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**Electronic Supplementary Information** 

# Systematic study on the effects of the structure of block copolymers of PEG and poly(ɛ-caprolactone-co-glycolic acid) on their temperatureresponsive sol-to-gel transition behavior

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# MATERIALS AND METHODS

#### Materials

Poly(ethylene glycol) (PEG) (molecular weight (MW) = 1000 g/mol (PEG<sub>1000</sub>), 1540 g/mol (PEG<sub>1540</sub>), and 2000 g/mol (PEG<sub>2000</sub>)),  $\varepsilon$ -caprolactone (CL), tin-2-ethylhexanoate (Sn(Oct)<sub>2</sub>), butyric acid anhydride, 4-dimethylaminopyridine (DMAP), anhydrous dichloromethane, as well as other chemicals and organic solvents were purchased from FUJIFILM Wako Pure Chemical Ind., Ltd. (Osaka, Japan). Mono-methoxy poly(ethylene glycol) (MeO-PEG) (MW = 550 g/mol, MeO-PEG<sub>550</sub>) and glycolide (GL) were obtained from Sigma-Aldrich Co. LLC (St. Louis, MO, USA). Hexamethylene diisocyanate (HMDI) and acetic anhydride (Ac<sub>2</sub>O) were purchased from Tokyo Chemical Industry Co. Ltd. (Tokyo, Japan). Branched PEG with four arms (MW = 5000 g/mol, 4-arm PEG<sub>5000</sub>) was supplied by NOF Co. (Tokyo, Japan). Water was purified using Millipore Elix UV3 direct-Q UV (Merck, Darmstadt, Germany).

#### Measurements

<sup>1</sup>H nuclear magnetic resonance (<sup>1</sup>H-NMR) spectra were obtained through an NMR spectrometer (400 MHz, JNM-GSX-400, JEOL, Tokyo, Japan) using deuterated solvent (CDCl<sub>3</sub>). The chemical shifts were calibrated using tetramethylsilane (TMS). The number average molecular weight ( $M_n$ ) of the polymers was calculated from <sup>1</sup>H-NMR spectra. The weight average molecular weight ( $M_w$ ) and polydispersity index ( $M_w/M_n$ ) of the polymers were determined using size exclusion chromatography (SEC) (column: TSKgel Multipore H<sub>XL</sub>-M × 2; detector: RI). The measurements were performed using dimethylformamide (DMF) as an eluent with a flow rate of 1.0 mL min<sup>-1</sup> at 40 °C using a series of PEGs as standards. Temperature-dependent rheological measurements of the sol-to-gel transition of the polymer solutions were performed using a dynamic rheometer (Thermo HAAKE RS600, Thermo Fisher Scientific, Waltham, MA, USA) according to a study reported previously.<sup>1</sup> Thermal analysis of the copolymers and solutions was conducted using a differential scanning calorimeter (DSC-60, Shimadzu) with sealed aluminum pans.

# Synthesis of PCGA-PEG-PCGA (ABA triblock copolymer)

ABA triblock copolymers of poly( $\varepsilon$ -caprolactone-*co*-glycolide) (PCGA) and PEG (PCGA-PEG-PCGA) were synthesized by ring-opening copolymerization of CL and GL in the presence of PEG as a macroinitiator and Sn(Oct)<sub>2</sub> as a catalyst, according to the method described in previous works (**Scheme S1**). <sup>1-3</sup> PEG<sub>1540</sub> (16.0 g, 10.4 mmol) was dried under *in vacuo* at 120 °C for 5 h in a 100-mL flask with a stopcock. After cooling to room temperature (r.t.), CL (34.4 g, 302 mmol), GL (6.04 g, 52.1 mmol), and Sn(Oct)<sub>2</sub> (151 mg, 372 µmol) were added to the flask and dried further *in vacuo* at r.t. for 12 h. Polymerization was carried out at 160 °C for 12 h by heating the flask in an oil bath. The product was purified by reprecipitation using chloroform (100 mL) as a good solvent and diethyl ether

(1000 mL) as a poor solvent to obtain the PCGA-PEG-PCGA triblock copolymer as a colorless liquid or white solid. The yield after reprecipitation was 52.3 g (92.7%). Various PCGA-PEG-PCGA triblock copolymers were synthesized by changing CL and GL feed amounts and the MW of PEG (using PEG<sub>1000</sub> or PEG<sub>2000</sub>). The products were analyzed using <sup>1</sup>H-NMR, and the CL/GA ratio (GL corresponds two GA units) was determined from the <sup>1</sup>H-NMR spectra. The obtained PCGA-PEG-PCGA triblock copolymers (ABA series) were given code names such as ABA1.5k-3.1k-3.4, where 1.5k = MW of a PEG segment, 3.1k = sum MW of two PCGA segments, and 3.4 = molar ratio of CL/GA in a PCGA segment. Typical <sup>1</sup>H-NMR spectra and SEC elution profiles for ABA triblock copolymers are shown in **Figures S1(a)–(k)**.

# Synthesis of PEG-PCGA-PEG (BAB triblock copolymer)

AB-type diblock copolymers of PEG and PCGA (PEG-PCGA) were synthesized by ring-opening copolymerization of CL and GL in the presence of MeO-PEG<sub>550</sub> as a macroinitiator and Sn(Oct)<sub>2</sub> as a catalyst according to the method described below (Scheme S2). MeO-PEG<sub>550</sub> (9.42 g, 17.1 mmol) was dried in vacuo at 120 °C for 5 h in a 100-mL flask with a stopcock. After cooling to r.t., CL (18.0 g, 158 mmol), GL (2.62 g, 22.5 mmol), and Sn(Oct)<sub>2</sub> (79.8 mg, 197 µmol) were added to the flask and dried further in vacuo at r.t. for 12 h. Polymerization was carried out at 160 °C for 12 h by heating the flask in an oil bath. The product was purified using the same reprecipitation method described above. A white solid or colorless liquid of PEG-PCGA diblock copolymer was obtained. Yield: 18.4 g (61.5%). Various PEG-PCGA diblock copolymers were synthesized by changing CL and GL feed amounts. The obtained PEG-PCGA (4.50 g, 2.14 mmol) was placed in a flask and dried in vacuo at 120 °C for 4 h. Anhydrous dichloromethane was added to the flask to dissolve PEG-PCGA. HMDI (183 mg, 1.09 mmol) was dissolved in anhydrous dichloromethane and refluxed at 70 °C for 6 h. The product was purified using the same reprecipitation method described above. A white solid (in some cases, a paleyellow solid or liquid) of the PEG-PCGA-PEG triblock copolymer was obtained. The yield after reprecipitation was 3.14 g (68.3%). The obtained PEG-PCGA-PEG triblock copolymers (BAB series) were given code names such as BAB1.1k-3.8k-3.9, where 1.1k = sum MW of two PEG segments and 3.8k = MW of the PCGA segment, and the molar ratio of CL/GA in the PCGA segment is 3.9. Typical <sup>1</sup>H-NMR spectra and SEC elution profiles for BAB triblock copolymers are shown in **Figures S2(a)**-(f).

#### Synthesis of ABA-Ac and ABA-Bu series

The introduction of an acetyl or butyryl group on the termini of PCGA-PEG-PCGA was carried out under an N<sub>2</sub> atmosphere, as shown in **Scheme S3**. The previously prepared ABA1.5k-3.1k-3.4 (0.87 g, 0.19 mmol) was placed in a 30-mL flask and dried *in vacuo* for 2h. Pyridine (3 mL) was added to the flask to dissolve the polymer. Ac<sub>2</sub>O (17.8  $\mu$ L, 0.19 mmol) and DMAP (33.0 mg, 0.27 mol) were

then added to the flask. The reaction mixture was stirred at 80 °C for 1 h. Pyridine, excess Ac<sub>2</sub>O, and acetic acid by-products were removed under reduced pressure. The product was purified by reprecipitation using chloroform (1.5 mL) as a good solvent and a mixture of diethyl ether and methanol (10/1, v/v) (30 mL) or diethyl ether (30 mL) as a poor solvent to give a white solid of the partially acetylated PCGA-PEG-PCGA, ABA-Ac series. The yield after reprecipitation was 0.53 g (67.1%). By varying the feed ratio of Ac<sub>2</sub>O or butyric acid anhydride, PCGA-PEG-PCGA polymers with various degrees of substitution (DS) of the acetyl or butyryl groups (ABA-Ac or ABA-Bu series) were synthesized. The products were analyzed using <sup>1</sup>H-NMR, and the DS values of the acetyl or butyryl groups were calculated from the <sup>1</sup>H-NMR spectra. The obtained partially acetylated or butyrylated PCGA-PEG-PCGAs were given code names such as ABA-Ac<sub>96</sub> or ABA-Bu<sub>91</sub>, where 96 or 91 = DS of acetyl or butyryl groups, respectively. Typical <sup>1</sup>H-NMR spectra and SEC elution profiles for ABA-Ac and ABA-Bu series are shown in **Figures S3(a)–(d), S4(a)–(b)**.

## Synthesis of 4-arm PEG-PCGA and partially acetylated 4-arm PEG-PCGA

4-Arm PEG-PCGA and partially acetylated 4-arm PEG-PCGA were synthesized using a method similar to that described above, using 4-arm PEG<sub>5000</sub> as a macroinitiator and subsequent acetylation (Scheme S4).

4-Arm PEG<sub>5000</sub> (1.49 g, 0.298 mmol) was dried *in vacuo* at 120 °C for 3 h in a 30-mL flask with a stopcock. After cooling to r.t., CL (3.29 g, 28.9 mmol), GL (0.577 g, 4.97 mmol), and Sn(Oct)<sub>2</sub> (17.9 mg, 44.2 µmol) were added to the flask. The flask was then cooled with liquid N<sub>2</sub> and dried overnight *in vacuo*. Polymerization was performed at 160 °C for 12 h by heating the flask in an oil bath. The product was purified by the same reprecipitation method. A white or pale-yellow solid of 4-arm PEG-PCGA was obtained. The yield after reprecipitation was 4.33g (80.3%). Various 4-arm PEG-PCGAs were synthesized by varying CL and GL feed amounts. The products were analyzed using <sup>1</sup>H-NMR, and the CL/GA ratio was determined from the <sup>1</sup>H-NMR spectra. The obtained 4-arm PEG-PCGAs (4-arm series) were given code names such as 4-arm5.0k-7.6k, where 5.0k = MW of 4-arm PEG and 7.6k = sum MW of four PCGA segments. Typical <sup>1</sup>H-NMR spectra and SEC elution profiles for 4-arm series are shown in **Figures S5(a)–(d)**.

The prepared 4-arm5.0k-7.6k (0.60 g, 0.047 mmol) was placed in a 30-mL flask and dried *in vacuo* for 2h. Pyridine (3 mL) was added to the flask to dissolve the polymer. Ac<sub>2</sub>O (26.8  $\mu$ L, 0.071 mmol) was then added. The reaction mixture was stirred at 80 °C for 1 h. Pyridine, excess Ac<sub>2</sub>O, and acetic acid by-products were removed under reduced pressure. The product was purified by the same precipitation method. A white solid, partially acetylated 4-arm PEG-PCGA, was obtained. The yield after reprecipitation was 0.56 g (93.1%). By varying the amount of Ac<sub>2</sub>O, partially acetylated 4-arm PEG-PCGA with various DS of acetyl groups (4-arm-Ac series) were synthesized. The products were analyzed using <sup>1</sup>H-NMR, and the DS values of the acetyl groups were calculated from the <sup>1</sup>H-NMR

spectra. The obtained partially acetylated 4-arm PEG-PCGAs (4-arm-Ac series) were named 4-arm- $Ac_{18}$ , where 18 = DS of the acetyl group. Typical <sup>1</sup>H-NMR spectra and SEC elution profiles for 4-arm-Ac series are shown in **Figures S6(a)–(d)**.

#### Morphology of the polymers

For the polymer morphology, photographs of the neat polymers (in dry state at r.t.) are shown in **Figures S7–12**. No photograph was recorded for the samples not shown.

### Confirmation of sol-to-gel transition behavior

The sol-to-gel transition behavior of the aqueous solutions of the polymers was investigated by a testtube inverting method.<sup>4</sup> A vial containing the polymer dissolved in phosphate-buffered saline (PBS, pH = 7.4) was immersed in a water bath at the desired temperature for 15 min, removed from the water bath, then inverted repeatedly within 30 sec to determine  $T_{gel}$  based on the criteria of "flow" (= sol) and "no flow" (= gel). This was repeated with a temperature increment of 1 °C per step. The measurements were repeated thrice at each temperature to determine the transition temperature in the phase diagram. Typical phase diagrams for the polymers are shown in **Figure S13**. Polymer concentrations were 20 wt.% for determine transition temperature in Table S1-S5, otherwise cited.

Temperature-dependent rheological measurements of the temperature-responsive sol-to-gel transition of the polymer solution in PBS (pH = 7.4) were performed using a dynamic rheometer (Thermo HAAKE RS600, Thermo Fisher Scientific, Waltham, MA, USA). A solvent trap was used to prevent solvent vaporization. The polymer concentration was 30 wt. %. Typically, each sample was placed between parallel plates (25 mm diameter and 1.0 mm gap) using a syringe. The data was collected under controlled stress (4.0 dyn/cm<sup>2</sup>) at a frequency of 1.0 rad/s. The heating rate was 0.5 °C/min. The storage modulus (G') and loss modulus (G') of the formulations were monitored in the range of 20-50 °C, and the gelation temperature ( $T_{gel}$ ) was defined as the crossover point from G' to G''. Typical rheological measurements are shown in **Figure S14**.

Code	MW of PEG (g/mol) <sup>a)</sup>	MW of PCGA (g/mol) <sup>b)</sup>	CL/GA (mol/mol) <sup>c)</sup>	Total $M_n$ (g/mol) <sup>d)</sup>	$M_{\rm w}/M_{\rm n}^{\rm e)}$	PCGA% <sup>f)</sup>	Morphology	$T_{\text{gel}}$ (°C) <sup>g)</sup>	Other transition temperature (°C) <sup>h)</sup>
BAB1.1k-2.1k-2.4		2100	2.4	3200	1.5	65.6	Viscous liquid	N.D.	53 <sup>i)</sup>
BAB1.1k-2.5k-4.4		2500	4.4	3600	1.6	69.4	Sticky solid	N.D.	52 <sup>i)</sup>
BAB1.1k-2.9k-3.6		2900	3.6	4000	1.5	72.5	Sticky solid	N.D.	52 <sup>i)</sup>
BAB1.1k-3.3k-1.5		3300	1.5	4400	1.5	75.0	Viscous liquid	47	53 <sup>j)</sup> , 57 <sup>i)</sup>
BAB1.1k-3.4k-3.5		3400	3.5	4500	1.3	75.6	Powdery solid	43	54 <sup>k)</sup>
BAB1.1k-3.7k-4.7	$550 \times 2$	3700	4.7	4800	1.3	77.1	Sticky solid	45	56 <sup>k)</sup>
BAB1.1k-3.8k-3.9		3800	3.9	4900	1.3	77.6	Powdery solid	42	58 <sup>k)</sup>
BAB1.1k-4.0k-3.1		4000	3.1	5100	1.5	78.4	Sticky solid	41	55 <sup>k</sup> )
BAB1.1k-4.3k-3.7		4300	3.7	5400	1.6	79.6	Powdery solid	N.D.	45 <sup>i)</sup>
BAB1.1k-4.9k-3.9		4900	3.9	6000	1.3	81.7	Powdery solid	N.D.	37 <sup>i)</sup>
BAB1.1k-5.6k-3.5		5600	3.5	6700	_	83.6	Powdery solid	- (insoluble)	- (insoluble)

Table S1. Characterization of PEG-PCGA-PEG triblock copolymers (BAB series)

a) as indicated by the supplier.

b) total molecular weight of PCGA segment connected with urethane bonds estimated by <sup>1</sup>H-NMR.

c) molar ratio of CL to GA found in the polymer, estimated by <sup>1</sup>H-NMR.

d) number-average molecular weight estimated by <sup>1</sup>H-NMR.

e) polydispersity index estimated by SEC.

f) Weight content of PCGA segment =  $[(MW \text{ of total polymer}) - (MW \text{ of PEG unit})] / (MW \text{ of total polymer}) \times 100 (\%)$ 

g) sol-to-gel transition temperature determined by test tube inverting method.

h) transition temperature other than sol-to-gel shown as i)-k).

i) sol-to-precipitate transition.

j) gel-to-sol transition.

k) gel-to-precipitate transition.

N.D.: not detected, -: not determined.

Code	Terminal group	DS of terminal groups (%) b)	Morphology	$T_{\text{gel}}(^{\mathbf{o}}\mathbf{C})^{c)}$	$T_{\rm sol}({}^{\mathbf{o}}{\rm C}){}^{\rm d)}$	$T_{\rm prec} (^{\rm o}C)^{\rm e)}$
ABA-Ac <sub>0</sub> (ABA1.5k-3.1k-3.4) <sup>a</sup>	None (-OH)	0	Powdery solid	42	45	53
ABA-Ac <sub>22</sub>		22.3	Powdery solid	40	45	57
ABA-Ac <sub>36</sub>	t- 1	36.5	Powdery solid	37	45	51
ABA-Ac <sub>52</sub>	acetyr	52.0	Powdery solid	36	46	54
ABA-Ac <sub>96</sub>		95.8	Sticky solid	34	48	51
ABA-Bu <sub>12</sub>		12.0	Powdery solid	39	44	53
ABA-Bu <sub>36</sub>		35.7	Powdery solid	35	44	50
ABA-Bu <sub>46</sub>		46.0	Powdery solid	33	46	50
ABA-Bu <sub>69</sub>	butyryl	68.7	Powdery solid	29	N.D.	38 <sup>f)</sup>
ABA-Bu <sub>82</sub>		82.3	Powdery solid	22	N.D.	35 <sup>f)</sup>
ABA-Bu <sub>89</sub>		89.2	Powdery solid	18	N.D.	34 <sup>f)</sup>
ABA-Bu <sub>91</sub>		91.3	Powdery solid	16	N.D.	34 <sup>f)</sup>

Table S2. Characterization of partially acetylated or butyrylated PCGA-PEG-PCGA triblock copolymers (ABA-Ac series and ABA-Bu series)

a) base polymer used to prepare ABA-Ac and ABA-Bu; MW of PEG = 1540, MW of PCGA =  $1550 \times 2$ , CL/GA ratio = 3.4, total  $M_n$  = 4640,  $M_w/M_n$  = 1.3

b) degree of substitution of acetyl or butyryl groups (%).

c) sol-to-gel transition temperature determined by test tube inverting method.

d) gel-to-sol transition.

e) sol-to-precipitate transition otherwise cited.

f) gel-to-precipitate transition.

N.D.: not detected.

Code <sup>a)</sup>	MW of PCGA (g/mol) <sup>b)</sup>	CL/GA (mol/mol) <sup>c)</sup>	Total <i>M</i> <sub>n</sub> (g/mol) <sup>d)</sup>	$M_{\rm w}/M_{\rm n}^{\rm e)}$	PCGA% <sup>f)</sup>	Morphology	$T_{\rm gel}({}^{\rm o}{\rm C})^{{ m g})}$	$T_{\rm prec}  (^{\rm o}{\rm C})^{\rm h)}$
4-arm5.0k-3.6k	900 × 4	4.1	8600	1.5	42	Sticky solid	N.D.	60 <sup>i)</sup>
4-arm5.0k-7.2k	$1800 \times 4$	3.6	12 200	1.6	59	Sticky solid	57	60
4-arm5.0k-7.6k	$1900 \times 4$	3.4	12 600	1.9	60	Powdery solid	53	57
4-arm5.0k-8.0k	$2000 \times 4$	4.1	13 000	1.7	62	Powdery solid	52	58
4-arm5.0k-8.8k	$2200 \times 4$	3.8	13 800	1.9	64	Powdery solid	63	66
4-arm5.0k-12.0k	$3000 \times 4$	3.4	17 000	2.2	71	Powdery solid	41	58
4-arm5.0k-12.4k	3100 × 4	3.3	17 400	2.0	71	Powdery solid	47	55
4-arm5.0k-13.6k	$3400 \times 4$	3.2	18 600	1.1	73	Powdery solid	42	51
4-arm5.0k-16.0k	$4000 \times 4$	3.2	21 000	1.1	76	Powdery solid	41	64
4-arm5.0k-20.0k	5000 × 4	4.0	25 000	2.5	80	Powdery solid	- (insoluble)	- (insoluble)

Table S3. Characterization of 4-arm PEG-PCGA copolymers (4-arm series)

a) MW of 4-arm PEG = 5000.

b) estimated by <sup>1</sup>H-NMR.

c) molar ratio of CL to GA found in the polymer, estimated by <sup>1</sup>H-NMR.

d) number-average molecular weight estimated by <sup>1</sup>H-NMR.

e) polydispersity index estimated by SEC.

f) weight content of PCGA segment = [(MW of total polymer) - (MW of PEG unit)] / (MW of total polymer) × 100 (%)

g) sol-to-gel transition temperature determined by test tube inverting method.

h) gel-to-precipitate transition otherwise cited.

i) sol-to-precipitate transition.

N.D.: not detected, -: not determined.

		-		
Code	DS of acetyl group <sup>b)</sup>	Morphology	$T_{\text{gel}}(^{\mathbf{o}}\mathrm{C})^{\mathrm{c})}$	$T_{\rm prec} (^{\rm o}C)^{\rm d}$
4-arm-Ac <sub>0</sub> (4-arm5.0k-7.6k) <sup>a)</sup>	0.0	Powdery solid	53	57
4-arm-Ac <sub>18</sub>	17.5	Powdery solid	49	62
4-arm-Ac <sub>38</sub>	37.6	Sticky solid	49	65
4-arm-Ac <sub>56</sub>	56.5	Sticky solid	46	65
4-arm-Ac <sub>66</sub>	65.7	Sticky solid	44	64
4-arm-Ac95	94.7	Powdery solid	44	61

Table S4. Characterization of partially acetylated 4-arm PEG-PCGA copolymers (4-arm-Ac series)

a) base polymer used to prepare 4-arm-Ac; MW of 4-arm PEG = 5000, MW of PCGA =  $1900 \times 4$ , CL/GA ratio = 3.4, total  $M_n = 12600$ ,  $M_w/M_n = 1.9$ 

b) degree of substitution of acetyl groups (%).

c) sol-to-gel transition temperature determined by test tube inverting method.

d) gel-to-precipitate transition.

Code	$mp_1$	$\Delta H_1$	$mp_2$	$\Delta H_2$	mp <sub>3</sub>	$\Delta H_3$	$\Delta H_{\rm total}$		CI %	Xc	$T_{\rm gel}$	Mambalagy
	[°C ]	[J/g]	[°C]	[J/g]	[°C ]]	[J/g]	[J/g]	PCGA70	CL70	[%]	[°C]	Worphology
ABA1.5k-3.1k-3.4	23.0	46.0	-	-	-	-	46.0	66.8	52.1	63.3	42	Powdery solid
ABA1.5k-3.5k-3.4	21.9	44.3	-	-	-	-	44.3	66.9	54.1	58.7	38	Powdery solid
ABA1.5k-3.8k-3.1	22.6	16.4	27.4	1.0	-	-	17.4	71.2	54.2	23.0	33	Powdery solid
ABA1.5k-3.8k-3.5	20.7	13.5	27.7	4.4	-	-	17.9	71.2	55.8	23.0	33	Powdery solid
ABA1.5k-4.0k-3.7	20.9	15.5	34.5	2.0	-	-	17.5	72.2	57.3	21.9	33	Powdery solid
ABA1.5k-4.6k-3.5	21.8	11.4	30.1	9.9	-	-	21.3	74.9	58.7	26.0	insoluble	Powdery solid
BAB1.1k-3.3k-1.5	9.6	0.4	-	-	-	-	0.4	75.0	45.0	0.6	47	Viscous liquid
BAB1.1k-3.4k-3.5	22.1	5.7	29.6	5.4	-	-	11.1	75.6	58.8	13.5	43	Powdery solid
BAB1.1k-3.8k-3.9	27.1	8.5	35.5	17.8	-	-	26.3	77.6	61,7	30.6	42	Powdery solid
BAB1.1k-4.0k-3.1	14.0	0.9	21.3	1.0	29.8	14.5	16.4	78.4	59.3	19.8	41	Sticky solid

**Table S5.** Results of differential scanning calorimeter (DSC) analysis for PCGA-PEG-PCGA triblock copolymers (ABA series) and PEG-PCGA-PEG triblock copolymers (BAB series)



Scheme S1. Synthesis of PCGA-PEG-PCGA triblock copolymers (ABA series).



Scheme S2. Synthesis of PEG-PCGA-PEG triblock copolymer (BAB series).



**Scheme S3.** Synthesis of partially acetylated or butyrylated PCGA-PEG-PCGA (ABA-Ac series and ABA-Bu series).



**Scheme S4.** Synthesis of 4-arm PEG-PCGA (4-arm series) and partially acetylated 4-arm PEG-PCGA (4-arm-Ac series).



Figure S1(a). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.5k-2.4k-3.6.



Figure S1(b). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.5k-3.0k-3.5.



Figure S1(c). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.5k-3.1k-3.4.



Figure S1(d). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.5k-3.5k-3.4.



Figure S1(e). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.5k-3.6k-4.8.



Figure S1(f). <sup>11</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.5k-3.8k-3.5.



Figure S1(g). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.5k-4.0k-3.7.



Figure S1(h). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.5k-4.6k-3.5.



Figure S1(i). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.0k-2.0k-4.1.



Figure S1(j). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA1.0k-2.6k-3.6.



Figure S1(k). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA2.0k-4.6k-2.7.



Figure S2(a). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for BAB1.1k-2.1k-2.4.



Figure S2(b). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for BAB1.1k-2.5k-4.4.



Figure S2(c). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for BAB1.1k-2.9k-3.6.



Figure S2(d). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for BAB1.1k-3.7k-4.7.



Figure S2(e). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for BAB1.1k-3.8k-3.9.



Figure S2(f). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for BAB1.1k-4.3k-3.7.



Figure S3(a). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA-Ac<sub>36</sub>.



Figure S3(b). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA-Ac<sub>52</sub>.



Figure S3(c). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA-Ac<sub>96</sub>.



Figure S3(d). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA-Bu<sub>36</sub>.



Figure S4(a). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA-Bu<sub>46</sub>.



Figure S4(b). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for ABA-Bu<sub>91</sub>.



Figure S5(a). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for 4-arm5.0k-3.6k.



Figure S5(b). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for 4-arm5.0k-7.6k.



**Figure S5(c).** <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for 4-arm5.0k-12.0k.



Figure S5(d). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for 4-arm5.0k-13.6k.



Figure S6(a). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for 4-arm-Ac<sub>18</sub>.



Figure S6(b). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for 4-arm-Ac<sub>56</sub>.



Figure S6(c). <sup>1</sup>H-NMR spectrum (in CDCl<sub>3</sub>) and elution profile of SEC for 4-arm-Ac<sub>95</sub>.



**Figure S7.** Photographs of PCGA-PEG-PCGA triblock copolymers (ABA1.5k series) synthesized with PEG<sub>1540</sub>.

#### ABA1.0k-1.8k-3.9 ABA1.0k-2.0k-3.7 ABA1.0k-2.0k-4.1







Sticky solid ABA1.0k-3.0k-3.4

Sticky solid



Sticky solid

Sticky solid

Figure S8. Photographs of PCGA-PEG-PCGA triblock copolymers (ABA1.0k series) synthesized with PEG<sub>1000</sub>.



Figure S9. Photographs of PEG-PCGA-PEG triblock copolymers (BAB series) synthesized.



**Figure S10.** Photographs of partially acetylated or butyrylated PCGA-PEG-PCGA (ABA-Ac series and ABA-Bu series) synthesized.







**Figure S12.** Photographs of partially acetylated 4-arm PEG-PCGA copolymers (4-arm-Ac series) synthesized.



Figure S13. Typical examples of phase diagram in PBS (pH = 7.4).





**Figure S14.** Typical examples of temperature-dependent storage (G', closed symbols) and loss moduli (G'', open symbols) estimated by reoplogical measuremants in PBS (pH = 7.4).



**Figure S15.** Effects of PCGA segment (PCGA%) content and CL/GA ratios on linear block copolymer morphology and transition modes.

PCGA% = [(MW of total polymer) - (MW of PEG unit)] / (MW of total polymer) × 100 (%)
Closed circle, triangle, and square: ABA1.5k series (MW of PEG = 1540); closed diamond:
ABA1.0k series (MW of PEG = 1000); double circle: ABA2.0k; open symbols: BAB series.
Circle: powdery solid, triangle and diamond: sticky solid; square: viscous liquid. Red, sol-to-gel; green, turbid sol-to-precipitate; blue, clear sol-to-precipitate; black, insoluble.



**Figure S16.** Relationship between gelation temperature ( $T_{gel}$ ) and molar ration of CL unit to GA unit (CL/GA) for ABA and BAB triblock copolymers with similar MW PCGA segment length. For ABA series, copolymers with MW of PEG = 1.5k and MW of PCGA = 3.6k - 4.0k were shown. For BAB series, copolymers with MW of PCGA = 3.3k - 4.0k were shown. Closed red circle: ABA1.5k series (MW of PEG = 1540), open red circle: BAB series.



**Figure S17.** Results of differential scanning calorimetry for (A) ABA1.5k series and (B) BAB1.1k series, and (C) plots of  $T_{gel}$  vs. crystallinity (Xc). Closed symbol: ABA series and open symbols: BAB series. For each plot, sequence and PCGA% and CL/GA ratio were indicated in the blackets. The original data were shown in **Table S6**.



**Figure S18.** Comparison of the effect of MW of one PCGA segment (g/mol) on the gelation temperature ( $T_{gel}$ ) for linear ABA triblock (ABA1.5k series) and 4-arm branched PEG-PCGA copolymers (4-arm series).

Circle: sol-to-gel transition, triangle: no gelation (sol-to-precipitate), square: insoluble. Blue: ABA1.5k series (MW of PEG = 1540); red:4-arm series.

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