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Supplemental Information: π -Facial Selectivity in the Diels-Alder Reaction of Glucosamine-based Chiral Furans and Maleimides

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Material and methods

S1. Reagents

All reagents were acquired from commercial sources and used without further purification. Specifically, D-glucosamine HCl was purchased from Carbosynth Ltd, Silica gel (ultra pure, 40-60 um, 60A), molecular sieves (4 Å), and platinum on activated carbon (5% Pt, not reduced) from Acros Organics, Celite 535 from Carl Roth, phenylboronic acid from AmBeed and *N*-phenylmaleimide from TCI. Thin-layer chromatography (TLC) was carried out on Merck silica gel 60 F254 aluminum plates. Visualization of compounds by TLC was done by potassium permanganate stain. Automated flash column chromatography cartridges were acquired from Screening Devices (UltraPure Irregular Silica Gel, 40 – 63 um, 60A, 5/1P). *N*-benzyloxycarbonyl glucosamine (**GlcNCbz 9**) was prepared according to literature.¹

(1) Carbohydr. Res., **1999**, 321, 176–189.

S2. Equipment

¹H-NMR and ¹³C-NMR analyses were performed on a Bruker Avance NEO (400 MHz) at 25 °C. High-resolution mass spectrometry (HRMS) was performed on a Thermo Scientific LTQ Orbitrap XL (FTMS). Infrared spectra (IR) spectra were recorded on a PerkinElmer Spectrum Two FT-IR Spectrometer. DSC was recorded on a TA instruments DSC Q20 and enantiomeric excess (*e.e.*) was determined by chiral HPLC on a waters Acquity UPC². Automated flash column chromatography was carried out on a Buchi Sepacore[®] Flash System X10 (Pumps: 601, control unit: 620, detector: 640, fraction collector: 660). Crystal data was collected at 200 K on a Bruker D8 Venture diffractometer equipped with multilayer optics for monochromatization and collimator, Mo K α radiation (λ = 0.71073 Å) and an Oxford Cryostream 800 unit.

S3. Chiral HPLC method for Di-HCF (10)

Chiral SFC Method	:	31875C UPC ²	10m Amy2	EIAN		
System	:	Waters Acquit	ty UPC ² syst	em with UV d	etector and QDA detector	
Column	:	Phenomenex	Lux Amylose	e-2 (3.0 x 150	mm; 3 μm)	
Mobile Phase A	:	CO ₂				
Mobile Phase B	:	Ethanol/Isopr	opanol/Acet	tonitrile/Amm	onium acetate (300 mL/300 mL/300 mL/1	39 g)
Pump Flow	:	1.0 mL/Min				
UV detection	:	210 nm				
Injection Volume	:	1.0 μL				
Total Run Time	:	10.0 Min				
Column Temperature	:	40 °C				
ABPR	:	138 bar				
Mass Detection	:	MS Scan ES po	ositive and n	egative		
Mass Range	:	100 – 600 Da				
Pump Program	:	Gradient				
		Time (Min)	%A	%B	Curve	

Time (Min)	%A	%B	Curve
Initial	98	2	Initial
6.0	60	40	6
9.0	60	40	6
9.1	98	2	6



S4. Crystallographic Data of DAPRED-Bz (16)

Crystals of DAPRED-Bz (**16**) were obtained from an ethanol solution (10 mg/mL) as tiny needles. A single crystal was coated with mineral oil, mounted on Mitegen MicroMounts with the aid of a microscope, and immediately placed in the low temperature nitrogen stream of the diffractometer. As crystals diffracted very weakly, data collection was only performed up to theta 25 degrees. Crystallographic data for **16** is presented in table 1.

The structure was solved, by using the Olex2² package by intrinsic phasing methods (SHELXT)³ and refined by least-squares against F^2 (SHELXL).⁴ All non-hydrogen atoms were anisotropically refined, while hydrogen atoms that were located in the different Fourier maps were isotropically refined, except those of the phenyl rings which were placed at idealized positions and refined using a riding model.

	DAPRED-Bz (16)
Formula	C ₃₁ H ₂₈ N ₂ O ₈
Μ	556.55
<i>T</i> [K]	200(2)
λ[Å]	0.71073
Crystal system	monoclinic
Space group	P2 ₁
<i>a</i> [Å]; α [°]	12.4038(19)
<i>b</i> [Å]; β [°]	6.0006(9); 105.886(4)
<i>c</i> [Å]; γ [°]	18.430(3)
<i>V</i> [Å ³]	1319.3(3)
Z	2
$ ho_{ m calcd} [m g cm^{-3}]$	1.401
μ [mm ⁻¹]	0.102
<i>F(000)</i>	584
Crystal size [mm ³]	0.15 x 0.098 x 0.04
θ range [deg]	3.40 to 25.02
Index ranges	14 to -14,
	7 to -7,
	21 to -21
Reflections collected	35365
Unique data	$4636(R_{int} = 0.145)$
Reflections $[I>2\sigma(I)]$	3286
Goodness-of-fit on F ²	1.035
Final R indices [I>2 σ (I)]	R1 = 0.045
	wR2 = 0.080
R indices (all data)	R1 = 0.086
Largest diff. peak/hole [e·Å-3]	WK2 = 0.096 0.164/-0.194

Table 1. Experimental data for the X-ray diffraction studies on DAPRED-Bz (16).

 ${}^{a}R1 = \Sigma ||F_{0}| - |F_{c}|| / [\Sigma |F_{0}|] wR2 = \{ [\Sigma w (F_{0}^{2} - F_{c}^{2})^{2}] / [\Sigma w (F_{0}^{2})^{2}] \}^{1/2}$

- (2) J. Appl. Cryst. 2009, 42, 339-341.
- (3) Acta Cryst. 2015, A71, 3-8
- (4) Acta Cryst. 2015, C71, 3-8

S5. Reversibility Study

In DMSO- d_6 (0.8 mL, 30 mg/mL) solutions were prepared of the *endo* and *exo* diastereomers, as mixtures with matching topologies, of the Diels Alder products of **Di-HCF** (**10**) with *N*-methylmaleimide and *N*-phenylmaleimide. The product distribution was determined at t = 0 by ¹H-NMR (table 2). The samples were kept at room temperature, 40 °C, 50 °C, 60 °C and 70 °C. After 24 h, the decrease in concentration of the corresponding isomers was determined by ¹H-NMR (table 3). The sum of the concentration of the two target isomers was normalized, and the isomer concentration decrease at each temperature was calculated (table 4, graph 1).

Dienophile	Di-HCF	Endo	Endo'	Exo	Exo'	Isomer sum
N-methylmaleimide	00/	400/	400/	0.1	20/	0.00/
<i>endo</i> -isomer	0%	49%	49%	0%	2%	98%
N-methylmaleimide		00/	00/	26%	740/	100%
<i>exo</i> -isomer	0%	0%	0%	20%	/4%	100%
N-phenylmaleimide		450/	500/	0.1	50/	050/
endo -isomer	0%	45%	50%	0%	5%	95%
N-phenylmaleimide		00/	00/	22%	770/	100%
<i>exo</i> -isomer	U%	0%	υ%	23%	11%	100%

Table 2. Isomer distribution at t = 0

Table 3. Isomer distribution at t = 24h

Diananhila	Temperatur	Di-	Endo	Endo'	Exo	Exo'	Isomer
Dieliopilie	е	HCF					sum
	RT	0%	49%	49%	0%	2%	98%
N-	40 °C	3%	47%	48%	0%	2%	95%
methylmaleimide	50 °C	5%	44%	46%	2%	3%	90%
endo-isomer	60 °C	7%	40%	42%	4%	7%	82%
	70 °C	8%	34%	37%	8%	13%	71%
	RT	0%	0%	0%	26%	74%	100%
N-	40 °C	0%	0%	0%	26%	74%	100%
methylmaleimide	50 °C	2%	0%	0%	26%	72%	98%
exo Endo-isomer	60 °C	4%	1%	2%	25%	68%	93%
	70 °C	5%	2%	3%	24%	65%	89%
	RT	0%	45%	50%	0%	5%	95%
N phonylmalaimida	40 °C	6%	39%	45%	0%	5%	84%
<i>N</i> -prienymalemiue	50 °C	9%	37%	42%	4%	8%	79%
endo-isomer	60 °C	12%	29%	35%	8%	16%	64%
	70 °C	14%	17%	21%	17%	31%	38%
N phonylmaloimida	RT	0%	0%	0%	23%	77%	100%
w-phenyimaleimide	40 °C	3%	0%	0%	23%	74%	97%
exo-isomer	50 °C	5%	2%	3%	22%	68%	90%

60 °C	8%	4%	6%	23%	60%	83%
70 °C	9%	8%	11%	24%	47%	71%

Time	N-	N-	N-	N-
&	methylmaleimid	methylmaleimid	phenylmaleimid	phenylmaleimid
Temperatur	е	е	е	е
е	endo -isomer	<i>exo</i> -isomer	endo -isomer	<i>exo</i> -isomer
t = 0	100%	100%	100%	100%
t = 24, RT	100%	100%	100%	100%
t = 24, 40 °C	97%	100%	88%	97%
t = 24, 50 °C	92%	98%	83%	90%
t = 24, 60 °C	84%	93%	67%	83%
t = 24, 70 °C	72%	89%	40%	71%

Table 4. Normalized data. Sum of target isomers set to 100% at t=0

Graph 1. Representation of the normalized data from table 4. **Left**: The DA product of **Di-HCF** (**10**) + N-methylmaleimide. **Right**: The DA product of **Di-HCF** (**10**) + N-phenylmaleimide





Experimental procedures

S6. Periodate mediated diol oxidation to facilitate NMR analysis

NMR sample preparation. The sample (10 mg substrate) was dissolved in a mixture of $DMSO-d_6 - D_2O$ (1:1. 0.8 mL). To the solution was added $NalO_4$ (10 mg, ca. 2 eq.) and the mixture was stirred at room temperature for 1 h. The white precipitate was removed by filtration over a disposable syringe filter (0.45 uM) and the filtrate was analyzed by ¹H-NMR.

S7. Synthesis of Di-HCF (10)

Benzyl (R)-(5-(1,2-dihydroxyethyl)furan-3-yl)carbamate (Di-HCF 10). A mixture was prepared of Nbenzyloxycarbonyl glucosamine (GlcNCbz 9) (95 g, 1 eq., 0.30 mol), phenylboronic acid (55 g, 1.5 eq., 0.45 mol), molsieves (500 g, 4 Å) and pyridine (1 L). Under stirring was added a solution of triflic acid (46 g, 27 mL, 1 eq., 0.30 mol) in pyridine (0.5 L). The mixture was stirred at reflux for 30 min and subsequently cooled to rt. Solid K₂CO₃ (84 g, 2 eq., 0.61 mol) and neopentyl glycol (63 g, 60 mL, 2 eq., 0.61 mol) were added. The suspension was stirred at rt for 1 h. The mixture was filtered over a glass filter (p2) and the filter residue was washed with ethyl acetate until colourless. The filtrate was concentrated to a volume of ca. 500 mL and the brown solution was poured into a suspension of heptanes (2.5 L) and celite (200 g). The solids were filtered off and washed with heptanes (1 L). The filtrate was disposed, and the residue was extracted with ethyl acetate (1.5 L), until colourless. The ethyl acetate filtrate was washed with citric acid (10%, aq., 5 x 500 mL), NaHCO₃ (sat., aq., 2 x 500 mL) and brine (500 mL). The organic layer was dried over Na₂SO₄ and concentrated in vacuo to afford a brown solid (68 g). The crude product was re-crystallized from DCE (300 mL), the solids were filtered off and the filter cake was washed with small volume of cold DCE. The filter dry solids were dried in vacuo (50 °C) to furnish a beige solid (42 g, yield: 50%, purity: 100% (qNMR), 97.4% ee). ¹H-NMR (400 MHz, DMSO-d6) δ 9.59 (s, 1H), 7.57 (s, 1H), 7.42 – 7.37 (m, 4H), 7.37 – 7.30 (m, 1H), 6.20 (s, 1H), 5.31 (d, 1H), 5.12 (s, 2H), 4.73 (t, 1H), 4.41 (q, 1H), 3.62 – 3.43 (m, 2H). ¹³C-NMR (101 MHz, DMSO-d₆) δ 155.59, 153.72, 137.13, 128.92, 128.68, 128.49, 128.46, 126.24, 101.94, 68.35, 66.38, 64.83. HRMS (ESI) calculated for C₁₄H₁₅NO₅ (M+H⁺):279.1056; found (M+H⁺): 279.1057.

S8. Synthesis of DAP (14)

Benzyl ((3aS,4S,7S,7aR)-7-((R)-1,2-dihydroxyethyl)-1,3-dioxo-2-phenyl-2,3,3a,4,7,7a-hexahydro-1H-4,7epoxyisoindol-5-yl)carbamate (DAP 14). A solution of N-phenylmaleimide (2.8 g, 1.5 eq., 16 mmol) in chloroform (10 mL) was cooled to -10 °C. To the yellow solution was added benzyl (R)-(5-(1,2-dihydroxyethyl)furan-3yl)carbamate (Di-HCF 10) (3.0 g, 1 eq., 11 mmol) and the resulting suspension was stirred at -10 °C for 16 h. The brown solution was loaded as such on a column cartridge (330 g) and purified by automated flash chromatography (dichloromethane-methanol (0% \rightarrow 6%), figure 1). The combined fractions with product (ca. 0.5 L) were washed with water (4x 300 mL) and brine-water (1:1) (300 mL). The organic phase was dried over Na₂SO₄ and concentrated *in vacuo* (35 °C) to afford the product as an off-white foam (2.8 g, yield: 58%). ¹H-NMR (400 MHz, DMSO- d_6) δ 9.92 (s, 1H), 7.53 – 7.45 (m, 2H), 7.45 – 7.38 (m, 5H), 7.38 – 7.31 (m, 1H), 7.24 – 7.19 (m, 2H), 5.93 (s, 1H), 5.21 (s, 1H), 5.13 (s, 2H), 5.12 (d, 1H), 4.53 (t, 1H), 3.98 (dd, 1H), 3.63 – 3.37 (m, 2H), 3.30 (d, 1H), 3.20 (d, 1H). ¹³C-NMR (101 MHz, DMSO- d_6) δ 175.67, 174.31, 153.50, 144.69, 136.70, 132.66, 129.41, 128.96, 128.83, 128.63, 128.57, 127.28, 110.48, 93.87, 80.53, 69.99, 66.77, 63.77, 50.96, 50.24. HRMS (ESI) calculated for C₂₄H₂₂N₂O₇ (M+H⁺): 474.1353; found (M+H⁺): 474.1344.



Figure 1. Automated flash chromatography of **DAP** (14). Eluent: DCM-MeOH ($0\% \rightarrow 6\%$).

S9. Synthesis of DAPRED (15)

Benzyl ((3aS,4S,5S,7S,7aR)-7-((R)-1,2-dihydroxyethyl)-1,3-dioxo-2-phenyloctahydro-1H-4,7-epoxyisoindol-5yl)carbamate (DAPRED 15). To a solution of benzyl ((3aS,4S,7S,7aR)-7-((R)-1,2-dihydroxyethyl)-1,3-dioxo-2-phenyl-2,3,3a,4,7,7a-hexahydro-1H-4,7-epoxyisoindol-5-yl)carbamate (DAP 14) (1.5 g, 1 eq, 3.3 mmol) in THF (20 mL) was added platinum/C (0.32 g, 5% wt., 0.025 eq., 83 µmol). The suspension was placed under a hydrogen atmosphere (1 atm, balloon) and was stirred for 1 h. The mixture was filtered over a disposable syringe filter (0.45 µM) and the filtrate was concentrated *in vacuo* afford a white foam (1.5 g, yield: quant.). ¹H-NMR (400 MHz, DMSO- d_6) δ 7.86 (d, 1H), 7.52 – 7.44 (m, 2H), 7.43 – 7.37 (m, 5H), 7.37 – 7.30 (m, 1H), 7.27 – 7.20 (m, 2H), 5.11 (d, 1H), 5.07 (d, 2H), 4.71 (d, 1H), 4.55 (t, 1H), 3.98 (dd, 1H), 3.93 (q, 1H), 3.58 (t, 2H), 3.48 (d, 1H), 3.15 (d, 1H), 2.10 (t, 1H), 1.51 (dd, 1H). ¹³C-NMR (101 MHz, DMSO- d_6) δ 177.29, 175.57, 156.48, 137.27, 133.00, 129.26, 128.90, 128.70, 128.48, 128.44, 127.28, 91.72, 80.21, 71.86, 66.21, 62.77, 52.13, 50.27, 46.96, 36.94. HRMS (ESI) calculated for C₂₄H₂₄N₂O₇ (M+H⁺): 454.1690; found (M+H⁺): 454.1690.

S10. Synthesis of DAPRED-Bz (16)

(R)-2-((3aR,4S,6S,7S,7aS)-6-(((Benzyloxy)carbonyl)amino)-1,3-dioxo-2-phenyloctahydro-4H-4,7-epoxyisoindol-4-

yl)-2-hydroxyethyl benzoate (DAPRED-Bz 16). To a solution of benzyl ((3aS,4S,5S,7S,7aR)-7-((R)-1,2-dihydroxyethyl)-1,3-dioxo-2-phenyloctahydro-1H-4,7-epoxyisoindol-5-yl)carbamate (DAPRED 15) (1.5 g, 1 eq.,3.3 mmol) and triethylamine (0.75 g, 1.0 mL, 2.25 eq., 7.5 mmol) in dichloromethane (20 mL) was added benzoyl chloride (0.93 g, 0.77 mL, 2 eq., 6.6 mmol), while maintaining the temperature below 25 °C. The clear mixture was stirred at room temperature for 1 h and subsequently diluted with ethyl acetate (200 mL). The solution was washed with citric acid (10%, aq., 2 x 150 mL), NaHCO₃ (sat., aq., 2 x 150 mL) and brine (150 mL). The organic phase was dried over Na₂SO₄ and concentrated *in vacuo* to afford an off-white foam. The crude product was purified by automated column chromatography (40 g column, heptanes-ethyl acetate (0% \rightarrow 75%)) to afford the product as a white solid (1.5 g, yield: 81%). ¹H-NMR (400 MHz, DMSO- *d*₆) δ 8.04 (d, 2H), 7.91 (d, 1H), 7.70 – 7.61 (m, 1H), 7.53 (t, 2H), 7.51 – 7.49 (m, 1H), 7.49 – 7.45 (m, 2H), 7.44 – 7.30 (m, 5H), 7.25 (d, 2H), 5.77 (d, 1H), 5.09 (s, 2H), 4.78 (d, 1H), 4.46 (td, 2H), 4.41 - 4.32 (m, 1H), 4.10 - 3.94 (m, 1H), 3.53 (d, 1H), 3.27 (d, 1H), 2.25 (t, 1H), 1.60 (dd, 1H). 13 C-NMR (101 MHz, DMSO- d_6) δ 177.23, 175.31, 166.23, 156.47, 137.23, 133.82, 132.91, 130.17, 129.83, 129.31, 129.16, 128.90, 128.79, 128.51, 128.46, 127.31, 90.95, 80.68, 68.17, 66.30, 66.24, 52.07, 50.49, 46.81, 36.50. HRMS (ESI) calculated for $C_{31}H_{28}N_2O_8$ (M+H⁺): 558.1952; found (M+H⁺): 558.1935.

S11. Synthesis of 18

N-((3aS,4S,7S,7aR)-7-((R)-1,2-dihydroxyethyl)-1,3-dioxo-2-phenyl-2,3,3a,4,7,7a-hexahydro-1H-4,7-epoxyisoindol-5-yl)acetamide (18). A solution of *N*-phenylmaleimide (2.8 g, 1.5 eq., 16 mmol) in chloroform (10 mL) was cooled to -10 °C. To the yellow solution was added dihydroxyethyl acetamidofuran (Di-HAF (1)) (2.0 g, 1 eq., 11 mmol) and DMSO (1.5 mL). The resulting suspension was stirred at -10 °C for 16 h. The brown solution was poured into a suspension of Celite (6 g) in diisopropyl ether (150 mL). The solids were filtered off and washed with diisopropyl ether (2 x 50 mL) and *tert*-butyl methyl ether (2 x 25 mL). The filter residue was ground to a fine powder with pestle and mortar and was used as solid load for purification by automated flash chromatography (330 g column, dichloromethane-methanol (0% →10%)). The combined fractions with product were extracted with water (3x 200 mL). The aqueous phase was saturated with sodium chloride and subsequently extracted with EtOAc (3 x 300 mL) The organic phase was dried over Na₂SO₄, diluted with toluene (0.5 L) and concentrated *in vacuo* (25 °C) to afford the product as a white solid (2.3 g, yield: 59%). ¹H-NMR (400 MHz, DMSO-*d*₆) δ 10.16 (s, 1H), 7.54 – 7.46 (m, 2H), 7.45 – 7.37 (m, 1H), 7.22 (d, 2H), 6.15 (s, 1H), 5.21 (s, 1H), 5.12 (d, 1H), 4.53 (t, 1H), 4.05 – 3.93 (m, 1H), 3.63 – 3.39 (m, 2H), 3.28 (d, 1H), 3.19 (d, 1H), 1.98 (s, 3H). ¹³C-NMR (101 MHz, DMSO-*d*₆) δ 175.77, 174.33, 168.86, 144.25, 132.66, 129.41, 128.83, 127.28, 112.28, 93.66, 80.71, 70.01, 63.80, 50.79, 50.26, 23.71. HRMS (ESI) calculated for C₁₈H₁₈N₂O₆ (M+H⁺): 359.1238; found (M+H⁺): 359.1230.

Characterization

S12. Periodate mediated diol oxidation Reaction mixture, *before & after*





S13. Di-HCF (10)

¹H-NMR of Di-HCF (**10**) in DMSO- d_6





DSC of Di-HCF (10)





HRMS of Di-HCF (10)



Chiral LC analysis of Di-HCF (10)



Channel Description PDA Ch1 210nm@4.8nm -Compens.

Processed Channel: PDA Ch1 210nm@4.8nm -Compens.

	RT (Min)	Area	Height	Area %
1	5.798	4162073	1121923	98.73
2	6.152	53498	15690	1.27

S14. DAP (14)

¹H-NMR of DAP (**14**) in DMSO- d_6





DSC of DAP (14)











¹H-NMR of DAPRED (**15**) in DMSO- d_6



¹³C-NMR of DAPRED (**15**) in DMSO- d_6



DSC of DAPRED (15)









¹H-NMR of DAPRED-Bz (**16**) in DMSO- d_6



¹³C-NMR of DAPRED (**16**) in DMSO- d_6



DSC of DAPRED-Bz (16)







S17. Compound **18**

¹H-NMR of **18** in DMSO- d_6



¹³C-NMR of **18** in DMSO- d_6



DSC of **18**





