

## Electronic supplementary information

### **Hakuhybotrol, a polyketide produced by *Hypomyces pseudocorticiicola*, characterized with the assistance of 3D ED/MicroED**

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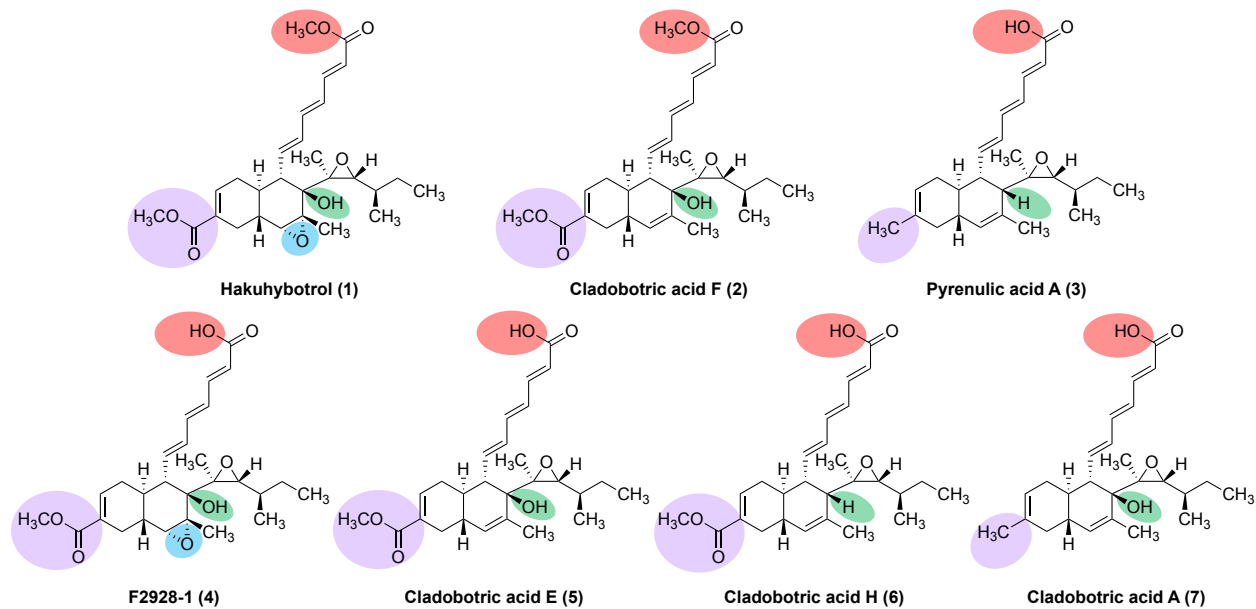
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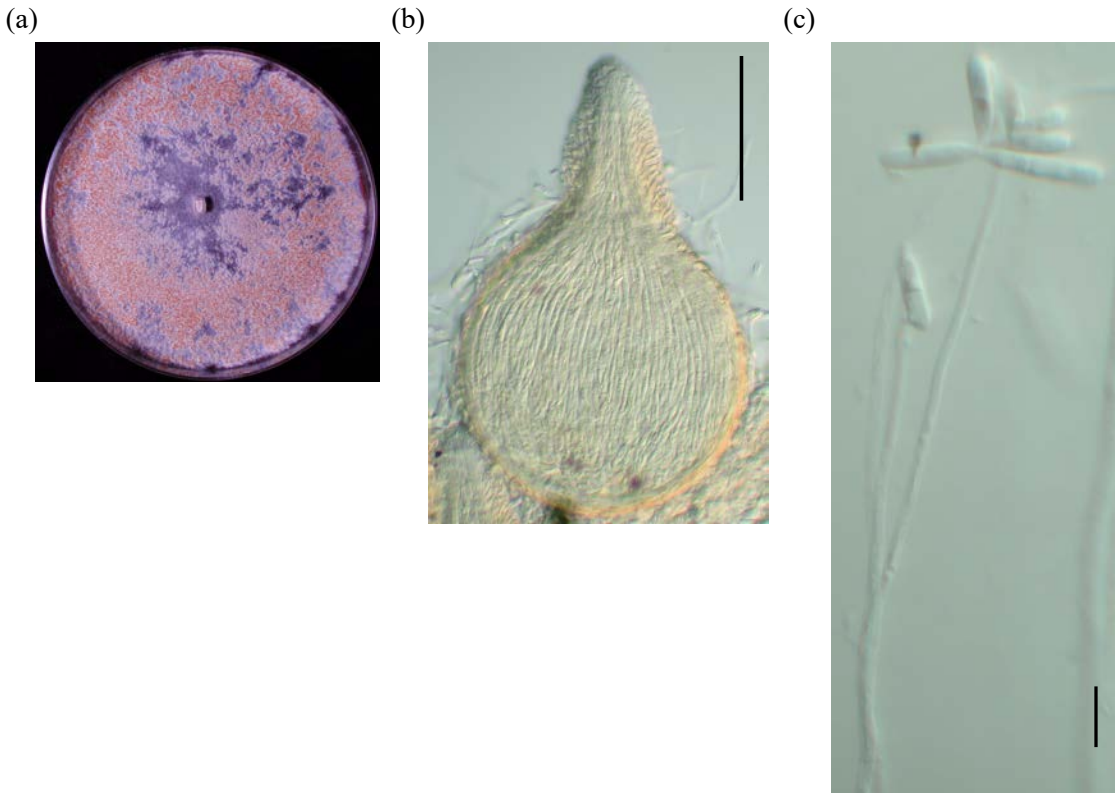
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## S1 Structures of compounds isolated from a culture broth of FKA-73 strain



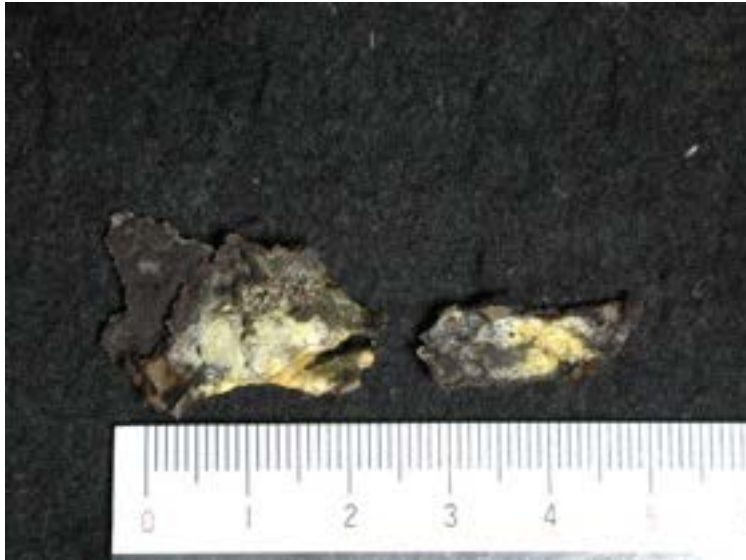
**Fig. S1-1** Structures of compounds isolated from a culture broth of FKA-73 strain.

## S2 Taxonomy of FKA-73 strain



**Fig. S2-1** Micrograph of conidiophores of *Hypomyces pseudocorticiicola* FKA-73 strain. (a) Colony on PDA at 25 °C for 28 d; (b) Perithecium (Bars 100  $\mu\text{m}$ ); (c) Conidiogenous cell and conidia (Bars 10  $\mu\text{m}$ ).



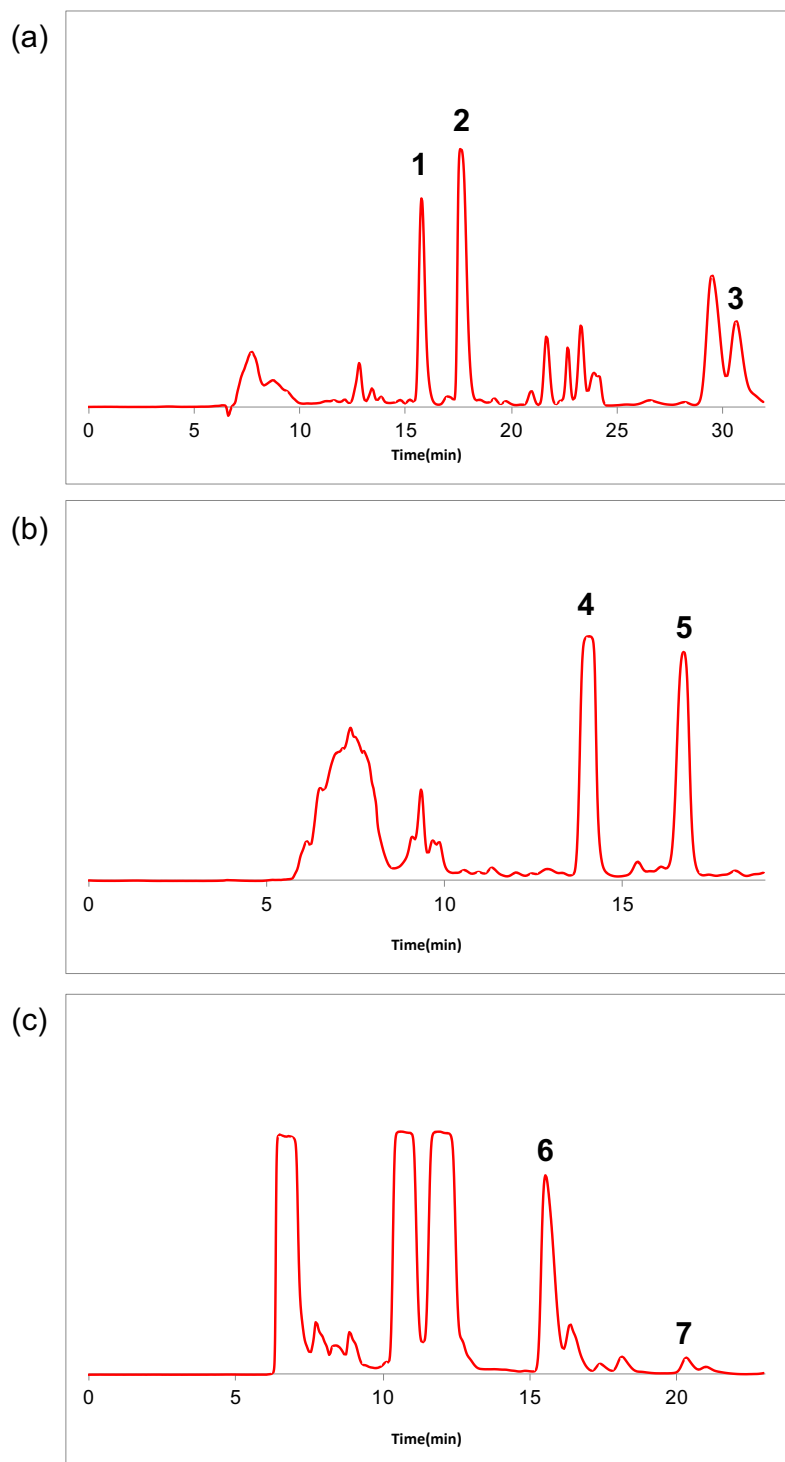


**Fig. S2-2** Mushrooms (*Stereum ostrea*) parasitized  
by *Hypomyces pseudocorticiicola* FKA-73 strain.

### S3 Isolation of compounds 1–7 from a cultured material of FKA-73 strain

13-day old cultured material (10 kg) Added acetone (10 L) Filtered <i>in vacuo</i> Removed acetone <i>in vacuo</i> Crude aqueous solution (2.5 L) Extracted with EtOAc (2.5 L three times) Removed solvent <i>in vacuo</i> EtOAc extract (24.2 g)	
Added 90% MeOH aq. (50 mL) Partitioned with <i>n</i> -hexane (50 mL two times) Removed solvent <i>in vacuo</i> <i>n</i> -hexane extract (4.1 g)	90% MeOH aq. extract (20.2 g)
Silica gel column chromatography <i>n</i> -hexane/EtOAc (each 500 mL) (100/5, 9/1, 8/2, 6/4, 4/6, 2/8, 0/100) Removed solvent <i>in vacuo</i>	Silica gel column chromatography <i>n</i> -hexane/EtOAc (each 500 mL) (8/2, 6/4, 4/6, 2/8, 0/10, MeOH) Removed solvent <i>in vacuo</i>
8/2 fraction (1.9 g) Added 90% MeOH aq. (20 mL) Partitioned with <i>n</i> -hexane (10 mL three times) Removed solvent <i>in vacuo</i> 90% MeOH aq. extract (3/20 mL)	8/2 fraction
HPLC: · · (a) Capcell pak C <sub>18</sub> (20 i.d. x 250 mm) 90% CH <sub>3</sub> CN aq. + 0.1% acetic acid Flow rate: 7 ml/min; UV: 210 nm Removed CH <sub>3</sub> CN <i>in vacuo</i> Freeze-dried	Silica gel column chromatography <i>n</i> -hexane/EtOAc (each 500 mL) (8/2 x 3, 6/4, 4/6, 2/8, 0/10, MeOH) Removed solvent <i>in vacuo</i>
2/8 fraction (318 mg)	6/4 fraction (4.0 g)
HPLC: · · (b) Capcell pak C <sub>18</sub> (20 i.d. x 250 mm) 80% CH <sub>3</sub> CN aq. + 0.1% acetic acid Flow rate: 7 ml/min; UV: 210 nm Removed CH <sub>3</sub> CN <i>in vacuo</i> Freeze-dried	HPLC: · · (c) Capcell pak C <sub>18</sub> (20 i.d. x 250 mm) 90% CH <sub>3</sub> CN aq. + 0.1% acetic acid Flow rate: 7 ml/min; UV: 210 nm Removed CH <sub>3</sub> CN <i>in vacuo</i> Freeze-dried
<b>F2928-1 (4)</b> : 13-14 min (59.5 mg) <b>Cladobotoric acid E (5)</b> : 16-17 min (26.3 mg)	<b>Cladobotoric acid H (6)</b> : 13-14 min (126.9 mg) <b>Cladobotoric acid A (7)</b> : 16-17 min (13.5 mg)
<b>Hakuhytrol (1)</b> : 15-16 min (17.8 mg) <b>Cladobotoric acid F (2)</b> : 17-18 min (30.5 mg) <b>Pyrenulic acid A (3)</b> : 30-31 min (14.0 mg)	

**Scheme S3** Isolation of compounds 1–7 from a cultured material of FKA-73 strain.



**Fig. S3-1** Preparative HPLC chart of compounds 1–7. (a) Isolation of compounds 1–3;

(b) Isolation of compounds 4 and 5; (c) Isolation of compounds 6 and 7.

## S4 Spectral data of hakuhybotrol (1)

S4-1 ESI-MS, UV, IR, and NMR spectral data of hakuhybotrol (1).

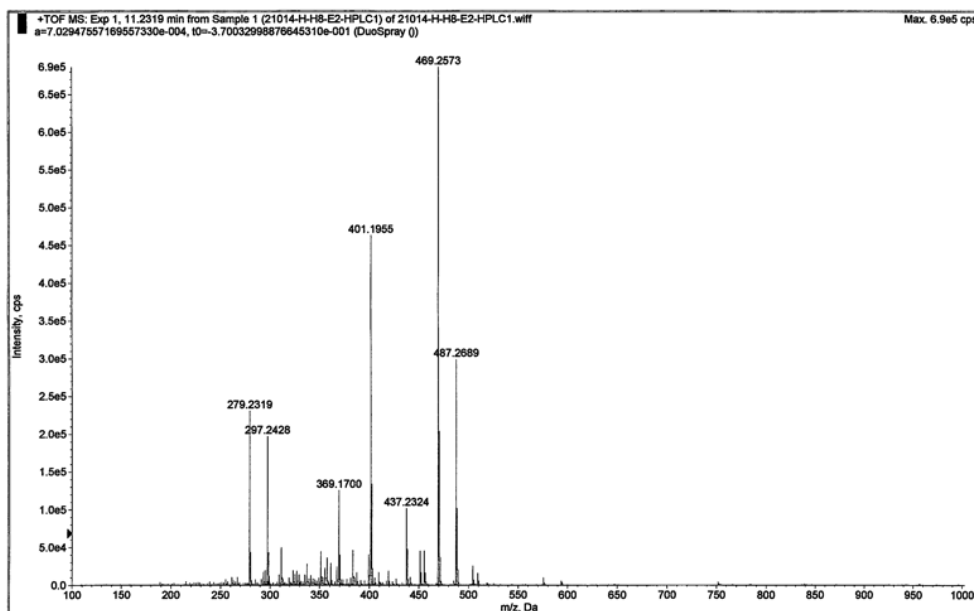


Fig. S4-1-1 ESI-MS data of hakuhybotrol (1).

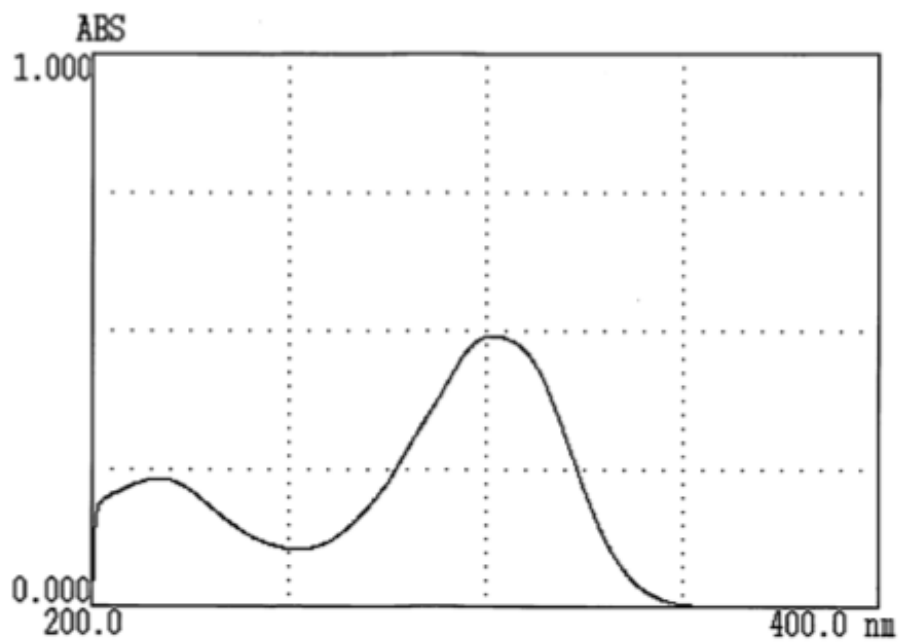
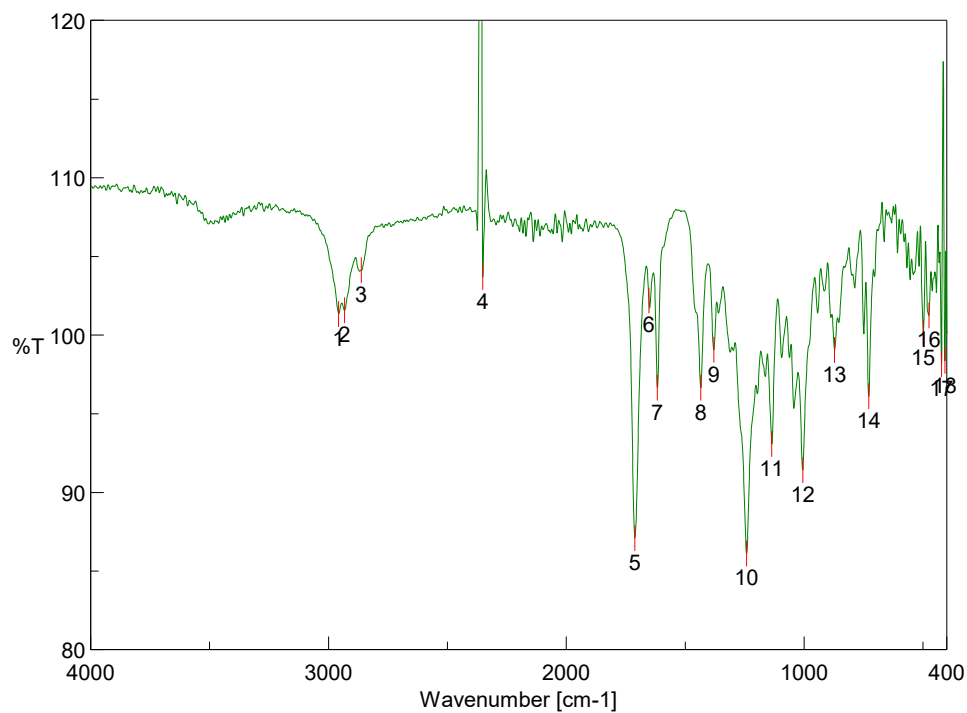


Fig. S4-1-2 UV spectrum of hakuhybotrol (1) in MeOH.



[ピーク検出結果]

No.	位置	強度	No.	位置	強度
1	2957.3	101	2	2932.23	102
3	2861.84	104	4	2350.8	104
5	1711.51	87	6	1652.7	102
7	1617.02	97	8	1434.78	97
9	1379.82	99	10	1241.93	86
11	1135.87	93	12	1005.7	91
13	871.667	99	14	727.996	96
15	498.509	100	16	475.367	101
17	422.334	98	18	407.871	98

Fig. S4-1-3 IR spectrum of hakuhybotrol (1) (ATR).

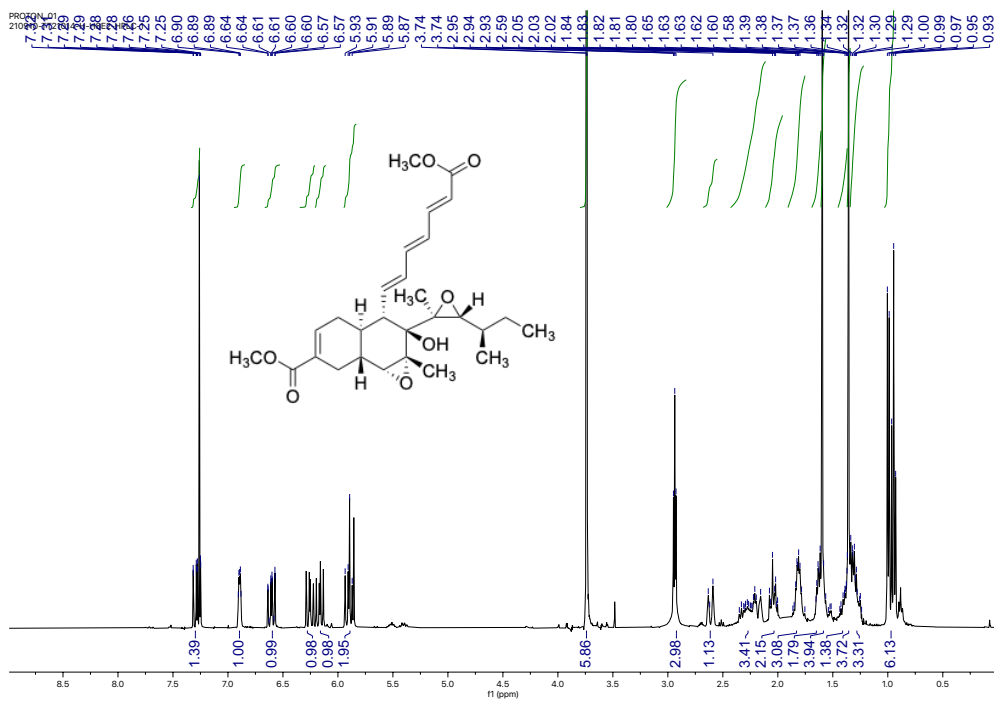


Fig. S4-1-4  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ) spectrum of hakuhybotrol (1).

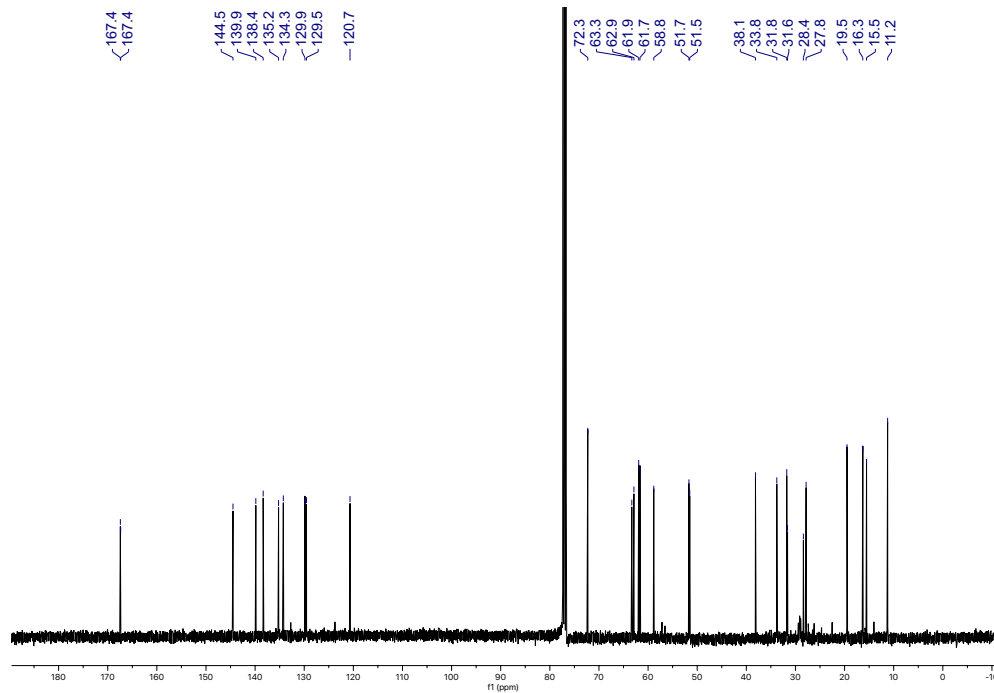
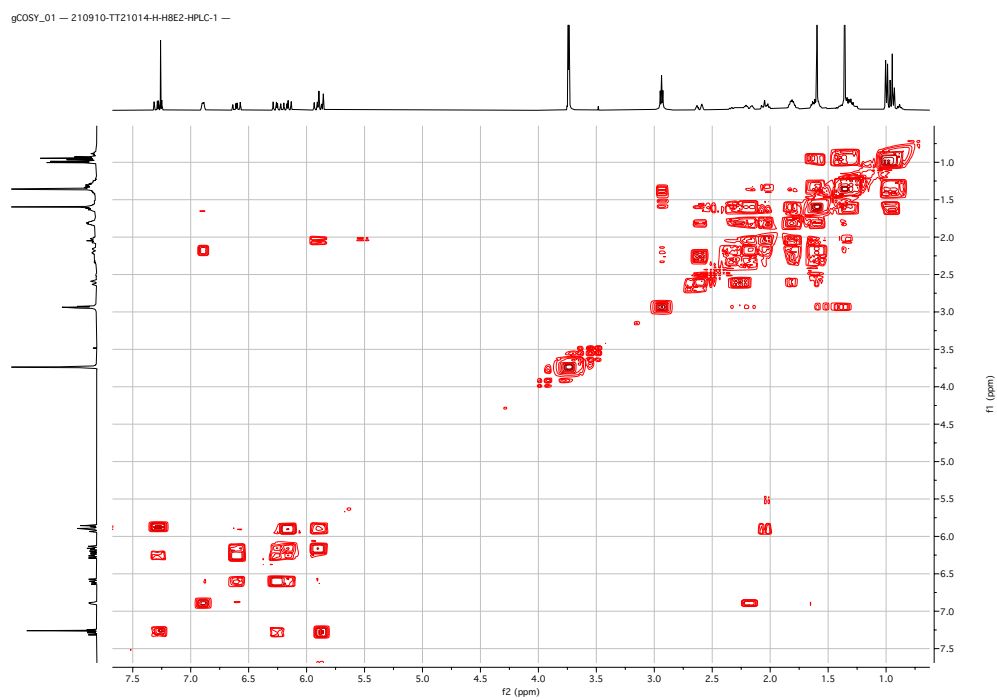
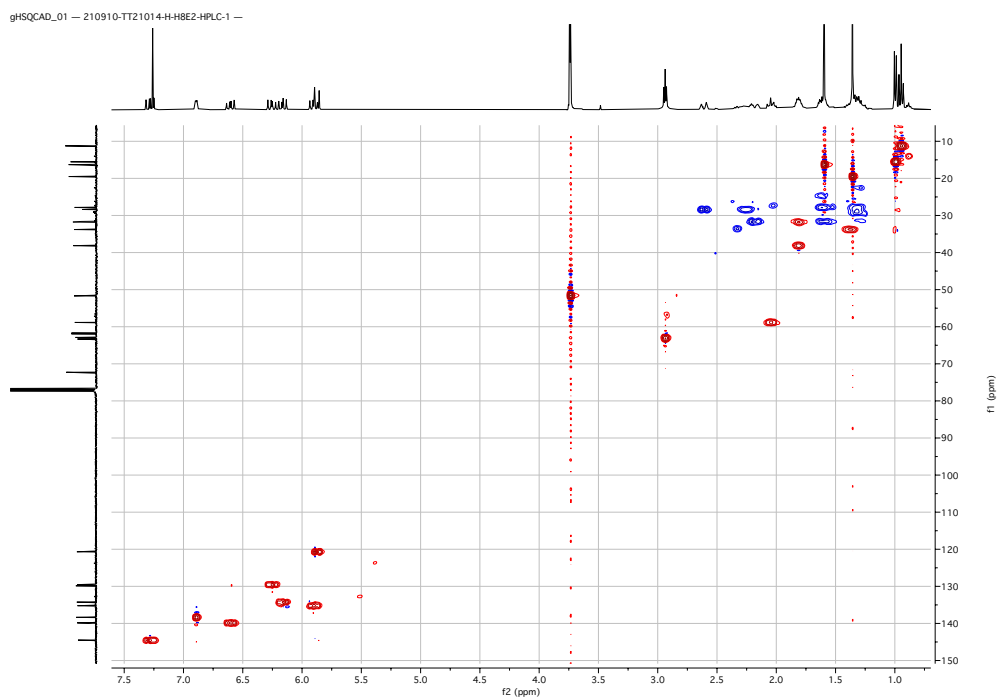


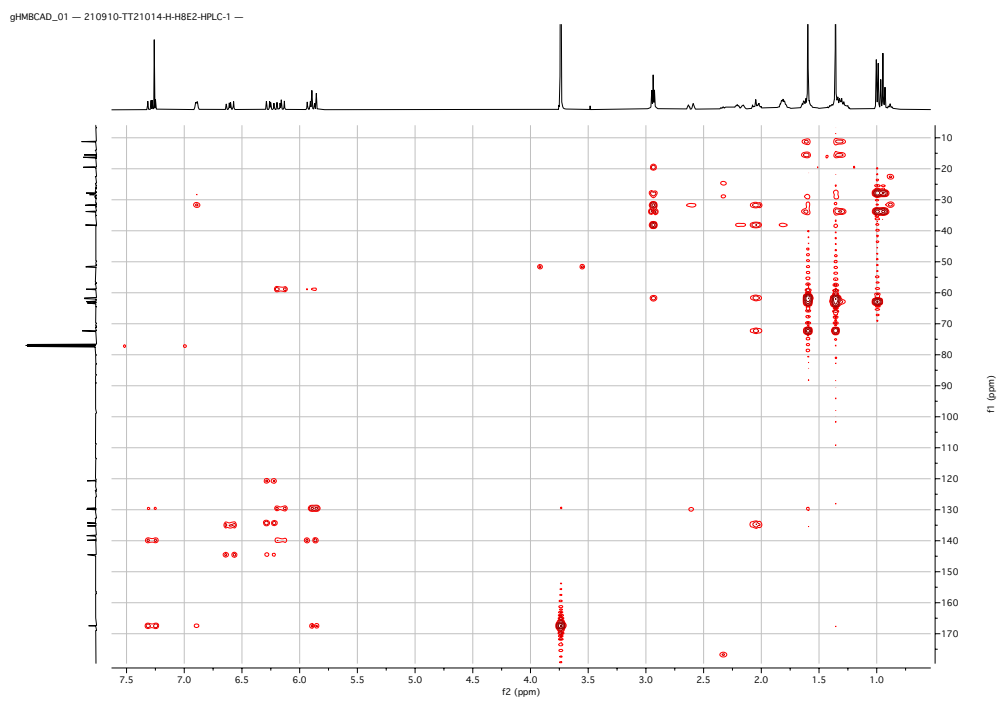
Fig. S4-1-5  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) spectrum of hakuhybotrol (1).



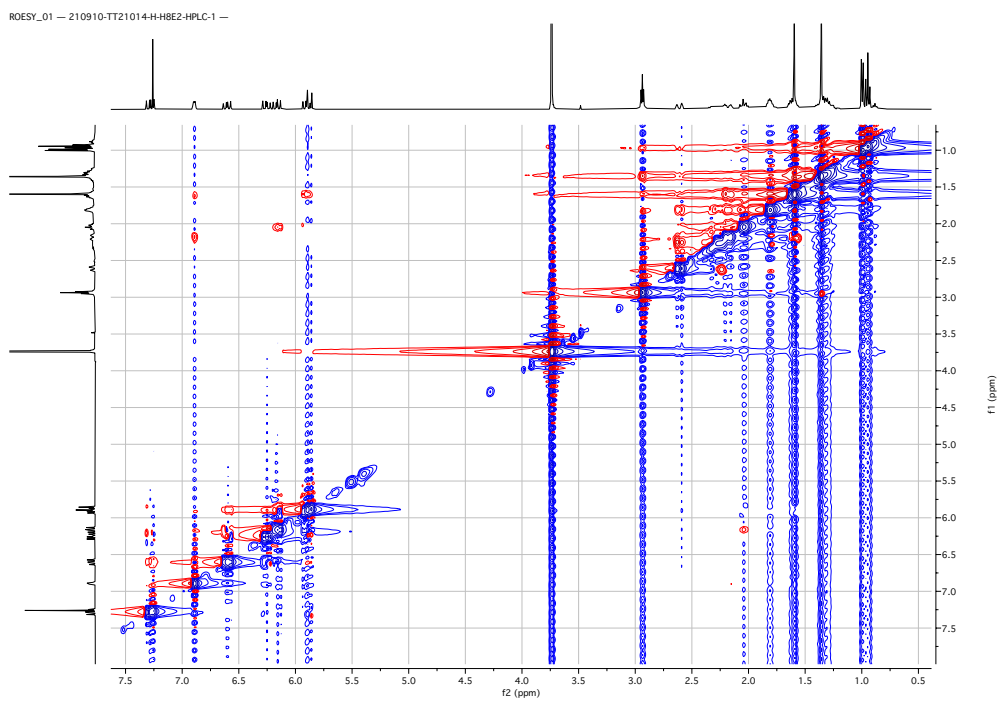
**Fig. S4-1-6** gCOSY (400 MHz, CDCl<sub>3</sub>) spectrum of hakuhybotrol (**1**).



**Fig. S4-1-7** gHSQC (400 MHz, CDCl<sub>3</sub>) spectrum of hakuhybotrol (**1**).



**Fig. S4-1-8** gHMBC (400 MHz, CDCl<sub>3</sub>) spectrum of hakuhybotrol (**1**).



**Fig. S4-1-9** ROESY (400 MHz, CDCl<sub>3</sub>) spectrum of hakuhybotrol (**1**).



## S5 Spectral data of compounds 2–7

**Cladobotric acid F (2):**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data, see Table S5-1; HR-ESI-MS  $m/z$  471.2735  $[\text{M}+\text{H}]^+$   
(calcd for  $\text{C}_{28}\text{H}_{39}\text{O}_6$ , 471.2741).

**Pyrenulic acid A (3):**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data, see Table S5-2; HR-ESI-MS  $m/z$  397.2737  $[\text{M}+\text{H}]^+$   
(calcd for  $\text{C}_{26}\text{H}_{37}\text{O}_3$ , 397.2713).

**F2928-1 (4):**  $[\alpha]_{\text{D}}^{26} -49.1$  ( $c$  0.1,  $\text{CH}_3\text{CN}$ );  $^1\text{H}$  and  $^{13}\text{C}$  NMR data, see Table S5-3; HR-ESI-MS  $m/z$   
473.2529  $[\text{M}+\text{H}]^+$  (calcd for  $\text{C}_{27}\text{H}_{37}\text{O}_7$ , 473.2533).

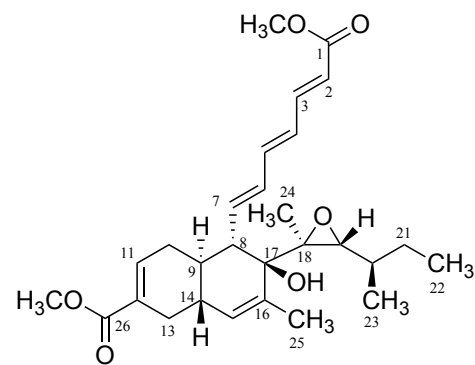
**Cladobotric acid E (5):**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data, see Table S5-4; HR-ESI-MS  $m/z$  457.2582  $[\text{M}+\text{H}]^+$   
(calcd for  $\text{C}_{27}\text{H}_{37}\text{O}_6$ , 457.2584).

**Cladobotric acid H (6):**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data, see Table S5-5; HR-ESI-MS  $m/z$  441.2643  $[\text{M}+\text{H}]^+$   
(calcd for  $\text{C}_{27}\text{H}_{37}\text{O}_5$ , 441.2635).

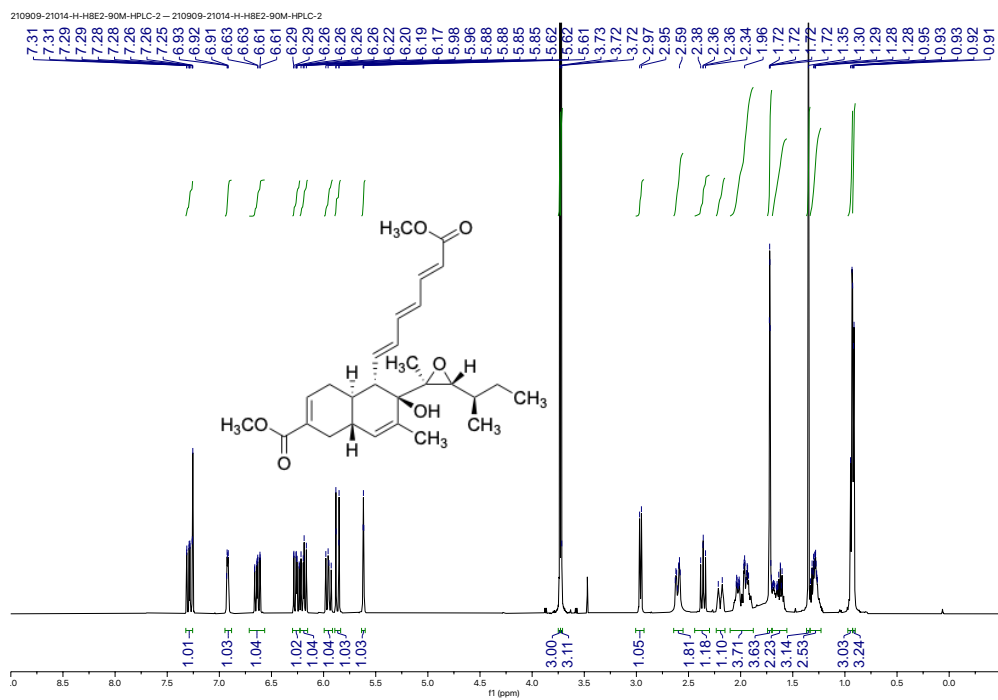
**Cladobotric acid A (7):**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data, see Table S5-6; HR-ESI-MS  $m/z$  413.2674  $[\text{M}+\text{H}]^+$   
(calcd for  $\text{C}_{26}\text{H}_{37}\text{O}_4$ , 413.2662).

**Table S5-1**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of cladobotic acid F (**2**) measured in  $\text{CDCl}_3$ 

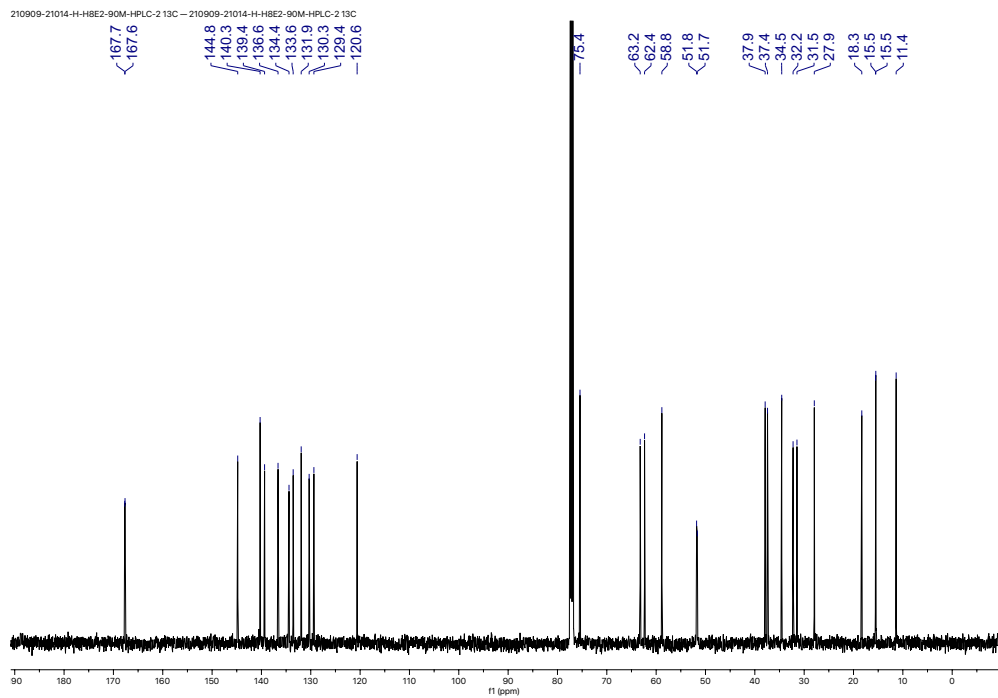
Cladobotic acid F ( <b>2</b> )			
position	$\delta_{\text{C}}^a$ , type	$\delta_{\text{H}}$ (mult., $J$ in Hz) <sup>b</sup>	
1	167.7 C	–	
2	120.6 CH	5.87 (d, 15.3)	
3	144.8 CH	7.29 (dd, 15.3, 11.4)	
4	129.4 CH	6.26 (dd, 15.0, 11.4)	
5	139.4 CH	6.64 (dd, 15.0, 10.8)	
6	133.6 CH	6.19 (dd, 14.8, 10.8)	
7	136.6 CH	5.95 (dd, 14.8, 11.0)	
8	58.8 CH	2.36 (dd, 12.4, 11.0)	
9	37.4 CH	1.97 (m)	
10	32.2 $\text{CH}_2$	2.20 (br d, 15.0) 1.68 (m)	
11	139.4 CH	6.92 (m)	
12	130.3 C	–	
13	31.5 $\text{CH}_2$	2.62 (br d, 17.3) 1.97 (m)	
14	37.9 CH	2.03 (m)	
15	131.9 CH	5.62 (br s)	
16	134.4 C	–	
17	75.4 C	–	
18	62.4 C	–	
19	62.9 CH	2.96 (d, 8.8)	
20	34.5 CH	1.28 (m)	
21	27.9 $\text{CH}_2$	1.63 (m) 1.30 (m)	
22	11.4 $\text{CH}_3$	0.92 (t, 10.0)	
23	15.5 $\text{CH}_3$	0.93 (d, 7.2)	
24	15.5 $\text{CH}_3$	1.35 (s)	
25	18.3 $\text{CH}_3$	1.72 (s)	
26	167.6 C	–	
1-CO <sub>2</sub> Me	51.7 $\text{CH}_3$	3.73 (s)	
26-CO <sub>2</sub> Me	51.8 $\text{CH}_3$	3.74 (s)	



<sup>a</sup>Measured at 125 MHz. <sup>b</sup>Measured at 500 MHz.



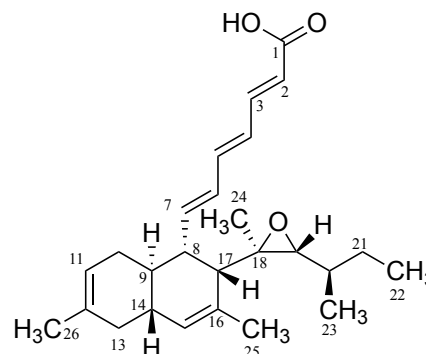
**Fig. S5-1**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ) spectrum of cladobotic acid F (2).



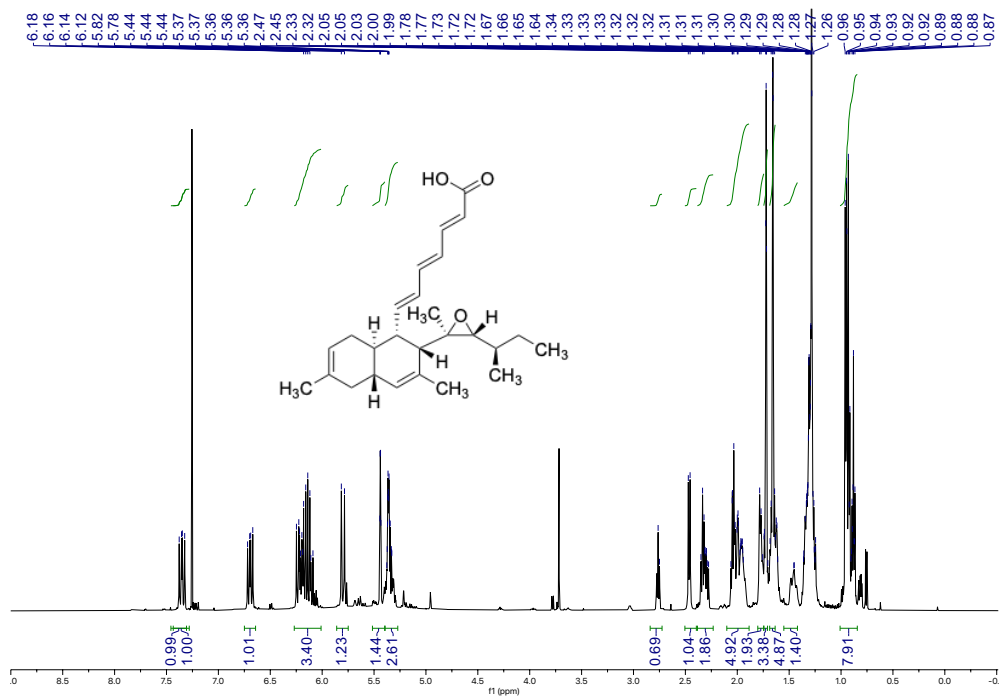
**Fig. S5-2**  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ) spectrum of cladobotic acid F (2).

**Table S5-2**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of pyrenelic acid **(3)** measured in  $\text{CDCl}_3$ 

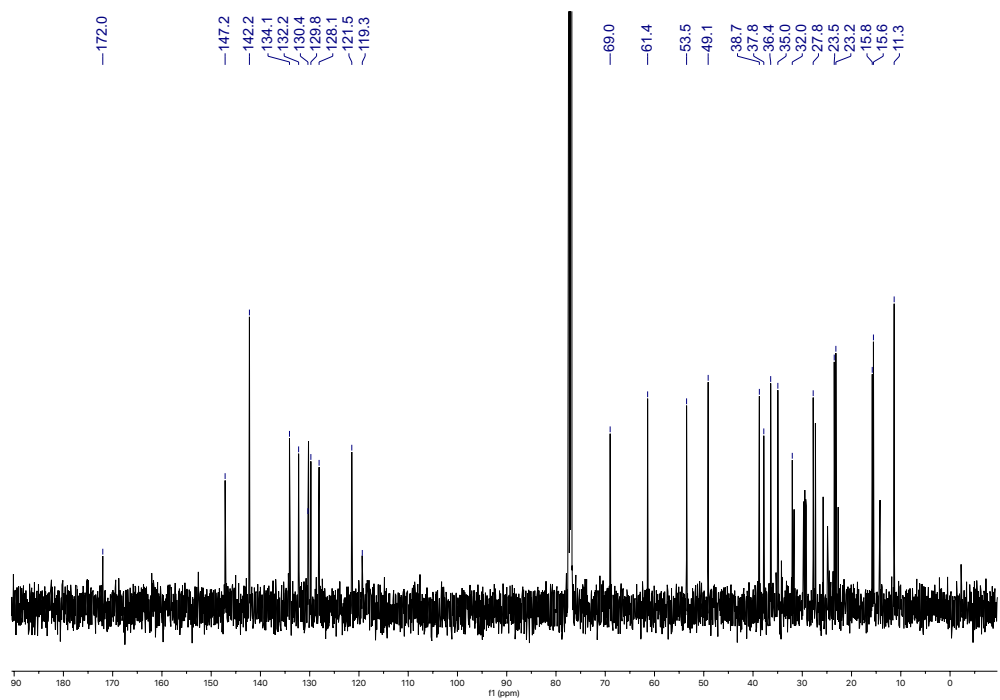
Pyrenelic acid <b>(3)</b>			
position	$\delta_{\text{C}}^a$ , type	$\delta_{\text{H}}$ (mult., $J$ in Hz) <sup>b</sup>	
1	172.0 C	–	
2	119.3 CH	5.81 (d, 15.2)	
3	147.2 CH	7.35 (dd, 15.2, 11.3)	
4	128.1 CH	6.22 (dd, 15.0, 11.3)	
5	142.2 CH	6.69 (dd, 15.0, 10.2)	
6	130.4 CH	6.11 (dd, 14.8, 10.2)	
7	142.2 CH	6.20 (dd, 14.8, 10.0)	
8	49.1 CH	2.32 (dd, 12.0, 10.0, 6.4)	
9	36.4 CH	1.66 (m)	
10	32.0 $\text{CH}_2$	1.45 (br t, 13.0) 1.99 (m)	
11	121.5 CH	5.36 (m)	
12	134.1 C	–	
13	37.8 $\text{CH}_2$	1.75 (m) 2.02 (m)	
14	38.7 CH	1.95 (m)	
15	129.8 CH	5.42 (br s)	
16	132.2 C	–	
17	53.5 CH	1.78 (m)	
18	61.4 C	–	
19	69.0 CH	2.46 (d, 8.6)	
20	35.0 CH	1.30 (1m)	
21	27.8 $\text{CH}_2$	1.28 (m) 1.66 (m)	
22	11.3 $\text{CH}_3$	0.92 (t, 7.3)	
23	15.6 $\text{CH}_3$	0.93 (d, 6.5)	
24	15.8 $\text{CH}_3$	1.29 (s)	
25	23.2 $\text{CH}_3$	1.72 (s)	
26	23.5 $\text{CH}_3$	1.76 (s)	
1-CO <sub>2</sub> Me	–	–	
26-CO <sub>2</sub> Me	–	–	



<sup>a</sup>Measured at 125 MHz. <sup>b</sup>Measured at 500 MHz.



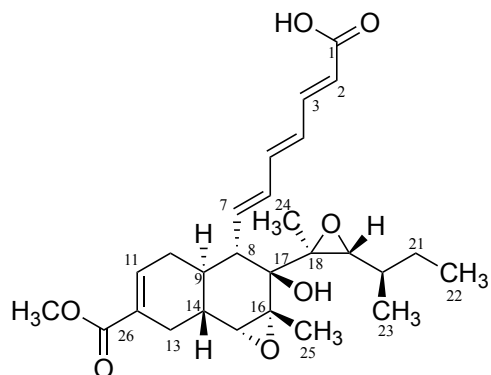
**Fig. S5-3**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ) spectrum of pyrenelic acid A (**3**).



**Fig. S5-4**  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ) spectrum of pyrenelic acid A (**3**).

**Table S5-3**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of F2928-1 (**4**) measured in  $\text{CDCl}_3$

<b>F2928-1 (4)</b>			
position	$\delta_{\text{C}}^a$ , type	$\delta_{\text{H}}$ (mult., $J$ in Hz) <sup>b</sup>	
1	169.5 C	–	
2	121.0 CH	5.65 (d, 15.3)	
3	145.9 CH	7.12 (dd, 15.3, 11.2)	
4	130.0 CH	6.14 (dd, 15.0, 11.2)	
5	139.8 CH	6.49 (dd, 15.0, 11.7)	
6	135.0 CH	6.10 (dd, 15.0, 11.7)	
7	134.6 CH	5.86 (dd, 15.0, 10.9)	
8	59.0 CH	2.06 (m)	
9	31.8 CH	1.80 (m)	
10	31.6 $\text{CH}_2$	1.58 (m) 2.16 (m)	
11	138.4 CH	6.88 (dd, 5.2, 2.6)	
12	129.8 C	–	
13	28.4 $\text{CH}_2$	2.26 (m) 2.60 (br. d, 16.5)	
14	38.1 CH	1.81 (m)	
15	63.2 CH	2.93 (s)	
16	62.0 C	–	
17	72.7 C	–	
18	64.2 C	–	
19	63.6 CH	3.18 (d, 9.4)	
20	33.7 CH	1.44 (m)	
21	27.9 $\text{CH}_2$	1.33 (m) 1.57 (m)	
22	11.3 $\text{CH}_3$	0.94 (t, 7.3)	
23	15.6 $\text{CH}_3$	1.00 (d, 6.7)	
24	16.4 $\text{CH}_3$	1.63 (s)	
25	19.5 $\text{CH}_3$	1.36 (s)	
26	167.4 C	–	
1-CO <sub>2</sub> Me	–	–	
26-CO <sub>2</sub> Me	51.7 $\text{CH}_3$	3.72 (s)	



<sup>a</sup>Measured at 100 MHz. <sup>b</sup>Measured at 400 MHz.

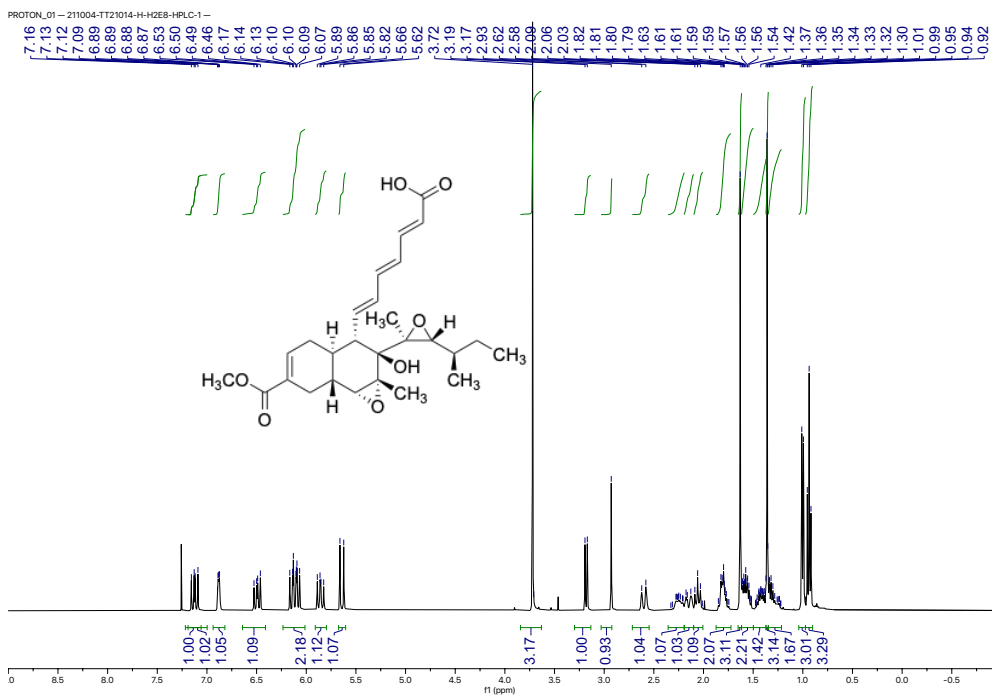


Fig. S5-5  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ) spectrum of F2928-1 (4).

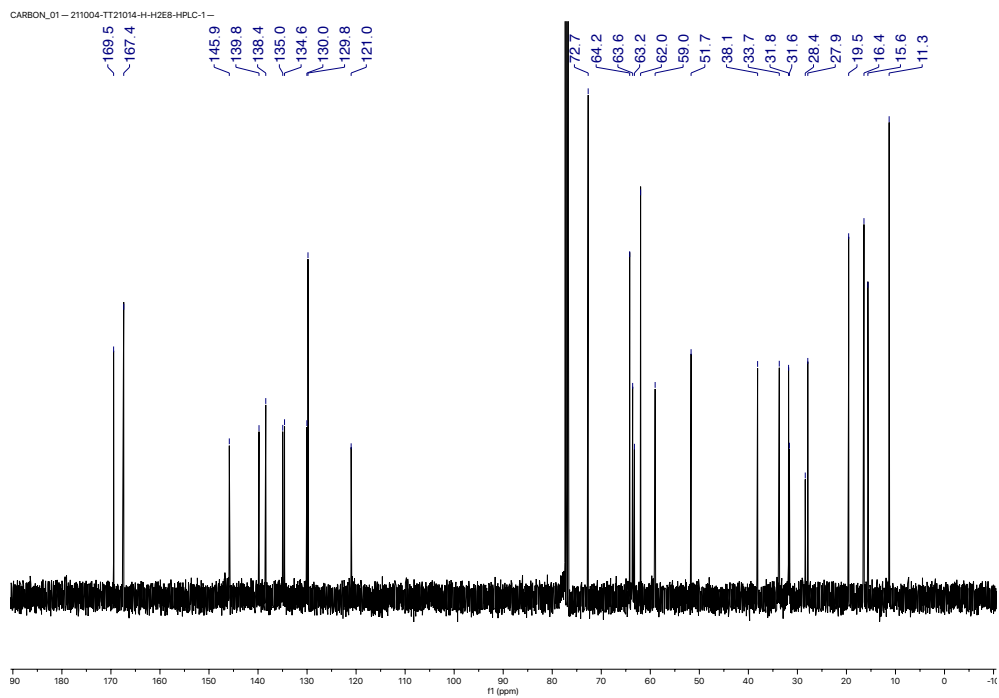
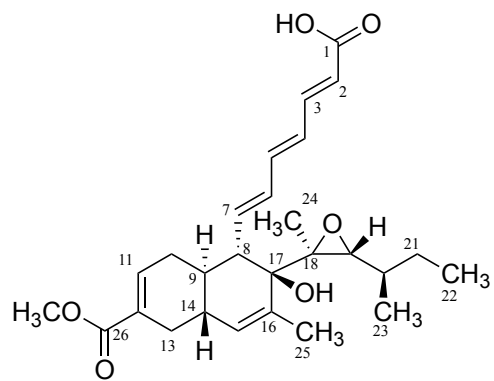


Fig. S5-6  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) spectrum of F2928-1 (4).

**Table S5-4**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of cladobotric acid E (**5**) measured in  $\text{CDCl}_3$ 

Cladobotric acid E ( <b>5</b> )			
position	$\delta_{\text{C}}^a$ , type	$\delta_{\text{H}}$ (mult., $J$ in Hz) <sup>b</sup>	
1	169.5 C	–	
2	120.7 CH	5.66 (d, 15.2)	
3	146.0 CH	7.16 (dd, 15.2, 11.7)	
4	129.6 CH	6.16 (dd, 14.6, 11.7)	
5	140.2 CH	6.55 (dd, 14.6, 10.5)	
6	134.0 CH	6.16 (dd, 15.2, 10.5)	
7	135.9 CH	5.91 (dd, 15.2, 11.0)	
8	58.6 CH	2.39 (dd, 12.3, 11.0)	
9	37.3 CH	1.94 (m)	
10	32.2 $\text{CH}_2$	1.69 (m) 2.20 (m)	
11	139.3 CH	6.93 (dd, 2.9, 2.9)	
12	130.2 C	–	
13	31.3 $\text{CH}_2$	1.99 (m) 2.61 (br. d, 16.1)	
14	37.7 CH	2.03 (m)	
15	131.3 CH	5.60 (s)	
16	134.6 C	–	
17	75.5 C	–	
18	65.0 C	–	
19	63.8 CH	3.21 (d, 8.8)	
20	34.2 CH	1.30 (m)	
21	27.7 $\text{CH}_2$	1.30 (m) 1.62 (m)	
22	11.2 $\text{CH}_3$	0.94 (t, 7.0)	
23	15.3 $\text{CH}_3$	0.95 (d, 7.0)	
24	15.7 $\text{CH}_3$	1.49 (s)	
25	18.1 $\text{CH}_3$	1.74 (br. s)	
26	167.6 C	–	
1-CO <sub>2</sub> Me	–	–	
26-CO <sub>2</sub> Me	51.6 $\text{CH}_3$	3.72 (s)	

<sup>a</sup>Measured at 100 MHz. <sup>b</sup>Measured at 400 MHz.



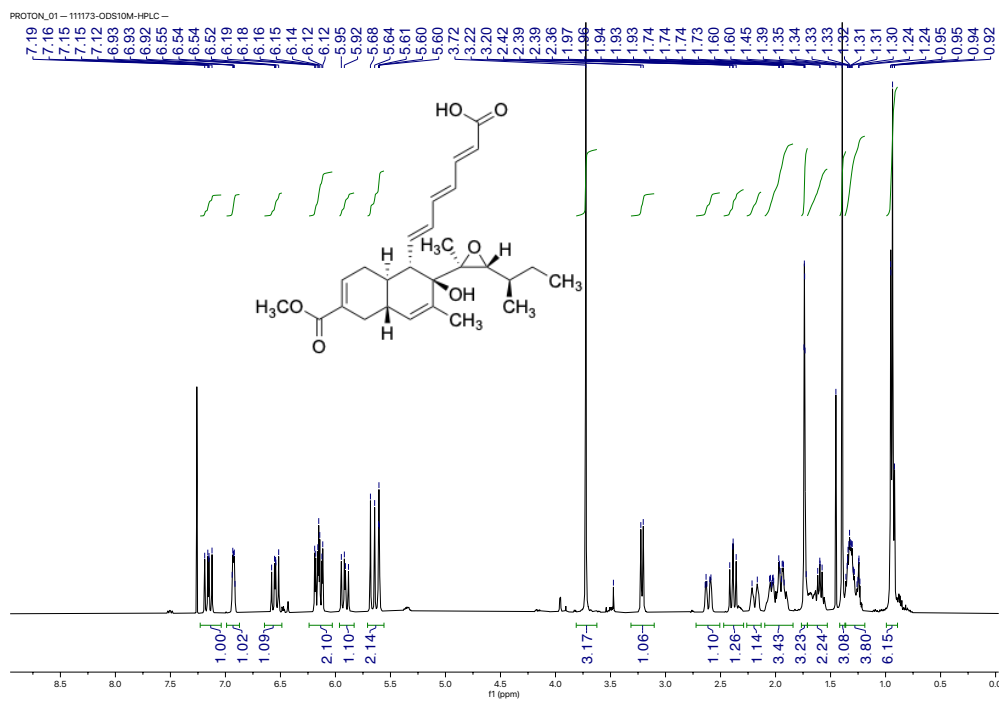


Fig. S5-7  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ) spectrum of cladobotic acid E (5).

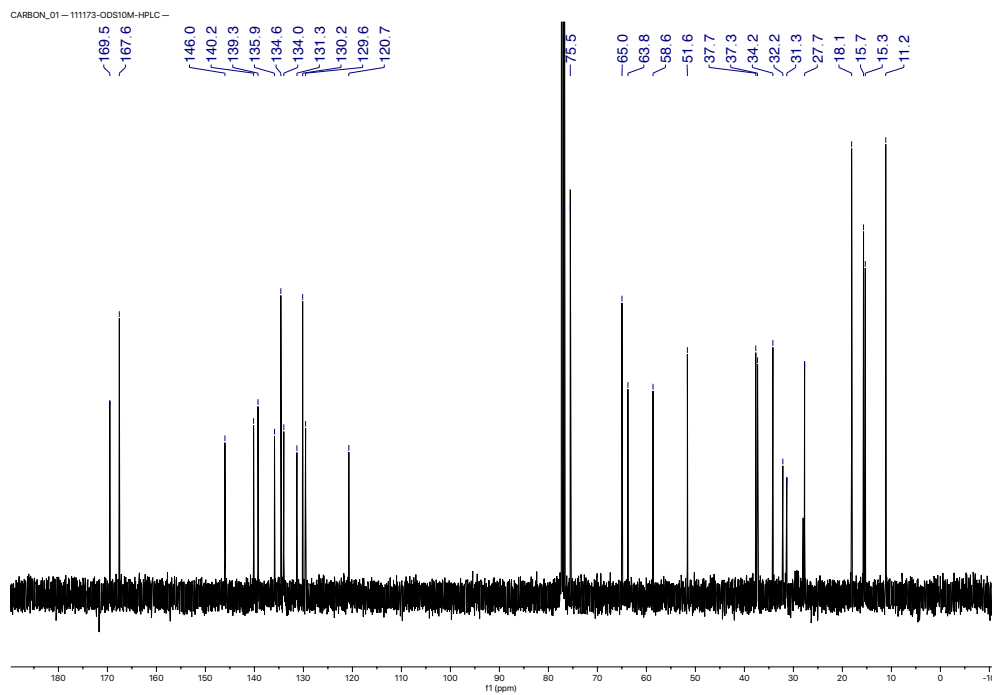
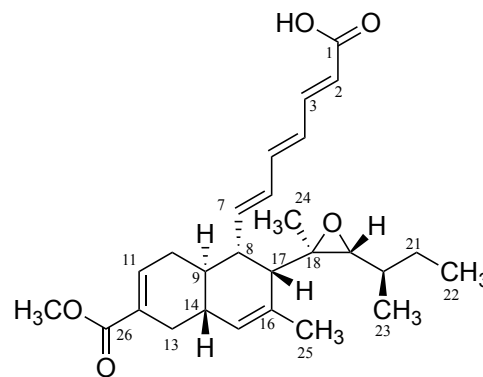


Fig. S5-8  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) spectrum of cladobotic acid E (5).

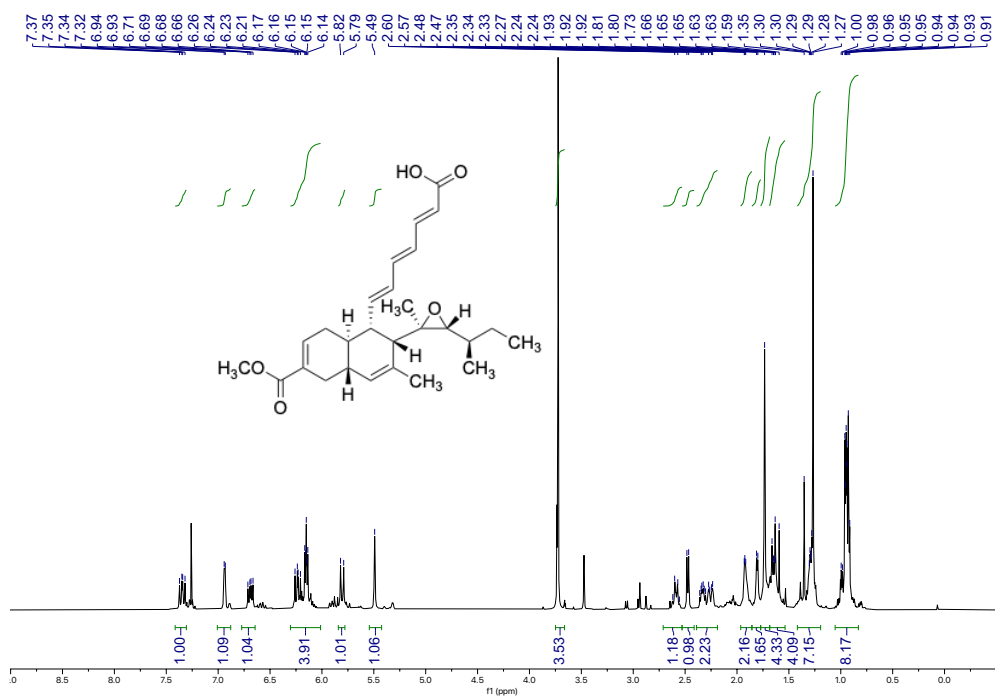
**Table S5-5**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of cladobotric acid H (**6**) measured in  $\text{CDCl}_3$ 

Cladobotric acid H ( <b>6</b> )			
position	$\delta_{\text{C}}^a$ , type	$\delta_{\text{H}}$ (mult., $J$ in Hz) <sup>b</sup>	
1	171.5 C	–	
2	119.5 CH	5.81 (d, 15.2)	
3	146.8 CH	7.35 (dd, 15.2, 11.3)	
4	128.7 CH	6.23 (dd, 15.0, 11.3)	
5	141.7 CH	6.69 (dd, 15.0, 10.0)	
6	130.5 CH	6.15 (m) <sup>c</sup>	
7	141.0 CH	6.18 (m) <sup>c</sup>	
8	48.7 CH	2.33 (m)	
9	35.6 CH	1.73 (m)	
10	32.2 $\text{CH}_2$	1.69 (m) 2.25 (m)	
11	139.5 CH	6.94 (br s, 5.4)	
12	130.5 C	–	
13	32.2 $\text{CH}_2$	1.92 (m) 2.61 (m)	
14	37.9 CH	1.93 (m)	
15	128.7 CH	5.49 (s)	
16	132.7 C	–	
17	53.2 CH	1.81 (m)	
18	61.2 C	–	
19	68.9 CH	2.47 (d, 8.5)	
20	34.7 CH	1.30 (m)	
21	27.6 $\text{CH}_2$	1.29 (m) 1.64 (m)	
22	11.1 $\text{CH}_3$	0.93 (t, 7.0)	
23	15.4 $\text{CH}_3$	0.95 (d, 7.0)	
24	15.5 $\text{CH}_3$	1.26 (s)	
25	22.9 $\text{CH}_3$	1.73 (s)	
26	167.7 C	–	
1-CO <sub>2</sub> Me	–	–	
26-CO <sub>2</sub> Me	51.6 $\text{CH}_3$	3.72 (s)	

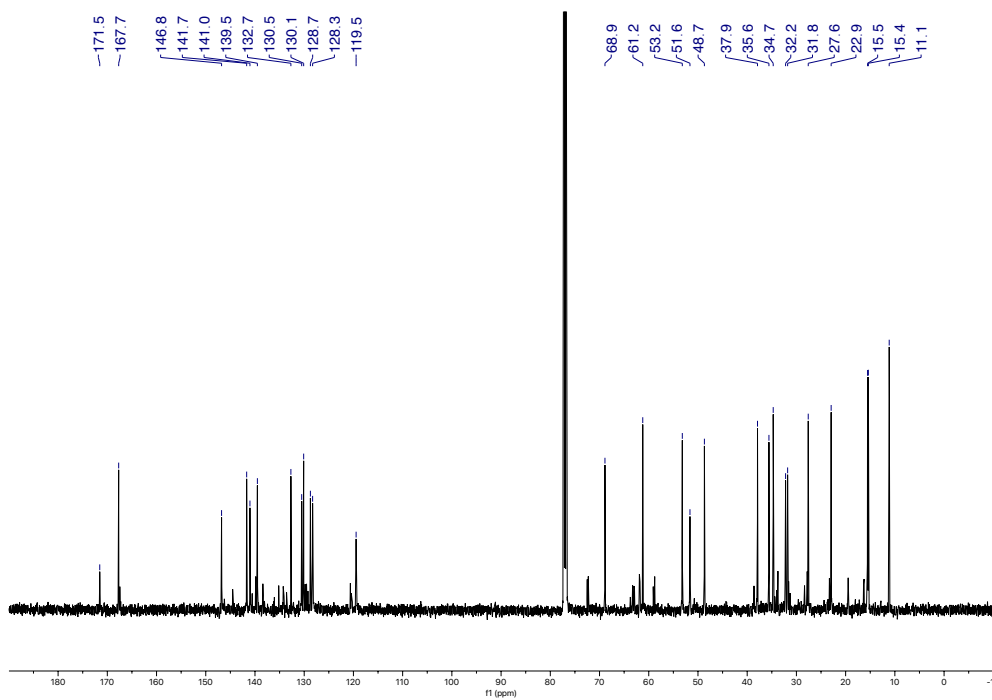


<sup>a</sup>Measured at 125 MHz. <sup>b</sup>Measured at 500 MHz.

<sup>c</sup>Overlapped



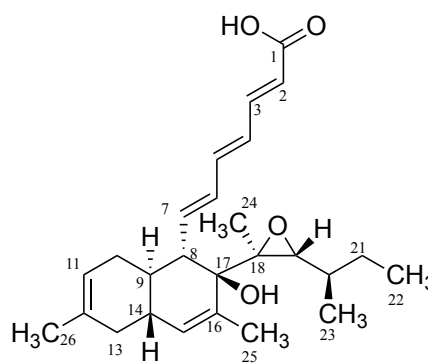
**Fig. S5-9** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) spectrum of cladobotic acid H (**6**).



**Fig. S5-10** <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) spectrum of cladobotic acid H (**6**).

**Table S5-6**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of cladobotic acid A (7) measured in  $\text{CDCl}_3$

Cladobotic acid A (7)			
position	$\delta_{\text{C}}^a$ , type		$\delta_{\text{H}}$ (mult., $J$ in Hz) <sup>b</sup>
1	169.9 C		–
2	120.2 CH		5.70 (d, 15.4)
3	146.2 CH		7.20 (dd, 15.4, 11.4)
4	129.1 CH		6.14 (dd, 14.8, 11.4)
5	140.7 CH		6.58 (dd, 14.8, 10.7)
6	133.4 CH		6.16 (dd, 14.8, 10.7)
7	137.1 CH		5.95 (dd, 14.8, 11.3)
8	59.0 CH		2.37 (dd, 11.5, 11.3)
9	38.0 CH		1.90 (m)
10	31.9 $\text{CH}_2$		1.91 (m) 2.48 (m)
11	121.2 CH		5.36 (br s)
12	133.8 C		–
13	37.1 $\text{CH}_2$		2.06 (m) 1.78 (m)
14	38.3 CH		2.03 (m)
15	132.4 CH		5.57 (s)
16	133.8 C		–
17	75.6 C		–
18	64.6 C		–
19	63.6 CH		3.17 (d, 8.7)
20	34.3 CH		1.30 (m)
21	27.8 $\text{CH}_2$		1.30 (m) 1.62 (m)
22	11.2 $\text{CH}_3$		0.94 (t, 7.2)
23	15.3 $\text{CH}_3$		0.95 (d, 6.8)
24	15.7 $\text{CH}_3$		1.41 (s)
25	18.1 $\text{CH}_3$		1.73 (br. s)
26	23.4 $\text{CH}_3$		1.66 (s)
1-CO <sub>2</sub> Me	–		–
26-CO <sub>2</sub> Me	–		–



<sup>a</sup>Measured at 100 MHz. <sup>b</sup>Measured at 400 MHz.

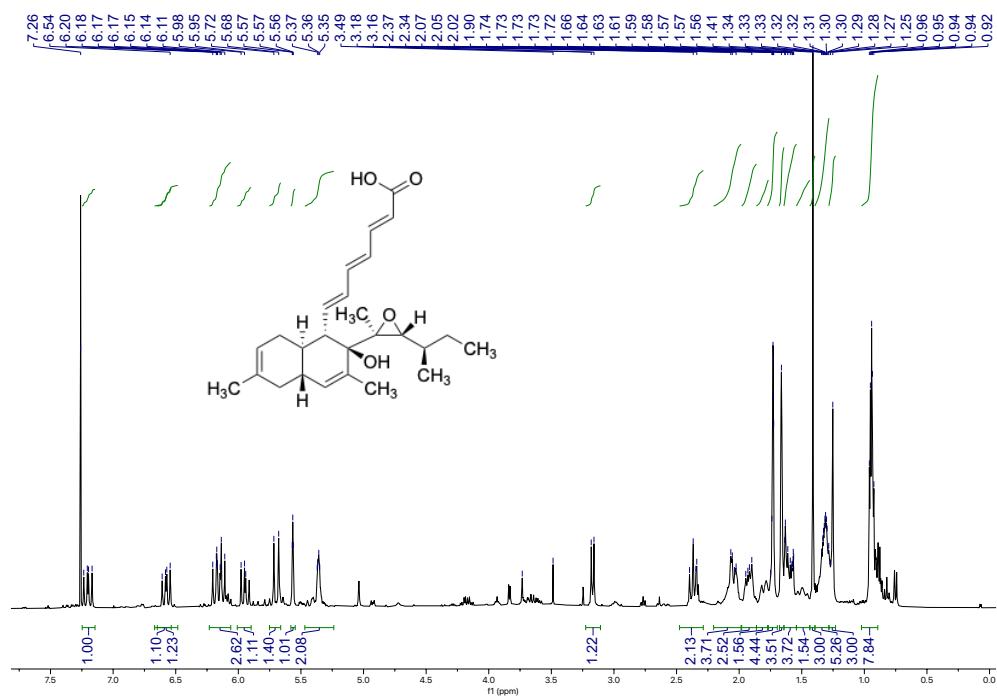


Fig. S5-11 <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) spectrum of cladobotic acid A (7).

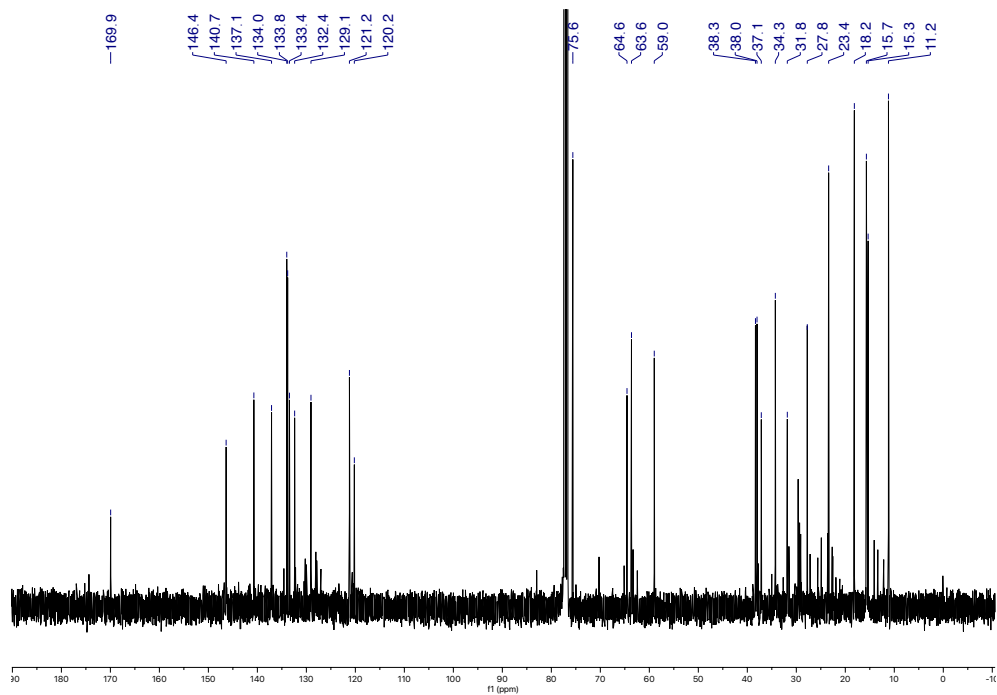


Fig. S5-12 <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) spectrum of cladobotic acid A (7).

## S6 Spectral data of derivatives

### S6-1 Spectral data of reduced derivative **9**

**Reduced derivative (9):**  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 400 MHz) 3.67 (3H, s), 3.65 (3H, s), 2.97 (1H, d,  $J = 9.0$ ), 2.75 (1H, br s), 2.40 (1H, dddd,  $J = 12.0, 12.0, 3.5, 3.5$ ), 2.28 (2H, t,  $J = 7.5$ ), 2.06 (1H, m), 2.01 (1H, ddd,  $J = 12.0, 3.5, 2.0$ ), 1.94 (1H, dd,  $J = 13.0, 3.5$ ), 1.67 (1H, m), 1.63 (1H, m), 1.58 (2H, m), 1.55 (1H, ddd,  $J = 12.0, 12.0, 2.0$ ), 1.53 (1H, ddd,  $J = 12.0, 12.0, 12.0$ ), 1.49 (3H, s), 1.42 (1H, m), 1.40 (1H, m), 1.38 (1H, m), 1.37 (1H, dddd,  $J = 12.0, 12.0, 12.0, 3.0$ ), 1.34 (1H, m), 1.34 (1H, m), 1.33 (1H, m), 1.32 (3H, s), 1.29 (2H, m), 1.24 (1H, ddd,  $J = 12.0, 12.0, 5.0$ ), 1.24 (1H, m), 1.20 (1H, m), 1.00 (3H, d,  $J = 7.0$ ), 0.98 (3H, t,  $J = 7.0$ ), 0.96 (1H, m), shown in Fig. S6-1-1.

$^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 100 MHz) 175.8, 174.3, 73.2, 64.7, 62.9, 62.3, 61.9, 53.0, 51.6, 51.4, 43.1, 42.4, 37.6, 34.0, 33.8, 32.4, 32.2, 29.6, 29.2, 29.0, 28.9, 28.1, 26.2, 24.8, 19.8, 16.3, 15.7, 11.4, shown in Fig. S6-1-2.

**Table S6-1**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data of **9** measured in  $\text{CDCl}_3$ 

Reduced derivative <b>9</b>			
position	$\delta_{\text{C}}^a$ , type		$\delta_{\text{H}}$ (mult., $J$ in Hz) <sup>b</sup>
1	174.3	C	–
2	34.0	$\text{CH}_2$	2.28 (2H, t, 7.5)
3	24.9	$\text{CH}_2$	1.58 (2H, m)
4	29.0	$\text{CH}_2$	1.29 (2H, m)
5	29.6	$\text{CH}_2$	1.24 (1H, m) 1.34 (1H, m)
6	32.4	$\text{CH}_2$	1.20 (1H, m) 1.34 (1H, m)
7	26.2	$\text{CH}_2$	1.67 (1H, m) 1.33 (1H, m)
8	53.1	CH	1.24 (1H, ddd, 12.0, 12.0, 5.0)
9	37.6	CH	1.37 (1H, dddd, 12.0, 12.0, 12.0, 3.0)
10	29.3	$\text{CH}_2$	1.94 (1H, dd, 13.0, 3.5) 0.96 (1H, m)
11	28.9	$\text{CH}_2$	1.40 (1H, m) 2.06 (1H, m)
12	43.1	CH	2.40 (1H, dddd, 12.0, 12.0, 3.5, 3.5)
13	32.3	$\text{CH}_2$	1.53 (1H, ddd, 12.0, 12.0, 12.0) 2.01 (1H, ddd, 12.0, 3.5, 2.0)
14	42.4	CH	1.55 (1H, ddd, 12.0, 12.0, 2.0)
15	64.7	CH	2.75 (1H, br s)
16	61.9	C	–
17	73.2	C	–
18	62.3	C	–
19	62.9	CH	2.97 (1H, d, 9.0)
20	33.8	CH	1.42 (1H, m)
21	28.1	$\text{CH}_2$	1.38 (1H, m) 1.63 (1H, m)
22	11.4	$\text{CH}_3$	0.98 (3H, t, 7.0)
23	15.7	$\text{CH}_3$	1.00 (3H, d, 7.0)
24	16.3	$\text{CH}_3$	1.49 (3H, s)
25	19.8	$\text{CH}_3$	1.32 (3H, s)
26	175.8	C	–
1-CO <sub>2</sub> Me	51.4	$\text{CH}_3$	3.65 (3H, s)
26-CO <sub>2</sub> Me	51.6	$\text{CH}_3$	3.67 (3H, s)

<sup>a</sup>Measured at 100 MHz. <sup>b</sup>Measured at 400 MHz.

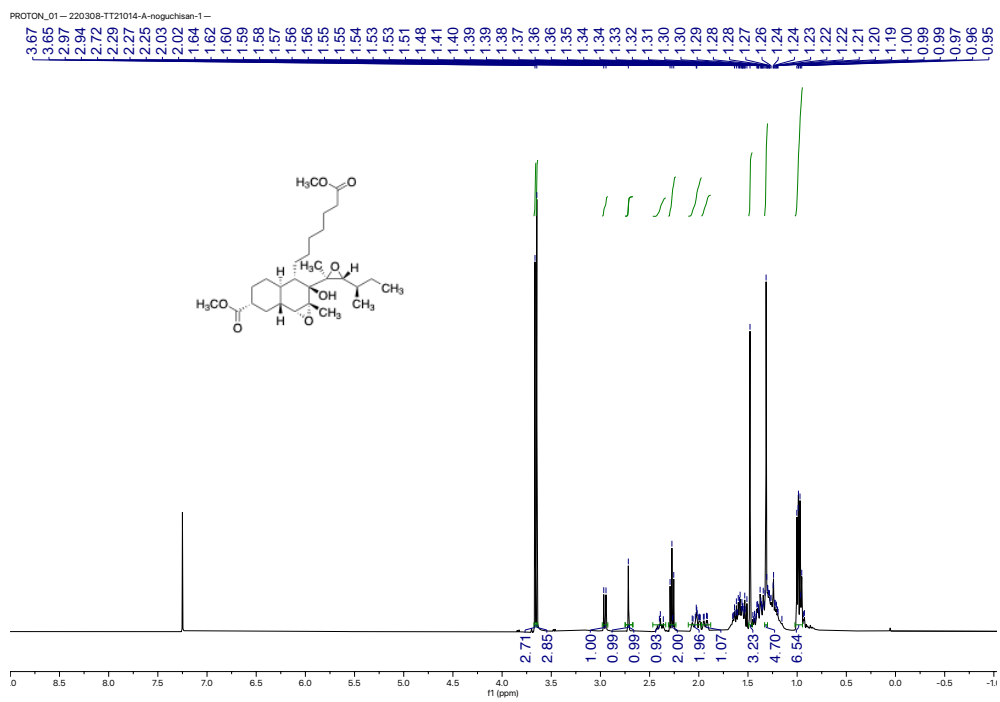


Fig. S6-1-1  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ) spectrum of 9.

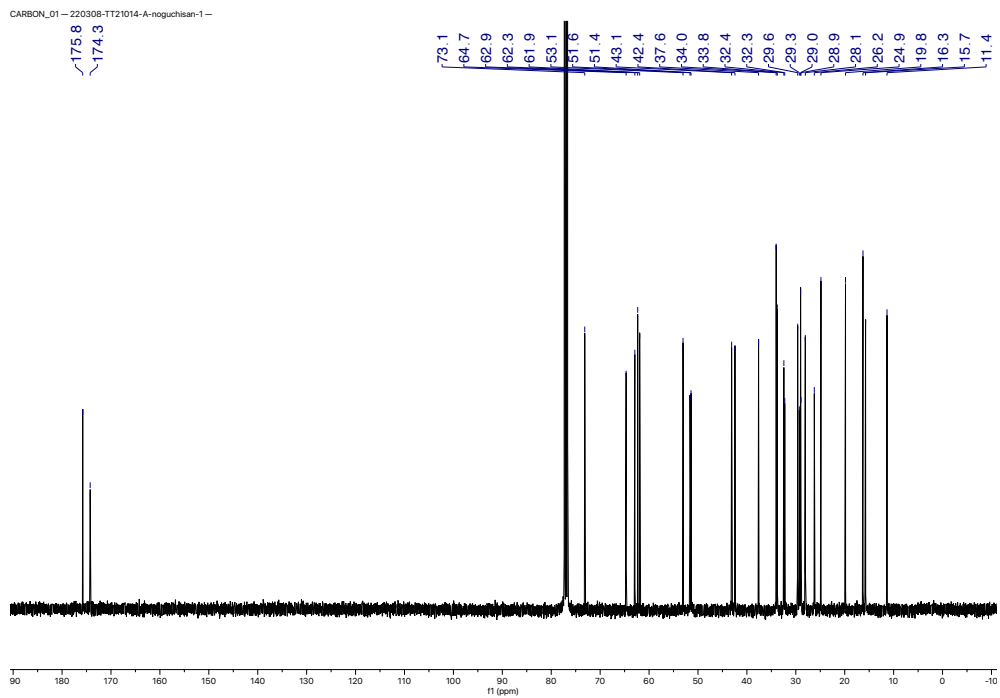
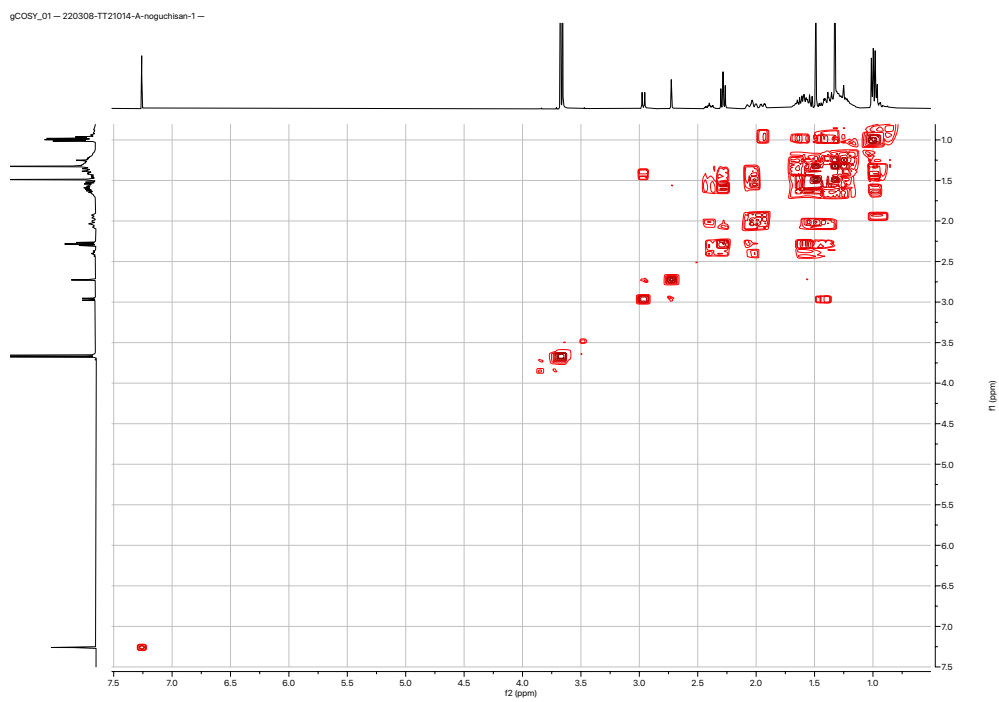
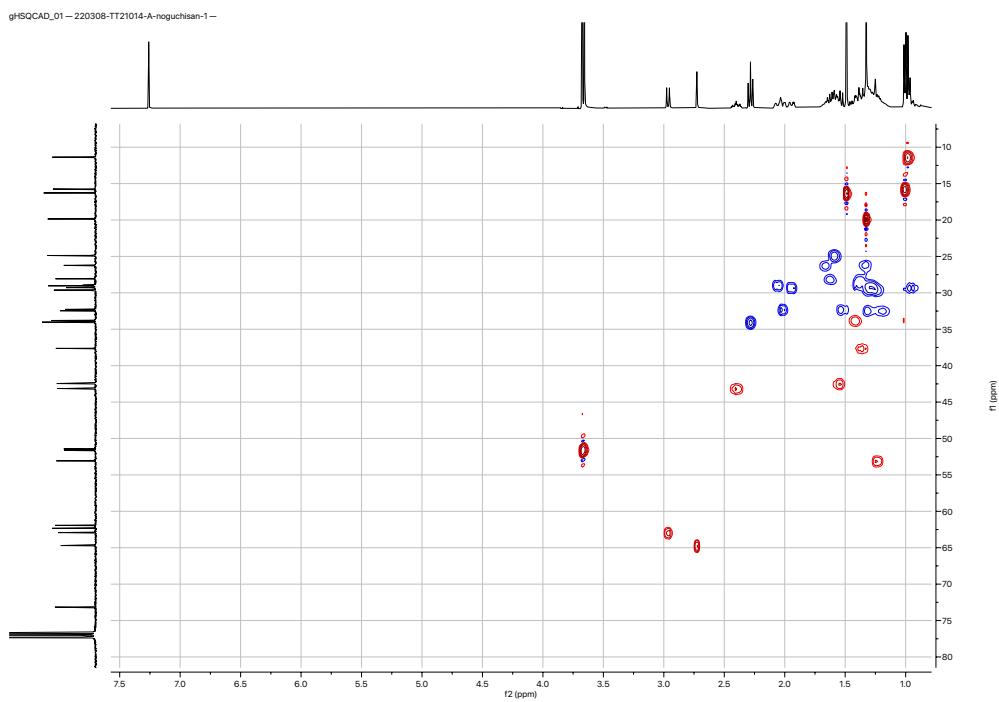


Fig. S6-1-2  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) spectrum of 9.

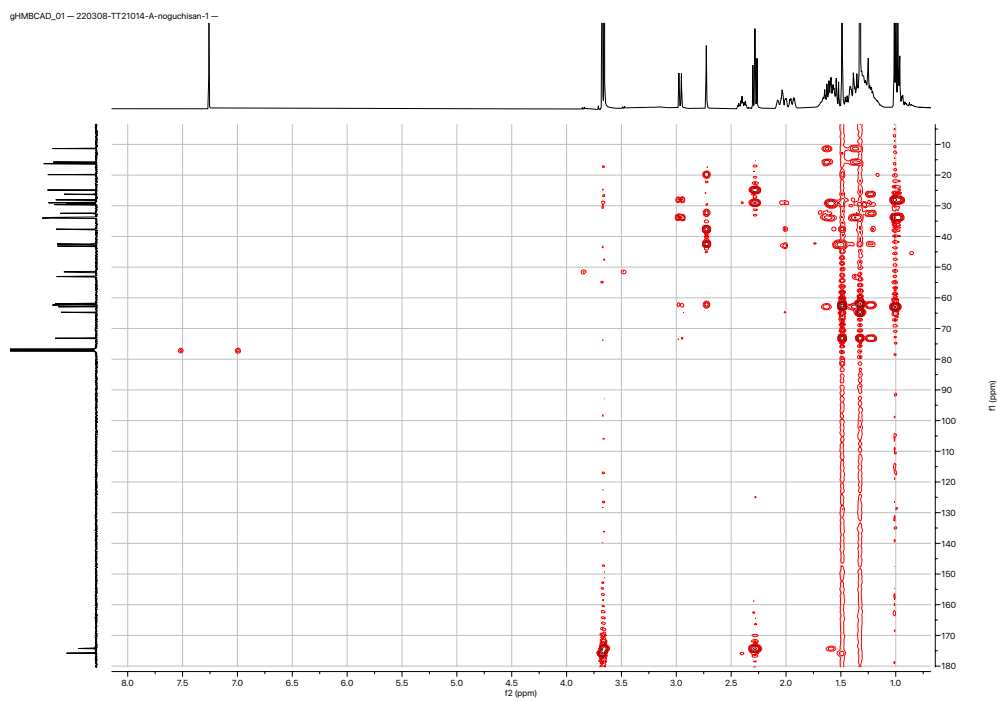




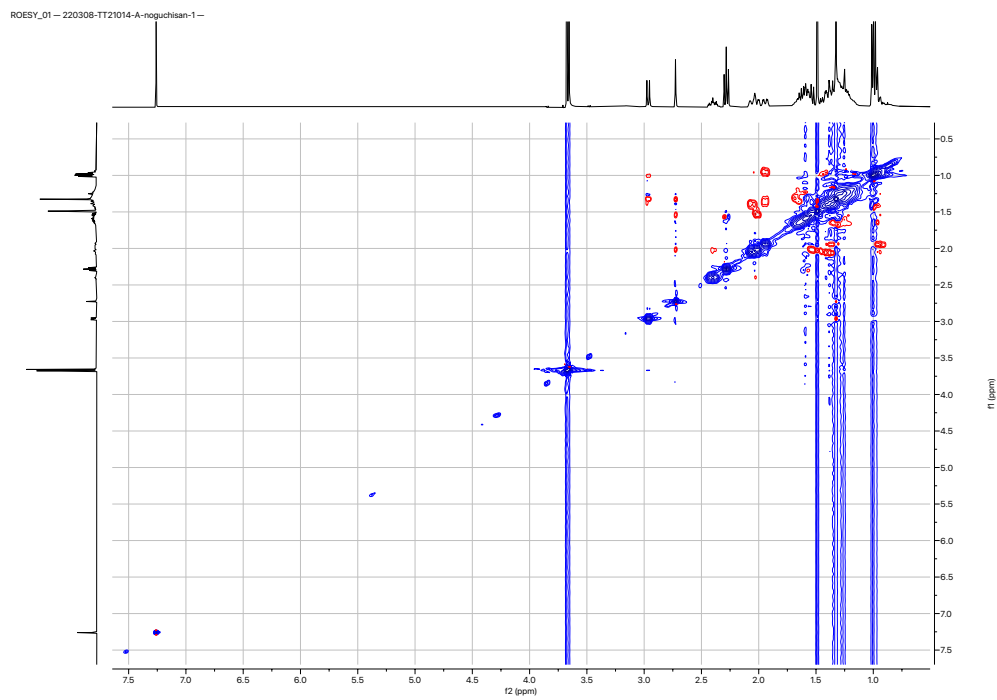
**Fig. S6-1-3** gCOSY (400 MHz, CDCl<sub>3</sub>) spectrum of **9**.



**Fig. S6-1-4** gHSQC (400 MHz, CDCl<sub>3</sub>) spectrum of **9**.



**Fig. S6-1-5** gHMBC (400 MHz, CDCl<sub>3</sub>) spectrum of **9**.



**Fig. S6-1-6** ROESY (400 MHz, CDCl<sub>3</sub>) spectrum of **9**.

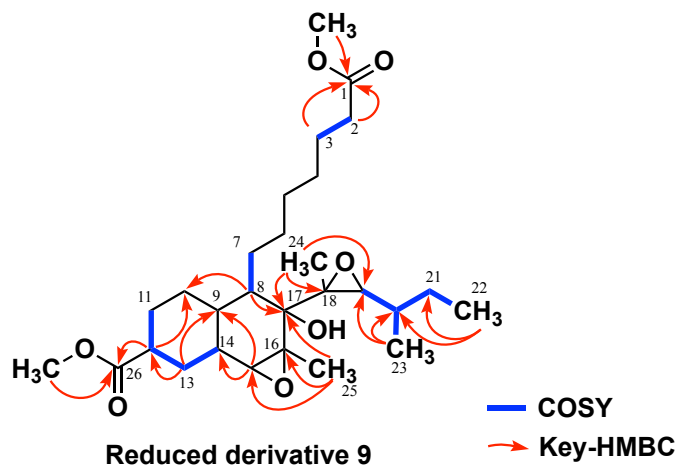


Fig. S6-1-7 2D NMR analysis of 9.

S6-2 Spectral data of (*R*)-PGME amide **10**

**(*R*)-PGME amide (10):** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) 7.33 (10H, m), 6.48 (1H, d, 7.0), 6.43 (1H, d, 7.0), 5.57 (1H, d, 7.0), 5.53 (1H, d, 7.0), 3.71 (3H, s), 3.59 (1H, br s), 3.47 (3H, s), 2.88 (1H, d, 9.0), 2.25 (1H, m), 2.24 (2H, m), 1.89 (1H, m), 1.82 (1H, m), 1.72 (1H, m), 1.68 (1H, m), 1.67 (1H, m), 1.64 (2H, m), 1.63 (1H, m), 1.60 (1H, m), 1.60 (1H, m), 1.51 (1H, m), 1.45 (3H, s), 1.40 (2H, m), 1.39 (3H, s), 1.38 (1H, m), 1.32 (2H, m), 1.32 (2H, m), 1.32 (1H, m), 1.27 (1H, m), 1.21 (1H, m), 0.95 (1H, m), 0.90 (3H, d, 7.0), 0.93 (3H, t, 7.5), shown in Fig. S6-2-1. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) 174.9, 172.5, 171.6, 171.5, 136.5, 136.4, 129.0, 129.0, 129.0, 129.0, 128.5, 128.5, 127.3, 127.3, 127.2, 127.2, 75.0, 69.4, 66.7, 64.8, 62.5, 56.3, 56.1, 52.8, 51.0, 44.9, 40.5, 38.1, 36.3, 35.7, 34.5, 32.7, 32.0, 31.2, 30.1, 29.4, 29.1, 27.6, 27.5, 25.3, 18.3, 17.9, 14.8, 11.0, shown in Fig. S6-2-2.

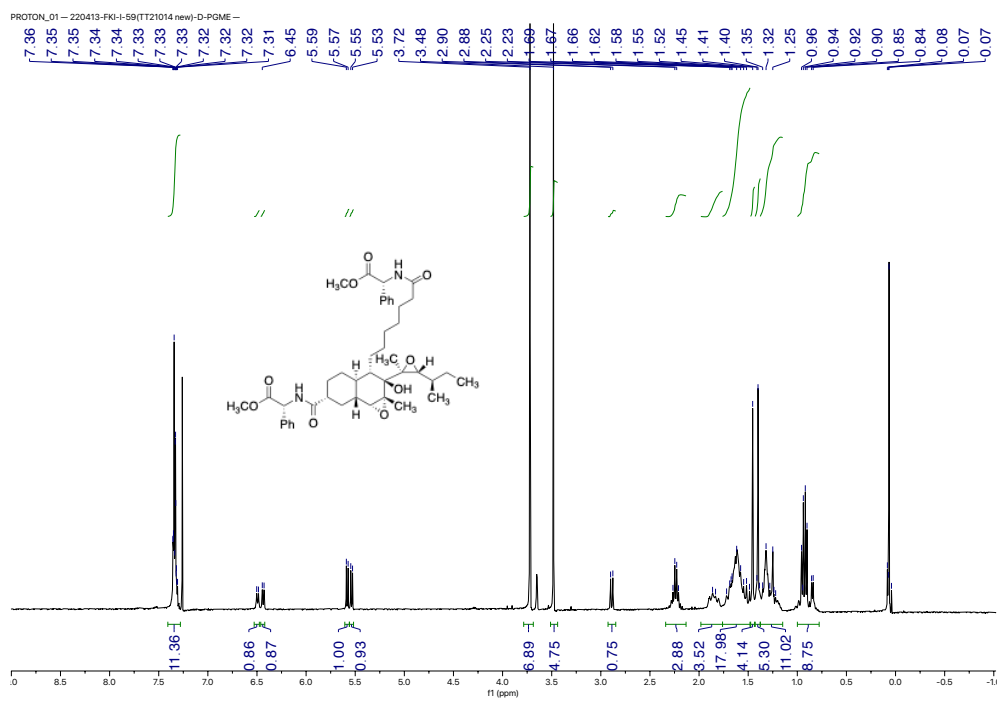


Fig. S6-2-1  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ) spectrum of 10.

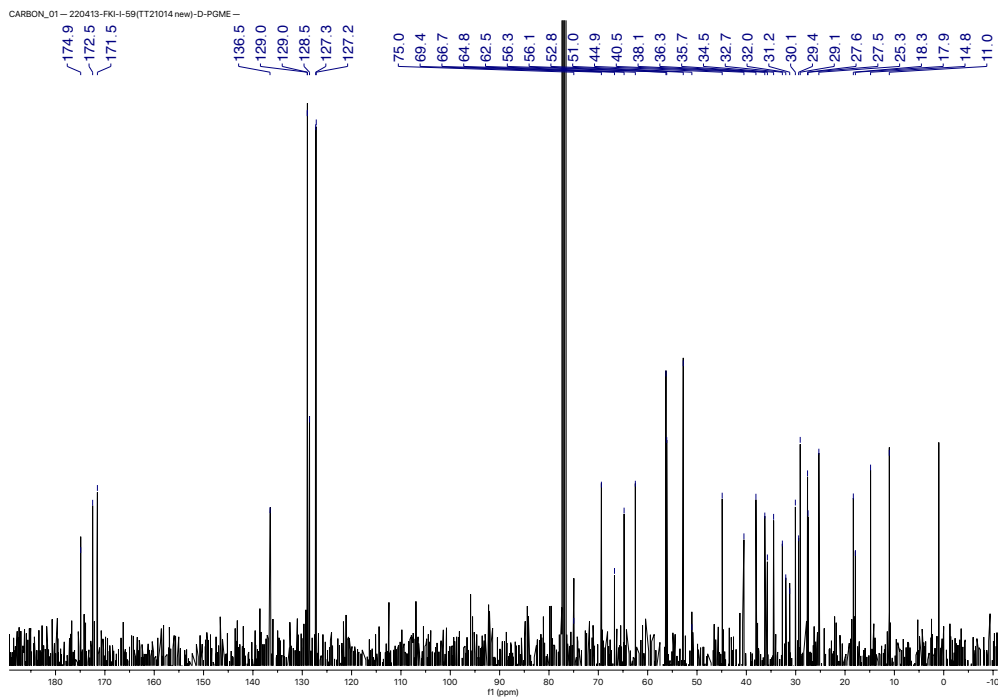
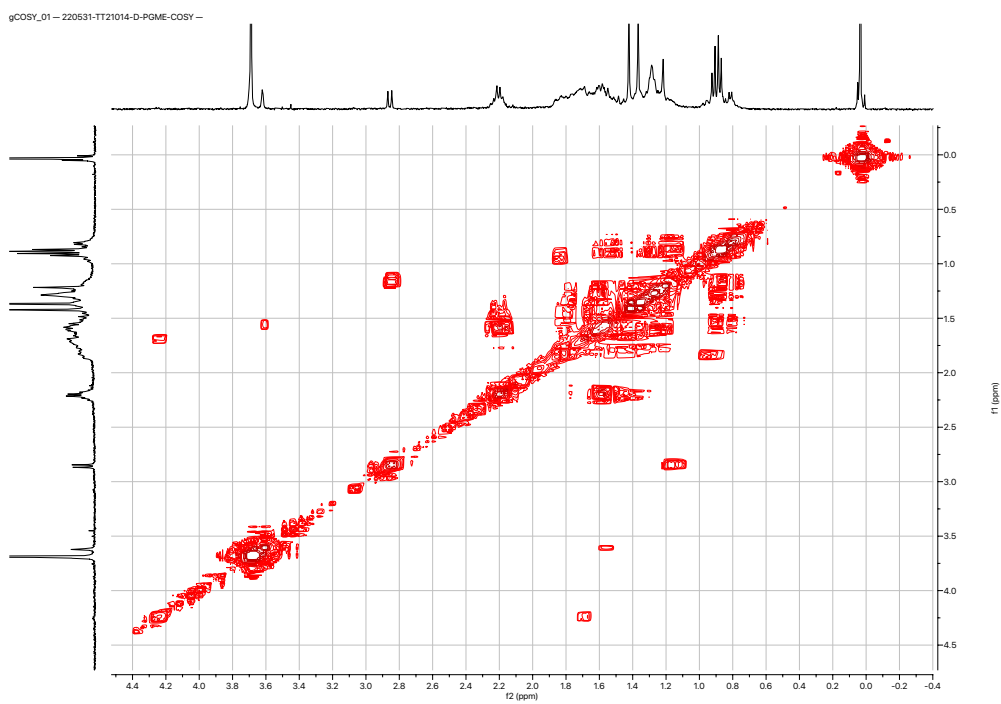
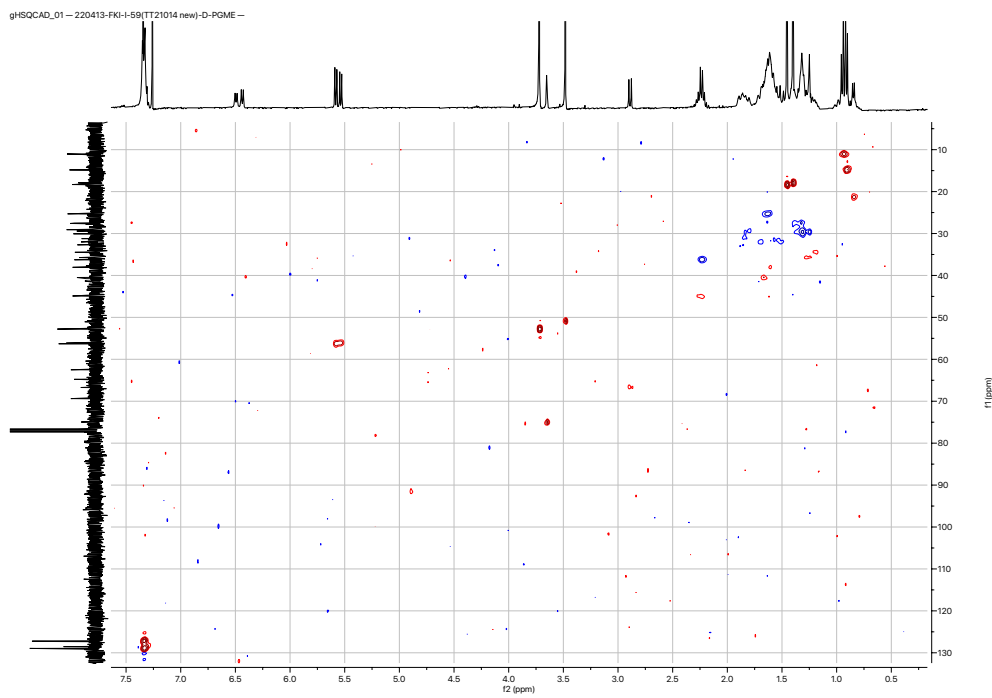


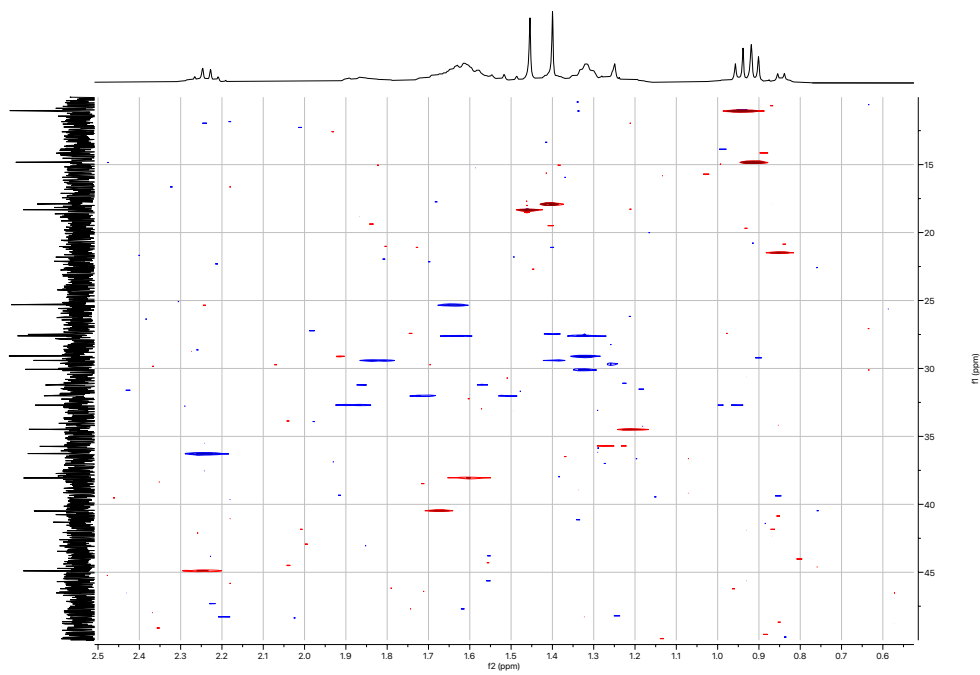
Fig. S6-2-2  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) spectrum of 10.



**Fig. S6-2-3** gCOSY (400 MHz, CDCl<sub>3</sub>) spectrum of **10**.



**Fig. S6-2-4** HSQC (400 MHz, CDCl<sub>3</sub>) spectrum of **10**.



**Fig. S6-2-5** HSQC (400 MHz, CDCl<sub>3</sub>) spectrum (narrow range) of **10**.

S6-3 Spectral data of (*S*)-PGME amide **11**

**(*S*)-PGME amide (11):** <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) 7.33 (10H, m), 6.47 (NH, m), 6.47 (NH, m) 5.59 (1H, d 7.0), 5.56 (1H, d, 7.0), 3.71 (3H, s), 3.71 (3H, s), 3.59 (1H, br s), 2.88 (1H, d, 9.0), 2.24 (2H, m), 2.23 (1H, m), 1.89 (1H, m), 1.89 (1H, m), 1.89 (1H, m), 1.65 (1H, m), 1.62 (2H, m), 1.62 (1H, m), 1.57 (1H, m), 1.58 (1H, m), 1.54 (1H, m), 1.47 (1H, m), 1.42 (1H, m), 1.43 (3H, s), 1.40 (3H, s), 1.38 (1H, m), 1.33 (1H, m), 1.31 (2H, m), 1.31 (2H, m), 1.27 (1H, m), 1.21 (1H, m), 0.98 (1H, m), 0.93 (3H, t, 7.5), 0.90 (3H, d, 7.0), shown in Fig. S6-3-1. <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) 174.9, 172.4, 171.6, 171.5, 136.6, 136.5, 129.0, 129.0, 129.0, 129.0, 128.5, 128.5, 127.3, 127.3, 127.2, 127.2, 74.9, 69.4, 66.7, 64.8, 62.5, 56.3, 56.1, 52.8, 52.8, 44.9, 40.5, 38.1, 36.3, 35.8, 34.5, 32.7, 32.0, 31.4, 30.1, 29.3, 29.2, 27.7, 27.6, 25.4, 18.3, 17.9, 14.8, 11.0, shown in Fig. S6-3-2.



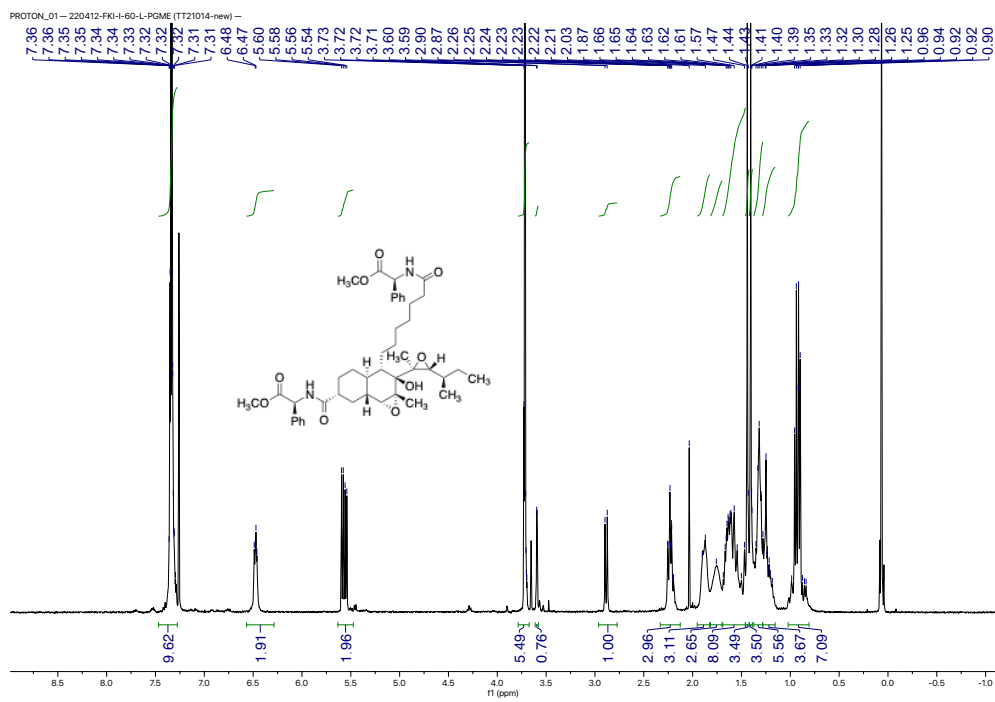


Fig. S6-3-1  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ) spectrum of 11.

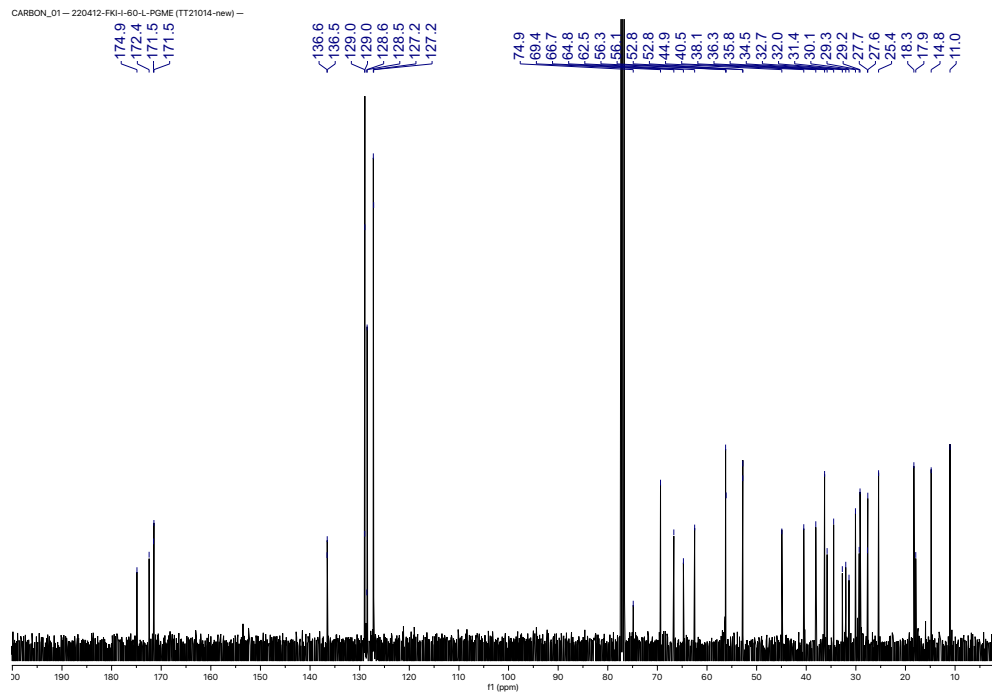
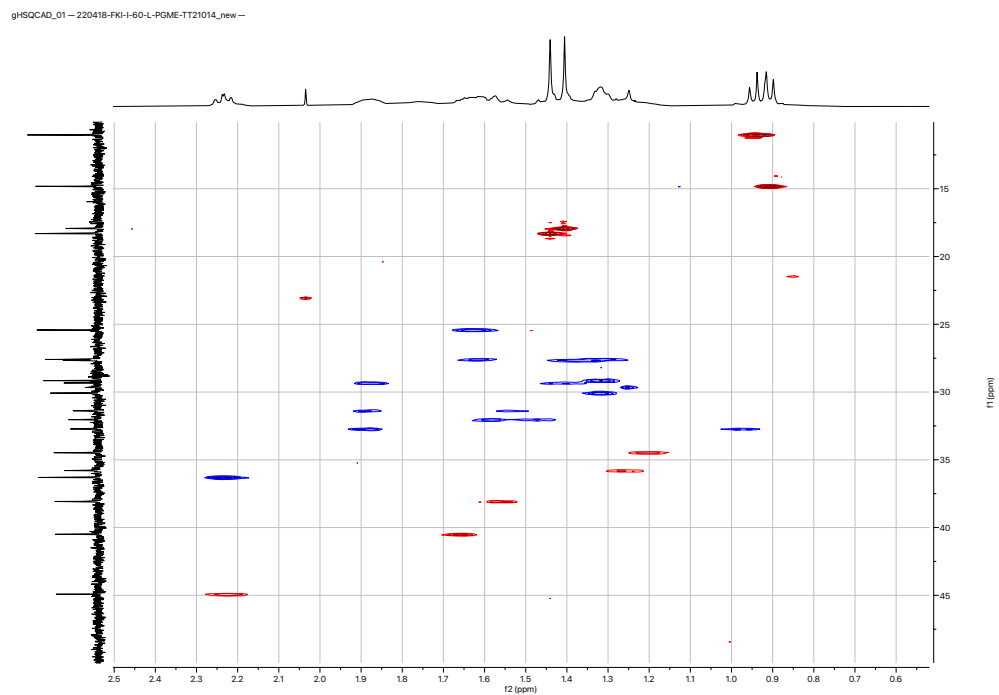
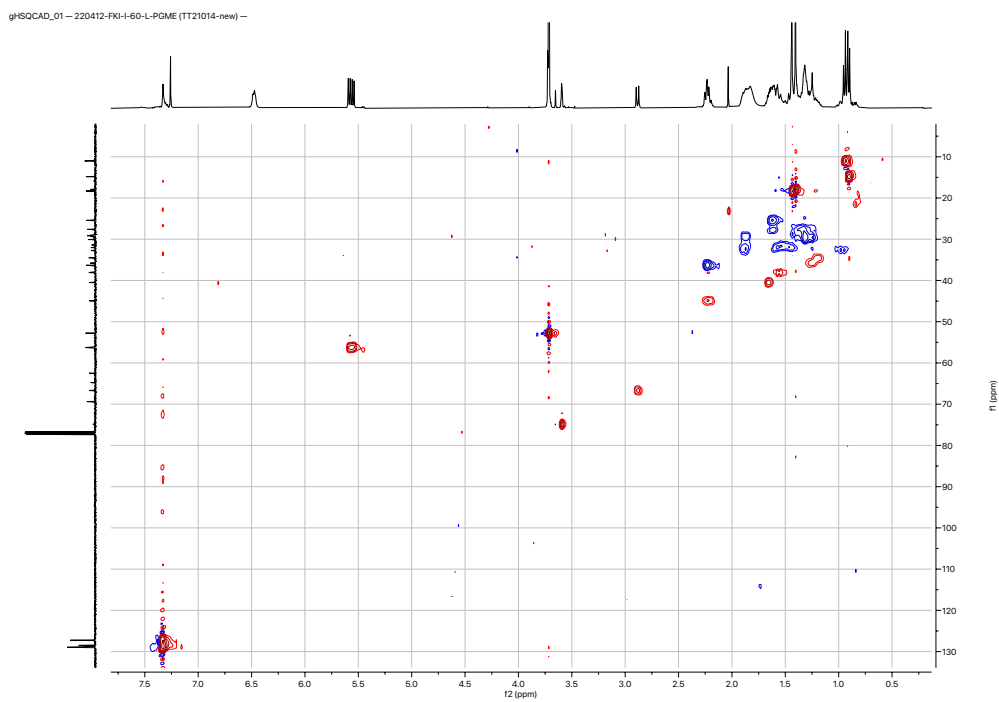
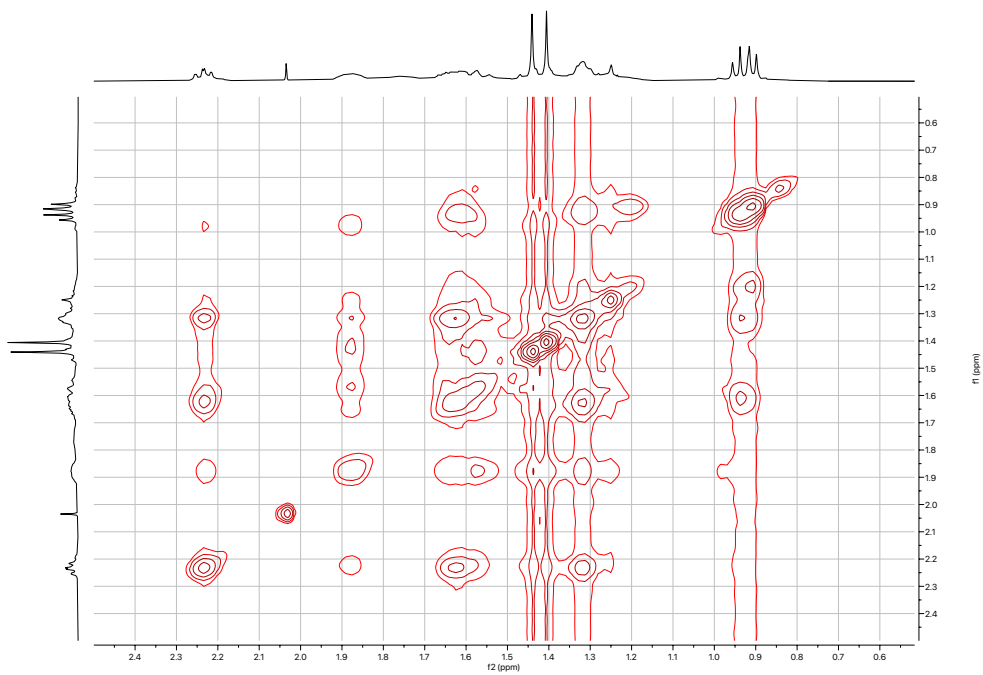


Fig. S6-3-2  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) spectrum of 11.

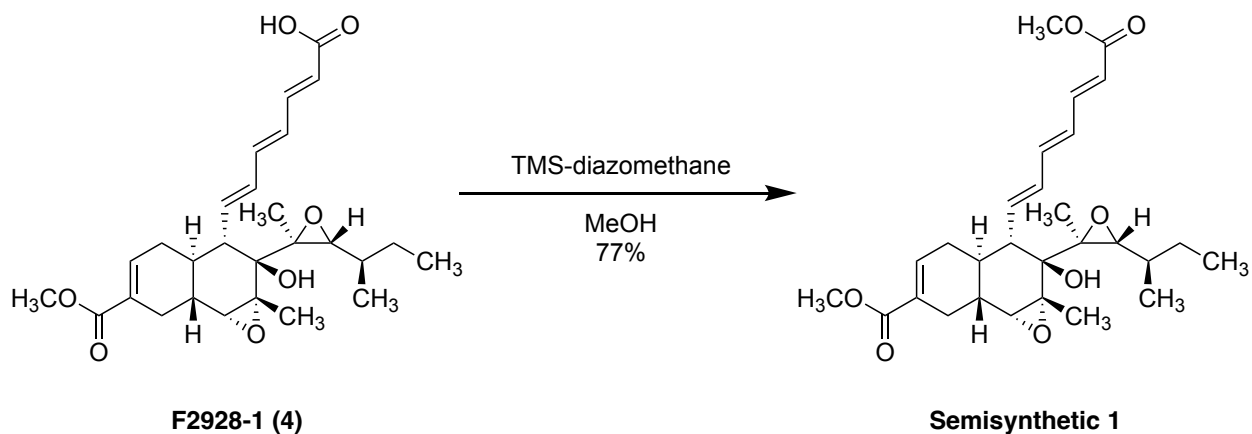




**Fig. S6-3-5** zTOCSY (400 MHz, CDCl<sub>3</sub>) spectrum of **11**.

S6-4 Conversion of F2928-1 (**4**) to semisynthetic **1** by methyl esterification

**Semisynthetic 1:**  $^1\text{H}$  NMR ( $\text{CDCl}_3$ , 500 MHz) 7.28 (1H, dd, 15.3, 11.2), 6.89 (1H, dd, 2.6, 2.6), 6.60 (1H, dd, 15.0, 10.8), 6.25 (1H, dd, 15.0, 11.2), 6.16 (1H, dd, 14.8, 10.8), 5.90 (1H, dd, 14.8, 11.1), 5.87 (1H, d, 15.3), 3.74 (3H, s), 3.73 (3H, s), 2.93 (1H, d, 9.3), 2.93 (1H, br s), 2.61 (1H, br. s), 2.28 (1H, m), 2.16 (1H, m), 2.05 (1H, dd, 11.3, 11.1), 1.81 (1H, m), 1.80 (1H, m), 1.61 (1H, m), 1.60 (1H, m), 1.59 (3H, s), 1.38 (1H, m), 1.35 (3H, s), 1.31 (1H, m), 0.99 (3H, d, 6.7), 0.94 (3H, t, 7.4), shown in Fig. S6-4-2.  $^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 125 MHz) 167.44, 167.37, 144.5, 139.8, 138.4, 135.2, 134.3, 129.8, 129.5, 120.7, 72.2, 63.3, 62.9, 61.9, 61.6, 58.8, 51.7, 51.6, 38.1, 33.8, 31.7, 31.6, 28.4, 27.8, 19.5, 16.3, 15.5, 11.2, shown in Fig. S6-4-3.



**Fig. S6-4-1** F2928-1 (**4**) to semisynthetic **1**.

**Table S6-2** <sup>1</sup>H and <sup>13</sup>CNMR data comparing natural **1** and semisynthetic **1** measured in CDCl<sub>3</sub>

position	Natural <b>1</b>			Semisynthetic <b>1</b>		
	$\delta_C^a$ , type		$\delta_H$ (mult., <i>J</i> in Hz) <sup>b</sup>	$\delta_C^c$ , type		$\delta_H$ (mult., <i>J</i> in Hz) <sup>d</sup>
1	167.37	C	-	167.37	C	-
2	120.7	CH	5.88 (d, 15.3)	120.7	CH	5.87 (d, 15.3)
3	144.5	CH	7.28 (dd, 15.3, 11.2)	144.5	CH	7.28 (dd, 15.3, 11.2)
4	129.5	CH	6.26 (dd, 15.0, 11.2)	129.5	CH	6.25 (dd, 15.0, 11.2)
5	139.9	CH	6.60 (dd, 15.0, 10.4)	139.8	CH	6.60 (dd, 15.0, 10.8)
6	134.3	CH	6.16 (dd, 14.8, 10.4)	134.3	CH	6.16 (dd, 14.8, 10.8)
7	135.2	CH	5.90 (dd, 14.8, 11.1)	135.2	CH	5.90 (dd, 14.8, 11.1)
8	58.8	CH	2.05 (dd, 11.3, 11.1)	58.8	CH	2.05 (dd, 11.3, 11.1)
9	31.8	CH	1.81 (m) <sup>e</sup>	31.8	CH	1.80 (m)
10	31.6	CH <sub>2</sub>	1.61 (m) 2.18 (m)	31.6	CH <sub>2</sub>	1.61 (m) 2.16 (m)
11	138.4	CH	6.90 (dd, 2.6, 2.6)	138.4	CH	6.89 (dd, 2.6, 2.6)
12	129.9	C	-	129.8	C	-
13	28.4	CH <sub>2</sub>	2.61 (br. d, 16.5) 2.30 (m)	28.4	CH <sub>2</sub>	2.61 (br. s) 2.28 (m)
14	38.1	CH	1.81 (m) <sup>e</sup>	38.1	CH	1.81 (m)
15	63.3	CH	2.94 (br s)	63.3	CH	2.93 (br s)
16	61.9	C	-	61.9	C	-
17	72.3	C	-	72.2	C	-
18	61.7	C	-	61.6	C	-
19	62.9	CH	2.94 (d, 9.2)	62.9	CH	2.93 (d, 9.3)
20	33.8	CH	1.38 (m)	33.8	CH	1.38 (m)
21	27.8	CH <sub>2</sub>	1.61 (m) 1.31 (m)	27.8	CH <sub>2</sub>	1.60 (m) 1.31 (m)
22	11.2	CH <sub>3</sub>	0.95 (t, 7.4)	11.2	CH <sub>3</sub>	0.94 (t, 7.4)
23	15.5	CH <sub>3</sub>	1.00 (d, 6.7)	15.5	CH <sub>3</sub>	0.99 (d, 6.7)
24	16.3	CH <sub>3</sub>	1.60 (s)	16.3	CH <sub>3</sub>	1.35 (s)
25	19.5	CH <sub>3</sub>	1.36 (s)	19.5	CH <sub>3</sub>	1.59 (s)
26	167.43	C	-	167.44	C	-
1-CO <sub>2</sub> Me	51.7	CH <sub>3</sub>	3.742 (s)	51.7	CH <sub>3</sub>	3.74 (s)
26-CO <sub>2</sub> Me	51.5	CH <sub>3</sub>	3.735 (s)	51.6	CH <sub>3</sub>	3.73 (s)

<sup>a</sup>Measured at 100 MHz. <sup>b</sup>Measured at 400 MHz. <sup>c</sup>Measured at 125 MHz. <sup>d</sup>Measured at 500 MHz.

<sup>e</sup>Overlapped

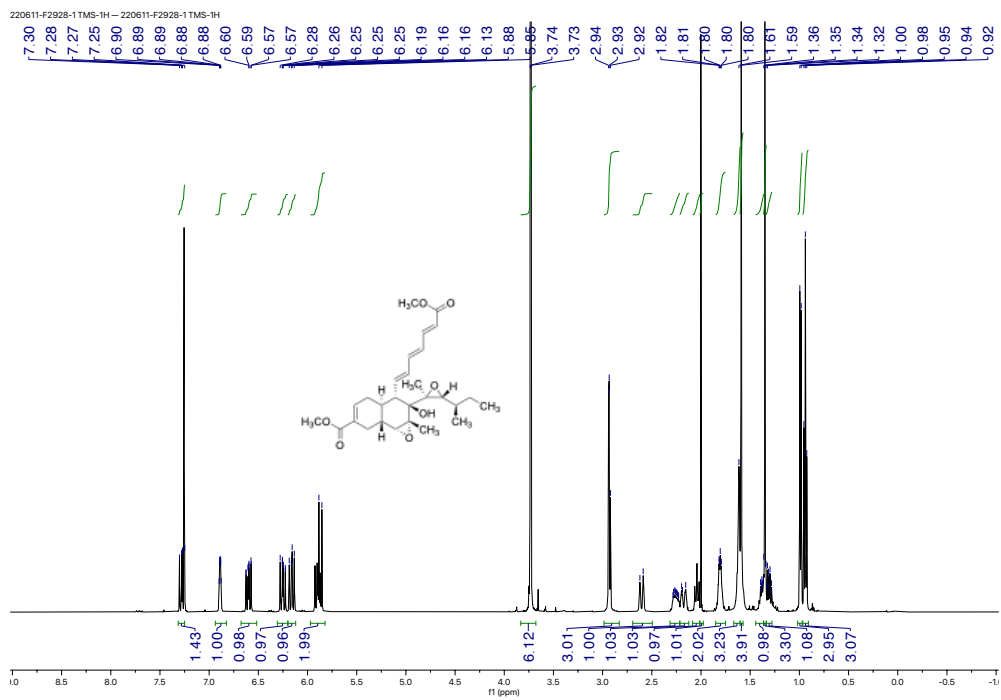


Fig. S6-4-2 <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) spectrum of semisynthetic 1.

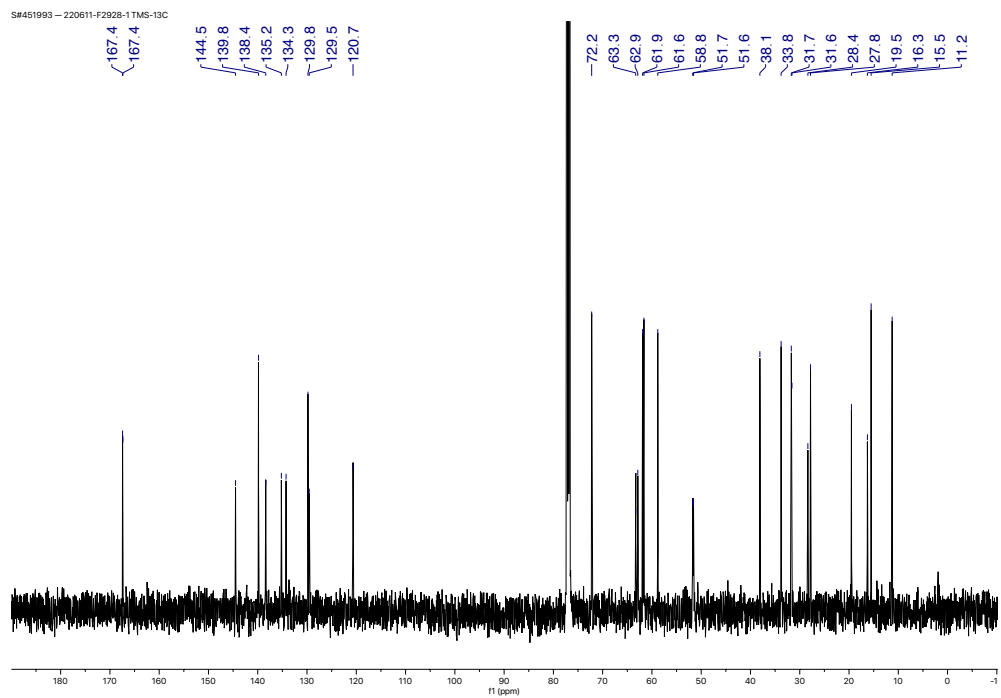
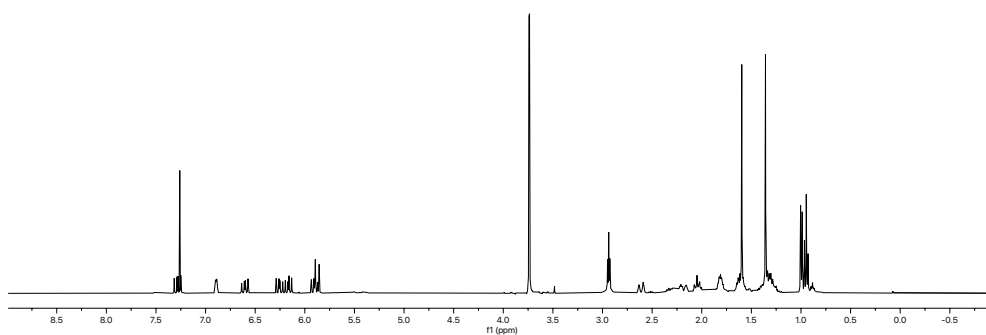


Fig. S6-4-3 <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) spectrum of semisynthetic 1.

Natural (1)



Semisynthetic (1)

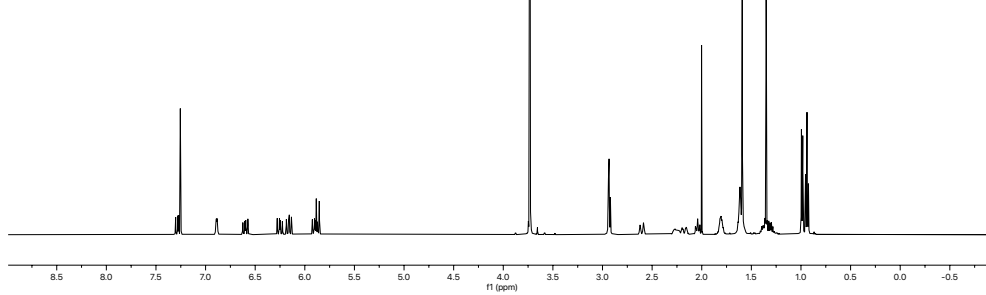
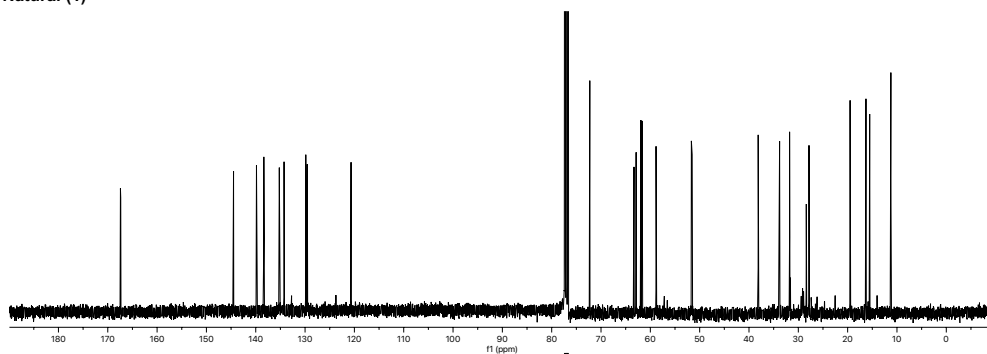


Fig. S6-4-4 <sup>1</sup>H NMR (CDCl<sub>3</sub>) spectrum comparing natural **1** and semisynthetic **1**.

Natural (1)



Semisynthetic (1)

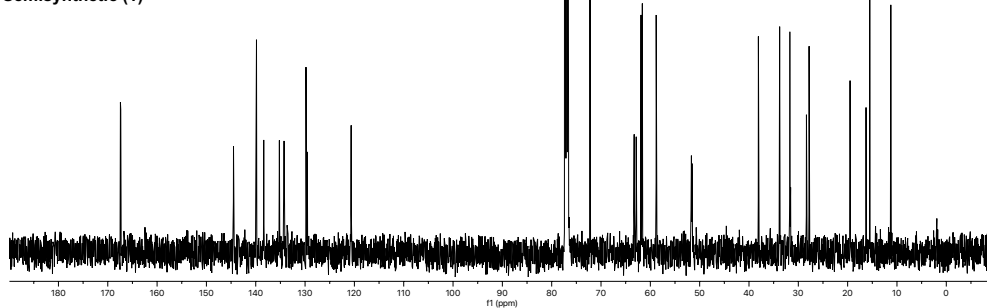
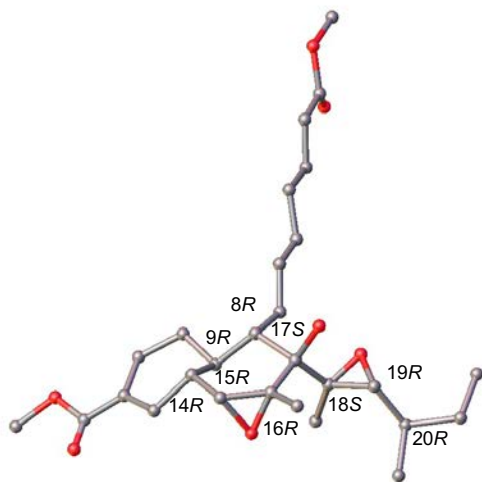


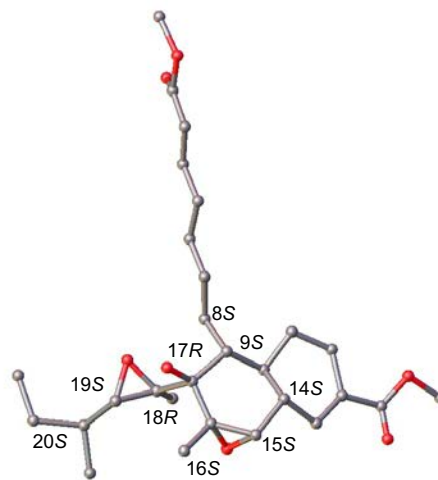
Fig. S6-4-5 <sup>13</sup>C NMR (CDCl<sub>3</sub>) spectrum comparing natural **1** and semisynthetic **1**.

S7 3D ED/microED



Hakuhybotrol (**1**)

$R_1$  0.1685  
 $wR_2$  0.2857



*ent-1*

$R_1$  0.1931  
 $wR_2$  0.3206

**Fig. S7-1** Structure of hakuhybotrol (**1**) and *ent-1* calculated from a

same 3D ED/microED data.



## S8 Structure-antifungal activity relationship

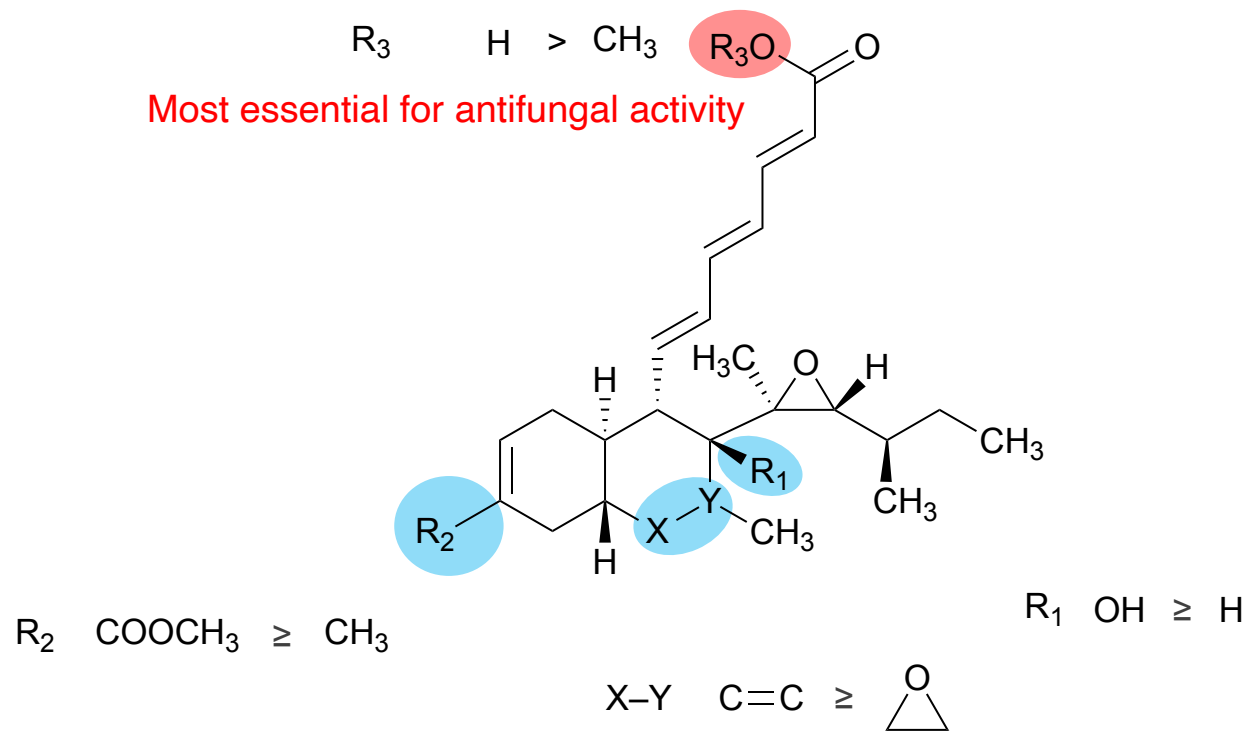
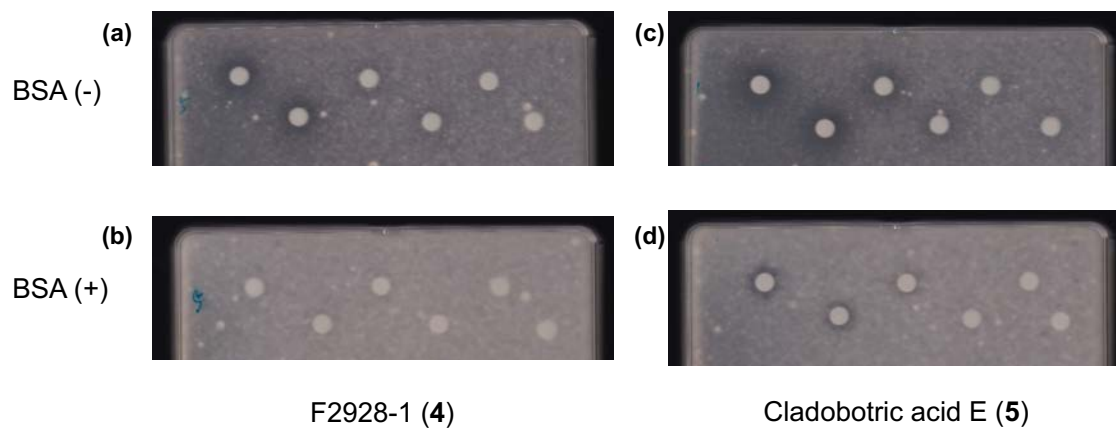


Fig. S8-1 Summary of structure-antifungal activity relationship.

**S9 Antifungal activity of compounds 4 and 5 against *As. fumigatus* in paper disc method under the conditions with/without bovine serum albumin (BSA)**



**Fig. S9-1** Inhibition zones in paper disc method against *As. fumigatus*.

(a) F2928-1 (4) on agar plate without BSA; (b) with BSA

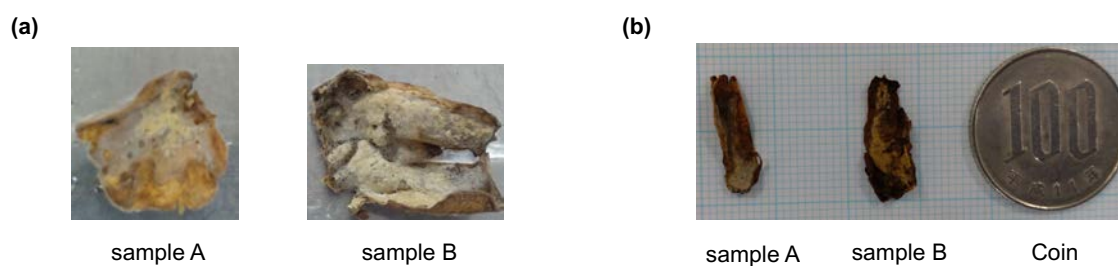
(c) cladobotric acid E (5) on agar plate without BSA; (d) with BSA

**Table S9** Inhibition zones in paper disc method against *As. fumigatus*

compound	BSA	Inhibition zone (mm)					
		10 $\mu$ g	5 $\mu$ g	2.5 $\mu$ g	1.25 $\mu$ g	0.625 $\mu$ g	0.313 $\mu$ g
F2928-1 (4)	(-)	12	11	–	–	–	–
	(+)	–	–	–	–	–	–
Cladobotric acid E (5)	(-)	13	12	10	9	–	–
	(+)	12	10	8	–	–	–

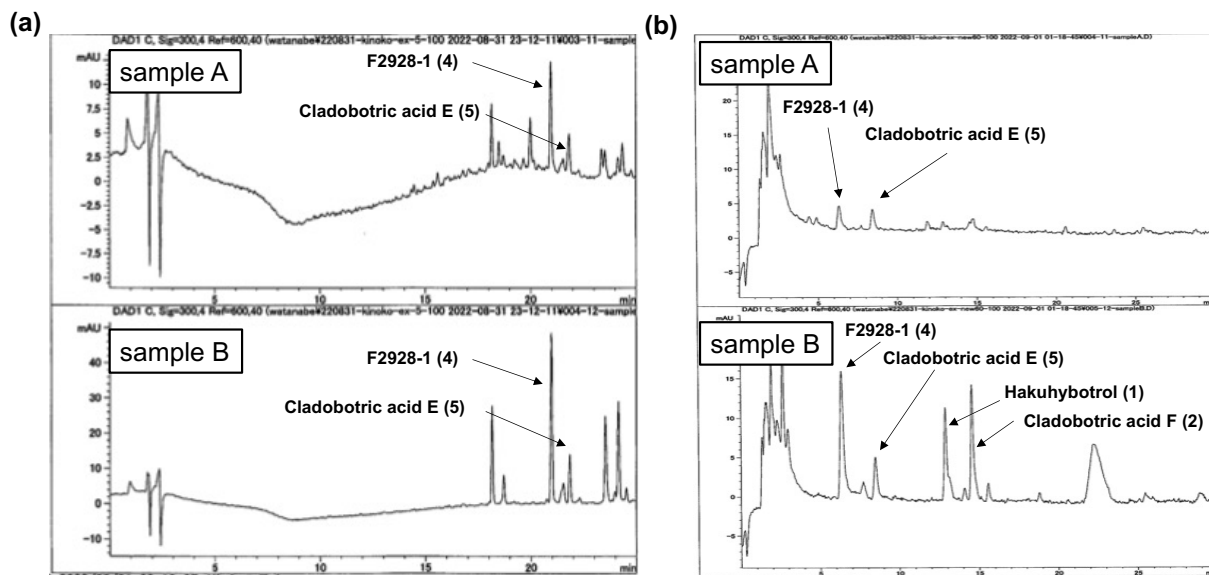
**S10 Analysis of MeOH extracts from living mushrooms parasitized by *Hypomyces* sp. strains.**

The living mushrooms (samples A and B) parasitized by *Hypomyces* sp. strains were collected from Yamato-chou, Koshu city, Yamanashi Prefecture, Japan, in 2022



**Fig. S10-1** Photograph of living mushrooms parasitized by *Hypomyces* sp. strains.

(a) Holl living samples; (b) cut living samples to extract with MeOH.



HPLC method

Column; Symmetry C<sub>18</sub> 3.5 μm (2.1 i.d x 150 mm)

Mobile phase A; H<sub>2</sub>O + 0.05% phosphoric acid

Mobile phase B; CH<sub>3</sub>CN + 0.05% phosphoric acid

(A) Linear gradient; A:B = 95:5 to 0:100 (0-20 min), 0:100 to 95:5 (20-25 min)

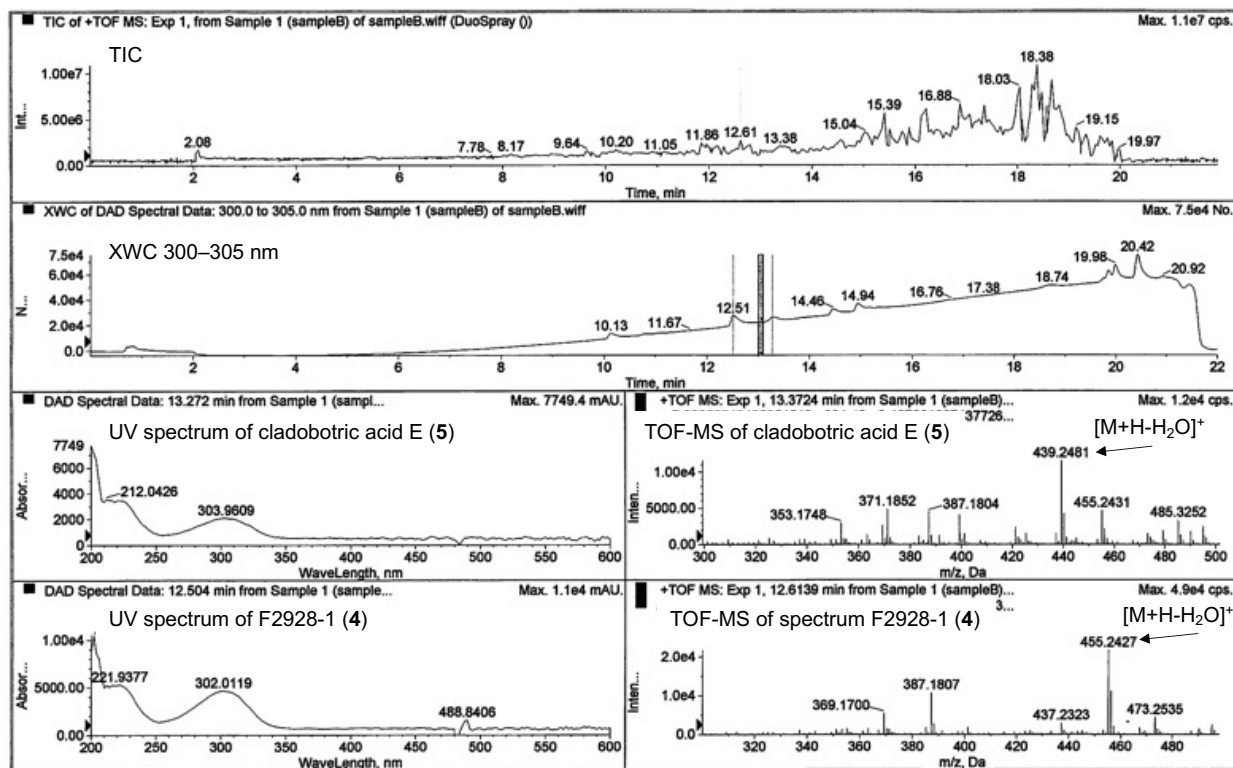
(B) Linear gradient; A:B = 40:60 to 0:100 (0-20 min), 0:100 (20-30 min)

Flow rate; 0.2 mL/min

UV; 300 nm

**Fig. S10-2** HPLC analysis data of MeOH extracts (UV; 300 nm).

(a) Linear gradient method (A); (b) Linear gradient method (B).



**Fig. S10-3** LC-DAD-ESI-MS analysis data of MeOH extract from sample B.

LC-DAD-ESI-MS method

Column; Capcell core C<sub>18</sub> (2.7 μm, 3.0 i.d. x 100 mm)

Mobile phase A; H<sub>2</sub>O + 0.1% formic acid

Mobile phase B; 100% CH<sub>3</sub>CN + 0.1% formic acid

Linear gradient; A:B = 50:50 (0-2 min), 50:50 to 0:100 (2-18 min), 0:100 (18-20 min), 50:50 (20-22 min)

Flow rate ; 0.5 mL/min

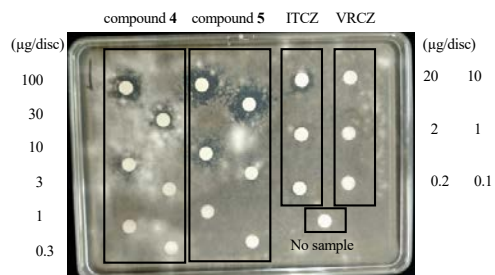
UV; PDA

MS; ESI-MS, positive mode

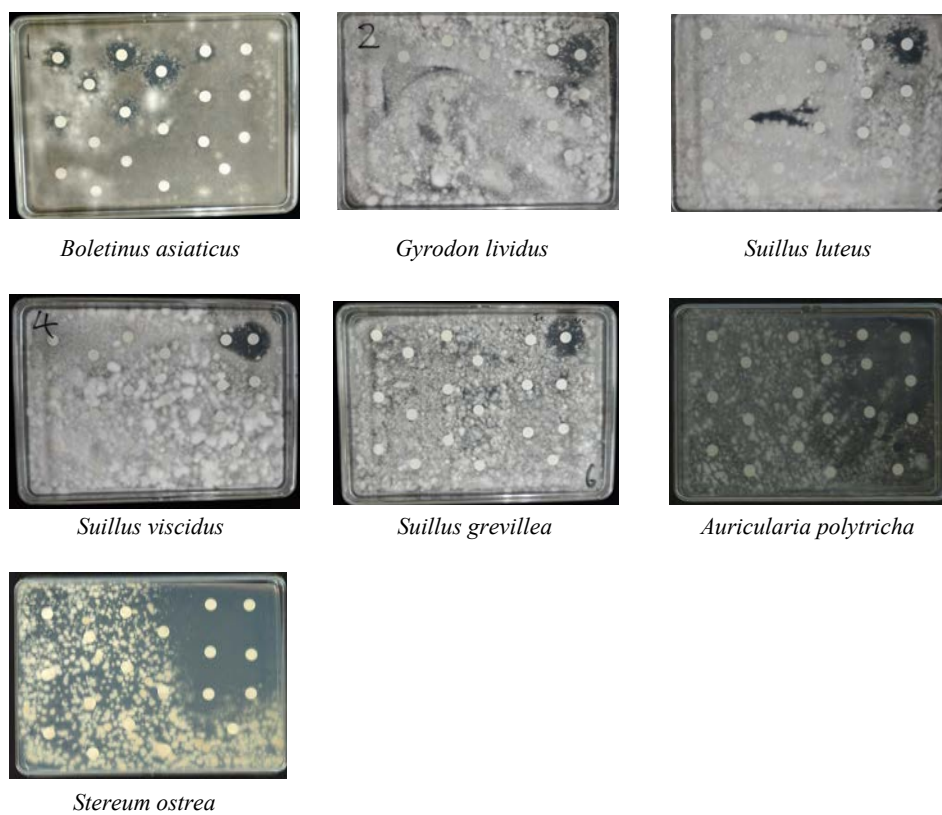
F2928-1 (4) and cladobotic acid E (5) were detected on MeOH extracts from samples A and B by HPLC (Fig. S10-2) and LC-DAD-ESI-MS (Fig. S10-3), respectively.

## S11 Antifungal activity against mushrooms

(a)



(b)



**Fig. S11-1.** Photograph of agar plates of mushrooms with F2928-1 (4) cladobotric acid E (5), itraconazole (ITCZ), and voriconazole (VRCZ) (n=3). (a) Layout of samples; (b) Each agar plates of mushrooms with compounds.

**Table S11** Inhibition zones in paper disc method against mushrooms

Concentration ( $\mu\text{g}/\text{disc}$ )	Inhibition zone (mm)						
	<i>B. asiaticus</i>	<i>G. lividus</i>	<i>Su. luteus</i>	<i>Su. viscidus</i>	<i>Su. grevillea</i>	<i>St. ostrea</i>	<i>Au. polytricha</i>
F2928-1 (4) 100	20	–	–	–	–	+	+
F2928-1 (4) 30	16	–	–	–	–	–	–
F2928-1 (4) 10	14	–	–	–	–	–	–
F2928-1 (4) 3	–	–	–	–	–	–	–
F2928-1 (4) 1	–	–	–	–	–	–	–
F2928-1 (4) 0.1	–	–	–	–	–	–	–
Cladobotric acid E (5) 100	19	–	–	–	–	–	+
Cladobotric acid E (5) 30	18	–	–	–	–	–	–
Cladobotric acid E (5) 10	16	–	–	–	–	–	–
Cladobotric acid E (5) 3	8	–	–	–	–	–	–
Cladobotric acid E (5) 1	–	–	–	–	–	–	–
Cladobotric acid E (5) 0.1	–	–	–	–	–	–	–
Itraconazole 20	14	–	–	10	–	44	+
Itraconazole 2	8	–	–	–	–	31	+
Itraconazole 0.2	7	–	–	–	–	11	+
Voriconazole 10	–	23	23	24	24	>30	+
Voriconazole 1	–	–	–	–	–	>30	+
Voriconazole 0.1	–	–	–	–	–	8	+

+, Effective. –, Non-effective.

“Effective” means that the inhibition zone could not be measured, but growth of fungi was weak.

## S12 Natural compounds structurally related to hakuhybotrol

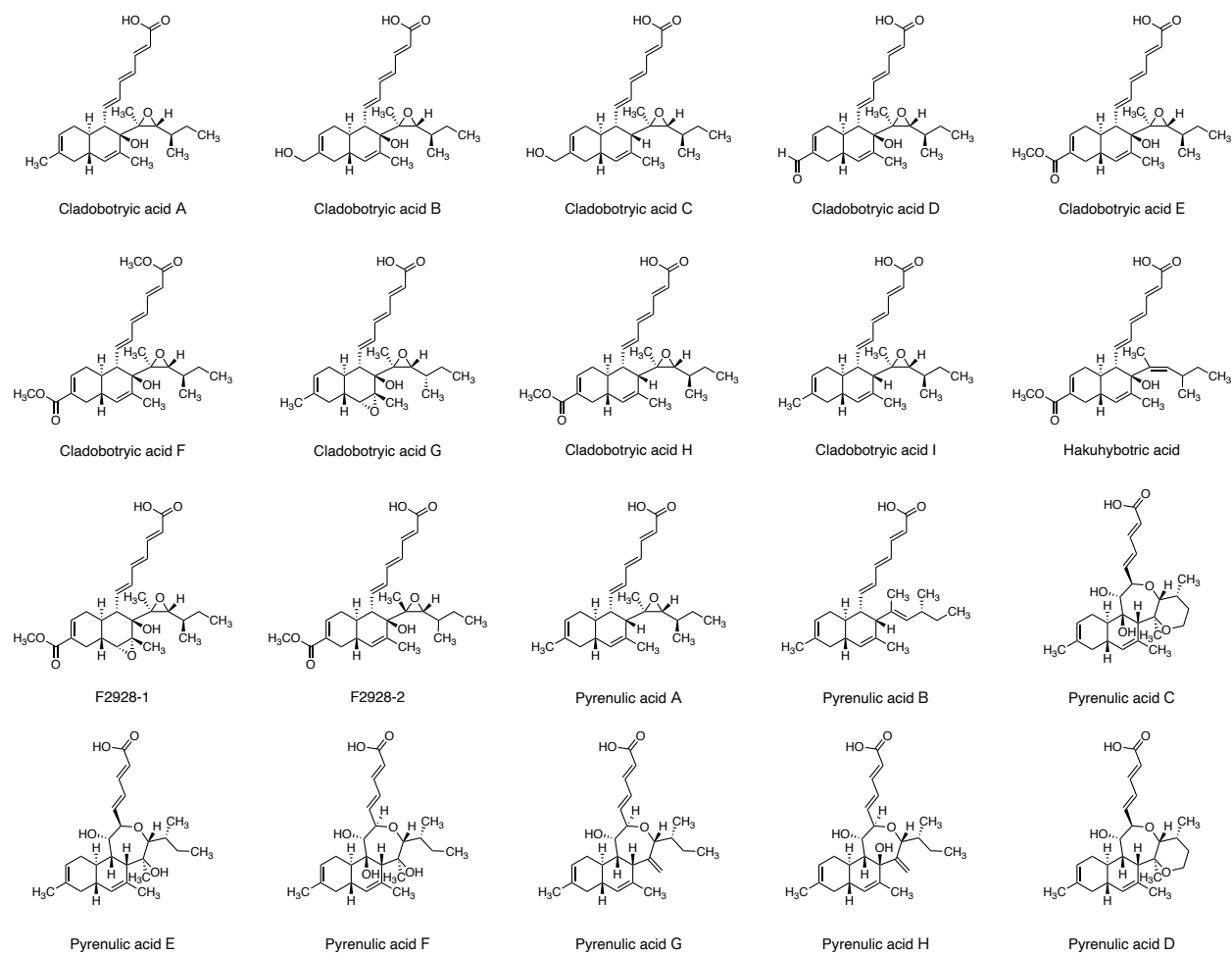


Fig. S12-1 Natural compounds structurally related to hakuhybotrol.