# Stereo-electronic Contributions in Yttrium-Mediated Stereoselective Ring-Opening Polymerization of Functional Racemic β-Lactones: ROP of 4-Alkoxymethylene-β-Propiolactones with Bulky Exocyclic Chains

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#### **General conditions**

#### **Material and methods**

All manipulations involving organometallic catalysts were performed under inert atmosphere (argon, <3 ppm O<sub>2</sub>) using standard Schlenk, vacuum line, and glovebox techniques. Toluene was freshly distillated from Na/benzophenone under argon and degassed thoroughly by freeze-thaw-vacuum cycles prior to use. Isopropyl alcohol was distilled over Mg turnings under argon atmosphere and kept over activated 3–4 Å molecular sieves. Proligands {ONNO<sup>*t*Bu2</sup>}H<sub>2</sub>, {ONNO<sup>*c*umyl2</sup>}H<sub>2</sub>, {ONNO<sup>Me2</sup>}H<sub>2</sub> and {ONNO<sup>*C*l2</sup>}H<sub>2</sub>, and precursor Y[N(SiHMe<sub>2</sub>)<sub>2</sub>]<sub>3</sub>(THF)<sub>1.5</sub>, used to prepare amido yttrium complexes **2a-d**, respectively, were synthesized according to the reported procedures.<sup>6a,13</sup>

#### Instrumentation and measurements

<sup>1</sup>H (500 and 400 MHz) and <sup>13</sup>C{<sup>1</sup>H} (125 MHz and 100 MHz) NMR spectra were recorded on Bruker Avance AM 500 and Ascend 400 spectrometers at 25 °C. <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra were referenced internally relative to SiMe<sub>4</sub> ( $\delta = 0$  ppm) using the residual solvent resonances.

Number-average molar mass ( $M_{n,SEC}$ ) and dispersity ( $D_M = M_w/M_n$ ) values of the PBPL<sup>FG</sup>s were determined by size-exclusion chromatography (SEC) in THF at 30 °C (flow rate = 1.0 mL.min<sup>-1</sup>) on a Polymer Laboratories PL50 apparatus equipped with a refractive index detector and a set of two ResiPore PLgel 3 µm MIXED-D 300 × 7.5 mm columns. The polymer samples were dissolved in THF (*ca.* 2 mg.mL<sup>-1</sup>). All elution curves were calibrated with polystyrene standards;  $M_{n,SEC}$  values of the PBPL<sup>FG</sup>s were uncorrected for the possible difference in hydrodynamic radius *vs.* that of polystyrene.

The molar mass of PBPL<sup>FG</sup> samples was also determined by <sup>1</sup>H NMR analysis in CDCl<sub>3</sub> from the relative intensities of the signals of the PBPL<sup>FG</sup> repeating unit methine hydrogen ( $\delta$ , ppm): 5.25 –OC*H*(CH<sub>2</sub>OCH(CH<sub>3</sub>)<sub>2</sub>), PBPL<sup>CH2O/Pr</sup>; 5.22 –OC*H*(CH<sub>2</sub>OC(CH<sub>3</sub>)<sub>3</sub>), PBPL<sup>CH2O/Bu</sup>; 5.20 –OC*H*(CH<sub>2</sub>OSi(CH<sub>3</sub>)<sub>2</sub>C(CH<sub>3</sub>)<sub>3</sub>), PBPL<sup>CH2OTBDMS</sup>; and of the isopropyl chain-end ( $\delta$ , ppm): 4.94–4.98 (CH<sub>3</sub>)<sub>2</sub>CHO–, 1.19–1.25 (CH<sub>3</sub>)<sub>2</sub>CHO–).

Monomer conversions were calculated from <sup>1</sup>H NMR spectra of the crude polymer samples in CDCl<sub>3</sub> by using the integration (Int.) ratios [Int.<sub>PBPL(FG)</sub> / (Int.<sub>PBPL(FG)</sub> + Int.<sub>BPL(FG)</sub>)] of the methine hydrogens of BPL<sup>FG</sup>s and PBPL<sup>FG</sup>s (corresponding methine hydrogen signal of the polymers (see above)), and of the monomers (d (ppm) 4.60 BPL<sup>CH2O/Pr</sup>, 4.55 BPL<sup>CH2O/Bu</sup>, and 4.65 BPL<sup>CH2OTBDMS</sup>). High resolution Matrix Assisted Laser Desorption Ionization - Time of Flight, MALDI-ToF, mass spectra of the polymers were recorded using an ULTRAFLEX III TOF/TOF spectrometer (Bruker Daltonik Gmbh, Bremen, Germany) in positive ionization mode. Spectra were recorded using reflectron mode and an accelerating voltage of 25 kV. A mixture of a freshly prepared solution of the polymer in THF or  $CH_2Cl_2$  (HPLC grade, 10 mg mL<sup>-1</sup>) and DCTB (*trans*-2-(3-(4-*tert*-butylphenyl)-2methyl-2-propenylidene) malononitrile, and a MeOH solution of the cationizing agent (NaI, 10 mg mL<sup>-1</sup>) were prepared. The solutions were combined in a 1:1:1 *v*/*v*/*v* ratio of matrix-to-sample-to-cationizing agent. The resulting solution (0.25–0.5 µL) was deposited onto the sample target (Prespotted AnchorChip PAC II 384 / 96 HCCA) and air or vacuum dried.

Differential scanning calorimetry (DSC) analyses were performed with a DSC2500 TA Instrument apparatus calibrated with indium using aluminum capsules (40  $\mu$ L). The thermograms were recorded under a continuous flow of helium (25 mL min<sup>-1</sup>) according to the following cycles: -80 to 200 °C at 10 °C min<sup>-1</sup>; 200 to -80 °C at 10 °C min<sup>-1</sup>; -80 °C for 5 min; -80 to 200 °C at 10 °C min<sup>-1</sup>; 200 to -80 °C at 10 °C min<sup>-1</sup>.

Synthesis and characterization of BPL<sup>CH2OR</sup> monomers. BPL<sup>CH2OR</sup> monomers were synthesized by carbonylation of the corresponding racemic or enantiopure glycidyl ethers (*rac-*/(*S*)-Glyc<sup>CH2OR</sup>) using a previously reported procedure.<sup>1</sup> All *rac*-BPL<sup>CH2OR</sup> and (*S*)-BPL<sup>CH2OR</sup> monomers were stored under argon at -27 °C.

*Rac/(S)*-BPL<sup>CH2O*i*Pr</sup>. Using [Salph(Cr(THF)<sub>2</sub>)][Co(CO)<sub>4</sub>] (431 mg, 0.47 mmol) and *rac*-Glyc<sup>CH2O*i*Pr</sup> (5.51 g, 47.51 mmol, 100 equiv), the carbonylation reaction afforded *rac*-BPL<sup>CH2O*i*Pr</sup> which was isolated following a double distillation using a Kügelrorh oven (180 °C, 0.1 torr) as a colorless viscous liquid (4.45 g, 65% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 25 °C)  $\delta$  (ppm):  $\delta$  4.60 (dtd, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 6 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 4 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 3 Hz, 1H), 3.74 (dd, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 12 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 3 Hz, 1H), 3.68 – 3.58 (m, 2H), 3.42 (dd, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 16 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 6 Hz, 1H), 3.35 (dd, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 16 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 4 Hz, 1H), 1.15 (d, *J* = 6 Hz, 6H) (Figure S1). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 25 °C)  $\delta$  (ppm): 72.6 (OCH(CH<sub>3</sub>)<sub>2</sub>), 69.6 (CHOC(O)), 67.4 (CH<sub>2</sub>OCH), 39.4 (CH<sub>2</sub>C(O)O), 21.8 (CH(*C*H<sub>3</sub>)<sub>2</sub>), 21.8 (CH(*C*H<sub>3</sub>)<sub>2</sub>) (Figure S2). Carbonylation of (*S*)-BPL<sup>CH2O*i*Pr</sup> was performed similarly but starting from (*S*)-Glyc<sup>CH2O*i*Pr</sup> and gave (*S*)-BPL<sup>CH2O*i*Pr</sup> as a colorless viscous liquid (4.3 g, 63% yield) that displayed NMR spectra identical to those of *rac*-BPL<sup>CH2O*i*Pr</sup>.

*Rac-/(S)*-BPL<sup>CH2O/Bu</sup> Using [Salph(Cr(THF)<sub>2</sub>)][Co(CO)<sub>4</sub>] (281 mg, 0.31 mmol) and *rac*-Glyc<sup>CH2O/Bu</sup> (4.03 g, 31.0 mmol, 100 equiv), the carbonylation reaction afforded *rac*-

BPL<sup>CH2O/Bu</sup> which was isolated following a double distillation using a Kügelrorh oven (190-200 °C, 0.1 torr) as a colorless viscous liquid (3.13 g, 64% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 25 °C)  $\delta$  (ppm):  $\delta$  4.55 (dtd, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 6 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 4 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 3 Hz, 1H), 3.66 (dd, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 11 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 3 Hz, 1H), 3.54 (dd, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 11 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 4 Hz, 1H), 3.37 (dd, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 16 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 6 Hz, 1H), 3.31 (dd, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 16 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 5 Hz, 1H), 1.15 (s, 9H) (Figure S1). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 25 °C)  $\delta$  (ppm):  $\delta$  167.9 (*C*=O), 74.3 (O*C*(CH<sub>3</sub>)<sub>3</sub>), 69.6 (*C*HOC(O)), 63.6 (*C*H<sub>2</sub>OC), 39.4 (*C*H<sub>2</sub>C(O)O), 27.2 (*C*(*C*H<sub>3</sub>)<sub>3</sub>) (Figure S2). ESI-MS *m*/*z*<sub>found</sub> = 181.0833 *vs*. *m*/*z*<sub>calculated</sub> = 181.0835. Carbonylation of (*S*)-BPL<sup>CH2O/Bu</sup> was performed similarly but starting from (*S*)-Glyc<sup>CH2O/Bu</sup> and gave (*S*)-BPL<sup>CH2O/Bu</sup> as a colorless viscous liquid (3.0 g, 61% yield) that displayed NMR spectra identical to those of *rac*-BPL<sup>CH2O/Bu</sup>.

*Rac-/(S)*-BPL<sup>CH2OTBDMS</sup>.<sup>2</sup> Using [Salph(Cr(THF)<sub>2</sub>)][Co(CO)<sub>4</sub>] (246 mg, 0.27 mmol) and *rac*-Glyc<sup>CH2OTBDMS</sup> (5.57 g, 29.64 mmol, 100 equiv) afforded *rac*-BPL<sup>CH2OTBDMS</sup> which was isolated following a double distillation using a Kügelrorh oven (140 °C, 0.1 torr) as a colorless viscous liquid (3.8 g, 60% yield). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>, 25 °C)  $\delta$  (ppm): 4.65–4.46 (m, 1H), 4.02 (dd, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 12 Hz, <sup>2</sup>*J*<sub>*H*-*H*</sub> = 3 Hz, 1H), 3.82 (dd, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 12 Hz, <sup>3</sup>*J*<sub>*H*-*H*</sub> = 3 Hz, 1H), 3.53–3.28 (m, 2H), 0.91 (s, 9H), 0.09 (s, 6H) (Figure S1). <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>, 25 °C)  $\delta$  (ppm): 171.7 (*C*=O), 74.8 (*C*HOC(O)), 63.9 (*C*H2OTBDMS), 37.4 (*C*H2C(O)O), 25.7 Si(CH<sub>3</sub>)<sub>2</sub>*C*(CH<sub>3</sub>)<sub>3</sub>, 15.7 Si(CH<sub>3</sub>)<sub>2</sub>*C*(*C*H<sub>3</sub>)<sub>3</sub>, -4.9 Si(*C*H<sub>3</sub>)<sub>2</sub>*C*(*C*H<sub>3</sub>)<sub>3</sub> (Figure S2). Carbonylation of (*S*)-BPL<sup>CH2OTBDMS</sup> was performed similarly but starting from (*S*)-Glyc<sup>CH2OTBDMS</sup> and gave (*S*)-BPL<sup>CH2OTBDMS</sup> as a colorless viscous liquid (3.65 g, 58% yield) that displayed NMR spectra identical to those of *rac*-BPL<sup>CH2OTBDMS</sup>.

### References

a) J. A. Schmidt, E. B. Lobkovsky and G. W. Coates, *J. Am. Chem. Soc.*, 2005, *127*, 11426–11435; b) J. W. Kramer, G. W. Coates, *Tetrahedron* 2008, *64*, 6973–6978.

<sup>&</sup>lt;sup>2</sup> R. M. Shakaroun, H. Li, P. Jéhan, A. Alaaeddine, J.-F. Carpentier, S. M. Guillaume, *Polym. Chem.*, **2020**, *11*, 2640–2652.



**Figure S1.** <sup>1</sup>H NMR spectra (500 MHz, CDCl<sub>3</sub>, 23 °C) of *rac*-BPL<sup>CH2OTBDMS</sup>, *rac*-BPL<sup>CH2OtBu</sup>, *rac*-BPL<sup>CH2OtPr</sup> monomers (from top to bottom).



**Figure S2.** J-MOD / <sup>13</sup>C{<sup>1</sup>H} NMR spectra (100 MHz, CDCl<sub>3</sub>, 25 °C) of *rac*-BPL<sup>CH2OTBDMS</sup>, *rac*-BPL<sup>CH2O*t*Bu</sup>, and *rac*-BPL<sup>CH2O*t*Pr</sup> monomers (from top to bottom).



**Figure S3**. <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>, 25 °C) (top) and J-MOD (125 MHz, CDCl<sub>3</sub>, 25 °C) (bottom) NMR spectra of a syndio-enriched PBPL<sup>CH2O*i*Pr</sup> prepared from the ROP of *rac*-BPL<sup>CH2O*i*Pr</sup> mediated by the **2b**/*i*PrOH (1:1) system (Table 1, entry 6).



**Figure S4.** <sup>1</sup>H-<sup>1</sup>H COSY NMR spectrum (500 MHz, CDCl<sub>3</sub>, 25 °C) of a syndio-enriched PBPL<sup>CH2O/Pr</sup> (Table 1, entry 6).



**Figure S5.** <sup>1</sup>H-<sup>13</sup>C HMBC NMR spectrum (500 MHz, CDCl<sub>3</sub>, 25 °C) of a syndio-enriched PBPL<sup>CH2O*i*Pr</sup> (Table 1, entry 6).



**Figure S6.** MALDI-ToF mass spectrum (DCTB matrix, ionized by Na<sup>+</sup>) of a PBPL<sup>CH2O/Pr</sup> (Table 1, entry 1), showing populations I (major, top) and II (minor, bottom) showing a repeating unit of m/z 144 corresponding to the BPL<sup>CH2O/Pr</sup> monomer unit. Right and middle zoomed regions correspond to the simulated (top) and experimental (bottom) spectra for populations I and II, respectively. The major population (I) corresponds to  $\alpha$ -isopropoxy, $\omega$ -hydroxyl telechelic PBPL<sup>CH2O/Pr</sup> chains ionized by Na<sup>+</sup>, as confirmed by the close match with the corresponding isotopic simulation of [(CH<sub>3</sub>)<sub>2</sub>CHO(COCH<sub>2</sub>CH(CH<sub>2</sub>OC<sub>3</sub>H<sub>7</sub>)O)<sub>n</sub>H]·Na<sup>+</sup> with, for example,  $m/z_{calculated} = 1379.7545$  *vs.*  $m/z_{found} = 1379.746$  for n = 9. The minor population (II) corresponds to  $\alpha$ -carboxylic, $\omega$ -hydroxy telechelic PBPL<sup>CH2O/Pr</sup> chains ionized by Na<sup>+</sup>, as confirmed by Na<sup>+</sup>, as confirmed by Na<sup>+</sup>, as confirmed by the close match with the corresponds to  $\alpha$ -carboxylic, $\omega$ -hydroxy telechelic PBPL<sup>CH2O/Pr</sup> chains ionized by Na<sup>+</sup>, as confirmed by the close match with the corresponds to  $\alpha$ -carboxylic, $\omega$ -hydroxy telechelic PBPL<sup>CH2O/Pr</sup> chains ionized by Na<sup>+</sup>, as confirmed by the close match with the sourcesponding isotopic simulation illustrated for [HO(C<sub>7</sub>H<sub>12</sub>O<sub>3</sub>)<sub>n</sub>H]·Na<sup>+</sup> with, for example,  $m/z_{calculated} = 1337.7076$  *vs.*  $m/z_{found} = 1337.709$  for n = 9. This latter population likely results from hydrolysis during the MS sample preparation/ionization.



**Figure S7.** <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>, 25 °C) (top) and J-MOD (125 MHz, CDCl<sub>3</sub>, 25 °C) (bottom) NMR spectra of a syndio-enriched PBPL<sup>CH2O*t*Bu</sup> prepared from the ROP of *rac*-BPL<sup>CH2O*t*Bu</sup> mediated by the **2a**/*i*PrOH system (Table 2, entry 11).



**Figure S8.** <sup>1</sup>H-<sup>1</sup>H COSY NMR spectrum (500 MHz, CDCl<sub>3</sub>, 25 °C) of a syndio-enriched PBPL<sup>CH2O/Bu</sup> (Table 2, entry 11).



**Figure S9.** <sup>1</sup>H-<sup>13</sup>C HMBC NMR spectrum (500 MHz, CDCl<sub>3</sub>, 25 °C) of a syndio-enriched PBPL<sup>CH2OrBu</sup> (Table 2, entry 11).



**Figure S10.** MALDI-ToF mass spectrum (DCTB matrix, ionized by Na<sup>+</sup>) of a PBPL<sup>CH2O/Bu</sup> (Table 2, entry 5). Right zoomed regions correspond to the simulated (top) and experimental (bottom) spectra of the main population (Table 2, entry 5). The expected population of macromolecules having a repeating unit of m/z 158, corresponding to  $\alpha$ -isopropoxy, $\omega$ -hydroxyl telechelic PBPL<sup>CH2O/Bu</sup> chains ionized by Na<sup>+</sup> is clearly observed. This is confirmed by the close match with the corresponding isotopic simulations, as illustrated for [(CH<sub>3</sub>)<sub>2</sub>CHO(COCH<sub>2</sub>CH(CH<sub>2</sub>OC<sub>4</sub>H<sub>9</sub>)O)<sub>n</sub>H]·Na<sup>+</sup> with, for example,  $m/z_{calculated}$  1189.7068 *vs.*  $m/z_{found}$  1189.698 for n = 7.



**Figure S11.** Variation of  $M_{n,NMR}$  O,  $M_{n,SEC}$   $\bigcirc$ , and  $M_{n,theo}$  (solid line) values of PBPL<sup>CH2O/Bu</sup> synthesized from the ROP of *rac*-BPL<sup>CH2O/Bu</sup> mediated by the **2b**/*i*PrOH (1:1) system as a function of the monomer loading/conversion (Table 2, entries 5–9).



**Figure S12.** <sup>1</sup>H (500 MHz, CDCl<sub>3</sub>, 25 °C) (top) and J-MOD (125 MHz, CDCl<sub>3</sub>, 25 °C) (bottom) NMR spectra of a syndio-enriched PBPL<sup>CH2OTBDMS</sup> prepared from the ROP of *rac*-BPL<sup>CH2OTBDMS</sup> mediated by the **2a**/*i*PrOH (1:1) system (Table 3, entry 7); \* stands for residual monomer, toluene and water resonances.



**Figure S13.** <sup>1</sup>H-<sup>1</sup>H COSY NMR spectrum (500 MHz, CDCl<sub>3</sub>, 25 °C) of a syndio-enriched PBPL<sup>CH2OTBDMS</sup> (Table 3, entry 7).



**Figure S14**. <sup>1</sup>H-<sup>13</sup>C HMBC NMR spectrum (500 MHz, CDCl<sub>3</sub>, 25 °C) of a syndio-enriched PBPL<sup>CH2OTBDMS</sup> (Table 3, entry 7).



**Figure S15.** MALDI-ToF mass spectrum (DCTB matrix, ionized by Na<sup>+</sup>) (top, full spectrum; bottom, detail of the region for DP = 7-9) of a low molar mass PBPL<sup>CH2OTBDMS</sup> ( $M_{n,NMR} = 4000$ ,  $M_{n,SEC} = 6300$  g.mol<sup>-1</sup>) prepared from the ROP of 25 equiv of *rac*-BPL<sup>CH2OTBDMS</sup> with the **2a**/*i*PrOH (1:1) system (entry not reported in Table 3). The major populations observed (**I**) corresponds to  $\alpha$ -isopropoxy, $\omega$ -hydroxyl telechelic PBPL<sup>CH2OTBDMS</sup> chains ionized by Na<sup>+</sup> (see the zoomed regions featuring the simulated (top) and experimental (bottom) spectra). The minor populations (**II-IV**) most likely arise from hydrolysis during the MS sample preparation/ionization.



**Figure S16.** Variation of  $M_{n,NMR}$  (2),  $M_{n,SEC}$   $\bigcirc$ , and  $M_{n,theo}$  (solid line) molar mass values of PBPL<sup>CH2OTBDMS</sup> synthesized from the ROP of *rac*-BPL<sup>CH2OTBDMS</sup> mediated by the **2b**/*i*PrOH (1:1) catalyst system as a function of the BPL<sup>CH2OTBDMS</sup> monomer loading/conversion (Table 3, entries 2–7).



**Figure S17.** DSC thermogram (heating rate = 10 °C min<sup>-1</sup>, second heating cycle –80 to 200 °C) of a syndio-enriched PBPL<sup>CH2O*i*Pr</sup> ( $P_r = 0.86$ ) prepared by ROP of *rac*-BPL<sup>CH2O*i*Pr</sup> with the **2b**/*i*PrOH system (Table 1, entry 8).



**Figure S18.** DSC thermogram (heating rate = 10 °C min<sup>-1</sup>, second heating cycle –80 to 200 °C) of a syndio-enriched PBPL<sup>CH2O/Bu</sup> ( $P_r = 0.84$ ) prepared by ROP of *rac*-BPL<sup>CH2O/Bu</sup> with the **2b**/*i*PrOH system (Table 2, entry 5).



**Figure S19.** DSC thermogram (heating rate = 10 °C min<sup>-1</sup>, second heating cycle –80 to 200 °C) of a syndio-enriched PBPL<sup>CH2OTBDMS</sup> ( $P_r = 0.81$ ) prepared by ROP of *rac*-BPL<sup>CH2OTBDMS</sup> with the **2b**/*i*PrOH system (Table 3, entry 6).



**Figure S20.** DSC thermogram (heating rate = 10 °C min<sup>-1</sup>, second heating cycle –80 to 200 °C) of a syndio-enriched PBPL<sup>CH2OTBDMS</sup> ( $P_r = 0.81$ ) prepared by ROP of *rac*-BPL<sup>CH2OTBDMS</sup> with the **2b**/*i*PrOH system (Table 3, entry 7).