1.8 Hz) ⁴
	C-7	—	—
	C-8	5.79 (d, <i>J</i> = 2.5 Hz) ⁴	5.93 (d, <i>J</i> = 1.8 Hz) ⁴
	C-9	—	—
	C-10	—	—
B	C-1'	—	—
	C-2'	6.59 (d, <i>J</i> = 1.9 Hz) ²	6.72 (d, <i>J</i> = 1.8 Hz) ²
	C-3'	—	—
	C-4'	—	—
	C-5'	6.62 (d, <i>J</i> = 8.0 Hz) ¹	6.75 (d, <i>J</i> = 7.8 Hz) ¹
	C-6'	6.35 (dd, <i>J</i> = 1.9, 8.0 Hz) ³	6.49 (dd, <i>J</i> = 1.8, 7.8 Hz) ³
Bottom Unit			
F	C-2	4.71 (brs)	4.84 (brs)
	C-3	3.95–3.85 (m)	4.09 (brm)
	C-4	2.85–2.70 (m) 2.60 (dd, <i>J</i> = 2.2, 17.3)	2.91 (dd, <i>J</i> = 5.4, 16.8 Hz) 2.73 (dd, <i>J</i> = 2.4, 16.8 Hz)
D	C-5	—	—
	C-6	6.00 (s)	6.13 (s)
	C-7	—	—
	C-8	—	—
	C-9	—	—
	C-10	—	—
E	C-1'	—	—
	C-2'	6.58 (d, <i>J</i> = 1.9 Hz) ²	6.71 (d, <i>J</i> = 1.8 Hz) ²
	C-3'	—	—
	C-4'	—	—
	C-5'	6.51 (d, <i>J</i> = 8.0 Hz) ¹	6.64 (d, <i>J</i> = 7.8 Hz) ¹
	C-6'	6.31 (dd, <i>J</i> = 1.9, 8.0 Hz) ³	6.45 (dd, <i>J</i> = 1.8, 7.8 Hz) ³

Note: a) Number 1,2,3 and 4 in each column may be interchanged. b) Symbol *a* in each column represents overlapped signals.

References:

1. H. Kawamoto, F. Nakatsubo and K. Murakami, *Syn. Commun.*, 1996, **26**, 531–534.
2. S. B. Wan, K. R. L. -Piwowar, D. J. Kuhn, D. Chen, Q. P. Dou and T. H. Chan, *Bioorg. Med. Chem.*, 2005, **13**, 2177–2185.
3. P. M. Tadross, C. D. Gilmore, P. Bugga, S. C. Virgil and B. M. Stoltz, *Org. Lett.*, 2010, **12**, 1224–1227.
4. K. Ohmori, T. Yano and K. Suzuki, *Org. Biomol. Chem.*, 2010, **8**, 2693–2696.
5. V. V. Betkekar, K. Suzuki and K. Ohmori, *Org. Bio. Chem.*, 2019, **17**, 9129–9134.
6. A. Saito, N. Nakajima, A. Tanaka and M. Ubukata, *Heterocycles*, 2003, **61**, 287–298.

¹H NMR of SI-3 (600 MHz CDCl₃)



Current Data Parameters
 NAME VB-BnPh
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 PROCNO 1

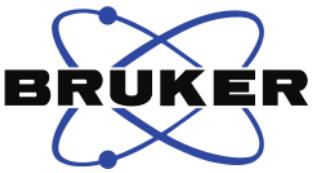
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 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 31.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 ======

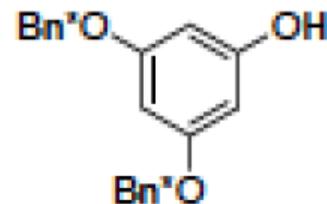
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 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

F2 - Processing parameters
 SI 65536
 SF 600.1300144 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C NMR of SI-3 (150 MHz, CDCl₃)



13C



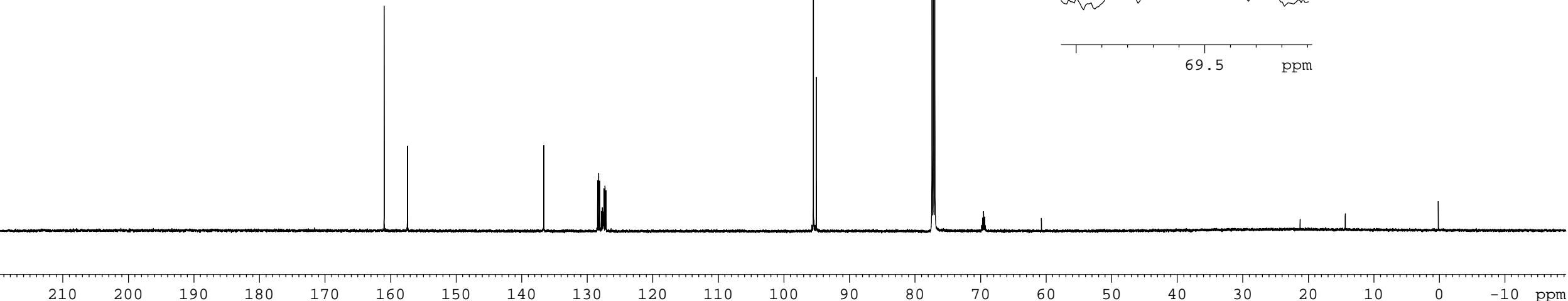
— 160.99
— 157.41

136.63
128.42
128.26
128.10
127.86
127.70
127.54
127.46
127.30
127.14

95.51
95.08

77.41
77.20
76.99
69.84
69.69
69.55
69.40
69.26

69.84
69.69
69.55
69.40
69.26



Current Data Parameters
NAME VB-BnPh
EXPNO 21
PROCNO 1

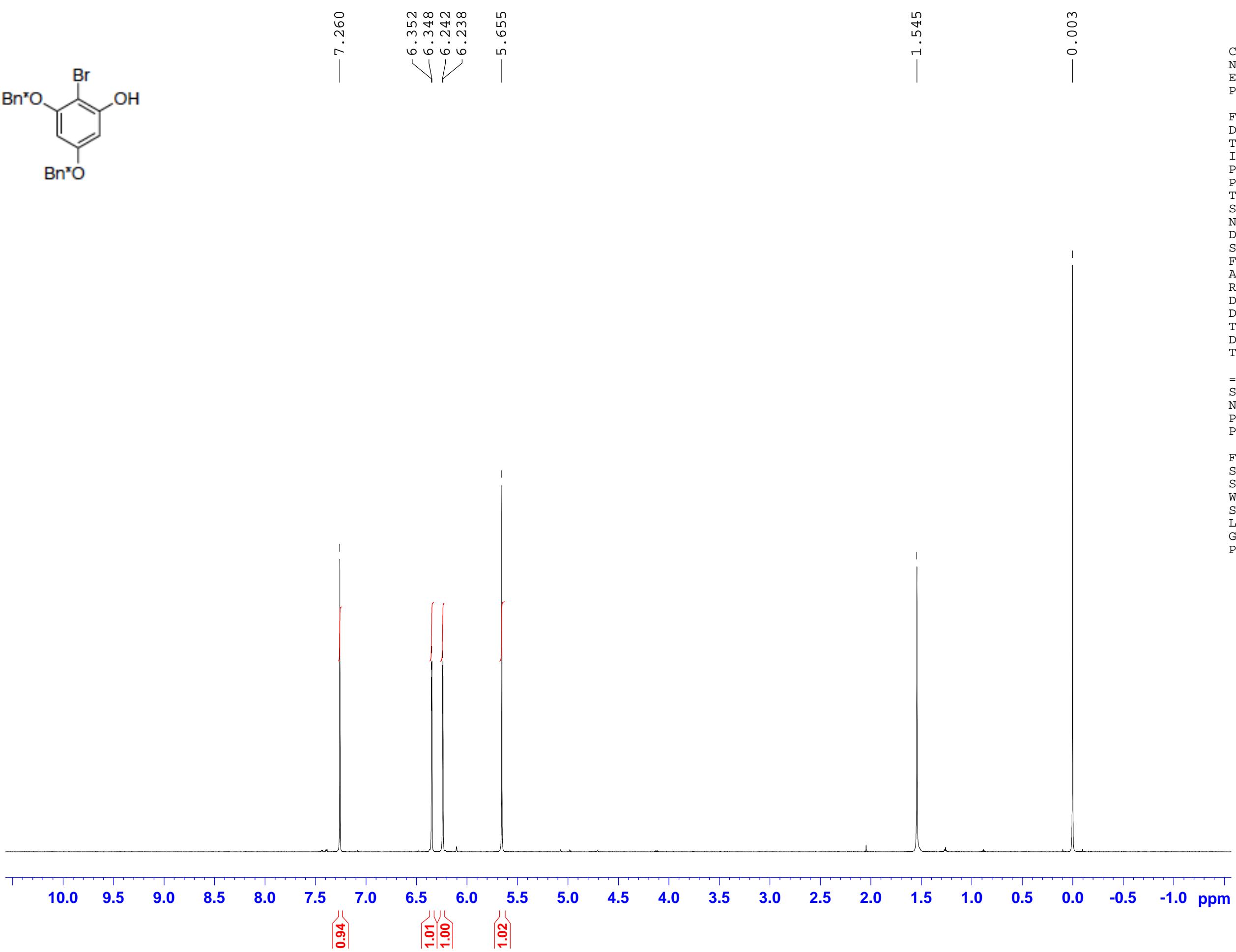
F2 - Acquisition Parameters
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Time 0.56
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PULPROG zpgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 4
SWH 36057.691 Hz
FIDRES 0.550197 Hz
AQ 0.9087659 sec
RG 175.56
DW 13.867 usec
DE 18.00 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 ====== SFO1 150.9178981 MHz
NUC1 13C
P1 10.00 usec
PLW1 80.00000000 W

===== CHANNEL f2 ====== SFO2 600.1324005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 70.00 usec
PLW2 13.43999958 W
PLW12 0.61714000 W
PLW13 0.31042001 W

F2 - Processing parameters
SI 32768
SF 150.9027843 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

¹H NMR of 5 (600 MHz, CDCl₃)



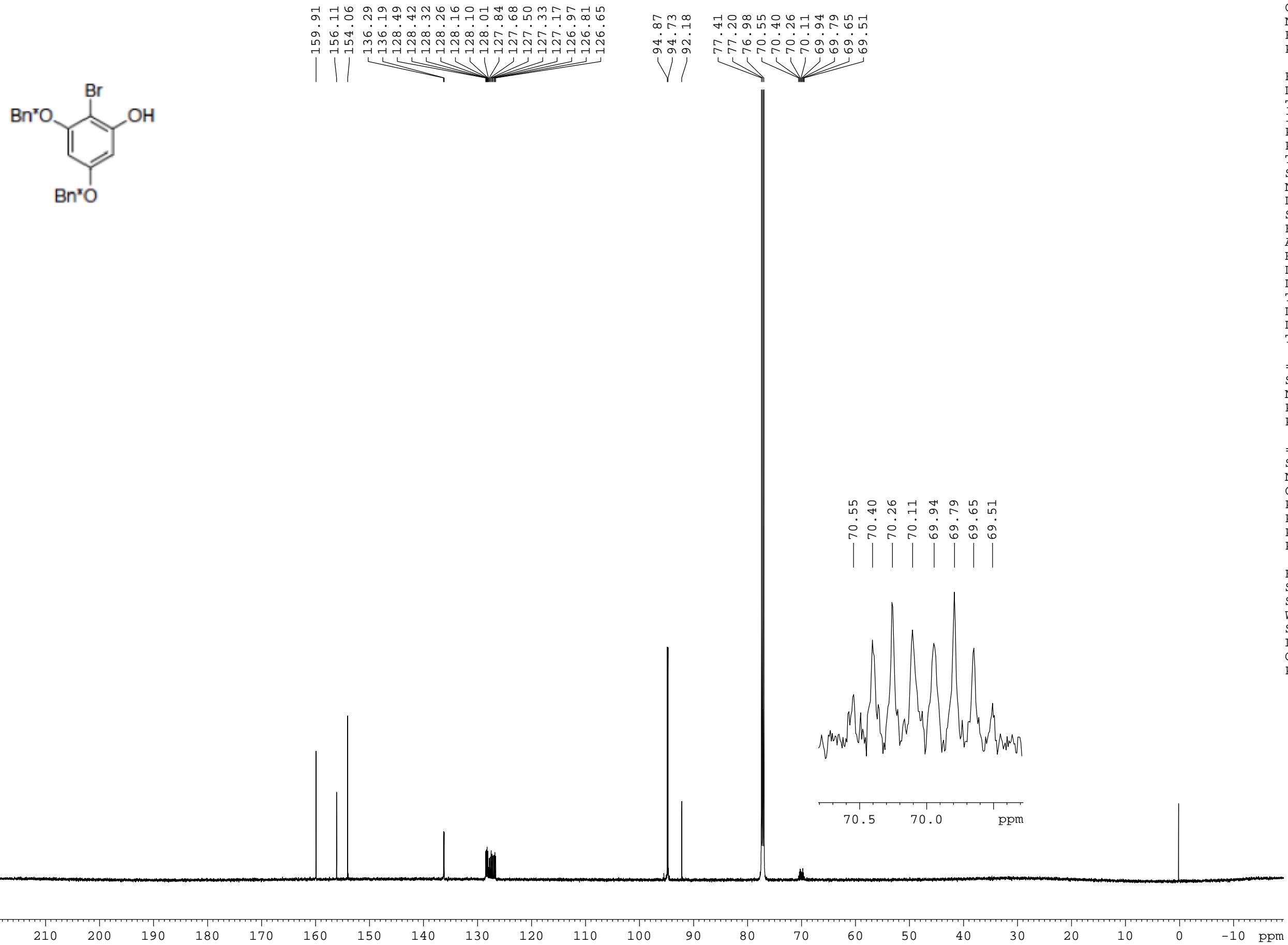
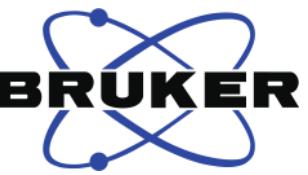
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PROCNO 1

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PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 12019.230 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 31.94
DW 41.600 usec
DE 10.00 usec
TE 298.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.00000000 W

F2 - Processing parameters
SI 65536
SF 600.1300149 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C NMR of 5 (150 MHz, CDCl₃)



¹H NMR of SI-5 (600 MHz, CDCl₃)

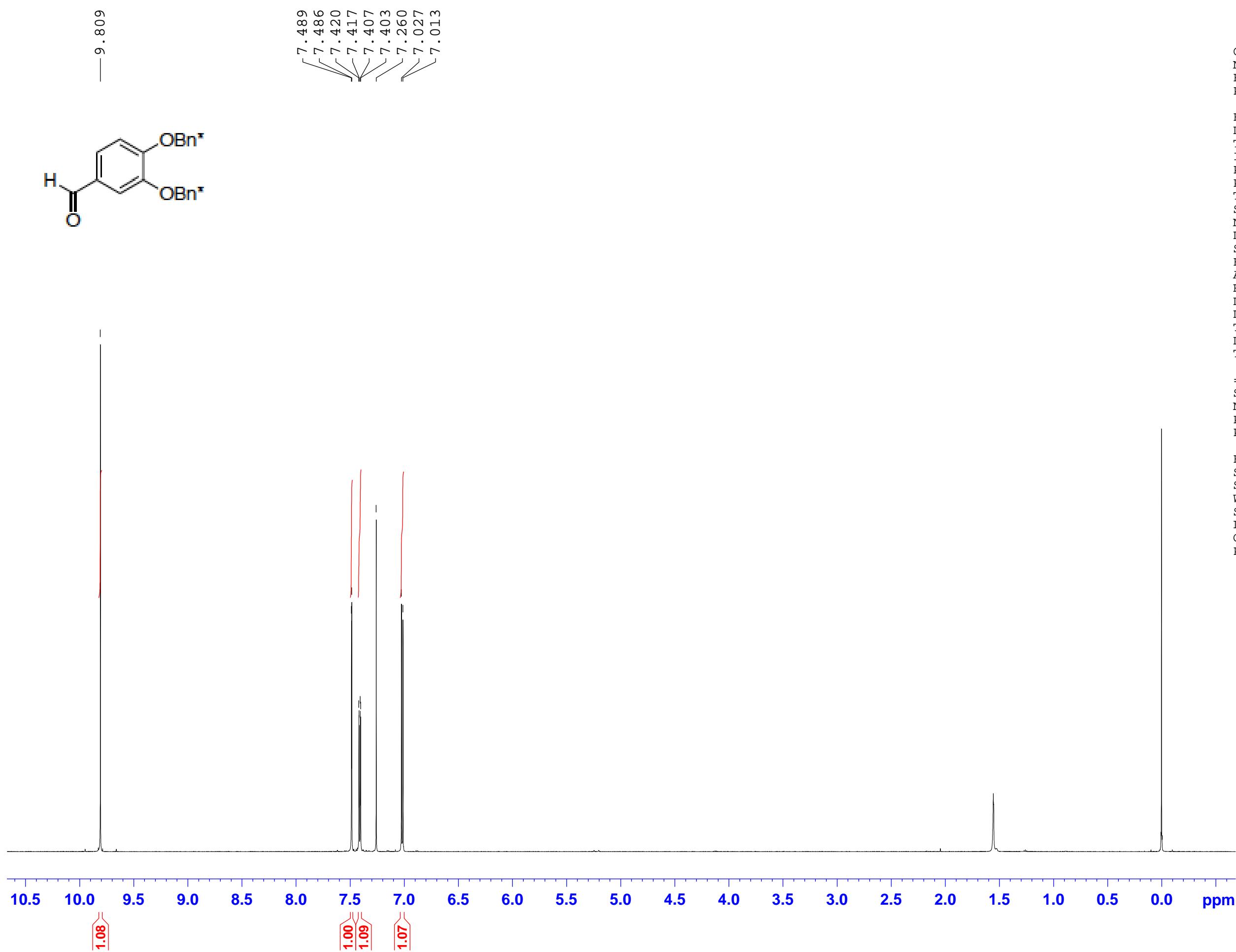


Current Data Parameters
 NAME VB-aldehyde
 EXPNO 10
 PROCNO 1

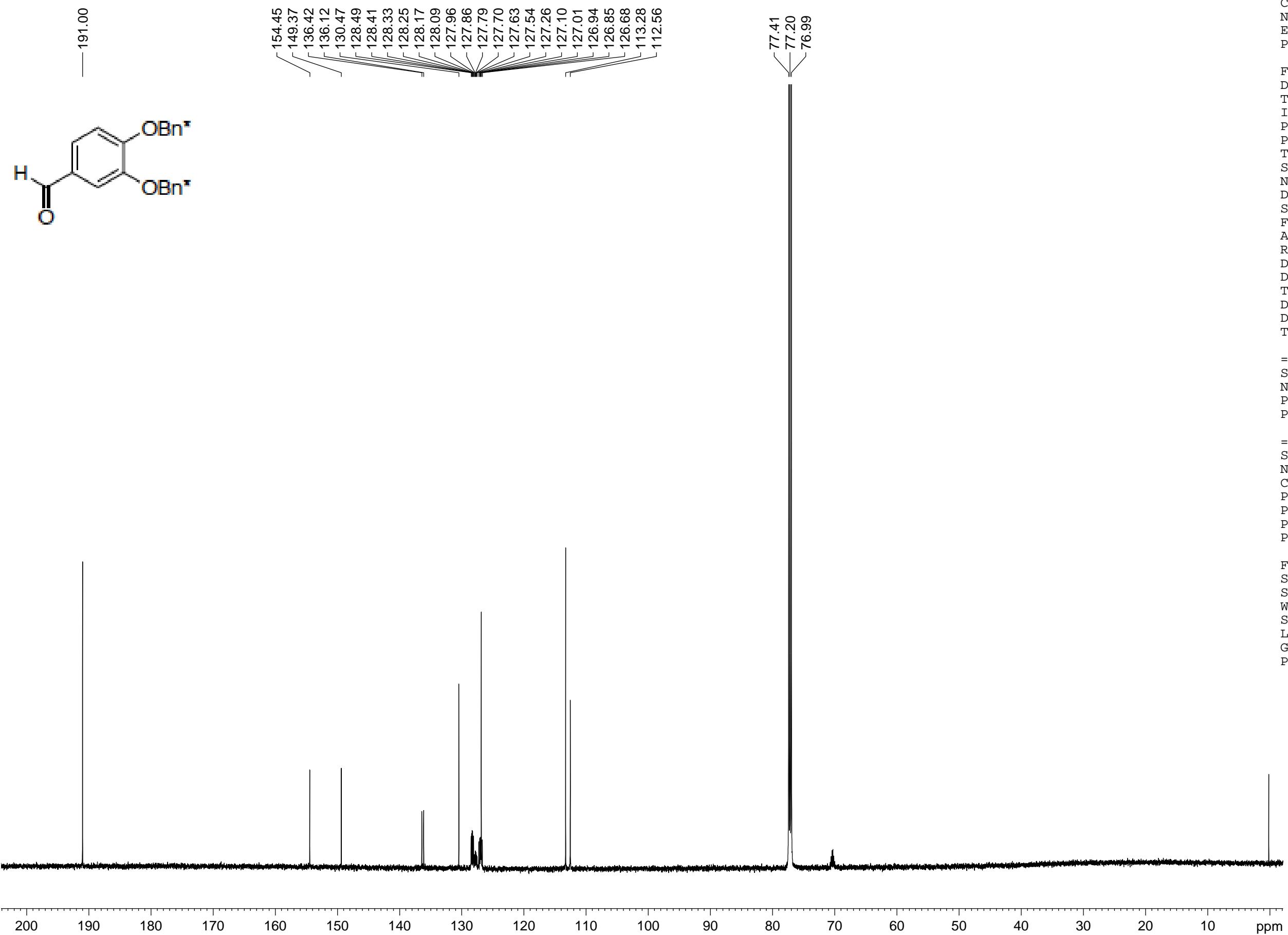
F2 - Acquisition Parameters
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 Time 15.44
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 31.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 ====== SFO1 600.1337060 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

F2 - Processing parameters
 SI 65536
 SF 600.1300144 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



¹³C NMR of SI-5 (150 MHz, CDCl₃)



Current Data Parameters
NAME VB-aldehyde
EXPNO 11
PROCNO 1

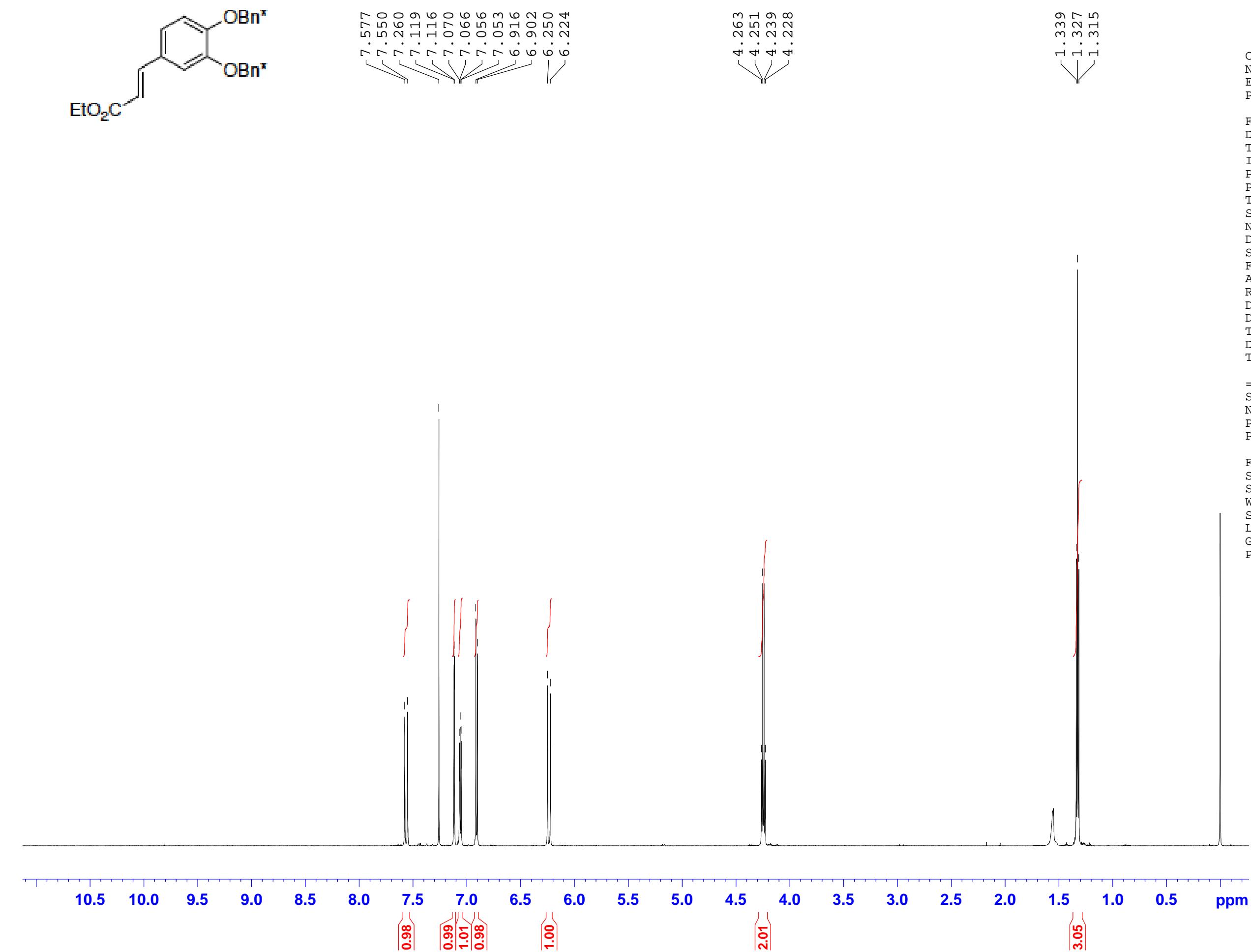
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Time 22.53
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PULPROG zpgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 4
SWH 36057.691 Hz
FIDRES 0.550197 Hz
AQ 0.9087659 sec
RG 175.56
DW 13.867 usec
DE 18.00 usec
TE 298.2 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 ======
SFO1 150.9178981 MHz
NUC1 ¹³C
P1 10.00 usec
PLW1 80.0000000 W

===== CHANNEL f2 ======
SFO2 600.1324005 MHz
NUC2 ¹H
CPDPRG[2] waltz16
PCPD2 70.00 usec
PLW2 13.43999958 W
PLW12 0.61714000 W
PLW13 0.31042001 W

F2 - Processing parameters
SI 32768
SF 150.9027831 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

¹H NMR of SI-6 (600 MHz, CDCl₃)



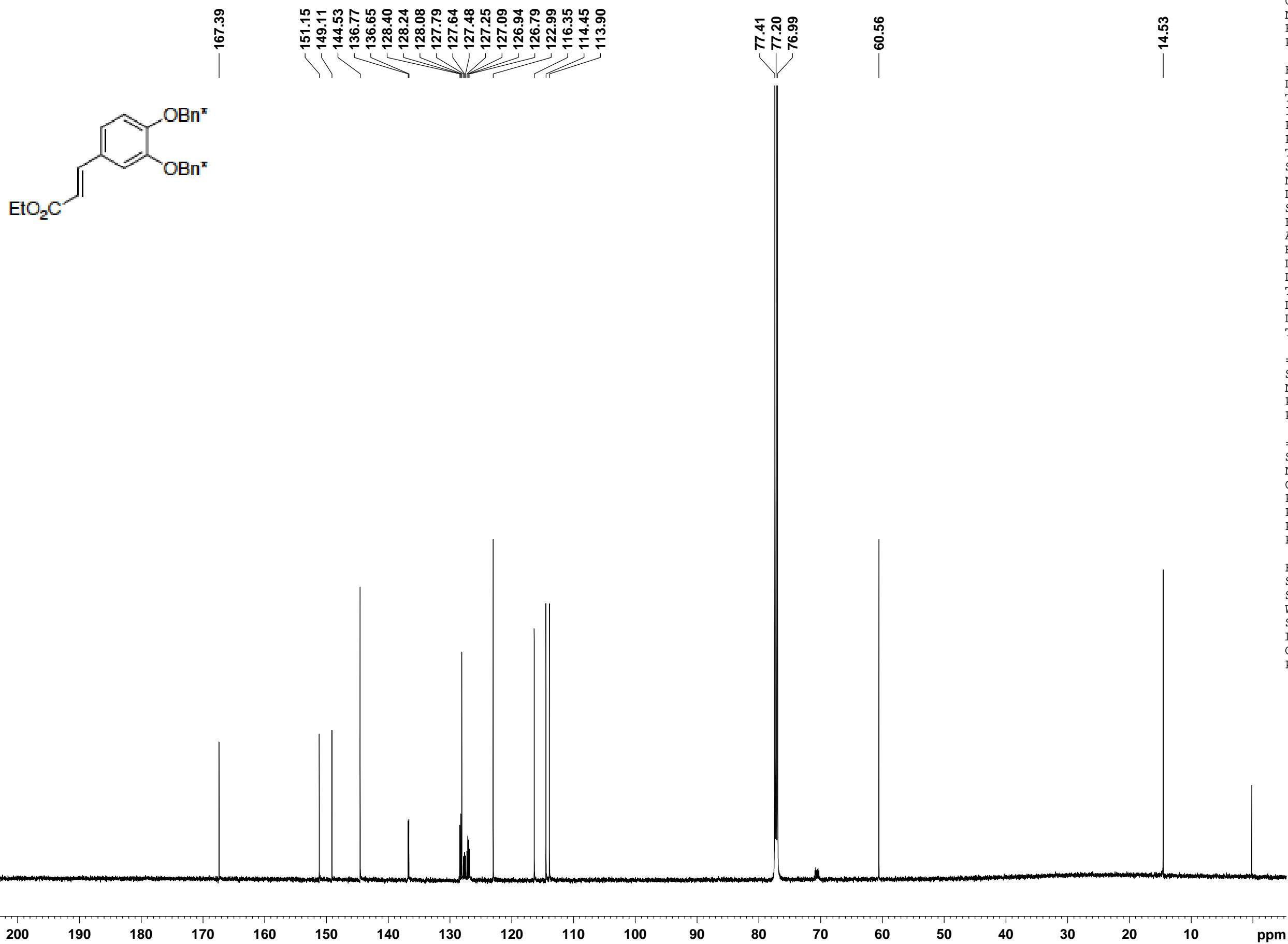
Current Data Parameters
NAME VB-ester
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
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Time 15.47
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PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 12019.230 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 31.94
DW 41.600 usec
DE 10.00 usec
TE 298.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.00000000 W

F2 - Processing parameters
SI 65536
SF 600.1300144 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C NMR of SI-6 (150 MHz, CDCl₃)



Current Data Parameters
 NAME VB-ester
 EXPNO 11
 PROCNO 1

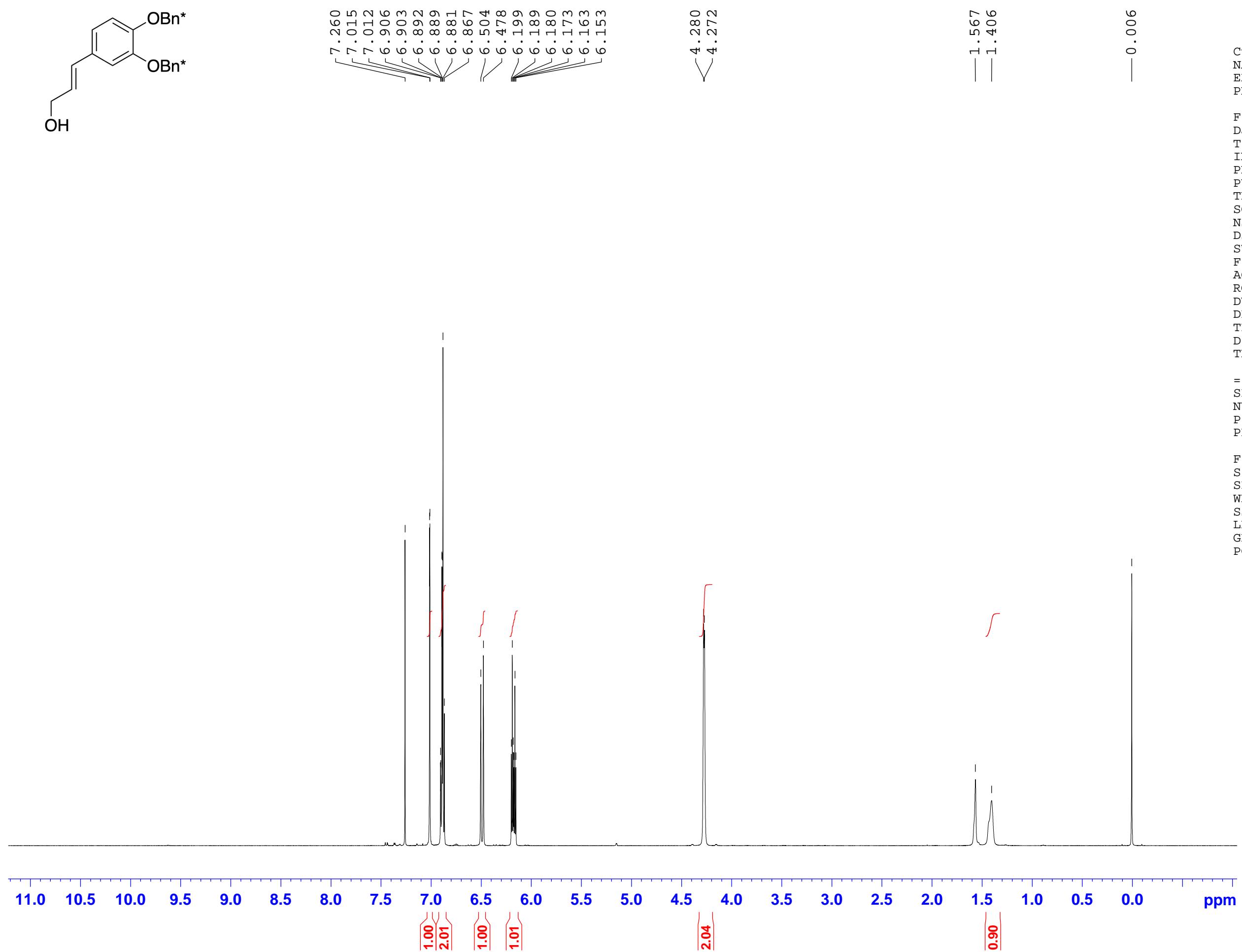
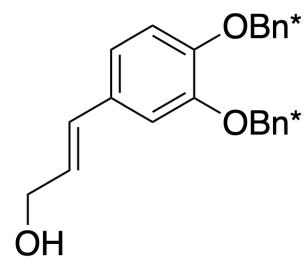
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 TD 65536
 SOLVENT CDCl₃
 NS 2000
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 175.56
 DW 13.867 usec
 DE 18.00 usec
 TE 298.2 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 ======
 SFO1 150.9178981 MHz
 NUC1 ¹³C
 P1 10.00 usec
 PLW1 80.0000000 W

===== CHANNEL f2 ======
 SFO2 600.1324005 MHz
 NUC2 ¹H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 13.43999958 W
 PLW12 0.61714000 W
 PLW13 0.31042001 W

F2 - Processing parameters
 SI 32768
 SF 150.9027829 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

¹H NMR of SI-7 (600 MHz, CDCl₃)





Current	Data	Parameters
NAME	VB-845	
EXPNO	20	
PROCNO	1	

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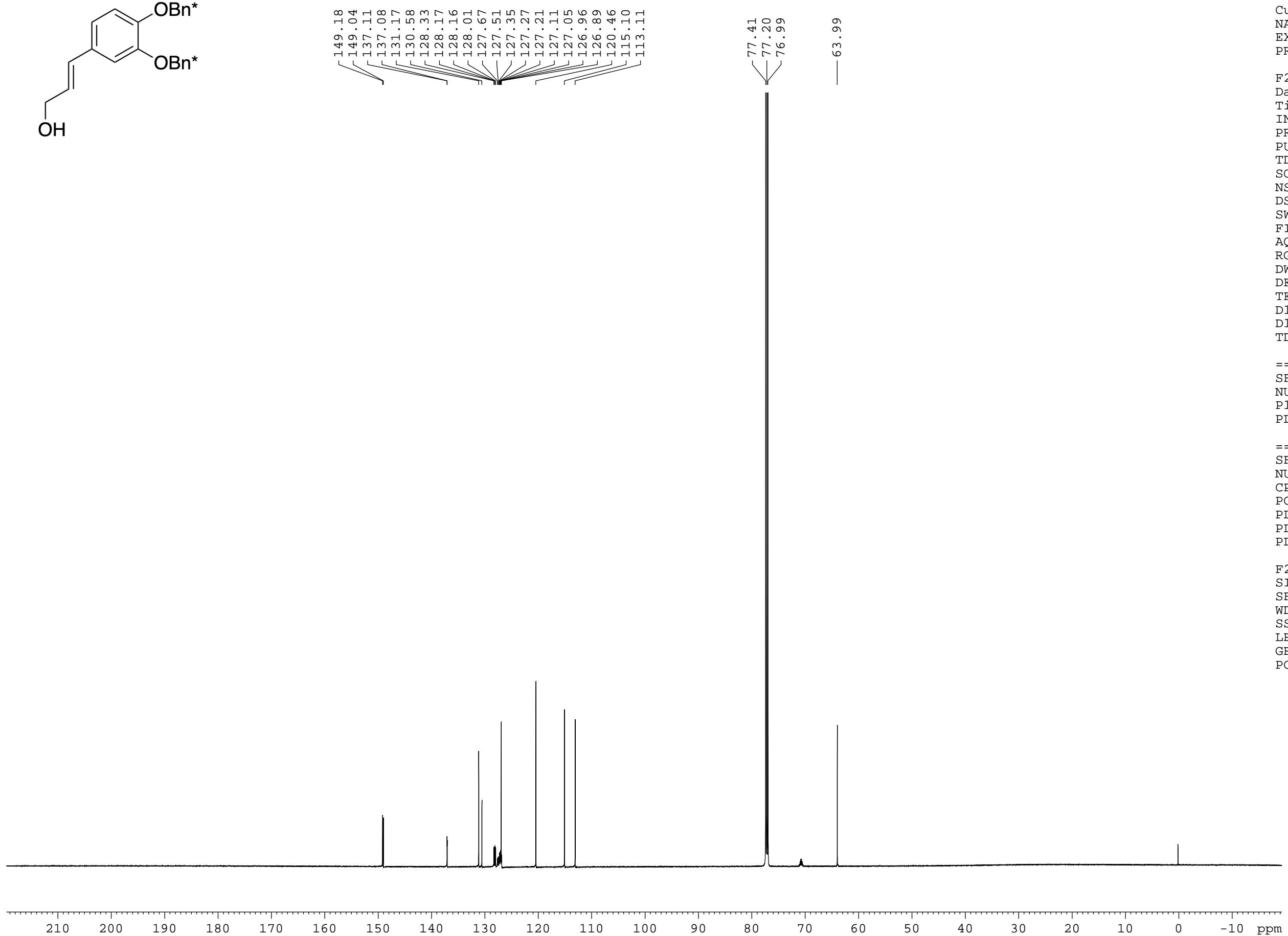
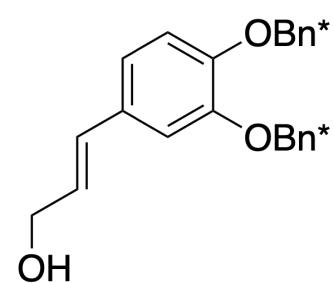
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Time            17.05
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PROBHD         5 mm CPPBBO BB
PULPROG        zg30
TD              65536
SOLVENT         CDCl3
NS              16
DS              2
SWH             12019.230 Hz
FIDRES        0.183399 Hz
AQ              2.7262976 sec
RG              31.94
DW              41.600 usec
DE              10.00 usec
TE              298.1 K
D1              1.00000000 sec
TD0                 1

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===== CHANNEL f1 ======
SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.000000000 W

```
F2 - Processing parameters
SI           65536
SF          600.1300144 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB          0
PC          1.00
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¹³C NMR of SI-7 (150 MHz, CDCl₃)



Current Data Parameters
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 PROCNO 1

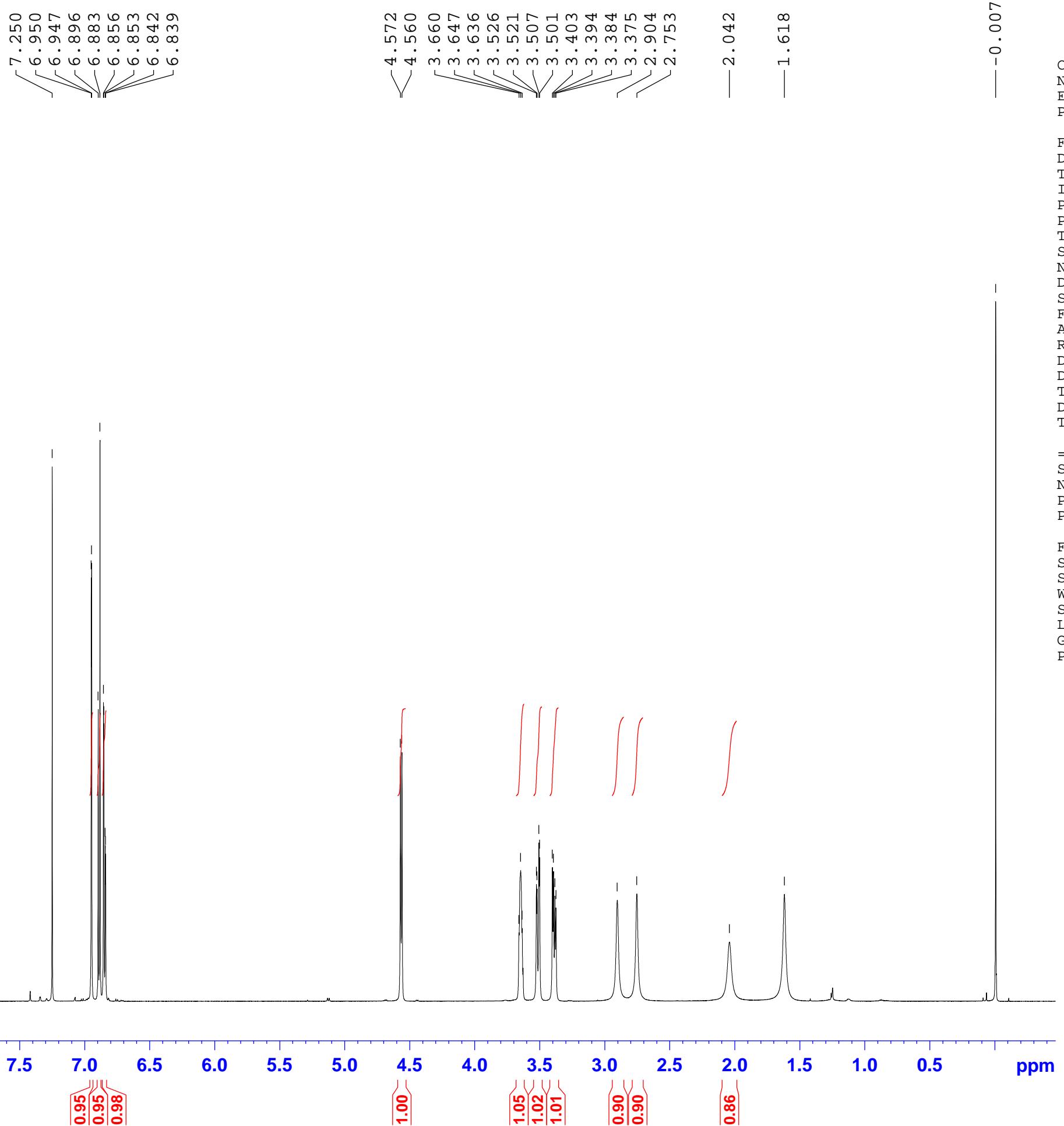
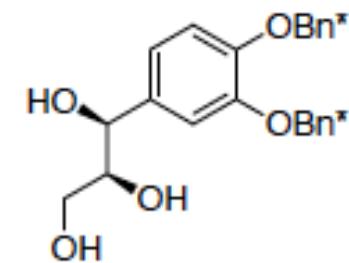
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 PULPROG zpgpg30
 TD 65536
 SOLVENT CDCl3
 NS 4200
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 175.56
 DW 13.867 usec
 DE 18.00 usec
 TE 298.1 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 ======
 SFO1 150.9178981 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 80.0000000 W

===== CHANNEL f2 ======
 SFO2 600.1324005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 13.43999958 W
 PLW12 0.61714000 W
 PLW13 0.31042001 W

F2 - Processing parameters
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 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

¹H NMR of SI-8 (600 MHz, CDCl₃)



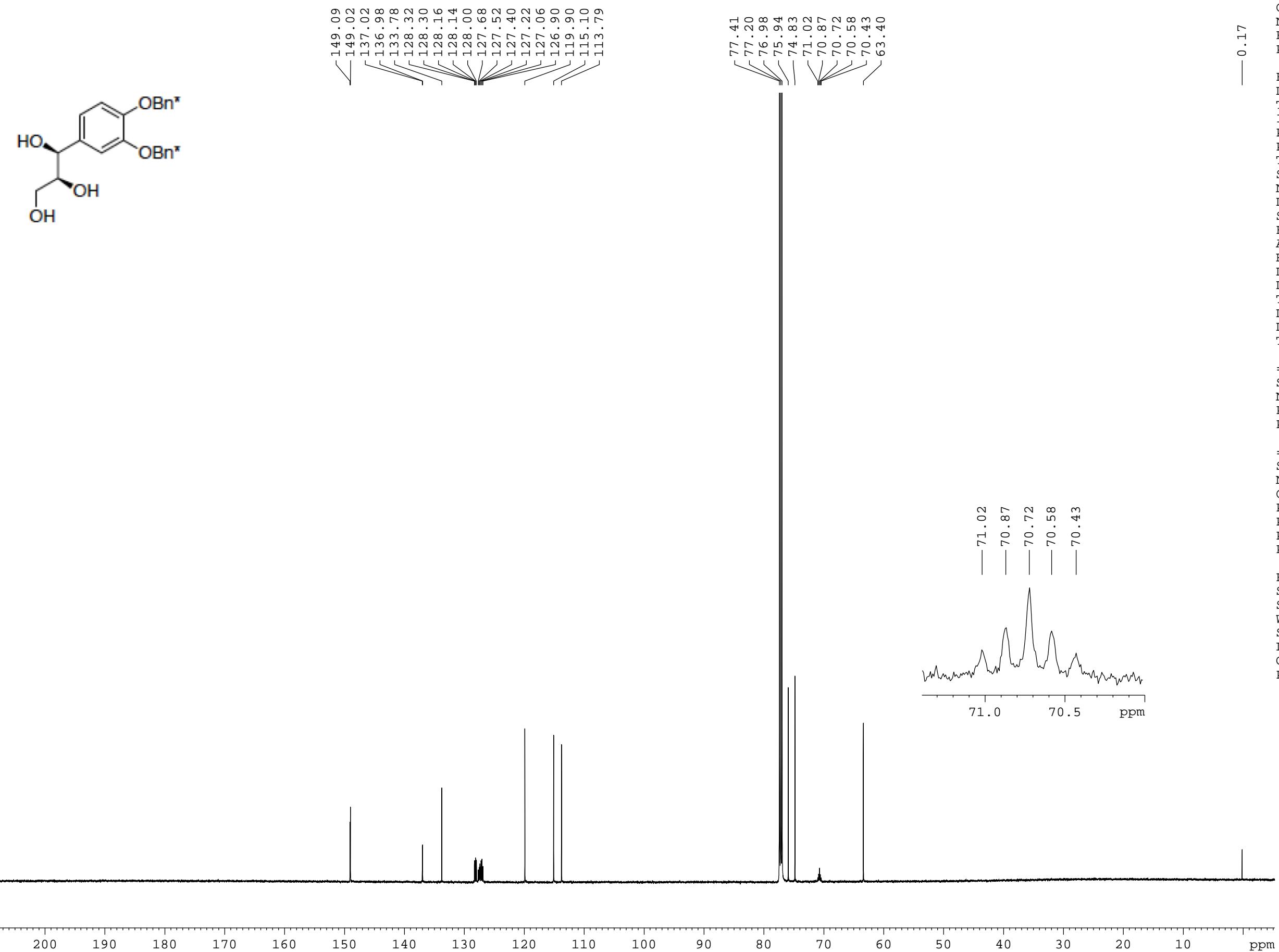
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 NAME VB-814
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 PROCNO 1

F2 - Acquisition Parameters
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 Time 19.53
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 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 31.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 ====== SFO1 600.1337060 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

F2 - Processing parameters
 SI 65536
 SF 600.1300205 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C NMR of SI-8 (150 MHz, CDCl₃)



Current Data Parameters
 NAME VB-814
 EXPNO 21
 PROCNO 1

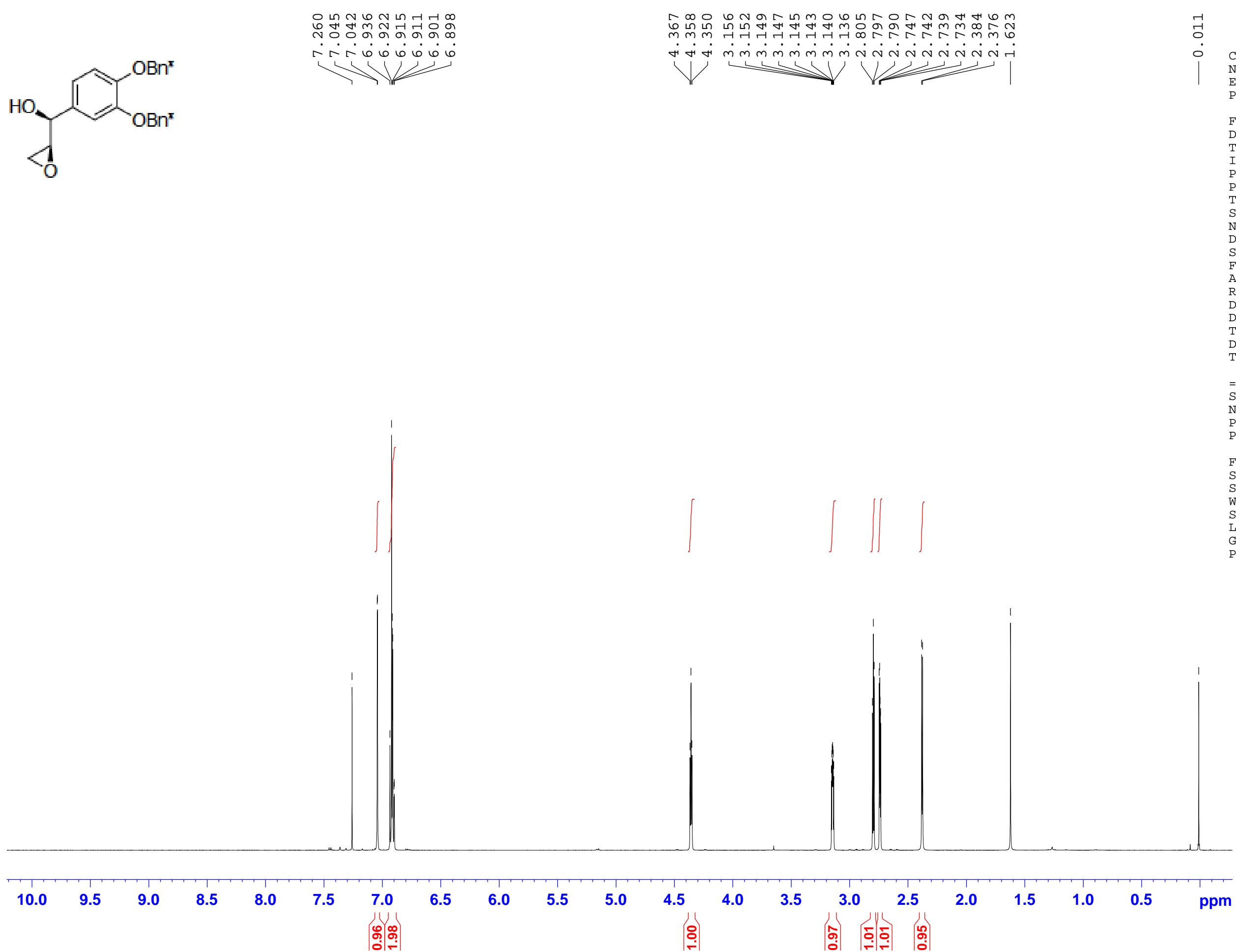
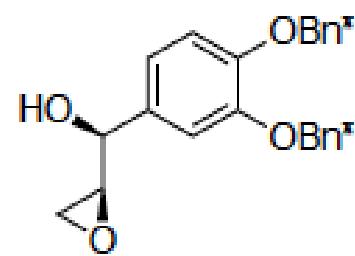
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 PULPROG zpgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1700
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 175.56
 DW 13.867 usec
 DE 18.00 usec
 TE 298.1 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 ======
 SFO1 150.9178981 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 80.0000000 W

===== CHANNEL f2 ======
 SFO2 600.1324005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 13.43999958 W
 PLW12 0.61714000 W
 PLW13 0.31042001 W

F2 - Processing parameters
 SI 32768
 SF 150.9027843 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

¹H NMR of 6 (600 MHz, CDCl₃)



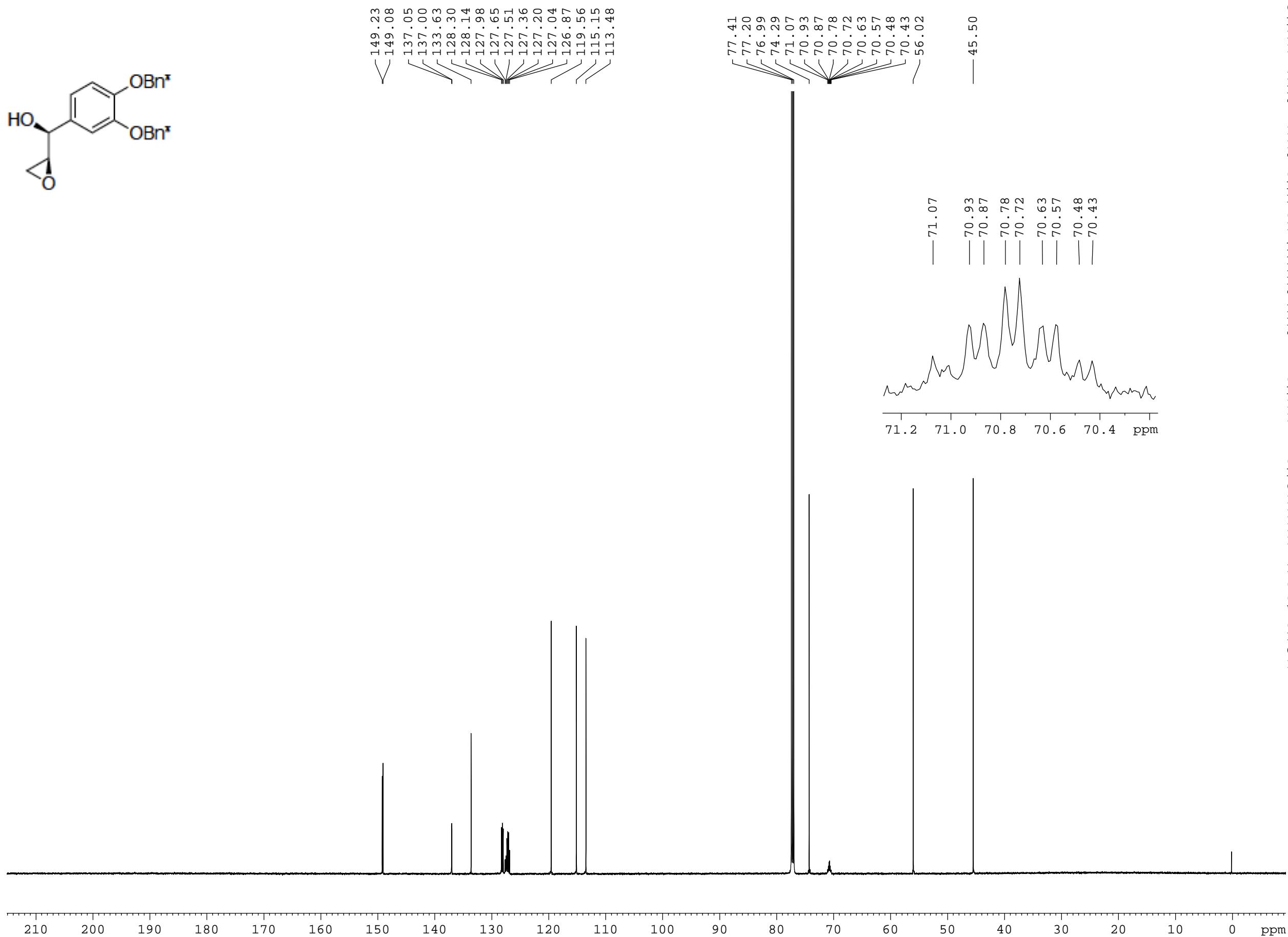
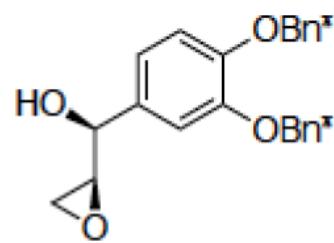
Current Data Parameters
NAME VB-818
EXPNO 20
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210531
Time 16.50
INSTRUM spect
PROBHD 5 mm CPPBBO BB
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 12019.230 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 17.5
DW 41.600 usec
DE 10.00 usec
TE 298.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 ====== SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.00000000 W

F2 - Processing parameters
SI 65536
SF 600.1300145 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C NMR of 6 (150 MHz, CDCl₃)



Current	Data	Parameters
NAME		VB-818
EXPNO		21
PROCNO		1

```

F2 - Acquisition Parameters
Date_           20210531
Time            22.53
INSTRUM        spect
PROBHD         5 mm CPPBBO BB
PULPROG        zgpg30
TD              65536
SOLVENT         CDC13
NS              1024
DS                  4
SWH             36057.691 Hz
FIDRES        0.550197 Hz
AQ              0.9087659 sec
RG                 175.56
DW              13.867 usec
DE                 18.00 usec
TE                 298.2 K
D1      2.000000000 sec
D11     0.030000000 sec
TD0                  1

```

```
===== CHANNEL f1 =====  
SFO1          150.9178981 MHz  
NUC1          13C  
P1            10.00 usec  
PI.W1         80 00000000 W
```

```

===== CHANNEL f2 =====
SFO2          600.1324005 MHz
NUC2          1H
CPDPRG[2      waltz16
PCPD2         70.00 usec
PLW2          13.43999958 W
PLW12         0.61714000 W
PLW13         0.31042001 W

```

```

F2 - Processing parameters
SI           32768
SF          150.9027873 MHz
WDW          EM
SSB           0
LB           1.00 Hz
GB           0
PC           1.40

```

¹H NMR of 7 (600 MHz, CDCl₃)

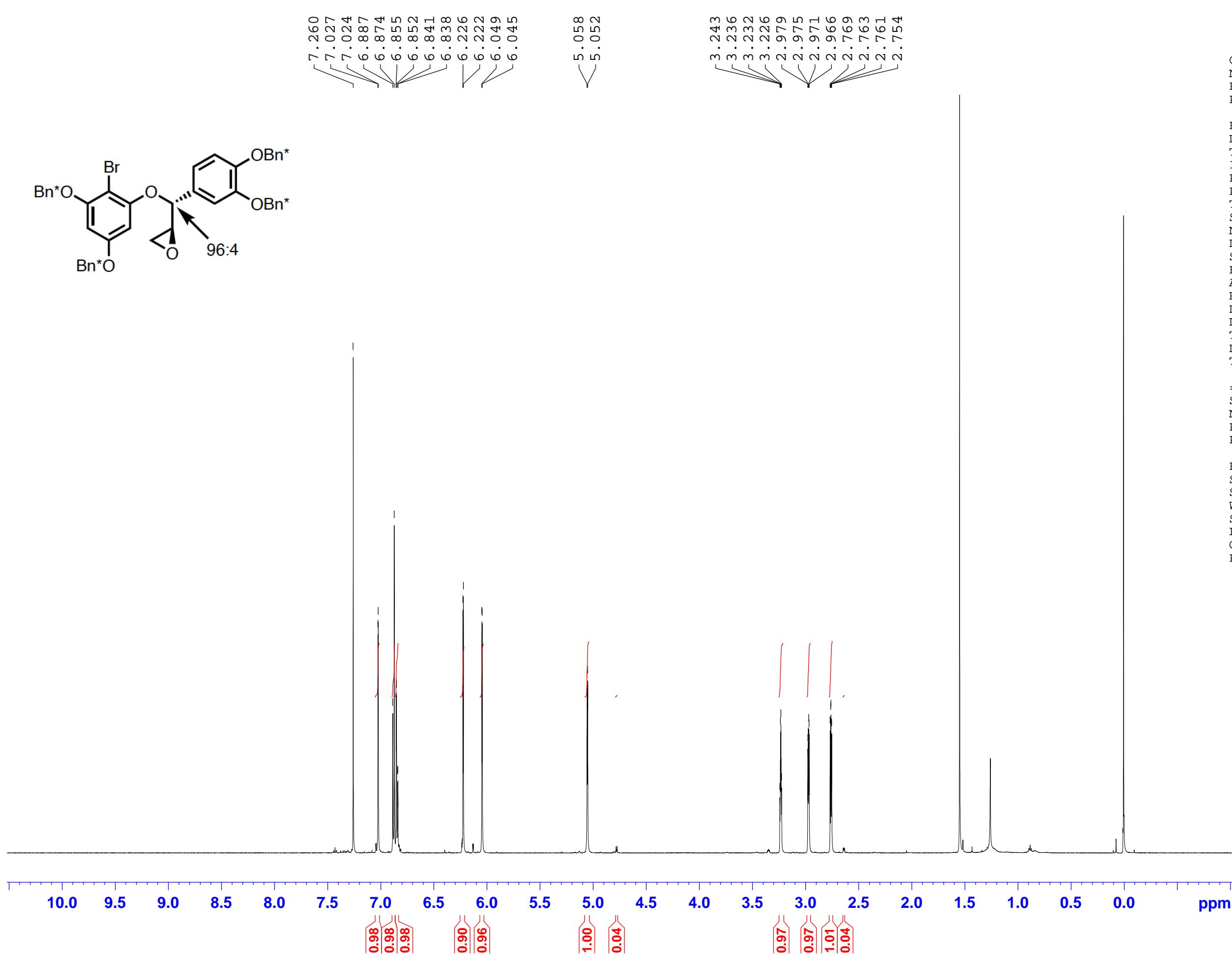
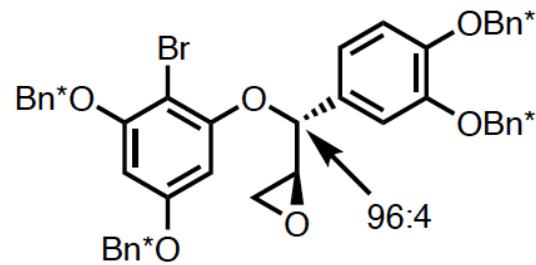


Current Data Parameters
 NAME VB-670
 EXPNO 30
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200929
 Time 18.53
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 31.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 600.1337060 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

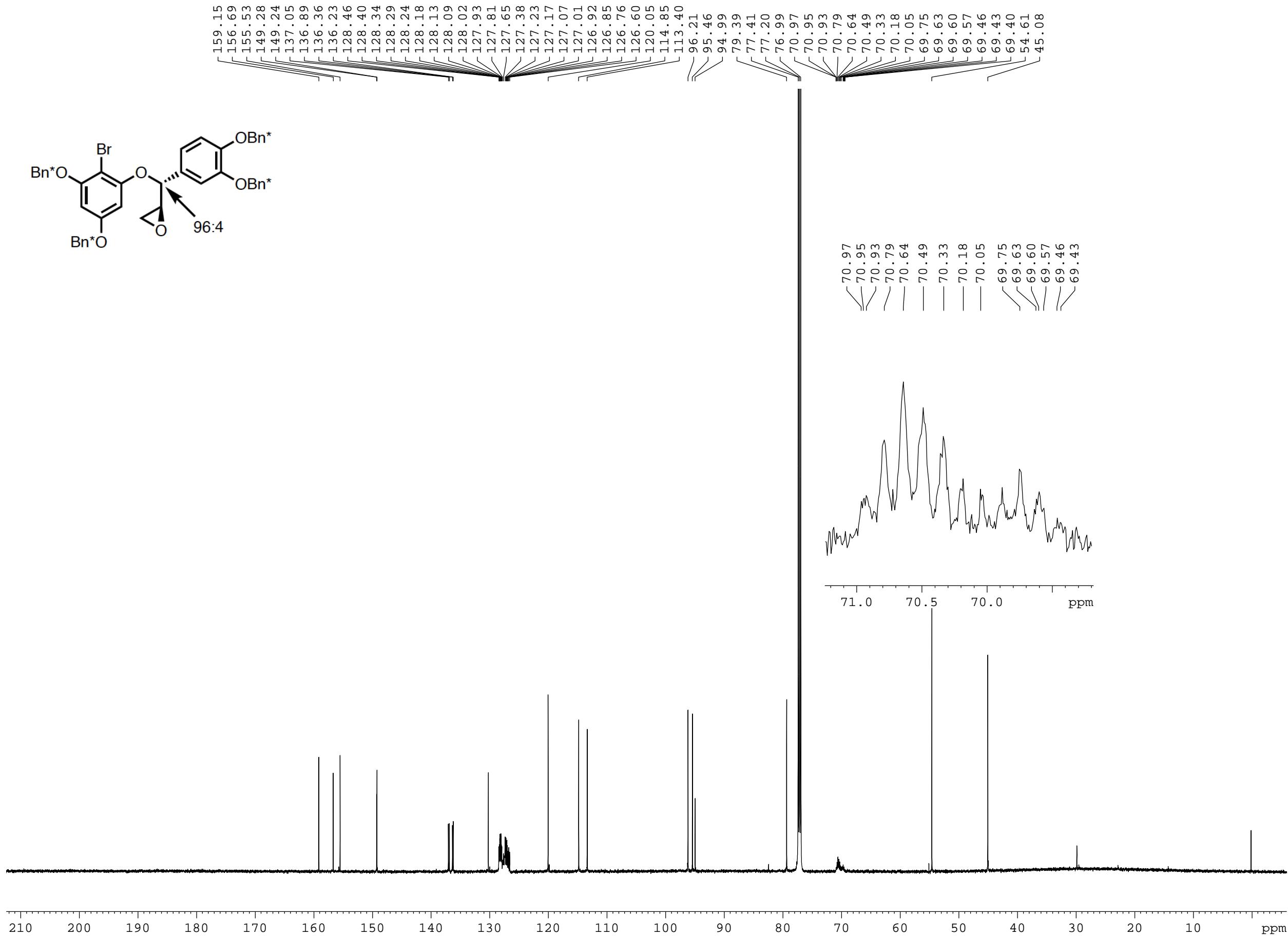
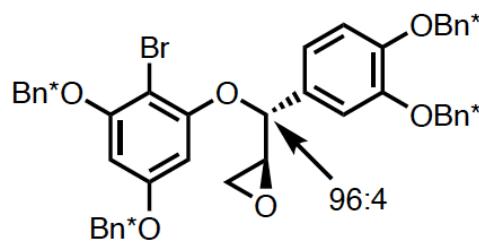
F2 - Processing parameters
 SI 65536
 SF 600.1300142 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



¹³C NMR of 7 (150 MHz, CDCl₃)



Bruker



Current Data Parameters
NAME VB-670
EXPNO 31
PROCNO 1

```

F2 - Acquisition Parameters
Date_           20200930
Time            9.06
INSTRUM        spect
PROBHD         5 mm CPPBBO BB
PULPROG        zgpg30
TD              65536
SOLVENT         CDCl3
NS              4800
DS                  4
SWH             36057.691 Hz
FIDRES        0.550197 Hz
AQ              0.9087659 sec
RG              175.56
DW              13.867 usec
DE              18.00 usec
TE              298.1 K
D1              2.00000000 sec
D11             0.03000000 sec
TD0                  1

```

```
===== CHANNEL f1 ======  
SFO1      150.9178981 MHz  
NUC1      13C  
P1        10.00 usec  
PLW1      80.000000000 W
```

```

===== CHANNEL f2 =====
SFO2          600.1324005 MHz
NUC2          1H
CPDPRG[2     waltz16
PCPD2         70.00 usec
PLW2          13.43999958 W
PLW12         0.61714000 W
PLW13         0.31042001 W

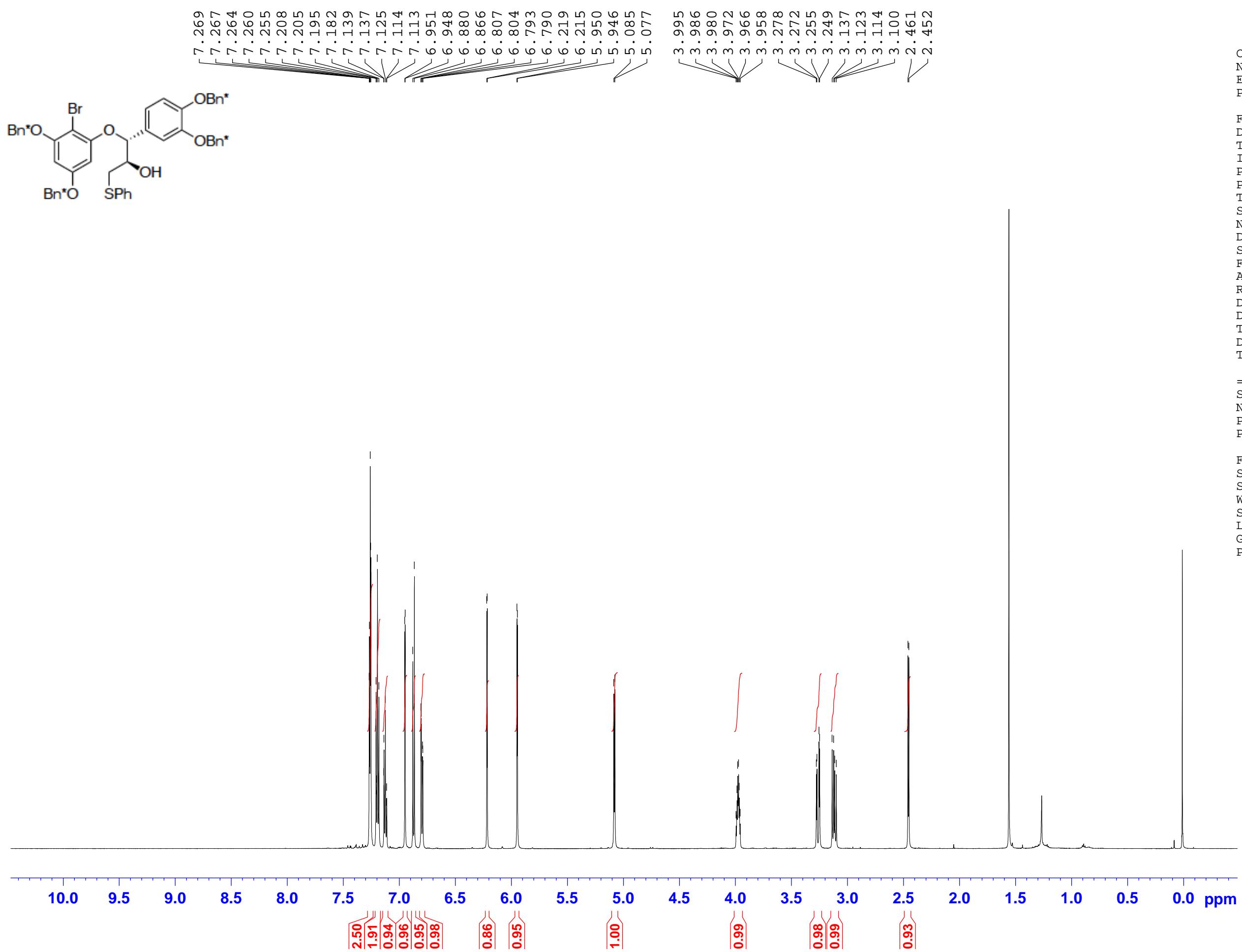
```

```

F2 - Processing parameters
SI           32768
SF          150.9027831 MHz
WDW          EM
SSB           0
LB           1.00 Hz
GB           0
PC           1.40

```

¹H NMR of 8 (600 MHz, CDCl₃)



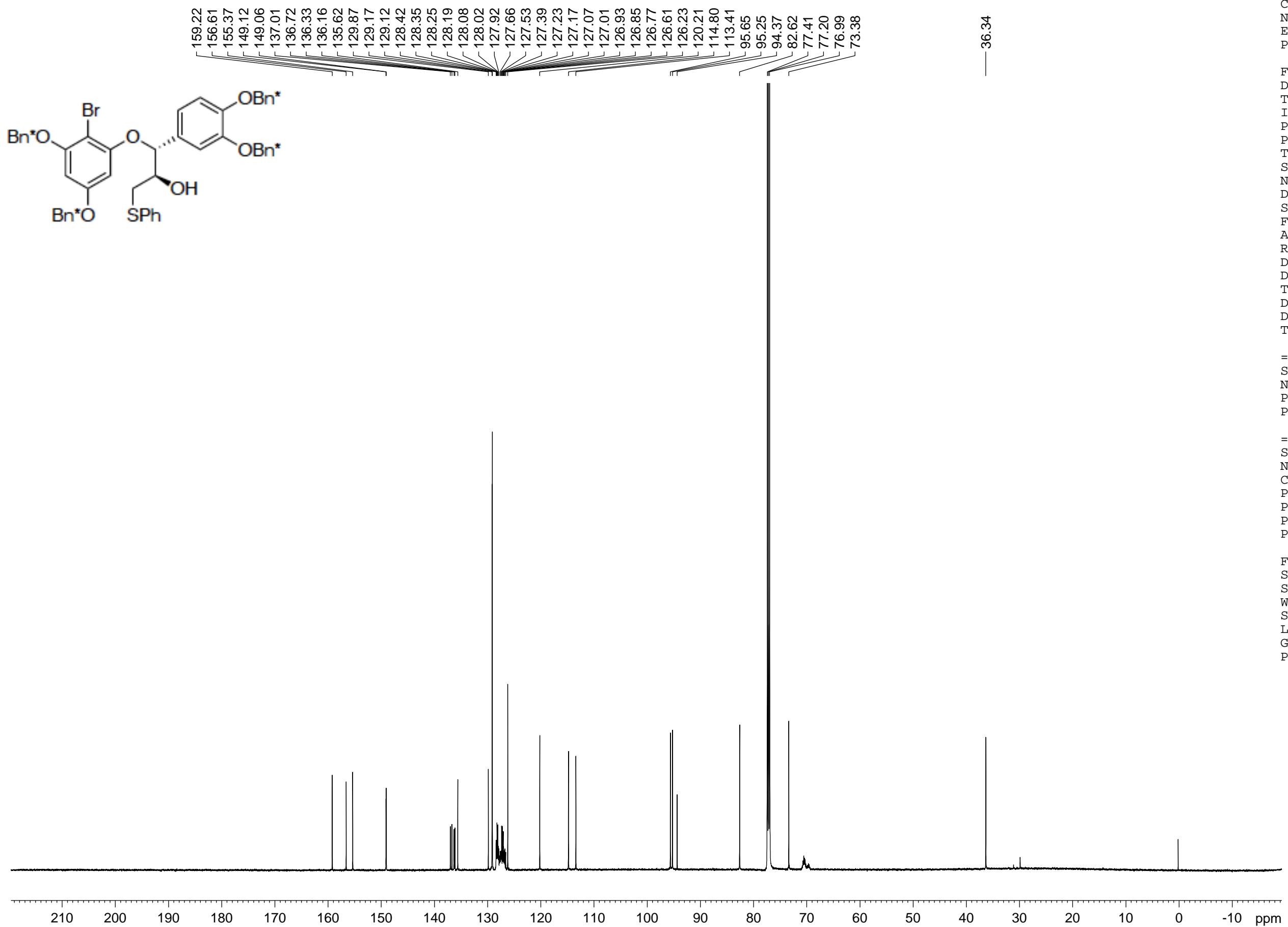
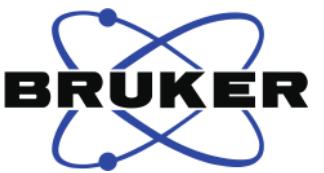
Current Data Parameters
NAME VB-671-2
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20201001
Time 11.04
INSTRUM spect
PROBHD 5 mm CPPBBO BB
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 12019.230 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 31.94
DW 41.600 usec
DE 10.00 usec
TE 298.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 ====== SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.00000000 W

F2 - Processing parameters
SI 65536
SF 600.1300142 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C NMR of 8 (150 MHz, CDCl₃)



Current Data Parameters
 NAME VB-671-2
 EXPNO 11
 PROCNO 1

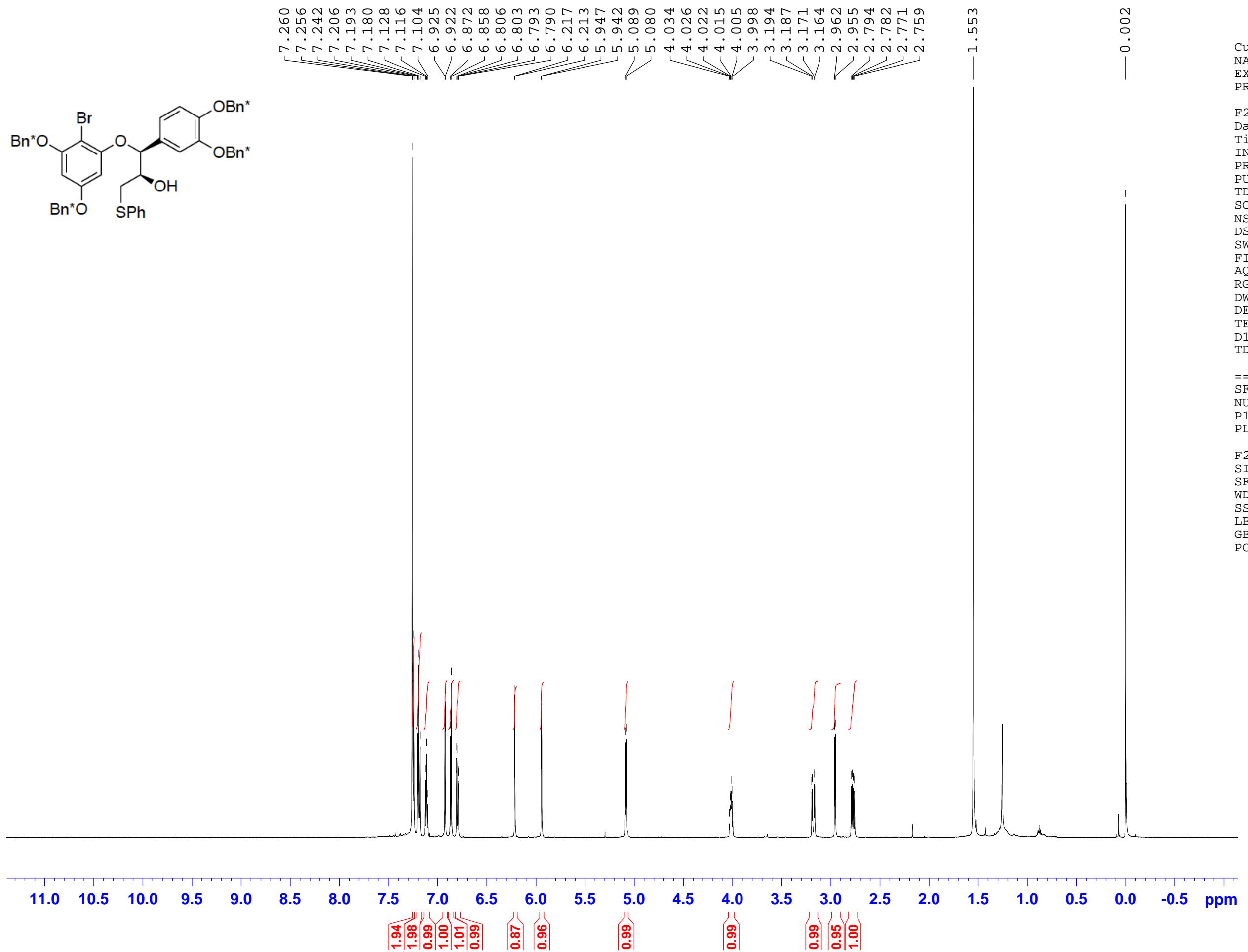
F2 - Acquisition Parameters
 Date_ 20201002
 Time 3.10
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 5000
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 175.56
 DW 13.867 usec
 DE 18.00 usec
 TE 298.1 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 ======
 SFO1 150.9178981 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 80.0000000 W

===== CHANNEL f2 ======
 SFO2 600.1324005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 13.43999958 W
 PLW12 0.61714000 W
 PLW13 0.31042001 W

F2 - Processing parameters
 SI 32768
 SF 150.9027854 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

¹H NMR of 2-*epi*-8 (600 MHz, CDCl₃)



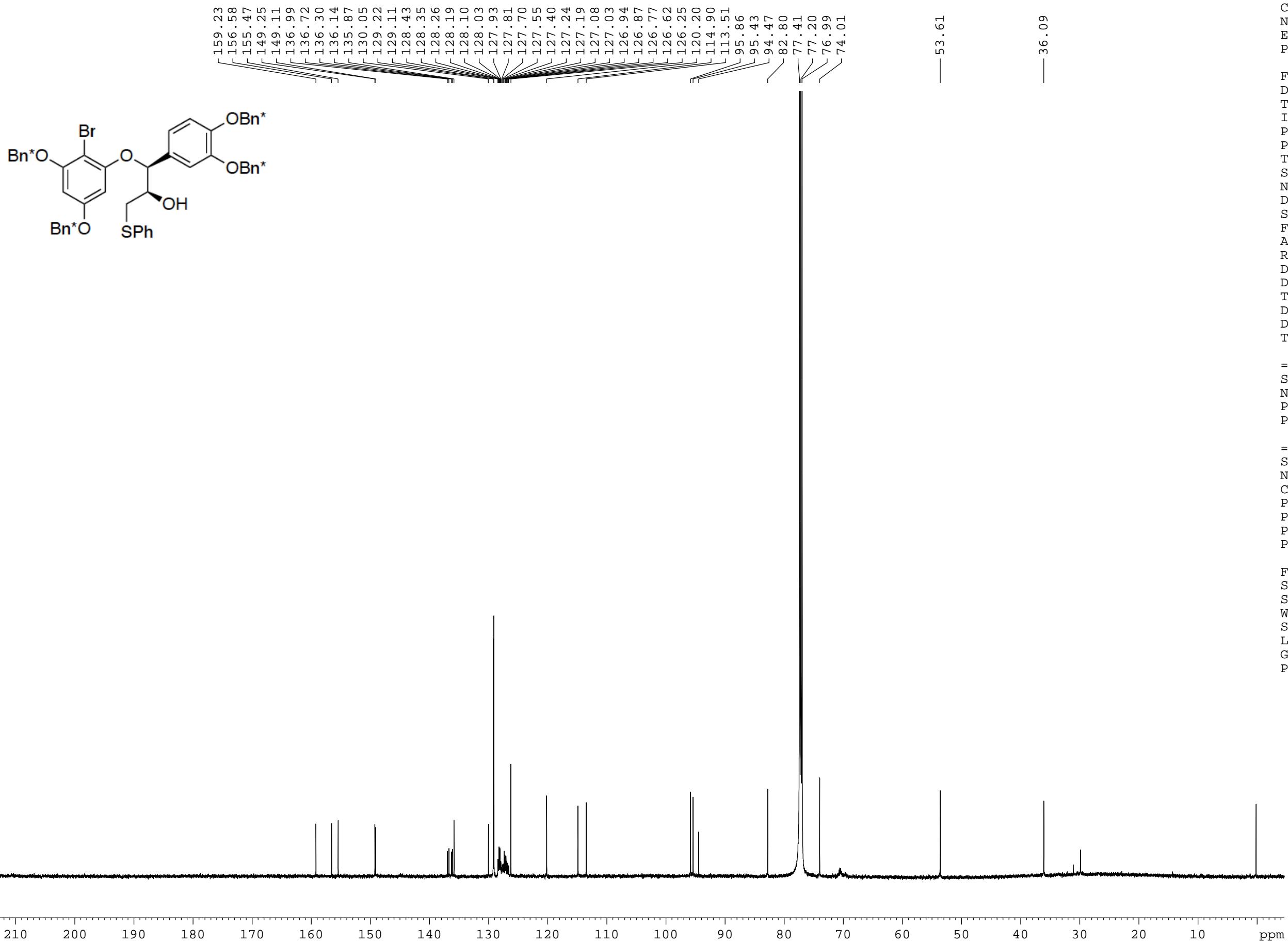
Current Data Parameters
 NAME VB-890
 EXPNO 30
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20211001
 Time 19.13
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 18.96
 DW 41.600 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 ====== SFO1 600.1337060 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

F2 - Processing parameters
 SI 65536
 SF 600.1300154 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C NMR of 2-*epi*-8 (150 MHz, CDCl₃)



Current Data Parameters
 NAME VB-890
 EXPNO 31
 PROCNO 1

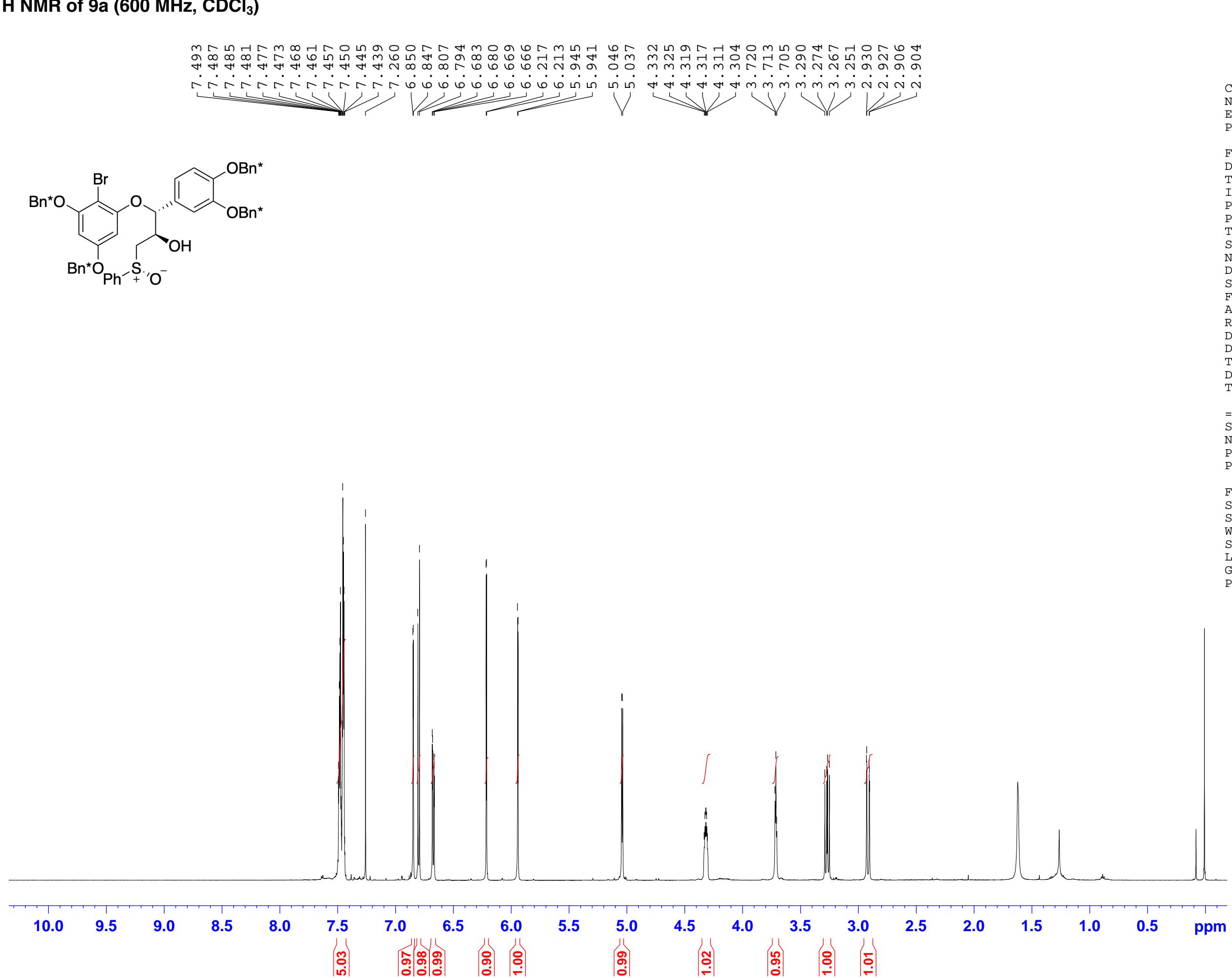
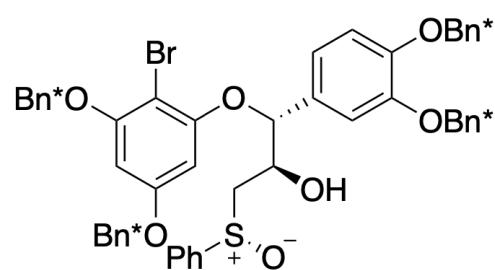
F2 - Acquisition Parameters
 Date_ 20211002
 Time 3.49
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zpgpg30
 TD 65536
 SOLVENT CDCl3
 NS 7000
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 175.56
 DW 13.867 usec
 DE 18.00 usec
 TE 298.0 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 ======
 SFO1 150.9178981 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 80.0000000 W

===== CHANNEL f2 ======
 SFO2 600.1324005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 13.43999958 W
 PLW12 0.61714000 W
 PLW13 0.31042001 W

F2 - Processing parameters
 SI 32768
 SF 150.9027825 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

¹H NMR of 9a (600 MHz, CDCl₃)



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Current	Data	Parameters
NAME	VB-824	
EXPNO	30	
PROCNO		1

```

F2 - Acquisition Parameters
Date_           20210610
Time            16.44
INSTRUM        spect
PROBHD         5 mm CPPBBO BB
PULPROG        zg30
TD              65536
SOLVENT         CDCl3
NS              16
DS              2
SWH             12019.230 Hz
FIDRES        0.183399 Hz
AQ              2.7262976 sec
RG              31.94
DW              41.600 usec
DE              10.00 usec
TE              298.1 K
D1              1.00000000 sec
TD0                 1

```

===== CHANNEL f1 =====
SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.000000000 W

```

F2 - Processing parameters
SI           65536
SF          600.1300144 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB          0
PC          1.00

```

¹³C NMR of 9a (150 MHz, CDCl₃)



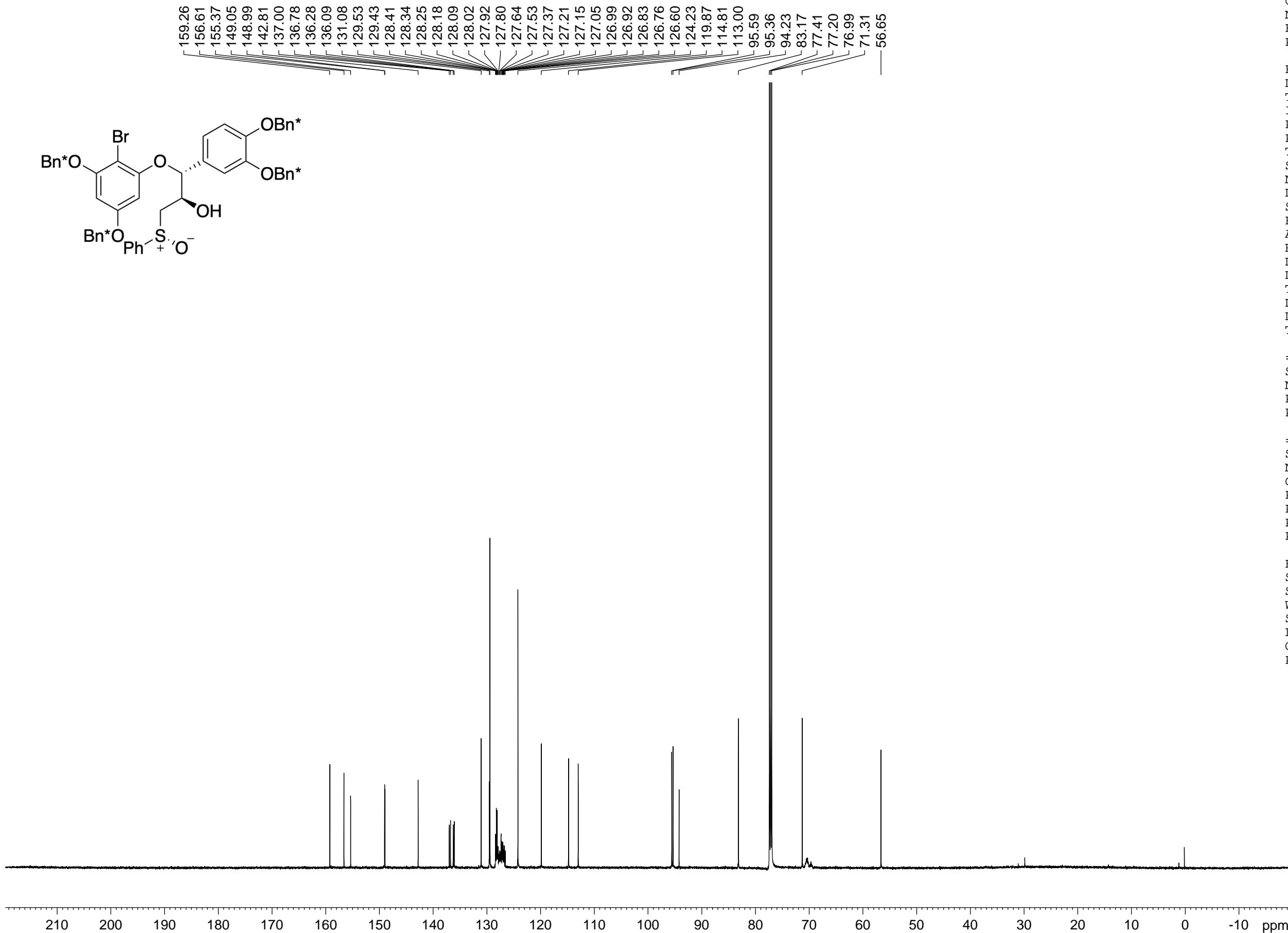
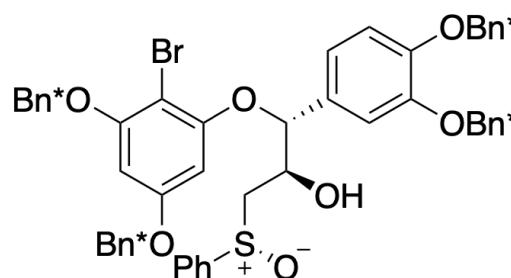
Current Data Parameters
 NAME VB-824
 EXPNO 31
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210610
 Time 23.41
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl₃
 NS 2000
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 175.56
 DW 13.867 usec
 DE 18.00 usec
 TE 298.2 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

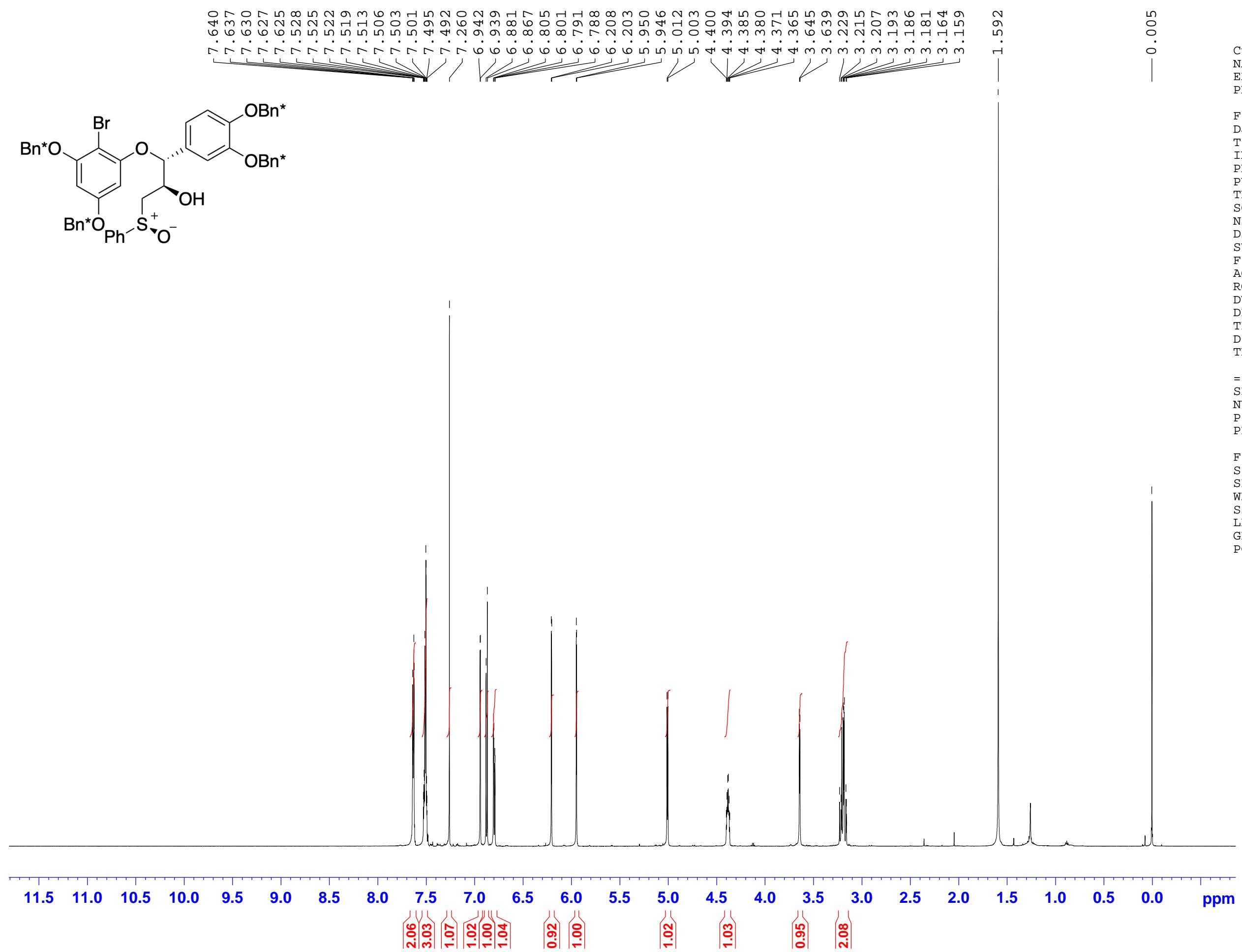
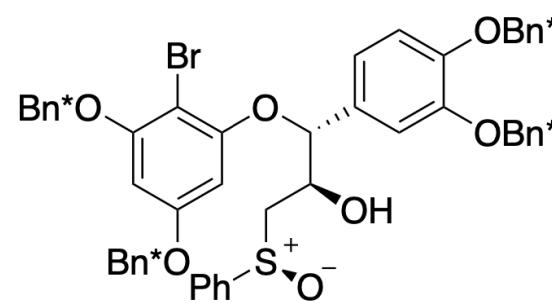
===== CHANNEL f1 ====== SFO1 150.9178981 MHz
 NUC1 ¹³C
 P1 10.00 usec
 PLW1 80.0000000 W

===== CHANNEL f2 ====== SFO2 600.1324005 MHz
 NUC2 ¹H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 13.43999958 W
 PLW12 0.61714000 W
 PLW13 0.31042001 W

F2 - Processing parameters
 SI 32768
 SF 150.9027860 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



¹H NMR of 9b (600 MHz, CDCl₃)



The Bruker logo consists of the word "BRUKER" in a bold, black, sans-serif font. Above the letter "B", there is a blue stylized atom symbol with three orbiting electrons.

Current	Data	Parameters
NAME	VB-847-2	
EXPNO	50	
PROCNO		1

```

F2 - Acquisition Parameters
Date_          20210906
Time           19.06
INSTRUM        spect
PROBHD         5 mm CPPBBO BB
PULPROG        zg30
TD             65536
SOLVENT        CDC13
NS              16
DS              2
SWH             12019.230 Hz
FIDRES        0.183399 Hz
AQ              2.7262976 sec
RG              31.94
DW              41.600 usec
DE              10.00 usec
TE              298.1 K
D1              1.00000000 sec
TD0                 1

```

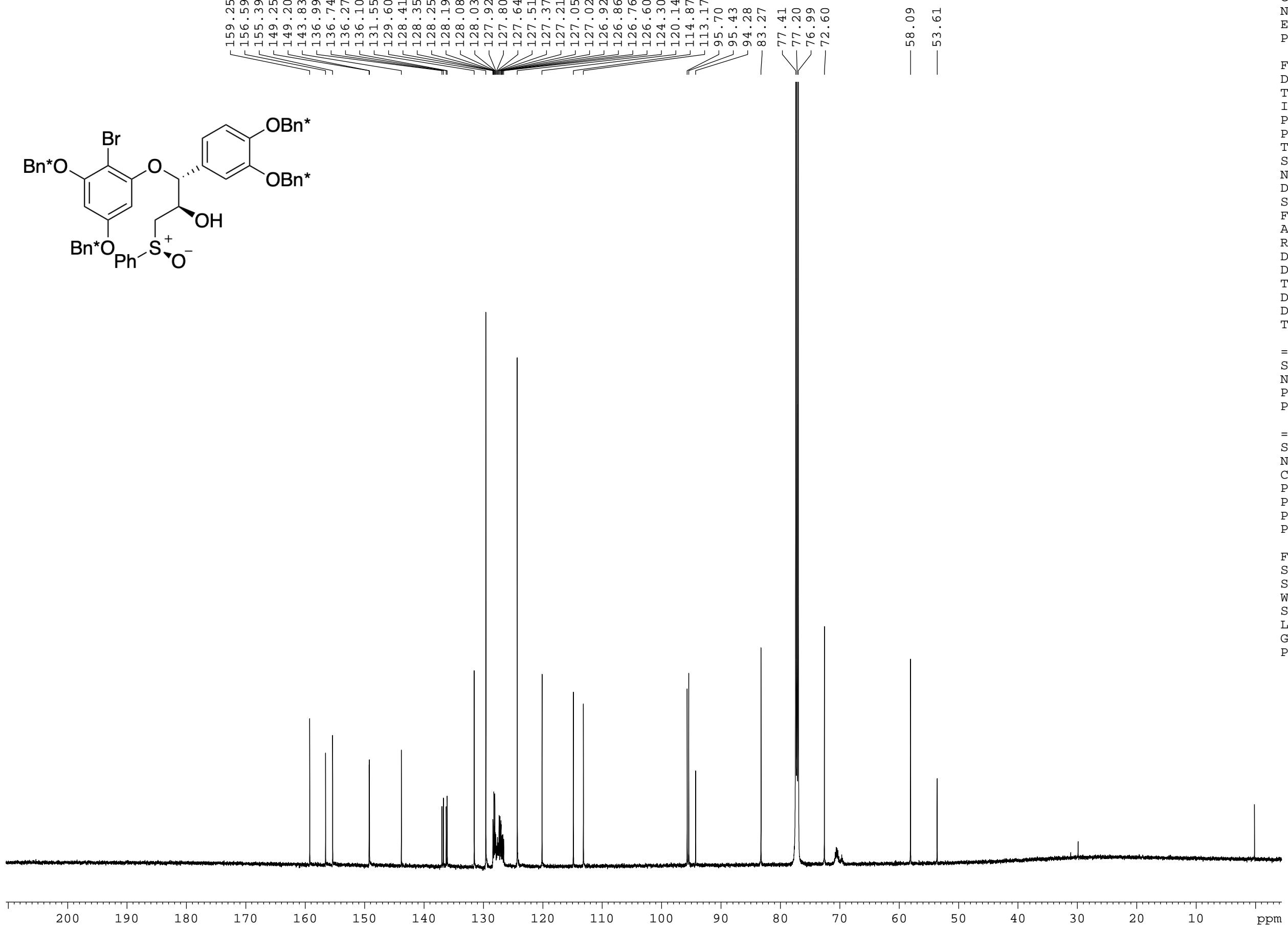
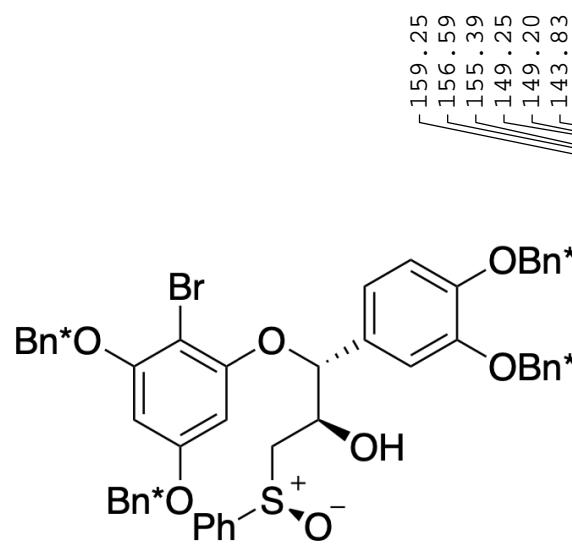
===== CHANNEL f1 ======
SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.00000000 W

F2 - Processing parameters
SI 65536
SF 600.1300147 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C NMR of 9b (150 MHz, CDCl₃)



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Current Data Parameters
NAME VB-847-2
EXPNO 51
PROCNO 1

```

F2 - Acquisition Parameters
Date_          20210907
Time           6.21
INSTRUM        spect
PROBHD         5 mm CPPBBO BB
PULPROG        zgpg30
TD             65536
SOLVENT        CDC13
NS             5000
DS              4
SWH            36057.691 Hz
FIDRES        0.550197 Hz
AQ             0.9087659 sec
RG              175.56
DW              13.867 usec
DE              18.00 usec
TE              298.1 K
D1             2.000000000 sec
D11            0.030000000 sec
TD0                 1

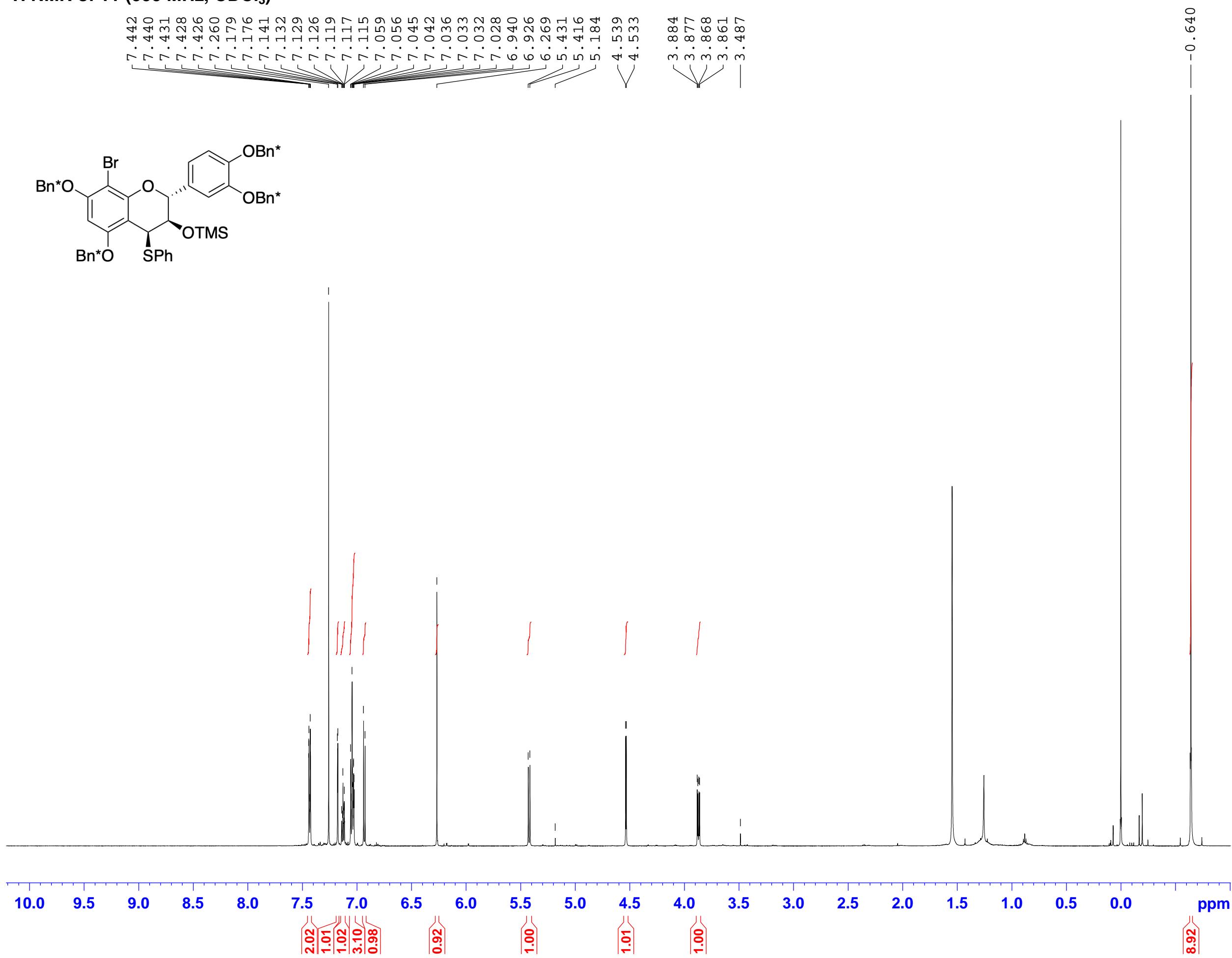
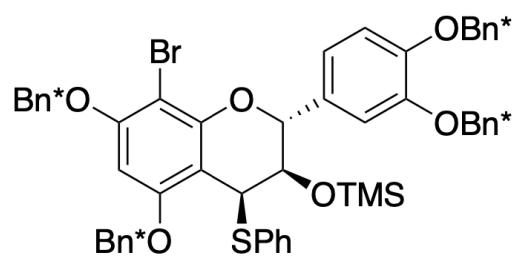
```

```
===== CHANNEL f1 =====
SFO1          150.9178981 MHz
NUC1          13C
P1            10.00 usec
PI.W1         80 000000000 W
```

```
===== CHANNEL f2 ======  
SFO2          600.1324005 MHz  
NUC2          1H  
CPDPRG[2      waltz16  
PCPD2         70.00 usec  
PLW2          13.43999958 W  
PLW12         0.61714000 W  
PLW13         0.31042001 W
```

F2 - Processing parameters
SI 32768
SF 150.9027843 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

¹H NMR of 11 (600 MHz, CDCl₃)





Current	Data	Parameters
NAME	VB-867-2A	
EXPNO	20	
PROCNO	1	

```

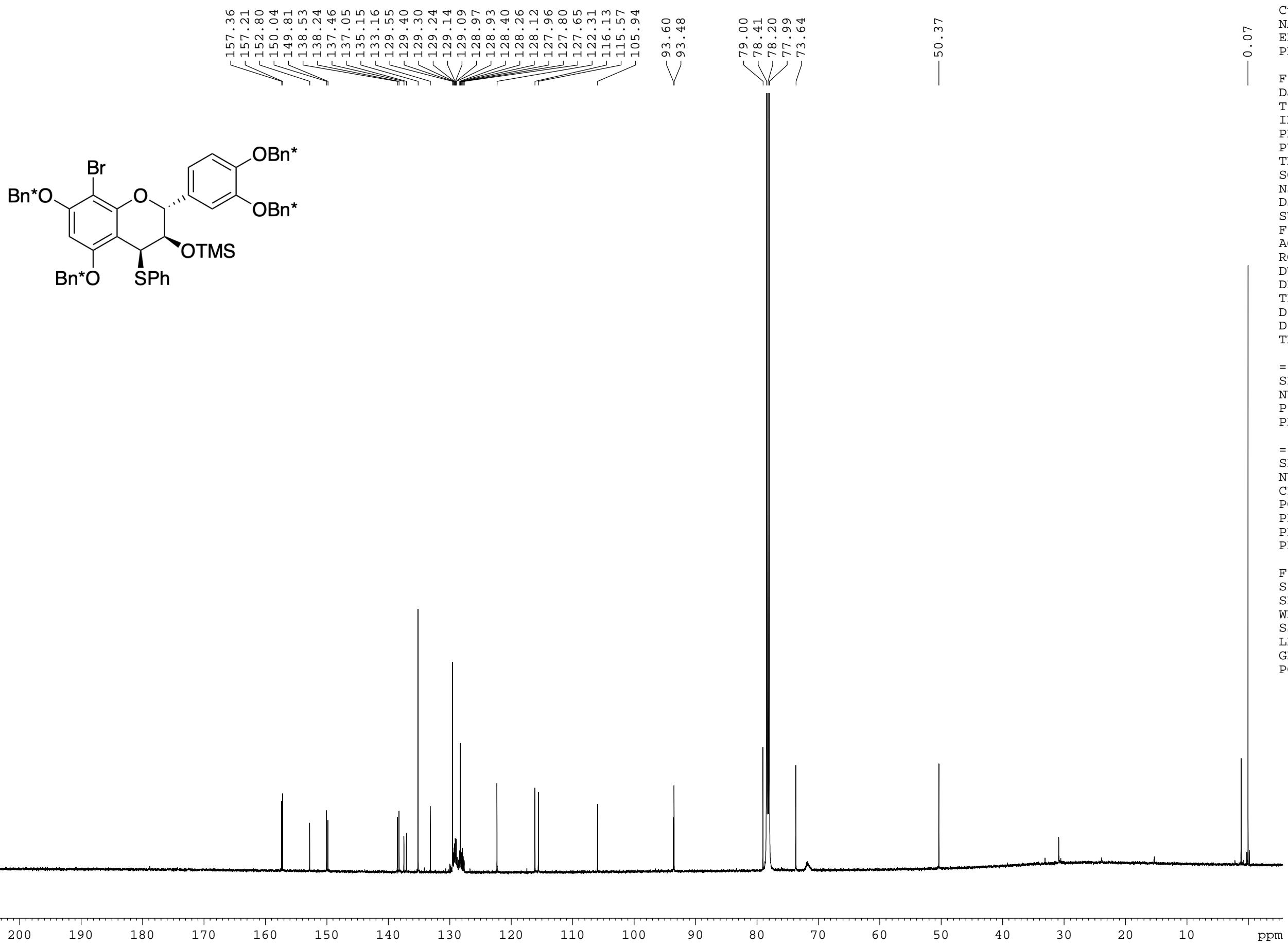
F2 - Acquisition Parameters
Date_          20210903
Time           18.49
INSTRUM        spect
PROBHD         5 mm CPPBBO BB
PULPROG        zg30
TD             65536
SOLVENT        CDC13
NS              16
DS               2
SWH             12019.230 Hz
FIDRES        0.183399 Hz
AQ             2.7262976 sec
RG              31.94
DW             41.600 usec
DE              10.00 usec
TE              298.1 K
D1             1.00000000 sec
TD0                 1

```

===== CHANNEL f1 ======
SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.000000000 W

```
F2 - Processing parameters
SI           65536
SF          600.1300144 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB          0
PC          1.00
```

¹³C NMR of 11 (150 MHz, CDCl₃)



Current Data Parameters
NAME VB-867-2A
EXPNO 33
PROCNO 1

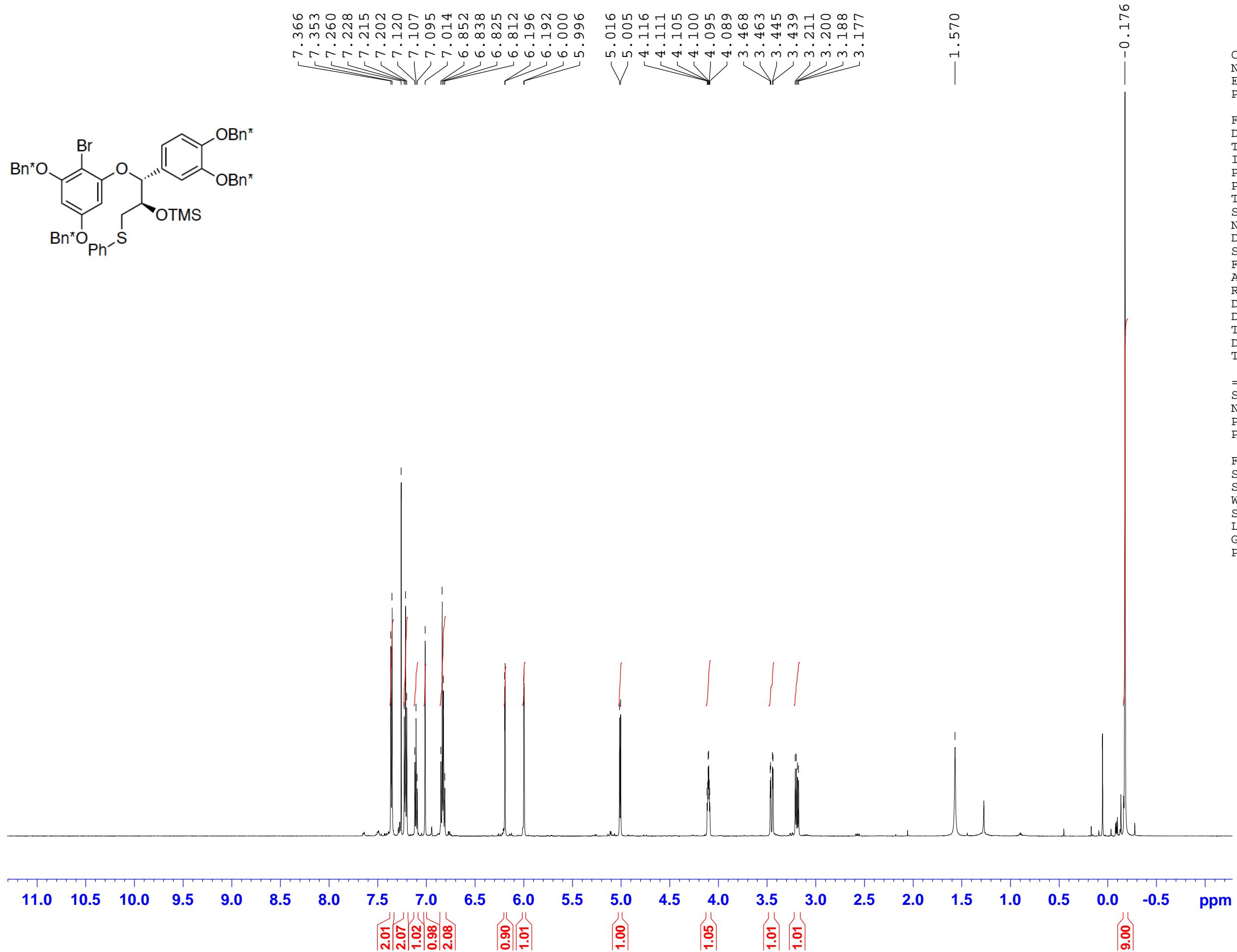
F2 - Acquisition Parameters
Date_ 20210904
Time 8.34
INSTRUM spect
PROBHD 5 mm CPPBBO BB
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 12240
DS 4
SWH 36057.691 Hz
FIDRES 0.550197 Hz
AQ 0.9087659 sec
RG 175.56
DW 13.867 usec
DE 18.00 usec
TE 298.2 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 ======
SFO1 150.9178981 MHz
NUC1 ¹³C
P1 10.00 usec
PLW1 80.0000000 W

===== CHANNEL f2 ======
SFO2 600.1324005 MHz
NUC2 ¹H
CPDPRG[2] waltz16
PCPD2 70.00 usec
PLW2 13.43999958 W
PLW12 0.61714000 W
PLW13 0.31042001 W

F2 - Processing parameters
SI 32768
SF 150.9026314 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

¹H NMR of 12 (600 MHz, CDCl₃)



Current Data Parameters
NAME VB-867-1
EXPNO 40
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210913
Time 21.55
INSTRUM spect
PROBHD 5 mm CPPBBO BB
PULPROG zg30
TD 65536
SOLVENT CDCl₃
NS 16
DS 2
SWH 12019.230 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 17.5
DW 41.600 usec
DE 10.00 usec
TE 298.2 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 ======

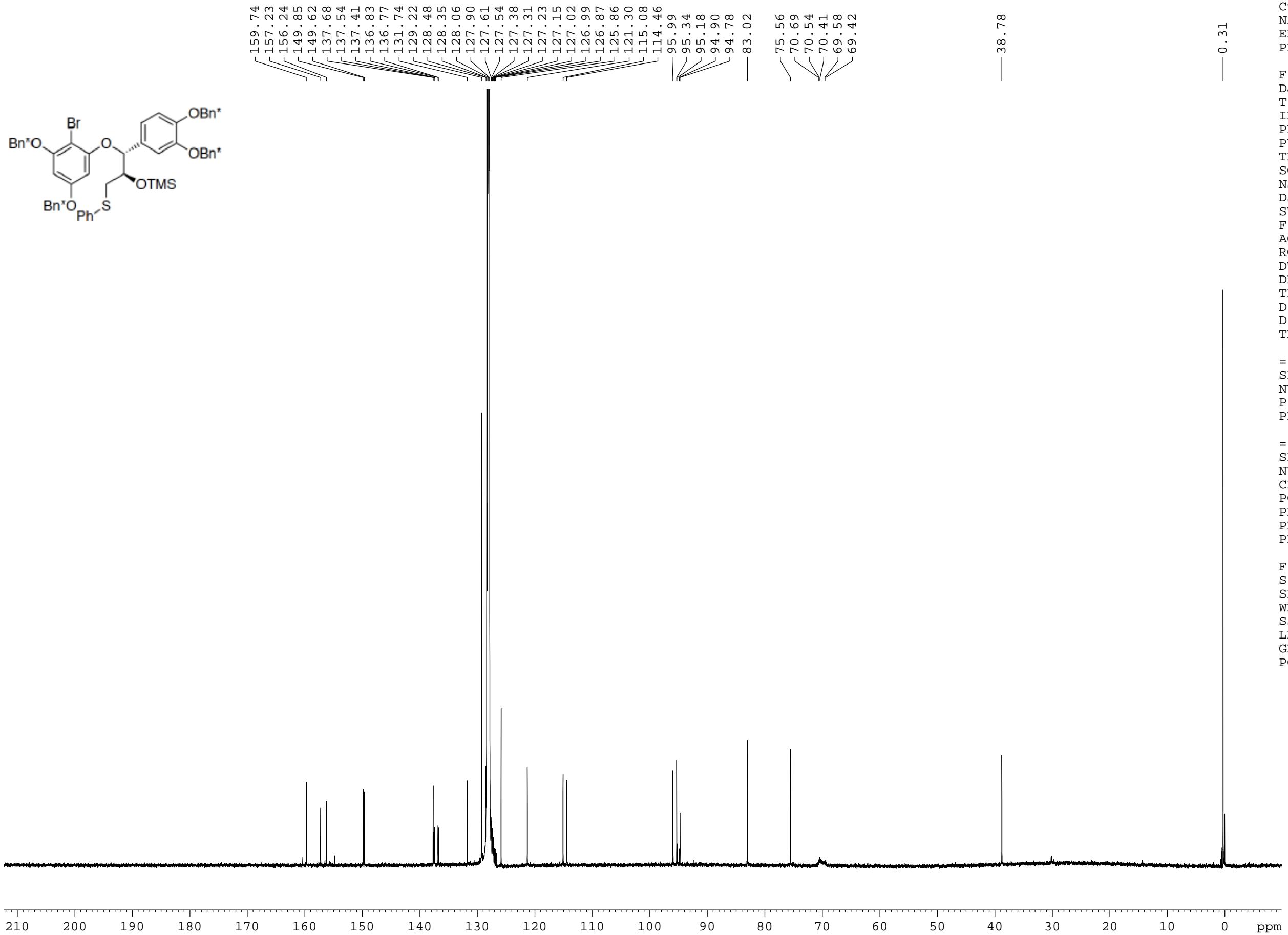
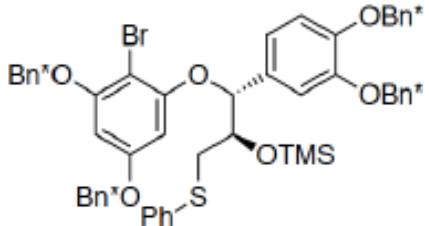
SFO1 600.1337060 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.00000000 W

F2 - Processing parameters
SI 65536
SF 600.1300145 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

¹³C NMR of 12 (150 MHz, C₆D₆)



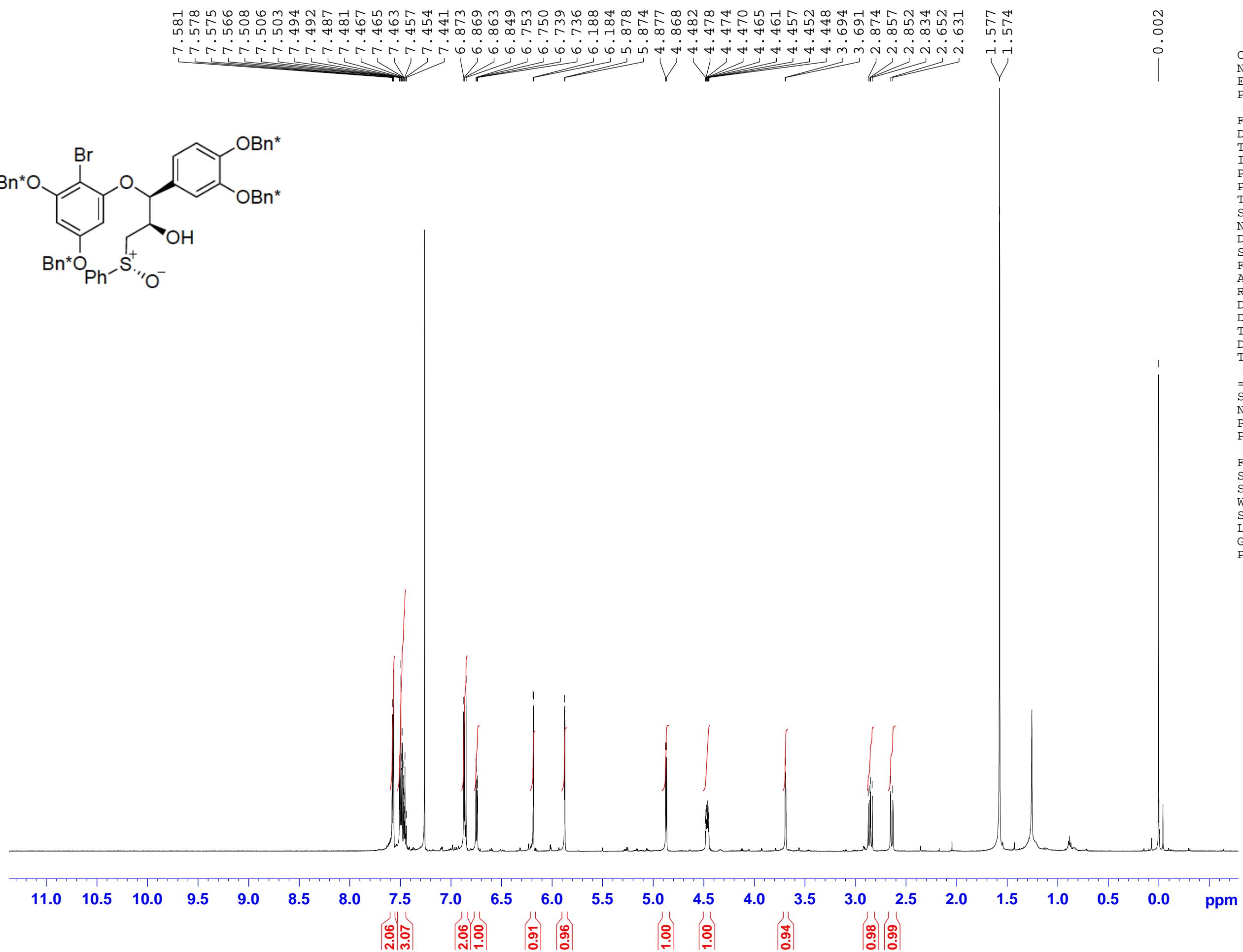
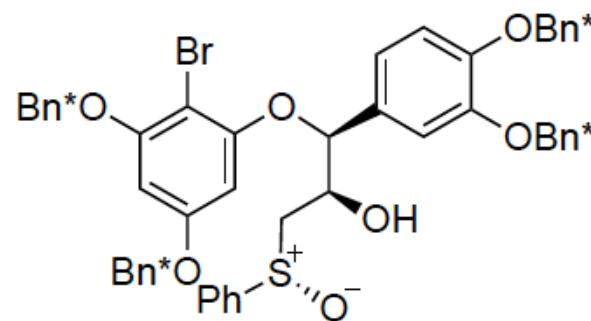
BRAKUER



Current Data Parameters

NAME	VB-867-1
EXPNO	71
PROCNO	1
F2 - Acquisition Parameters	
Date_	20220716
Time	2.12
INSTRUM	spect
PROBHD	5 mm CPPBBO BB
PULPROG	zgpg30
TD	65536
SOLVENT	C6D6
NS	5000
DS	4
SWH	36057.691 Hz
FIDRES	0.550197 Hz
AQ	0.9087659 sec
RG	175.56
DW	13.867 usec
DE	18.00 usec
TE	298.1 K
D1	2.00000000 sec
D11	0.03000000 sec
TD0	1
===== CHANNEL f1 =====	
SFO1	150.9178981 MHz
NUC1	13C
P1	10.00 usec
PLW1	80.00000000 W
===== CHANNEL f2 =====	
SFO2	600.1324005 MHz
NUC2	1H
CPDPRG[2]	waltz16
PCPD2	70.00 usec
PLW2	13.43999958 W
PLW12	0.61714000 W
PLW13	0.31042001 W
F2 - Processing parameters	
SI	32768
SF	150.9027533 MHz
WDW	EM
SSB	0
LB	1.00 Hz
GB	0
PC	1.40

¹H NMR of 2-*epi*-9a (600 MHz, CDCl₃)



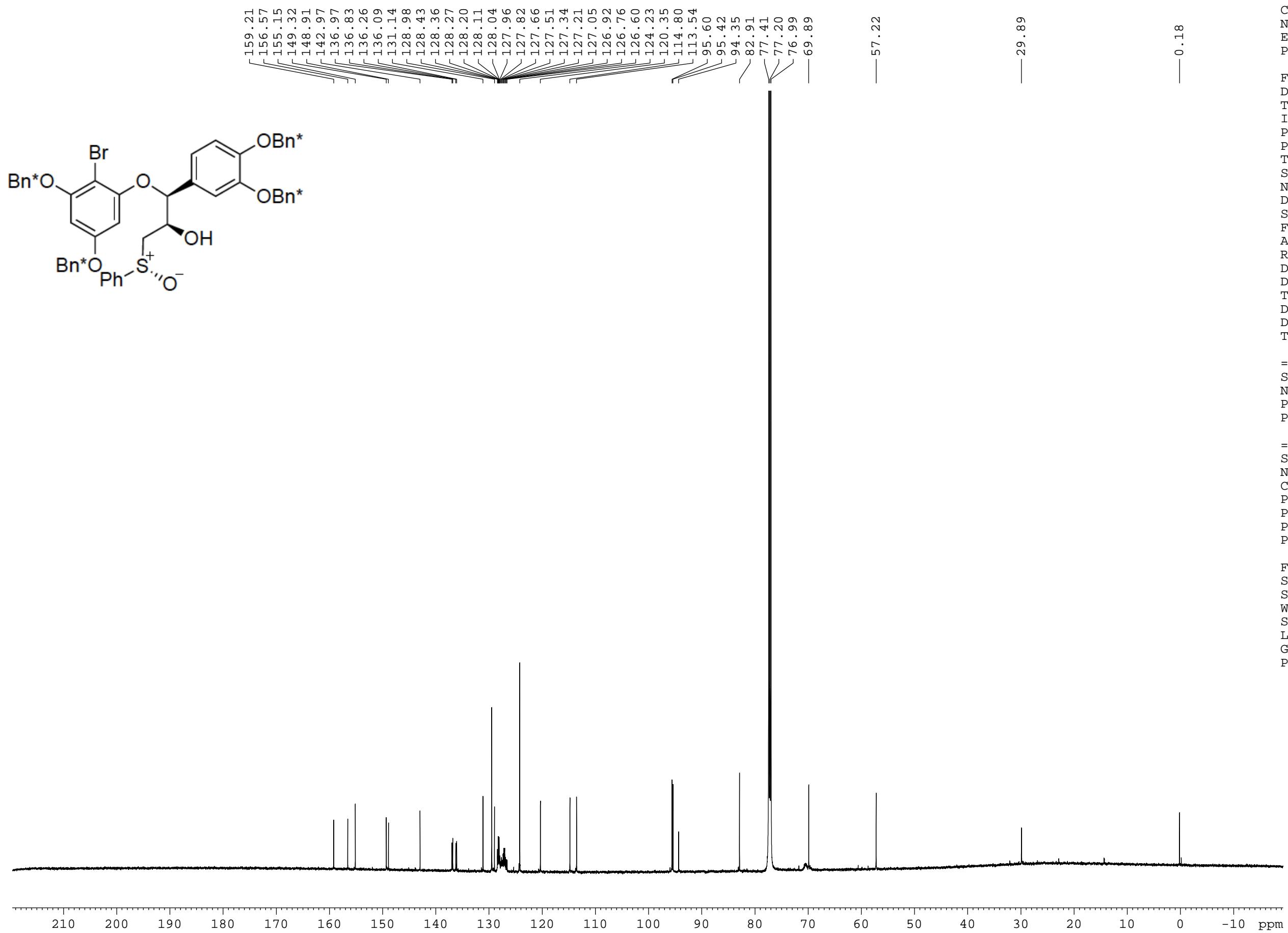

BRUKER
 Current Data Parameters
 TNAME VB-888-1A
 EXPNO 10
 PROCNO 1

 2 - Acquisition Parameters
 Date_ 20211006
 Time 15.38
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 JS 16
 DS 2
 SWH 12019.230 Hz
 TIDRES 0.183399 Hz
 Q 2.7262976 sec
 G 31.94
 W 41.600 usec
 E 10.00 usec
 TE 298.2 K
 D1 1.00000000 sec
 D0 1

 ===== CHANNEL f1 =====
 F01 600.1337060 MHz
 JUC1 1H
 I1 12.00 usec
 LW1 21.00000000 W

```
T2 - Processing parameters
      I          65536
      SF         600.1300148 MHz
      JDW        EM
      SSB        0
      SB         0.30 Hz
      SB        0
      PC         1.00
```

¹³C NMR of 2-*epi*-9a (150 MHz, CDCl₃)



Current Data Parameters
 NAME VB-888-1A
 EXPNO 12
 PROCNO 1

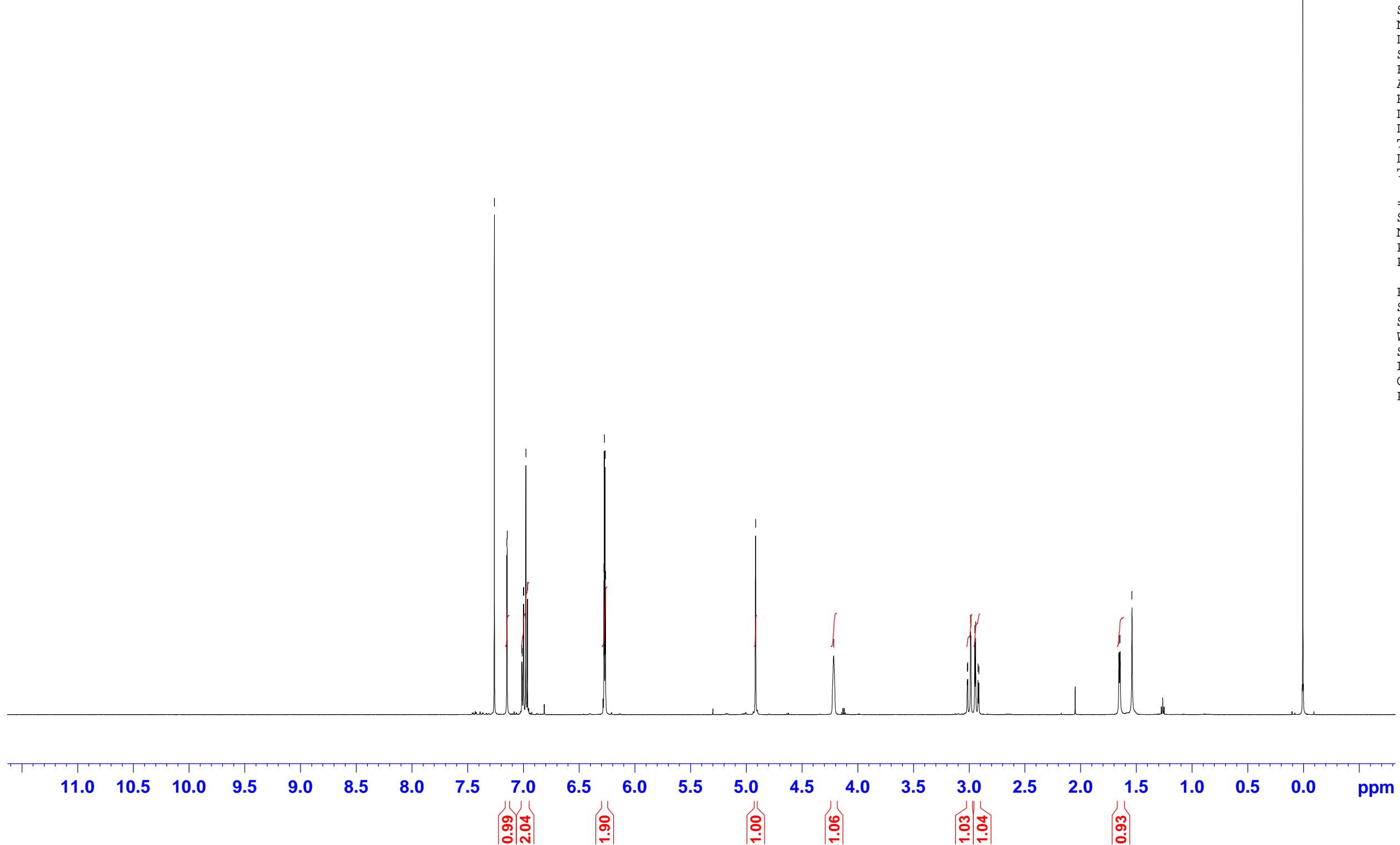
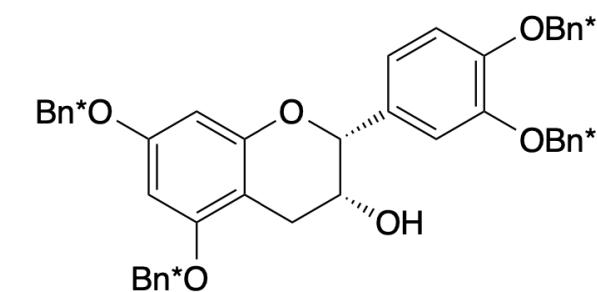
F2 - Acquisition Parameters
 Date_ 20211007
 Time 5.03
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl₃
 NS 8500
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 175.56
 DW 13.867 usec
 DE 18.00 usec
 TE 298.2 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 ======
 SFO1 150.9178981 MHz
 NUC1 ¹³C
 P1 10.00 usec
 PLW1 80.0000000 W

===== CHANNEL f2 ======
 SFO2 600.1324005 MHz
 NUC2 ¹H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 13.43999958 W
 PLW12 0.61714000 W
 PLW13 0.31042001 W

F2 - Processing parameters
 SI 32768
 SF 150.9027830 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

¹H NMR of 15 (600 MHz, CDCl₃)



Current Data Parameters
 NAME VB-Bn4EC
 EXPNO 20
 PROCNO 1

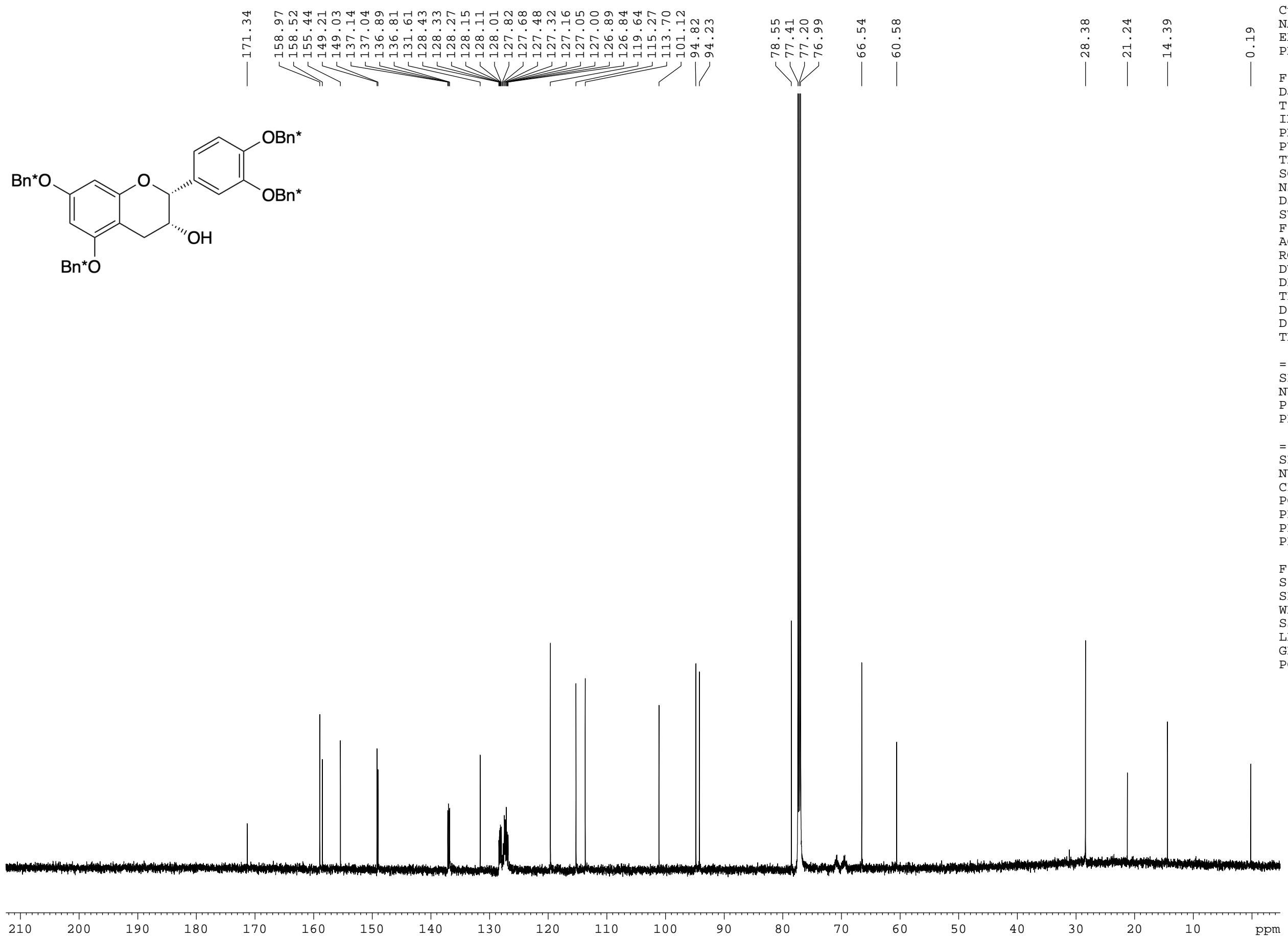
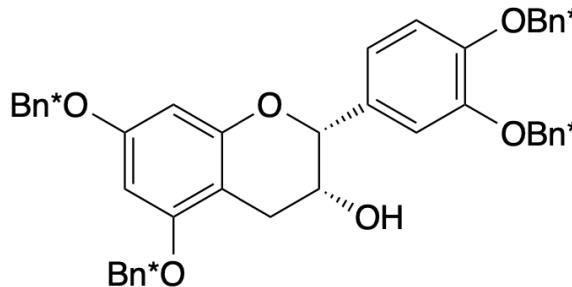
F2 - Acquisition Parameters
 Date_ 20211221
 Time 18.12
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 31.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 ====== SFO1 600.1337060 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

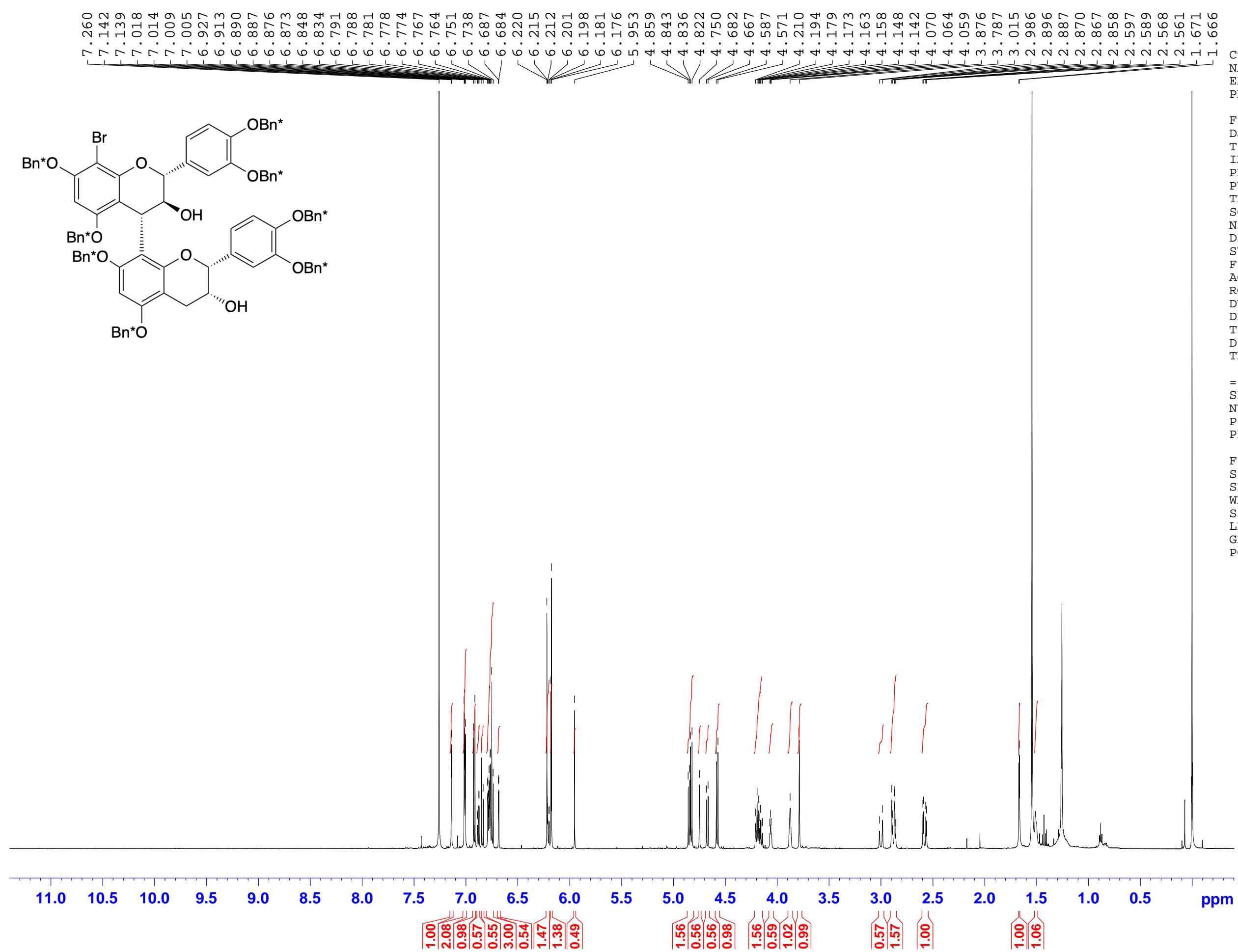
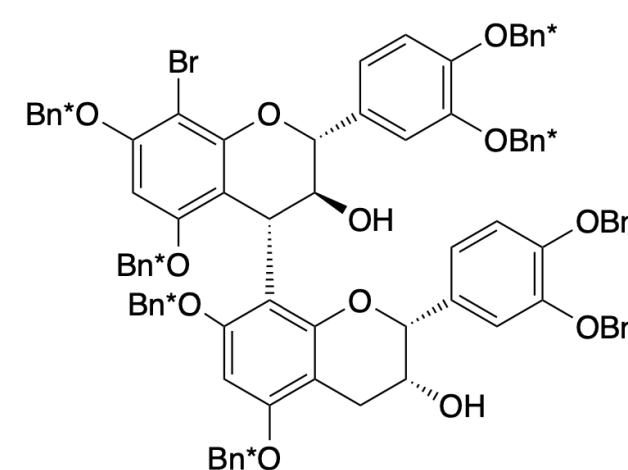
F2 - Processing parameters
 SI 65536
 SF 600.1300144 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C NMR of 15 (150 MHz, CDCl₃)





¹H NMR of 16 (600 MHz, CDCl₃)





Current	Data	Parameters
NAME	VB-861-2A1	
EXPNO	23	
PROCNO		1

```

F2 - Acquisition Parameters
Date_           20211221
Time            12.27
INSTRUM         spect
PROBHD          5 mm CPPBBO BB
PULPROG        zg30
TD              65536
SOLVENT         CDC13
NS              100
DS              2
SWH             12019.230 Hz
FIDRES         0.183399 Hz
AQ              2.7262976 sec
RG              31.94
DW              41.600 usec
DE              10.00 usec
TE              298.1 K
D1              1.00000000 sec
TD0             1

```

```
===== CHANNEL f1 ======  
SFO1      600.1337060 MHz  
NUC1      1H  
P1        12.00 usec  
PLW1      21.000000000 W
```

```

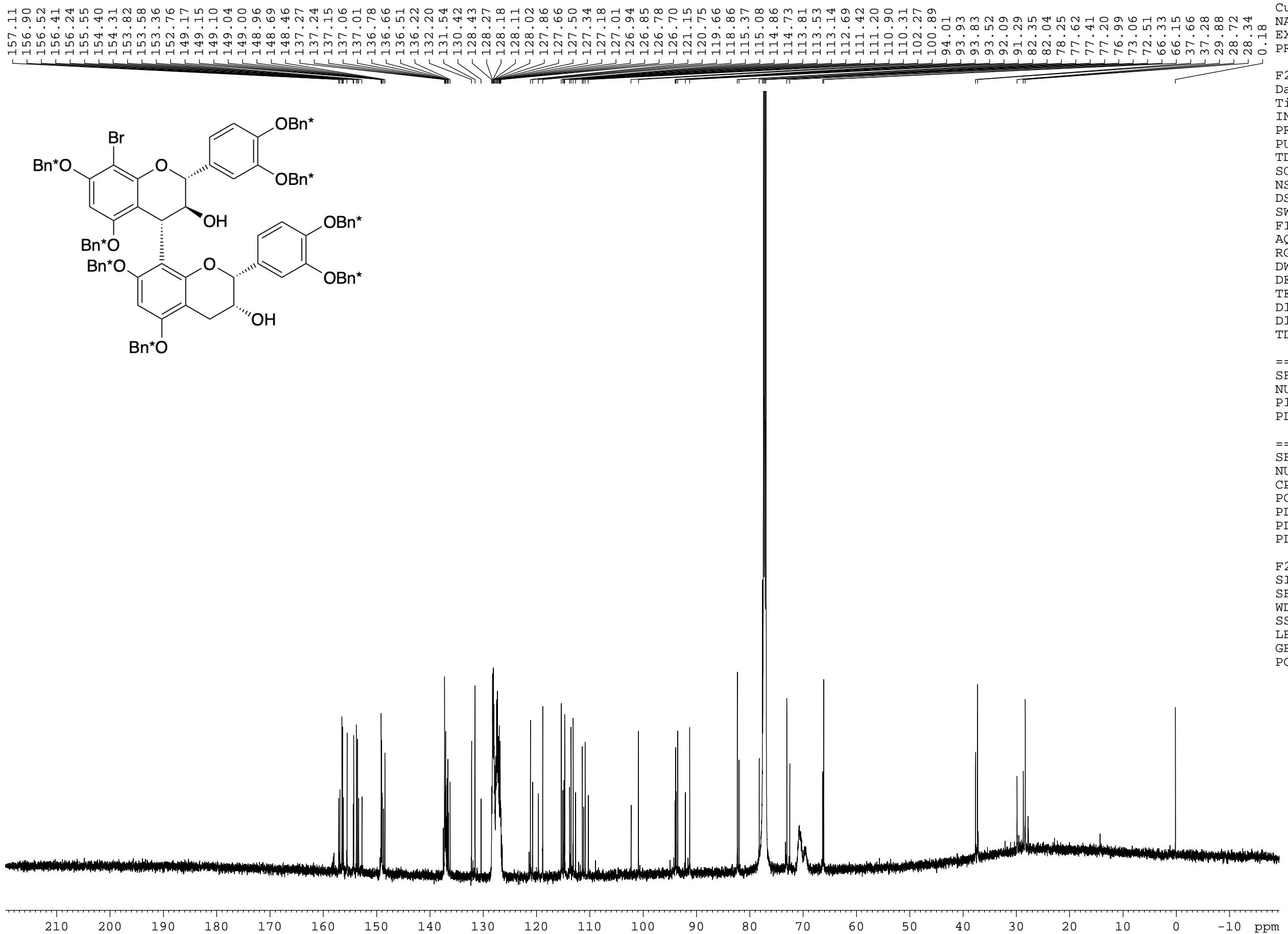
F2 - Processing parameters
SI           65536
SF          600.1300145 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB          0
PC          1.00

```

¹³C NMR of 16 (150 MHz, CDCl₃)



BRUKER



```

Current Data Parameters
NAME VB-CA-EC-2
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters
Date_ 20210908
Time 4.49
INSTRUM spect
PROBHD 5 mm CPPBBO BB
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 7000
DS 4
SWH 36057.691 Hz
FIDRES 0.550197 Hz
AQ 0.9087659 sec
RG 175.56
DW 13.867 usec
DE 18.00 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 150.9178981 MHz
NUC1 13C
P1 10.00 usec
PLW1 80.00000000 W

===== CHANNEL f2 =====
SFO2 600.1324005 MHz
NUC2 1H
CPDPKG[2] waltz16
PCPD2 70.00 usec
PLW2 13.43999958 W
PLW12 0.61714000 W
PLW13 0.31042001 W

F2 - Processing parameters
SI 32768
SF 150.9027847 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

```

¹H NMR of 4-*epi*-16 (600 MHz, CDCl₃)

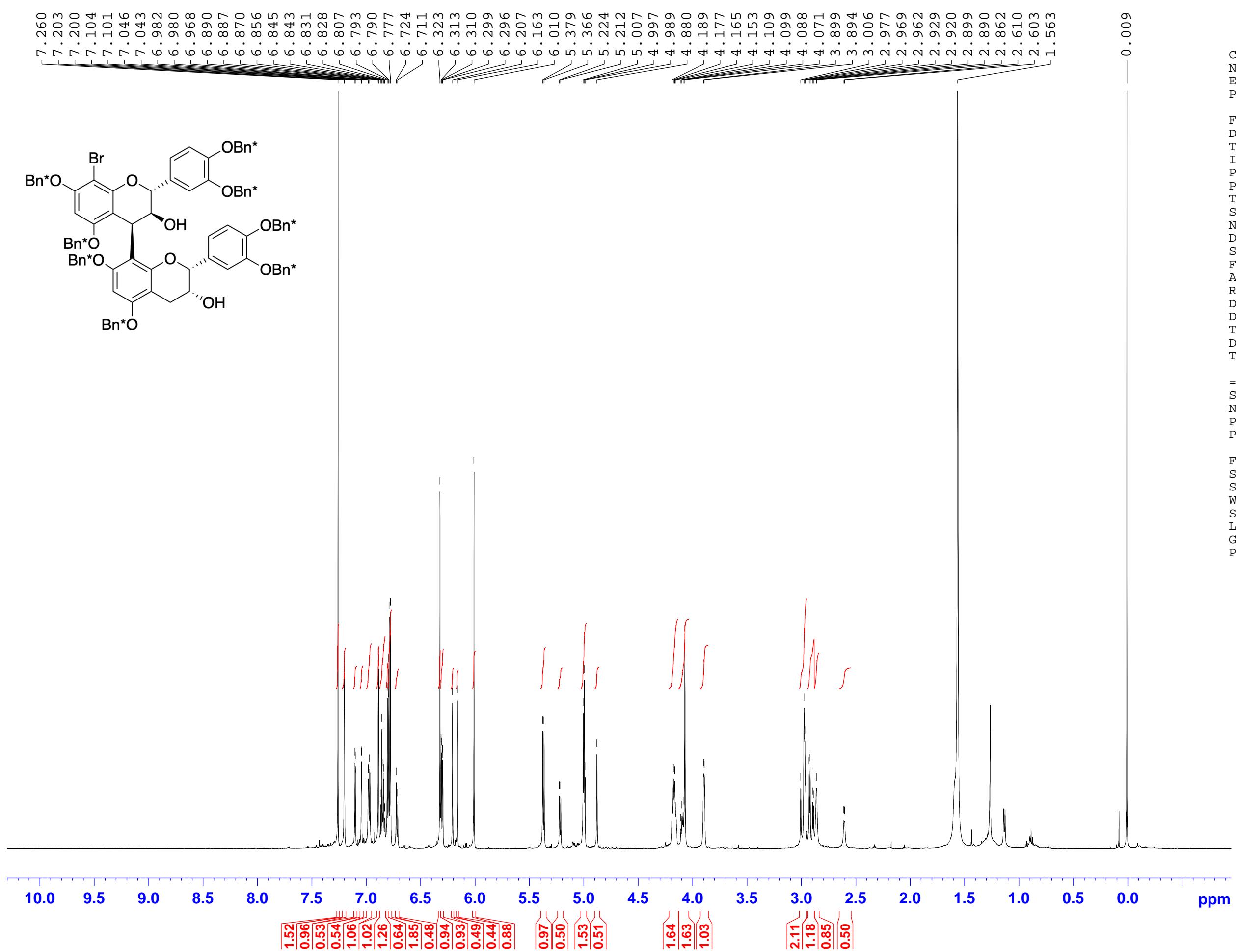


Current Data Parameters
 NAME VB-861-1
 EXPNO 22
 PROCNO 1

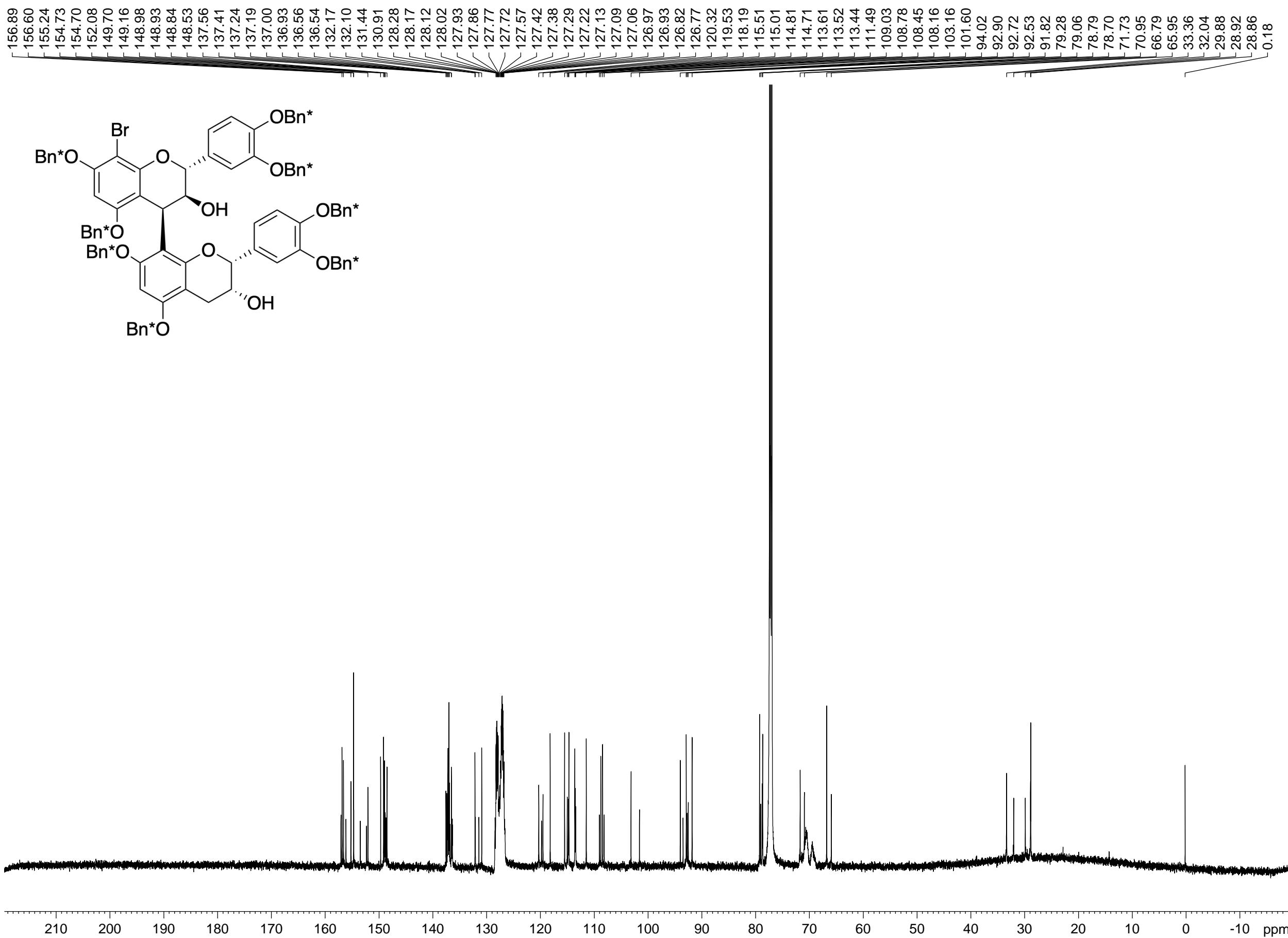
F2 - Acquisition Parameters
 Date_ 20210728
 Time 15.28
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 100
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 31.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 ====== SFO1 600.1337060 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

F2 - Processing parameters
 SI 65536
 SF 600.1300150 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



¹³C NMR of 4-*epi*-16 (150 MHz, CDCl₃)



Current Data Parameters
NAME VB-861-1
EXPNO 23
PROCNO 1

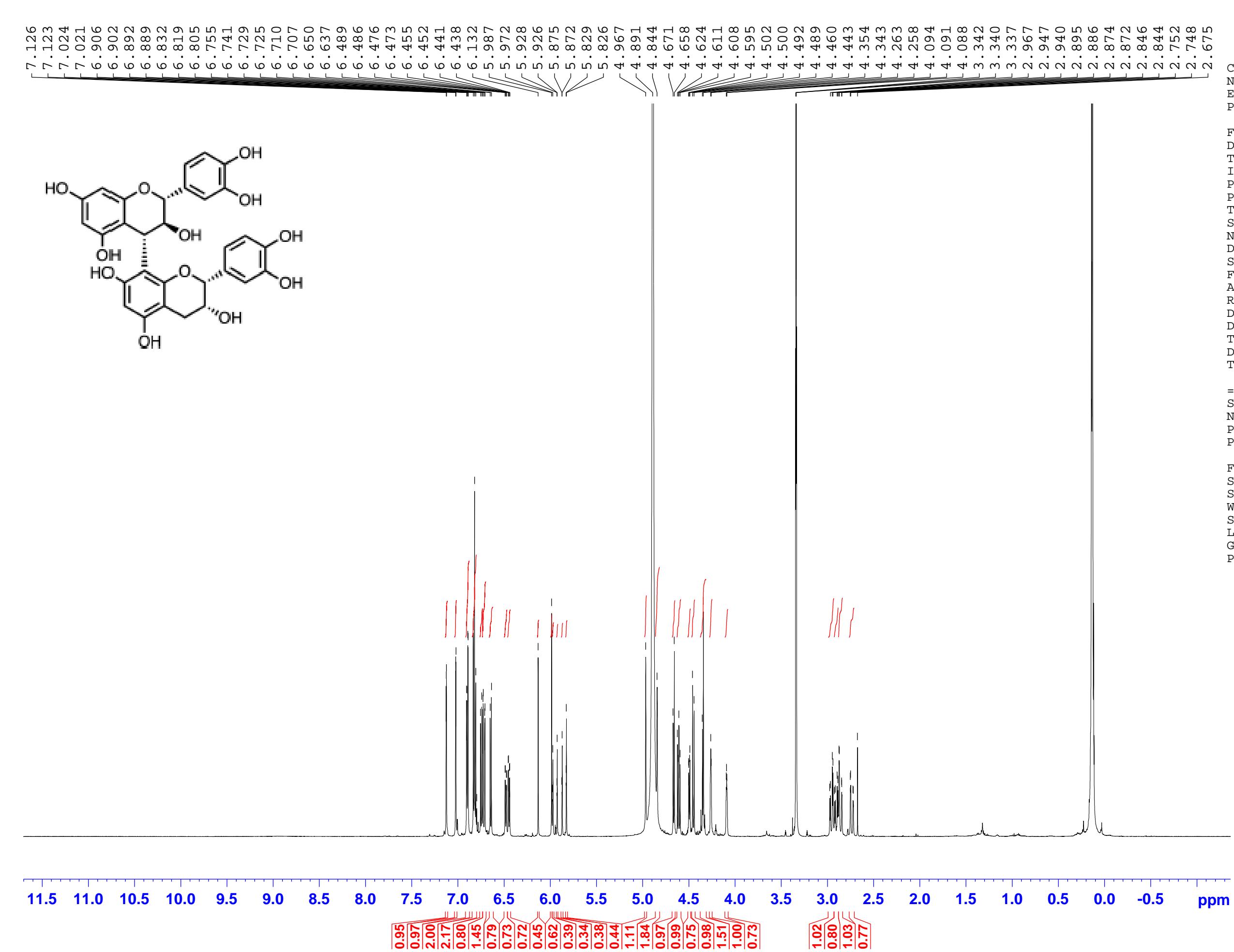
F2 - Acquisition Parameters
Date_ 20210729
Time 7.06
INSTRUM spect
PROBHD 5 mm CPPBBO BB
PULPROG zgpg30
TD 65536
SOLVENT CDCl₃
NS 8500
DS 4
SWH 36057.691 Hz
FIDRES 0.550197 Hz
AQ 0.9087659 sec
RG 175.56
DW 13.867 usec
DE 18.00 usec
TE 298.2 K
D1 2.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 ======
SFO1 150.9178981 MHz
NUC1 ¹³C
P1 10.00 usec
PLW1 80.00000000 W

===== CHANNEL f2 ======
SFO2 600.1324005 MHz
NUC2 ¹H
CPDPRG[2] waltz16
PCPD2 70.00 usec
PLW2 13.43999958 W
PLW12 0.61714000 W
PLW13 0.31042001 W

F2 - Processing parameters
SI 32768
SF 150.9027849 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

¹H NMR of 2 (600 MHz, CDCl₃)



BRUKER

Current Data Parameters
 NAME VB-865
 EXPNO 11
 PROCNO 1

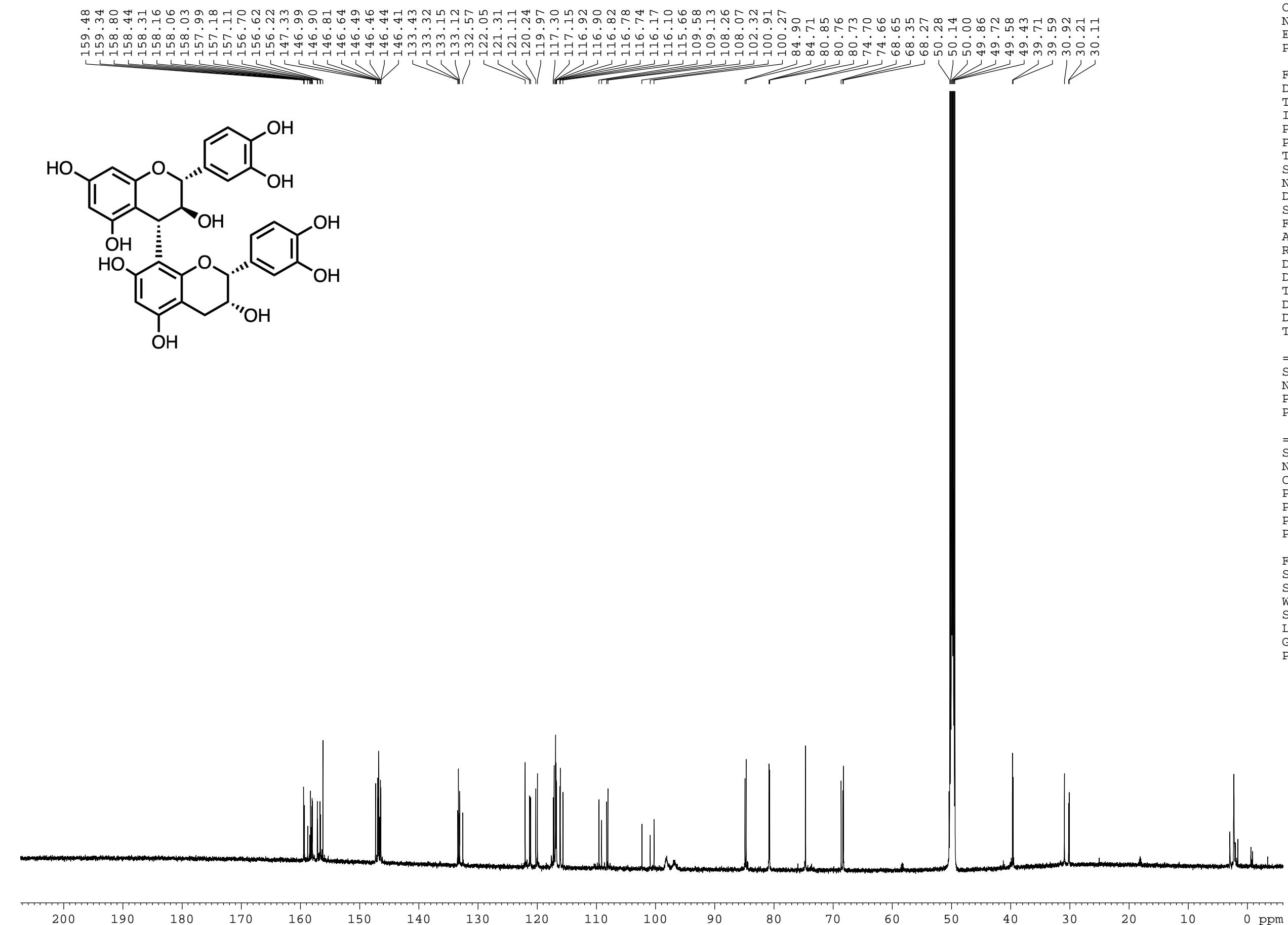
F2 - Acquisition Parameters
 Date_ 20210803
 Time 11.55
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT MeOD
 NS 100
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 31.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 ======

SFO1 600.1337060 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

F2 - Processing parameters
 SI 65536
 SF 600.1299933 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

¹³C NMR of 2 (150 MHz, CD₃OD)



Current Data Parameters
 NAME VB-865
 EXPNO 13
 PROCNO 1

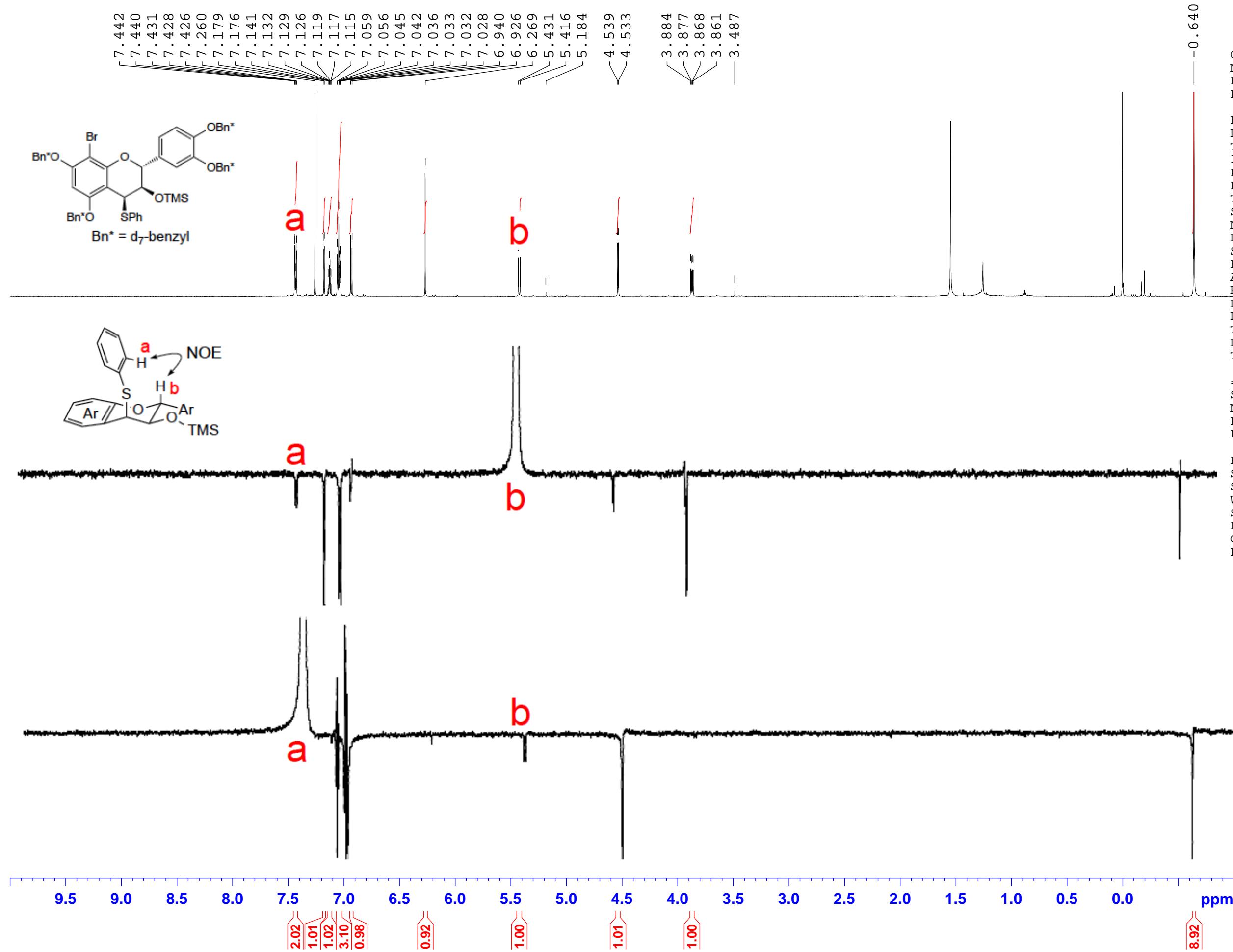
F2 - Acquisition Parameters
 Date_ 20210804
 Time 4.15
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zgpg30
 TD 65536
 SOLVENT MeOD
 NS 7500
 DS 4
 SWH 36057.691 Hz
 FIDRES 0.550197 Hz
 AQ 0.9087659 sec
 RG 175.56
 DW 13.867 usec
 DE 18.00 usec
 TE 298.1 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 ======
 SFO1 150.9178981 MHz
 NUC1 ¹³C
 P1 10.00 usec
 PLW1 80.0000000 W

===== CHANNEL f2 ======
 SFO2 600.1324005 MHz
 NUC2 ¹H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 13.43999958 W
 PLW12 0.61714000 W
 PLW13 0.31042001 W

F2 - Processing parameters
 SI 32768
 SF 150.9024684 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

NOE of 11



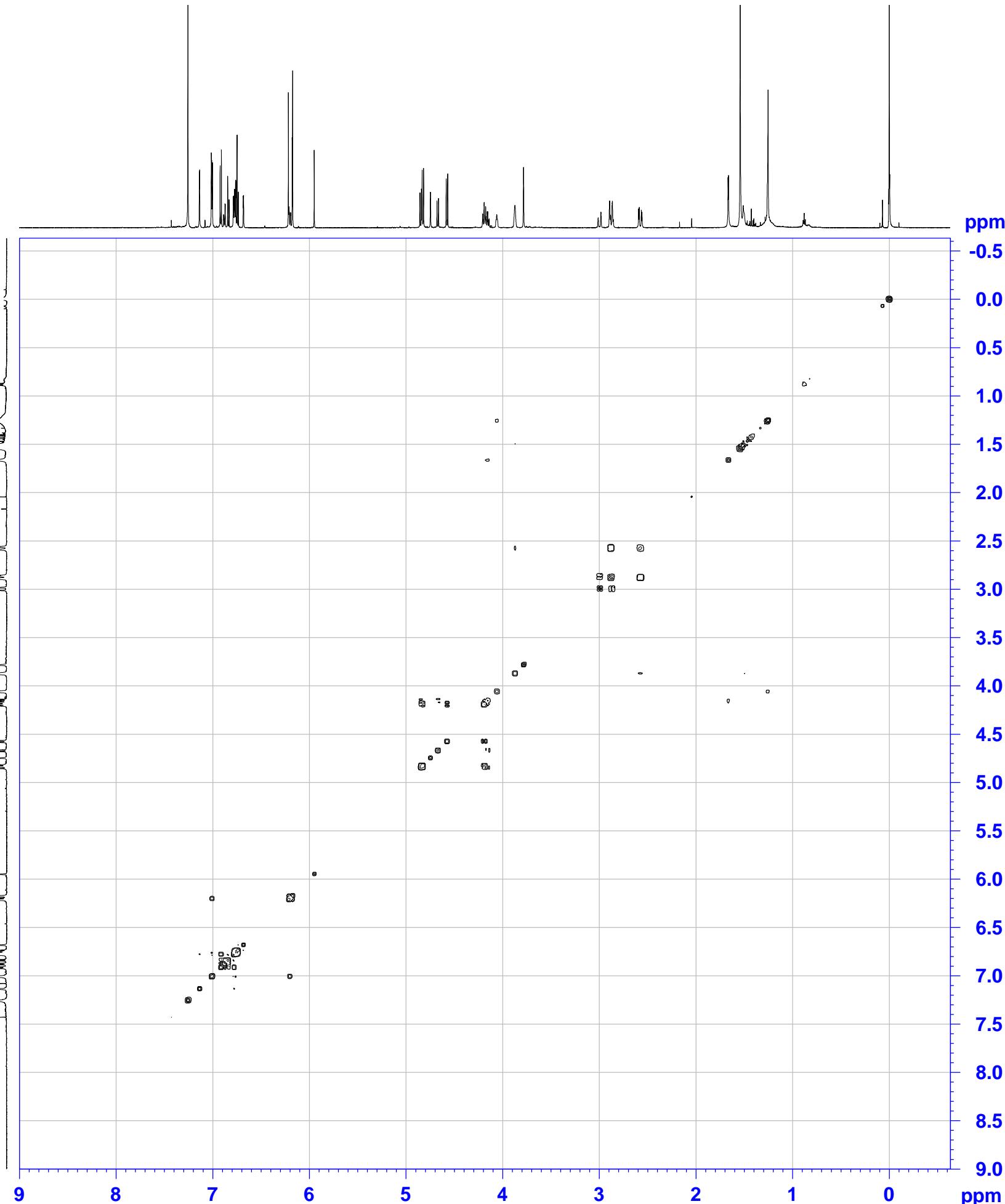
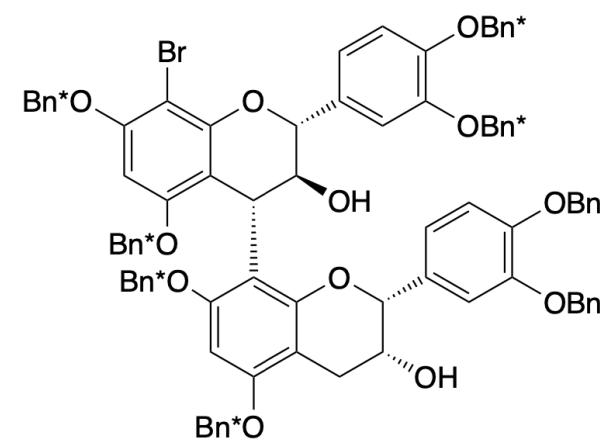
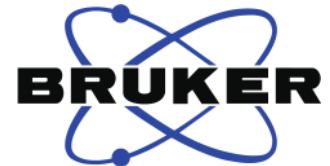
Current Data Parameters
 NAME VB-867-2A
 EXPNO 20
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20210903
 Time 18.49
 INSTRUM spect
 PROBHD 5 mm CPPBBO BB
 PULPROG zg30
 TD 65536
 SOLVENT CDCl₃
 NS 16
 DS 2
 SWH 12019.230 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 31.94
 DW 41.600 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 ====== SFO1 600.1337060 MHz
 NUC1 1H
 P1 12.00 usec
 PLW1 21.00000000 W

F2 - Processing parameters
 SI 65536
 SF 600.1300144 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

COSY of 16 (600 MHz, CDCl₃)



Current Data Parameters
NAME VB-861-2A1
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters
Date_ 20211221
Time 12.02
INSTRUM spect
PROBHD 5 mm CPPBBO BB
PULPROG cosygpppqr
TD 2048
SOLVENT CDCl₃
NS 1
DS 8
SWH 5154.639 Hz
FIDRES 2.516914 Hz
AQ 0.1986560 sec
RG 18.96
DW 97.000 usec
DE 10.00 usec
TE 298.0 K
D0 0.00000300 sec
D1 1.92913902 sec
D11 0.03000000 sec
D12 0.00002000 sec
D13 0.00000400 sec
D16 0.00020000 sec
INO 0.00019400 sec

===== CHANNEL f1 ======
SFO1 600.1322140 MHz
NUC1 1H
P0 12.00 usec
P1 12.00 usec
P17 2500.00 usec
PLW1 21.00000000 W
PLW10 4.83839989 W

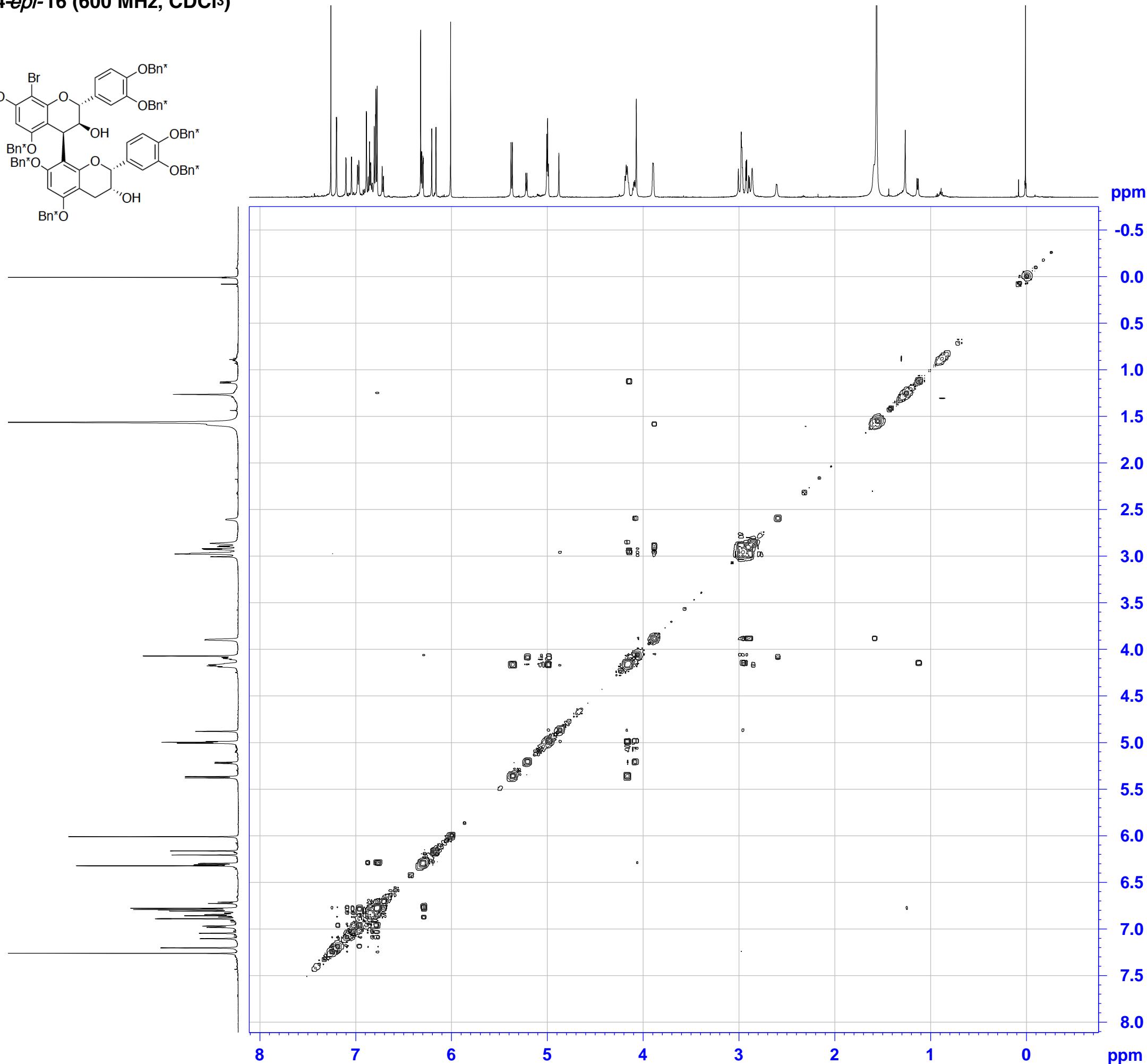
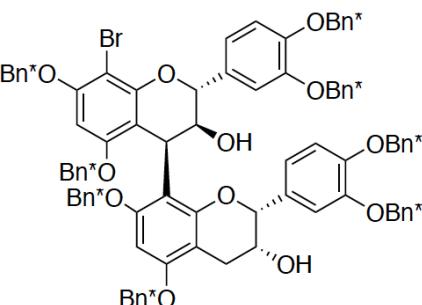
===== GRADIENT CHANNEL =====
GPNAME[1] SMSQ10.100
GPZ1 10.00 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 128
SFO1 600.1322 MHz
FIDRES 40.270618 Hz
SW 8.589 ppm
FnMODE QF

F2 - Processing parameters
SI 1024
SF 600.1300161 MHz
WDW QSINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 600.1300161 MHz
WDW QSINE
SSB 0
LB 0 Hz
GB 0

COSY of 4-*epi*-16 (600 MHz, CDCl³)





Current	Data	Parameters
NAME	VB-861-1	
EXPNO	21	
PROCNO	1	

```

F2 - Acquisition Parameters
Date_          20210728
Time           15.19
INSTRUM       spect
PROBHD        5 mm CPPBBO BB
PULPROG       cosygpppqf
TD             2048
SOLVENT        CDC13
NS              1
DS              8
SWH            5319.149 Hz
FIDRES        2.597241 Hz
AQ             0.1925120 sec
RG              18.96
DW             94.000 usec
DE              10.00 usec
TE              298.1 K
D0             0.00000300 sec
D1             1.93528295 sec
D11            0.03000000 sec
D12            0.00002000 sec
D13            0.00000400 sec
D16            0.00020000 sec
TIN0           0.00018800 sec

```

```
===== CHANNEL f1 =====
SFO1          600.1322288 MHz
NUC1           1H
P0            12.00 usec
P1            12.00 usec
P17           2500.00000 usec
PLW1           21.0000000 W
PLW10          4.83839989 W
```

===== GRADIENT CHANNEL =====
GPNAME[1] SMSQ10.100
GPZ1 10.00 %
P16 1000.00 usec

```

F1 - Acquisition parameters
TD           128
SFO1        600.1322 MHz
FIDRES      41.555851 Hz
SW          8.863 ppm
FpMODE      OF

```

F2 - Processing parameters
SI 1024
SF 600.1300208 MHz
WDW QSINE
SSB 0

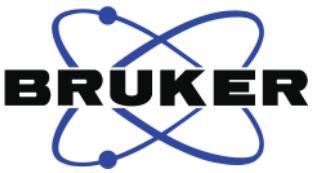
```

LB          0 Hz
GB          0
PC          1.40

F1 - Processing parameters
SI          1024
MC2         QF
SF          600.1300208 MHz
WDW         QSINE
SSB          0
LB          0 Hz

```

COSY of 2 (600 MHz, CD₃OD)



Current Data Parameters
NAME VB-865
EXPNO 12
PROCNO 1

F2 - Acquisition Parameters
Date 20210803
Time 11.56
INSTRUM spect
PROBHD 5 mm CPPBBO BB
PULPROG cosygpppqr
TD 2048
SOLVENT MeOD
NS 1
DS 8
SWH 5000.000 Hz
FIDRES 2.441406 Hz
AQ 0.2048000 sec
RG 17.5
DW 100.000 usec
DE 10.00 usec
TE 298.2 K
D0 0.00000300 sec
D1 1.92299497 sec
D11 0.03000000 sec
D12 0.00002000 sec
D13 0.00000400 sec
D16 0.00020000 sec
INO 0.00020000 sec

===== CHANNEL f1 ======

SFO1 600.1321047 MHz
NUC1 1H
P0 12.00 usec
P1 12.00 usec
P17 2500.00 usec
PLW1 21.0000000 W
PLW10 4.83839989 W

===== GRADIENT CHANNEL =====

GPNAM[1] SMSQ10.100
GPZ1 10.00 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 128
SFO1 600.1321 MHz
FIDRES 39.062500 Hz
SW 8.331 ppm
FnMODE QF

F2 - Processing parameters
SI 1024
SF 600.1300723 MHz
WDW QSINE
SSB 0
LB 0 Hz
GB 0
PC 1.40

F1 - Processing parameters
SI 1024
MC2 QF
SF 600.1300723 MHz
WDW QSINE
SSB 0
LB 0 Hz
GB 0

